

1 Supplemental Material

2 **Differential role of TGF- β in extracellular matrix**
3 **regulation during *Trypanosoma cruzi* - host cell**
4 **interaction**

5

6 **Tatiana Araújo Silva, Luis Felipe de Carvalho Ferreira, Mirian Claudia de Souza Pereira and**
7 **Claudia Magalhães Calvet*.**

8

9 Cellular Ultrastructure Laboratory, Oswaldo Cruz Institute, FIOCRUZ, Rio de Janeiro, RJ, 21040-360, Brazil

10

11 *Corresponding author: Dr Claudia M. Calvet, Laboratório de Ultraestrutura Celular, Instituto Oswaldo
12 Cruz, FIOCRUZ, Av. Brasil 4365, Manguinhos, 21040-362 Rio de Janeiro, RJ, Brazil, e-mail:
13 cmcalvet@ioc.fiocruz.br

14

15

16

17

18

19

20

21

22

23

24

25

26

27

28

29

30

31

32

33

34

35

36

37

38

39

40

41

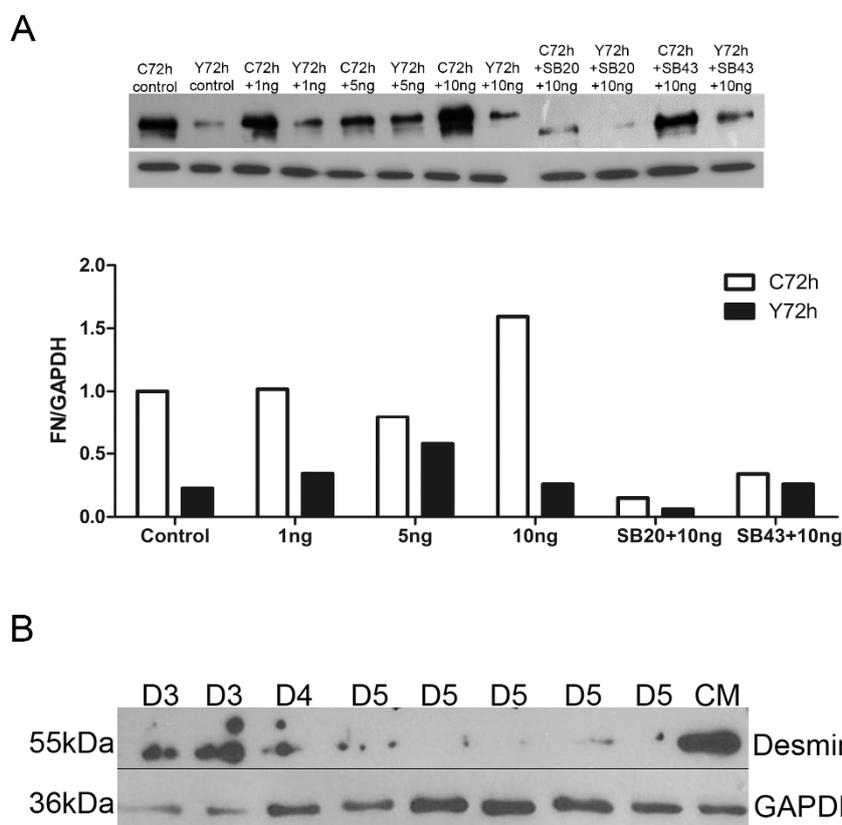
42

43

44 **Characterization of cardiac fibroblasts.**

45 Cardiac fibroblasts have the largest cell nucleus when compared to
46 cardiomyocyte (CM), this morphology being a distinctive feature in this cell
47 type. Initially, CF purification obtained from CM cultures was evaluated only by
48 morphology. CF obtained in the third dissociation of CM cultures were infected
49 with *T. cruzi* and treated with TGF- β for 48 hours and were subjected to Western
50 blot to analyze the FN expression under these conditions. Our data revealed an
51 increase in FN expression only with 10 ng/ml TGF- β . Furthermore, third
52 dissociation from CM cultures had reduced FN expression compared to normal
53 control even after the addition of TGF- β . The addition of inhibitors SB431542 and
54 SB203580 (SMADS pathway and p38 MAPK inhibitor respectively) in this culture
55 led to inhibition of FN stimulation by TGF- β , since in infected cultures treated
56 with 10 ng/ml TGF- β (48h) reduced FN expression after pre-treatment with
57 inhibitors signaling (S1A) was seen. The data obtained in the third dissociation
58 from CM culture extracts with the FN expression correspond with that of
59 cardiomyocytes [1], suggesting that CF purification was needed adjustments and
60 that FN response could not be displayed properly when CF is present in CM
61 culture, suggesting that morphological analysis under the microscope do not
62 guarantee good purification of CF.

63 Thus, to assess the degree of CF purification from CM cultures, desmin, a
64 specific cytoskeletal protein of cardiomyocytes, was evaluated in CF extracts
65 after different dissociations. CF extracts obtained in the third and fourth
66 dissociation revealed that cardiomyocytes are still present in CF culture, since the
67 extracts also had high expression of desmin, demonstrating that these
68 dissociations are not viable for conducting experimental tests. Pure CF was
69 obtained only from the fifth dissociation from CM culture in the absence of
70 cardiomyocytes, with desmin undetectable in this culture only after the 5th
71 passage (S1B).



72 **S1 - FN expression in cells obtained in the third CM culture passage. (A)**
 73 **Control cultures show FN stimulation only with the treatment with 10 ng/ml**
 74 **TGF- β (48h).** Doses of 1ng/ml and 5ng/ml did not induce increase of FN
 75 expression. *T. cruzi* infection leads to a reduction in FN expression after addition
 76 of TGF- β . Inhibitory action of SMADs and p38 MAPK pathway prevents increase
 77 in FN expression even after stimulation with 10 ng/ml. These results obtained
 78 with cell extracts after third CM culture passage are similar to previously
 79 published data in cardiomyocyte cultures under the same conditions. **(B)**
 80 **Characterization of the culture of heart fibroblasts.** Rate of CF purification from
 81 cardiac culture. The degree of CF purification was evaluated by detection of
 82 desmin in cell extracts obtained in the 3rd, 4th and 5th dissociation. CF
 83 purification was obtained from the fifth dissociation, as shown by the absence of
 84 desmin.

86

87 **References**

- 88 1. Calvet, C.M.; Oliveira, F.O.R.; Araújo-Jorge, T.C.; Pereira, M.C.S.
 89 Regulation of extracellular matrix expression and distribution in
 90 *Trypanosoma cruzi*-infected cardiomyocytes. *Int. J. Med. Microbiol.* **2009**,
 91 299, 301–12.

92

93