

Viral strategies of manipulating autophagy to benefit infection

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Abstract

Autophagy is an evolutionarily conserved central process in host metabolism. Among its major functions are conservation of energy during starvation, recycling organelles, and turnover of long-lived proteins. Besides, autophagy plays a critical role in removing intracellular pathogens and very likely represents a primordial intrinsic cellular defence mechanism. More recent findings indicate that it has not only retained its ability to degrade intracellular pathogens, but also functions to augment and fine tune antiviral immune responses. Interestingly, viruses have also co-evolved strategies to manipulate this pathway and use it to their advantage. Particularly intriguing is infection-dependent activation of autophagy with positive stranded (+)RNA virus infections, which benefit from the pathway without succumbing to lysosomal degradation. In this review we summarise recent data on viral manipulation of autophagy, with a particular emphasis on +RNA viruses and highlight key unanswered questions in the field that we believe merit further attention.

1. Introduction

RNA viruses have evolutionarily constrained genome sizes. At the same time they have co-evolved efficient means to manipulate host cellular processes to acquire nutrients while evading immune detection. Multifunctional viral proteins, molecular mimicry of host components, and the intrinsically high mutagenicity of their RNA genome converge to dysregulate host cellular pathways, and exploit metabolic processes to their advantage. One such target is the autophagy machinery. While this cellular degradative process has been historically described to restrict growth of many intracellular pathogens including bacteria, parasites and viruses, many have evolved mechanisms to circumvent and even actively benefit from it.

Autophagy is initiated by sequestration of cytoplasmic proteins and damaged organelles into crescent-shaped double-membrane vesicles known as isolation membranes, long-debated on their membrane source [1]. Current consensus on the source of autophagosomal membranes is the endoplasmic reticulum [2]. Once contents are captured, the immature isolation membranes expand to forming autophagosomes, which subsequently fuse with lysosomes, thus forming autolysosomes. The contents undergo degradation within the autolysosomes to enable recycling during starvation. About 30 genes have been reported to participate in the process of autophagosomal degradation.

Apart from turnover of organelles and primarily long-lived proteins, autophagy is a powerful tool that operates to defend host cells against intracellular pathogens, delivering trapped bacterial or viral products to lysosomes for degradation. Besides, it is equipped to clear invasive pathogens through induction of CD8⁺ T-cell responses, and also initiates a primordial innate immune response by cooperating with pattern recognition receptor signalling to induce interferon production. This was recently described in an elegant study in the context of DNA virus infection, where cyclic GMP-AMP (cGAMP) and STING-dependent activation of autophagy was necessary to remove viral DNA from the cytosol [3]. However, in an ongoing evolutionary arms race, most pathogens have acquired the ability to hijack and subvert autophagy to evade degradation through this pathway. More remarkably, +RNA viruses have adapted to not only protect themselves from autophagic elimination but even harness the machinery to their own benefit, as will be covered in more detail in the subsequent sections.

2. Subversion of autophagy by +RNA viruses

Among the +RNA viruses, data on favourable versus detrimental impact of autophagy is particularly confounding. A link between autophagosomes and virus-induced vesicles was proposed by George Palade by EM imaging of poliovirus containing vesicles that resembled autophagosomal membranes [4]. Over the past few decades, a growing body of research has defined the critical role of this pathway in facilitating infection by numerous +RNA viruses, including poliovirus (PV) [5,6], Coxsackievirus B3 (CVB3) [7,8], CVB4 [9], Enterovirus 71 (EV71) [10], Human rhinovirus (HRV) [11], Foot-and-mouth disease virus (FMDV) [12], encephalomyocarditis virus (EMCV) [13], Dengue virus (DENV) [14,15], Zika virus (ZIKV) [16,17], Hepatitis C virus (HCV) [18], Mouse hepatitis virus (MHV), Newcastle disease virus (NDV)[19], Severe and acute respiratory syndrome coronavirus (SARS-CoV) [20], Chikungunya virus (ChikV) [21], and Japanese encephalitis virus (JEV) [22]. In many of the above cases, pharmacological or genetic manipulation of autophagy *in vitro* confirmed an inhibition in replication and/or spread of these viruses, whereas induction of autophagy resulted in increased production of progeny virions [19,23].

Current evidence indicates that many, if not all +RNA viruses depend on the initiation of the autophagic pathway for their optimal production. This is counterintuitive, since these viruses replicate in the cytosol, and autophagy serves to promote degradation of cytosolic contents. Therefore, it is evident that +RNA viruses in particular have evolved sophisticated mechanisms to circumvent or exploit this pathway. Not surprisingly, most +RNA viruses also trigger massive membrane remodelling within infected host cells to create membrane delineated structures, often referred to as replication organelles, vesicle packets, convoluted membranes or double membrane vesicles, depending on their morphology and ultrastructure [16,24]. Whether these replication organelles are pseudo-autophagosomes, has long been a point of contention.

Several genome-wide screens, e.g with CRISPR/Cas9 libraries, haploid KBM7 cells, and shRNA depletions, as well as proteomic studies have universally indicated the involvement of the autophagy pathway in +RNA virus infections [23,25,26]. However, among the 30 odd autophagy-related genes, the functional contribution of the individual components in virus infection is far from clear. A recent targeted CRISPR/Cas9 screen uncovered a fairly diverse range of involvement among autophagic components in three +RNA virus infections – PV, DENV and ZIKV [27]. The authors reported that all three viruses employed multiple proteins of the autophagy pathway while bypassing others, and

each virus used a unique set of initiation components. A common feature among the tested viruses underscored the requirement of the LC3 protein but not its canonical cellular lipidation process, where LC3 was recruited to virally induced membranes by alternative means. This study highlights the importance of assessing the pathway in its entirety when seeking to understand how pathogens co-opt it for purposes of genome replication and spread, as well as to identify universal drug targets. Many different mechanisms have been proposed on how autophagy is manipulated to facilitate infection while preventing degradation of +RNA viruses (**Figure 1**). While in no way exhaustive, the following sections cover the salient features that are recurrent among several viral genera:

2.1 Partial/selective autophagy

The physical hallmark of the autophagy pathway is the formation of LC3+ cytosolic double-membrane vesicles, also often observed in +RNA virus infections. One of the long-running debates is whether these replication organelles are themselves immature autophagosomes or take advantage of the same machinery for their biogenesis. However, canonical autophagic vesicles are part of a degradative pathway, where they fuse with lysosomes for their contents to be hydrolysed by proteases and lipases. Viruses from different families appear to possess a diverse set of strategies to prevent this from happening. Flaviviruses, such as DENV and ZIKV have been reported to trigger autophagy on the one hand, while utilising the ER as a focal point for generating their replication organelles and assembly of progeny virions. Consequently, both viruses have evolved means to suppress ER-turnover via reticulophagy. The ER-localised reticulophagy receptor FAM134B was identified as a restriction factor for both DENV and ZIKV. RNAi-depletion of FAM134B significantly enhanced DENV and ZIKV replication at an early stage of the viral life cycle. The virus-encoded NS3 protease from several flaviviruses directly cleaved FAM134B at a single site within its reticulon homology domain to selectively suppress ER degradation [28], underscoring a sophisticated mechanism to differentially regulate specific arms of autophagy.

Coxsackievirus B3 (CVB3), an enterovirus belonging to the *Picornaviridae* family, is known to rely on autophagosome formation for optimal replication [29,30]; however, both *in vitro* and *in vivo* evidence suggest that during infection amphisome maturation and autophagic protein degradation are inhibited. An increase in autophagosomal abundance concomitant with a decrease in autophagic flux was reported with CVB3, prompting the hypothesis that infection selectively triggers autophagosome formation while preventing

the terminal stages in degradation [5,31]. The molecular determinants and mechanism by which CVB3 limits autophagic degradation is currently unknown. Interestingly, treatment of CVB3-infected cells with inhibitors of autophagosome maturation resulted in increased virus production, indicating that canonical autophagy was not completely blocked in virus-infected cells, and at least a population of the virus remained sensitive. A similar finding was reported more recently with rotavirus where virus replication benefited from induction of autophagy while blocking degradation [32]. As with CVB3, the mechanism by which rotaviruses specifically inhibit autolysosomal degradation has not been elucidated, emphasising the importance of identifying the specific virus or host components that prevent degradation to provide fundamental insights on autophagic regulation in general. The case with HCV infection is more convoluted on account of contradictory data: whereas GFP-RFP-LC3 expressing cells infected with HCV displayed a complete maturation of autophagosomes followed by fusion with lysosomes [33,34], at least one other study reported that HCV replication restricted autophagosomal fusion [35]. Yet another study reported that the autophagy pathway in its entirety was necessary during HCV-infection for optimal replication; however, the advantage derived from it was primarily due to suppression of innate immune responses [34]. Blocking fusion between autophagosomes and lysosomes has also been reported to increase DENV2 yield [36]. However, this effect may be viral serotype-specific, since inhibiting lysosome fusion reduced DENV3 production [37]. The mechanisms by which autophagy favors DENV production was recently described where rather than replication, assembly and release of progeny virions was affected by blocking autophagy mediated lipid droplet hydrolysis [14,38,39].

With Coronaviruses, initiation of autophagy appears to be through the ER-derived, PtdIns3P-enriched omegasomes that normally operate during starvation. Infectious bronchitis virus (IBV) - an avian coronavirus responsible for major losses to the poultry industry, is one such example where a significant portion of the genome encodes nonstructural proteins (NSP) dedicated to virus replication. Expression of NSP6 proteins resulted in increased levels of PtdIns3P on ER membranes, recruitment of PtdIns3P effector protein WIPI2 and the generation of autophagosomes directly from the ER [40,41]. However, when compared to the properties of starvation-induced autophagosomes, those generated by Coronavirus infection or NSP6 proteins presented significant differences. NSP6-induced autophagosomes displayed limited ability to undergo expansion, preventing formation of large autolysosomes, and hence circumvented degradation of viral particles through the lysosomal pathway [41]. Results obtained with IBV, SARS and MHV

nsp6 was recapitulated with Middle eastern respiratory syndrome coronavirus (MERS-CoV), where a similar phenomenon was observed for nsp6 [42].

2.2 Autophagosomes as viral replication sites

A distinctive feature shared by +RNA viruses is to assemble and replicate on intracellular membranes, which have been proposed to offer a two-fold advantage: (a) scaffold for anchoring and concentrating the replication complexes and (b) to insulate dsRNA intermediates from innate sensing by cytosolic pattern recognition receptors [43]. The replication complexes are typically composed of the viral RNA-dependent RNA polymerase, accessory non-structural proteins, viral RNA, and host factors. Specialised autophagosomes, sometimes referred to as amphisomes (formed upon fusion with endosomes), omegasomes, and EDEMosomes (both ER-derived) have been hypothesised to function as replication organelles [41,44,45]. Given the resemblance of virus-triggered double membrane vesicles with that of autophagosomes, it is plausible that generation of replication organelles are a result of mechanisms similar to autophagy [46-49]. Like autophagosomal membranes, many virus-induced vesicles are believed to be ER-derived. Recent data from several different studies have provided direct evidence of the association between viral replication complexes and autophagosome structures as summarized in Table 1. Poliovirus (PV) vesicle clusters were found to contain LC3 and lysosomal markers, reminiscent of autolysosomes, and colocalised with the PV replication complex [5,50]. Furthermore, formation of infectious PV progeny virions was reported to depend on vesicular acidification, prompting the hypothesis that particle assembly, genome replication and virion maturation occurred in bona fide autophagosomal vesicles [44].

Among the *Flaviviridae* family, several non-structural proteins have been observed in LC3+ vesicles. DENV non-structural protein NS1 and dsRNA were reported to co-localise with LC3 and ribosomal proteins [36,37]. Another study described the induction of LC3+ vesicles, which colocalised with NS4A in infected cells, or when transfected with a combination of DENV NS4A and NS4B [14]. This was independently corroborated by ZIKV infection in human fetal neural stem cells where expression of NS4A and NS4B were sufficient to block neurogenesis and promote autophagy, displaying partial colocalisation with LC3+ vesicles [51]. Ultrastructural analysis of ChikV virions also suggested their location in the lumen of autophagosome-like vacuoles [21].

Similar to other members of this family, HCV infection induces massive intracellular membrane rearrangements. Competing hypotheses have been proposed as to whether autophagosomes themselves serve as sites for HCV replication. By sucrose gradient analysis, LC3-II was found to co-sediment with HCV RNA and non-structural proteins NS3 and NS5A [52]. However, in a separate study confocal microscopy showed little evidence of co-localisation of LC3 or Atg5 with HCV proteins [53,54]. Along the same lines, depletion of either LAMP2 or Rab7, which allowed accumulation of autophagosomes by preventing fusion with lysosomes inhibited HCV viral replication, also suggesting that they are not the major sites for HCV genome replication [34], while multiple reports indicate that lipid droplets are the more likely sites for replication and assembly, as reviewed elsewhere [38,55].

Conflicting evidence also exists for MHV-induced replication compartments. On the one hand, MHV replication complexes were found to be associated with LC3 and Atg12 in embryonic stem cell lines [20]. On the other hand in primary macrophages and murine embryonic fibroblasts MHV replication did not require the autophagy gene Atg5 [56]. Differences in permissiveness to infection often exists between primary cells and transformed cell lines, as does viral tropism towards distinct cell types, either of which can account for these experimental discrepancies. Among other +RNA viruses, immunoelectron microscopy demonstrated co-localisation of EV71 capsid protein VP1 with autophagosomes in virus-infected mouse neurons [57]. Similarly, during EMCV infection, colocalisation of non-structural protein 3A and capsid protein VP1 was visualised by confocal and immunoelectron microscopy [58]. Colocalisation of non-structural proteins 2B, 2C, and 3A with LC3, and structural protein VP1 with Atg5 were also reported in FMDV-infected cells [12]. Although direct evidence of the association of viral replication complexes with autophagosomes is lacking for CVB3, impaired maturation of autolysosomes brought about through pharmacological or genetic inhibition increased the accumulation of autophagosomes in virus-infected cells resulting in enhanced viral replication [29,30]. These data implicated autophagosomes as virus anchoring and replication sites during CVB3 replication.

2.3 Autophagy as a mechanism to facilitate virus assembly

Delineating the process of viral assembly from replication is technically challenging, especially since both processes would very likely occur in concert at the same sites. Although utilisation of autophagosomes as assembly sites has been recorded for some

DNA-viruses e.g., Hepatitis B virus (HBV) [59], experimental data with +RNA viruses are scant. However, at least for DENV, recent evidence indicates that autophagy might actually assist in assembly of progeny virions, without them serving as replication organelles. One of the initial studies describing the induction of autophagy in flavivirus infections was performed by Lee et al [60]. The authors demonstrated that DENV2 infection in hepatocytes induced autophagy; targeting with either the inhibitor 3-methyladenine (3MA) or siRNAs against autophagy genes compromised infection. DENV-induced autophagosomes colocalised with LAMP1, a marker of lysosomal fusion, which was independently validated by immunofluorescence assays and pharmacological inhibition. Following this initial characterisation, a more mechanistic study emerged describing the role of selective autophagy facilitating hydrolysis of lipid droplets in an infection-specific manner [15].

Apart from the relatively non-specific bulk macroautophagy, cellular organelles are turned over through several types of selective autophagy. This phenomenon occurs under normal physiological conditions and is hypothesised to initiate a physiological response to appropriately address a specific stress. In the context of DENV infection, a type of selective autophagy of lipid storage organelles (lipid droplets) referred to as lipophagy was described that hydrolyses neutral fat deposits to free fatty acids and cholesterol, and supplements cellular energy reservoirs [61,62]. Heaton et al performed a targeted siRNA screen to identify cellular cofactors of DENV2 replication in hepatocytes, which revealed, among others, genes involved in the induction of autophagy [63], and were further characterised to reveal that DENV induced autophagosomes not only acquired LAMP1, but underwent complete maturation to become autolysosomes [15]. These did not colocalise with markers of the viral replication complex, suggesting that they may play an indirect, non-structural role in DENV replication. A subsequent study described the involvement of AUP1, a type-III membrane protein, in the initiation of virus-induced autophagy. AUP1 was regulated by monoubiquitin modification, where infection or a combined expression of viral NS4A and NS4B were necessary and sufficient to generate the unmodified form of AUP1 – a step that was critical in induction of this pathway. Interestingly, loss of AUP1 did not affect viral replication; however, impaired autophagy was accompanied by degradation of viral proteins through the proteasomal pathway resulting in significantly reduced production of progeny virions, supporting a specific role of AUP1-dependent lipophagy in assembly of virus particles [14]. These results were in

agreement with an independent report demonstrating expression of Dengue NS4A was sufficient to trigger autophagy and protect against cell death.

2.4 Non-lytic viral transmission by secretory autophagy

Several reports on mechanisms of secretion and cell-to-cell transfer of intracellular pathogens indicate non-degradative autophagic vesicles as an efficient mode of transport. An interesting recent study with *Mycobacterium* demonstrated that autophagosomes chaperone an organelle referred to as the “ejectosome”, facilitating cell-to-cell spread of cytosolic bacteria [64]. Secretory autophagy is a newly discovered pathway in which autophagosomes fuse with the plasma membrane instead of lysosomes and release single membrane vesicles containing cytosolic content into the extracellular milieu [65].

Non-degradative autophagy has been suggested to facilitate non-lytic egress of some +RNA viruses. The initial characterisation was with enteroviruses, which appear to exploit this pathway to exit cells, and are released into the extracellular environment as particle populations contained within vesicles [66]. Clusters of enteroviral particles were packaged with phosphatidyl serine into autophagic vesicles, which enabled efficient transfer to primary macrophages, significantly enhancing viral infectivity. This revealed a novel mode of transport where viral genomes were transferred *en bloc* to recipient cells, facilitating genetic cooperativity and enhancing infection. This mode of transfer had previously also been noted for CVB3, where a recombinant fluorescent virus was released into the extracellular medium in microvesicles containing autophagic markers [67].

Poliovirus is often considered a lytic virus; however, non-lytic release of poliovirus has also been reported [6]. Reduced levels of viral particles in extracellular medium in autophagy-deficient cells correlated with inhibition of non-lytic release of autophagic vesicles. More recently, an important role of the secretory autophagy pathway was implicated in ZIKV vertical transmission as well as cell-to-cell spread of DENV. ZIKV-induced autophagic activity in human trophoblasts restricted by pharmacological inhibition, or by deficiency in an essential autophagy gene, Atg16l1, limited ZIKV vertical transmission and improved placental and fetal outcomes, which supported a role for autophagic secretion in the process [68]. Along the same lines, it was hypothesised that DENV might evade neutralising antibodies and increase viral spread by exploiting autophagic vesicles for delivery to the extracellular medium [69]. Double staining of DENV E antigen and LC3 in a close-contact co-culture experimental set-up verified secretion of DENV-containing autophagic vesicles from donor cells, which were subsequently taken

up by recipient cells. In a parallel study, maturation of infectious DENV virus particles was attributed to this process, when cleavage of Pr peptide from PrM by the furin protease was prevented upon blocking autophagy [70]. Further investigation is needed to provide more direct evidence on the mechanism, regulation and molecular determinants of secretory autophagy in facilitating viral release.

2.5 Suppression of innate antiviral responses through activation of autophagy

Activation of autophagy represents a fairly ubiquitous response to eliminate intracellular pathogens. Several studies have described mechanisms where pathogen recognition receptors trigger this response upon detection of microbe-specific pathogen associated molecular patterns. A diverse set of pathogens including bacteria, viruses and parasites have provided corroborating evidence supporting this phenomenon. Whether this process occurs in parallel to non-degradative autophagy, or the cross-talk that might exist between the two flavours of autophagy during +RNA viral infection merits further investigation.

Recent studies have shed light on how autophagy offers an advantage to HCV infection by suppressing innate immune responses [34,71]. This contrasts with previous data on HCV-induced incomplete autophagy and defined a pathway where the entire process from initiation through lysosomal degradation is necessary for HCV replication largely to suppress anti-viral innate immune response. In HCV-infected cells, interferon- β (IFN- β) production could be modulated by UPR-mediated autophagy; activation of this pathway reduced IFN- β production and vice-versa [72]. Inhibition of autophagy by suppressing Beclin-1 or Atg7 reduced HCV replication, which was accompanied by the activation of IFN signaling. A similar phenomenon was also proposed for DENV where activation of autophagy not only facilitated viral replication, but also suppressed IFN-I production, suggesting that both viruses may share the same mechanism to evade innate immune responses. Interestingly, in West Nile virus infections, perturbation of intracellular cholesterol levels dictated IFN-I responses [73]. Although the role of autophagy in WNV infection has been contested, replication was reported to occur independent of autophagy by multiple groups – a clear difference from other flaviviruses; however, whether it plays a role in regulating free cholesterol and fatty acid levels is yet to be determined. Together, these studies indicate a critical mechanism by which flaviviruses may avoid innate immune responses through activating the host autophagy pathway.

2.6 Induction of autophagy to facilitate cell survival

The link between autophagy and apoptosis has been defined extensively in various contexts including cancer, neurodegenerative disorders, and infectious diseases [74]. A more comprehensive understanding on circumventing cell death has been recorded for bacterial rather than viral infections, amongst which most are DNA viruses. Premature cell death can function as an anti-viral host mechanism by providing an unfavorable environment and shorter timeframes for viral propagation. Induction of autophagy has often been linked to inhibition of apoptosis [75,76]. Both DENV2 and a murine flavivirus-induced autophagy was reported to prevent apoptosis mediated via the viral NS4A protein [77]. Knockdown of autophagy-related gene expression abolished the protective role of autophagy against cell death and resulted in reduced viral replication. Apart from DENV, cross-talk between autophagy and apoptosis was also reported in CVB4 infection, where suppression of autophagy by 3MA triggered caspase activation and vice-versa [78].

3. Impact of virus-triggered autophagy on antigen presentation

An interesting question that arises out of the induction of autophagy in the context of DENV/ZIKV infection is the process of antigen presentation. Monocytes and monocyte-derived cells are a major target of flaviviruses, where antigen presentation is facilitated by autophagy. This implies that while autophagy favours production of viral progeny, it should simultaneously increase viral antigen presentation and T-cell responses, thus generating neutralising antibodies and promoting cytotoxic T-cell killing. Information on these seemingly contradictory processes is currently limited. However, DENV-infected human monocyte-derived DCs fail to upregulate MHC and co-stimulatory molecules and have an impaired ability to polarize CD4⁺ Th type 1 (Th1) effector properties [79], contributing to inefficient adaptive immune responses observed in patients. In bulk cultures of dendritic cells, exposure to DENV augments MHC-I and MHC-II expression in non-infected bystander cells; however, infected monocyte-derived dendritic cells display an inhibition in this process within the same cultures [80]. In clinical studies gene expression analyses of DENV patients revealed that severe cases expressed lower levels of genes linked to antigen processing, presentation and T-cell activation compared to mild cases. Another related flavivirus, Japanese Encephalitis virus, inhibits expression of MHC-I and induces functional impairment of DCs, resulting in poor CD8⁺ T cell responses [81]. Thus impaired antigen presentation and functionality of virus-infected DCs may reflect a viral immune escape strategy to dampen T-cell responses and impact disease severity.

One study reported that DENV activated autophagy only during the early infection stage, suggesting a biphasic response of autophagy to DENV infection, where it shifted from a supporting to an antiviral role at later time points [82]. These results might enable us to reconcile how flaviviruses have evolved strategies to manipulate this pathway while subverting T-cell based immune responses. A quantitative and time resolved analyses of this process in virus-infected cells might shed light on its utilisation in the benefit versus detriment towards virus production.

4. Autophagy-mediated restriction of +RNA virus infections

Activation of autophagy in the presence of intracellular pathogens is a fairly universal cellular response. Xenophagy as an intrinsic defence mechanism was first described through electron microscopy studies upon visualising HSV-1 and cytomegalovirus inside autophagosomes [83].

Among viruses, autophagic protection has been recorded in a wide-ranging species and genera [84-86]. The mechanism of autophagy-mediated restriction of +RNA viruses is less well-documented on account of its proviral influence in most cases. However, there are instances where autophagic degradation of virions (virophagy) or viral proteins has been observed, especially in neurons where it is a critical form of antiviral defence. During Sindbis virus (SINV) infection, Beclin 1 and p62-dependent degradation of the capsid protein protects against SINV-mediated encephalitis [87,88]. Moreover, ATG5 deficiency results in delayed SINV clearance and accumulation of the autophagy receptor p62. More recently, Fanconi anaemia group C protein (FANCC) was found to interact with the SINV capsid protein and facilitate virophagy [26]. Picornaviruses, such as PV and HRV permeabilise endosomes to release their genome into the cytosol. This step is detected by galectin 8, which restricts viral infection by initiating degradation of the viral RNA genome [89]. As counterstrategy, the host protein HRAS-like suppressor 3 (PLA2G16) is exploited by the virus to enable genome delivery. CVB3, also belonging to the same family, undergoes p62-dependent degradation and uses the viral protease 2A to cleave p62 and inhibit virophagy [90]. Interestingly, although HCV has been demonstrated to induce autophagy to its own advantage by multiple groups, one study demonstrated that an ER transmembrane protein, SCOTIN, interacted with the viral protein NS5A, resulting in its autophagic degradation to suppress viral replication [91]. Among flaviviruses, autophagosomal degradation of neurotropic viruses has been recorded. In the drosophila brain, ZIKV infection triggered NF κ B-dependent

inflammatory signaling, inducing expression of dSTING which subsequently restricted infection by upregulating autophagy. Defective or absence of autophagy resulted in increased infection in the fly brain and death [92]. STING-dependent induction of protective autophagy was independently reported for DNA viruses and sea anemone, supporting an evolutionarily conserved role for STING in microbial autophagy [3].

5. Future perspectives

Despite major advances in elucidating molecular determinants of the autophagy pathway, the rules that govern its utilisation during infections are far from obvious. A complex interplay between viral manipulation and host innate immunity dictates disease outcomes. Autophagy is expected to restrict viral infections at multiple levels by eliminating viruses, regulating inflammatory responses and promoting antigen presentation. However, +RNA viruses have co-evolved to manipulate autophagy for immune evasion, replication, assembly and release from infected cells. The repertoire of universal and distinct mechanisms that these viruses draw on to interfere with autophagy are striking, often targeting the same pathway in unique ways with different functional implications. Distinct viral strategies fine-tune the process to simultaneously escape destruction while capitalising on the structural and nutrient benefits that autophagy provides. Further understanding of the contribution of autophagy to the different stages in the viral life-cycle, subversion of its antigen presentation function and innate immune responses is necessary to delineate its function in immunity against viral infections versus viral pathogenesis.

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Figure Legends

Figure 1. Mechanisms of viral subversion of autophagy

Schematic illustration of the different pathways of selective autophagy that are triggered upon +RNA virus infections. Initiation of autophagosomes is through formation of an isolation membrane most likely derived from the ER; depending on the molecular composition and function, they may form either omegasomes, EDEMosomes or amphisomes. Turnover of organelles occur through convergence of specialised autophagosomes with lysosomes for their selective degradation: ER via reticulophagy; mitochondria via mitophagy; lipid droplets via lipophagy and virions or viral proteins via virophagy. Viruses that are known to upregulation specific autophagosomal pathways are depicted in black, those that suppress specific types or steps of autophagy are depicted in red.

Table 1. Viral protein interaction with components of the host autophagy machinery

Virus	Viral protein	Interaction with autophagy machinery	Reference
Flaviviridae			
DENV	NS1	Colocalises with dsRNA, autophagic vacuoles and cathepsin D	[36,37]
	NS4A	Induces PI3K-dependent autophagy and protect cell death from stress; redistributes to LC3+ vesicles	[14,77]
WNV	NS4A, NS4B	Single amino acid substitution in either protein is sufficient to upregulate autophagy	[93]
ZIKV	NS1	Possible transport through autophagic secretion	[94]
	NS4A, NS4B	Supresses Akt-mTOR signaling to induce autophagy	[51]
JEV	NS1	Colocalises with endogenous LC3	[95]
DENV, WNV, ZIKV	NS2B, NS3	Cleaves FAM134B to interfere with reticulophagy	[28]
HCV	NS4B	Redistributes LC3 to punctuate structures	[96]
	NS5A	Degraded in autophagosomes by SCOTIN Upregulates Beclin 1 and activates mTOR signaling pathway	[91] [97]
	NS5B	Interacts with autophagy elongation complex (ATG5-12/16L1) for proper membranous web formation at replication sites	[98]
Coronaviridae			
HCoV-NL63	NSP3	PLP2 domain interacts with LC3 and Beclin 1 to induce autophagosomes while blocking lysosomal fusion	[99]
MHV	NSP2, NSP3	Colocalises but does not interact with LC3-I	[100]
IBV, SARS, MHV	NSP6	Induces Atg5 and LC3 dependent, starvation-independent autophagosomes	[41,101]
Picornaviridae			
CVB3	2A	Cleaves p62/SQSTM1 to disrupt selective autophagy and NFκB signaling	[102]
	2B	Autophagy-inducing motif at 36-83 amino acids	[103]
EV-D68	3C	Cleaves autophagosomal SNARE, SNAP29	[104]
EV71	VP1	Colocalises with LC3 to form autophagosome-like vesicles	[57]
FMDV	VP1	Colocalises with Atg5 and p62	[12]
	VP2	Induces autophagy through HSPB1 and EIF2S1-ATF4 pathway	[105]
PV	2B	Induces autophagy	[106]
	2B, 2C, 3A	Colocalises with LC3	[12]
	VP1	Colocalises with LC3, postulated to release virus through autophagosome-like vesicles	[5]
	2BC, 3A	Induces formation of LC3 coated double membrane vesicles	[107]
	3AB	Induces formation of autophagosome-like double-membrane liposomes	[108]

Table1: Summary of Interactions between proteins from positive strand RNA viruses and host autophagy machinery.

References

- [1] S.T. Shibutani, T. Yoshimori, A current perspective of autophagosome biogenesis, *Nature Publishing Group*. 24 (2014) 58–68. doi:10.1038/cr.2013.159.
- [2] D. Mijaljica, M. Prescott, R.J. Devenish, Endoplasmic reticulum and Golgi complex: Contributions to, and turnover by, autophagy, *Traffic*. 7 (2006) 1590–1595. doi:10.1111/j.1600-0854.2006.00495.x.
- [3] X. Gui, H. Yang, T. Li, X. Tan, P. Shi, M. Li, et al., Autophagy induction via STING trafficking is a primordial function of the cGAS pathway, *Nature*. 567 (2019) 262–266. doi:10.1038/s41586-019-1006-9.
- [4] S. DALES, H.J. EGGERS, I. TAMM, G.E. PALADE, ELECTRON MICROSCOPIC STUDY OF THE FORMATION OF POLIOVIRUS, *Virology*. 26 (1965) 379–389.
- [5] W.T. Jackson, T.H. Giddings, M.P. Taylor, S. Mulinyawe, M. Rabinovitch, R.R. Kopito, et al., Subversion of cellular autophagosomal machinery by RNA viruses, *PLoS Biol*. 3 (2005) e156. doi:10.1371/journal.pbio.0030156.
- [6] S.W. Bird, N.D. Maynard, M.W. Covert, K. Kirkegaard, Nonlytic viral spread enhanced by autophagy components, *Proceedings of the National Academy of Sciences*. 111 (2014) 13081–13086. doi:10.1073/pnas.1401437111.
- [7] M. Alirezaei, C.T. Flynn, M.R. Wood, S. Harkins, J.L. Whitton, Coxsackievirus can exploit LC3 in both autophagy-dependent and -independent manners in vivo, *Autophagy*. 11 (2015) 1389–1407. doi:10.1080/15548627.2015.1063769.
- [8] Y. Mohamud, J. Shi, J. Qu, T. Poon, Y.C. Xue, H. Deng, et al., Enteroviral Infection Inhibits Autophagic Flux via Disruption of the SNARE Complex to Enhance Viral Replication, *CellReports*. 22 (2018) 3292–3303. doi:10.1016/j.celrep.2018.02.090.
- [9] S.Y. Yoon, Y.E. Ha, J.E. Choi, J. Ahn, H. Lee, H.-S. Kweon, et al., Coxsackievirus B4 uses autophagy for replication after calpain activation in rat primary neurons, *Journal of Virology*. 82 (2008) 11976–11978. doi:10.1128/JVI.01028-08.
- [10] Y.-R. Lee, P.-S. Wang, J.-R. Wang, H.-S. Liu, Enterovirus 71-induced autophagy increases viral replication and pathogenesis in a suckling mouse model, *Journal of Biomedical Science*. 21 (2014) 80. doi:10.1186/s12929-014-0080-4.
- [11] K.A. Klein, W.T. Jackson, Human rhinovirus 2 induces the autophagic pathway and replicates more efficiently in autophagic cells, *Journal of Virology*. 85 (2011) 9651–9654. doi:10.1128/JVI.00316-11.
- [12] V. O'Donnell, J.M. Pacheco, M. LaRocco, T. Burrage, W. Jackson, L.L. Rodriguez, et al., Foot-and-mouth disease virus utilizes an autophagic pathway during viral replication, *Virology*. 410 (2011) 142–150. doi:10.1016/j.virol.2010.10.042.
- [13] L. Hou, X. Ge, L. Xin, L. Zhou, X. Guo, H. Yang, Nonstructural proteins 2C and 3D are involved in autophagy as induced by the encephalomyocarditis virus, *Virology*. 410 (2011) 142–150. doi:10.1186/1743-422X-11-156.
- [14] J. Zhang, Y. Lan, M.Y. Li, M.M. Lamers, M. Fusade-Boyer, E. Klemm, et al., Flaviviruses Exploit the Lipid Droplet Protein AUP1 to Trigger Lipophagy and Drive Virus Production, *Cell Host & Microbe*. 23 (2018) 819–831.e5. doi:10.1016/j.chom.2018.05.005.
- [15] N.S. Heaton, G. Randall, Dengue virus-induced autophagy regulates lipid metabolism, *Cell Host & Microbe*. 8 (2010) 422–432.

- doi:10.1016/j.chom.2010.10.006.
- [16] M. Cortese, S. Goellner, E.G. Acosta, C.J. Neufeldt, O. Oleksiuk, M. Lampe, et al., Ultrastructural Characterization of Zika Virus Replication Factories, *CellReports*. 18 (2017) 2113–2123. doi:10.1016/j.celrep.2017.02.014.
- [17] D.K. Offerdahl, D.W. Dorward, B.T. Hansen, M.E. Bloom, Cytoarchitecture of Zika virus infection in human neuroblastoma and *Aedes albopictus* cell lines, *Virology*. 501 (2017) 54–62. doi:10.1016/j.virol.2016.11.002.
- [18] M.D. Hansen, I.B. Johnsen, K.A. Stiberg, T. Sherstova, T. Wakita, G.M. Richard, et al., Hepatitis C virus triggers Golgi fragmentation and autophagy through the immunity-related GTPase M, *Proceedings of the National Academy of Sciences*. 114 (2017) E3462–E3471. doi:10.1073/pnas.1616683114.
- [19] Y. Sun, S. Yu, N. Ding, C. Meng, S. Meng, S. Zhang, et al., Autophagy benefits the replication of Newcastle disease virus in chicken cells and tissues, *Journal of Virology*. 88 (2014) 525–537. doi:10.1128/JVI.01849-13.
- [20] E. Prentice, W.G. Jerome, T. Yoshimori, N. Mizushima, M.R. Denison, Coronavirus replication complex formation utilizes components of cellular autophagy, *J Biol Chem*. 279 (2004) 10136–10141. doi:10.1074/jbc.M306124200.
- [21] P. Krejbich-Trotot, B. Gay, G. Li-Pat-Yuen, J.-J. Hoarau, M.-C. Jaffar-Bandjee, L. Briant, et al., Chikungunya triggers an autophagic process which promotes viral replication, *Virology*. 432 (2011) 432. doi:10.1016/j.virol.2011.08.032.
- [22] M. Sharma, S. Bhattacharyya, K.B. Sharma, S. Chauhan, S. Asthana, M.Z. Abdin, et al., Japanese encephalitis virus activates autophagy through XBP1 and ATF6 ER stress sensors in neuronal cells, *Journal of General Virology*. 98 (2017) 1027–1039. doi:10.1099/jgv.0.000792.
- [23] A. Orvedahl, R. Sumpter, G. Xiao, A. Ng, Z. Zou, Y. Tang, et al., Image-based genome-wide siRNA screen identifies selective autophagy factors, *Nature*. 480 (2011) 113–117. doi:10.1038/nature10546.
- [24] D. Paul, R. Bartenschlager, Flaviviridae Replication Organelles: Oh, What a Tangled Web We Weave, *Annu Rev Virol*. 2 (2015) 289–310. doi:10.1146/annurev-virology-100114-055007.
- [25] G. Savidis, W.M. McDougall, P. Meraner, J.M. Perreira, J.M. Portmann, G. Trincucci, et al., Identification of Zika Virus and Dengue Virus Dependency Factors using Functional Genomics, *CellReports*. 16 (2016) 232–246. doi:10.1016/j.celrep.2016.06.028.
- [26] R. Sumpter, S. Sirasanagandla, Á.F. Fernández, Y. Wei, X. Dong, L. Franco, et al., Fanconi Anemia Proteins Function in Mitophagy and Immunity, *Cell*. 165 (2016) 867–881. doi:10.1016/j.cell.2016.04.006.
- [27] E. Abernathy, R. Mateo, K. Majzoub, N. van Buuren, S.W. Bird, J.E. Carette, et al., Differential and convergent utilization of autophagy components by positive-strand RNA viruses, *PLoS Biol*. 17 (2019) e2006926. doi:10.1371/journal.pbio.2006926.
- [28] N.J. Lennemann, C.B. Coyne, Dengue and Zika viruses subvert reticulophagy by NS2B3-mediated cleavage of FAM134B, *Autophagy*. 13 (2017) 322–332. doi:10.1080/15548627.2016.1265192.
- [29] C.C. Kemball, M. Alirezai, C.T. Flynn, M.R. Wood, S. Harkins, W.B. Kiosses, et al., Coxsackievirus infection induces autophagy-like vesicles and megaphagosomes in pancreatic acinar cells in vivo, *Journal of Virology*. 84 (2010) 12110–12124. doi:10.1128/JVI.01417-10.
- [30] J. Wong, J. Zhang, X. Si, G. Gao, I. Mao, B.M. McManus, et al., Autophagosome supports coxsackievirus B3 replication in host cells, *Journal of Virology*. 82

- (2008) 9143–9153. doi:10.1128/JVI.00641-08.
- [31] C.C. Kemball, M. Alirezaei, C.T. Flynn, M.R. Wood, S. Harkins, W.B. Kiosses, et al., Coxsackievirus infection induces autophagy-like vesicles and megaphagosomes in pancreatic acinar cells in vivo, *Journal of Virology*. 84 (2010) 12110–12124. doi:10.1128/JVI.01417-10.
- [32] S.E. Crawford, J.M. Hyser, B. Utama, M.K. Estes, Autophagy hijacked through viroporin-activated calcium/calmodulin-dependent kinase kinase- β signaling is required for rotavirus replication, *Proceedings of the National Academy of Sciences*. 109 (2012) E3405–13. doi:10.1073/pnas.1216539109.
- [33] S. Kimura, T. Noda, T. Yoshimori, Dissection of the autophagosome maturation process by a novel reporter protein, tandem fluorescently-tagged LC3, *Autophagy*. 3 (2007) 452–460. doi:10.4161/auto.4451.
- [34] P.-Y. Ke, S.S.L. Chen, Activation of the unfolded protein response and autophagy after hepatitis C virus infection suppresses innate antiviral immunity in vitro, *J. Clin. Invest.* 121 (2011) 37–56. doi:10.1172/JCI41474.
- [35] D. Sir, W.-L. Chen, J. Choi, T. Wakita, T.S.B. Yen, J.-H.J. Ou, Induction of incomplete autophagic response by hepatitis C virus via the unfolded protein response, *Hepatology*. 48 (2008) 1054–1061. doi:10.1002/hep.22464.
- [36] M. Panyasrivanit, A. Khakpoor, N. Wikan, D.R. Smith, Co-localization of constituents of the dengue virus translation and replication machinery with amphisomes, *J. Gen. Virol.* 90 (2009) 448–456. doi:10.1099/vir.0.005355-0.
- [37] A. Khakpoor, M. Panyasrivanit, N. Wikan, D.R. Smith, A role for autophagolysosomes in dengue virus 3 production in HepG2 cells, *J. Gen. Virol.* 90 (2009) 1093–1103. doi:10.1099/vir.0.007914-0.
- [38] J. Zhang, Y. Lan, S. Sanyal, Modulation of Lipid Droplet Metabolism-A Potential Target for Therapeutic Intervention in Flaviviridae Infections, *Front Microbiol.* 8 (2017) 2286. doi:10.3389/fmicb.2017.02286.
- [39] N.S. Heaton, G. Randall, Dengue Virus and Autophagy, *Viruses*. 3 (2011) 1332–1341. doi:10.3390/v3081332.
- [40] M.M. Angelini, M. Akhlaghpour, B.W. Neuman, M.J. Buchmeier, Severe acute respiratory syndrome coronavirus nonstructural proteins 3, 4, and 6 induce double-membrane vesicles, *mBio*. 4 (2013) 1967. doi:10.1128/mBio.00524-13.
- [41] E.M. Cottam, M.C. Whelband, T. Wileman, Coronavirus NSP6 restricts autophagosome expansion, *Autophagy*. 10 (2014) 1426–1441. doi:10.4161/auto.29309.
- [42] A. Lundin, R. Dijkman, T. Bergström, N. Kann, B. Adamiak, C. Hannoun, et al., Targeting membrane-bound viral RNA synthesis reveals potent inhibition of diverse coronaviruses including the middle East respiratory syndrome virus, *PLoS Pathog.* 10 (2014) e1004166. doi:10.1371/journal.ppat.1004166.
- [43] S. Miller, J. Krijnse-Locker, Modification of intracellular membrane structures for virus replication, *Nat Rev Microbiol.* 6 (2008) 363–374. doi:10.1038/nrmicro1890.
- [44] A.L. Richards, W.T. Jackson, Intracellular vesicle acidification promotes maturation of infectious poliovirus particles, *PLoS Pathog.* 8 (2012) e1003046. doi:10.1371/journal.ppat.1003046.
- [45] A.L. Richards, J.A.P. Soares-Martins, G.T. Riddell, W.T. Jackson, Generation of unique poliovirus RNA replication organelles, *mBio*. 5 (2014) e00833–13. doi:10.1128/mBio.00833-13.
- [46] D. Paul, R. Bartenschlager, Architecture and biogenesis of plus-strand RNA virus replication factories, *World J Virol.* 2 (2013) 32–48. doi:10.5501/wjv.v2.i2.32.

- [47] R. Gosert, A. Kanjanahaluethai, D. Egger, K. Bienz, S.C. Baker, RNA replication of mouse hepatitis virus takes place at double-membrane vesicles, *Journal of Virology*. 76 (2002) 3697–3708. doi:10.1128/JVI.76.8.3697-3708.2002.
- [48] E.J. Snijder, H. van Tol, N. Roos, K.W. Pedersen, Non-structural proteins 2 and 3 interact to modify host cell membranes during the formation of the arterivirus replication complex, *J. Gen. Virol.* 82 (2001) 985–994. doi:10.1099/0022-1317-82-5-985.
- [49] C.S. Goldsmith, K.M. Tatti, T.G. Ksiazek, P.E. Rollin, J.A. Comer, W.W. Lee, et al., Ultrastructural characterization of SARS coronavirus, *Emerging Infect. Dis.* 10 (2004) 320–326. doi:10.3201/eid1002.030913.
- [50] D.A. Suhy, T.H. Giddings, K. Kirkegaard, Remodeling the endoplasmic reticulum by poliovirus infection and by individual viral proteins: an autophagy-like origin for virus-induced vesicles, *Journal of Virology*. 74 (2000) 8953–8965.
- [51] Q. Liang, Z. Luo, J. Zeng, W. Chen, S.-S. Foo, S.-A. Lee, et al., Zika Virus NS4A and NS4B Proteins Deregulate Akt-mTOR Signaling in Human Fetal Neural Stem Cells to Inhibit Neurogenesis and Induce Autophagy, *Cell Stem Cell*. 19 (2016) 663–671. doi:10.1016/j.stem.2016.07.019.
- [52] P. Ferraris, E. Blanchard, P. Roingeard, Ultrastructural and biochemical analyses of hepatitis C virus-associated host cell membranes, *Journal of General Virology*. 91 (2010) 2230–2237. doi:10.1099/vir.0.022186-0.
- [53] M. Dreux, P. Gastaminza, S.F. Wieland, F.V. Chisari, The autophagy machinery is required to initiate hepatitis C virus replication, *Proceedings of the National Academy of Sciences*. 106 (2009) 14046–14051. doi:10.1073/pnas.0907344106.
- [54] I. Tanida, M. Fukasawa, T. Ueno, E. Kominami, T. Wakita, K. Hanada, Knockdown of autophagy-related gene decreases the production of infectious hepatitis C virus particles, *Autophagy*. 5 (2009) 937–945. doi:10.4161/auto.5.7.9243.
- [55] A. Filipe, J. McLauchlan, Hepatitis C virus and lipid droplets: finding a niche, *Trends Mol Med*. 21 (2015) 34–42. doi:10.1016/j.molmed.2014.11.003.
- [56] Z. Zhao, L.B. Thackray, B.C. Miller, T.M. Lynn, M.M. Becker, E. Ward, et al., Coronavirus replication does not require the autophagy gene ATG5, *Autophagy*. 3 (2007) 581–585. doi:10.4161/auto.4782.
- [57] S.-C. Huang, C.-L. Chang, P.-S. Wang, Y. Tsai, H.-S. Liu, Enterovirus 71-induced autophagy detected in vitro and in vivo promotes viral replication, *J Med Virol*. 81 (2009) 1241–1252. doi:10.1002/jmv.21502.
- [58] Y. Zhang, Z. Li, X. Ge, X. Guo, H. Yang, Autophagy promotes the replication of encephalomyocarditis virus in host cells, *Autophagy*. 7 (2011) 613–628. doi:10.4161/auto.7.6.15267.
- [59] T. Döring, L. Zeyen, C. Bartusch, R. Prange, Hepatitis B Virus Subverts the Autophagy Elongation Complex Atg5-12/16L1 and Does Not Require Atg8/LC3 Lipidation for Viral Maturation, *Journal of Virology*. 92 (2018) 101. doi:10.1128/JVI.01513-17.
- [60] Y.-R. Lee, H.-Y. Lei, M.-T. Liu, J.-R. Wang, S.-H. Chen, Y.-F. Jiang-Shieh, et al., Autophagic machinery activated by dengue virus enhances virus replication, *Virology*. 374 (2008) 240–248. doi:10.1016/j.virol.2008.02.016.
- [61] K. Liu, M.J. Czaja, Regulation of lipid stores and metabolism by lipophagy, *Cell Death & Differentiation*. 20 (2013) 3–11. doi:10.1038/cdd.2012.63.
- [62] R. Singh, S. Kaushik, Y. Wang, Y. Xiang, I. Novak, M. Komatsu, et al., Autophagy regulates lipid metabolism, *Nature*. 458 (2009) 1131–1135. doi:10.1038/nature07976.

- [63] N.S. Heaton, R. Perera, K.L. Berger, S. Khadka, D.J. Lacount, R.J. Kuhn, et al., Dengue virus nonstructural protein 3 redistributes fatty acid synthase to sites of viral replication and increases cellular fatty acid synthesis, *Proceedings of the National Academy of Sciences*. 107 (2010) 17345–17350. doi:10.1073/pnas.1010811107.
- [64] L. Gerstenmaier, R. Pilla, L. Herrmann, H. Herrmann, M. Prado, G.J. Villafano, et al., The autophagic machinery ensures nonlytic transmission of mycobacteria, *Proceedings of the National Academy of Sciences*. 112 (2015) E687–92. doi:10.1073/pnas.1423318112.
- [65] M. Ponpuak, M.A. Mandell, T. Kimura, S. Chauhan, C. Cleyrat, V. Deretic, Secretory autophagy, *Curr Opin Cell Biol*. 35 (2015) 106–116. doi:10.1016/j.ceb.2015.04.016.
- [66] Y.-H. Chen, W. Du, M.C. Hagemeijer, P.M. Takvorian, C. Pau, A. Cali, et al., Phosphatidylserine vesicles enable efficient en bloc transmission of enteroviruses, *Cell*. 160 (2015) 619–630. doi:10.1016/j.cell.2015.01.032.
- [67] S.M. Robinson, G. Tsueng, J. Sin, V. Mangale, S. Rahawi, L.L. McIntyre, et al., Coxsackievirus B exits the host cell in shed microvesicles displaying autophagosomal markers, *PLoS Pathog*. 10 (2014) e1004045. doi:10.1371/journal.ppat.1004045.
- [68] B. Cao, L.A. Parnell, M.S. Diamond, I.U. Mysorekar, Inhibition of autophagy limits vertical transmission of Zika virus in pregnant mice, *Journal of Experimental Medicine*. 214 (2017) 2303–2313. doi:10.1084/jem.20170957.
- [69] Y.-W. Wu, C. Mettling, S.-R. Wu, C.-Y. Yu, G.C. Perng, Y.-S. Lin, et al., Autophagy-associated dengue vesicles promote viral transmission avoiding antibody neutralization, *Sci. Rep.* 6 (2016) 32243. doi:10.1038/srep32243.
- [70] R. Mateo, C.M. Nagamine, J. Spagnolo, E. Méndez, M. Rahe, M. Gale, et al., Inhibition of cellular autophagy deranges dengue virion maturation, *Journal of Virology*. 87 (2013) 1312–1321. doi:10.1128/JVI.02177-12.
- [71] S. Shrivastava, A. Raychoudhuri, R. Steele, R. Ray, R.B. Ray, Knockdown of autophagy enhances the innate immune response in hepatitis C virus-infected hepatocytes, *Hepatology*. 53 (2011) 406–414. doi:10.1002/hep.24073.
- [72] P.-Y. Ke, S.S.L. Chen, Activation of the unfolded protein response and autophagy after hepatitis C virus infection suppresses innate antiviral immunity in vitro, *Journal of Clinical Investigation*. 121 (2011) 37–56. doi:10.1172/JCI41474DS1.
- [73] J.M. Mackenzie, A.A. Khromykh, R.G. Parton, Cholesterol manipulation by West Nile virus perturbs the cellular immune response, *Cell Host & Microbe*. 2 (2007) 229–239. doi:10.1016/j.chom.2007.09.003.
- [74] B. Levine, J. Yuan, Autophagy in cell death: an innocent convict? *Journal of Clinical Investigation*. 115 (2005) 2679–2688. doi:10.1172/JCI26390.
- [75] G. Kroemer, M. Jäättelä, Lysosomes and autophagy in cell death control, *Nat. Rev. Cancer*. 5 (2005) 886–897. doi:10.1038/nrc1738.
- [76] B. Levine, S.C. Sinha, G. Kroemer, Bcl-2 family members: Dual regulators of apoptosis and autophagy, *Autophagy*. 4 (2008) 600–606. doi:10.4161/auto.6260.
- [77] J.E. McLean, A. Wudzinska, E. Datan, D. Quaglino, Z. Zakeri, Flavivirus NS4A-induced autophagy protects cells against death and enhances virus replication, *Journal of Biological Chemistry*. 286 (2011) 22147–22159. doi:10.1074/jbc.M110.192500.
- [78] S.Y. Yoon, Y.E. Ha, J.E. Choi, J. Ahn, H. Lee, D.H. Kim, Autophagy in coxsackievirus-infected neurons, *Autophagy*. 5 (2009) 388–389.

- doi:10.4161/auto.5.3.7723.
- [79] A.J. Chase, F.A. Medina, J.L. Muñoz-Jordan, Impairment of CD4+ T Cell Polarization by Dengue Virus–Infected Dendritic Cells, *J. Infect. Dis.* 203 (2011) 1763–1774. doi:10.1093/infdis/jir197.
- [80] D.R. Palmer, P. Sun, C. Celluzzi, J. Bisbing, S. Pang, W. Sun, et al., Differential effects of dengue virus on infected and bystander dendritic cells, *Journal of Virology*. 79 (2005) 2432–2439. doi:10.1128/JVI.79.4.2432-2439.2005.
- [81] A.G. Aleyas, Y.W. Han, J.A. George, B. Kim, K. Kim, C.-K. Lee, et al., Multifront assault on antigen presentation by Japanese encephalitis virus subverts CD8+ T cell responses, *The Journal of Immunology*. 185 (2010) 1429–1441. doi:10.4049/jimmunol.0902536.
- [82] P. Metz, A. Chiramel, L. Chatel-Chaix, G. Alvisi, P. Bankhead, R. Mora-Rodríguez, et al., Dengue Virus Inhibition of Autophagic Flux and Dependency of Viral Replication on Proteasomal Degradation of the Autophagy Receptor p62, *Journal of Virology*. 89 (2015) 8026–8041. doi:10.1128/JVI.00787-15.
- [83] J.D. Smith, E. de Harven, Herpes simplex virus and human cytomegalovirus replication in WI-38 cells. III. Cytochemical localization of lysosomal enzymes in infected cells, *Journal of Virology*. 26 (1978) 102–109.
- [84] S. Shelly, N. Lukinova, S. Bambina, A. Berman, S. Cherry, Autophagy is an essential component of *Drosophila* immunity against vesicular stomatitis virus, *Immunity*. 30 (2009) 588–598. doi:10.1016/j.immuni.2009.02.009.
- [85] A. Orvedahl, D. Alexander, Z. Tallóczy, Q. Sun, Y. Wei, W. Zhang, et al., HSV-1 ICP34.5 confers neurovirulence by targeting the Beclin 1 autophagy protein, *Cell Host & Microbe*. 1 (2007) 23–35. doi:10.1016/j.chom.2006.12.001.
- [86] G.B. Kyei, C. Dinkins, A.S. Davis, E. Roberts, S.B. Singh, C. Dong, et al., Autophagy pathway intersects with HIV-1 biosynthesis and regulates viral yields in macrophages, *J Cell Biol*. 186 (2009) 255–268. doi:10.1083/jcb.200903070.
- [87] X.H. Liang, L.K. Kleeman, H.H. Jiang, G. Gordon, J.E. Goldman, G. Berry, et al., Protection against fatal Sindbis virus encephalitis by beclin, a novel Bcl-2-interacting protein, *Journal of Virology*. 72 (1998) 8586–8596.
- [88] A. Orvedahl, S. MacPherson, R. Sumpter, Z. Tallóczy, Z. Zou, B. Levine, Autophagy protects against Sindbis virus infection of the central nervous system, *Cell Host & Microbe*. 7 (2010) 115–127. doi:10.1016/j.chom.2010.01.007.
- [89] J. Staring, E. von Castelmur, V.A. Blomen, L.G. van den Hengel, M. Brockmann, J. Baggen, et al., PLA2G16 represents a switch between entry and clearance of Picornaviridae, *Nature*. 541 (2017) 412–416. doi:10.1038/nature21032.
- [90] Y. Mohamud, J. Qu, Y.C. Xue, H. Liu, H. Deng, H. Luo, CALCOCO2/NDP52 and SQSTM1/p62 differentially regulate coxsackievirus B3 propagation, *Cell Death & Differentiation*. (2018). doi:10.1038/s41418-018-0185-5.
- [91] N. Kim, M.-J. Kim, P.S. Sung, Y.C. Bae, E.-C. Shin, J.-Y. Yoo, Interferon-inducible protein SCOTIN interferes with HCV replication through the autolysosomal degradation of NS5A, *Nat Commun*. 7 (2016) 10631. doi:10.1038/ncomms10631.
- [92] Y. Liu, B. Gordesky-Gold, M. Leney-Greene, N.L. Weinbren, M. Tudor, S. Cherry, Inflammation-Induced, STING-Dependent Autophagy Restricts Zika Virus Infection in the *Drosophila* Brain, *Cell Host & Microbe*. 24 (2018) 57–68.e3. doi:10.1016/j.chom.2018.05.022.
- [93] A.-B. Blázquez, M.A. Martín-Acebes, J.-C. Saiz, Amino acid substitutions in the non-structural proteins 4A or 4B modulate the induction of autophagy in West Nile virus infected cells independently of the activation of the unfolded protein

- response, *Front Microbiol.* 5 (2014) 797. doi:10.3389/fmicb.2014.00797.
- [94] Z.-W. Zhang, Z.-L. Li, S. Yuan, The Role of Secretory Autophagy in Zika Virus Transfer through the Placental Barrier, *Front Cell Infect Microbiol.* 6 (2016) 206. doi:10.3389/fcimb.2016.00206.
- [95] M. Sharma, S. Bhattacharyya, M. Nain, M. Kaur, V. Sood, V. Gupta, et al., Japanese encephalitis virus replication is negatively regulated by autophagy and occurs on LC3-I- and EDEM1-containing membranes, *Autophagy.* 10 (2014) 1637–1651. doi:10.4161/auto.29455.
- [96] W.-C. Su, T.-C. Chao, Y.-L. Huang, S.-C. Weng, K.-S. Jeng, M.M.C. Lai, Rab5 and class III phosphoinositide 3-kinase Vps34 are involved in hepatitis C virus NS4B-induced autophagy, *Journal of Virology.* 85 (2011) 10561–10571. doi:10.1128/JVI.00173-11.
- [97] S. Shrivastava, J. Bhanja Chowdhury, R. Steele, R. Ray, R.B. Ray, Hepatitis C virus upregulates Beclin1 for induction of autophagy and activates mTOR signaling, *Journal of Virology.* 86 (2012) 8705–8712. doi:10.1128/JVI.00616-12.
- [98] A.M. Fahmy, P. Labonté, The autophagy elongation complex (ATG5-12/16L1) positively regulates HCV replication and is required for wild-type membranous web formation, *Sci. Rep.* 7 (2017) 40351. doi:10.1038/srep40351.
- [99] X. Chen, K. Wang, Y. Xing, J. Tu, X. Yang, Q. Zhao, et al., Coronavirus membrane-associated papain-like proteases induce autophagy through interacting with Beclin1 to negatively regulate antiviral innate immunity, *Protein Cell.* 5 (2014) 912–927. doi:10.1007/s13238-014-0104-6.
- [100] F. Reggiori, I. Monastyrska, M.H. Verheije, T. Cali, M. Ulasli, S. Bianchi, et al., Coronaviruses Hijack the LC3-I-positive EDEMosomes, ER-derived vesicles exporting short-lived ERAD regulators, for replication, *Cell Host & Microbe.* 7 (2010) 500–508. doi:10.1016/j.chom.2010.05.013.
- [101] E.M. Cottam, H.J. Maier, M. Manifava, L.C. Vaux, P. Chandra-Schoenfelder, W. Gerner, et al., Coronavirus nsp6 proteins generate autophagosomes from the endoplasmic reticulum via an omegasome intermediate, *Autophagy.* 7 (2011) 1335–1347. doi:10.4161/auto.7.11.16642.
- [102] J. Shi, J. Wong, P. Piesik, G. Fung, J. Zhang, J. Jagdeo, et al., Cleavage of sequestosome 1/p62 by an enteroviral protease results in disrupted selective autophagy and impaired NFKB signaling, *Autophagy.* 9 (2013) 1591–1603. doi:10.4161/auto.26059.
- [103] H. Wu, X. Zhai, Y. Chen, R. Wang, L. Lin, S. Chen, et al., Protein 2B of Coxsackievirus B3 Induces Autophagy Relying on Its Transmembrane Hydrophobic Sequences, *Viruses.* 8 (2016). doi:10.3390/v8050131.
- [104] A.K. Corona, H.M. Saulsbery, A.F. Corona Velazquez, W.T. Jackson, Enteroviruses Remodel Autophagic Trafficking through Regulation of Host SNARE Proteins to Promote Virus Replication and Cell Exit, *CellReports.* 22 (2018) 3304–3314. doi:10.1016/j.celrep.2018.03.003.
- [105] P. Sun, S. Zhang, X. Qin, X. Chang, X. Cui, H. Li, et al., Foot-and-mouth disease virus capsid protein VP2 activates the cellular EIF2S1-ATF4 pathway and induces autophagy via HSPB1, *Autophagy.* 14 (2018) 336–346. doi:10.1080/15548627.2017.1405187.
- [106] D. Ao, H.-C. Guo, S.-Q. Sun, D.-H. Sun, T.S. Fung, Y.-Q. Wei, et al., Viroporin Activity of the Foot-and-Mouth Disease Virus Non-Structural 2B Protein, *PLoS ONE.* 10 (2015) e0125828. doi:10.1371/journal.pone.0125828.
- [107] M.P. Taylor, K. Kirkegaard, Modification of cellular autophagy protein LC3 by poliovirus, *Journal of Virology.* 81 (2007) 12543–12553. doi:10.1128/JVI.00755-07.

- [108] J. Wang, J.B. Ptacek, K. Kirkegaard, E. Bullitt, Double-membraned liposomes sculpted by poliovirus 3AB protein, *Journal of Biological Chemistry*. 288 (2013) 27287–27298. doi:10.1074/jbc.M113.498899.