

1 Article

2 Curcumin Attenuates Lead-Induced Cerebellar 3 Toxicity in Rats via Inhibition of Oxidative Stress 4 and Chelating Activity

5 Kabeer Abubakar^{1,3*}, Maryam Muhammad Mailafiya^{1,3}, Abubakar Danmaigoro⁵, Samaila Musa
6 Chiroma^{1,6}, Ezamin Bin Abdul Rahim^{4*}, and Zuki Abu Bakar @ Zakaria²

7 1 Department of Human Anatomy, Faculty of Medicine and Health Sciences, University Putra Malaysia,
8 43400 Serdang, Selangor Darul Ehsan, Malaysia; maryam.mailafia@gmail.com (M.M.M.);
9 musasamailachiroma@yahoo.com (S.M.C.)

10 2 Department of Preclinical Sciences Faculty of Veterinary Medicine, University Putra Malaysia, 43400
11 Serdang, Selangor Darul Ehsan, Malaysia; zuki@upm.edu.my (M.Z.A.)

12 3 Department of Human Anatomy, College of Medical Sciences, Federal University Lafia, P.M.B 146,
13 Akunza, Lafia, Nasarawa State, Nigeria.

14 4 Department of Radiology, Faculty of Medicine and Health Sciences, University Putra Malaysia, 43400
15 Serdang, Selangor Darul Ehsan, Malaysia; (E.A.R.)

16 5 Department of Veterinary Anatomy, Faculty of Veterinary Medicine, Usman Danfodiyo University,
17 P.M.B 2346, Sokoto, Nigeria; abubakar.danmaigoro@udusok.edu.ng (A.D.)

18 6 Department of Human Anatomy, Faculty of Basic Medical Sciences, University of Maiduguri, Borno
19 State, Nigeria,

20 * Correspondence: drezahar@gmail.com (E.A.R.); Tel.: +60196846933 and kabeernakhadee@yahoo.com
21 (K.A.); Tel.: +60182018569.

22 First Author's ORCID: <https://orcid.org/0000-0001-7552-0383>

23 **Abstract:** Lead (Pb) is a toxic environmental heavy metal that induces serious clinical defect on all
24 organs with the nervous system being the primary target. Curcumin is the main active constituent of
25 turmeric rhizome (*Curcuma longa*) with strong antioxidant and anti-inflammatory properties. This
26 study is aimed at evaluating the therapeutic potentials of curcumin on Pb-induced neurotoxicity.
27 Thirty six male Sprague Dawley rats were randomly assigned into five (5) groups with 12 rats in the
28 control (normal saline) and 6 rats for the lead treated group (LTG) (50 mg/kg lead acetate for 4 weeks),
29 recovery group (RC) (50 mg/kg lead acetate for 4 weeks), treatment group 1 (Cur100) (50 mg/kg lead
30 acetate for 4 weeks, followed by 100 mg/kg curcumin for 4 weeks) and treatment group 2 (Cur200)
31 (50 mg/kg lead acetate for 4 weeks, followed by 200 mg/kg curcumin for 4 weeks) groups each. All
32 experimental groups received the oral treatments through orogastric-tube on alternate days. Motor
33 function was assessed using horizontal bar method while Pb concentration in the cerebellum of the
34 rats were evaluated using ICP-MS techniques. Pb-administered rats showed significant decrease in
35 motor scores, SOD activity with increase MDA levels and Pb concentration in their cerebellum with
36 marked alterations in the histological architecture of the cerebellar cortex layers. However, treatment
37 with curcumin improved the motor score, reduced Pb concentration in the cerebellum and
38 ameliorates the markers of oxidative stress as well as restored the histological architecture of the
39 cerebellum. The results this in study suggested that curcumin attenuates Pb-induced neurotoxicity
40 via inhibition of oxidative stress and chelating activity.

41 **Keywords:** Curcumin; Lead toxicity; ICP-MS; Horizontal bar; Motor coordination; Oxidative stress;
42 Cerebellum.

43 1. Introduction

44 Lead (Pb) is a ubiquitous environmental neurotoxin that induces several physiological,
45 behavioral and biochemical abnormalities in humans and animals [1]. Pb toxicity remain a common
46 problem in both developing and industrialized countries due to unavoidable environmental and
47 occupational exposure resulting to about 600,000 new cases of intellectual disabilities in children and

48 account for about 143,000 deaths per year [1,2]. Although data from Adults Blood Lead Epidemiology
49 and Surveillance (ABLES) program indicates a significant decrease in incidence of blood lead levels
50 (BLLs) among adults' industrial workers, occupational exposure Pb remained a public health
51 concern accounting for about 94% Pb exposures [3]. Occupational exposure to Pb is linked with
52 several health consequences such as cognitive impairment, reproductive disorders, hypertension,
53 motor dysfunction, cancer, hepatotoxicity, nephrotoxicity, and mortality [4,5].

54 The mechanism of Pb-toxicity is due to its ability to induce oxidative stress via disruption the
55 oxidant/antioxidant balance mechanism in the cells [6]. Pb as a ubiquitous toxin is known to induce
56 oxidative stress by increasing the generation of reactive oxygen species (ROS) such as hydroxyl
57 radicals, lipid peroxides, superoxide radicals and hydrogen peroxide [6]. Pb-toxicity has long been
58 linked with impaired motor function particularly deficits in visuomotor coordination among adult
59 industrial workers and children exposed to Pb [7]. However, studies on the effect of Pb exposure on
60 visuomotor integration among Yugoslavian and urban African children shows that blood lead levels
61 (BLLs) contributed significantly to poorer fine motor skills as well as gross motor speed [7,8]. Animal
62 models, when treated with Pb display increased oxidative stress, cognitive impairments,
63 degeneration of neurons, deficits in motor coordination and mortality [9–11].

64 The field of phytotherapy is growing more interest in the use of phytochemical compounds such
65 as curcumin due to its wide pharmacological safety and antioxidant properties against heavy metal
66 induced-toxicity since the application of standard drugs such as chelators in the treatment and
67 management of heavy metal poisoning have shown several side effects, such as brain damage,
68 anemia, liver and kidney diseases, anaphylactic shock and others [12]. Curcumin is the main natural
69 polyphenol in the rhizome of turmeric (*Curcuma longa*), that belong to the family of ginger
70 (*Zingiberaceae*) which is widely used as traditional medicine and food in Asia [13]. It is lipophilic,
71 phenolic and water insoluble compound which has antioxidant, anti-inflammatory and anticancer
72 properties [14]. Curcumin has a strong antioxidant properties through increased production of
73 antioxidant enzymes resulting in to the scavenging of excess ROS produced and inhibition of lipid
74 peroxidation [15]. Curcumin as a lipophilic compound has the potency to cross the blood-brain
75 barrier (BBB) and bind to plaques in the brain, thus inhibiting amyloid-B peptide aggregation in
76 Alzheimer's disease [16]. Post-treatment with curcumin in subarachnoid hemorrhage (SAH) induced
77 mice preserved the integrity of the BBB and improves brain functions via down regulation of Matrix
78 Metalloproteinase 9 (MMP-9) and inhibition of microglia cells as well as reducing water content in the
79 brain [17].

80 The neuroprotective mechanism of curcumin on neurodegenerative disorders is mainly due to
81 its ability to bind redox-active transition metal ions such as Mn^{2+} , Fe^{2+} , Cu^{2+} , and Zn^{2+} to form active
82 and tight complexes [18]. Additionally, curcumin has a wide range of pharmacological activities and
83 safety margin, it is identified as a natural drug against neurodegenerative disorders [18]. However,
84 there is paucity of knowledge on the therapeutic potentials of curcumin in Pb induced cerebellar
85 toxicity. Therefore, the present study aimed at investigating the neurotherapeutic potentials of
86 curcumin on Pb-induced cerebellar toxicity in rats.

87 2. Materials and Methods

88 2.1. Chemicals

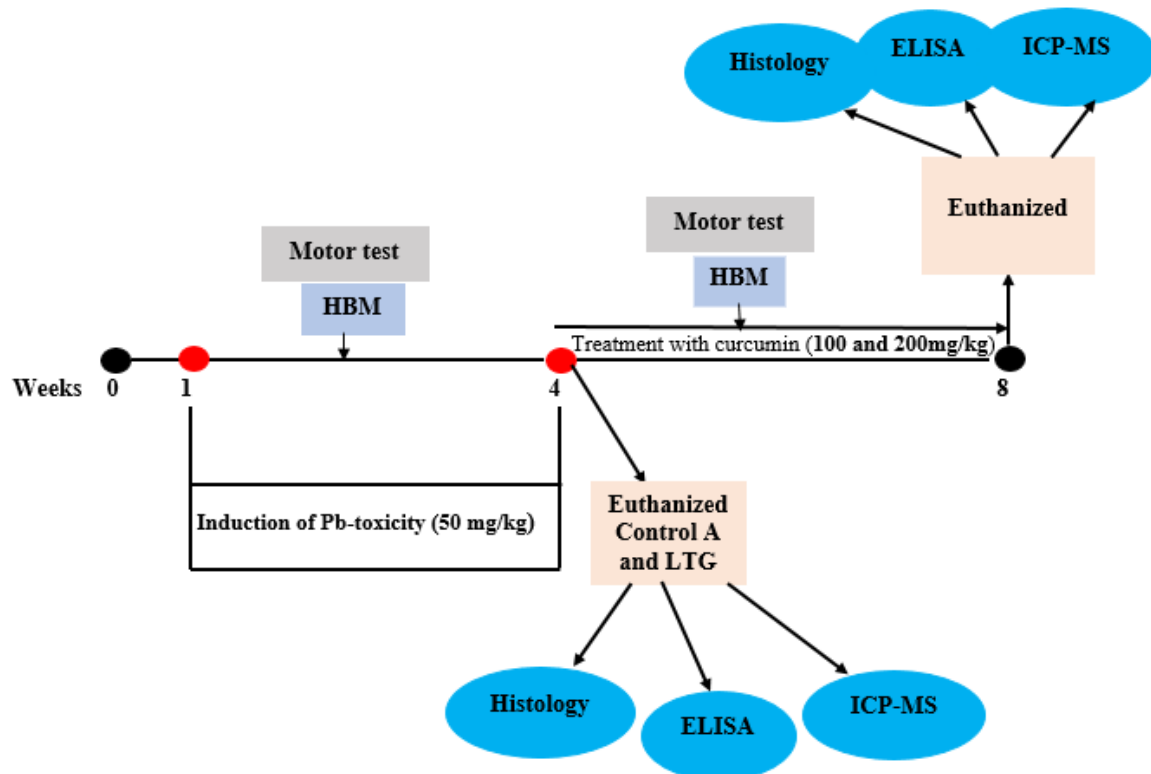
89 Lead acetate, (CH₃CO₂)₂Pb.3H₂O, 99%) and animal feed were purchased from Sigma-Aldrich
90 (St. Louis, MO USA) while MDA ELISA kit and SOD assay kit were purchased from Elabscience
91 (Houston, TX, USA). Curcumin, sodium acetate, aniline solution and colophonium were purchased
92 from Apical Scientific Sdn. Bhd. Malaysia. All other chemicals used were of high analytical grade.

93 2.2. *Animals*

94 Thirty six (36) male Sprague Dawley rats weighing 200-250g were obtained from the Animal
95 Breeding Unit, Faculty of Veterinary Medicine, Universiti Putra Malaysia (UPM). The rats were kept
96 in plastic cages and maintained at room temperature of 25°C ± 2°C with a 12 h light- dark cycle, all
97 rats had free access to food and water during the study period. The rats were allowed to acclimatize
98 for one week prior to the experiment in the animal house, Faculty of Medicine and Health Science,
99 UPM. The animal management and handling procedures was performed based on the recommended
100 institutional animal care and use committee (IACUC) guidelines with the reference number
101 **UPM/IACUC/AUP-R038/2018**, approved on 19th September 2018.

102 2.3. *Experimental design*

103 The rats were randomly divided into five (5) groups (Groups A, B, C, D, and E) with six (6) rats
104 each in group B, C, D and E, while group A consist of twelve (12) rats. Group A (normal control
105 group), received normal saline orally for the whole period of the experiment (8 weeks), Group B
106 designated as the lead treated group (**LTG**), receive 50 mg/kg of Pb acetate orally for 4 weeks
107 (induction of lead toxicity), group C designated as recovery group (**RC**) receive 50 mg/kg of Pb acetate
108 orally for 4 weeks and left with no treatment for 4 weeks, Group D also known as the treatment group
109 1 (**Cur100**) received 50 mg/kg of Pb acetate for 4 weeks orally followed by 100 mg/kg of curcumin for
110 another 4 weeks and group E designated as treatment group 2 (**Cur200**) received 50 mg/kg of Pb
111 acetate for 4 weeks orally followed by 200 mg/kg of curcumin for another 4 weeks. At the end of 4
112 weeks of oral administration of Pb acetate, six rats each from group A and B were euthanatized to
113 confirm the Pb-toxicity via histopathological examination. At the end of the experiment (at week 8)
114 all the rats were sacrificed and tissues were harvested for biochemical and histopathological
115 examinations. Motor activity and weight of the rats was evaluated weekly (figure 1). The dose of Pb
116 acetate [19,20] and curcumin [21] was selected based on previous studies.



117

118 **Figure 1.** Schematic representation of experimental design. HBM-Horizontal bar method, LTG-lead
 119 treated group, ICP-MS-inductive coupled plasma spectrometry.

120 2.4. Motor activity

121 Motor activity of the rats was evaluated using the horizontal bar method.

122 2.4.1. Horizontal bar method

123 Horizontal bar method is a test that measures forelimb strength and coordination in rodents. It
 124 involves the use of 2mm diameter metal bars, 38cm long suspended horizontally above 49cm in
 125 height with support at two end by a laboratory clamp and a padded surface to ensure soft landing of
 126 the experimental animals. The rats were placed on the center of the metal bar by handling them
 127 through their tail to ensure that only the forepaws grasp the metal bar. The falling time of the rats
 128 were recorded by a stopwatch and subsequently translated into scores as described by Deacon [22].

129 2.5. Oxidative stress biomarkers analysis

130 5.5.1. Determination of protein concentration

131 The total protein concentrations of the cerebellar homogenates were measured using the
 132 Bicinchronic acid assay (BCA assay). Bovine serum albumin (BSA) (2 mg/mL) was used as standard
 133 with a working range between 125-2000 $\mu\text{g/mL}$

134 2.5.2. Antioxidant enzyme activity analysis

135 Superoxide dismutase (SOD) enzyme activity was determined in the rats' cerebellum
 136 homogenates using SOD Assay kit (Elabscience, E-BC-K020). The reaction mixture consist of 20 μL of
 137 tissue homogenates, 20 μL of enzyme working solution and 200 μL of substrate application solution

138 fully mixed and incubated at 37°C for 20minutes. The absorbance was measured at wave-length
139 450nm, using micro-plate reader and the results expressed as mgprot/mL.

140 2.5.3. Malondialdehyde (MDA) analysis

141 Malondialdehyde level was determined from the rats cerebellum using MDA ELISA kit (E-EL-
142 006, Elabscience) using the principle of competitive-ELISA and the test was conducted according to
143 manufacturer's manual (Elabscience, USA). Tissue pieces were washed, weighed and finally
144 homogenized in PBS in a ratio of 1:9 with a glass homogenizer on ice. The homogenates were then
145 centrifuged for 5minutes at 5000xg to get the supernatant. The obtained supernatants were used to
146 analyze the MDA using a micro-plate reader at wave-length 450nm. The results for MDA were
147 expressed as ng/mL.

148 5.6. Inductive coupled plasma mass spectrometry (ICP-MS)

149 2.6.1. Sample preparation

150 Harvested cerebellum from the rats was digested with 65% nitric acid using microwave reaction
151 system (Anton Paar, Multiwave PRO). About 0.5 g of the samples were placed in a Teflon vessel and
152 4 mL of nitric acid were added, the samples were then transferred into the microwave oven for 60
153 minutes to obtain a free-contaminated clear digested solution, the solution was diluted with an
154 atomic water to 25 mL in agreement with the digestion protocol as proposed by Rattanachongkiat *et*
155 *al.*, (2004) and Simsek *et al.*, (2017).

156 2.6.2. Sample analysis with ICP-MS

157 Pb was determined with an Agilent 7700x inductively coupled plasma mass spectrometer
158 (Agilent Technologies, Barcelona, Spain) equipped with a Micro Mist nebulizer (Glass Expansion,
159 Melbourne, Australia). Table 1 shows parameters and operation conditions of Agilent 7700x ICPMS.
160 The results were quantified using external calibration standards. Each sample was digested and
161 analyzed in duplicate. Quality control (QC) was performed by analyzing from 20 ppb of calibration
162 standard for every three samples.

163

164 **Table 1.** Work specification resume for ICP-MS Agilent 7700x and measurement parameters.

165

Parameters	Values	Units
RF Power	1550	W
RF Matching	1.6	V
Sample depth	9.5	mm
Torch	-0.1	mm
Torch-V	0	mm
Argon Gas Flow Rate	15	L/min
Carrier Gas Flow	1.01	L/min
Make up Gas Flow	0.15	L/min
Auxiliary Gas Flow Rate	0.58	L/min

Sample Uptake Rate	0.3	rps
Sample Uptake Rate	100	$\mu\text{L}/\text{min}$
Sampling Depth	6 - 7.6	mm
Spray chamber temperature	2	$^{\circ}\text{C}$
Nebulizer Pump	0.1	rps
Integration Time	3	s
Internal Standard ($^{103}\text{Rh},^{208}\text{Bi}$)	200	ppb

166

167 *2.7. Histopathological examination and scoring*

168 The rats' cerebellum were fixed in 10% buffered formalin and prepared for 5 days and
 169 subsequently prepared for histological examination. $5\mu\text{m}$ paraffin sections of the cerebellar tissue
 170 were cut with a microtome (Leica 2235 Microtome, USA) mounted and stained on glass slide with
 171 hematoxylin and eosin (H&E) or toluidine blue stain [25] for histochemistry examination under a
 172 light microscope (Leica DM4M, NY USA). Micrographs were captured using Moticam Pro 282A
 173 5.0MP (Motic images Software Plus 2.0 TWAIN, Hong Kong).

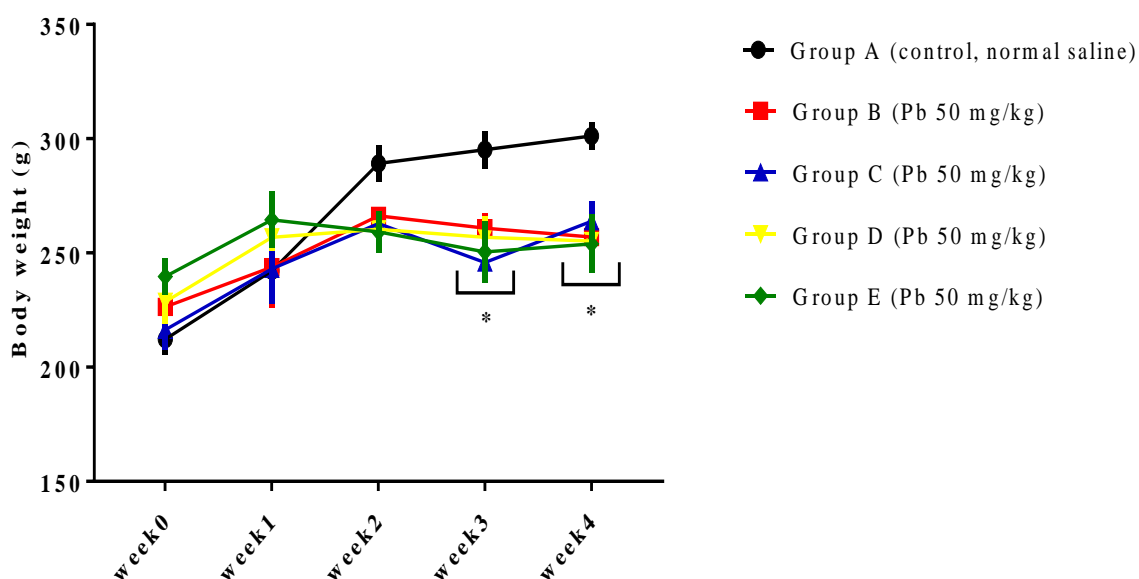
174 The numbers of necrotic Purkinje cells in the cerebellum were quantified in all the rats groups,
 175 using an image analyzer software (Motic images Software Plus 2.0 TWAIN, Hong Kong) in the
 176 histopathology department, faculty of veterinary medicine, UPM. Quantification of the necrotic
 177 Purkinje cells was done at x400 magnification in ten (10) non overlapping fields from six (6) different
 178 sections from 3 rats of each groups. The average values of the necrotic Purkinje cells for the 10 non
 179 overlapping fields were calculated for each section and analyzed with GraphPad prism.

180 *2.8. Statistical analysis*

181 The data obtained from this present study were presented as the mean \pm standard error of mean
 182 (SEM), using unpaired independent student t-test, one-way ANOVA and two-way ANOVA, where
 183 $p < 0.05$ is considered significant using GraphPad prism (GraphPad Prism software, Inc. Version 6.01,
 184 San Diego, California, USA).

185 **3. Results**186 **Induction of Pb acetate toxicity in rats**187 *3.1. Effect of Pb acetate on body weight of rats during Pb toxicity induction*

188 In order to evaluate the effect of Pb acetate on body weight of the experimental rats two-way
 189 ANOVA was employed and the results indicates statistically significant interaction between
 190 treatment effect of Pb acetate on body weight and weeks of induction of Pb-toxicity, [F (16, 125) =
 191 3.01, $p = 0.0003$]. Tukey's post hoc comparison indicates a significant decrease ($p < 0.05$) in body weight
 192 in rats of Group B, C, D and E in week3 and week4 respectively when compared to the body weight
 193 of rats in Group A as shown in figure 2.

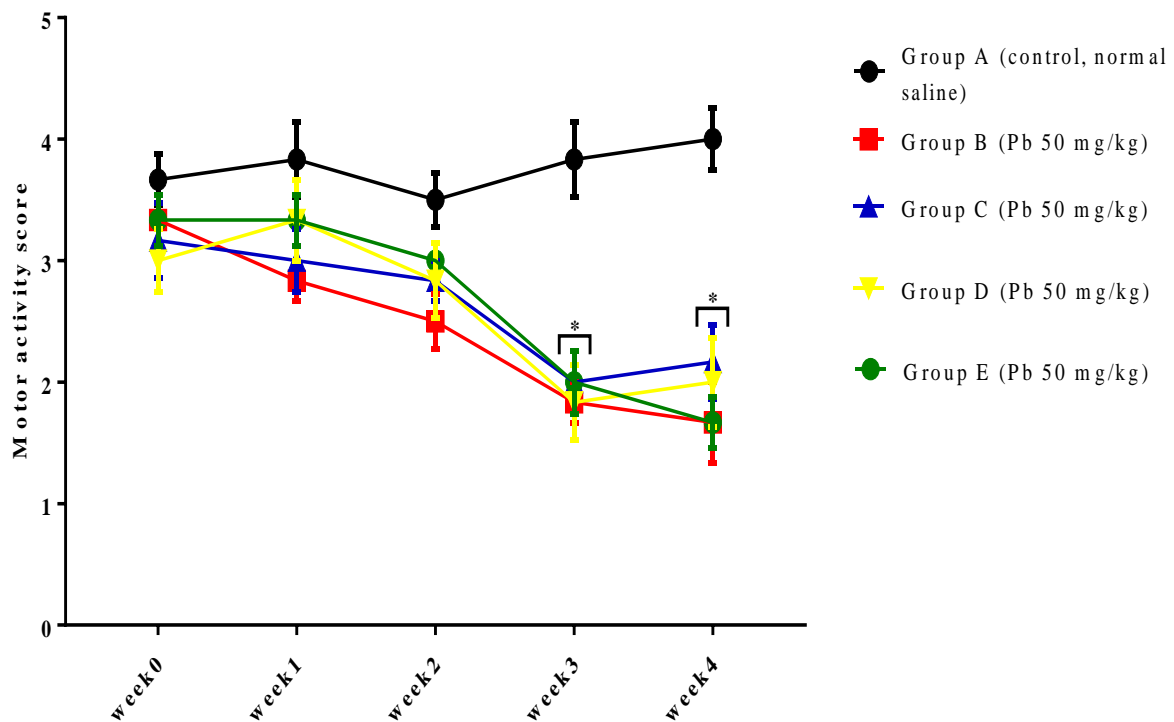


194

195 **Figure 2.** Effect of Pb toxicity on body weight after four (4) weeks of oral administration lead acetate
 196 during induction of Pb toxicity. Data are represented as mean \pm SEM, n = 6. * p < 0.05 vs. control.

197 3.2. Effect of Pb acetate on motor score and coordination in rats during Pb toxicity induction

198 In order to evaluate the effect of oral administration of Pb acetate on motor function of rats,
 199 horizontal bar test was performed. The results indicates statistically significant interaction between
 200 treatment effect of oral administration of Pb acetate and weeks of treatment [$F(16, 125) = 2.23$,
 201 $p = 0.0072$] in motor score of among the rat groups. Tukey's post hoc comparison showed significant
 202 decreases ($p < 0.05$) in motor score of Pb-administered rats in their ability to maintain a hand grip
 203 balance on 2mm horizontal bar at week3 and week4 in rats of Group B, C, D and E compared with
 204 the motor score of rats in Group A as seen in figure 3.



205

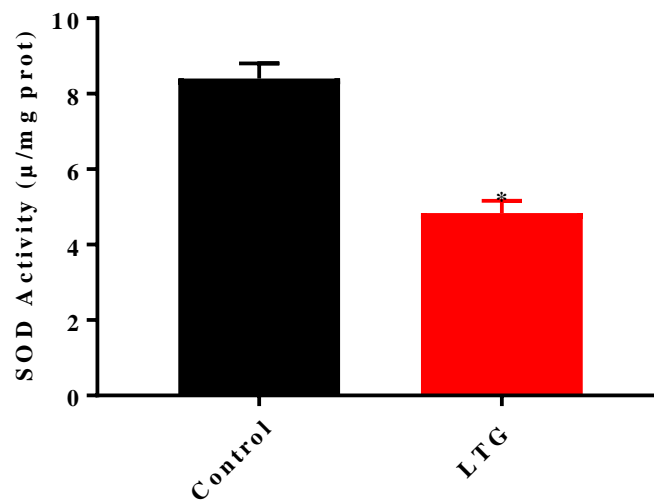
206 **Figure 3.** Effect of oral administration of Pb acetate on motor score of rats during induction of Pb
 207 toxicity. Values were presented as mean \pm SEM (n = 6), * $p < 0.05$ vs. control.

208 3.3. Effect of Pb acetate on oxidative stress status of cerebellum in rats of control and LTG groups during Pb
 209 toxicity induction

210 In order to confirm the induction of Pb toxicity in the rats, the oxidative stress status in their
 211 cerebellum were assessed through unpaired independent student t-test.

212 3.3.1. SOD activity

213 Results from the unpaired independent student t-test (figure 4) showed statistically significant
 214 differences of SOD activity in the cerebellum of rats in the control ($M=8.4$, $SEM=0.4$) and LTG
 215 ($M=4.833$, $SEM=0.3283$); $t(4) = 6.892$, $p = 0.0023$ after treatment with Pb acetate.



216

217 **Figure 4.** Effect of Pb acetate administration on SOD activity in cerebellum of rats after Pb toxicity
218 induction. Data were express as mean \pm SEM, n = 3, * $p < 0.005$ vs. control.

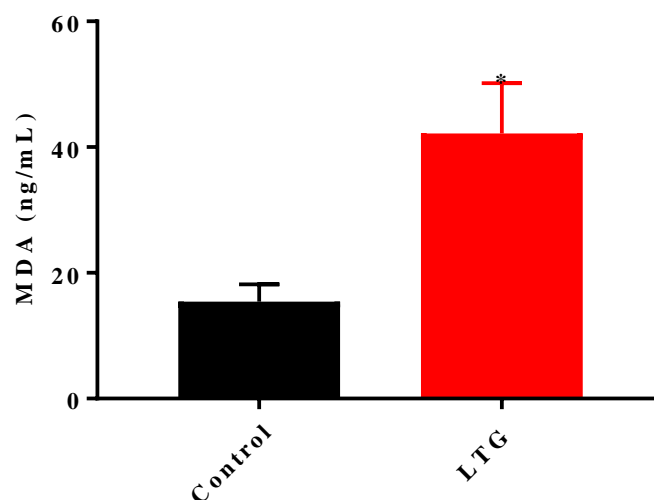
219

220 3.3.2. MDA level

221 Unpaired independent student t-test results in figure 5 indicates statistically significant
222 differences in MDA level in cerebellum of rats in the control ($M=15.4$, SEM = 2.794) and LTG ($M=42.19$,
223 SEM=7.979); $t(4)=3.169$, $p = 0.0339$ after oral administration of lead acetate for four (4) weeks.

224

225



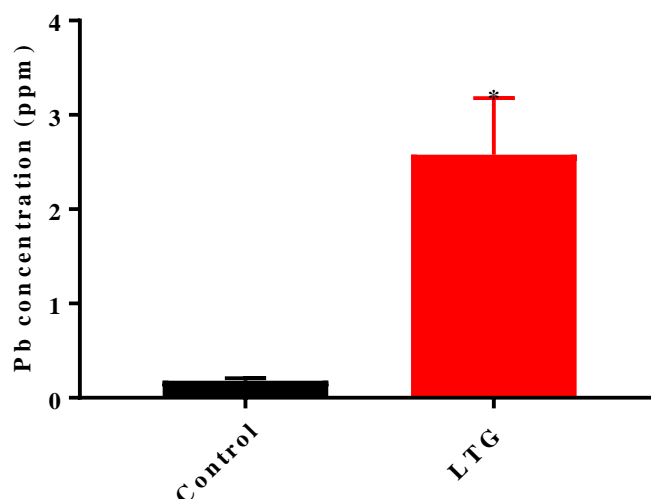
226

227 **Figure 5.** Effect of Pb acetate administration on MDA level in cerebellum of rats after Pb toxicity
228 induction. Data were express as mean \pm SEM, n = 3, * $p < 0.005$ vs. control.

229 3.4. Determination of Pb concentration in the cerebellum of rats in control and LTG groups after Pb toxicity
230 induction.

231 To determine the concentration of Pb in the rat's cerebellum, an ICP-MS techniques was used.
232 Unpaired independent student t-test was employed to analyze the difference of Pb concentration in
233 the cerebellum of rats in the control and LTG after oral administration of Pb acetate. The results
234 indicates statistically significant differences in Pb concentration in the cerebellum of rats in the control
235 ($M=0.2$, $SEM=0.024$) and LTG ($M=2.58$, $SEM=0.6009$); $t(4) = 3.981$, $p=0.0164$ (figure 6).

236



237

238 **Figure 6.** Concentration (ppm) of Pb in rats' cerebellum after induction of Pb toxicity for four (4)
239 weeks. Data are represented as mean \pm SEM, $n = 3$. * $p < 0.05$ vs. control.

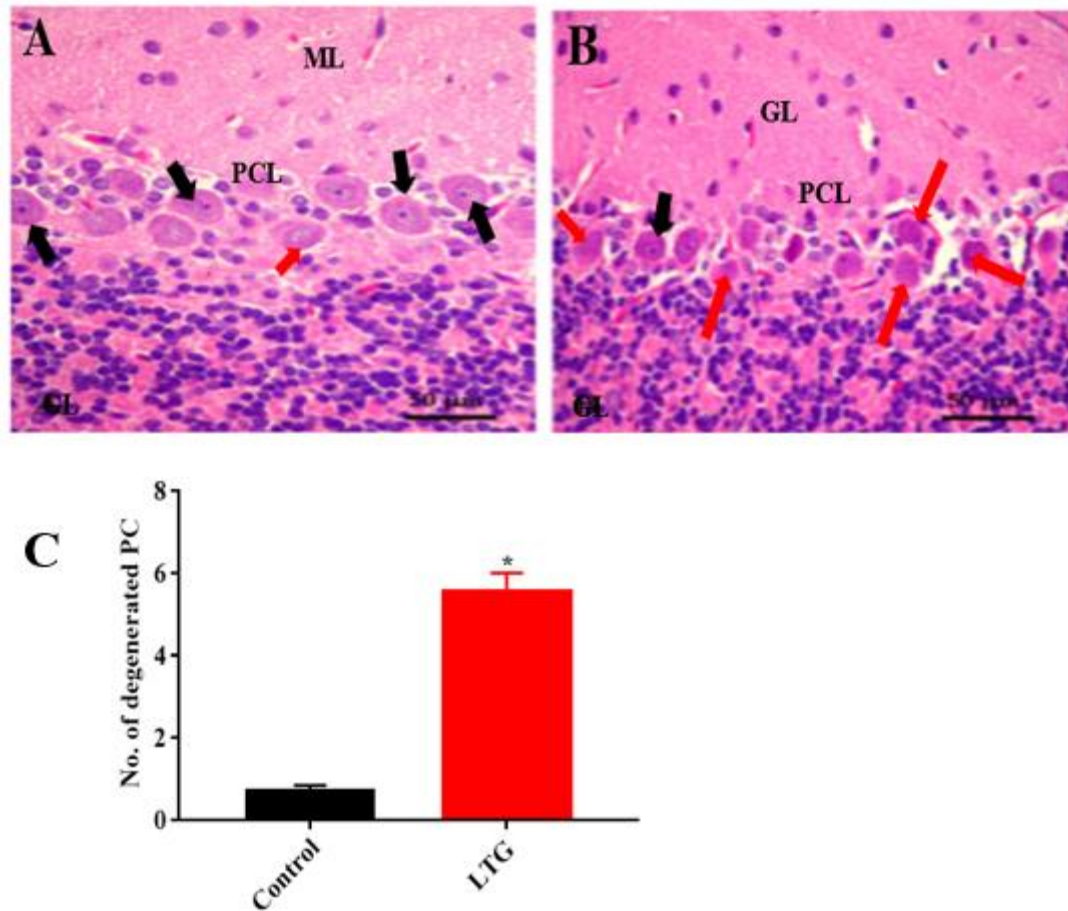
240 3.5. Effect of Pb-acetate on histology of cerebellum of rats in the control and LTG groups after the induction of
241 Pb toxicity

242 3.5.1. Cerebellum stained with hematoxylin and eosin (H&E)

243 The histopathological examination of the cerebellar tissue of Pb-administered rats using H&E
244 stain revealed histological alteration of the cerebellar cortex layers with shrinkage and degeneration
245 of the purkinje and molecular layer cells with scattered glia cells. The purkinje cells exhibited darkly
246 stained nuclei with eosinophilic cytoplasm and empty spaces between them indicating degeneration
247 of the purkinje cells (figure 7B) compared with the control rats (figure 7A).

248 Further, in order to evaluate the effect of Pb-acetate on purkinje cells of the cerebellum,
249 nonparametric t-test was used. Mann-Whitney test indicated that the number of degenerated
250 purkinje cells was statistically significantly greater in the cerebellum of rats in the LTG ($M = 5.5$)
251 group, when compared to the control group ($M = 0.73$), $U = 0$, $p = 0.0022$ (figure 7C).

252



253

254

255

256

257

258

259

Figure 7. Photomicrograph of (A) Control Cerebellar cortex showing the layers of the cerebellum (ML) molecular layer, (PCL) middle purkinje cells layer with large pyriform shape (**black arrow**) and inner granular layer (GL) with aggregation of granular cells. (B) LTG group showing marked degeneration of purkinje cells (**red arrow**). (C) Semi quantitative representation of degenerated Purkinje cells of the control and LTG groups: * $p < 0.05$ vs. control, H&E x400, scale bar 50 μm , $n = 6$.

260 3.5.2. Cerebellum stained with toluidine blue

261

262

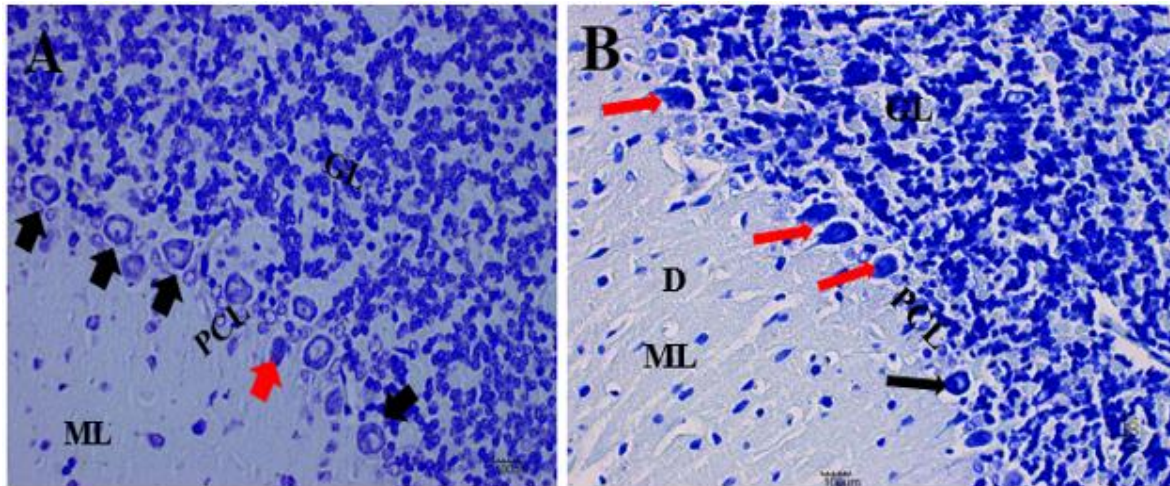
263

264

265

266

In order to further confirm the neurodegenerative effect of Pb-acetate induction on the cerebellum of rats, toluidine blue staining was also performed. The histological results from the cerebellum of rats in the LTG indicates degeneration of the cells of the molecular layer and distortion of the purkinje cells layer with the purkinje cells having a darkly stained cytoplasm and distorted nuclei with empty space between them indicating loss of the purkinje cells (figure 8B) compared with rats in the control (figure 8A).

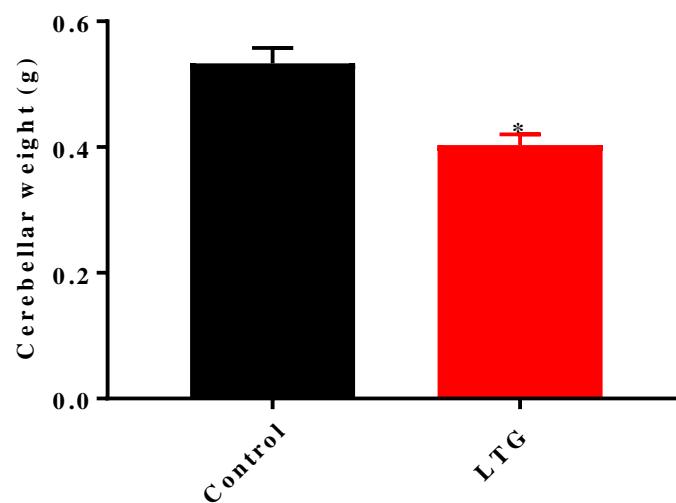


267
268
269
270
271
272
273
274
275

Figure 8. Photomicrograph sections of the cerebellum from rat's groups (A) Control showing molecular layer (ML), the granular layer (GL) and Purkinje cells layer (PCL). The Purkinje cells (black arrow) with prominent nuclei. (B) LTG showing molecular layer (ML) with degenerated area of cells (D), Purkinje cells with darkly stained cytoplasm and distorted nuclei leaving spaces between them (red arrow). The granular layer (GL) indicates deeply stained cells with vacuolation (Toluidine blue x200 scale bar = 100µm).

276 3.5.3. Effect of Pb acetate on the weight of the cerebellum of control and LTG rats groups after Pb
277 toxicity induction

278 Unpaired t-test was employed to evaluate the effect of Pb acetate administration on cerebellar
279 weight in rats of the control and LTG groups. The results showed statistically significant differences
280 of cerebellar weight in the control group of rats ($M=0.5333$, $SEM=0.02418$), when compared to LTG
281 groups of rat ($M=0.4033$, $SEM=0.1706$); $t(10)=4.393$, $p=0.0013$ (figure 9).

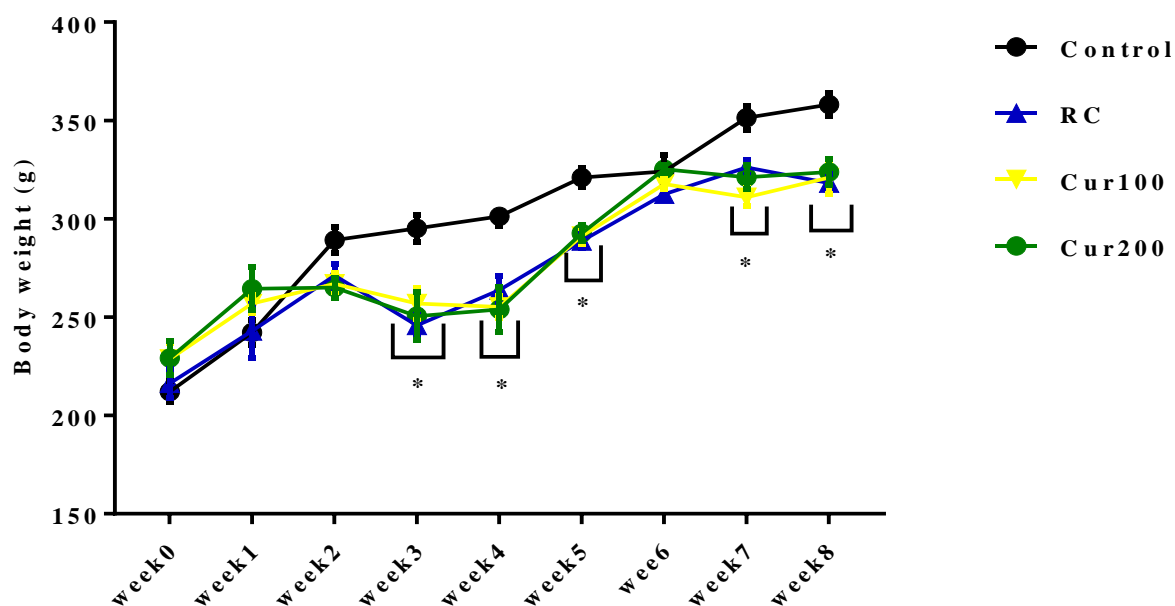


282
283
284
285

Figure 9. Effect of Pb acetate on the weight of the cerebellum of control and LTG. Data are represented as mean ± SEM, n=6, * $p<0.05$ vs. control

286 **Treatments of Pb acetate induced rats with curcumin**287 *3.6. Effect of curcumin on the body weight of Pb induced rats*

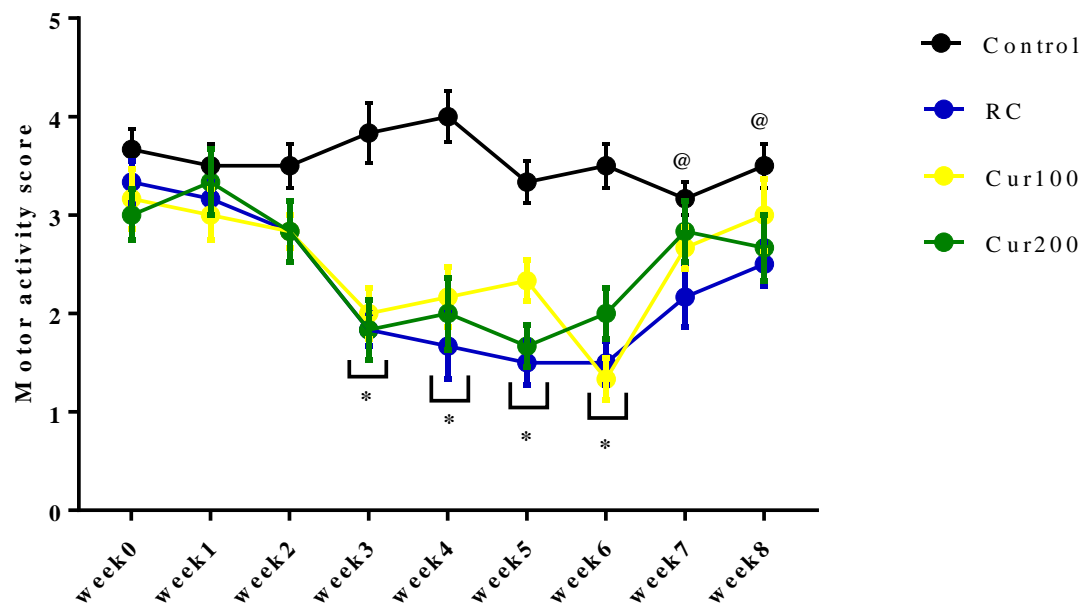
288 Two-way ANOVA showed statistically significant interaction between treatment effect of
 289 curcumin on body weight of rats and the weeks of administration, [F (24,180) = 3.242, $p=0.0001$].
 290 Tukey's post hoc test indicates a significant decrease ($p<0.05$) in body weight of rats in the RC, Cur100
 291 and Cur200 groups in week3, week4, week5, week7 and week8 respectively when compared to body
 292 weight of rats in the control as shown on figure 10.



293 **Figure 10.** Effect of oral administration of curcumin on body weight of Pb acetate induced rats. Data
 294 are represented as mean \pm SEM, $n = 6$. * $p<0.05$ vs. control.
 295
 296

297 *3.7. Curcumin ameliorates Pb-induced alteration of motor coordination of rats in horizontal bar test.*

298 Two-way ANOVA showed statistically significant interaction between treatment effect of oral
 299 administration of curcumin and weeks of treatment, [F (24, 180) = 2.448, $p=0.0004$] in the motor score
 300 of Pb induced rats treated with curcumin (figure 11). Tukey's post hoc comparison indicates
 301 statistically significant decrease ($p<0.05$) in motor score of rats in the RC, Cur100 and Cur200 in the
 302 ability to maintain a hand grip balance on the 2mm horizontal bar in week3, week4, week5 and week6
 303 when compared the control group of rats. Additionally, Tukey's post hoc test further reveal a
 304 significant increase ($p<0.05$) in the motor score of rats in the control group in week7 and week8, when
 305 compared to motor score of rats in the RC group rats.



306

307 **Figure 11.** Effect of oral admiration of curcumin on Pb induced rats in horizontal bar test. Values
 308 were presented as mean \pm SEM (n = 6), * $p < 0.05$ vs. control, @ $p < 0.05$ vs. RC.

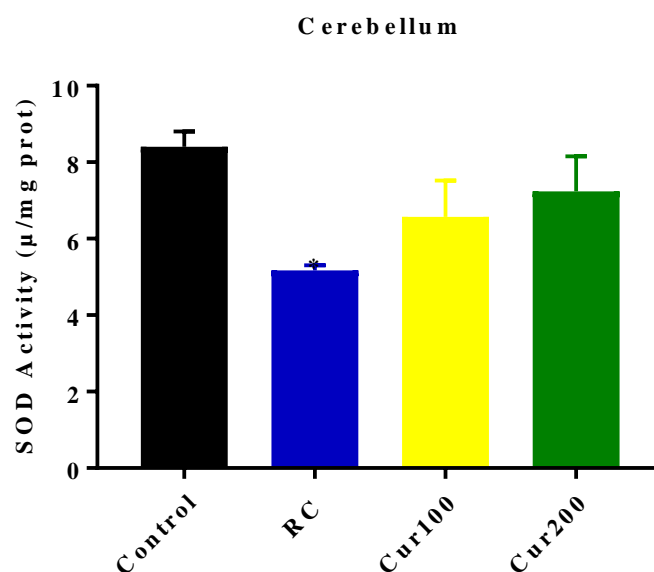
309

310 3.8. Curcumin reverse Pb induced oxidative stress in rats' cerebellum.

311 In order to assess the antioxidant properties of curcumin on Pb-induced oxidative stress in rats,
 312 the cerebellar homogenates of the rats were analyzed for SOD activity and MDA levels.

313 3.8.1. SOD activity

314 As shown in figure 12, one-way ANOVA revealed statistically significant differences in SOD
 315 activity in the cerebellum of different rats groups [F (3, 8) = 3.768, $p = 0.0493$]. Tukey's post hoc test
 316 showed statistically significant decrease in SOD activity in cerebellum of the RC (5.167 ± 0.133 , $p =$
 317 0.0443) when compared with the control group of rats (8.4 ± 0.4).

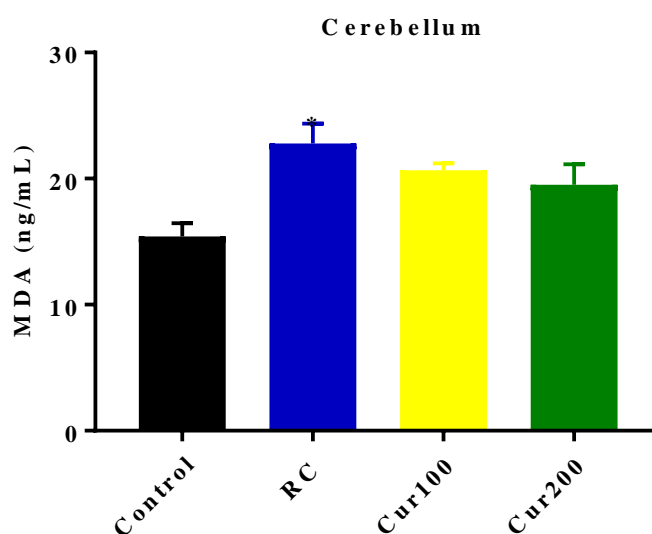


318

319 **Figure 12.** Effect of curcumin on SOD activity in the cerebellum of Pb induced rats. Data were express
320 as mean \pm SEM, n = 3, * $p < 0.05$ vs. control.
321

322 3.8.2. MDA level

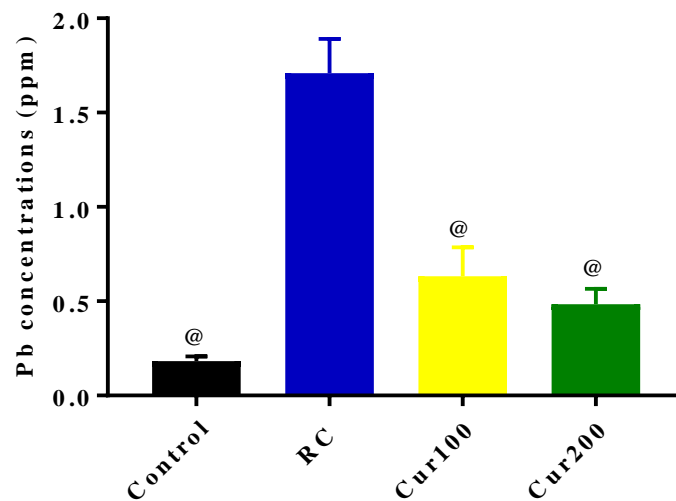
323 One-way ANOVA showed statistically significant differences of MDA level in the cerebellum
324 of all the rats groups, [F (3, 8) = 5.844, $p = 0.0205$]. Tukey's comparison test revealed significant increase
325 in MDA level in the cerebellum of rats from RC group (22.78 ± 1.579 , $p = 0.0153$), when compared to
326 the control group of rats (15.4 ± 1.062) as shown in figure 13.



327 **Figure 13.** Effect of curcumin on MDA level in the cerebellum of Pb induced rats. Data were express
328 as mean \pm SEM, n = 3, * $p < 0.05$ vs. control.
329
330

331 3.9. Chelating potentials of curcumin on Pb induced toxicity in rats

332 The results obtained from ICP-MS analysis was subjected to one-way ANOVA to evaluate the
333 mean Pb concentration in the cerebellum of rats in the control, RC, Cur100 and Cur200. Results
334 obtained showed statistically significant differences in Pb concentration in the cerebellum of different
335 rats groups, [F (3, 8) = 27.55, $p = 0.0001$]. Tukey's post hoc comparison indicated significant decreases
336 in Pb concentration in the cerebellum of rats in the control (0.1828 ± 0.02414 , $p = 0.0001$), Cur100
337 (0.6319 ± 0.1545 , $p = 0.0014$) and Cur200 (0.4848 ± 0.08147 , $p = 0.006$), when compared to rats in the RC
338 group (figure 14).
339



340

341 **Figure 14.** Concentration of Pb in rats' cerebellum after oral administration of curcumin and
342 withdrawal of Pb acetate for four (4) weeks. Data are represented as mean \pm SEM, n=3. @ $p < 0.05$ vs.
343 RC.

344

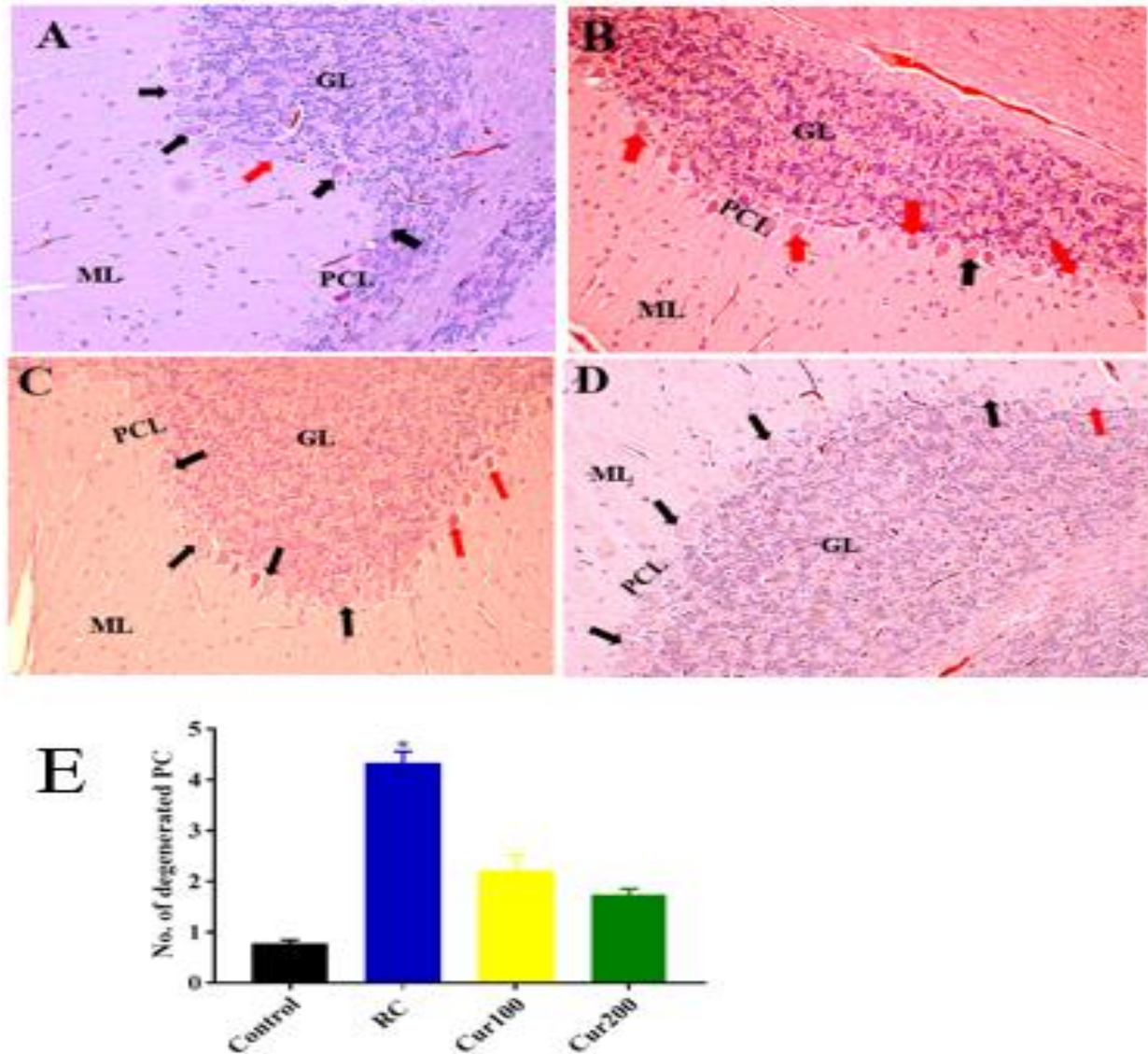
345 3.10. Curcumin attenuates Pb induced cerebellar damage in rats cerebellum

346 2.10.1. H&E staining

347 The histology results from the cerebellum of rats in the RC groups of rats stained with H&E after
348 withdrawal of Pb-acetate indicated no recovery when compared to rats in control group (figure 15A).
349 The RC group cerebellum revealed eosinophilic purkinje cells with dark and irregular nuclei with the
350 molecular layer appeared to have scattered glia cells with perineural spaces, thus the granular layer
351 appeared to have normal histological appearance (figure 15B).

352 Oral administration of curcumin for four (4) weeks attenuated the pathological changes in the
353 cerebellum of rats in the Cur100 (figure 15C) and Cur200 (figure 15D)

354 In addition, semi quantitative analysis of the purkinje cells of the cerebellum revealed
355 statistically significant differences in degenerated purkinje cells of the experimental rats, [H (3) =
356 19.75, $p = 0.0002$], with a mean rank of 3.583 for control, 21.5 for RC, 13.83 for Cur100 and 11.08 for
357 Cur200. Dunn's multiple comparison test further showed statistically significant increases ($4.333 \pm$
358 0.2246 , $p=0.0001$) in degenerated purkinje cells in the cerebellum of the RC group of rats, when
359 compared to the rats in the control group (figure 15E).



360

361 **Figure 15.** Photomicrograph sections of the cerebellum of rat's groups (A) control indicating layers
 362 of the cerebellar cortex, the molecular layer (ML) with glia cells (**red arrow**), middle Purkinje
 363 layer (PCL) with Purkinje cell having a large pyriform shape (**black arrow**) and inner granular
 364 layer with aggregation of granular cells (GL). (B) RC showing eosinophilic Purkinje cells with
 365 irregular dark cytoplasm (**red arrow**) and scattered glia cells in the molecular layer (ML). (C)
 366 Cur100 showing the Purkinje cells with prominent nuclei and regular shape (**black arrow**) and
 367 the molecular and granular layer appear normal. (D) Cur200 showing normal pyriform shaped
 368 purkinje cells with prominent nuclei (**black arrow**) with normal granular and molecular layer.
 369 (E) Semi quantitative representation of degenerated Purkinje cells: * $p < 0.005$ vs. control, H&E x200, scale bar 100 μm .

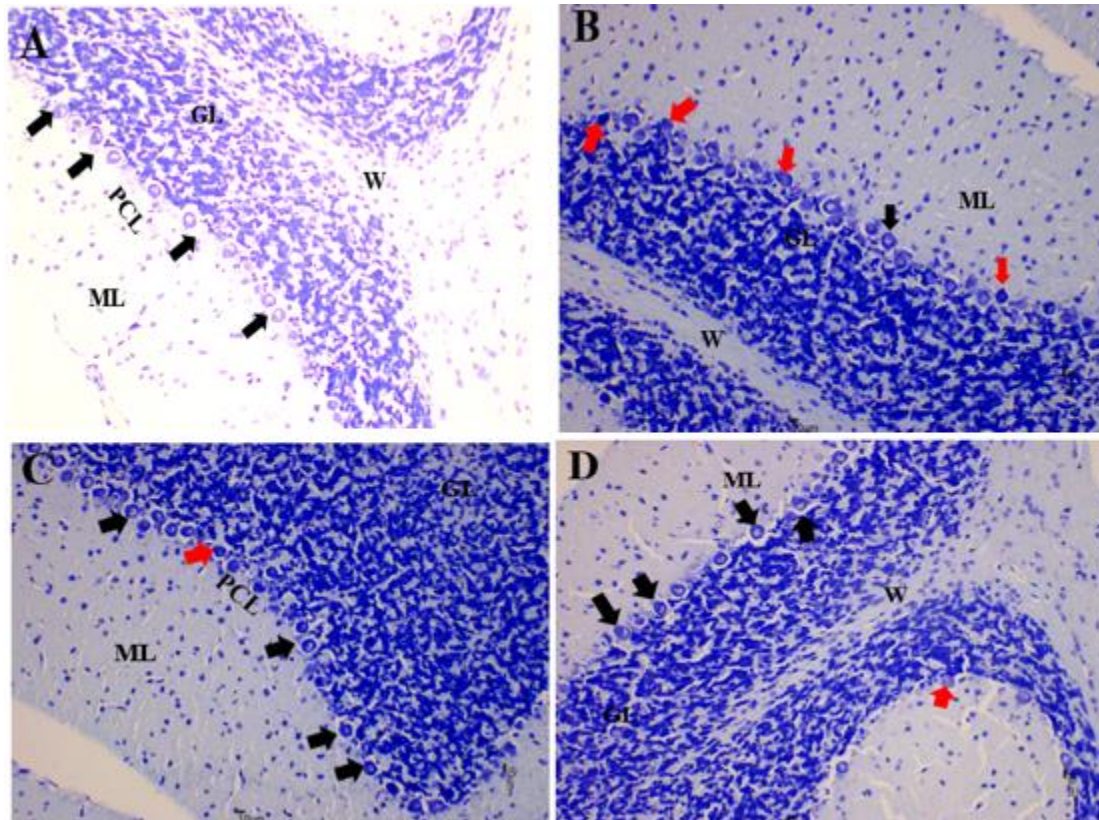
370 3.10.2. Toluidine blue staining

371 The photomicrograph section from the cerebellum of the control group of rats indicates normal
 372 histological structure of the cerebellum with the three layers of the cerebellar cortex. Purkinje
 373 cells appeared to have a regular prominent central nucleus with an apical dendrites. Granular and
 374 molecular layer showed normal cells with dark stained nuclei (figure 16A).

375 Photomicrograph section from the cerebellum of RC group of rats revealed alteration in the purkinje
 376 cells layer with the purkinje cells appeared to have darkly stained cytoplasm with irregular nuclei.
 377 The molecular layer showed scattered glial cells, although the granular layer appeared to be normal
 378 with granular cells (figure 16B).

379 The photomicrograph section from the Cur100 and Cur200 showed restoration of the purkinje
 380 cells layer with healthy purkinje cells. The molecular and granular layers appeared to be normal with
 381 normal cells (figure 16C and figure 16D).

382

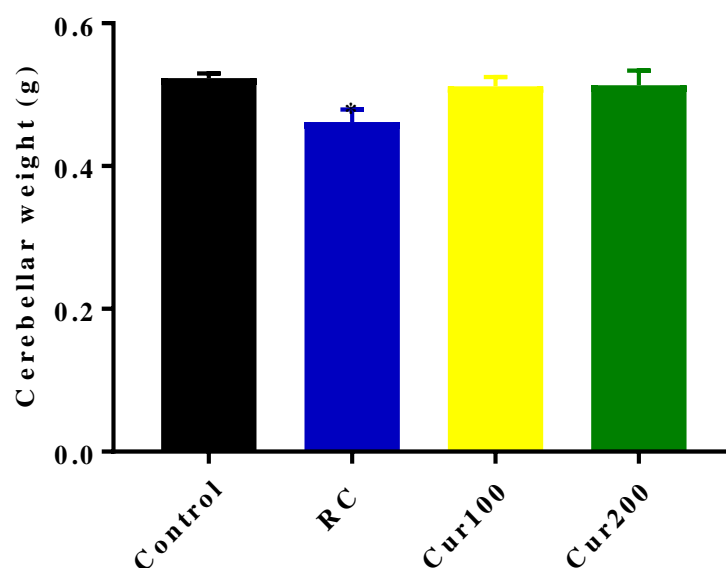


383

384 **Figure 16.** Photomicrograph sections of the cerebellum from rat's groups (A) Control showing the
 385 white matter (W) and the three layers of the cerebellar cortex, molecular layer (ML), the granular
 386 layer (GL), the purkinje cells layer (PCL) with the Purkinje cells (**black arrow**) with prominent nuclei.
 387 (B) RC showing the purkinje cells having deeply stained cytoplasm with distorted shape (**red arrow**)
 388 and the molecular layer showing scattered glia cells (ML). (C) Cur100 and (D) Cur100 showing the
 389 restoration of the molecular (ML) and granular layer (GL), the Purkinje cells layer (PCL) appeared to
 390 be arranged in a linear pattern with the Purkinje cells (**black arrow**) having prominent nuclei and
 391 regular shape. Toluidine blue, x200 scale bar = 100µm.

392 3.10.3. Effects of curcumin administration on cerebellar weight

393 One-way ANOVA indicates statistically significant differences in weight of the cerebellum
 394 among the groups of rats [F (3, 20) = 3.195, $p = 0.457$]. Tukey's post hoc comparison revealed statistically
 395 significant decreases in the cerebellum weight of RC group of rats (0.4617 ± 0.01778 , $p = 0.0486$), when
 396 compared to control group as shown in figure 17.



397

398 **Figure 17.** Effect of curcumin administration on cerebellar weight of Pb induced rats. Data are
399 represented as mean \pm SEM, n=6. *p<0.05 vs. control.

400

401 4. Discussion

402 Preceding studies have documented the toxic effects of Pb on biological systems resulting into several
403 pathologies and clinical implications with morbidity on almost all the organs with the brain, kidney
404 and liver serving as primary targets [12,26,27]. These pathological alterations may include increase
405 oxidative stress [9], neurological deficit [28], decrease motor coordination and cognitive deficit
406 [29,30]. Alteration in astrocyte maturation, degeneration of neural cells [28,30], renal dysfunction,
407 degenerative changes in tubular epithelium and hepatotoxicity [2,31] were also reported in Pb
408 poisoning both in humans and different rodents species. In the present study, rat model of Pb toxicity
409 was used to investigate the therapeutic potentials of curcumin on fundamental mechanism of motor
410 dysfunctions and neurodegeneration caused by Pb toxicity. The Pb induced rats developed a
411 remarkable deficiency in cerebellum dependent horizontal bar test for motor functions, increased
412 oxidative stress, besides marked degeneration of cells in molecular and purkinje cell layers of the
413 cerebellum and high concentration of Pb in their cerebellum. However, treatment with curcumin
414 irrespective of the dose given attenuated the above aforementioned alterations and aberrations
415 caused by Pb acetate induced toxicity in the rats.

416 Worthy to note in this study, is the choice of male Sprague Dawley rats for all the
417 experimentations. As female sex hormones such as estrogen, prolactin and progesterone may
418 influence cognitive function, emotion and motor behavior in rats, because their cyclical hormonal
419 changes usually comes with mood swing. Hence, female rats were less preferred by the authors for
420 pharmacological testing of phytochemical substance like curcumin in order not to be a confounding
421 factor [32–34]. Therefore, healthy male Sprague Dawley rats were used in this present study to
422 evaluate the neurotherapeutic effects of oral administration of curcumin on Pb-induced toxicity of
423 motor function and coordination.

424 Coordination of motor behavior and cognitive function depend on the integrity of the nervous
425 system and core psychological functions of the animals [22,35]. The cerebellum is a delicate structure
426 that is vulnerable to intoxication and poisoning with the cerebellar circuit especially the purkinje cells
427 susceptible to injury after exposure to environmental toxins such as Pb [36]. Further, the cerebellum
428 plays a vital role in the unification of motor and sensory functions, any lesion on the cerebellum may
429 resulted into deformation of motor coordination and balance [37]. In this study decrease in motor
430 score on horizontal bar method test were observed in Pb induced rats which is in agreement with the
431 previous works documented by Nehru and Sidhu [38], Barkur and Bairy [39], and Sabbar *et al.* [40]
432 who also reported decrease in cognitive and motor function in rat models of Pb toxicity. Mason *et al.*
433 [7] also reported that Pb exposure affect motor function such as deficits in visuomotor coordination
434 in adult Pb workers and children exposed to Pb. The mechanism in which Pb execute this decrease
435 motor function could be due to its ability to induce oxidative stress in rats exposed to Pb [6]. However,
436 the present study observed increase in motor score of rats treated with curcumin regardless of the
437 dose given. Moore *et al.* [41], also observed a similar findings and attributed it to the antioxidant and
438 anti-inflammatory properties of curcumin on motor function. Chongtham and Agrawal [42] also
439 reported that curcumin ameliorates disease symptoms in *Drosophila* model of Huntington disease
440 (HD). However, supplementation of diet with curcumin for 12 weeks in healthy aging population
441 does not influenced their motor performance [43].

442 Alterations of oxidative status either by overproduction of oxidant or deficit in antioxidant
443 activity could be one of the direct consequences of Pb-toxicity and poisoning in living organisms [6].
444 Preceding studies indicates that oxidative stress is linked to the pathogenesis of Pb-toxicity resulting
445 in lipid peroxidation [6], neurodegeneration [44], oxidation of hemoglobin [45] and impairment of
446 fundamental biological cellular process such as cell adhesion, intra and inter cellular signaling, ionic
447 transportation, enzyme regulation, neurotransmitters release and apoptosis [12]. The fundamental
448 oxidant that plays vital role in redox reaction includes hydroxyl radical (OH), hydroxyl anion (OH⁻),
449 hydrogen peroxide (H₂O₂), nitric oxide (NO) and peroxynitrite (ONOO⁻) [46]. MDA or thiobarbituric
450 acid-reactive species (TBARS) is the end product of lipid peroxidation that plays a vital role in lipid
451 membrane damage in cells due to increase reactive oxygen species (ROS) [6]. The present study
452 revealed an increased level of MDA and decreased SOD activity in the cerebellum of Pb-induced rats.
453 However, these alterations were ameliorated by treatment with curcumin irrespective of the dose
454 given. These findings were in agreement with the previous studies that have reported the antioxidant
455 properties of curcumin [47–49].

456 Inductive coupled plasma mass spectrometry (ICP-MS) is a robust technique for molecular
457 analysis of element and physiochemical compound with separation techniques that makes it suitable
458 for detecting elements in pharmaceutical research and for specific investigation of element present in
459 molecule [50]. ICP-MS is a multi-element system techniques in the analysis of biological fluids with
460 greater sensitivity and selectivity when compared to inductive coupled plasma-optical emission
461 spectrometry (ICP-OES) and graphite furnace atomic absorption spectrometry (GF-AAS) [24].
462 Concentration of trace elements beyond physiological limits in organs can be toxic in both animals
463 and humans. Likewise, concentration of heavy metals such as Pb in the biological system are known
464 to be toxic thus, affecting biochemical reactions [24]. Further, bones remain a vital site for Pb
465 accumulation after exposure, although circulating Pb in the blood can be distributed to various vital
466 organs such as the brain, kidney and liver [51,52]. This present study decided to use ICP-MS for its

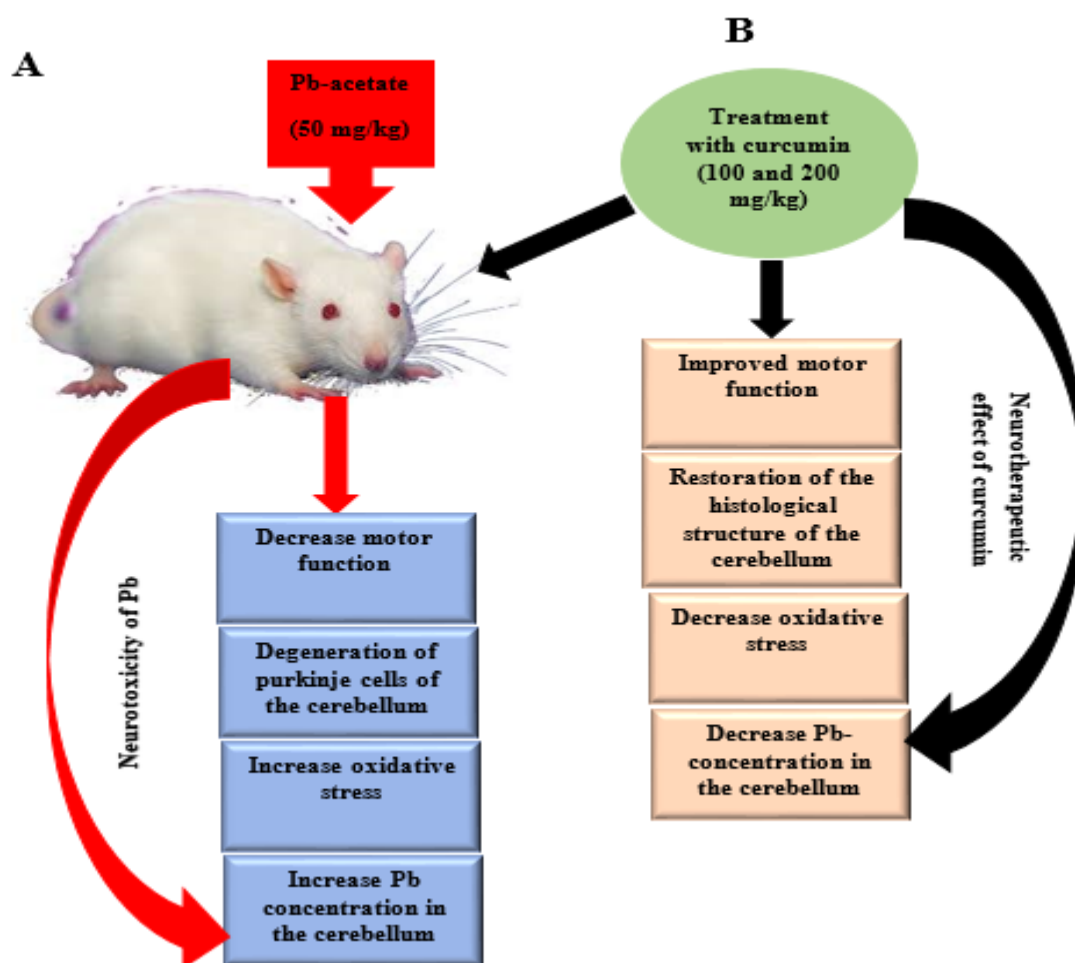
467 superiority over other techniques and found significantly high concentration of Pb in the cerebellum
468 of Pb-induced rats when compared to their control counterparts. The results is in agreement with the
469 previous works of Sousa *et al.* [52] and Simsek *et al.* [24] that reported increased concentration of Pb
470 in the brains of Pb-induced rats. Noteworthy, is administration of curcumin to Pb induced rats
471 decreased the concentration of Pb in their cerebellum. These findings were in accordance with the
472 previous studies of Mary *et al.* [18] and Shen *et al.* [53], who reported the chelating properties of
473 curcumin.

474 In the present study, multiple pathological lesions were observed in the cerebellum of Pb
475 induced rats. These changes includes neuronal damage and alteration of the histological architecture
476 of the cerebellar cortex. The purkinje cells exhibited eosinophilic cytoplasm with darkly stained and
477 irregular nuclei leaving empty spaces between them. Semi quantitative analysis of purkinje cells
478 showed significant increase ($P<0.05$) in the number of necrotic purkinje cells. Further, Pb-induced rats
479 showed decreased ($P<0.05$) cerebellar weight, which could be attributed to the marked degeneration
480 of the cerebellar cells observed. These results are in accordance with several preceding studies
481 [9,26,37,54,55] that reported similar findings of neuronal degeneration in Pb and other heavy metals
482 toxicity on the cerebellum with more consequences on the purkinje cells due to sensitivity of the
483 purkinje cells layer. However, treatment with curcumin reverses the above mentioned Pb-induced
484 morphological aberrations in the rats. This could be due to the antioxidant properties of curcumin as
485 seen in the present study or combined antioxidant and anti-inflammatory role of curcumin in
486 neurotoxicity and neurodegenerative diseases as documented by previous studies [16,56,57].

487 The most important step in dealing with Pb-toxicity is to avoid exposure to the sources of Pb
488 contamination which might not always be feasible. Therefore, there exists standard drugs chelators
489 in the markets that are being used in terms of heavy metal poisoning, however, they are expensive
490 and exhibits a lot of adverse effects [12]. This study revealed that withdrawal of Pb acetate alone was
491 not enough to restore the damages inflicted by Pb-toxicity in rats of the RC group. The persistent
492 changes observed after the Pb withdrawal includes, morphological aberrations of the cerebellum,
493 decrease in motor score on horizontal bar test, oxidative stress as well as high concentration of Pb the
494 cerebellum on ICP-MS analysis when compared to the curcumin treated groups. These could be due
495 to specific kinetics, where portion of Pb absorbed is accumulated in the bone [11,58,59], thus
496 gradually released to soft tissues such as the brain, kidney and liver [58–60]. Similar findings were
497 reported by Omobowale [61]; Nehru and Sidhu [38]; and Khalaf *et al.* [62] after withdrawal of Pb
498 exposure in rats model of Pb-toxicity.

499 5. Conclusions

500 In consideration to the results obtained from this study, Pb-toxicity resulted into decreased motor
501 functions, increased oxidative stress, aberrations in the histological structure of the cerebellum as
502 well as accumulation of Pb in the cerebellum of affected rats. Treatment with curcumin regardless to
503 the dose attenuates the abnormalities caused by Pb-toxicity, which could be due to the antioxidant
504 and chelating properties of curcumin (figure 18). It is noteworthy, to conclude that curcumin could
505 be develop as natural drug for treatment of Pb-toxicity due to its therapeutic potentials and wide
506 pharmacological safety margin. Finally, the authors recommend future research on longer treatment
507 periods with to assess the effectiveness of its long term preclinical applications in Pb-induced toxicity
508 and neurodegeneration.



509

510 **Figure 18.** Proposed mechanism of Pb-induced cerebellar toxicity and the ameliorative effects of
 511 curcumin.

512

513 Abbreviations

514 ABLES: adults blood lead epidemiology and surveillance; BLLs: blood lead levels; Pb: lead; Pb-acetate:
 515 lead acetate; ROS: reactive oxygen species; BBB: blood brain barrier; SAH: subarachnoid hemorrhage;
 516 MDA: malondialdehyde; MMP-9: matrix Metalloproteinase 9; Mn^{2+} : manganese ion; Fe^{2+} : iron ion; Cu^{2+} :
 517 copper ion, Zn^{2+} : zinc ion, HBM: horizontal bar method, BCA: Bicinchoninic acid assay, BSA: bovine
 518 serum albumin; SOD: superoxide dismutase; OD: optical density; ICP-MS: inductive coupled plasma
 519 spectrometry; QC: quality control; ANOVA: analysis of variance; H&E: hematoxylin and eosin; HD:
 520 Huntington disease.

521 **Author Contributions:** “conceptualization, K.A.; E.A.R and Md.Z.A.; methodology K.A.; A.D.; E.A.R and
 522 Md.Z.A, validation, S.M.C; A.D.; and Md.Z.A.; formal analysis K.A.; A.D. and M.M.M. investigation, K.A.;
 523 M.M.M.; A.D.; E.A.R, and Md.Z.A.; data curation, K.A; M.M.M.; S.M.C.; and; A.D; writing—original draft
 524 preparation, K.A.; writing—review and editing, K.A.; M.M.M.; A.D. and S.M.C.; supervision, E.A.R.; Md. Z.A.

525 **Funding:** This research was funded by Universiti Putra Malaysia, (Grant number GP-IPS 9663600).

526 **Acknowledgments:** The authors would like to acknowledge Universiti Putra Malaysia for funding this research
 527 project (Grant number GP-IPS 9663600). The authors would also like to acknowledge the support and effort of
 528 Mr. Saipulzaman Ali and Mrs. Jamila Histopathology department, faculty of veterinary medicine, UPM.

529 **Conflicts of Interest:** The authors declare no conflict of interest.

530

531 **References**

- 532 1. El-Tantawy, W.H. Antioxidant effects of Spirulina supplement against lead acetate-induced hepatic
533 injury in rats. *J. Tradit. Complement. Med.* **2016**, *6*, 327–331.
- 534 2. Abdel-Zaher, A.O.; Abd-ellatief, R.B.; Aboulhagag, N.A.; Farghaly, H.S.M.; AL-Wasei, F.M.M. The
535 interrelationship between gasotransmitters and lead-induced renal toxicity in rats. *Toxicol. Lett.* **2019**,
536 *310*, 39–50.
- 537 3. Shaffer, R.M.; Gilbert, S.G. Reducing occupational lead exposures: Strengthened standards for a healthy
538 workforce. *Neurotoxicology* **2017**.
- 539 4. Kosnett, M.J.; Wedeen, R.P.; Rothenberg, S.J.; Hipkins, K.L.; Materna, B.L.; Schwartz, B.S.; Hu, H.; Woolf,
540 A. Recommendations for medical management of adult lead exposure. *Environ. Health Perspect.* **2007**,
541 *115*, 463–471.
- 542 5. Chowdhury, R.; Ebel Sarnat, S.; Darrow, L.; McClellan, W.; Steenland, K. Mortality among participants
543 in a lead surveillance program. *Environ. Res.* **2014**, *132*, 100–104.
- 544 6. Flora, G.; Gupta, D.; Tiwari, A. Toxicity of lead: A review with recent updates. *Interdiscip. Toxicol.* **2012**,
545 *5*, 47–58.
- 546 7. Mason, L.H.; Harp, J.P.; Han, D.Y.; Mason, L.H.; Harp, J.P.; Han, D.Y. Pb neurotoxicity:
547 Neuropsychological effects of lead toxicity. *Biomed Res. Int.* **2014**, *2014*, 1–8.
- 548 8. Chiodo, L.M.; Jacobson, S.W.; Jacobson, J.L. Neurodevelopmental effects of postnatal lead exposure at
549 very low levels. *Neurotoxicol. Teratol.* **2004**, *26*, 359–371.
- 550 9. Radad, K.; Hassanein, K.; Al-Shraim, M.; Moldzio, R.; Rausch, W.D. Thymoquinone ameliorates lead-
551 induced brain damage in Sprague Dawley rats. *Exp. Toxicol. Pathol.* **2014**, *66*, 13–17.
- 552 10. Andjelkovic, M.; Djordjevic, A.B.; Antonijevic, E.; Antonijevic, B.; Stanic, M.; Kotur-Stevuljevic, J.;
553 Spasojevic-Kalimanovska, V.; Jovanovic, M.; Boricic, N.; Wallace, D.; et al. Toxic effect of acute cadmium
554 and lead exposure in rat blood, liver, and kidney. *Int. J. Environ. Res. Public Health* **2019**, *16*.
- 555 11. Ishii, C.; Nakayama, S.M.M.; Kataba, A.; Ikenaka, Y.; Saito, K.; Watanabe, Y.; Makino, Y.; Matsukawa,
556 T.; Kubota, A.; Yokoyama, K.; et al. Characterization and imaging of lead distribution in bones of lead-
557 exposed birds by ICP-MS and LA-ICP-MS. *Chemosphere* **2018**, *212*, 994–1001.
- 558 12. Kabeer, A.; Mailafiya, M.M.; Danmaigoro, A.; Rahim, E.A.; Bakar, Z.A. Therapeutic potential of
559 curcumin against lead-induced toxicity : A review. *Biomed. Res. Ther.* **2019**, *6*, 3053–3066.
- 560 13. Ghosh, S.S.; Gehr, T.W.B.; Ghosh, S. Curcumin and chronic kidney disease (CKD): Major mode of action
561 through stimulating endogenous intestinal alkaline phosphatase. *Molecules* **2014**, *19*, 20139–20156.
- 562 14. Hwang, E.S.; Lim, S.M.; Woo, E.J.; Kim, H.B.; Lee, S.; Choi, B.K.; Kwon, O.I.; Kim, H.J.; Kim, J.W.; Kyung,
563 E.J. Evaluation of Hepatoprotective Effect of Curcumin on Liver Cirrhosis Using a Combination of
564 Biochemical Analysis and Magnetic Resonance-Based Electrical Conductivity Imaging. *Mediators
565 Inflamm.* **2018**, *2018*, 1–9.
- 566 15. Hewlings, S.; Kalman, D. Curcumin: A Review of Its' Effects on Human Health. *Foods* **2017**, *6*, 92.
- 567 16. Shi, L.-Y.; Zhang, L.; Li, H.; Liu, T.-L.; Lai, J.-C.; Wu, Z.-B.; Qin, J. Protective effects of Curcumin on
568 acrolein-induced neurotoxicity in HT22 mouse hippocampal cells. *Pharmacol. Reports* **2018**.
- 569 17. Yuan, J.; Liu, W.; Zhu, H.; Zhang, X.; Feng, Y.; Chen, Y.; Feng, H.; Lin, J. Curcumin attenuates blood-
570 brain barrier disruption after subarachnoid hemorrhage in mice. *J. Surg. Res.* **2017**, *207*, 85–91.
- 571 18. Mary, C.P.V.; Vijayakumar, S.; Shankar, R. Metal chelating ability and antioxidant properties of
572 Curcumin-metal complexes – A DFT approach. *J. Mol. Graph. Model.* **2017**, *79*, 1–14.
- 573 19. JO, O. Healing and Prophylactic Effects of Moringa oleifera Leaf Extract on Lead Induced Damage to

- 574 Haematological and Bone Marrow Elements in Adult Wistar Rat Models. *J. Aquac. Res. Dev.* **2013**, *01*, 1–
575 5.
- 576 20. Ekanem, A.U.; Kwari, H.D.; Garba, S.H.; Salami, H.A. Effect of Lead Acetate on Spleen and Blood
577 Parameters in Albino Rats. *IOSR J. Dent. Med. Sci. Ver. I* **2015**, *14*, 2279–861.
- 578 21. Zhang, Y.; Fang, M.; Sun, Y.; Zhang, T.; Shi, N.; Li, J.; Jin, L.; Liu, K.; Fu, J. Curcumin attenuates cerebral
579 ischemia injury in Sprague–Dawley rats and PC12 cells by suppressing overactivated autophagy. *J.*
580 *Photochem. Photobiol. B Biol.* **2018**, *184*, 1–6.
- 581 22. Deacon, R.M.J. Measuring Motor Coordination in Mice. *J. Vis. Exp.* **2013**, 1–8.
- 582 23. Rattanachongkiat, S.; Millward, G.E.; Foulkes, M.E. Determination of arsenic species in fish, crustacean
583 and sediment samples from Thailand using high performance liquid chromatography (HPLC) coupled
584 with inductively coupled plasma mass spectrometry (ICP-MS). **2004**.
- 585 24. Simsek, N.; Akinci, L.; Alan, H.; Gecör, O.; Özcan, Ü. Determination of trace elements in kidneys, livers
586 and brains of rats with sealer implants by ICP-MS. *Biotechnol. Biotechnol. Equip.* **2017**, *31*, 397–402.
- 587 25. Costa, P.M. *Staining Protocols*; 2018; ISBN 9780128120323.
- 588 26. Mohammed Raouf, G.A.; Vaibhav, K.; Khan, A.; Tabassum, R.; Ahmed, M.E.; Javed, H.; Chander, K.;
589 Islam, F.; Siddiqui, M.S. Terminalia arjuna bark extract inhibits histological alterations by mitigating
590 oxidative stress in lead intoxicated mice. *Orient. Pharm. Exp. Med.* **2013**, *13*, 253–265.
- 591 27. Aldahmash, B.A.; El-Nagar, D.M. Antioxidant effects of captopril against lead acetate-induced hepatic
592 and splenic tissue toxicity in Swiss albino mice. *Saudi J. Biol. Sci.* **2016**, *23*, 667–673.
- 593 28. Sanders, T.; Liu, Y.; Buchner, V.; Tchounwou, P.B. Neurotoxic Effects and Biomarkers of Lead Exposure:
594 A Review. *Rev. Environ. Health* **2009**, *24*, 15–45.
- 595 29. Brochin, R.; Leone, S.; Phillips, D.; Shepard, N.; Zisa, D.; Angerio, A. The Cellular Effect of Lead
596 Poisoning and Its Clinical Picture. *Georg. Undergrad. J. Heal. Sci.* **2008**, *5*, 1–8.
- 597 30. Garza, A.; Vega, R.; Soto, E. Cellular mechanisms of lead neurotoxicity. *Med Sci Monit* **2006**, *12*, RA57-
598 A65.
- 599 31. Offor, S.J.; Mbagwu, H.O.C.; Orisakwe, O.E. Lead induced hepato-renal damage in male albino rats and
600 effects of activated charcoal. *Front. Pharmacol.* **2017**, *8*, 1–10.
- 601 32. Chiroma, S.; Baharuldin, M.; Mat Taib, C.; Amom, Z.; Jagadeesan, S.; Ilham Adenan, M.; Mahdi, O.;
602 Moklas, M. Protective Effects of Centella asiatica on Cognitive Deficits Induced by D-gal/AlCl₃ via
603 Inhibition of Oxidative Stress and Attenuation of Acetylcholinesterase Level. *Toxics* **2019**, *7*, 19.
- 604 33. Llana, D. Progesterone and estrogen influence impulsive burying and avoidant freezing behavior of
605 OVX rats. *J. Pharmacol. Exp. Ther.* **2009**, *93*, 337–342.
- 606 34. Frye, C.A.; Llana, D.C.; Walf, A.A. Progesterone can enhance consolidation and/or performance in
607 spatial, object and working memory tasks in Long-Evans rats. *Anim. Behav.* **2009**, *78*, 279–286.
- 608 35. Adolph, K.E.; Franchak, J.M. The development of motor behavior. *Wiley Interdiscip. Rev. Cogn. Sci.* **2017**,
609 *8*, 1–30.
- 610 36. Manto, M. *Toxic agents causing cerebellar ataxias*; 1st ed.; Elsevier B.V., 2011; Vol. 103;.
- 611 37. El-Eraky El-Azab, N.; M. El-Mahalaway, A.; Sabry, D. Effect of Methyl Mercury on the Cerebellar Cortex
612 of Rats and the Possible Neuroprotective Role of Mesenchymal Stem Cells Conditioned Medium.
613 Histological and Immunohistochemical Study. *J. Gastrointest. Cancer Stromal Tumors* **2018**, *08*.
- 614 38. Nehru, B.; Sidhu, P. Neurotoxic effects of differential doses of lead on rat brain followed by recovery. *J.*
615 *Trace Elem. Exp. Med.* **2002**, *15*, 131–140.
- 616 39. Barkur, R.R.; Bairy, L.K. Histological study on hippocampus, amygdala and cerebellum following low

- 617 lead exposure during prenatal and postnatal brain development in rats. *Toxicol. Ind. Health* **2014**, *32*,
618 1052–1063.
- 619 40. Sabbar, M.; Delaville, C.; De Deurwaerdère, P.; Lakhdar-Ghazal, N.; Benazzouz, A. Lead-induced
620 atypical Parkinsonism in rats: Behavioral, electrophysiological, and neurochemical evidence for a role
621 of noradrenaline depletion. *Front. Neurosci.* **2018**, *12*, 1–13.
- 622 41. Moore, T.L.; Bowley, B.G.E.; Shultz, P.L.; Calderazzo, S.M.; Shobin, E.J.; Uprety, A.R.; Rosene, D.L.;
623 Moss, M.B. Oral curcumin supplementation improves fine motor function in the middle-aged rhesus
624 monkey. *Somatosens. Mot. Res.* **2018**, *35*, 1–10.
- 625 42. Chongtham, A.; Agrawal, N. Curcumin modulates cell death and is protective in Huntington's disease
626 model. *Sci. Rep.* **2016**, *6*, 1–10.
- 627 43. Santos-Parker, J.R.; Lubieniecki, K.L.; Rossman, M.J.; Van Ark, H.J.; Bassett, C.J.; Strahler, T.R.; Chonchol,
628 M.B.; Justice, J.N.; Seals, D.R. Curcumin supplementation and motor-cognitive function in healthy
629 middle-aged and older adults. *Nutr. Heal. Aging* **2018**, *4*, 323–333.
- 630 44. Breitenbach, M. Oxidative stress and neurodegeneration: the yeast model system. *Front. Biosci.* **2013**, *18*,
631 1174.
- 632 45. Patrick, L. Lead Toxicity , A Review of the Literature . Part L ' Exposure , Evaluation , and Treatment.
633 *Altern. Med. Rev.* **2006**, *11*.
- 634 46. Swomley, A.M.; Butterfield, D.A. Oxidative stress in Alzheimer disease and mild cognitive impairment:
635 evidence from human data provided by redox proteomics. *Arch. Toxicol.* **2015**, *89*, 1669–1680.
- 636 47. Ali, B.H.; Al-Salam, S.; Al Suleimani, Y.; Al Kalbani, J.; Al Bahlani, S.; Ashique, M.; Manoj, P.; Al Dhahli,
637 B.; Al Abri, N.; Naser, H.T.; et al. Curcumin Ameliorates Kidney Function and Oxidative Stress in
638 Experimental Chronic Kidney Disease. *Basic Clin. Pharmacol. Toxicol.* **2018**, *122*, 65–73.
- 639 48. Alisi, I.O.; Uzairu, A.; Abechi, S.E.; Idris, S.O. Evaluation of the antioxidant properties of curcumin
640 derivatives by genetic function algorithm. *J. Adv. Res.* **2018**, *12*, 47–54.
- 641 49. Priyadarsini, K.I.; Maity, D.K.; Naik, G.H.; Kumar, M.S.; Unnikrishnan, M.K.; Satav, J.G.; Mohan, H. Role
642 of phenolic O-H and methylene hydrogen on the free radical reactions and antioxidant activity of
643 curcumin. *Free Radic. Biol. Med.* **2003**, *35*, 475–484.
- 644 50. Ammerman, J.; Huang, C.; Sailstad, J.; Wieling, J.; Whitmire, M.L.; Wright, D.; De Lisio, P.; Keenan, F.;
645 Mccurdy, E.; Woods, B.; et al. Technical aspects of inductively coupled plasma bioanalysis techniques.
646 *Bioanalysis* **2013**, *5*, 1831–1841.
- 647 51. Renner, R. Exposure on tap: drinking water as an overlooked source of lead. *Environ. Health Perspect.*
648 **2010**, *118*.
- 649 52. Sousa, R.A. De; Sabarense, C.M.; Prado, G.L.P.; Metze, K.; Cadore, S. Lead biomonitoring in different
650 organs of lead intoxicated rats employing GF AAS and different sample preparations. *Talanta* **2013**, *104*,
651 90–96.
- 652 53. Shen, L.; Zhang, H.Y.; Ji, H.F. A theoretical study on Cu(II)-chelating properties of curcumin and its
653 implications for curcumin as a multipotent agent to combat Alzheimer's disease. *J. Mol. Struct.*
654 *THEOCHEM* **2005**, *757*, 199–202.
- 655 54. Sidhu, P.; Nehru, B. Lead Intoxication: Histological and Oxidative Damage in Rat Cerebrum and
656 Cerebellum. *J. Trace Elem. Exp. Med.* **2004**, *17*, 45–53.
- 657 55. Nam, S.M.; Seo, J.S.; Nahm, S.S.; Chang, B.J. Effects of ascorbic acid on osteopontin expression and axonal
658 myelination in the developing cerebellum of lead-exposed rat pups. *Int. J. Environ. Res. Public Health*
659 **2019**, *16*.

- 660 56. Wang, R.; Li, Y.H.; Xu, Y.; Li, Y.B.; Wu, H.L.; Guo, H.; Zhang, J.Z.; Zhang, J.J.; Pan, X.Y.; Li, X.J. Curcumin
661 produces neuroprotective effects via activating brain-derived neurotrophic factor/TrkB-dependent
662 MAPK and PI-3K cascades in rodent cortical neurons. *Prog. Neuro-Psychopharmacology Biol. Psychiatry*
663 **2010**, *34*, 147–153.
- 664 57. Samarghandian, S.; Azimi-nezhad, M.; Farkhondeh, T. ScienceDirect Anti-oxidative effects of curcumin
665 on immobilization-induced oxidative stress in rat brain , liver and kidney. *Biomed. Pharmacother.* **2017**,
666 *87*, 223–229.
- 667 58. Wani, A.L.; Ara, A.; Usmani, J.A. Lead toxicity: A review. *Interdiscip. Toxicol.* **2015**, *8*.
- 668 59. Mushak, P. *Lead exposure in human populations: Lead toxicokinetics and biomarkers of lead exposure.*; 2011; Vol.
669 10; ISBN 9780444515544.
- 670 60. Barbosa, F.; Tanus-Santos, J.E.; Gerlach, R.F.; Parsons, P.J. A Critical Review of Biomarkers Used for
671 Monitoring Human Exposure to Lead: Advantages, Limitations, and Future Needs. *Environ. Health*
672 *Perspect.* **2005**, *113*, 1669–1674.
- 673 61. Omobowale, T.O.; Oyagbemi, A.A.; Akinrinde, A.S.; Saba, A.B.; Daramola, O.T.; Ogunpolu, B.S.;
674 Olopade, J.O. Failure of recovery from lead induced hepatotoxicity and disruption of erythrocyte
675 antioxidant defence system in Wistar rats. *Environ. Toxicol. Pharmacol.* **2014**, *37*, 1202–1211.
- 676 62. Khalaf, A.A.; Moselhy, W.A.; Abdel-Hamed, M.I. The protective effect of green tea extract on lead
677 induced oxidative and DNA damage on rat brain. *Neurotoxicology* **2012**, *33*, 280–289.
678