

Supplementary information

		Subgroups ^a	Similarity (%) ^b
OsWRKY5	WRKYGQKMAKGNPCPRAYRCTMASQCPVRKQVQRCAEDKSLILITTYEGTHSH		
AtWRKY18	WRKYGQKVTRDNPSPRAYFRCSFAPSCPVKKKVQRSAEDPSLIVATYEGTHNH	IIa	35/53 (66)
AtWRKY40	WRKYGQKVTRDNPSPRAYFKCACAPSCSVKKKVQRSVEDQSVLIVATYEGEHNH	IIa	31/53 (58)
AtWRKY6	WRKYGQKMAKGNPCPRAYRCTMATGCPVRKQVQRCAEDRSILITTYEGNHNH	IIb	48/53 (91)
AtWRKY47	WRKYGQKMAKGNPCPRAYRCTMAVGCVPVRKQVQRCAEDTTILITTYEGNHNH	IIb	46/53 (87)
AtWRKY8	WRKYGQKAVRNSPYPRSYRCTTQ-KCNVKKRVERSYQDPTVVITTYESQHNH	IIc	29/53 (55)
AtWRKY49	WRKYGQKSIKNSPNPRSYRKCTNP-ICNAKKQVERSIDESNTYILITTYEGFHH	IIc	27/53 (51)
AtWRKY7	WRKYGQKPIKGSFHPRGYYKCSSVRGCPARKHVERALDDAMMLIVTYEGDHNH	IId	30/53 (57)
AtWRKY17	WRKYGQKPIKGSFHPRGYYKCTFRGCPARKHVERALDDSTMLIVTYEGEHRH	IId	30/53 (57)
AtWRKY14	WRKYGQKPIKGSFFPRGYRCSSSKGCSARKQVERSRTPDNMLVITYTSEHNH	IIe	28/53 (53)
AtWRKY29	WRKYGQKPIKGSFYPRSYRCSSSKGCLARKQVERNPQNPEKFTITYTNEHNH	IIe	26/53 (49)
	WRKYGQK C C H H		

Fig. S1. Amino acid sequence alignments of WRKY domains between OsWRKY5 and group II AtWRKY proteins.

Amino acid sequences of OsWRKY5 and group II AtWRKY proteins were obtained from the Rice Genome Annotation Project (<http://rice.plantbiology.msu.edu/>) and The Arabidopsis Information Resource (TAIR, <https://www.arabidopsis.org/>), respectively. Sequence alignment was performed using ClustalW with default parameters. Sequences are as follows: OsWRKY5, Os05g04640; AtWRKY18, At4g31800; AtWRKY40, At1g80840; AtWRKY6, At1g62300; AtWRKY47, At4g01720; AtWRKY8, At5g46350; AtWRKY49, At5g43290; AtWRKY7, At4g24240; AtWRKY17, At2g24570; AtWRKY14, At1g30650; AtWRKY29, At4g23550. Black boxes represent amino acids of AtWRKY proteins that are identical to those of OsWRKY5. Red and blue boxes represent the conserved WRKYGQK sequence and zinc finger motif of the WRKY domain, respectively. ^aEulgem et al. (2000) classified group II AtWRKY proteins into five subgroups [37]. ^bAmino acid similarity of AtWRKY proteins compared with OsWRKY5. Os, *Oryza sativa*; At, *Arabidopsis thaliana*.

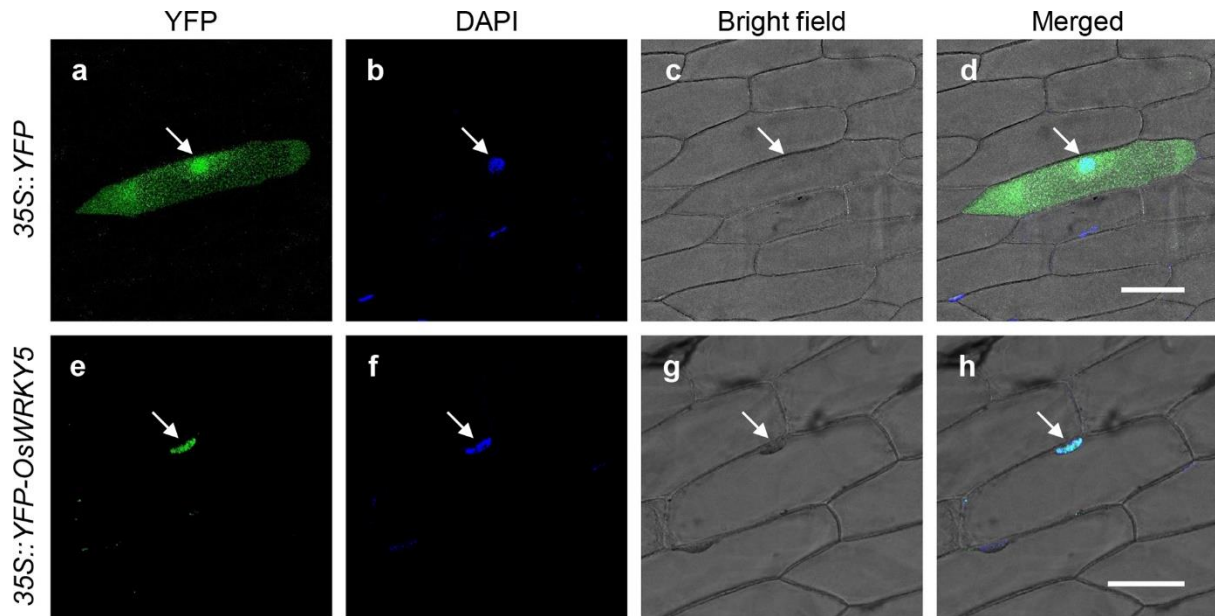


Fig. S2. Subcellular localization of OsWRKY5.

YFP-fused OsWRKY5 proteins transiently expressed in onion epidermal cells. Upper panels show fluorescence from the YFP control (**a-d**), which was distributed throughout the cell. Lower panels show the fluorescent signal of YFP-OsWRKY5 exclusively localized to the nucleus (**e-h**). Nuclei were stained with 4',6-diamidino-2-phenylindole dihydrochloride (DAPI). Experiments were repeated twice with similar results. White arrows indicate the position of the nucleus. Bars = 100 μ m.

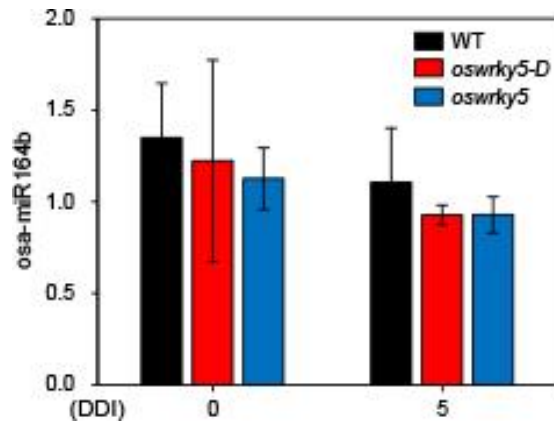


Fig. S3. Endogenous *osa-miR164b* levels in *oswrky5-D* and *oswrky5*.

Total RNA was extracted from detached leaves of WT and mutant lines (*oswrky5-D* and *oswrky5*) at 0 and 5 DDI under DIS as shown in Figure 2B. First-strand cDNA was synthesized by stem-loop pulsed RT-PCR. Transcript levels of *osa-miR164b* were determined by RT-qPCR and normalized using that of U6 snRNA. Mean and SD values were obtained from more than three biological replicates. No significant difference was found between WT and mutant lines by Student's *t*-test. DDI, day(s) of dark incubation.

Table S1. Primers used in this study.

A. Primers for verification of T-DNA insertion		
Primer names	Left primers (5' → 3')	Right primers (5' → 3')
PFG_3A-15928	CATTAAAGCTGGACCAGATGG	AACCACTTGCGGATTAATGC
PFG_3A-06060	TTGGATGCCTGATTAAGGTTG	CCGTTCTTGACATTACACAC
pGA2715	CTAGAGTCGAGAATTCAGTACA	TTGGGGTTTCTACAGGACGTAAC
B. Primers for subcellular localization		
Gene	Forward primers (5' → 3')	Reverse primers (5' → 3')
<i>OsWRKY5</i>	ATGGAGATGATGGTGCAGAAGCA	TCAGGTGGGAGACGTGCCGCAA
C. Primers for RT-qPCR		
Genes	Forward primers (5' → 3')	Reverse primers (5' → 3')
<i>OsWRKY5</i>	GGCTCCAATGATCAGTGATGGA	AGCCATTGTGCATCGGTAGT
<i>SGR</i>	AGGGGTGGTACAACAAGCTG	GCTCCTTGCGGAAGATGTAG
<i>NYC3</i>	TGTCGTTGCCATGTGAAGAT	TTGGTCACGCCACAAATCTA
<i>OsPAO</i>	GGAAATCCTAGCCAAGAAGTGTTG	CGCAGGAATCCCAGCAGTT
<i>Osh69</i>	CCACAACACGGATAACTT	GGTGAACACTATGGAACA
<i>Osh36</i>	GCACGGAGGCGAACGA	TTGAGCGGTAGCACCCATT
<i>Osl85</i>	GAGCAACGGCGTGGAGA	GCGGCGGTAGAGGAGATG
<i>OsNAP</i>	CAAGAAGCCGAACGGTTC	GTTAGAGTGGAGCAGCAT
<i>OsNAC2</i>	CAACTCCTGGAGAGCTGCAA	GATCTCCGGGTTACGTCG
<i>OsNCED3</i>	GTGGTGCTCGACAAGGAGAA	CAGAGGTGGAAGCAGAAGCA
<i>OsNCED4</i>	GAGGTACGACTTCCATGGGC	TTGAGGTACGGCTTGACAC
<i>OsNCED5</i>	CCCAGCTTGAAGCTTTTGCT	ACAACACTGCAACTATCCCTATCACT
<i>OsUBQ5</i>	ACCACTTCGACCGCCACTACT	ACGCCTAAGCCTGCTGGTT
D. Primers for pulsed RT-qPCR of miR164b		
Primer names	Forward primers (5' → 3')	Reverse primers (5' → 3')
miR164b_pulsed RT	GTCGTATCCAGTGCAGGGTCCGAGGTATTCGCACTGGATACGACTGCACG	
miR164b_qPCR	GCTCTGGAGAAGCAGGGC	GTGCAGGGTCCGAGGT
U6 snRNA_qPCR	CAACGGATATCTCGGCTCT	CAACTTGCGTTCAAAGACTC
E. Primers for yeast one-hybrid assay		
Primer names	Forward primers (5' → 3') ^a	Reverse primers (5' → 3') ^a
<i>OsWRKY5_GAD424</i>	<u>GAATTC</u> ATGGAGATGATGGTGCAG	<u>CTGCAGTC</u> AGGTGGGAGACGTGCCG
<i>_EcoRI and PstI</i>	AAGCAACGA	CAA
<i>OsNAP-1_EcoRI and XbaI</i>	<u>GAATTC</u> AGGTGTGAAAACAAATAA GA	<u>CTCGAGA</u> ATAGTACCGCTGTGGTGA A
<i>OsNAP-2_EcoRI and XbaI</i>	<u>GAATTC</u> ATCACGTCGTTTTTCAACT A	<u>CTCGAGGT</u> ACCAAGGTGCTAAGATA C

OsNAC2-1_ <i>SalI</i> and <i>XhoI</i>	<u>GTCGACT</u> GTGTTTCAGTTTGTCTCTT C	<u>CTCGAGG</u> TTAACAAGCCAGAACAAA C
OsNAC2-2_ <i>SalI</i> and <i>XhoI</i>	<u>GTCGACC</u> GGGGACATTTTCAGACGT TT	<u>CTCGAGT</u> GGTTTTGTGGGGCTTAGA A
OsNAC2-3_ <i>SalI</i> and <i>XhoI</i>	<u>GTCGACCC</u> ACTGCTATTACACAAT AG	<u>CTCGAGAT</u> CTCCCAGGAGATAAGCC A
OsNAC2-4_ <i>SalI</i> and <i>XhoI</i>	<u>GTCGACTT</u> CTCGGTTGCTGCTCGGC T-	<u>CTCGAGGG</u> CTAGTGATCCATCAGAT C
OsNAC2-5_ <i>SalI</i> and <i>XhoI</i>	<u>GTCGACCT</u> AGCTAGTACTCCATCC GT	<u>CTCGAGC</u> AGGTTACTACTCCCTCCAT

^a The underlined nucleotides represent the restriction site for restriction enzymes.