

1 Article

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Strategies for the Surface Modification with Ag

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Shaped Nanoparticles: Electrocatalytical

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Enhancement of Screen Printed Electrodes for the

5

Detection of Heavy Metals

6 **Karina Torres-Rivero^{1,2*}, Lourdes Torralba-Cadena¹, Alexandra Espriu-Gascon^{1,2}, Ignasi Casas^{1,2},**
7 **Julio Bastos-Arrieta^{3*}, Antonio Florido^{1,2}**8 ¹ Departament d'Enginyeria Química, Escola d'Enginyeria de Barcelona Est (EEBE), Universitat Politècnica
9 de Catalunya, BarcelonaTEch (UPC), Av. Eduard Maristany 16, 08019 Barcelona, Spain;
10 ltc_86@hotmail.com (L.T.-C.); alexandra.espriu@upc.edu (A.E.-G.); ignasi.casas@upc.edu (I.C.);
11 antonio.florido@upc.edu (A.F.)12 ² Barcelona Research Center for Multiscale Science and Engineering, Av. Eduard Maristany 16,
13 08019 Barcelona, Spain14 ³ Physical Chemistry TU Dresden, Zellescher Weg 19, 01069 Dresden, Germany

15 * Correspondence: julio.bastos@tu-dresden.de (J.B.-A.); karina.torres.rivero@upc.edu (K.T.-R.)

16 **Abstract:** The screen-printed carbon nanofibers electrodes (SPCNFE) represent an alternative with
17 great acceptance due to their results, as well as their low impact for the environment. In order to
18 improve their performance, in the present work they were modified with silver nanoparticles (Ag-
19 NPs) and electrochemically characterized by using anodic stripping voltammetry. From the Ag-NPs
20 synthesis, silver seeds (Ag-NS) and silver nanoprisms (Ag-NPr) were obtained. The Ag-NPs
21 formation was confirmed by micrographs where Ag-NPs with diameters of 12.20 ± 0.04 nm for Ag-
22 NS, and 20.40 ± 0.09 nm for Ag-NPr were observed. The electrodes were modified by using three
23 different deposition methods: drop-casting, spin-coating and *in-situ* approaches. It was observed
24 that the last methodology showed a low amount of Ag-NS deposited on the electrode surface and a
25 deep alteration of this surface. Those facts suggested that the *in situ* synthesis methodology were
26 not appropriate for the determination of heavy metals and it was discarded. The incorporation of
27 the nanoparticles by spin-coating and drop-casting strategies showed different spatial distribution
28 on the electrode surface as proved by scanning electron microscopy. The electrodes modified by
29 these strategies, were evaluated for the cadmium(II) and lead(II) detection using differential pulse
30 anodic stripping voltammetry, obtaining detection limit values of 2.1 and $2.8 \mu\text{g L}^{-1}$, respectively.
31 The overall results showed that the incorporation route does not change directly the electrocatalytic
32 effect of the nanoparticles, but the shape of these nanoparticles (spherical for seeds and triangular
33 for prisms) has a preferential electrocatalytical enhancement over Cd(II) or Pb(II).34 **Keywords:** screen printed electrodes; Ag nanoparticles; drop-casting; spin-coating; nanoprisms;
35 heavy metals; Differential pulse anodic stripping voltammetric; electrocatalysis
3637

1. Introduction

38 In order to ensure water quality, World Health Organization (WHO) has published the
39 maximum allowed concentration of pollutants in drinking water [1]. Among these pollutants, the
40 concentration of heavy metal ions (HMI) is of special concern due to their toxicity and
41 bioaccumulation.42 As examples, As, Cr, Hg, Pb and Cd concentrations must be under 10, 50, 1, 10 and $3 \mu\text{g L}^{-1}$,
43 respectively [1]. Such low values need to be determined by means of highly sensitive techniques like
44 Flameless Atomic Adsorption Spectrophotometry (FAAS) [2], Inductively Coupled Plasma Optical

45 Emission Spectroscopy (ICP-OES) [3] and Inductively Coupled Plasma Mass Spectrometry (ICP-MS)
46 [4], among others, nevertheless require expensive equipment and specialized technicians. These facts
47 increases both the analysis time and the operation costs.

48 In order to avoid those inconveniences, alternative methodologies can be used for the
49 quantification of HMI as, for instance, electrochemical voltammetry. This technique offers, among
50 other advantages, a low cost equipment, easy to handle, a relatively fast analysis, and it is suitable to
51 be used as a portable quantification device [5–9]. Thus, regarding the determination of HMI, the
52 improvement of the electrochemical performance and sensitivity to HMI are required.

53 Analysts have invested big efforts into the design of new electrochemical sensors by taking
54 advantage of the electrocatalytic effect of nanomaterials. This is aimed by the incorporation of
55 metallic nanoparticles (MNPs) [10], oxide nanoparticles (ONPs) or carbon nanoallotropes (as carbon
56 nanotubes or graphene) in the electrode components. These nanomaterials reduce the electron
57 transfer resistance at the electrode surface which leads to the decrease of the electron-transfer limited
58 process and consequently the response of the electrode at low analyte concentration is catalyzed [11–
59 13]. For example, it has been reported that with these modifications, the HMI concentration can be
60 determined at values lower than $1 \mu\text{g}\cdot\text{L}^{-1}$ which meets the WHO requirements for heavy metal
61 quantification in drinking water samples.

62 More specifically, the incorporation of nanoparticles (NPs) on the sensor surface has shown good
63 results for determination of As [14–16], Cr [17], Cu [10,18,19], Pb [10,18–20], Cd [5], Hg [7,21,22]. Most
64 of these works used a screen printed electrode (SPE) modified with different types of NPs (Pt-NPs,
65 Ag-NPs or Au-NPs) as electrocatalysts. The main advantages of using SPE are their versatility and
66 the fact that they can be used as one-use disposable sensor (avoiding any possible carryover
67 contamination from previous measurements). Additionally, they are cost efficient and easily tunable
68 as well as suitable to be incorporated in portable devices.

69 There are different approaches to the modification of electrochemical sensors with
70 nanomaterials. For instance, the modification used by Sanllorente-Méndez et al. [15] and Domínguez-
71 Renedo et al. [17] was based on the electrochemical reduction of a PtCl_6^{2-} solution by applying two
72 different potentials (+0.5 V for 0.01 s and -0.7 V for 10 s) to prepare Pt-NPs directly on the SPE. The
73 main disadvantages of these procedures are that the distribution of the NPs among the electrode
74 surface cannot be controlled and agglomeration of NPs is very common, influencing directly the
75 electrochemical response of the sensor.

76 On the other hand, Li et al. [7] carried out a dip coating strategy, by immersing the electrode into
77 an Au-NPs colloidal solution overnight and then the authors dried the electrode at 80 °C. A similar
78 process was used by Pérez-Ràfols et al. [10], as they drop-casted Ag-NPs on the SPE surface, with a
79 later drying stage of 30 min at 50°C. In this case, the particle size distribution and morphology can be
80 regulated by the NPs preparation, but there is still a lack control of the final distribution of the NPs
81 on the modified surface.

82 Muñoz et al. [23] carried out the incorporation of NPs to carbon nanotubes/epoxy
83 nanocomposite electrodes by physical and chemical approaches. These included the *in situ*
84 modification of the transducer material (with control on NPs spatial distribution); the physical mixing
85 of NPs into the electrode matrix and drop attachment (drop-casting) of a NPs containing solutions.
86 The overall results showed an enhancement of the electrochemical response to hydrogen peroxide as
87 analyte.

88 In this sense, the aim of this work is to evaluate the feasibility of different strategies to modify
89 commercial screen-printed carbon nanofibers electrodes (SPCNFE) with shaped silver nanoparticles
90 and therefore, compare the electrocatalytic effect of these NPs on the determination of Cd(II) and
91 Pb(II) in water samples. More specifically, three procedures of Ag-NPs deposition are considered:
92 drop-casting (DC), *in situ* (IS) and spin-coating (SC). Besides, the characterization of the NPs and the
93 SPCNFE surface by means of electron microscopy is described. Finally, the electrochemical
94 characterization by Differential Pulse Anodic Stripping Voltammetry (DPASV) of the modified
95 sensors is also presented.

96 **2. Materials and Methods**

97 2.1. Reagents and solutions

98 All chemicals were analytical grade and were used with no additional purification. Different
99 reagents used for the Ag-NPs synthesis (sodium citrate, sodium polystyrene sulfonic acid (SPSS), and
100 silver nitrate) were supplied by Sigma-Aldrich (Munich, Germany), sodium borohydride by Panreac
101 (Barcelona, Spain) and ascorbic acid by Scharlab (Barcelona, Spain). Lead(II) nitrate and cadmium(II)
102 nitrate were purchased from Fluka (Buchs, Switzerland) and VWR International LTD (Radnor, PA,
103 USA), respectively. A $2072 \mu\text{g L}^{-1}$ of Pb(II) and a $1124 \mu\text{g L}^{-1}$ of Cd(II) solutions (corresponding to 10^{-5}
104 mol L⁻¹) were prepared by sequential dilution from a 1000 mg L^{-1} stock solution. Metal solutions
105 were standardized by ICP-AES, Perkin Elmer model Optima 3200 (Waltham, MA, USA) or by ICP-
106 MS, Agilent model 7500cx (Santa Clara, CA, USA). A 0.1 mol L^{-1} acetate buffer solution (pH 4.5)
107 prepared from acetic acid (Merck, Munich, Germany) and sodium acetate (Panreac, Barcelona, Spain)
108 was used as electrolyte and for constant pH. All solutions were made with ultrapure water ($18.2 \text{ M}\Omega\text{-cm}$)
109 obtained from a Milli-Q plus 185 system (Millipore, Burlington, MA, USA).

110 2.2. Electrodes

111 Commercial SPCNFE (Dropsens®, ref. 110CNF, Llanera, Spain) including working, counter and
112 reference electrodes were used. The working electrode consisted on a disk of 4 mm diameter where
113 the carbon nanofiber surface was modified with the silver nanoparticles (Ag-NPs-SPCNFE).

114 2.3. Nanoparticle Synthesis

115 Ag-nanoseeds (Ag-NS) and Ag-nanoprisms (Ag-NPr) were prepared as follows, based on the
116 methodology described somewhere else [24,25].

117 *Preparation of Ag-NS*

118 Ag-NS were obtained by mixing under continuous stirring 5 mL of 2.5 mmol L^{-1} sodium citrate,
119 0.25 mL of 500 mg L^{-1} PSSS and 0.3 mL of 10 mmol L^{-1} sodium borohydride. Afterwards, a solution
120 of 0.5 mmol L^{-1} silver nitrate was continuously added to the previous solution at 2 mL min^{-1} rate by
121 using a syringe pump from Kd Scientific, model KDS 510 (Holliston, MA, USA).

122 *Preparation of Ag-NPr*

123 Several aliquots in the range 400-1600 μL of the Ag-NS previously obtained were mixed with 5
124 mL of Milli-Q water and 75 μL of 10 mmol L^{-1} ascorbic acid. Then, 3 mL of 0.5 mmol L^{-1} silver nitrate
125 were continuously added to each aliquot at 1 mL min^{-1} . Finally, 0.5 mL of 25 mmol L^{-1} sodium citrate
126 was added to each solution in order to stabilize the obtained Ag-NPr.

127 2.4. Electrode Modification

128 *Drop-casting methodology*

129 The same methodology and conditions as previously studied by Pérez-Ràfols [10] were used in
130 the modification of the SPCNFE by both Ag-NS and Ag-NPr. The deposition methodology consisted
131 in placing 40 μL of the Ag-NPs aqueous solution on the working electrode surface of the SPCNFE
132 and evaporate the solvent by heating it at 50°C for 30 minutes. This procedure avoids damage to the
133 Ag-NPs or electrical parts of the electrode and, at the same time, it ensures the removal of water.

134 *Spin-coating methodology*

135 Ag-NS and Ag-NPr were also used in the SPCNFE modification by the spin-coating
136 methodology. A spin coater WS-650-8B from Laurell Technologies Corporation (North Wales, PA,
137 USA) was used. The SPCNFE was attached to the center part of the spin coater by the vacuum system.
138 Then, the different Ag-NPs were added onto the center part of the working electrode by placing 20
139 μL of the colloidal solution. The SPCNFE was then stirred under nitrogen atmosphere at 2000 rpm
140 for 3 minutes inside the spin coater. A second 20 μL drop of Ag-NP was placed onto the same place
141 and the same stirring methodology was performed again to the electrode.

142 *In-situ Nanoparticle synthesis on electrode surface*

143 In this methodology a bare SPCNFE electrode was dipped in different beakers containing the
144 following solutions: first it was immersed in a 3 mol·L⁻¹ nitric acid for 1 h; afterwards in a 1 mol·L⁻¹
145 sodium chloride; later in a 0.1 mol·L⁻¹ silver nitrate and finally in a 0.2 mol·L⁻¹ freshly prepared sodium
146 borohydride, all of them for 30 minutes[23].

147 2.5. Characterization of the Ag-NPs and of the SPCNFE surface

148 *UV/Vis Spectroscopy*

149 The UV/Vis spectra showing the Surface Plasmon Resonance (SPR) of the Ag-NS and Ag-NPr
150 colloidal solutions were recorded by using an Agilent-Hewlett Packard spectrophotometer, model
151 8453 (Waldbronn, Germany).

152 *Electron Microscopy Characterization*

153 *Scanning Electron Microscopy (SEM)*

154 Colloid solutions of Ag-NS and Ag-NPr nanoparticles were characterized by using a Gemini
155 SEM from ZEISS® (Jena, Germany). Ag-NPs samples were prepared as in a previous work [26]. In
156 addition, the surface of the SPCNFE electrodes was also studied by the SEM before and after the Ag-
157 NPs deposition. This aimed to determine the NPs presence and their spatial distribution on the
158 SPCNFE regarding the modification strategy: drop-casting, spin-coating or *in-situ*.

159 *Transmission Electron Microscopy (TEM)*

160 For further characterization of the Ag-NPs synthetized in this work, a JEM-2010 TEM from JOUL
161 ® (Tokyo, Japan) was used. Ag-NPs samples were also conditioned as in a previous work [26]. From
162 the TEM images, the size distribution of the obtained Ag-NPs was determined and the size
163 distribution histograms were calculated as before [26] using the Image-J version 1.51m software.

164 2.6. Electrochemical Characterization of Ag-NPs-SPCNFE

165 Voltammetric studies of the SPCNFE modified electrodes were performed with a Multi
166 Autolab/M204 - Modular Multi Potentiostat/Galvanostat and a personal computer with NOVA 2.1
167 software package to control the potentiostat as well as for the required data treatment, all from
168 Metrohm (Herisau, Switzerland).

169 Cyclic voltammograms of bare and NPs modified electrodes were obtained in acetic acid/acetate
170 buffer solution by scanning the potential from -1.00 to +1.00 V at a scan rate of 0.01 V·s⁻¹ and a step
171 potential of 0.00244 V.

172 Differential pulse anodic stripping voltammetric (DPASV) measurements by using Ag-NPs-
173 SPCNFE for the determination of Pb(II) and Cd(II) ions were performed. A deposition potential (E_d)
174 of -1.40 V applied under stirring conditions during a deposition time (t_d) of 180 of seconds and
175 followed by a rest period (t_r) of 5 seconds were used. DPASV measurements were carried out under
176 the following conditions: a scanning potential range from -1.40 to 0.00 V, a step potential of 5 mV, a
177 pulse time of 50 ms and a pulse amplitude of 50 mV. All experiments were performed at room
178 temperature (22 ± 1 °C) and without oxygen removal.

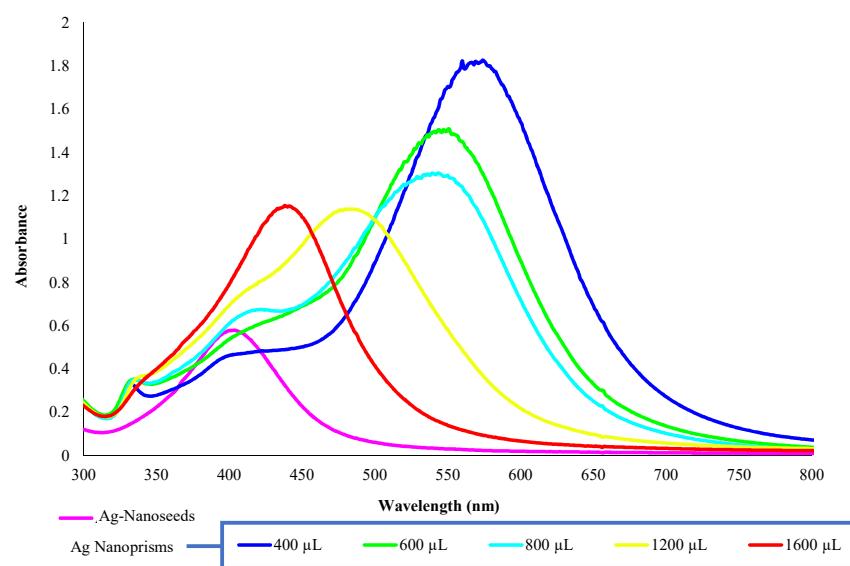
179 Measurements of Pb(II) and Cd(II) ions by DPASV were performed using bare electrodes and
180 for each Ag-NPs-SPCNFE prepared by the different modification methodologies. Initially, the
181 electrodes were calibrated for each HMI. For this purpose, increasing concentrations of Pb(II) and
182 Cd(II) solutions were added to an initial 40 mL of acetic/acetate buffer solution. Calibration samples
183 ranged from 1.0 to 100.0 and from 1.0 to 75.0 µg L⁻¹ for Pb(II) and Cd(II), respectively.

184 3. Results and Discussion

185 3.1. UV/Vis Spectroscopy Characterization

186 The formation of the Ag-nanoparticles, perceived by simple visual observation of the color, was
187 monitored by the UV-Vis spectra of the different colloidal solutions. Thus, to the initial Ag-NS

188 suspension, different volumes ranging between 400 and 1600 μL of the Ag-NS colloidal solution were
 189 added. Figure 1 shows the spectra obtained for the Ag-NS and for Ag-NPr synthesized by using the
 190 different volumes of Ag-seed solution already mentioned. It can be observed that the Ag-NPr
 191 nanoparticles with less amount of seed solution (400 μL) showed an absorbance peak at 570 nm
 192 further away from to the initial seeds' absorbance peak at 405 nm. However, colloidal solutions with
 193 higher amounts of the added Ag-NS solution presented absorbance bands that shifted to lower
 194 wavelengths getting closer to the original Ag-NS solution (see Figure 1). The obtained wavelengths
 195 agreed with previously reported values where silver colloids exhibit maximum absorbance within
 196 the range 400–500 nm due to Surface Plasmon Resonance (SPR) [27–29]. It must be mentioned that
 197 the peak absorbance of the 400 μL Ag-NS solution was higher than all the others and when higher
 198 volumes of Ag-NS solution were added, peaks heights decreased to values getting closer to the ones
 199 presented by the initial Ag-NS solution. Moreover, the shift (from 405 nm in the initial Ag-NS
 200 solution) towards larger wavelengths (570 nm in 400 μL solution) could also indicate an increase in
 201 the mean diameter of Ag-nanoparticles [30,31]. For all this, the Ag-NS and the Ag-NPr obtained from
 202 a volume of 400 μL of Ag-NS solution were used in the next studies.

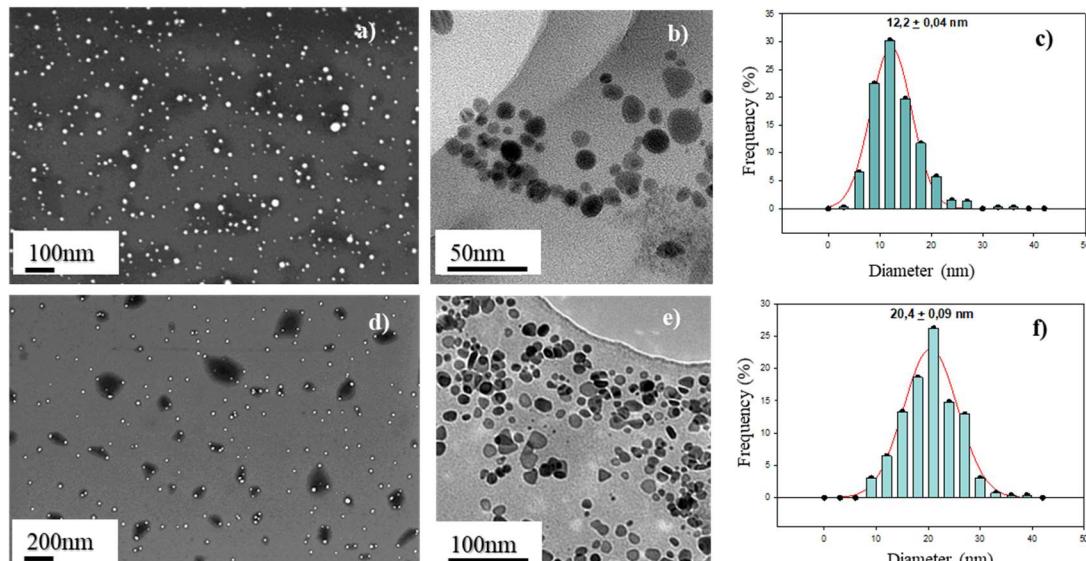


203
 204 **Figure 1.** UV/Vis spectra obtained for the Ag-nanoseeds and for Ag-nanoprisms synthesized by using
 205 different volumes of Ag-seed solution: 400, 600, 800, 1200 and 1600 μL .
 206

207 3.2. Electron Microscopy Characterization

208 3.2.1. Characterization of Ag-nanoparticles by TEM and SEM

209 Figure 2 shows the electron microscopy characterization of the studied Ag-NPs samples. The
 210 SEM image of the Ag-NS can be observed in Figure 2a (white dots), confirming the effectivity of the
 211 synthesis procedure followed. Additionally, Figure 2c shows the size distribution histogram obtained
 212 from a total of 400 Ag-NS. The nanoparticle counting was performed following the same procedure
 213 and using the same software as reported previously [26]. These results show that the Ag-NS obtained
 214 present an average diameter of 12.2 ± 0.4 nm.



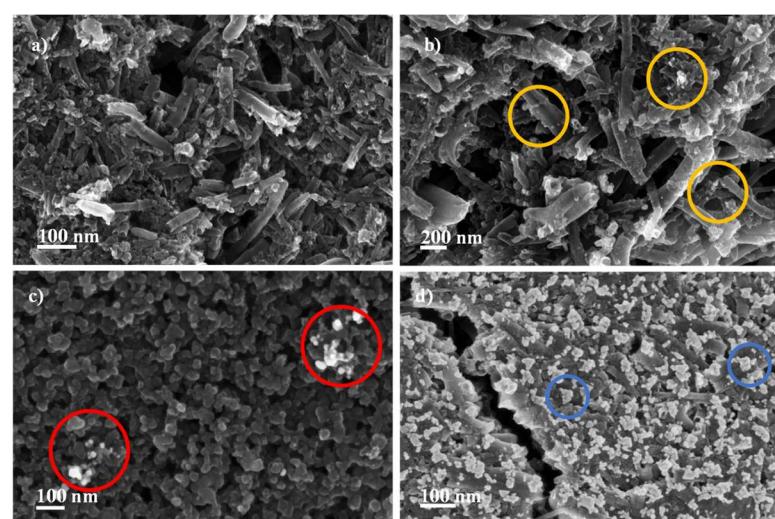
215
216 **Figure 2.** Characterization of the Ag-nanoseeds (a, b, c) and Ag-nanoprisms (d, e, f). SEM images (a,
217 d); TEM images (b, e) and Size Distribution Histograms (c, f), respectively.
218

219 From the TEM image presented in Figure 2b it can be deduced that most of Ag-NS present a
220 spherical shape. These structures are in good agreement with reported shapes of Ag-NPs [24,25].

221 The SEM resulting image of the Ag-NPr obtained can be observed as white dots in Figure 2d.
222 The diameter distribution histogram of the Ag-NPr (see Figure 2f) show that the average particle size
223 was 20.4 ± 0.09 nm. Compared with the Ag-NS, Ag-NPr are bigger and present a slight wider size
224 distribution than the Ag-NS. These results confirmed the transformation of Ag-NS to Ag-NPr as
225 described previously [24,25]

226 3.2.2. Electrode Characterization by SEM

227 In Figure 3, the SEM InLens images of the different electrodes obtained as explained above is
228 presented in order to compare the final surface obtained. The Ag-NS were located into the carbon
229 fibers, identified as circled white spots in the images of the Figures 3 b, c and d.
230



231
232 **Figure 3.** SEM InLens images of the electrodes used in this work. a) Bare commercial SPCNFE; b) Ag-
233 NS-SPCNFE modified by drop-casting method; c) Ag-NS-SPCNFE modified by *in-situ* synthetic
234 method; d) Ag-NS-SPCNFE modified by spin-coating method. Examples of Ag-nanoparticle location
235 is indicated with colored circles.
236

237 Figure 3c shows the Ag-NPs incorporation by the *in situ* methodology. It can be observed that
238 the modification of the electrode surface was not acceptable due the low amount of Ag-NS deposited.

239 In addition, the mechanical resistance of the SPCNFE was compromised as some changes on the
 240 substrate were observed (change of color of the electrode connectors) that could be caused by the
 241 NaBH₄. Additionally, blank electrodes were prepared by the IS approach and their response signal
 242 was lost in all cases, showing that this modification strategy was not appropriate to functionalize the
 243 electrode surface with NPs. Therefore, the IS strategy was not used for the determination of heavy
 244 metals.

245 The SEM images of the electrodes prepared by drop-casting of Ag-NS (Figure 3b), indicated in
 246 this case a successful modification of the electrode, with Ag-NS visible over the electrode surface.
 247 Finally, the electrodes prepared by the spin-coating approach showed a strong modification of the
 248 SPCNFE electrode (Figure 3d) with a larger amount of deposited Ag-NPs as compared with the two
 249 previous methodologies. This effect could be attributed to a better and uniform dispersion of the
 250 nanoparticle suspension over the electrode substrate, due to the speed of the spray coating process
 251 itself. Based on the characterizations performed, it was decided to continue the work only using the
 252 two electrodes with successful incorporation of the NPs to the electrode surface.

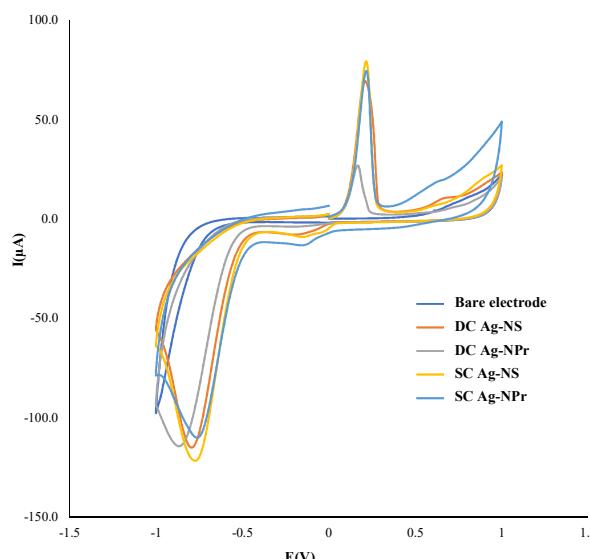
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 254 3.3. Electrochemical characterization of the electrodes

255 3.3.1. Preliminary Studies of the SPCNFE modification with Ag-nanoparticles

256 In order to determine if the modification of SPCNFE with Ag-nanoparticles resulted in the
 257 enhancement of their electrochemical response, Cyclic Voltammetry (CV) and Differential Pulse
 258 Anodic Stripping Voltammetry (DPASV) were performed.

259 First, the cyclic voltammograms of both the bare and the Ag-NPs modified electrodes were
 260 carried out, as seen in Figure 4. It can be seen that the current intensity of the oxidation peaks at 0.2
 261 V (Ag-NPs) of the Ag-NS-SPCNFE and Ag-NPr-SPCNFE are very similar, suggesting that the Ag-
 262 NPs concentration on the WE surface would be comparable, which is important for the evaluation of
 263 both strategies.

264 It seems that SC strategy offers a more consistent approach in terms of amount of NPs
 265 incorporated to the SPCNFE. DC strategy depends on factors like solvent evaporation and
 266 appropriate location of the drop, this can explain the different signal obtained regarding the amount
 267 of NPs incorporated, although for the case of DC- Ag-NPr, the signals agree accordingly to the SC
 268 strategy.

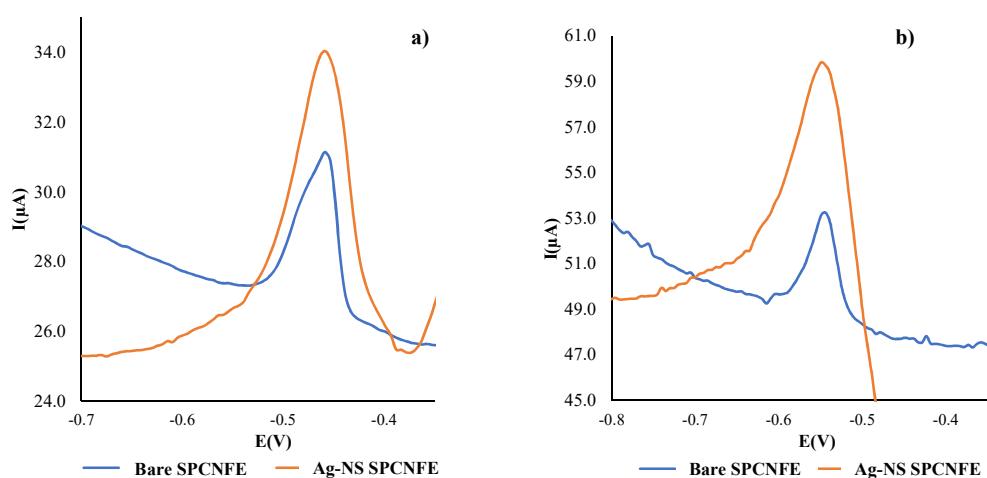


269
 270 **Figure 4:** Cyclic voltammograms in acetic acid/acetate buffer at pH 4.5 of Ag-nanoseeds and Ag-
 271 nanoprisms based SPCNFE electrodes obtained by using either drop-casting (DC) or spin-coating
 272 (SC) methodologies.

273
 274 As seen in Figure 4, current peaks obtained in the case of Ag-NS-SPCNFE were higher than in
 275 the case of Ag-NPr-SPCNFE. Even though this effect might be caused by the fact that the amount of

276 Ag-NPs on both electrode surface could be different due to the different Ag-NPs concentration on
 277 the colloidal solutions, another reason could be related to the specific surface area of the Ag-NPs. As
 278 it was shown in Figure 2, smaller nanoparticles were obtained in the case of Ag-NS than in Ag-NPr
 279 and this would probably result in a better response of the Ag-NS-SPCNFE

280 In order to study the electrochemical response comparing a bare SPCNFE electrode and a Ag-
 281 NPs-SPCNFE in the determination of Pb(II) or Cd(II), differential pulse anodic stripping
 282 voltammetry (DPASV) measurements were performed in solutions containing either 70.0 $\mu\text{g L}^{-1}$ of
 283 Pb(II) or Cd(II). From the results obtained (see Figure 5), it can be concluded that the modification of
 284 the electrodes by Ag-NPs caused a significant increase in the electrode response and would be an
 285 interesting alternative in the determination of both metal ions.
 286



287
 288 **Figure 5.** Differential pulse anodic stripping voltammetry (DPASV) measurements of non-modified
 289 SPCNFE and Ag-nanoseeds-SPCNFE electrodes obtained for a 70 $\mu\text{g L}^{-1}$ of a) lead(II) or b)
 290 cadmium(II) ions. Experimental conditions: acetic acid/acetate buffer pH 4.5 with an E_d of -1.40 V
 291 applied during a t_d of 180 seconds and employing a step potential of 5 mV, a pulse time of 50 ms and
 292 a pulse amplitude of 50 mV.
 293

294 3.3.2. Study of Ag-NS-SPCNFE electrodes obtained by either drop-casting (DC) or spin-coating 295 (SC) methodologies

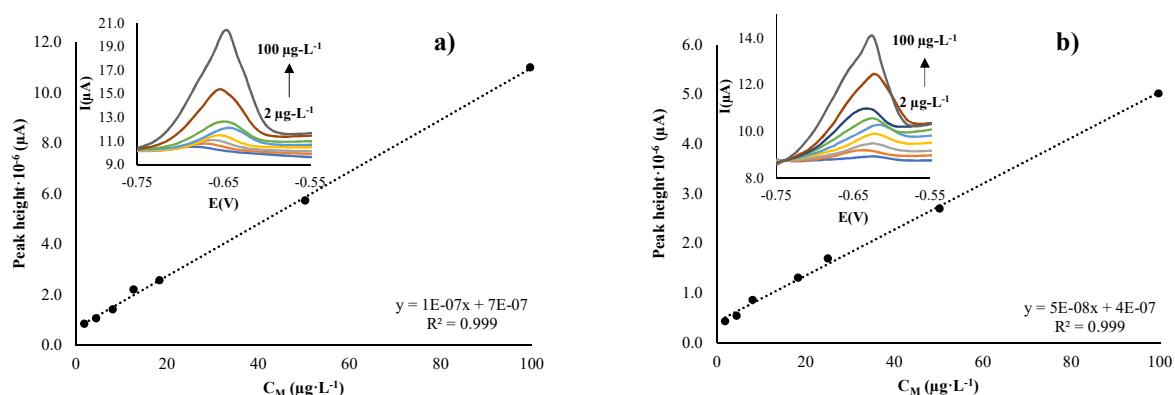
296 Single calibration curves by DPASV were performed by increasing the concentration of Pb(II)
 297 and Cd(II) in the range from 1.0 to 100.0 and from 1.0 to 75.0 $\mu\text{g L}^{-1}$, respectively. The same procedure
 298 was followed for either DC or SC electrodes. From the data obtained, detection limits were
 299 determined by using the Miller & Miller procedure [32,33].

300 Results of the calibration parameters as detection limits (LOD), linear ranges and linearity are
 301 listed in Table I. The limit of quantification (LOQ) is considered as the lower value of the linear range.
 302 As an example, the voltammogram related to the Pb(II) response and the corresponding calibration
 303 plot of Ag-NS-SPCNFE for DC and SC deposition methods are shown in Figure 6.
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314 **Table I.** Calibration parameters as limits of detection (LOD), linear ranges and linearity obtained for
 315 the determination of Pb(II) and Cd(II) on Ag-nanoseeds-SPCNFE and Ag-nanoprisms-SPCNFE and
 316 for drop-casting (DC) and spin-coating (SC) methodologies.

Deposition/Ag-NPs	Pb(II)			Cd(II)		
	LOD ($\mu\text{g}\cdot\text{L}^{-1}$)	Linear Range ($\mu\text{g}\cdot\text{L}^{-1}$)	R^2	LOD ($\mu\text{g}\cdot\text{L}^{-1}$)	Linear Range ($\mu\text{g}\cdot\text{L}^{-1}$)	R^2
DC Ag-nanoseeds	3.3	10.9-99.6	0.9990	3.7	12.2-73.4	0.9923
SC Ag-nanoseeds	2.8	9.4-99.6	0.9990	2.4	8.1-73.4	0.9976
DC Ag-nanoprisms	3.1	10.3-18.3	0.9840	2.2	7.4-73.4	0.9980
SC Ag-nanoprisms	3.4	11.3-50.3	0.9911	2.1	6.9-73.4	0.9976

317
318



319 **Figure 6.** Calibration curves and DPASV measurements (insets) for Pb(II) calibration obtained with
 320 the Ag-nanoseeds-SPCNFE electrodes using a) drop-casting and b) spin-coating methodologies.
 321 Same experimental conditions as in Figure 4.

322
323

324 It can be seen from the DPASV curves presented in Figure 6 that a stable oxidizing peak appears
 325 around -0.64 V, which corresponds with the increasing concentration of Pb(II) in both approaches.
 326 Comparing the results obtained with Ag-NS with the ones obtained with the bare electrode, it can be
 327 concluded that the electrode electrocatalytic response is clearly enhanced. This result agrees with
 328 previous reported works that stated the effect of Ag-NPs on the increased sensitivity and analytical
 329 features of the modified SPCNFE [10,26].

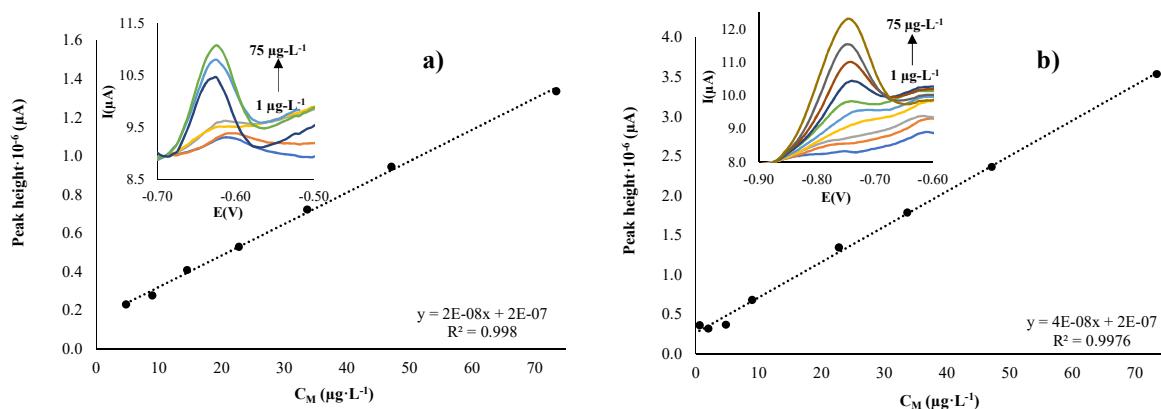
330 Table I shows that in all cases, electrodes presented good and similar performance in terms of
 331 LOD with slightly better values in the case of the spin-coating method. In terms of metal ions
 332 response, it can be observed that Ag-NS electrodes showed better linear regressions and wider linear
 333 ranges in the Pb(II) calibrations than in the Cd(II) response. This behavior was obtained for both
 334 modification strategies, and it seems that the electrocatalytic enhancement relies only on the
 335 presence of the NPs but it does not seem to be associated to the incorporation route. As it was seen
 336 in SEM images, in SC deposition most of the NPs are homogeneously distributed on the surface of the
 337 electrode (externally localized), while for DC, it seems that there is some diffusion into the matrix
 338 (internally localized). SC is a common technique used to produce uniform thin films (e.g. of organic
 339 materials) with customized thicknesses of the order of micrometers and nanometers. For the case of
 340 the preparation of NPs films, the coating depends on the NPs concentration. One crucial parameter
 341 to control in SC is the spin acceleration, which drives the fluid thinning, solvent evaporation and
 342 consequently, the film formation [34]. On the other hand, the DC deposition depends on the
 343 evaporation rate due to the temperature and the uniform coating during the casting. It seems that

344 interaction between the casting and evaporation in DC leads to the NPs penetration into the nanofiber
 345 matrix.

346 In all cases, the LODs values obtained for both deposition methods are close to $3 \mu\text{g L}^{-1}$. It is
 347 important to point out that these values are below or at least at the same order of magnitude of the
 348 legislated values in the case of Pb(II) and Cd(II) concentrations in drinking waters [1].
 349

350 3.3.3. Study of Ag-NPr-SPCNFE electrodes prepared by either drop-casting (DC) or spin-coating
 351 (SC) methodologies

352 By following a similar procedure as before, Ag-NPr electrodes were studied and their response
 353 to Pb(II) and Cd(II) concentrations was determined. Results of the calibration parameters are
 354 presented in Table I. From this data, it can be determined that slightly better electrochemical
 355 performance was observed in the case of Cd(II) response for this kind of Ag-NPs. In Figure 7 it can
 356 be observed a stable and measurable signal when Cd(II) concentration is increased. This performance
 357 was seen in both DC and SC methodologies, and the current signal improved in a relevant way
 358 compared with the bare electrode.



359
 360 **Figure 7.** Calibration curves and DPASV measurements (insets) for Cd(II) calibration obtained with
 361 the Ag-nanoprisms-SPCNFE electrodes using a) drop-casting and b) spin-coating methodologies.
 362 Same experimental conditions as in Figure 4.
 363

364 From the data presented in Table I, it can be pointed out that Ag-NPr electrodes presented
 365 slightly better LOD values, better linear regressions and wider linear ranges in the Cd(II) calibrations
 366 than in the Pb(II) response. However, non-significant differences were seen in this case in terms of
 367 drop-casting or spin-coating methodologies. This would confirm the results mentioned for Ag-NS-
 368 SPCNFE that the modification methodology would not be a determinant factor in the enhancement
 369 of the response of the electrodes.

370 Additionally, as it was already mentioned, LODs obtained for Ag-NPr-SPCNFE in both
 371 deposition methods and for both studied ions were around $3 \mu\text{g L}^{-1}$. These values are also below the
 372 legislated values for the determination of Pb(II) and Cd(II) concentration in drinking waters.
 373 Nevertheless, a comparison among all the response characteristics for both Ag-NPs studied could
 374 conclude that Ag-NS would be more suitable in the determination of Pb(II), meanwhile Ag-NPr
 375 would result more appropriate to determine Cd(II). This can be mainly attributed to the different
 376 reactivity that these NPs can have due to their shape.

377
 378

379 **4. Conclusions**

380 In this work, three different deposition methodologies: *in situ*, drop-casting and spin-coating
381 have been evaluated as feasible strategies for the modification of screen-printed carbon nanofibers
382 electrodes (SPCNFE) with two different shaped nanoparticles: Ag-NS and Ag-NPr. For this, the
383 formation of each type of Ag-NPs was monitored by UV-Vis. Moreover, electron microscopy was
384 used in the characterization of their size, shape and distribution on the SPCNFE surface. The obtained
385 Ag-NS presented an average diameter of 12.2 ± 0.04 nm and, in general, showed a spherical shape.
386 On the other hand, in the case of Ag-NPr, the obtained average particle size was 20.4 ± 0.09 nm and,
387 among a variety of shapes, mostly showed a triangular shape. The electrode modification approaches
388 were studied by Scanning Electron Microscopy. In two of them, drop-casting and spin-coating, SEM
389 images indicated a correct modification of the electrode, with Ag-NPs embedded inside or all over
390 the carbon nanofibers. Additionally, in the case of the spin-coating methodology, SEM images
391 showed a strong modification of the SPCNFE with many and more uniformed Ag-nanoparticles
392 deposited due to the high speed of the spin coater. Finally, the *in situ* methodology showed a low
393 amount of Ag-NS deposited on the electrode surface and a deep alteration of this surface. Those facts
394 suggested that the *in situ* synthesis methodology were not appropriate for the electrode modification
395 and it was discarded.

396 Finally, electrochemical characterizations of the Ag-NPs-SPCNFE and their application in the
397 determination of Pb(II) and Cd(II) ions were carried out. The results obtained in the case of Ag-NS
398 and Ag-NPr based SPCNFE electrodes for both studied metal ions and for the DC and SC deposition
399 strategies indicated appropriate and similar performances in terms of LOD with values around $3 \mu\text{g}$
400 L^{-1} . In terms of metal ions response, it was observed that Ag-NS electrodes showed better linear
401 regressions and wider linear ranges in the case of Pb(II) calibrations, meanwhile for Ag-NPr
402 electrodes better results were obtained for Cd(II) response. This behavior was obtained for both DC
403 and SC methodologies. A comparison among all the response characteristics for both Ag-NPs studied
404 could conclude that Ag-NS would be more suitable in the determination of Pb(II), while Ag-NPr
405 would result more appropriate to determine Cd(II). On the other hand, an evaluation of the effect of
406 the modification methodology on the Ag-NPs-SPCNFE response indicated that using DC or SC
407 deposition methods seemed not to be a parameter that enhanced the response of the SPCNFE
408 electrodes, even though better Ag-NPs distribution was observed by SEM images in the case of SC
409 methodology. Therefore, it can be concluded that the spatial distribution of the NPs in the SPCNFE
410 does not play a significant effect on the detection, under the conditions of the present study.
411 However, their presence is the one giving the electrocatalytic enhancement, so the electrode substrate
412 acts as the only component during the electrochemical detection. Nevertheless, SC approach can be
413 considered more time and cost efficient, which makes it a more environmentally friendly approach
414 for the modification of electrodes.

415 Additionally, LODs obtained in both deposition methods, Ag-NPs and for both studied ions
416 (around $3 \mu\text{g L}^{-1}$) are below or at least the same order of the legislated values in the case of Pb(II) and
417 Cd(II) concentration in drinking waters. For all this, Ag-NPs-SPCNFE could be an accurate, portable
418 and sensible analytical system for the determination of Pb(II) or Cd(II) in natural waters.

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