

1 Article

2 Oxidative/antioxidative status in patients after 3 myocardial infarction and in those without 4 cardiovascular event depending on anthropometric 5 factors associated with overweight and obesity

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20 **Abstract:** Obesity is one of the factors leading to the development of atherosclerosis. This metabolic
21 disorder is associated with an increased production of reactive oxygen species, which affect the
22 oxidative stress level. The aim of this study was to evaluate oxidative/antioxidative status and to
23 investigate the correlation between redox markers and anthropometric parameters and body
24 composition in adult patients after myocardial infarction and in individuals without a
25 cardiovascular event in the past. Descriptive data on socio-demographic, clinical, and
26 anthropometric features and blood samples were collected and categorized into two equal groups:
27 after myocardial infarction (study group (SG), n = 80) and without a cardiovascular event (control
28 group (CG), n = 80). The oxidative/antioxidative status was assessed in plasma on the basis of total
29 oxidative/capacitive status (PerOx), total antioxidative status/capacity (ImAnOx), and oxidized
30 low-density lipoprotein (oxLDL). OxLDL was significantly higher in the CG group compared to the
31 SG group ($p = 0.02$). No significant differences were found with regard to PerOx and ImAnOx
32 values between the studied groups. Significant positive correlation between PerOx and percentage
33 of adipose tissue (FM [%]) and body adiposity index (BAI) was found in the two studied groups.
34 ImAnOx significantly positively correlated with VAI in SG and FM% in CG. OxLDL negatively
35 correlated with body mass index and waist to hip circumference ratio in CG. The total
36 oxidative/antioxidative status is related to the amount of adipose tissue and the BAI of the subjects.
37 It was observed that it correlates more frequently with the visceral distribution of body fat.

38 **Keywords:** oxidative status; antioxidant status; oxidative stress; cardiovascular diseases;
39 overweight, obesity

40

41 1. Introduction

42 Cardiovascular diseases (CVDs) constitute a major health problem worldwide, and reducing
43 mortality due to these diseases and their consequences is one of the important priorities for health
44 care industries in the developed countries [1]. Atherosclerosis underlies most CVDs and is a disease

45 that is characterized by multifactorial etiopathogenesis [2]. The origin and course of atherosclerosis
46 is strongly influenced by oxidative stress in the blood vessel wall, which is defined as an imbalance
47 between the production of reactive oxygen species (ROS) in cells and the antioxidant capacity of the
48 organism [3]. Oxidative stress is responsible for causing oxidative damage to lipids, proteins, and
49 nucleic acids as well as results in the modification of their structures and functional properties [4,5].
50 ROS include small and highly reactive compounds such as free radicals containing an unpaired
51 electron, i.e., a peroxide anion (O₂⁻) and hydroxyl radical (OH[•]), and hydrogen peroxide (H₂O₂),
52 which is not a free radical. Physiologically, ROS participate in many important cellular processes
53 such as growth, proliferation, differentiation, apoptosis, gene expression regulation, protein
54 phosphorylation, and immune defense mechanisms. However, in case of oxidative/antioxidative
55 balance disorders, ROS is produced in excess and contribute to many pathological processes
56 involved in the formation and progression of atherosclerosis. There are three types of ROS activities.
57 Oxidative stress induces strong oxidation and causes damage to proteins, lipids, phospholipids, cell
58 membranes, and DNA, thus contributing to cell dysfunction and destruction. ROS produced in
59 excess react with nitric oxide (NO), which impairs the bioavailability of NO and thus weakens the
60 vasodilatation function of the endothelium. Moreover, ROS modulate the activity of many cellular
61 proteins and signal pathways (redox signaling), thus inducing specific acute or chronic changes in
62 the phenotype and functioning of the cells [6]. The sources of ROS in the cell include: respiratory
63 chain, xanthine oxidase, uncoupled endothelial NO synthase, cyclooxygenase, myeloperoxidase, or
64 lipoxidase [7]. Among these sources, enzymes belonging to the NADPH oxidase family seem to be
65 the key enzymes responsible for ROS production in the cells of vascular walls [6]. Many studies
66 demonstrated that oxidative stress is associated with CVDs and that many atherosclerosis risk
67 factors influence the level of oxidative stress [8–10].

68 Obesity, i.e., an excessive accumulation of adipose tissue, is considered to be one of the CVD
69 risk factors and at the same time a factor influencing oxidative stress level [11,12]. Adipose tissue,
70 the organ that affects energy homeostasis of the organism, is mainly composed of adipocytes and
71 other cells (e.g., fibroblasts, fibroblastic pre-adipocytes, and endothelial and immune cells) that
72 secrete hormones and cytokines (adipokines or adipocytokines), which subsequently exert
73 endocrine, paracrine, and autocrine effects in the organism. Production of excess energy results in
74 energy accumulation in adipocytes, hypertrophy of adipose tissue, and its hyperplasia. Obesity
75 alters metabolic and endocrine functions of adipose tissue and leads to increased release of
76 hormones, fatty acids, and proinflammatory molecules that contribute to obesity-related
77 complications [13]. In physiological states, and even more so in pathological ones, adipokines induce
78 ROS production, affecting oxidative stress formation. Several mechanisms are involved in the
79 development of oxidative stress in obese individuals, and the mechanisms related to oxidative stress
80 formation are strongly associated with pro-inflammatory processes that cause endothelial damage
81 and excessive formation of free radicals. The presence of excessive adipose tissue was found to be a
82 source of pro-inflammatory cytokines, including tumor necrosis factor alpha, interleukin (IL)-1 β and
83 IL-6 [14], C-reactive protein, leptin, and resistin [15].

84 The formation of ROS and NO is an inseparable phenomenon accompanying biochemical
85 changes occurring in the human body, which under hemostasis conditions has developed
86 mechanisms to protect biomolecules against harmful effects of free radicals. Protective functions are
87 performed by enzymes like peroxide dismutase, catalase, and glutathione peroxidase; water;
88 fat-soluble antioxidants (e.g., glutathione, ascorbate (vitamin C), α -tocopherol (vitamin E), and
89 β -carotene); and endogenous antioxidants (e.g., albumin, bilirubin, and uric acid) [16,17]. Studies
90 show that mitochondria of white adipose tissues, especially in obese individuals, are the main sites
91 of ROS generation with participation of an increased expression of NADPH and decreased expression
92 of antioxidant enzymes [18]. Oxidative damage to important cellular structures is one of the factors
93 responsible for the development of obesity-related complications such as atherosclerosis,
94 hypertension, insulin resistance, and type 2 diabetes [19,20].

95 Many markers are currently used to evaluate oxidative and antioxidative status in an
96 individual. These include total oxidant capacity (TOC), total antioxidant capacity (TAC), oxidative

97 stress index, which expresses the TOC/TAC ratio, and oxidized low-density lipoproteins (oxLDLs),
98 which are lipid peroxidation metabolites [21–24]. Although there are many studies on oxidative
99 stress in obese adults in the literature, to our knowledge there are no reports on this topic in the
100 group of patients who were hospitalized during the beginning phase of cardiovascular rehabilitation
101 program following myocardial infarction and were continuing with motor, diet, and
102 pharmacological therapy. Therefore, the aim of this study was to assess the oxidative/antioxidative
103 status and to investigate the correlation between redox markers and anthropometric parameters and
104 body composition in adult patients after an episode of myocardial infarction and in those who did
105 not suffer from a cardiovascular event previously. The first hypothesis was that people without a
106 cardiovascular event in the past have higher levels of antioxidative markers and lower levels of
107 oxidative ones. The second hypothesis was that the oxidative/antioxidative status is associated with
108 obesity parameters, especially those describing the visceral distribution of adipose tissue.

109 2. Materials and Methods

110 2.1. Study design and population

111 A cross-sectional study was conducted during the period from August to December 2017
112 among 160 adults divided into two equal groups: study group and control group. Study group (SG)
113 included patients who after suffering from a myocardial infarction were hospitalized during the
114 early period of cardiac rehabilitation (up to 14 days after discharge from full revascularization) and
115 who continued with physiotherapy, dietary, and pharmacological therapy in the “Uzdrowisko
116 Nałęczów” S.A. Health Resort in Nałęczów and in the Railway Health Resort Hospital in Nałęczów
117 (in eastern Poland). The study included patients from consecutive rehabilitation periods, which
118 were held at 21- or 28-day intervals. All patients in this group were treated with primary
119 percutaneous coronary intervention after the first myocardial infarction. The criteria for inclusion in
120 the SG were as follows: age 40–65 years, condition after myocardial infarction, professionally active
121 persons, and provide written consent to participate in the study. Exclusion criteria were as follows:
122 renal failure, cancer, history of pulmonary or rheumatic diseases, and age under 40 years and over
123 65 years. Other exclusion criteria included factors that may influence oxidative status, such as
124 infection (e.g., respiratory tract infections) and antioxidant vitamin intake. Research manuscripts
125 reporting large datasets that are deposited in a publicly available database should specify where the
126 data have been deposited and provide the relevant accession numbers. If the accession numbers
127 have not yet been obtained at the time of submission, please state that they will be provided during
128 review. They must be provided prior to publication.

129 Control group (CG) included professionally active adults without a cardiovascular event in the
130 past but reporting for follow-up examinations to the occupational physician as part of periodic
131 examinations. Respondents were recruited from the Provincial Center for Occupational Medicine of
132 the Center for Prophylaxis and Therapy in Lublin (eastern Poland). The criteria for inclusion in the
133 study were as follows: age between 40 and 65 years, no history of a cardiovascular event, low
134 10-year risk of a cardiovascular event (SCORE index < 5) [25], professional status (working person),
135 no chronic diseases (renal failure, cancer, rheumatism, and lung diseases), no cardiovascular
136 ailments that could suggest the existence of atherosclerotic CVD, no acceptance of preparations
137 modifying the risk of atherosclerosis, and receiving no therapy for hypertension, pre-diabetes
138 mellitus, and hypercholesterolemia. The exclusion criteria were as follows: active infection, intake of
139 drugs or dietary supplements that may affect oxidative status (e.g., vitamins), and long-term use of
140 fruit and vegetable diets only [26].

141 During the initial visit, all participants were investigated using an extensive questionnaire,
142 interview, physical examination, and performing additional tests (anthropometric measurements),
143 and the next day after the patient was prepared (12 hours of fasting), laboratory tests were carried
144 out.

145 The research project received a positive opinion from the Bioethics Committee at the Medical
146 University of Lublin (KE-0254/197/2017) and was conducted in accordance with the Helsinki

147 Declaration. All respondents were presented with the purpose of the study and then asked for
148 written consent to participate in the study.

149 2.2. Anthropometric measurements

150 Anthropometric measurements of body height and weight were performed in all patients.
151 Height was measured with the accuracy of 0.1 cm using an altimeter and body weight was measured
152 without shoes and topwear using a platform scale with the accuracy of 0.1 kg. Then, the body mass
153 index (BMI) index, defined as body weight in kilograms (kg) divided by the height in square meters
154 (kg/m^2), was calculated for all subjects [27]. Non-flexible measuring tape was used to measure the
155 waist circumference (WC), between the lower edge of the ribbed arch and the upper comb of the hip
156 bone, and the hip circumference (HC), at the level of the curve of the larger femur. Both
157 measurements were taken in a standing position. Then, the ratios of waist to hip circumference
158 (WHR) and waist to height circumference (WHtR) were calculated [28].

159 The percentage of adipose tissue content (FM [%]) was assessed using an electrical
160 bioimpedance method with a body composition analyzer (OMRON Model BF306) according to the
161 manufacturer's algorithm.

162 Visceral adiposity index (VAI) and body adiposity index (BAI) were calculated for all
163 respondents on the basis of anthropometric measurements and biochemical results, but VAI was
164 calculated separately for women and men. VAI for women = $[\text{WC}/(36.58 + (1.89 \times \text{BMI}))] \times (\text{TG} \times 0.81)$
165 $\times (1.52/\text{HDL})$; VAI for men = $[\text{WC}/(39.68 + (1.88 \times \text{BMI}))] \times (\text{TG}/1.03) \times (1.31/\text{HDL})$ [29], where TG
166 represents triglycerides and HDL represents high-density lipoprotein. $\text{BAI} = [\text{HC} (\text{cm}) / \text{height} (\text{m})$
167 $1.5] - 18$ [30,31].

168 2.3. Blood sample

169 Blood samples were taken from the elbow vein of fasting subjects in the morning (7:00–9:00 am)
170 after an overnight rest in two tubes with clotting activator and separating agent (granules) and
171 delivered to the laboratory within 1 hour. Samples were stored at 4°C until the blood sample was
172 delivered to the laboratory. The plasma was separated by centrifugation at a rate of 3.000 rpm for 10
173 minutes. The serum from one tube was used for biochemical assays, while serum from the second
174 sample was transferred to Eppendorf tubes immediately after centrifugation, then frozen at -80°C,
175 and stored until the markers of oxidative/antioxidative status were determined.

176 Centrifuged serum from one of the tubes was used to determine the lipid profile (total
177 cholesterol (TC), triglycerides (TG), high-density lipoprotein cholesterol (HDL-C)), serum glucose,
178 and creatinine using standard laboratory methods. Low-density lipoprotein cholesterol was
179 calculated from the Friedewald formula (when $\text{TG} < 400 \text{ mg}/\text{dL}$), estimated glomerular filtration rate
180 from the Cockcroft-Gault formula, and non-HDL from the non-HDL formula = $\text{TC} [\text{mg}/\text{dL}] - \text{HDL-C}$
181 $[\text{mg}/\text{dL}]$ [32].

182 2.4. Determination of oxidative/antioxidative status markers

183 Oxidative stress markers such as total oxidative/capacitive status (PerOx (TOS/TOC)) and total
184 antioxidative status/capacity (ImAnOx (TAS/TAC)) were evaluated using photometric technique.
185 Quantitative immunoenzymatic method was used to measure oxidized LDL (oxLDL). The
186 determinations were performed by ELISA tests using the original reagents (Immun Diagnostik,
187 Bensheim, Germany).

188 2.5. Statistical analysis

189 Continuous variables are summarized by means \pm standard deviations or by median and
190 interquartile range (q1–q3). Categorical data are presented as number and percentage (%).
191 Comparison of variables of interest between clinical and control study groups was done using *t*-test
192 or Mann–Whitney test for continuous variables and using chi-squared test for categorical variables.
193 Due to skewness in the antioxidant values of oxLDL, PerOX values (right-skewed) were natural

194 logarithm transformed and the ImAnOx values (left-skewed) were squared. Associations between
 195 log oxLDL, log PerOx, and square of ImAnOx and anthropometric variables were assessed by
 196 Pearson correlations. Additionally series of liner regression models were performed to take into
 197 account the influence of gender (model I), age (model II), or smoking status (model III) on tested
 198 relationships. All analysis were performed in whole sample and in study and control group
 199 (supplementary file). Statistical analysis was performed using SPSS (version 25) software. A p -value
 200 < 0.05 was considered to be statistically significant.

201 3. Results

202 3.1. Characteristics of Participants

203 Table 1 presents the characteristics of the studied groups. The study involved 160 patients who
 204 are divided into two groups. The mean age in SG was 53.34 ± 4.74 years, and in CG it was 49.25 ± 6.23
 205 years. No significant differences were observed in terms of gender, marital status, and family history
 206 of CVD in the studied groups ($p > 0.05$). Taking weight parameters into account, SG was
 207 characterized by significantly higher BMI, WHR, WHtR, and VAI values ($p < 0.05$) compared to those
 208 observed in CG.

209 **Table 1.** Baseline characteristics of the study population.

Variables	Study group ($n = 80$)	Control group ($n = 80$)	p
Demographic and clinical data:			
Age [years] ^b	53.34 ± 4.74	49.25 ± 6.23	< 0.001
Sex (Female vs. Male) ^a	22 (27.5) vs. 58 (72.5)	28 (35) vs. 52(65)	0.32
Marital status (free vs. in relationship) ^a	16 (20) vs. 64 (80)	11 (13.7) vs. 69 (86.3)	0.398
Clinical variables:			
Family history of CVD - on the mother's side ^a	46 (57.5)	42 (52.5)	0.634
Family history of CVD - on the father's side ^a	49 (61.3)	39 (48.8)	0.153
Diabetes ^a	19 (24)	1 (1,3)	< 0.001
Arterial hypertension ^a	54 (68)	9 (11)	< 0.001
Smoking ^a	25 (31.25)	9 (11.25)	< 0.001
Anthropometric variables:			
BMI [kg/m ²] ^b	28.89 ± 4.91	26.64 ± 4.04	0.002
WC [cm] ^b	103.9 ± 12.48	93.36 ± 12.50	< 0.001
HC [cm] ^b	105.0 ± 10.83	102.9 ± 7.72	0.15
WHR ^b	0.99 ± 0.08	0.91 ± 0.09	< 0.001
WHtR ^b	0.60 ± 0.07	0.54 ± 0.07	< 0.001
FM [%] ^b	31.08 ± 7.71	29.65 ± 7.05	0.22
VAI ^b	2.12 ± 1.56	1.06 ± 0.7	< 0.001
BAI ^b	28.63 ± 6.44	27.79 ± 4.73	0.35

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Table 1. Cont.

Variables	Study group (n = 80)	Control group (n = 80)	p
Biochemical parameters:			
Total cholesterol [mg/dL] ^b	147.81 ± 37	221.98 ± 44.2	< 0.001
Triglyceride [mg/dL] ^c	134.5 (103.25 - 174.35)	102.9 (73.4 - 141.15)	< 0.001
HDL-C [mg/dL] ^b	46.16 ± 11.50	64.05 ± 19.14	< 0.001
non-HDL [mg/dL] ^b	101.5 ± 34.31	157.9 ± 47.22	< 0,001
LDL-C [mg/dL] ^b	70.24 ± 25.86	134.81 ± 42.37	< 0.001
Glucose [mg/dL] ^c	102.5 (97.0-112.0)	102 (97.5-109.5)	0.78
Creatinine [mg/dL] ^b	0.85 ± 0.15	0.86 ± 0.18	0.97
eGFR [ml/min/1.73 m ²] ^b	93.5 ± 12.56	95.4 ± 13.33	0.35

212 Date are presented as: ^a n(%); ^b mean ± SD; ^c median (Q1-Q3).

213 BMI: body mass index; WC: waist circumference; HC: hip circumference; WHR: waist to hip ratio; WHtR:
 214 waist-to-height ratio; FM [%]: body fat percentage; VAI: visceral adiposity index; BAI: body adiposity index;
 215 HDL-C: high-density lipoprotein; LDL-C: low-density lipoprotein.

216 3.2. Oxidative/antioxidative status

217 Table 2 shows the oxidative/antioxidative status in the studied groups. There were no
 218 significant differences in PerOx levels between SG and CG patients, although the median value of
 219 this parameter was higher in CG and amounted to 718.01 μmol/L compared to SG, in which the
 220 median value was 699.44 μmol/L. Similarly, no significant differences were observed in the ImAnOx
 221 levels between the groups (p = 0.35). However, the higher median of this parameter was
 222 characteristic for SG (293.8 μmol/L) compared to CG (276.09 μmol/L). On the other hand, oxLDL was
 223 significantly higher in the group of patients without a cardiovascular event than in the group of
 224 patients after myocardial infarction (p = 0.02).

225 Figure 1 and Figure 2 illustrate a comparison of the oxidative / antioxidative status distribution
 226 in gender groups and depending on the smoking status of the entire sample. Gender significantly
 227 differentiated the overall oxidation / capacity (PerOx) status, higher values were found in women
 228 compared to men (p < 0.001). In other cases no significant differences were found.

229 **Table 2.** Oxidative/antioxidative status markers comparison of studied groups.

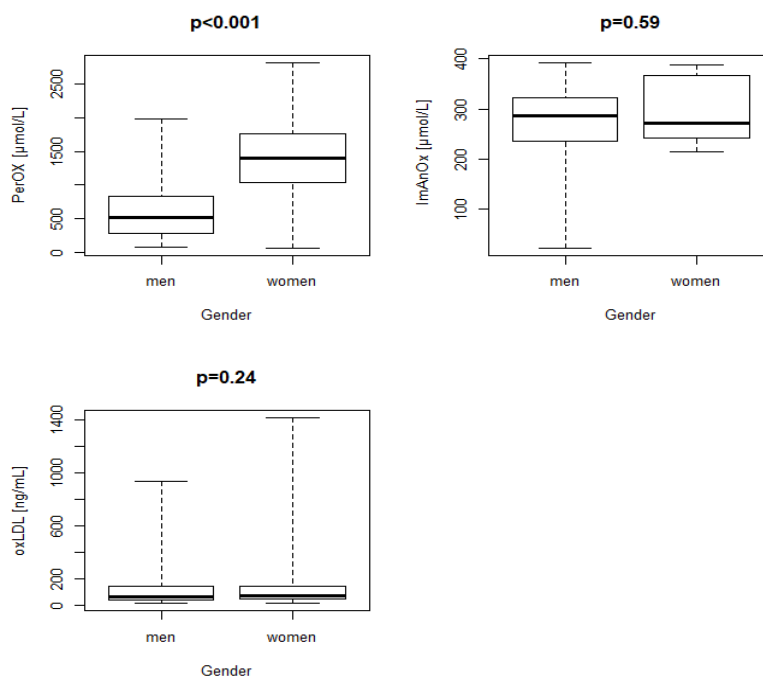
Variables	Study group (n = 80)	Control group (n = 80)	p
PerOx (TOS/TOC) [μmol/L]	699.44 (379.71 - 1152.36)	718.01 (298.35 - 1274.61)	0.77
ImAnOx (TAS/TAC) [μmol/L]	293.8 (246.52 - 317.92)	276.09 (233.76 - 333.33)	0.35
oxLDL [ng/mL]	54.25 (36.09 - 119.34)	75.91 (49.38 - 143.32)	0.02

230 PerOx (TOS/TOC): total oxidative status/capacity; ImAnOx (TAS/TAC): total antioxidative status/capacity;
 231 oxLDL: oxidized low-density lipoprotein.

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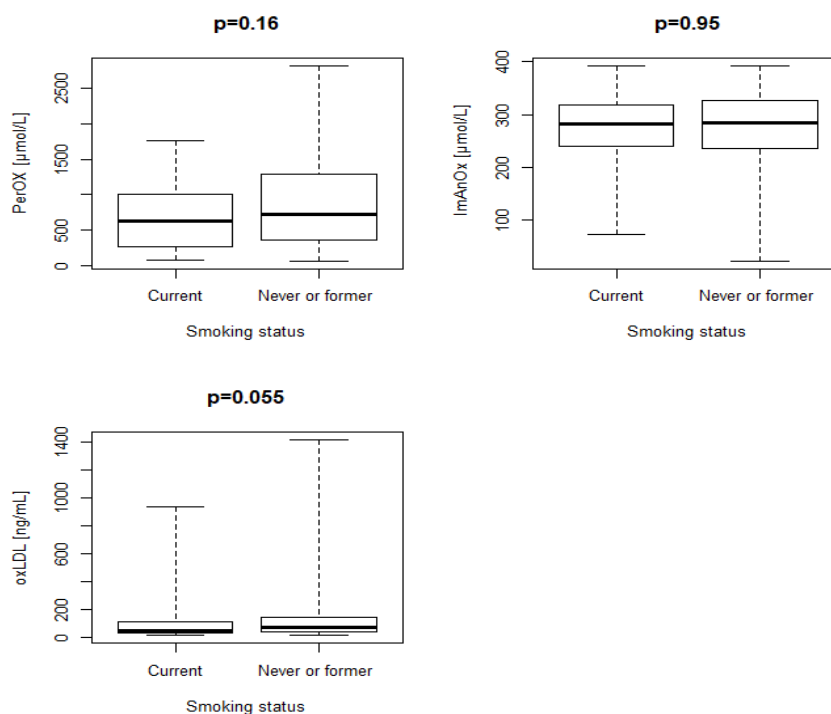
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Figure 1. Comparison of the distribution of oxidation / antioxidative status parameters between men and women ($n = 160$).

PerOx: total oxidative status/capacity; ImAnOx: total antioxidative status/capacity; oxLDL: oxidized low-density lipoprotein.



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Figure 2. Comparison of the distribution of oxidation/antioxidative status parameters between current and never or former smokers ($n = 160$).

PerOx: total oxidative status/capacity; ImAnOx: total antioxidative status/capacity; oxLDL: oxidized low-density lipoprotein.

246 3.3. Correlation between oxidative/antioxidative status and anthropometric measures

247 Table 3 shows the correlation between oxidative/antioxidative status and anthropometric
 248 factors. Significant positive correlations between PerOx and FM [%] (SG: $r = 0.400$, $p < 0.001$; CG:
 249 $r = 0.337$, $p = 0.002$) and BAI (SG: $r = 0.335$, $p = 0.002$; CG: $r = 0.309$, $p = 0.005$) were found in the
 250 two studied groups and among all the respondents (FM [%]: $r = 0.365$, $p < 0.001$; BAI: $r = 0.316$, p
 251 < 0.001). Significant negative correlation between PerOx and WHR ($r = -0.281$, $p = 0.01$) and VAI
 252 ($r = -0.327$, $p = 0.003$) was observed in CG and in the whole examined population (WHR: $r =$
 253 -0.206 , $p = 0.009$; VAI: $r = -0.214$, $p = 0.006$). ImAnOx TAS/TAC significantly positively correlated
 254 with VAI ($r = 0.391$, $p < 0.001$) in SG, with FM [%] ($r = 0.235$, $p = 0.03$) in CG, and with these two
 255 parameters in the whole group (VAI: $r = 0.18$, $p = 0.02$; FM [%]: $r = 0.154$, $p = 0.05$). OxLDL
 256 negatively correlated with WHR ($r = -0.243$, $p = 0.03$) in CG and WHR ($r = -0.210$, $p = 0.009$) in the
 257 whole examined group. After adjustment for age or smoking status, the results were similar to
 258 those obtained in the simple linear regression models (Tables S1-S3). However, after adjustment
 259 for gender, the relationships between PerOx and WHR, FM [%] and BAI lost statistical
 260 significance.

261 **Table 3.** Correlation between oxidative/antioxidative status and anthropometric measures.

Variables	Log PerOx (TOS/TOC) [$\mu\text{mol/L}$]		ImAnOx (TAS/TAC) [$\mu\text{mol/L}$]		Log oxLDL [ng/mL]	
	r	p	r	p	r	p
Study group (n = 80)						
BMI	0.118	0.30	0.114	0.31	0.047	0.68
WHR	-0.148	0.19	0.176	0.117	-0.092	0.41
WHtR	0.180	0.11	0.079	0.48	0.064	0.57
FM [%]	0.400	< 0.001	0.039	0.73	0.105	0.35
VAI	-0.122	0.28	0.391	< 0.001	-0.038	0.74
BAI	0.335	0.002	-0.056	0.62	0.127	0.26
Control group (n = 80)						
BMI	-0.093	0.41	0.004	0.97	-0.216	0.06
WHR	-0.281	0.01	0.056	0.62	-0.243	0.03
WHtR	-0.082	0.47	0.120	0.30	-0.186	0.11
FM [%]	0.337	0.002	0.235	0.03	0.036	0.75
VAI	-0.327	0.003	-0.007	0.95	-0.071	0.54
BAI	0.309	0.005	0.152	0.184	0.037	0.75
Total (n = 160)						
BMI	0.015	0.85	0.071	0.38	-0.096	0.23
WHR	-0.206	0.009	0.128	0.10	-0.210	0.009
WHtR	0.04	0.61	0.126	0.116	-0.098	0.22
FM [%]	0.365	< 0.001	0.154	0.05	0.059	0.46
VAI	-0.214	0.006	0.18	0.02	-0.087	0.28
BAI	0.316	< 0.001	0.055	0.49	0.08	0.32

262 BMI: body mass index; WHR: waist to hip ratio; WHtR: waist-to-height ratio; FM [%]: body fat percentage; VAI:
 263 visceral adiposity index; BAI: body adiposity index; PerOx (TOS/TOC): total oxidative status/capacity; ImAnOx
 264 (TAS/TAC): total antioxidative status/capacity; oxLDL: oxidized low-density lipoprotein.

265 4. Discussion

266 The relationship between oxidative stress and CVD is of immense interest to many researchers.
267 It was demonstrated that both excessive oxidative stress and inadequate antioxidative defense
268 mechanisms may cause an early onset of CVD [33]. Increased oxidative stress markers act in synergy
269 with standard risk factors for CVDs [34,35]. In this cross-sectional study, we evaluated the
270 oxidative/antioxidative status in parallel groups and examined the correlation between redox
271 markers and anthropometric parameters and body composition in the group of adult patients
272 undergoing cardiac rehabilitation after suffering from myocardial infarction and in individuals who
273 had not suffered from a cardiovascular event in the past. This is probably the first study to evaluate
274 the oxidative/antioxidative status of a group of people who enrolled in a cardiovascular
275 rehabilitation program following myocardial infarction and in those without a cardiovascular event,
276 during the subclinical development of atherosclerosis, and to identify which patients require
277 primary prevention (CG) or secondary prevention (SG) based on the risk of cardiovascular
278 complications. The results of our study did not confirm the first hypothesis. The markers of
279 oxidative/antioxidative status were better indicated in SG than in CG. The second hypothesis was
280 confirmed because our study proved that visceral fat distribution is correlated with
281 oxidative/antioxidative status, so management should place particular emphasis on weight
282 reduction.

283 Our study showed that lipid peroxidation (oxLDL) was much stronger in CG than in SG
284 patients. The result we obtained was completely different from the results obtained by
285 Dominguez-Rodriguez et al. [36], who evaluated nighttime oxLDL and melatonin levels in patients
286 with acute coronary syndrome (ACS) and healthy subjects without symptomatic atherosclerosis. In
287 this study, the group of patients with ACS had higher levels of oxLDL and lower levels of melatonin
288 than CG patients. However, in the Renko et al.'s [37] study conducted in a group of 120 men after
289 myocardial infarction and 250 men in the CG, no significant differences between groups were found
290 in the oxLDL level. The results of our study were obtained in a group of patients who after
291 suffering from myocardial infarction were receiving early cardiological rehabilitation, including
292 physiotherapy, pharmacotherapy, and dietary treatment. As the literature indicates, the occurrence
293 of myocardial infarction leads to the modification of lifestyle-related behaviors to a more
294 pro-healthy one, especially in the context of dietary changes [38]. The change of the diet to the one
295 containing less fat resulted in a decrease in oxLDL levels in the group consisting of obese women,
296 which was demonstrated by Elizabeth et al. [39] in their study, as well as in the patients who
297 participate regularly in Nordic walking, in the study by Cebula et al. [40]. Moreover, higher oxLDL
298 levels in CG in our study may be associated with higher levels of LDL-C in this group, because the
299 processes involved in the modification of this cholesterol fraction play a key role in the formation of
300 atherosclerotic plaque [41]. The above arguments may also influence the results obtained with
301 respect to other markers of oxidative status (PerOX), which was lower in SG, and antioxidative
302 status (ImAnOx), which was higher in CG, although these differences were not statistically
303 significant ($p > 0.05$). In our study, CG is characterized by higher blood lipid indices compared to
304 SG (TC, $p < 0.001$; LDL-C, $p < 0.001$; and non-HDL, $p < 0.001$). The available literature data indicate a
305 close correlation between the development of atherosclerotic plaque, determined by the thickness of
306 the intima-media complex, and the concentration of not only TC but also its individual lipid
307 fractions in blood [42–45]. Selection of respondents for CG was conditioned by non-administration
308 of preparations modifying the risk of atherosclerosis; therefore, it can be assumed that high lipid
309 levels correlate with the development of atherosclerotic plaque, and the changes in vascular
310 endothelium are reflected in oxidative/antioxidative disorders.

311 Drugs taken by the patient are very important in determining the level of oxidative stress. The
312 respondents after myocardial infarction were staying in the Health Resort Hospital and each of the
313 respondents took their medicines systematically. Some of the drugs are known to influence the level
314 of oxidative stress. Aspirin, statins, angiotensin converting enzyme II (Ang II) inhibitor, and
315 metformin are commonly used in the secondary prevention of cardiovascular events and treatment
316 of concomitant diseases; they also exhibit many pleiotropic effects [46]. The Paseban et al.'s [47]

317 study showed that the combined use of the above-mentioned drugs enhances their antioxidant effect.
318 Aspirin has an antioxidant effect by reducing the production of free radicals such as peroxide and
319 prevents a decrease in the activity of antioxidant enzymes (catalase and peroxide dismutase) [48].
320 Statins, due to their antioxidant activity by inhibition of NAD(P)H oxidase and active exchange of
321 free radicals [49–51], reduce chronic inflammation [52] and thus reduce oxidative stress [53]. Ang II
322 inhibitor may selectively reduce Ang II, endothelin, and oxidative stress levels, which may
323 potentially play a role in the lowering of blood pressure [54]. Moreover, in our sample, 24% of SG
324 patients are diabetics. Metformin is the first hypoglycemic drug used in the treatment of patients
325 with type 2 diabetes, and it has been shown to reduce ROS by enhancing the activity of antioxidant
326 enzymes [55].

327 Studying the relationship between gender and oxidative/antioxidant status is vital as oxidative
328 stress is a factor in the development of numerous diseases, including cardiovascular diseases, and at
329 the same time, these diseases occur differently in men and women. It was proved that oxidative
330 stress was lower in male rats than in females [56]. Ide et al. [57] showed that biomarkers of oxidative
331 stress in vivo were higher in young men than in women of the same age. In other studies, it was
332 observed that ROS production was higher in blood vessel cells in men than in women [58], and
333 women were found to have the greater antioxidant potential [59]. Other studies indicate that there is
334 a difference in the expression and/or activity of antioxidant enzymes between men and women.
335 These enzymes are present in various body tissues. With regard to SOD, there is no uniform
336 consensus on gender differences, although it is suggested that there may be variances in different
337 tissues. Chen et al. [60] showed that the level of SOD activity in brain tissue and lungs was higher in
338 female mice, but there were no significant differences in the level of SOD activity between females
339 and males in the kidney and heart. However, Barp et al. [56] established that female rats had a higher
340 level of SOD activity in the heart than males. Interestingly, they also found that after castration, the
341 level of SOD activity in both male and female rats was significantly reduced compared to the control
342 group. The results of the cited researchers indicate that there may be a relationship between sex
343 hormones and the level of SOD activity. However, some studies have not shown differences in the
344 level of SOD activity between men and women; therefore, there are some differences regarding the
345 association of SOD activity and gender [57, 61]. Chan et al. [60] revealed that catalase activity in both
346 female and male mice was the same in the brain, lungs, and heart, but higher in females in a kidney.
347 However, some studies did not demonstrate differences in the level of catalase activity between men
348 and women [56, 57, 62]. Thus, the studies cited above indicate that gender and sex hormones do not
349 affect catalase activity, and thus, the degradation of hydrogen peroxide [63]. Several studies have
350 shown that GPx activity was lower in women than in men [56, 57, 60], although there was no
351 significant change in GPx levels after castration, suggesting that sex hormones may not affect GPx
352 [56]. The fact that GPx levels were lower in women seems counterintuitive because women are
353 thought to be less prone to oxidative stress than men. This observation suggests that women possess
354 other mechanisms to protect themselves against oxidative stress. Although there may be some
355 differences in the level of antioxidant enzyme activity between men and women, as discussed earlier,
356 the difference in antioxidant properties is probably due to estrogen [63]. This sex hormone acts as an
357 antioxidant, scavenging free radicals due to the presence of a phenolic hydroxyl group [56]. In the
358 presented research, the overall oxidation/capacitance status (PerOx) was higher in women than in
359 men. The explanations for this result can be seen in the menopausal or postmenopausal period in
360 which the women were examined, which reduced the concentration of estrogens in the studied
361 group.

362 Obesity is considered to be an important risk factor leading to the development of many
363 diseases such as ischemic heart disease, diabetes, hypertension, dyslipidemia, stroke, and some
364 types of cancer [64,65]. This metabolic disorder is associated with an increased ROS production and
365 oxidative stress formation. Increased ROS production in obese individuals is associated with
366 excessive supply of macronutrients in diet, mitochondrial dysfunction, excessive ROS production at
367 the endoplasmic reticulum level, and inflammatory response [66,67]. Obesity is one of the factors
368 leading to oxidative stress, which in turn leads to atherosclerosis of vessels and, consequently, may

369 lead to myocardial infarction. Amirkhizi et al. [68] conducted a study on obese women and found
370 that obesity, even in the absence of smoking, diabetes, kidney, and liver diseases, may reduce
371 protective antioxidant mechanisms by increasing systemic oxidative stress.

372 In our study, anthropometric and physiological measurements (WHR, FM [%], VAI, and BAI)
373 significantly correlated with the oxidative/antioxidative status in SG and CG patients, as well as in
374 the entire sample. Moreover, we discovered several significant correlations between VAI and BAI
375 and oxidative/antioxidative status in both groups. In our study, the oxidative/antioxidative status
376 was more often correlated with FM%, VAI, and BAI than with BMI. Although BMI is used to
377 measure overweight and obesity, it does not take into account factors such as body size and adipose
378 tissue distribution. Studies suggest that abdominal obesity is more strongly associated with chronic
379 diseases [69,70], because visceral fat secretes several metabolites that cause chronic diseases [71,72].
380 Chrysohoou et al. [73] concluded in their study that the total oxidative capacity (TAC) was
381 significantly correlated with the central obesity index rather than with obesity. Visceral fat
382 accumulation (measured by computed tomography), as a factor associated with enhanced oxidative
383 state, was also demonstrated by Araki et al. [74]. The importance of adipose tissue distribution in the
384 context of oxidative stress in obesity was demonstrated by other authors who found a correlation
385 between anthropometric parameters (WC and BMI) and the level of oxidative stress activation
386 [75,76].

387 4.1. Limitations

388 Our study has a few limitations. First, it is a cross-sectional study, so neither temporality nor
389 causality can be established. Second is the inclusion of a relatively small number of patients from a
390 single center, especially with regard to patients qualified to be included in SG. The third limitation of
391 our study is the use of bioelectrical impedance analysis (BIA), which is an indirect method for the
392 evaluation of body composition. However, comparative studies have shown significant correlations
393 between BIA data and body composition measured by densitometry (dual-energy X-ray
394 absorptiometry (DXA)), which is the golden standard for this type of analysis [77]. Unfortunately,
395 DXA is associated with exposure to X-rays, which limits the regular use of this method. The fourth
396 limitation of our study was the non-inclusion of drugs taken by patients in the analysis of oxidative
397 stress levels in particular groups, although the main aim of the study was to assess the parameters
398 related to obesity and oxidative/antioxidative status, but these drugs may have influenced the
399 results of the study.

400 5. Conclusions

401 To sum up, our study shows that total oxidative/antioxidative status is related to the adipose
402 tissue content and skewness BAI of the subjects, but it was observed that it correlates more
403 frequently with the visceral distribution of adipose tissue. Given the significant association of
404 visceral fat distribution with oxidative/antioxidative status, further studies are needed to investigate
405 the impact of fat distribution on the risk of cardiometabolic diseases, especially in conjunction with
406 other common cardiovascular risk factors.

407 **Supplementary Materials:** The following are available online at www.mdpi.com/xxx/s1, **Table S1:**
408 Relationship between oxidative/antioxidative status and anthropometric measures in study group, **Table S2:**
409 Relationship between oxidative/antioxidative status and anthropometric measures in control group: **Table S3:**
410 Relationship between oxidative/antioxidative status and anthropometric measures in all participants.

411 **Author Contributions:** G.J.N. and B.Š. developed the concept of the study. G.J.N. and M.P. analyzed the data
412 and contributed to its interpretation. G.J.N. interpreted the data and wrote the original draft. G.J.N., B.Š. and
413 A.P. were involved in writing, reviewing and editing the manuscript. G.J.N., M. C-K. and E R.-D. were
414 responsible for funding acquisition and supervision. All authors were involved in critically revising the
415 manuscript, and have given their approval to the manuscript submitted.

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