

Evidence for immortality and autonomy as criteria for neoplasms has not been shown for the lesions resulting from most animal models of carcinogenesis

Xixi Dou^{1*}, Mingjuan Lei², Haiyan Zhou³, Lichan Chen², Lucas Zellmer⁴, Qingwen Jia¹, Daizhou Zhang¹, Ningzhi Xu^{5*}, Chengguang Wang^{6*}, and Dezhong Joshua Liao^{7*}

¹Shandong Provincial Key Laboratory of Transmucosal and Transdermal Drug Delivery, Shandong Freda Pharmaceutical Group Co., Ltd., Jinan 250101, Shandong Province, China

²Hormel Institute, University of Minnesota, Austin, MN 55912

³Clinical Research Center, Guizhou Medical University Hospital, Guiyang 550004, Guizhou Province, China

⁴Masonic Cancer Center, University of Minnesota, 435 E. River Road, Minneapolis, MN 55455, USA

⁵Laboratory of Cell and Molecular Biology & State Key Laboratory of Molecular Oncology, National Cancer Center/Cancer Hospital, Chinese Academy of Medical Sciences and Peking Union Medical College, Beijing 100021, China.

⁶Tianjin LIPOGEN gene technology Ltd., #238 Baidi Road, Nankai District, Tianjin 300192, China

⁷Department of Pathology, the Second Hospital of Guizhou University of Traditional Chinese Medicine, 32 Fei-shan Street, Guiyang 550001, Guizhou Province, China

*Addresses for correspondence:

Xixi Dou

Shandong Provincial Key Laboratory of Transmucosal and Transdermal Drug Delivery

Shandong Freda Pharmaceutical Group Co., Ltd.

Jinan 250101, Shandong Province

China

Email: xrl@sdfmg.com

Ningzhi Xu

Laboratory of Cell and Molecular Biology

National Cancer Center/Cancer Hospital

Chinese Academy of Medical Sciences and Peking Union Medical College

Beijing 100021, China

Email: xuningzhi@cicams.ac.cn

Chengguang Wang

Tianjin LIPOGEN gene technology Ltd.

#238 Baidi Road, Nankai District, Tianjin 300192

China

Email: wang@lipogenes.com

Joshua Liao

Department of Pathology

The second Hospital

Guizhou University of Traditional Chinese Medicine

Guiyang 550001, Guizhou Province

China

Email: djliao@gyctcm.edu.cn

Abstract

Modern research into carcinogenesis has undergone three phases. Surgeons and pathologists started the first phase and established autopsy and biopsy as routine pathology services, in turn establishing morphological traits for tumors and establishing immortality and autonomy as indispensable criteria for neoplasms. A century ago medical doctors and biologists initiated “experimental cancer research” as the second phase, in which they, with help from chemists, established many chemical-induced animal models of carcinogenesis. In this phase, the two-hit theory and stepwise carcinogenesis of “initiation-promotion” or “initiation-promotion-progression” were established, with an illustrious finding that outgrowths induced in animals depend on the inducers, and thus are not authentically neoplastic, until late stages. For the last 40 years, molecular biologists have gradually dominated the carcinogenesis research fraternity and have established numerous genetically-modified animal models of carcinogenesis. However, evidence has not been provided for immortality and autonomy of the lesions from most of these models. Probably, many peers had already collected the lesions from animals for analyses of “cancer” mechanisms before the lesions became autonomous. We herein review monumental work of many predecessors to reinforce that evidence for immortality and autonomy is essential for confirming a neoplastic nature. We extrapolate that immortality and autonomy are established early during sporadic human carcinogenesis, unlike the late establishment in all animal models. It is imperative to resume many forerunners’ work by determining the genetic bases for initiation, promotion and progression, the genetic bases for immortality and autonomy, and which animal models are, in fact, good for identifying such genetic bases.

Keywords:

Transgenic; Cancer; Carcinogenesis; Immortality; Autonomy; cancer stem cells; senescence

Running title: Many tumors induced in animals are inauthentic

Introduction

Cancer research has been going on for 2,700 years [1], although systematic study of cancer might not be started until 1775 when Pott reported his surgical observation of cancers in several body sites [2]. Therefore, its consequent literature is so enormous in size that it becomes a problem for researchers. Actually, even in 1955, Alexander Haddow (1907-1976) had pointed this out, writing that “the mere abundance of the data ...presents a growing problem, towards which there are two extreme types of reaction: first, that of the happy researcher who is content to ignore the original literature, and to rely upon others for his information; and secondly, the reaction of those whom the literature totally enslaves” [3]. Few of today’s researchers peruse the ancient literature, due to their many tiers of stress, such as dwindling funding and increasing difficulty in obtaining tenured faculty positions, besides the sheer volume of the literature to read. As a repercussion, cancer research manifests a discontinuous growth, just like cancer itself that is a discontinuous growth from its normal parental cell, meaning that few of today’s cancer students know and address the questions raised by their predecessors.

We write this perspective article to review some seminal findings by different trailblazers in “carcinogenesis research”, which is herein defined as the study 1) on the procedures that convert a normal cell to a malignant one and then to more malignant states, and 2) on the mechanisms underlying these steps. The clinical aspects of cancer will not be discussed to avoid digression. The idiom of “tumorigenesis”, which is of broader scope as it also covers the formation of benign tumors, is used sometimes, partly because many animal models produce both benign and malignant lesions. In our opinion, modern research on tumorigenesis, mainly carcinogenesis, has undergone three phases. The first one was the epoch of Rudolf Ludwig Carl Virchow (1821-1902), who proposed the theory that cancer formation is a result of chronic irritation [4-7], mainly chronic inflammation. The cancer research fraternity in this phase was dominated by surgeons and pathologists, who established autopsy and biopsy as the routine pathology practice that leads to the establishment of the morphological traits of tumors [8]. The second phase had its inception roughly at the beginning of the 20th century and was coined by cancer pathologist Harold Leroy Stewart (1908-1998) as an epoch of “experimental cancer research” [8]. In this lengthy second incarnation, medical doctors and biologists, with great help from chemists, established and characterized many animal models of chemical-induced tumorigenesis. With these models, they established the two-hit theory [9], mutation theory [10-12] and clonal evolution theory [13-16], as well as the multi-stage [17-19], i.e. initiation-promotion [20-25] or initiation-promotion-progression [26-29], models of carcinogenesis. Moreover, animal models of irradiation-induced carcinogenesis also emerged during this period [30].

Starting about 40 years ago when molecular cloning techniques began to disperse swiftly over the whole of biomedical research, molecular biologists, many lacking strict training and clinical experience in surgical pathology or oncology, have gradually replaced medical doctors and traditional biologists to now dominate the fraternity of carcinogenesis research [31, 32], thus moving “experimental cancer research” into a new phase. In this latest incarnation, molecular biologists have established numerous genetically manipulated animal models of carcinogenesis and in vitro systems of neoplastic transformation of normal cells. With these in vivo and in vitro models, we now know deeper mechanisms of how genes regulate behaviors of normal and neoplastic cells. Indeed, thanks to these achievements by molecular biologists, we now enjoy enormous amounts of information and great details on molecular signaling pathways for almost all physiological functions and pathological alterations in the human body. However, few of the genetic animal models established so far address the traditional multiple stages of “initiation-promotion-progression” [28], leaving those mavens who are familiar with their predecessors’ work to wonder how to couple the stepwise biological changes observed previously with the molecular alterations seen in these genetic models. Moreover, few of the publications reporting these genetic models provide material evidence for immortality and autonomy of the resulting lesions. To warrant this statement, we encourage readers to search published reports of these genetic models for “immortal”, “autonomous” or similar keywords, to see how many of them describe these properties of the resulting lesions. By reviewing work of many forerunners, most being preeminent cancer pathologists, we attempt in this essay to reinforce immortality and autonomy as the cardinal, yet long-neglected, criteria to qualify outgrowths as neoplasms, even the benign ones.

Many chemical-induced tumors in animals depend on the chemical until late stages

To our knowledge, the first experimentally induced tumors in animals were reported, in the German literature, by Fischer in 1906 (B. Fischer, Münch. med Wochnschr. 1906; 42: 2041). According to Davis [33, 34] and Vasiliev [35], Fischer showed that subcutaneous injections of Scarlet Red into the ears of rabbits induced papillomas, which are benign tumors, but the tumors regressed upon discontinuation of the injections, although they could be quickly re-induced with Scarlet Red. According to Davis [33, 34], these phenomena were confirmed by Helmholtz in 1907 and by Werner in 1908. Between 1914 and 1924, Katsusaburo Yamagiwa (1863-1930), after he left Virchow and returned to Japan [36, 37], was able to induce papillomas and papilocarcinomas in rabbits’ ears by painting the ears with coal tar, with metastases to nearby lymph nodes in some cases. Yamagiwa also found that the tumors, including some carcinomas with metastases, regressed after cessation of the tar-painting, but repainting could quickly

cause the recurrence of the tumors [38, 39]. Yamagiwa thus concluded that “carcinomas do not develop as carcinomas from the beginning, and do not always continue as carcinomas” [39]. This “do not always continue” is the first statement in the literature, to our knowledge, stating that induced cancer can disappear spontaneously. During the 1930s and 1940s, Peyton Rous (1879-1970), a Nobel laureate, had also performed a series of studies on tar-induced tumors in rabbits’ ears, and observed the regression of the lesions upon tar discontinuation and their quick reappearance upon tar repainting [25, 40, 41]. Actually, according to Rous, Des Ligneris had already confirmed in 1930 that, “...a second period of tarring brings out warts sooner than the first” [40]. Realizing that the reversible lesions could not be authentic neoplasms, Rous described them as warts, which is a hyperplastic type of lesion, and wrote in 1940 that, “...it will be seen that the tar warts of rabbits are tumors by all of the standard criteria except two. They have no capacity for independent growth like that exhibited by most (but not all) classical tumors; and the changes in their cells may conceivably be reversible since they often become smaller and vanish [40]. The two unmet criteria in Rous’ observations, i.e. “no capacity for independent growth” and being “reversible”, are later referred to by many cancer savants as “autonomy” and “immortality”. Rous further wrote that, “...in the current definition of a tumor no allowance is made for neoplasms which depend upon favoring factors for existence, and it cannot be used to rule them out” [40]. Here, the lesion inducer is dubbed as “favoring factors”.

Chronic treatment of rats with 7,12-dimethylbenz(a)anthracene (DMBA) could induce mammary tumors, but maintenance of the tumors is dependent on the continuous administration of DMBA [42-45]. Continuous feeding of rats with 3-methylcholanthrene could induce palpable mammary tumors as early as the 20th day from the start of the experiment [46]. Painting the skin of C57 brown mice with 3-methylcholanthrene could also induce palpable tumors as early as the 31st day [47]. However, 15 of the 22 induced skin tumors regressed completely upon cessation of the inducer, with hair eventually growing in the epilated areas; of the seven papillomas that remained, three eventually evolved to lesions with malignant morphology and four persisted [47]. Similarly, a large number of papillomas could be induced by painting the skin of albino mice with 3:4-benzopyrene, but the tumors actually sloughed off, and only a few continuously progressed to carcinomas [48]. After having studied successive stages of carcinogenesis [49-52], Rusch wrote in the 1950 that carcinogenesis generally consisted of “induction, reversibility and progression” [53], which clearly points out that the lesions can be immortal and autonomous only at a late stage. The typical inducer-dependency until a late stage can be exemplified by the skin carcinogenesis model presented by Berenblum in 1947 [21], which, in Haddow’s words, “proceeds from the normal epithelium first to an early non-specific hyperplasia, second to a specific pre-neoplastic hyperplasia, and

then to the emergence of papillomata, and how later stages can be recognized in the progressive growth of such papillomata, their conversion into carcinoma, and the uncontrolled growth of the latter...This general sequence takes place equally well whether exposure to the carcinogen is continued or not" [54].

Hormone-induced tumors in animals are also inducer-dependent until late stages

Chronic treatment of rats and mice with estrogens can induce cancers in the bladder and mammary glands as well as benign tumors in the pituitary and testes [55-70], and can also induce uterine tumors [71]. The ACI strain (August strain crossed with Copenhagen strain, also called AxC) of rats may be more susceptible than other strains in the induction of the mammary and pituitary tumors [56, 72], but we once found that about one-fourth of the females lacked one side of the uterus and ovary (DJ Liao's unpublished data), suggesting that the ACI strain may bear a recessive mutation in a relevant but not yet identified gene. Treatment of mice with estrogen or with both estrogen and androgen can induce benign and malignant tumors in the cervix and vagina, and the malignant tumors are transplantable to other mice treated with the hormones [73-79]. Administration of androgens to rats can induce prostate [80-84] and uterine [85, 86] cancers. Concomitant treatment of rats with estrogen and androgen can induce mammary and prostate cancers much more quickly than administration of androgen or estrogen alone [80-84, 87-92], and can induce uterine leiomyomas as well [93]. Administration of estrogen to hamsters can induce malignant renal tumors with abdominal metastases [94-99], while administration of both estrogen and androgen to hamsters can induce malignant tumors in the kidneys and can induce benign and malignant tumors in the uterus, in the skin, and in the epididymal tail and adjacent ductus deferens [80, 100, 101]. In addition, gonadal and gonadotrophic hormones have also been shown to possess the ability to induce endocrine cell tumors in ovaries or testes [68, 102-105].

Estrogen-induced mammary cancer as well as pituitary and testicular tumors have been known, ever since the 1930s, to regress partially or completely upon withdrawal of the hormone, and the tumors can sustain themselves without estrogen treatment only at very late stages [55, 65, 78, 106-123] (and DJ Liao's personal experience). Initially, estrogen-induced pituitary tumors can be transplantable only to the animals treated with estrogens, but not to the untreated animals, evincing their dependency on an excessive amount of estrogen [55]. However, they can eventually evolve to estrogen-independency [124, 125]. The Nobel laureate Charles B. Huggins (1901-1997) had shown in both animal studies and human clinics that castration or treatment with estrogens could cause regression of prostate cancer at certain stages, signifying that this cancer is hormone-dependent until a late stage [126-128]. In the words of Jacob Furth (1896-1979), a renowned pathologist [129-131], "this (prostate) tumor is an example of a growth in

man with a spectrum ranging from conditioned to highly autonomous type. The cases of Huggins that were controlled by castration (that is, removal of sources of androgens) may be regarded as dependent; those which partially or temporarily regressed after castration or estrogen treatment, as partially dependent; those not influenced by such therapy, as autonomous" [132]. Similarly, it is well known, with the earliest reference adduced herein dating back to 1919 [133], that human breast cancer at certain stages is estrogen dependent, and thus is often treated with antiestrogens. Estrogen-induced renal tumors in hamsters, including their abdominal metastases, will regress upon cessation of the estrogen treatment unless the tumors are at very advanced stages [134-137] (and DJ Liao's empirical knowledge). Initially, these renal tumors can be transplantable only to those hamsters that are treated with estrogen, but not to the untreated hamsters, bespeaking that the tumors still depend on an excessive amount of estrogen. However, these nonautonomous tumors can be converted to autonomous ones by manipulating the estrogen treatment of the recipient animals [97, 137, 138].

Treatment of mice with iodine-131 (I-131) can induce pituitary tumors that secrete thyroid stimulating hormone (TSH) [55, 139-142]. This is because I-131 damages the thyroid and thus decreases the levels of thyroid hormones, which in turn stimulates proliferation of TSH secretory cells in the pituitary with formation of TSH-secreting adenomas as the sequel [143-147]. By the same principle, partial thyroidectomy of rats and mice can cause pituitary adenomas as well [148-153]. The tumors can be transplantable [154]; initially only to those mice treated with thiouracil or other goitrogenic compounds that induce TSH, and then gradually to normal mice [155], which again shows the trajectory of "initial dependence and then autonomy", as said by Furth in 1953 [132].

Thyroid neoplasms can be induced in mice by treatment with thiouracil or other goitrogenic compounds that block thyroid hormone secretion and, in turn, induce TSH secretion from the pituitary [148, 151, 152, 156-166]. It is also the high level of TSH that induces the thyroid tumors, similar to the aforesaid induction of pituitary tumors. By the same principle, I-131 can induce thyroid tumors as well [145, 167-170]. Initially, the tumors can be transplantable only to the mice treated with thiouracil, but in a course of subpassages, the tumor cells can eventually be transplantable to normal mice, which, once more, shows the conversion from hormone (TSH) dependent to independent states [132, 148, 151, 152, 157]. These thyroid tumors, though dependent on TSH, often metastasize to lymph nodes [132] and the lungs [132, 148, 159]. In 1953, Furth wrote that "conditions can be created whereby uncontrolled proliferation of one cell type is obtained, resulting in a tumor-like growth. Manipulations attaining this need not involve any intrinsic alteration in cells causing them to behave as cancer cells. Whether or not such tumors and the similar human metastasizing thyroid adenomas are considered neoplastic depends

on the definition of a neoplasm. In our terminology such thyroid tumors are conditioned neoplasms. In the course of subpassages in thiouracil-treated mice the dependent growths give rise to autonomous growths which possess individual features of their own and can be grafted on normal mice. Thyroid adenomas induced by TSH-secreting pituitary tumors are indistinguishable from those induced by thiouracil" [132]. Here, Furth used "tumor-like growth", "behave as cancer cells" and "conditional neoplasms" to express his reservation in considering the induced pituitary and thyroid tumors, even the spontaneous human thyroid tumors, as authentic; despite their ability to metastasize. In his opinion, "dependent tumors are those in which apparently normal cells proliferate in an altered host; autonomous tumors are those in which permanently altered cells proliferate in normal hosts" [132], although, based on our training in human pathology, we opine that dependent tumor cells are not normal but are hyperplastic.

Immortality and autonomy had already become indispensable criteria for neoplasms over a century ago

To our knowledge, the abovementioned studies on chemical- or hormone-induced outgrowths are among the earliest ones that point out the problem of "inducer-dependency" and set immortality and autonomy as criteria for neoplasms in experimental animals. Actually, before these animal studies became well recognized, some German pathology textbooks published in the 1910s had already outlined these criteria for human neoplasms, according to Rous [40]. Jacob Furth [132] and transcendent pathologist James Ewing (1866-1943) [171] considered that all tumors, including those benign ones that would not progress to malignancy, should be in some form of autonomy. Haddow wrote in the 1947 that, "...we now know that, while constitutional and genetic factors can greatly influence susceptibility to cancer, and many even determine the site of its spontaneous occurrence, the disease is one of the individual cells as a separate organism and with no relation to the needs of the body as a whole. It is this which gives cancer its unique position in pathology, accounts for its intractable nature, and explains its growth, in Paget's words 'irrespective of the maintenance of the rest of the body, discordant from its normal type, and with no seeming purpose' (Paget, 1853)" [54]. The quoted words of Paget had already, in 1853, pointed out tumors' autonomous nature. Indeed, according to Haddow [54] and Knauss et al [172], a cancer has long been regarded as a new race or new strain of organism, which is another way of describing autonomy, dating back to 1897 by David Hansemann, 1903 by G. Hauser (Beitr. Path. Anat., 1903; 33, 1) and 1926 by Menetrier. Many other former pundits also described carcinogenesis as an atavistic procedure, further pointing out that the resulting "new race of organism" is evolutionarily-lower on the life-tree than its host animal [173-176].

What do “immortality” and “autonomy” really mean pertaining to a neoplasm? Immortality of tumor cells does not mean that each cell can survive forever. On the contrary, “that cancer cells are often sick cells and die young is known to every pathologist”, as Rous had pointed out in 1941 [40]. Instead, immortality betokens that a tumor can survive as a “newly developed independent organism” [40] that parasitizes the host patient [173, 174] and forever maintains its life by continuous replication of its cells, just like bacteria that maintain their strains by ongoing cell propagation, as we described before [175, 177, 178]. Harry S.N. Greene (1904-1969), a preeminent surgical pathologist at Yale University, elaborated on the autonomy by writing in 1951 that, “...the definition of a tumor as an autonomous growth has enjoyed persistent popularity in textbooks of pathology. In such definitions the adjective ‘autonomous’ is employed to express the idea of independence with respect to two different particulars. One of these relates to freedom from the laws restraining and coordinating normal tissue growth, and the other concerns release from the necessity of a continued stimulus” [179]. According to Furth’s translation [132], in an article in the German language from 1951, Bungeler considered that a dependence seen in a large variety of outgrowths in humans indicates that the outgrowths are not true tumors. More critically, there is no transition between the dependent and autonomous outgrowths, according to Furth’s translation of Bungeler’s words [132]. This “no transition” means that whether an outgrowth is autonomous or not is a black-and-white demarcation between neoplasms and non-neoplasms without leaving any room for us to confuse or admix the two. Describing human cancer’s properties, Emmanuel Farber (1918-2014), an illustrious cancer pathologist and former chair of the American Association of Cancer Research, also accentuated autonomy as a cornerstone of cancer biology [180]. Notwithstanding, it also needs to be pointed out that autonomy of tumor cells may be achieved in part via non-autonomous mechanisms, i.e. various interactions with other cell types [181-183].

Some tumors from genetically manipulated animals are inducer-dependent as well

The c-myc gene or a mutant of the k-ras gene in transgenic mice can induce malignant tumors in all target tissues or organs studied so far, such as in the liver, pancreas, lungs and mammary glands, as we have shown or reviewed before [116, 184-191]. However, many of these tumors have been shown to regress upon turning off the transgene and can be sustained without the expression of the transgene only at very advanced stages, although, once they have regressed, they can be quickly re-induced through reactivation of the transgene [191-200]. Xmrk, c-myc, mutant k-ras, or SV40 large T oncogene can also induce liver cancer in transgenic Zebrafish, and, again, the tumors will regress after inactivation of the transgene [201-206]. Conversely, inactivation of the tumor suppressor gene p53 via conditional knockout

can beget tumor formation, but reactivation of the p53 leads to regression of the tumors [205-211]. This phenomenon of “regression upon the inducer withdrawal and quick repopulation upon reintroduction of the inducer” is a full reflection of the same phenomenon seen in the chemical- or hormone-induced carcinogenesis in animals described above, and has become a rationale for targeting therapy in cancer [212-214]. Our contemporaries in the third phase of carcinogenesis research consider “regression upon inducer withdrawal” as “oncogene addiction” and “tumor dormancy” as the reason for “the tumor repopulation upon reintroduction of the inducer” [192, 196, 199, 212-218], but, peculiarly, without mentioning the same phenomenon observed by our predecessors.

Most animal models have not yet been tested for the trajectory of “induction, reversibility and progression”

There are still many animal models of carcinogenesis induced by chemicals or hormones that have not yet been determined for inducer-dependency. Even worse, only a few genetically-modified models, as described above, have been tested for the dependency, whereas the remaining vast majority have not yet been tested, presumably because many molecular biologists have not realized that immortality and autonomy are prerequisite criteria for neoplasms. For visceral tumor induction, like N-nitrosobis(2-hydroxypropyl)amine-instigated lung tumors [219], this situation is especially true, because the determination requires sacrifice of the animals. We surmise that most of the undetermined animal models may also show an inducer dependency until a late stage, with their carcinogenic procedures following the trajectory of “induction, reversibility and progression” described by Rusch in 1950 [53]. Considering that c-myc and mutant k-ras are the most potent oncogenes and the lesions they induced already manifest such dependency, probably other genetic models will show this trajectory as well. Nevertheless, this conjecture needs to be substantiated by studying untested animal models, especially the new ones to be established in the future, using a conditional transgenic or knockout approach.

Spontaneous regression of human neoplasms occurs but is rare

In humans, spontaneous regression of a solid tumor or remission of a liquid neoplasm, such as leukemia or myeloma, is extremely rare, but it is recurrently shown in case reports [220-235]. The frequency of spontaneous regression or remission is actually uncertain, as most relevant studies adduced only old figures that are between 1/60,000-100,000 cases [222, 236, 237]. The frequency may vary greatly among different cancer types. To our knowledge, malignant melanoma may have the highest rate of spontaneous regression [238-244], as it has been reported that 10-50% of cutaneous malignant

melanoma cases show partial or complete regression without a treatment [245, 246], and that even 0.23% of the metastatic cases show spontaneous regression [245].

Using very strict criteria, there may not be pure spontaneous regression or remission, because in reality probably no patient will do absolutely nothing for his illness. In our cogitation, spontaneous regression or remission may occur in one of four scenarios: 1) Some patients' self-management or self-treatment towards the neoplasm is actually effective, although their doctors may not realize or appreciate it. 2) The patients may have experienced severe infection, especially a febrile one, since a host of studies have shown that infection or fever may be an effective cancer remedy [236, 247-266], as we have reviewed before [175, 267]. 3) The neoplasm is inauthentic, either because of a misdiagnosis or because it has not yet evolved to an authentic one, as to be expanded upon below. 4) The spontaneous regression or remission is genuine via a currently unclear mechanism, but, after the above three scenarios are culled away, its frequency is much lower than what is reported in the literature.

Ewing once summarized spontaneous regression of tumors and suggested some possible mechanisms. He wrote in 1935 that "...moles and neurofibromas become sclerosed, myomas calcify, chondromas ossify. Very few cancer cells may be found in the bodies of subjects dead of scirrhus carcinomas; gastric cancer may end in linitis plastica; lymphosarcoma is sometimes fatal without leaving demonstrable tumor tissue in the body. Partial removal may prove such an insult to the momentum of growth that ovarian cancer, chorio-adenoma, lymphosarcoma, and some adenomatous tumors may regress after such an experience" [268]. Since malignant tumors keep randomly mutating, theoretically some mutations may be good ones that direct the cells towards differentiation. Conversely, since the most common genetic changes found in tumors are large chromosomal deletions [269, 270], genomic damage that is too severe may lead to the loss of those genes required for cell survival. Moreover, some pernicious mutants may undergo mutation again, back to the wild type or to a better version, which presumably has reverse evolution as its essence [271] and may cause differentiation of the cells. This so-called "back mutation" or "reverse mutation" is occasionally discerned in drosophila [272] as well as in some human genetic diseases [273-276] and in some cancers treated with chemotherapeutic agents [277-279].

Inauthenticity of the tumors described above as the third scenario may also be a reason for spontaneous regression. For instance, hepatomas, and even hepatocellular carcinomas, in women chronically using estrogen-rich oral contraceptives, with regression upon termination of the contraceptive use, were frequently reported in the 1970s-1980s [280-286], which substantiates the human relevance of estrogen-induced hepatomas in rodents reported in the 1950s-1960s [287-289]. The regression upon hormone cessation intimates that estrogen serves as an inducer to incessantly coerce hepatocytes into

neoplastic appearance without really bearing the genetic mutation(s) responsible for immortality and autonomy. As another example, low-grade lymphomas can result from infection by *Helicobacter pylori* (HP). These tumors are basically curable by eradication of the bacteria with antibiotic treatment [290-295], but, if left untreated, some of them will progress and become incurable, as reviewed by Park and Koo [296]. Similarly, Chronic HTLV-I (human T cell lymphotropic virus type I) infection may spawn adult T cell leukemia or lymphoma, but the neoplasm can be well controlled or even cured by antiviral treatment against HTLV-1 [297-299]. To us, these properties of these bacterium- or virus-caused outgrowths resemble those induced in many animal models described above, and thus are not authentically neoplastic at their early time point, although their diagnoses meet pathological criteria for lymphoma or leukemia and they, if left untreated, will eventually evolve to genuine neoplasms. Or, we can take a non-pathological definition of cancer proposed by Robert Axelrod, who majored in political science but became a prominent cancer ecologist [300], that incipient cancer cells might just have been partly transformed, and not yet fully malignant, thus requiring collaboration with each other for survival and for collective presentation of a cancer phenotype [301].

Loss of allegiance to the animal's body is the essence of a cell's immortality and autonomy

Sporadic tumors can only be derived from those cell types that are renewable, i.e. have a lifelong ability to replicate, but not from nonrenewable cell types, i.e. those that have lost their replicative ability in adulthood, such as neurons and cardiac myocytes [132, 302]. We tag those highly renewable cell types as “anabolic” for their great susceptibility to cancers, and those nonrenewable cell types as “catabolic” for their role in the development of type 2 diabetes [303]. The reason for why only renewable cell types can develop to neoplasms is that DNA mutation needs to be perpetuated by at least one round of DNA replication, and to be passed to filial cells via cellular divisions. The permanence becomes possible is because fitness testing of cells is usually conducted after the mutation is made permanently heritable [304]. Even for those renewable cells, it will take about one-fourth to one-third of the lifespan to complete the procedure of sporadic carcinogenesis, which is about 20-30 years for human beings [180], although it could take 50 years by others' estimation [9]. Therefore, the aforesaid tumors induced by 3-methylcholanthrene in just 3-4 weeks [46, 47] cannot be authentically neoplastic, since the lifespan of experimental mice and rats is three years or longer, although their counterparts in the wild live much shorter [12]. Indeed, we have not been aware of any rodent model in which a genuine cancer can be induced in a space less than a few months, except those genetic models in which the genetic manipulation

has already been effective during an embryonic stage, thus mimicking a pediatric (but not a sporadic) carcinogenesis, as to be enlarged upon late.

All cell types in a given human individual have a physiological total number. For instance, the white blood cell count in a normal adult should be somewhere between 4,000-8,000 cells per cubic microliter of blood. For renewable cell types, if the cell number is decreased for some reason, the body will trigger cell proliferation to restore the physiological number, which is usually coined as “regeneration”. On the other hand, if the number is higher than normal, as seen in over-regeneration that often happens following a regeneration procedure, the body will goad some of the cells into apoptosis to avoid cell redundancy. This denotes that all cells in an animal are allegiant to the animal’s body, as we described before [175, 178, 305-308], or “conform with the law of organisms”, as put by Rous in 1941 [40]. This allegiance of all normal cells to the animal’s body as the “law of organisms” allows the animal’s body to require some renewable cells to sacrifice their lives for the body’s ultimate interest. A good example is that white blood cells are put on the frontier by the body to fight against infectious micropathogens and die in the battle, so that the body as a whole can survive [307, 308]. However, sometimes some renewable cells, such as select bone marrow cells, epidermal keratinocytes and mucal cells in the gastric-intestinal tract, have lost their altruism, usually due to acquisition of tumor-driving mutations that make the cells egocentric. These selfish cells want to survive such stress as micropathogen infection, over-regeneration-triggered apoptosis, etc., and become independent of the body, i.e. become autonomous. Reiterated, this disloyalty or loss of loyalty to the animal’s body is the essence of, or the reason for, autonomy of some cells. “Fail to conform with the law of organisms” as said by Rous [40], or “become autonomous” as outlined by Ewing to be the pathological concept of tumor [268], was set as “the signature of a genuine neoplasm” by Borst in 1903 [309] and has, until today, been an salient feature of tumors, including the benign ones.

Leslie Foulds (1902-1974) split growth rate into “the responsive” component and “the intrinsic” component, and the total growth of cells is the sum of the two [132]. He wrote in 1953 that “all cells which can give rise to cancer possess the ability to multiply at a given rate, provided the environmental conditions are constant. They also have the capacity to respond to nutritional and hormonal growth factors, temperature, pH, etc. The intrinsic growth rate of normal cells is in general low; their responsive growth rate is high. The cancerous change goes with acquisition of a greater intrinsic growth rate and diminished responsiveness; the more malignant a cell, the greater the intrinsic and the less the responsive growth” [132]. Translated into today’s language of cancer research, “the responsive growth” is the regenerative type of cell proliferation that is controlled by the animal’s body and dwindles away during

carcinogenesis, whereas “the intrinsic growth” is the autonomous proliferation that is controlled by the cells themselves and is strengthened during carcinogenesis.

Animal models can generally be dichotomized

Animal models established since the 1910s have evolved from using, as the inducer, a single agent, such as Scarlet red or tar, to using a complex regimen or a combination of several manipulations. Nevertheless, we try to split all animal models into two groups, based on whether or not the inducer is a potent genotoxic agent, although there are many intermediate models in which the inducer is a combination of both genotoxic and non-genotoxic agents. In one group, the inducers are potent mutagenic agents or regimens, including genotoxic chemicals, irradiations, and those genetic manipulations that impair DNA repair. In this group of animal models, mutation(s) responsible for the initiation, the first step of carcinogenesis, occur very early. An exemplary one is the Solt-Farber’s “resistant hepatocyte” model of hepatocarcinogenesis in the rat (Fig 1) [310, 311], or our modified version of it in which the promoting agent 2-acetylaminofluorene is routed via gavage instead of by feeding *ad libitum* [312-315]. Carcinogenesis in this group follows a trajectory of “initiation-promotion” or “initiation-promotion-progression”, as detailed by Farber [26-28, 316, 317]. It is clear that the gene(s) and their mutation(s) responsible for initiation are not those responsible for immortality and autonomy. This can be discerned in the Solt-Farber model wherein spontaneous proliferation, which reflects immortality and autonomy, occurs only in the lesions coined by Farber as “phenotype 4” that appear months after the establishment of initiated cells and after the completion of the carcinogenic regimen (Fig 1) [317].

The other group of animal models uses nonmutagenic agents as the inducers, which in the literature are often dubbed as “epigenetic carcinogens or agents” [318-323], “non-genotoxic carcinogens” [322, 323], or “cocarcinogens” [21, 324-326]. In our opinion, carcinogenesis in this group often incepts with promotion, but not with initiation, unlike that in the aforesaid group. This is because in this group, the nongenotoxic inducer cannot cause mutation(s) to establish initiated cells at an early time point, and thus the early proliferative lesions are not of initiated cells. Alternatively, there is some evidence suggesting that initiation might not involve mutations [181, 327-334], which might be the case in this group of animal models. Initiated cells are established much later by stochastically occurring mutation(s) during the relentless proliferation of normal cells induced by the inducer. In our opinion, which still awaits experimental corroboration, lesions turn to be authentically neoplastic, especially malignant, much later in these animal models than in those involving potent mutagens as the inducers.

Why do lesions depend on the inducer and what does it mean?

Although the two groups of animal models described above differ in the potency of initiation, they all provide potent promotion, the essence of which is to drive cell replication, and they all can induce malignant tumors at a high incidence eventually. During promotion, the inducers function as aggressive coercers of the target cells, forcing them to replicate robustly and to manifest neoplastic, often malignant, cellular and histological morphologies with or without aggressive behaviors. In other words, an ominous morphology with or without diabolical behaviors occurs neither because mutation(s) responsible for immortality and autonomy have occurred nor as a repercussion of mutation(s) responsible for neoplastic morphologies and behaviors. It occurs simply under the inducer's duress, and thus the lesion disappears once the duress ends, because the lesion's cells still have allegiance to the animal's body. This remaining allegiance bespeaks the untransformed or non-autonomous nature, categorizes the lesion to hyperplasia, and is the reason for why the lesion's cells regress via apoptosis after the coercion is removed, so that the host organ or tissue does not have cell redundancy, as we have expounded before for what apoptosis really is [175, 178, 189, 305-308, 335]. Actually, we infer that the lesions may manifest a higher rate of apoptosis than their normal surrounding tissues because of their hyperplastic nature, although probably most, if not all, of the inducers may have a function to suppress apoptosis of the hyperplastic cells as part of their coercive mechanism. In an analogy, nice people may commit outrageous crimes if they are coerced, but they may still have good hearts and may become model citizens again once they are unfettered from the duress. However, if a person is incessantly coerced to commit crimes one after another, he may eventually evolve into a real criminal. The inducers prod cells into relentless proliferation while restraining them from committing apoptosis, which together will sooner or later lead to the genetic mutations responsible for immortality and autonomy as well as for neoplastic morphology and behaviors, making the lesions truly neoplastic. In conclusion, judging lawbreakers simply by the crimes they have committed is not completely right, and the reasons behind their criminal activities need to be taken into consideration. Similarly, diagnosis of outgrowths induced in animals should not solely rest on the pathological morphology. This "coercion hypothesis", proposed by us a few years ago [336] and recently [32] on the essence of animal models of carcinogenesis, deserves experimental testing.

Immortality and autonomy may occur early in most human lesions but occur late in animal models

When are immortality and autonomy established during a lengthy tumorigenesis in humans? It is an enthralling brainteaser, so far without a clear answer. For several reasons we infer that they occur at an early time point (Fig 2). First, spontaneous regression of tumors, many of which are diagnosed at early

stages, is rare and thus nearly all tumors are considered immortal. Second, in our pathology service and in the literature, we occasionally encounter very tiny malignant tumors in patients, in most cases found fortuitously. Albeit the tiny tumor had already been surgically removed, some patients still died of its metastases years later, which substantiates the malignant authenticity of the primary tumor. Third, autopsies of humans that died of various causes found about 3-27% of the bodies had an occult pituitary adenoma [337-341] (and DJ Liao's empirical knowledge), and magnetic resonance imaging of normal human volunteers found this tumor in about 10% of normal persons [342]. Similarly, it has also been known since 1934 that a large number of men over 40 years of ages have occult prostate adenomas or adenocarcinomas, although many of the lesions do not develop to clinical cancer before the men die from other reasons [343-345]. Nevertheless, more tangible proofs for the speculative early-establishment of immortality and autonomy are still wanting. In humans, tumor-promoting momentum is much weaker, including the thrust provided by those relatively potent promoters such as chronic viral hepatitis, compared with that provided in various animal models. Therefore, human lesions grow and progress much more slowly, allowing immortality and autonomy to occur much earlier with respect to the size of the lesions, and allowing the neoplastic transformation to occur as the result of some relevant mutation(s), long before the patients feel something wrong and go to see their doctors. This is partly because a lengthier course allows accumulation of more haphazardly-occurring mutations, including the one(s) required for immortality and autonomy, if we accepted the notion that tumors, especially cancers, occur as repercussions of mutations that have cell-autonomous modes of action [17-19, 328, 329, 346-353]. Reiterated, because in a sporadic human carcinogenesis the promoting impetus provided by the ascribed factors such as cigarette smoking or chronic hepatitis is weak, the course is lengthy, which allows mutations to occur in a random and spontaneous manner as additional factors to contribute to the carcinogenesis.

The undeniable fact that immortality and autonomy occur only in an advanced stage in many, if not most or all, animal models announces that few, if at all, animal models established so far can reflect most cases of sporadic carcinogenesis in humans, germane to the time point for the establishment of immortality and autonomy (Fig 2), no matter how much the lesions' morphologies resemble their human counterparts. Fortunately, many animal models still nicely reflect some rare human situations in which immortality and autonomy are established in a late stage. For example, familial colorectal polyps that will sooner or later progress to cancer are developed due to inherited mutations in some genes, like the APC (adenomatous polyposis coli) gene [354-358]. The constant presence of the mutation serves as a durative coercion on colorectal mucal cells, keeping them in an unremitting state of proliferation to form polyps.

These polyps are considered in pathology as premalignant lesions, based on their morphology and on the fact that cancer likely ensues. Notwithstanding, we are curious about whether the polyps would regress if we have a way to correct the mutation. Probably, from the point of immortality and autonomy, the polyps are just “preneoplastic”, an elegant jargon used by Haddow [54] and Rubin [359], or are “precursor lesions”, another good appellation used by Farber [360]. Other good examples of late establishment of immortality and autonomy in human outgrowths include the abovementioned hepatomas and hepatocellular carcinomas caused by long-term consumption of oral contraceptives [280-285], as well as lymphomas or leukemia caused by HP [290-296] or HTLV-1 [297-299], as these lesions at a relative early time point can be cured with removal of the inducer. It has also been suspected that some thyroid tumors may not be immortal and will probably not evolve to authentic neoplasm until the patients die of other causes [361], which dovetails with Forth’s opinion in 1953 [132].

Tissue culture and transplantation once were major methods to determine immortality and autonomy

As aforementioned, even over a century ago whether or not a patient’s tumor was immortal and autonomous had been a concern of, and thus often tested by, surgical pathologists, because they had realized that morphological traits should not be the only criteria, and the tumor’s behavior should also be considered, for an infallible diagnosis of cancer. The tests had been conducted, ever since 1901 [362, 363], mainly by culturing surgically removed tumor tissues in dishes or by transplanting the tissues to animals, the two modern techniques aforesaid. Actually, a technique involving both transplantation and culture is to inoculate tumor cells into a fertile egg and then hatch it [364-374], which is the parentage of some modern chick embryo assays for cancer studies [375-387], such as the chick heart invasion assay [388-393]. The rationale for using tissue culture is that all neoplastic cells in a tumor are immortal and can self-renew to forever maintain the tumor as a “new organism” by incessant cell division. Even after the patient has died, the “organism” can be maintained as cell lines. Indeed, the Hela cell line was established in 1951 from the cervical cancer of the late patient Henrietta Lacks [394], and is still widely used today in biomedical research. Therefore, the essence of this approach is to test tumor cells’ immortality or self-renewal ability. With this regard, in many cases those “transformed” cells that can be reversed back to the normal in cell culture, as mentioned by Harry Rubin and Andrew Rubin [395], may not have reached a truly neoplastic state.

Tumor tissue transplantation to animals, since it was started by Hanau in 1889 [396], has been heavily used in cancer research, as extensively reviewed many decades ago [396-411]. The rationale for using this approach to determine a tumor’s authenticity is to observe the tumor cells’ behaviors, mainly autonomy.

Greene was one of the pioneers in this line of work during the 1940-1960s [179, 412-419]. He elegantly selected the brain or the anterior chamber of an eye of animals as the recipient site. As shown in table 1, transplantations of animal tumors can generally be divided into five different types [179, 413], i.e. 1) autologous transplantation, or transfer back elsewhere in the same animal; 2) homologous I transplantation, or transfer to a tumor-bearing animal of the same species; 3) homologous II transplantation, or transfer to a normal animal of the same species; 4) heterologous I transplantation, or transfer to a tumor-bearing animal of a different species; and 5) heterologous II transplantation, or transfer to a normal animal of a different species. Obviously, transplantation of human tumors can only be done heterologously.

A seminal finding by Greene et al. in the 1940s is that some cancers are not transplantable to normal animals but are transplantable to animals that bear a spontaneous tumor, especially one of the same tissue origin [179, 413]. For instance, the Brown-Pearce rabbit tumor typically does not grow in normal C3H mice but it grows rapidly in those C3H mice bearing spontaneous tumors [414], and Rous chicken sarcoma grows upon subcutaneous transfer to tumor-bearing C3H mice but growths have not been observed in normal C3H mice [179]. These results led Greene to a conclusion that the factors affecting the take of transplanted tumors “are constitutional in distribution and are not localized at the site of the primary growth” [412]. As exceptions, lymphoblastic leukemia and lymphosarcomas are graftable to every normal genetically compatible host but do not produce tumors in the anterior chamber of an eye of an alien host, showing difference from other solid tumors [179]. The difference between normal and tumor-bearing hosts in response to a tumor graft suggests that tumor-bearing animals possess some factors affecting the graft’s survival. A possible interpretation is that the spontaneous tumor preexisting in the host has already suppressed the host’s immune function that is supposed to reject the graft. Studies of these inhibitory effects of the tumor-bearing host on the grafted tumors have later been extended to the studies on the interaction between normal cells and tumor cells, not only in vivo but also in vitro [420-429], as has been reviewed by us [302], by Rubin [430-433], by Aktipis [434, 435] and by Thomas et al [436-438] from different slants. For instance, it has been shown that normal cells suppress the growth of adjacent tumor cells in culture [439] and in skin grafts on mice [440]. Unfortunately, identifying these tumor or host factors has largely been neglected, although it is important because manipulation of these factors may be helpful in curing cancer.

Another trailblazing finding by Greene et al. in the 1940s is that the malignant tumors that have acquired an ability to metastasize are heterologously transplantable, i.e. can be transplanted to host animals of a different species, whereas cancers that are still incapable of metastasizing cannot [416, 418],

as has been tested for many human tumors [413, 416, 418, 419]. Based on these observations, Greene concluded that “cancer is not a sudden transformation of normal cells but, on the contrary, represents the final step in a development process”. He also concluded that only those lesions which can metastasize are fully autonomous and can be regarded as cancers, whereas those which have not yet possessed this ability are still conditionally autonomous and thus should not be regarded as malignancy [416, 418]. Although in pathology textbooks metastasis is not a required criterion for diagnosis of a malignancy, it is the only reliable yardstick to distinguish malignant neoplasms from benign ones [441]. Unfortunately, as a practical matter, clinicians cannot put a hold onto their diagnosis for months or years until they have seen whether or not their patients develop metastatic lesions. Considering that even today, compared with Green’s epoch, in the surgical pathology service we still do not have a simpler or more reliable approach to determine whether a primary tumor removed from a patient has encompassed the ability to metastasize, it is a pity that Greene’s simple but reliable test has not been routinely used in clinical service until now. Probably, a graft into an animal’s eye may be an ethical concern, and thus a constraint, today.

What do the two genetic hits do in carcinogenesis, and is a third hit needed?

The traditional “two-hit” theory [9] may hold true in many animal models, if we accept the century-old concepts that carcinogenesis results from genetic alterations and that cancer cells owe their properties to mutations [430, 442, 443], although the concepts may not necessarily be true and, actually, continue receiving arguments [181, 334, 395]. The first genetic hit is responsible for the establishment of initiated cells that differ from their surrounding uninitiated cells in the response to the promoting environment. In Rubin’s opinion, the cells in the skin papillomas produced in the aforementioned animal models that regress upon withdrawal of the inducer are initiated ones [431]. According to Farber [26-29, 316, 317, 360, 444-449], in most cases promoting agents or regimens cause “mitoinhibition”, i.e. inhibition of mitosis or proliferation, of normal cells, whereas initiated cells are resistant to this inhibitory effect (Fig 1) [186]. Actually, in cell culture, a condition disfavoring cell growth, such as a lower serum concentration or a cell confluence, is more potent in driving neoplastic transformation as well [269]. Therefore, in a promoting environment, probably also in the human [269], only initiated cells can robustly proliferate to form lesions, especially when many of their adjacent normal cells die and the organ or tissue has a strong demand for a compensatory regeneration [186, 187, 450]. This “mitoinhibiton” theory is accordant with the hypothesis of Rozhok and DeGregori that cancer occurs more often in old age because normal cells in the young have a stronger capacity of proliferation, thus being less “mitoinhibited” and providing a weaker promoting impetus, whereas the situation in the old is the opposite [12], making the spontaneously-

occurring initiated cells need to undergo a more robust compensatory proliferation. The molecular mechanisms of promotion via mitoinhibition still remain enshrouded. We extrapolate, sans evidence though, that mitoinhibited normal cells promote proliferation of initiated cells in part via a mechanism identical to that used by senescent cells to promote carcinogenesis of their adjacent cells, since senescence is a state of permanent growth arrest [451-455], i.e. “mitoinhibition”. This mechanism is coined as SASP (senescence-associated secretory phenotype) [456-458], and its effect on carcinogenesis has been extensively reviewed in the literature [452, 459-467].

In our meditation, the second hit is responsible for creating immortality and autonomy, thus establishing neoplastic cells, benign or malignant. This second hit occurs in a later promotion stage of the “initiation-promotion” models or in the progression stage of the “initiation-promotion-progression” models. In sporadic carcinogenesis in humans, initiated cells may also exist, although they are technically difficult to identify. Nevertheless, “preneoplastic” cells in humans may have already experienced the first hit, while “pre-cancerous cells”, which may or may not be benign, may have also experienced the second hit, in our opinion. Genotoxic agents or regimens can quickly establish initiation but have no way to establish immortality and autonomy until a late stage. Indeed, so far there has not been any animal model of sporadic carcinogenesis suggesting an early establishment of immortality and autonomy. While this fact remains to be an enchanting mystery, it insinuates that animal cells put more guards on the genes relevant to immortality and autonomy, compared to the guards on the genes relevant to initiation, meaning that the first hit is easy, but the second is difficult. Actually, so far no exogenous agent, whether a chemical, an irradiation or whatsoever, has been identified that can break through the second defensive line of cells to directly cause mutations to establish immortality and autonomy. A lengthy promotive period in all animal models of sporadic carcinogenesis established so far evinces that this breaking-through can only be made by currently-unknown intrinsic factor(s), although our manipulations can instigate the factor(s). Probably, one day we may identify the relevant genes and in turn find a way to break through this defensive line.

In some carcinogenic procedures wherein a malignancy does not require a benign lesion as a precursor and thus a second hit is sufficient, the mutation(s) responsible for immortality and autonomy may also be responsible for malignant morphologies and behaviors (Fig 3). However, in other animal models and human situations, the mutations for establishing immortality and autonomy may not be the ones for establishing malignant morphologies and behaviors, since benign neoplasms have also experienced the second hit. Therefore, in these situations a third genetic hit may be required to establish malignant morphologies and behaviors (Fig 3). Of course, malignant neoplasms continue evolving via the

fourth, fifth and subsequent hits to be more and more heinous, but this is irrelevant to the issue described here.

An old question: how many mutations are needed to complete a carcinogenesis?

The target or targets of the three genetic hits remain unknown to us. Initiation created by the first hit likely involves only one or several genes, since initiated cells are morphologically indistinguishable from uninitiated ones [27, 29, 317, 468]. Immortality and autonomy created by the second hit may involve only one or several genes as well. This conjecture is based on the observation in our pathology service that many benign tumor cells, such as uterine leiomyoma cells, are quite similar to, basically indistinguishable from, their normal counterparts in cellular morphology. These types of benign tumors can only be diagnosed by their macroscopic and histological traits, and not by their cellular morphology. If mutations in many genes are involved, cellular morphology should change. Moreover, immortalization of a mortal cell to establish a cell line has been proven to be easy, especially in vitro [116, 469, 470]. It is even easier when the cell has a small rodent parentage, presumably because these rodent cells have their telomerase constantly “on” and have only a single barrier to immortalization controlled by the RB (retinoblastoma protein) pathway [469, 471, 472]. For instance, targeting both the p16ink4 and c-myc genes can immortalize human mammary epithelial cells [473], and the IgEGF and SV40T bi-transgenes can immortalize mouse cells [474]. Actually, even just knocking out the p53 gene alone can immortalize mouse hepatocytes [474]. The third hit, if it is needed, may occur to more genes compared to the first two, since cellular and histological morphologies and behaviors can vary greatly among different cancer cases. Nevertheless, the sum of the three hits may still be congruent with the estimation by Armitage and Doll in 1954 [19] and by Vogestein in 1993 [475] that carcinogenesis requires only six or seven mutations. Liquid cancers such as leukemia may require even fewer mutations and thus may be relatively easier to cure, generally speaking, as to be explained later. A caveat is that different hits in different cancer cases may involve different genes, making the sum of “initiator genes”, “immortalizer genes” or “malignant morphology responsible genes” large, and the sum of all three even larger. This feature, dubbed as cancer heterogeneity, is a major reason why such a large number of genes have been found to be cancer-relevant.

Are immortality and autonomy independent of each other?

In 1983 Land et al showed that a mutant ras gene could transform embryonic fibroblasts without immortalizing the cells, and immortalization of the transformed cells could later be achieved by an ectopic expression of the c-myc or a viral oncogene [476]. Concomitant expression of the CDK4 gene and a mutant

ras can transform cells, as evidenced by the facts that the cells can form colonies in soft agar and develop into invasive tumors in animals; however, the “transformed” cells remain mortal as evidenced by their limited passages in culture [477]. These and other studies show that transformation and immortality can be extricated from each other. However, this segregation is not discerned in some tests with traditional approaches involving chemical carcinogens, such as the “Syrian hamster embryo cell transformation assay” (the so-called SHE assay) [478, 479], and conflicts with the opinion of Newbold et al that immortality is an early and prerequisite step of transformation [480-484]. This discrepancy among different studies with different approaches makes the reliability of the criteria for in vitro transformation [116, 485] dubious to us, because in human situations immortality and autonomy are inextricable from each other. Moreover, in vitro transformation is often reversible [395], which also collides with the aforementioned fact that human cancers rarely regress spontaneously. Probably, a gene like a mutant ras can coerce cells into behavior like being transformed in vitro without really transforming them by mutating the relevant genes, just like variegated inducers of carcinogenesis in animal models described earlier in this essay, although such mutations and true transformation are corollaries of the incessant coercion. In other words, ectopic expression of an oncogene or some other in vitro genetic manipulation may just function as a coercer to the cells, i.e. resemble the inducers in animal models that coerce the target cells in the animals into manifesting neoplastic morphology. Moreover, although malignant tumor cells dissected from patients or animals may indeed form colonies in soft agar, which is an in vitro test for transformation established by Freedman in 1974 [485] and has been widely-used, the other way around may not always be correct, meaning that forming colonies in soft agar may not necessarily be a token of a transformed, i.e. malignant, state. In our opinion, if an in vitro study shows that immortality and autonomy are extricable, more tenable proofs for the neoplastic state may be needed. This is to say that once the cells are shown to be capable of forming colonies in soft agar, they need to be tested for immortality before we can announce that they have been transformed. Unfortunately, few published studies of in vitro transformation show this additional evidence.

We still lack a good strategy to determine molecular pathways leading normal cells to cancers

Not only the genes mediating in the three genetic hits described above, but also the molecular pathways regulated by these genes, remain unknown. One of the reasons is that we have been encountering a logical plight for decades regarding our research strategies and approaches, as repeatedly pointed out by us before [32, 175, 336]: The results from the approaches we used, such as genetic engineering, can only tell us that certain manipulations or alterations, like concomitant overexpression of

the c-myc and a k-ras mutant, and the ensuing cascade of molecular changes, can cause cellular immortality and autonomy, i.e. neoplastic transformation. However, we still do not know whether cells in humans or in untreated animals spontaneously develop to neoplasms really via these cascades of changes. In an analogy, we have built the highway Interstate-95 (I-95) and know that Mr. Trump can go from New York City (NYC) to Washington DC by taking it, but we do not actually know whether this is indeed the path, but not others, he took. If we cannot come up with a novel strategy to solve this tribulation, our attempt to learn why and how some cells in humans become neoplastic will continue to be prodigal financially and in effort, because we will continue identifying (more correctly, creating) many more molecular pathways leading normal cells to neoplasms, besides the many pathways already known [486], while remaining unable to hold any particular pathway(s) accountable for sporadic carcinogenesis in humans. Restated, we are creating, but not identifying or discovering, pathways by such as creating otherwise non-existing transgenic mice, and assume that these man-made paths are the carcinogenic procedures occurring in patients' bodies. In another analogy, we already have many paths leading from NYC to DC but will endlessly build many more while remaining unable to know which path(s) Mr. Trump took or will take. Probably, we have been upending things or putting the cart before the horse in our research.

Many animal models are overpraised, due to neglect of the immortality issue

As aforementioned, evidence for immortality and autonomy has not been provided for the lesions resulting from the vast majority of genetically-manipulated animal models of carcinogenesis. This is an uncomfortable but undeniable flaw of the relevant studies, although we should somewhat be content with the enormous amount of information provided by these models on the functions and underlying mechanisms of the genes manipulated. It is possible that many peers have already harvested the "cancers" from their host animals for the mechanistic analyses before the lesions, probably large in size, have evolved to genuine neoplasms. This slip is made probably because the surgical pathologists involved made their diagnoses solely based on their experience in human tumor pathology, unlike their predecessors such as Greene and Furth who were familiar with the inducer-dependency of animals' lesions. Actually, although human tumor pathology is generally reliable in reflecting the neoplasms' prognosis, as proven by over a century of clinical practice, the lesion's behavior should be taken as an additional criterion for diagnosis. Greene had pointed it out in 1947 by saying that "the problem of cancer is primarily a problem of behavior. A pathologist who examines tumor tissue under the microscope may observe significant details of form and structure, but he can never determine its malignancy from its appearance alone; only

by its behavior in the living body can malignant tissue be unmistakably identified. Of two tumors with cells that look exactly alike, one may remain static or even disappear while the other inexorably spreads and kills the patient. Unfortunately many kinds and conditions of tissue which are not malignant bear a remarkable resemblance to cancer” [413]. This extra criterion is especially needed in diagnosis of outgrowths induced in animals because they differ greatly from their human counterparts in two particular aspects. First, immortality and autonomy may not have been established when the lesions are harvested from the animals. Therefore, these animal “cancers”, even large in size, can be cured easily, simply by withdrawal of the inducer or by chirurgic removal, whereas most human cancers are not curable, at least not so easily. Second, “malignant” tumors in many animal models, such as mammary tumors from the MMTV-c-myc transgenic mice we have used [184, 188], do not metastasize within the lifespan of the animals, which starkly contrasts most human cancers that, if untreated, will metastasize. Some tumors from animal models can metastasize, but the metastases may still be inducer-dependent. Although this speculation has not been broadly tested, examples have been described herein, like the abdominal metastases from estrogen-induced renal tumors in hamsters [134-136] (and also DJ Liao’s own experience), and the lung metastases from TSH-dependent thyroid tumors [132, 155, 159]. In a nutshell, the mortality and non-autonomy, the inducer-dependency, and the inability to metastasize are telltale evidence that many animal cancers are easily curable and thus are disarming, which starkly contrasts with most human cancers. An overarching theme in the second phase of carcinogenesis research had already been to determine immortality and autonomy of the lesions induced in animals, which, unfortunately, had been abandoned about three decades ago when the research entered into its third incarnation by shifting from chemical-induced to genetically-manipulated carcinogenesis. Therefore, there is an exigency to reincarnate this task.

Neglecting immortality causes confusion on aging-caused cell death in outgrowths

Normal animal cells undergo aging and eventually die of it [303, 307, 487-497], and so do cells in overt lesions from many animal models, which of course occurs more often in morphologically-benign than in morphologically-malignant ones [492, 494-496]. We define cell death via aging as “senescent death” (SD) [177], because normal cells have a lifespan [480-484, 498] and because, ever since it was first observed in 1965, this senescent phenomenon has been immediately linked to aging [499, 500]. Indeed, a host of studies have shown that aging and senescence are highly interrelated [12, 452-455, 500-509], although senescence itself is defined as a permanent growth arrest that does not necessarily lead to death of the cell [451-455]. This type of cell death has established “cancer cell senescence” as a popular new

research bailiwick [307], although many relevant studies do not involve lesions from animals but, peculiarly, use human cancer cell lines that are immortal. Because its essence is “dies from aging”, the “cancer cell senescence” concept is illogical and against the immortality and autonomy criteria of neoplastic cells, as it connotes that immortal cells still undergo aging and eventually die of it. In our logic, neoplastic cells are immortal and thus cannot undergo senescence and eventually die from it, whereas cells that can undergo senescence and die from it cannot be regarded as neoplastic no matter how much their morphology and behavior resemble those of neoplastic cells. In all those lesions that are morphologically malignant and even metastatic but are still inducer-dependent, such as the TSH-dependent thyroid tumors and their metastases [132, 148, 159], cells may undergo aging and die via the SD mechanism because they have not yet been authentically neoplastic.

Neglecting immortality causes confusion on cancer stem cells

In the past 20 years or so, “cancer stem cell” (CSC) has become a new popular research province, although the definition of CSC remains erratic or, in the words of Dumont et al [510, 511], fuzzy and evolving. Indeed, its concept in the literature has never been lucid in distinguishing CSCs on the one hand from normal stem cells and on the other hand from ordinary cancer cells [512]. The initial CSC concept was derived from some observations in the 1990s that many leukemia cells showed different extents of differentiation, leading to the theory that neoplasms might be derived from transformed undifferentiated pluripotent stem cells [513-515]. This concept annotates CSCs as those organ- or tissue-specific stem cells that somehow go awry, presumably due to mutation(s), and evolve to cancers [487, 516-518], although sometimes they are also called “transformed stem cells” [519]. According to a denomination of carcinogenic mechanisms, in a renewable cell type a stem cell that has gone awry may stop differentiation during an embryonic stage or during a tissue regenerative procedure, and continues proliferating to form a neoplasm, as we described before [302, 336]. Actually, this “stop-differentiation” mechanism is presumably a reason why nonrenewable cell types still develop childhood neoplasms: Pediatric tumorigenesis, instigated by such reasons as in utero exposure to a carcinogen [520], had already incepted during an embryonic stage when the cells still had their replicative ability [302], and might occur via a stop-differentiation mechanism. For this reason, we have suggested that molecular biologists should be wary of using those DNA elements that are activated during an embryonic stage [185, 191], such as the Mist-1 promoter [521, 522], as the promoters to drive transgenes, because the resulting transgenic animal models of carcinogenesis mimic only formation of childhood cancer via this “stop-differentiation” mechanism, whereas most cancers in humans are sporadic [185, 191].

With the number of CSC studies soaring, CSCs seem to have been redefined. In Clarke's words, "...a subset of cancer cells within some tumors, the so-called cancer stem cells, may drive the growth and metastasis of these tumors" [515]. More detailed by Chiodi, "in many types of cancers a subset of cells shows peculiar characteristics, such as the ability to induce tumors when engrafted into host animals, self-renew and being immortal, and give rise to a differentiated progeny. These cells have been defined as cancer stem cells (CSCs) or tumor initiating cells" [523]. Similarly, in the words of Weinberg's group, "the CSC hypothesis posits the existence of subpopulations of neoplastic cells within a tumor that exhibit an elevated ability to seed new tumors upon experimental implantation in appropriate animal hosts" [524]. By this definition, which is widely used in many publications [321, 516, 525-529], CSC is a diminutive group of cancer cells in the cancer mass, but not the single normal stem cell that undergoes a carcinogenic procedure to evolve to a cancer. Indeed, Weinberg's group clearly separates these two types of stem cells by saying "evidence is accumulating that both normal and fully neoplastic cell populations harbor subpopulations of stem cells (SC) that can both self-renew and spawn more differentiated progeny" [530]. This definition hints slyly that except a tiny fraction of cells in a cancer mass, the vast majority of cancer cells are not immortal and are not able to self-renew, which is obviously against the definition of neoplasm in all pathology textbooks published since the 1900s.

Benign and malignant cells relentlessly undergo symmetrical binary fission, just like bacterial cells that unremittingly divide without a clear distinction between somatic line and germ-line, to maintain their strains, although some pundits consider that bacterial cells also undergo asymmetrical division and thus undergo aging as well [531-533], probably in part for maintaining their vitality [534]. Moreover, malignant cells are highly plastic and can differentiate to various cell types [513, 535]. For instance, quite different types of cancer of epithelial origin [536-546], and even pre-cancerous lesions [547], manifest bone histology, or osseous metaplasia in pathological phraseology. Therefore, a strong pluripotent ability should not be used to dichotomize cancer cells to CSC and non-CSC groups. It is true that in cancers many cells die at a much higher rate than their surrounding normal counterparts due to various stressors, such as insufficient oxygen or nutrient nourishment or overly severe genetic damage [349, 350, 548]. The opposite is also true that in a cancer mass some cells' ability of self-renewal via symmetrical division is much more potent than that of the others. Actually, many ancient studies, started by Furth and Kahn in 1937 [549], have already shown that single cancer cells in late progression stages were highly transplantable and could grow rapidly in recipient animals [550-554], as extensively reviewed by George Klein six decades ago [397]. However, these quantitative differences simply reflect the well-known heterogeneity of malignant cells [433, 555, 556], which is largely ascribed to the stemness of some cells

[557] or great genetic variation among most cells [548, 558], and should not be used to split cells into CSC and non-CSC groups either. More critically, “ability of self-renewal vs inability of self-renewal” is actually “immortality vs mortality”, which is a black-white demarcation between neoplasms and non-neoplasms and thus should not be used again as the demarcation between CSCs and non-CSCs. We proffer that, since the above-defined CSCs differ from other cancer cells only quantitatively in such as the competency of self-renewal, metastasis, therapy-resistance, etc., clear quantitative parameters in these vicious behaviors should be established to separate those highly-competent cells, tagged as CSCs or not, from those less-competent counterparts, just like the establishment of the normal ranges for blood pressure, blood sugar, etc., as criteria for diagnosis. Identification of biomarkers for these cells, as performed by many cancer students now, is part of this line of work. Once these quantitative parameters have been set as criteria, these heinous cells can be more easily identified and studied for their behaviors in the aspects of chemotherapy, metastasis, patients’ prognosis, etc., without calling them CSCs. For example, it is unnecessary to call “CD44(+)/CD24(-) and ALDH1(+)” breast cancer cells as CSC [559-561], since “CD44(+)/CD24(-) and ALDH1(+)” defines them more specifically and clearly than the equivocal “CSC”.

It needs to be mentioned that many other studies prefer not to provide a pellucid CSC definition, such as the one by Lapinska et al [562], but, instead, to sway between the two different definitions described above. A few others, such as Chaffer and Weinberg [530], consider that CSCs may be derived from tissue-specific normal stem cells in a stepwise manner, which elegantly accommodates the second CSC definition into the first one described above but still indirectly suggests that only CSCs, and not the vast majority of cancer cells in the lump, can self-renew, and thus still denies immortality as an indispensable criterion for a neoplastic state.

Where lies the demarcation between benign and malignant neoplasms?

Not only have immortality and autonomy as a lucid demarcation between non-neoplastic and neoplastic cells been neglected by many of today’s cancer students, including some pundits, but also the demarcation between benign and malignant neoplasms has often been blurred. For instance, most of “cancer hallmarks” described by Hanahan and Weinberg [563, 564] are actually marks of tumors, including the benign ones, but not of cancer, as pointed out first by Lazebnik [441] and later by us [336]. Actually, Blagosklonny’s comment is more frank and straightforward: “...hallmarks can be observed without cancer” [565]. In our rumination, a genetic criterion to segregate the two is: Benign neoplasms do not bear mutation(s) at mutator gene(s), i.e. the gene(s) some alteration(s) of which allow mutations to occur to other genes. In contrast, malignant neoplasms bear mutation(s) at the mutator gene(s), and thus

accumulation of more mutations in more genes is a corollary of the unremitting cell replication, which, in turn, is the genetic basis for tumor cells to continue to progress to more malignant states [302]. Of course, benign neoplastic cells are also immortal and thus keep replicating, which increases the chance for new mutations to occur. Once new mutations occur to mutator gene(s), the benign neoplasm gets a chance to progress to malignancy, which is a reason for why benign neoplasms are at risk for progression.

Malignant neoplasms have three behaviors distinguishable from their benign counterparts, i.e. 1) encroachment into their normal surrounding tissue, which can be considered as local metastases, 2) consumption of their normal surrounding tissue, partly via cannibalism at the cellular level [566-570], and 3) metastasis to distant body sites. However, none of these three is unique to malignant cells, as some normal cells have these behaviors as well [302, 571] and progression of solid tumors does not always follow the stepwise procedure of “growth, invasion and then metastasis” [302, 572]: Trophoblasts make inroads into the uterine wall to establish gestation and may even encroach into the blood vessels and home in on the lungs of the mother and many organs of the newborn [572]; the function of osteoclasts is to eat up bone tissue; and many bone-marrow-derived or thymus-derived cells can enter into the blood or lymphatic circulation and home in on almost all body sites. Probably because these normal cells already possess the ability to migrate to and home in on most body sites, i.e. to “metastasize”, in all pathology textbooks all the neoplasms of the bone-marrow and lymphatic origins, without exception, are classified as malignancy. Actually, this property seems to have its good aspect, as liquid cancers may bear fewer mutations and be easier to cure than most solid cancers, generally speaking, because they inherit from their parental normal cells the ability to metastasize, i.e. to migrate to and home in on most body sites, and thus do not need additional mutation(s) for acquiring this ability. Bearing these bits of knowledge in mind, many “surprising findings” in animal models are actually not so surprising, such as the observations that epithelial cells can be evasive and disseminating and can enter into the bloodstream before they form primary tumors [573], that cancer cells can enter into the blood circulation before invading adjacent stroma [574], and that mammary epithelial cells can be manipulated to metastasize and colonize in the lungs before they are malignantly transformed [575, 576].

Concluding remarks

Over a century ago, pathologists had set immortality and autonomy as indispensable criteria for neoplasms, including the benign ones. It has been known ever since the 1910s that many, if not all or most, outgrowths resulting from experimental animals are inducer-dependent until very late stages. Unfortunately, indubitable evidence for the immortality and autonomy has not been provided for the

lesions resulting from most genetically manipulated animal models of carcinogenesis at the time when the lesions are collected from the animals. Although these lesions have indeed provided us with sheer amount of information on the functions and underlying mechanisms of the manipulated genes, especially on the aspects of cell proliferation and cell death, the lack of this needful evidence still makes us apprehensive, as much confusion on the behaviors of these animal lesions may be caused by their neoplastic inauthenticity. Furth's opinion should be taken that autonomy has two categories: "conditional" and "full" [132]. "Conditional" should be confined within the physiological range with a physiological factor as the conditioner. For example, breast and prostate cancers are genuine, although castration, which decreases the corresponding hormone to a level below the physiological one, can cause their regression. However, the mammary and prostate tumors in experimental animals have not yet become authentic if they still require a continuous hormone treatment to maintain a pharmacological or toxicological level of the hormone for their sustenance. "Full autonomy" means that sustainment of the tumors does not require a physiological factor as a conditioner. For instance, breast and prostate cancers can sustain and continue to progress after castration. Since achievements obtained many decades ago in the second incarnation of carcinogenesis research had already set immortality and autonomy as cardinal criteria for neoplasms and had identified initiation, promotion and progression stages of carcinogenesis, we should, with numerous genetic models created, continue on identifying the genomic bases for immortality and autonomy and for initiation, promotion and progression. Future carcinogenesis studies need to embark on the following brainteasers: 1) What is the genomic basis for immortality or autonomy? 2) What is the genomic basis for initiation, promotion or progression, or for the first, second or third genetic hit outlined herein? 3) How are immortality and autonomy related to initiation, promotion and progression? 4) Which are good animal models for us to use in tackling the above three tasks? Concerns raised in this essay are obviously provocative but deserve reconsideration by cancer research gurus, especially those at the pinnacle.

Declarations

Ethics approval and consent to participate: not applicable.

Consent for publication: All authors agree on the publication.

Availability of data and material: This is a review and no unpublished data and materials are involved.

Competing interests: Authors declare no interest conflicts.

Funding: This work was supported by a grant from the Chinese Natural Science Foundation to D. Joshua Liao (grant No. 81660501).

Authors' contributions: XD drafted the manuscript. ML, HZ, and LC prepared the table and figures, participated in discussion and revised the manuscript. LZ revised the manuscript and performed English editing. QJ and DZ participated in discussions. NX, CW and DJL formulated the concepts. DJL finalized the manuscript.

Acknowledgements: We would like to thank Dr. Fred Bogott at Austin Medical Center, Mayo Clinic in Austin, Minnesota, USA, for his excellent English editing of this manuscript.

Abbreviations

APC, adenomatous polyposis coli; ACI, August strain crossed with Copenhagen strain; CSC, cancer stem cell; DC, Washington DC; DMBA, 7,12-dimethylbenz(a)anthracene; HTLV-1, human T cell lymphotropic virus type I; HP, helicobacter pylori; MMTV, mouse mammary tumor virus long terminal repeat; RB, retinoblastoma protein; NYC, New York City, SASP, senescence-associated secretory phenotype; SD, senescent death; SHE, Syrian hamster embryo cell transformation assay; TSH, thyroid stimulating hormone.

References

1. Faguet, G. B. A brief history of cancer: age-old milestones underlying our current knowledge database. *Int. J Cancer* **2015**, 136:2022-2036.
2. Pott P (1775). Chirurgical observations relative to the cataract, the polypus of the nose, the cancer of the scrotum, the different kinds of ruptures, and the mortification of the toes and feet. *Natl. Cancer Inst. Monogr.* (Reprinted) **1963**, 10:7-13.
3. Haddow, A. Neoplastic diseases (cancer). *Annu. Rev. Med.* **1955**, 6:153-186.
4. Schmidt, A.; Weber, O. F. In memoriam of Rudolf virchow: a historical retrospective including aspects of inflammation, infection and neoplasia. *Contrib. Microbiol.* **2006**, 13:1-15.
5. Cuddihy, J. Rudolf Ludwig Carl Virchow. *Cancer Cells* **1991**, 3:110-112.
6. Morton, L. T. Rudolf Ludwig Carl Virchow (1821-1902): bibliography. *J Med. Biogr.* **1993**, 1:46-47.
7. Wullstein, H. L.; Hellmer, L. Rudolf Ludwig Carl Virchow (1821 to 1902). *Arch. Otolaryngol.* **1970**, 92:299-301.
8. STEWART, H. L. The cancer investigator. *Cancer Res.* **1959**, 19:804-818.
9. Loeb, L. A.; Harris, C. C. Advances in chemical carcinogenesis: a historical review and prospective. *Cancer Res.* **2008**, 68:6863-6872.
10. NORDLING, C. O. Evidence regarding the multiple mutation theory of the cancer-inducing mechanism. *Acta Genet. Stat. Med.* **1955**, 5:93-104.
11. NORDLING, C. O. A new theory on cancer-inducing mechanism. *Br. J Cancer* **1953**, 7:68-72.
12. Rozhok, A. I.; DeGregori, J. The evolution of lifespan and age-dependent cancer risk. *Trends Cancer* **2016**, 2:552-560.
13. Nowell P., H. D. A minute chromosome in human chronic granulocytic leukemia. *Science* **1960**, 132:1497.
14. Nowell, P. C.; HUNGERFORD, D. A. Chromosome studies on normal and leukemic human leukocytes. *J. Natl. Cancer Inst.* **1960**, 25:85-109.
15. Nowell, P. C. The minute chromosome (Ph1) in chronic granulocytic leukemia. *Blut* **1962**, 8:65-66.

16. Nowell, P. C. The clonal evolution of tumor cell populations. *Science* **1976**, 194:23-28.
17. Armitage, P.; Doll, R. The age distribution of cancer and a multi-stage theory of carcinogenesis. 1954. *Int. J Epidemiol.* **2004**, 33:1174-1179.
18. Armitage, P.; Doll, R. A two-stage theory of carcinogenesis in relation to the age distribution of human cancer. *Br. J Cancer* **1957**, 11:161-169.
19. Armitage, P.; Doll, R. The age distribution of cancer and a multi-stage theory of carcinogenesis. *Br. J Cancer* **1954**, 8:1-12.
20. Berenblum, I.; SHUBIK, P. A new, quantitative, approach to the study of the stages of chemical carcinogenesis in the mouse's skin. *Br. J Cancer* **1947**, 1:383-391.
21. Berenblum, I. Cocarcinogenesis. *Br. Med. Bull.* **1947**, 4:343-345.
22. Berenblum, I.; TRAININ, N. Possible two-stage mechanism in experimental leukemogenesis. *Science* **1960**, 132:40-41.
23. Berenblum, I. Established principles and unresolved problems in carcinogenesis. *J Natl. Cancer Inst.* **1978**, 60:723-726.
24. Berenblum, I. New ideas on the biological stages of carcinogenesis. *Acta Unio. Int. Contra. Cancrum.* **1954**, 10:21-24.
25. FRIEDEWALD, W. F.; Rous, P. THE INITIATING AND PROMOTING ELEMENTS IN TUMOR PRODUCTION : AN ANALYSIS OF THE EFFECTS OF TAR, BENZOPYRENE, AND METHYLCHOLANTHRENE ON RABBIT SKIN. *J Exp. Med.* **1944**, 80:101-126.
26. Farber, E. Carcinogenesis--cellular evolution as a unifying thread: Presidential address. *Cancer Res.* **1973**, 33:2537-2550.
27. Farber, E.; Cameron, R. The sequential analysis of cancer development. *Adv. Cancer Res.* **1980**, 31:125-226.
28. Farber, E. Cancer development and its natural history. A cancer prevention perspective. *Cancer* **1988**, 62:1676-1679.
29. Farber, E. Hepatocyte proliferation in stepwise development of experimental liver cell cancer. *Dig. Dis. Sci.* **1991**, 36:973-978.
30. FURTH, J.; TULLIS, J. L. Carcinogenesis by radioactive substances. *Cancer Res.* **1956**, 16:5-21.
31. Fais, S. A nonmainstream approach against cancer. *J Enzyme Inhib. Med. Chem.* **2016**, 31:882-889.
32. He, Y.; Yuan, C.; Chen, L.; Liu, Y.; Zhou, H.; Xu, N.; Liao, D. J. While it is not deliberate, much of today's biomedical research contains logical and technical flaws, showing a need for corrective action. *Int. J Med. Sci.* **2018**, 15:309-322.
33. Davis, J. S. III. The Effect of Scarlet Red, in Various Combinations, upon the Epitheliation of Granulating Surfaces. *Ann. Surg.* **1910**, 51:40-51.
34. Davis, J. S. X. A Further Note on the Clinical Use of Scarlet Red and its Component, Amidoazotoluol, in Stimulating the Epitheliation of Granulating Surfaces. *Ann. Surg.* **1911**, 53:702-719.
35. VASILIEV, J. M.; CHEUNG, A. B. Evolution of epithelial proliferation induced by scarlet red in the skin of normal and carcinogen-treated rabbits. *Br. J. Cancer* **1962**, 16:238-245.
36. Katsusaburo Yamagiwa (1863-1930). *CA Cancer J Clin.* **1977**, 27:172-173.
37. The 90-year anniversary of Katsusaburo Yamagiwa's carcinogenesis. Proceedings of the 1st International Awaji Liver Symposium. December 1, 2005. Awaji Island, Japan. *Oncology* **2007**, 72 Suppl 1:1-140.
38. Yamagiwa, K.; Ichikawa, K. Experimental study of the pathogenesis of carcinoma. *CA Cancer J. Clin.* **1977**, 27:174-181.
39. Fujiki, H. Gist of Dr. Katsusaburo Yamagiwa's papers entitled "Experimental study on the pathogenesis of epithelial tumors" (I to VI reports). *Cancer Sci.* **2014**, 105:143-149.
40. Rous, P.; Kidd, J. G. CONDITIONAL NEOPLASMS AND SUBTHRESHOLD NEOPLASTIC STATES : A STUDY OF THE TAR TUMORS OF RABBITS. *J. Exp. Med.* **1941**, 73:365-390.

41. FRIEDEWALD, W. F.; Rous, P. The pathogenesis of deferred cancer; a study of the after-effects of methylcholanthrene upon rabbit skin. *J Exp. Med.* **1950**, 91:459-484.
42. Haslam, S. Z.; Bern, H. A. Histopathogenesis of 7,12-dimethylbenz(a)anthracene-induced rat mammary tumors. *Proc. Natl. Acad. Sci. U. S. A* **1977**, 74:4020-4024.
43. McGuire, W. L.; Chamness, G. C.; Costlow, M. E.; Shepherd, R. E. Hormone dependence in breast cancer. *Metabolism* **1974**, 23:75-100.
44. Mobbs, B. G. Uptake of (3H)oestradiol by dimethylbenzanthracene-induced rat mammary tumours regressing spontaneously or after ovariectomy. *J. Endocrinol.* **1969**, 44:463-464.
45. YOUNG, S.; COWAN, D. M. Spontaneous regression of induced mammary tumours in rats. *Br. J. Cancer* **1963**, 17:85-89.
46. Huggins, C.; BRIZIARELLI, G.; SUTTON, H., Jr. Rapid induction of mammary carcinoma in the rat and the influence of hormones on the tumors. *J. Exp. Med.* **1959**, 109:25-42.
47. Mider, G. B.; Morton, J. J. Skin tumors following a single application of methylcholanthrene in C57 brown mice. *Am. J. Pathol.* **1939**, 15:299-302.
48. Cabot, S.; Shear, N.; Shear, M. J.; Perrault, A. Studies in carcinogenesis: XI. Development of skin tumors in mice painted with 3:4-benzpyrene and creosote oil fractions. *Am. J. Pathol.* **1940**, 16:301-312.
49. RUSCH, H. P. A theory on carcinogenesis. *Med. Bull. (Ann. Arbor)* **1956**, 22:501-516.
50. RUSCH, H. P. Carcinogenesis; a facet of living processes. *Cancer Res.* **1954**, 14:407-417.
51. RUSCH, H. P.; KLINE, B. E. Influence of interrupted carcinogenic treatment on tumor formation. *Proc. Soc. Exp. Biol. Med.* **1948**, 69:90-95.
52. RUSCH, H. P.; KLINE, B. E. Further evidence for successive stages in the formation of neoplasms. *Arch. Pathol. (Chic.)* **1946**, 42:445-454.
53. RUSCH, H. P. Stages in cancer research. *Tex. Rep. Biol. Med.* **1950**, 8:207-214.
54. Haddow, A. Mode of action of chemical carcinogens. *Br. Med. Bull.* **1947**, 4:331-342.
55. Anderson, E.; CLIFTON, K. H.; FURTH, J.; GADSDEN, E. L. Autonomous mammatropic pituitary tumors in mice; their somatotropic features and responsiveness to estrogens. *Cancer Res.* **1956**, 16:600-607.
56. Colletti, J. A.; Leland-Wavrin, K. M.; Kurz, S. G.; Hickman, M. P.; Seiler, N. L.; Samanas, N. B.; Eckert, Q. A.; Dennison, K. L.; Ding, L.; Schaffer, B. S.; Shull, J. D. Validation of six genetic determinants of susceptibility to estrogen-induced mammary cancer in the rat and assessment of their relevance to breast cancer risk in humans. *G3. (Bethesda.)* **2014**, 4:1385-1394.
57. Dennison, K. L.; Samanas, N. B.; Harenda, Q. E.; Hickman, M. P.; Seiler, N. L.; Ding, L.; Shull, J. D. Development and characterization of a novel rat model of estrogen-induced mammary cancer. *Endocr. Relat Cancer* **2015**, 22:239-248.
58. DUNNING, W. F.; CURTIS, M. R.; SEGALOFF, A. Strain differences in response to diethylstilbestrol and the induction of mammary gland, adrenal and bladder cancer in the rat. *J Mich. State Med Soc.* **1948**, 47:305-315.
59. DUNNING, W. F.; CURTIS, M. R.; MADSEN, M. E. Diethylstilbestrol-induced mammary gland and bladder cancer in reciprocal F1 hybrids between two inbred lines of rats. *Acta Unio. Int. Contra. Cancrum.* **1951**, 7:238-245.
60. DUNNING, W. F.; CURTIS, M. R.; SEGALOFF, A. Strain differences in response to estrone and the induction of mammary gland, adrenal, and bladder cancer in rats. *Cancer Res.* **1953**, 13:147-152.
61. FELS, E. [Pituitary tumors produced by a synthetic estrogen]. *Rev. Soc. Argent Biol.* **1950**, 26:38-43.
62. GARDNER, W. U.; Strong, L. C. Strain-limited Development of Tumors of the Pituitary Gland in Mice Receiving Estrogens. *Yale J Biol. Med* **1940**, 12:543-548.
63. GARDNER, W. U. Studies on steroid hormones in experimental carcinogenesis. *Recent Prog. Horm. Res.* **1947**, 1:217-259.
64. GARDNER, W. U. Steroid hormones in the induction of cancer. *Cancer Res.* **1947**, 7:37-52.

65. GARDNER, W. U. Studies on ovarian and pituitary tumorigenesis. *J Natl. Cancer Inst.* **1954**, 15:693-709.
66. RICHARDSON, F. L. Incidence of mammary and pituitary tumors in hybrid mice treated with stilbestrol for varying periods. *J Natl. Cancer Inst.* **1957**, 18:813-829.
67. Sabatino, M. E.; Petiti, J. P.; Sosa, L., V; Perez, P. A.; Gutierrez, S.; Leimgruber, C.; Latini, A.; Torres, A. I.; De Paul, A. L. Evidence of cellular senescence during the development of estrogen-induced pituitary tumors. *Endocr. Relat Cancer* **2015**, 22:299-317.
68. TWOMBLY, G. H.; MEISEL, D.; STOUT, A. P. Leydig-cell tumors induced experimentally in the rat. *Cancer* **1949**, 2:884-892.
69. Zondek, B. Hypophyseal tumors induced by estrogenic hormone. *Am. J Cancer* 1938, 33:555-559.
70. Zondek, B. Oestrogens and tumour genesis. *Acta radiol.* **1947**, 28:433-450.
71. FERRIGNO, M.; GARDNER, W. U. Unusual neoplastic lesions of the uterine horns of estrogen-treated mice. *J Natl. Cancer Inst.* **1956**, 17:601-613.
72. Schaffer, B. S.; Lachel, C. M.; Pennington, K. L.; Murrin, C. R.; Strecker, T. E.; Tochacek, M.; Gould, K. A.; Meza, J. L.; McComb, R. D.; Shull, J. D. Genetic bases of estrogen-induced tumorigenesis in the rat: mapping of loci controlling susceptibility to mammary cancer in a Brown Norway x ACI intercross. *Cancer Res.* **2006**, 66:7793-7800.
73. GARDNER, W. U. Sensitivity of the vagina to estrogen: genetic and transmitted differences. *Ann. N. Y. Acad. Sci.* **1959**, 83:145-159.
74. GARDNER, W. U. Experimental induction of uterine cervical and vaginal cancer in mice. *Cancer Res.* **1959**, 19:170-176.
75. GARDNER, W. U. Carcinoma of the uterine cervix and upper vagina: induction under experimental conditions in mice. *Ann. N. Y. Acad. Sci.* **1959**, 75:543-564.
76. GARDNER, W. U.; Allen, E. Malignant and Non-malignant Uterine and Vaginal Lesions in Mice Receiving Estrogens and Estrogens and Androgens Simultaneously. *Yale J Biol. Med.* **1939**, 12:213-234.
77. PAN, S. C.; GARDNER, W. U. Carcinomas of the uterine cervix and vagina in estrogen- and androgen-treated hybrid mice. *Cancer Res.* **1948**, 8:337-345.
78. GARDNER, W. U. Hormonal aspects of experimental tumorigenesis. *Adv. Cancer Res.* **1953**, 1:173-232.
79. GARDNER, W. U. Uterine cervical and vaginal cancers in experimental animals. *Acta Unio. Int. Contra. Cancrum.* **1961**, 17:905-909.
80. Liao, D. J.; Dickson, R. B. Roles of androgens in the development, growth, and carcinogenesis of the mammary gland. *J. Steroid Biochem. Mol. Biol.* **2002**, 80:175-189.
81. Noble, R. L. The development of prostatic adenocarcinoma in Nb rats following prolonged sex hormone administration. *Cancer Res.* **1977**, 37:1929-1933.
82. Noble, R. L. Sex steroids as a cause of adenocarcinoma of the dorsal prostate in Nb rats, and their influence on the growth of transplants. *Oncology* **1977**, 34:138-141.
83. Noble, R. L. Production of Nb rat carcinoma of the dorsal prostate and response of estrogen-dependent transplants to sex hormones and tamoxifen. *Cancer Res.* **1980**, 40:3547-3550.
84. Noble, R. L. Prostate carcinoma of the Nb rat in relation to hormones. *Int. Rev. Exp. Pathol.* **1982**, 23:113-159.
85. van Nie, R. Biological aspects of the genesis of uterine tumours induced in mice by testosterone. *Jaarb. Kankeronderz. Kankerbestrijd. Ned.* **1964**, 14:247-252.
86. van, N. I. E.; BENEDETTI, E. L.; Muhlbock, O. A carcinogenic action of testosterone, provoking uterine tumours in mice. *Nature* **1961**, 192:1303.
87. Liao, D. Z.; Pantazis, C. G.; Hou, X.; Li, S. A. Promotion of estrogen-induced mammary gland carcinogenesis by androgen in the male Noble rat: probable mediation by steroid receptors. *Carcinogenesis* **1998**, 19:2173-2180.

88. Xie, B.; Tsao, S. W.; Wong, Y. C. Induction of high incidence of mammary tumour in female Noble rats with a combination of 17beta-oestradiol and testosterone. *Carcinogenesis* **1999**, 20:1069-1078.
89. Xie, B.; Tsao, S. W.; Wong, Y. C. Sex hormone-induced mammary carcinogenesis in female noble rats: the role of androgens. *Carcinogenesis* **1999**, 20:1597-1606.
90. Xie, B.; Tsao, S. W.; Wong, Y. C. Sex hormone-induced mammary carcinogenesis in female Noble rats: expression of TGF-beta1 and its receptors, TGF-alpha, and EGF-R in mammary carcinogenesis. *Breast Cancer Res. Treat.* **1999**, 58:227-239.
91. Xie, B.; Tsao, S. W.; Wong, Y. C. Sex hormone-induced mammary carcinogenesis in the female Noble rats: expression of bcl-2 and bax in hormonal mammary carcinogenesis. *Breast Cancer Res. Treat.* **2000**, 61:45-57.
92. Liao, D. J.; Dickson, R. B. Steroid hormone-growth factor interactions in proliferative controls of the mammary gland and breast cancer: a rapidly evolving perspective. *J. Steroid Biochem. Mol. Biol.* **2002**, 80:135-136.
93. Zhang, J.; Sun, Y.; Liu, Y.; Sun, Y.; Liao, D. J. Synergistic effects of androgen and estrogen on the mouse uterus and mammary gland. *Oncol. Rep.* **2004**, 12:709-716.
94. Kirkman, H.; BACON, R. L. Malignant renal tumors in male hamsters (*Cricetus auratus*) treated with estrogen. *Cancer Res.* **1950**, 10:122-124.
95. Kirkman, H.; BACON, R. L. Estrogen-induced tumors of the kidney. II. Effect of dose, administration, type of estrogen, and age on the induction of renal tumors in intact male golden hamsters. *J. Natl. Cancer Inst.* **1952**, 13:757-771.
96. Kirkman, H.; BACON, R. L. Estrogen-induced tumors of the kidney. I. Incidence of renal tumors in intact and gonadectomized male golden hamsters treated with diethylstilbestrol. *J. Natl. Cancer Inst.* **1952**, 13:745-755.
97. Kirkman, H.; Algard, F. T. Autonomous variants of an androgen/estrogen-induced and -dependent ductus deferens leiomyosarcoma of the Syrian hamster. *Cancer Res.* **1970**, 30:35-40.
98. Liao, D. Z.; Hou, X.; Bai, S.; Li, S. A.; Li, J. J. Unusual deregulation of cell cycle components in early and frank estrogen-induced renal neoplasias in the Syrian hamster. *Carcinogenesis* **2000**, 21:2167-2173.
99. MATTHEWS, V. S.; Kirkman, H.; BACON, R. L. Kidney damage in the golden hamster following chronic administration of diethylstilbestrol and sesame oil. *Proc. Soc. Exp. Biol. Med.* **1947**, 66:195.
100. Kirkman, H.; Algard, F. T. Characteristics of an androgen-estrogen-induced uterine smooth muscle cell tumor of the Syrian hamster. *Cancer Res.* **1970**, 30:794-800.
101. Kirkman, H. Hormone-related tumors in Syrian hamsters. *Prog. Exp. Tumor Res.* **1972**, 16:201-240.
102. FURTH, J. Transplantability of induced granulosa cell tumors and of luteoma in mice; secondary effects of these growths. *Proc. Soc. Exp. Biol. Med.* **1946**, 61:212-214.
103. LI, M. H.; GARDNER, W. U. Further studies on the pathogenesis of ovarian tumors in mice. *Cancer Res.* **1949**, 9:35-41.
104. PECKHAM, B. M.; GREENE, R. R. Experimentally produced granulosa-cell tumors in rats. *Cancer Res.* **1952**, 12:25-29.
105. PECKHAM, B. M.; GREENE, R. R. Experimentally produced granulosa-cell tumors in rabbits. *Cancer Res.* **1952**, 12:654-656.
106. Burrows, H. Carcinoma mammae occurring in a male mouse under continued treatment with oestrin. *Am. J. Cancer* **1935**, 24:613-616.
107. Cutts, J. H. Estrogen-induced breast cancer in the rat. *Proc. Can. Cancer Conf.* **1966**, 6:50-68.
108. Cutts, J. H.; Froude, G. C. Regression of estrone-induced mammary tumors in the rat. *Cancer Res.* **1968**, 28:2413-2418.
109. Cutts, J. H. Unusual response to androgen of estrogen-dependent mammary tumors. *J. Natl. Cancer Inst.* **1969**, 42:485-488.

110. Cutts, J. H. Enzyme activities in regressing estrone-induced mammary tumors of the rat. *Cancer Res.* **1973**, 33:1235-1237.
111. Geschickter, C. F.; Lewis, D.; Hartman, C. G. Tumors of the breast related to the oestrin hormone. *Am. J. Cancer* **1934**, 21:828-859.
112. Huggins, C. Endocrine-induced regression of cancers. *Cancer Res.* **1967**, 27:1925-1930.
113. Huggins, C. Endocrine-induced regression of cancers. *Am. J. Surg.* **1978**, 136:233-238.
114. Mceuen, C. S. Occurrence of cancer in rats treated with oestrone. *Am. J. Cancer* **1938**, 34:184-195.
115. Noble, R. L.; Cutts, J. H. Mammary tumors of the rat: a review. *Cancer Res.* **1959**, 19:1125-1139.
116. Wang, C.; Lisanti, M. P.; Liao, D. J. Reviewing once more the c-myc and Ras collaboration: converging at the cyclin D1-CDK4 complex and challenging basic concepts of cancer biology. *Cell Cycle* **2011**, 10:57-67.
117. Noble, R. L.; Hochachka, B. C.; King, D. Spontaneous and estrogen-produced tumors in Nb rats and their behavior after transplantation. *Cancer Res.* **1975**, 35:766-780.
118. GARDNER, W. U. Steroids in experimental carcinogenesis. *Cancer* **1957**, 10:726-730.
119. ANDERVONT, H. B.; CANTER, H. Y.; SHIMKIN, M. B. Effect of discontinued estrogenic stimulation upon the development and growth of testicular tumors in mice. *J. Natl. Cancer Inst.* **1957**, 18:1-39.
120. ANDERVONT, H. B.; SHIMKIN, M. B.; CANTER, H. Y. Some factors involved in the induction or growth of testicular tumors in BALB/c mice. *J. Natl. Cancer Inst.* **1960**, 25:1083-1096.
121. ANDERVONT, H. B.; SHIMKIN, M. B.; CANTER, H. Y. The growth of estrogen-induced interstitial-cell testicular tumors in BALB/c mice. *J. Natl. Cancer Inst.* **1960**, 24:1219-1237.
122. ANDERVONT, H. B.; SHIMKIN, M. B.; CANTER, H. Y. Testicular tumors in mice after removal of stilbestrol-cholesterol pellets. *Acta Unio. Int. Contra. Cancrum.* **1961**, 17:105-112.
123. FURTH, J. Discussion of problems related to hormonal factors in initiating and maintaining tumor growth. *Cancer Res.* **1957**, 17:454-463.
124. GARDNER, W. U. Hormonal imbalances in tumorigenesis. *Cancer Res.* **1948**, 8:397-411.
125. DUNNING, W. F. Response of some isologously transplanted rat neoplasms to steroids. *Ann. N. Y. Acad. Sci.* **1958**, 76:696-704.
126. Huggins, C. Prostatic cancer treated by orchietomy; the five year results. *J. Am. Med. Assoc.* **1946**, 131:576-581.
127. Huggins, C.; BERGENSTAL, D. M. Inhibition of human mammary and prostatic cancers by adrenalectomy. *Cancer Res.* **1952**, 12:134-141.
128. Huggins, C.; Hodges, C. V. Studies on prostatic cancer: I. The effect of castration, of estrogen and of androgen injection on serum phosphatases in metastatic carcinoma of the prostate. **1941.** *J. Urol.* 2002, 168:9-12.
129. Angevine, D. M. Significant events in the life of Jacob Furth. *Cancer Res.* **1966**, 26:351-356.
130. Weinhouse, S.; Furth, J. J. Jacob Furth - September 20, 1896-July 23, 1979. *Biogr. Mem. Natl. Acad. Sci.* **1992**, 62:167-197.
131. FURTH, J. The making and missing of discoveries: an autobiographical essay. *Cancer Res.* **1976**, 36:871-880.
132. FURTH, J. Conditioned and autonomous neoplasms: a review. *Cancer Res.* **1953**, 13:477-492.
133. Loeb, L. Further Investigations on the Origin of Tumors in Mice : VI. Internal Secretion as a Factor in the Origin of Tumors. *J. Med Res.* **1919**, 40:477-496.
134. BLOOM, H. J.; BAKER, W. H.; DUKES, C. E.; MITCHLEY, B. C. HORMONE-DEPENDENT TUMOURS OF THE KIDNEY. II. EFFECT OF ENDOCRINE ABLATION PROCEDURES ON THE TRANSPLANTED OESTROGEN-INDUCED RENAL TUMOUR OF THE SYRIAN HAMSTER. *Br. J. Cancer* **1963**, 17:646-656.
135. BLOOM, H. J.; DUKES, C. E.; MITCHLEY, B. C. HORMONE-DEPENDENT TUMOURS OF THE KIDNEY. I. THE OESTROGEN-INDUCED RENAL TUMOUR OF THE SYRIAN HAMSTER. HORMONE TREATMENT AND POSSIBLE RELATIONSHIP TO CARCINOMA OF THE KIDNEY IN MAN. *Br. J. Cancer* **1963**, 17:611-645.

136. Kirkman, H. Steroid tumorigenesis. *Cancer* **1957**, 10:757-764.
137. Kirkman, H. Autonomous derivatives of estrogen-induced renal carcinomas and spontaneous renal tumors in the Syrian hamster. *Cancer Res.* **1974**, 34:2728-2744.
138. Kirkman, H.; Chesterman, F. C. Additional data on transplanted tumours of the golden hamster. *Prog. Exp. Tumor Res.* **1972**, 16:580-621.
139. UPTON, A. C.; FURTH, J. Induction of pituitary tumors by means of ionizing irradiation. *Proc. Soc. Exp. Biol. Med.* **1953**, 84:255-257.
140. GORBMAN, A. Tumorous growths in the pituitary and trachea following radiotoxic dosages of I131. *Proc. Soc. Exp. Biol. Med.* **1949**, 71:237-240.
141. GORBMAN, A. Pituitary tumors in rodents following changes in thyroid function: a review. *Cancer Res.* **1956**, 16:99-105.
142. GORBMAN, A. Factors influencing development of hypophyseal tumors in mice after treatment with radioactive iodine. *Proc. Soc. Exp. Biol. Med.* **1952**, 80:538-540.
143. DENT, J. N.; GADSDEN, E. L.; FURTH, J. On the relation between thyroid depression and pituitary tumor induction in mice. *Cancer Res.* **1955**, 15:70-75.
144. DENT, J. N.; GADSDEN, E. L.; FURTH, J. Further studies on induction and growth of thyrotropic pituitary tumors in mice. *Cancer Res.* **1956**, 16:171-174.
145. FURTH, J.; BURNETT, W. T., Jr. Hormone-secreting transplantable neoplasms of the pituitary induced by I131. *Proc. Soc. Exp. Biol. Med.* **1951**, 78:222-224.
146. FURTH, J.; GADSDEN, E. L.; BURNETT, W. T., Jr. Autonomous transplantable pituitary tumors arising in growths dependent on absence of the thyroid gland. *Proc. Soc. Exp. Biol. Med.* **1952**, 80:4-7.
147. FURTH, J.; BURNETT, W. T., Jr.; GADSDEN, E. L. Quantitative relationship between thyroid function and growth of pituitary tumors secreting TSH. *Cancer Res.* **1953**, 13:298-307.
148. BIELSCHOWSKY, F. Chronic iodine deficiency as cause of neoplasia in thyroid and pituitary of aged rats. *Br. J. Cancer* **1953**, 7:203-213.
149. BIELSCHOWSKY, F.; HALL, W. H. Carcinogenesis in the thyroidectomized rat. *Br. J. Cancer* **1953**, 7:358-366.
150. BIELSCHOWSKY, F. Functional acidophilic tumours of the pituitary of the rat. *Br. J. Cancer* **1954**, 8:154-160.
151. BIELSCHOWSKY, F. Neoplasia and internal environment. *Br. J. Cancer* **1955**, 9:80-116.
152. BIELSCHOWSKY, F.; HORNING, E. S. Aspects of endocrine carcinogenesis. *Br. Med. Bull.* **1958**, 14:106-115.
153. FURTH, J.; DENT, J. N.; BURNETT, W. T., Jr.; GADSDEN, E. L. The mechanism of induction and the characteristics of pituitary tumors induced by thyroidectomy. *J. Clin. Endocrinol. Metab.* **1955**, 15:81-97.
154. HALMI, N. S.; GUDE, W. D. The morphogenesis of pituitary tumors induced by radiothyroidectomy in the mouse and the effects of their transplantation on the pituitary body of the host. *Am. J. Pathol.* **1954**, 30:403-419.
155. BIELSCHOWSKY, F. The role of thyroxine deficiency in the formation of experimental tumours of the thyroid. *Br. J. Cancer* **1949**, 3:547-9.
156. BIELSCHOWSKY, F.; GRIESBACH, W. E.; . Studies on experimental goitre; the transplant-ability of experimental thyroid tumours of the rat. *Br. J. Cancer* **1949**, 3:541-6.
157. BIELSCHOWSKY, F. Hormonal factors in neoplasia of the thyroid. *Acta Unio. Int. Contra. Cancrum.* **1960**, 16:133-137.
158. MONEY, W. L.; GODWIN, J. T.; RAWSON, R. W. The experimental production of thyroid tumors in the rat by the administration of sodium-5-iodo-2-thiouracil. *Cancer* **1957**, 10:690-697.

159. MORRIS, H. P.; DALTON, A. J.; GREEN, C. D. Malignant thyroid tumors occurring in the mouse after prolonged hormonal imbalance during the ingestion of thiouracil. *J. Clin. Endocrinol. Metab.* **1951**, 11:1281-1295.
160. MORRIS, H. P.; GREEN, C. D. The role of thiouracil in the induction, growth, and transplantability of mouse thyroid tumors. *Science* **1951**, 114:44-46.
161. MORRIS, H. P. Experimental thyroid tumors. *Brookhaven. Symp. Biol.* **1955**, 7:192-218.
162. PURVES, H. D.; GRIESBACH, W. E. Studies on experimental goitre; thyroid carcinomata in rats treated with thiourea. *Br. J. Exp. Pathol.* **1946**, 27:294-297.
163. PURVES, H. D.; GRIESBACH, W. E. Studies on experimental goitre; thyroid tumours in rats treated with thiourea. *Br. J. Exp. Pathol.* **1947**, 28:46-53.
164. DALTON, A. J.; MORRIS, H. P.; STRIEBICH, M. J.; DUBNIK, C. S. Histologic changes in strain C mice following long-term ingestion of thiouracil. *J. Natl. Cancer Inst.* **1950**, 11:391-413.
165. DALTON, A. J.; MORRIS, H. P.; DUBNIK, C. Change in the thyroid and other organs in mice receiving thiouracil. *Fed. Proc.* **1946**, 5:219-230.
166. MORRIS, H. P.; GREEN, C. D.; DALTON, A. J. The effect of ingestion of thiouracil on strain C mice. *J. Natl. Cancer Inst.* **1951**, 11:805-815.
167. GOLDBERG, R. C.; LINDSAY, S.; NICHOLS, C. W., Jr.; CHAIKOFF, I. L. INDUCTION OF NEOPLASMS IN THYROID GLANDS OF RATS BY SUBTOTAL THYROIDECTOMY AND BY THE INJECTION OF ONE MICROCURIE OF I-131. *Cancer Res.* **1964**, 24:35-43.
168. GOLDBERG, R. C.; CHAIKOFF, I. L. Induction of thyroid cancer in the rat by radioactive iodine. *AMA. Arch. Pathol.* **1952**, 53:22-28.
169. GOLDBERG, R. C.; CHAIKOFF, I. L. Development of thyroid neoplasms in the rat following a single injection of radioactive iodine. *Proc. Soc. Exp. Biol. Med.* **1951**, 76:563-566.
170. FURTH, J. Thyroid-pituitary tumorigenesis. *J. Natl. Cancer Inst.* **1954**, 15:687-691.
171. Brand, R. A. Biographical sketch: James Stephen Ewing, MD (1844-1943). *Clin. Orthop. Relat Res.* **2012**, 470:639-641.
172. Knauss, S.; Klein, A. From aneuploidy to cancer: the evolution of a new species? *J. Biosci.* **2012**, 37:211-220.
173. Huxley, J. S. Cancer biology: Comparative and genetic. *Biol. Rev.* **1956**, 31:474-513.
174. Vincent, M. D. Cancer: beyond speciation. *Adv. Cancer Res.* **2011**, 112:283-350.
175. Zhang, J.; Lou, XM.; Jin, LY.; Zhou, RJ.; Liu, SQ.; Xu, NZ.; Liao, DJ. Necrosis, and then stress induced necrosis-like cell death, but not apoptosis, should be the preferred cell death mode for chemotherapy: clearance of a few misconceptions. *Oncoscience* **2014**, 1:407-422.
176. Duesberg, P.; Mandrioli, D.; McCormack, A.; Nicholson, J. M. Is carcinogenesis a form of speciation? *Cell Cycle* **2011**, 10:2100-2114.
177. Liu, X.; Yang, W.; Guan, Z.; Yu, W.; Fan, B.; Xu, N.; Liao, D. J. There are only four basic modes of cell death, although there are many ad-hoc variants adapted to different situations. *Cell Biosci.* **2018**, 8:6-
doi: 10.1186/s13578-018-0206-6.
178. Shi, M.; Zhou, H.; Lei, M.; Chen, L.; Zellmer, L.; He, Y.; Yang, W.; Xu, N.; Liao, D. J. Spontaneous Cancers, But Not Many Induced Ones in Animals, Resemble Semi-New Organisms that Possess a Unique Programmed Cell Death Mode Different from Apoptosis, Senescent Death, Necrosis and Stress-Induced Cell Death. *J. Cancer* **2018**, 9:4726-4735.
179. GREENE, H. S. A conception of tumor autonomy based on transplantation studies: a review. *Cancer Res.* **1951**, 11:899-903.
180. Farber, E. Origins of human cancers. *Toxicol. Pathol.* **1985**, 13:86-89.
181. Rubin, H. What keeps cells in tissues behaving normally in the face of myriad mutations? *Bioessays* **2006**, 28:515-524.

182. Tomlinson, I. P.; Bodmer, W. F. Modelling the consequences of interactions between tumour cells. *Br. J. Cancer* **1997**, 75:157-160.
183. Weber, R. J.; Desai, T. A.; Gartner, Z. J. Non-autonomous cell proliferation in the mammary gland and cancer. *Curr. Opin. Cell Biol.* **2017**, 45:55-61.
184. Liao, D. J.; Dickson, R. B. c-Myc in breast cancer. *Endocr. Relat. Cancer* **2000**, 7:143-164.
185. Liao, J. D.; Adsay, N. V.; Khannani, F.; Grignon, D.; Thakur, A.; Sarkar, F. H. Histological complexities of pancreatic lesions from transgenic mouse models are consistent with biological and morphological heterogeneity of human pancreatic cancer. *Histol. Histopathol.* **2007**, 22:661-676.
186. Lou, X.; Zhang, J.; Liu, S.; Xu, N.; Liao, D. J. The other side of the coin: The tumor-suppressive aspect of oncogenes and the oncogenic aspect of tumor-suppressive genes, such as those along the CCND-CDK4/6-RB axis. *Cell Cycle* **2014**, 13:1677-1693.
187. Wang, C.; Tai, Y.; Lisanti, M. P.; Liao, D. J. c-Myc induction of programmed cell death may contribute to carcinogenesis: a perspective inspired by several concepts of chemical carcinogenesis. *Cancer Biol. Ther.* **2011**, 11:615-626.
188. Liao, D. J.; Natarajan, G.; Deming, S. L.; Jamerson, M. H.; Johnson, M.; Chepko, G.; Dickson, R. B. Cell cycle basis for the onset and progression of c-Myc-induced, TGFalpha-enhanced mouse mammary gland carcinogenesis. *Oncogene* **2000**, 19:1307-1317.
189. Liao, D. J.; Dickson, R. B. Cell death in MMTV-c-myc transgenic mouse mammary tumors may not be typical apoptosis. *Lab. Invest.* **2003**, 83:1437-1449.
190. Liao, D. J.; Wang, Y.; Wu, J.; Adsay, N. V.; Grignon, D.; Khanani, F.; Sarkar, F. H. Characterization of pancreatic lesions from MT-tgfa, Ela-myc and MT-tgfa/Ela-myc single and double transgenic mice. *J. Carcinog.* **2006**, 5:DOI: 10.1186/1477-3163-5-19.
191. Ma, Y.; Jia, Y.; Chen, L.; Ezeogu, L.; Yu, B.; Xu, N.; Liao, D. J. Weaknesses and Pitfalls of Using Mice and Rats in Cancer Chemoprevention Studies. *J. Cancer* **2015**, 6:1058-1065.
192. Arvanitis, C.; Felsher, D. W. Conditional transgenic models define how MYC initiates and maintains tumorigenesis. *Semin. Cancer Biol.* **2006**, 16:313-317.
193. D'Cruz, C. M.; Gunther, E. J.; Boxer, R. B.; Hartman, J. L.; Sintasath, L.; Moody, S. E.; Cox, J. D.; Ha, S. I.; Belka, G. K.; Golant, A.; Cardiff, R. D.; Chodosh, L. A. c-MYC induces mammary tumorigenesis by means of a preferred pathway involving spontaneous Kras2 mutations. *Nat. Med.* **2001**, 7:235-239.
194. Felsher, D. W.; Bishop, J. M. Reversible tumorigenesis by MYC in hematopoietic lineages. *Mol. Cell* **1999**, 4:199-207.
195. Fisher, G. H.; Wellen, S. L.; Klimstra, D.; Lenczowski, J. M.; Tichelaar, J. W.; Lizak, M. J.; Whitsett, J. A.; Koretsky, A.; Varmus, H. E. Induction and apoptotic regression of lung adenocarcinomas by regulation of a K-Ras transgene in the presence and absence of tumor suppressor genes. *Genes Dev.* **2001**, 15:3249-3262.
196. Shachaf, C. M.; Kopelman, A. M.; Arvanitis, C.; Karlsson, A.; Beer, S.; Mandl, S.; Bachmann, M. H.; Borowsky, A. D.; Ruebner, B.; Cardiff, R. D.; Yang, Q.; Bishop, J. M.; Contag, C. H.; Felsher, D. W. MYC inactivation uncovers pluripotent differentiation and tumour dormancy in hepatocellular cancer. *Nature* **2004**, 431:1112-1117.
197. Tilli, M. T.; Furth, P. A. Conditional mouse models demonstrate oncogene-dependent differences in tumor maintenance and recurrence. *Breast Cancer Res.* **2003**, 5:202-205.
198. Tran, P. T.; Fan, A. C.; Bendapudi, P. K.; Koh, S.; Komatsubara, K.; Chen, J.; Horng, G.; Bellovin, D. I.; Giuriato, S.; Wang, C. S.; Whitsett, J. A.; Felsher, D. W. Combined Inactivation of MYC and K-Ras oncogenes reverses tumorigenesis in lung adenocarcinomas and lymphomas. *PLoS. One* **2008**, 3:e2125- doi: 10.1371/journal.pone.0002125.
199. Shachaf, C. M.; Felsher, D. W. Rehabilitation of cancer through oncogene inactivation. *Trends Mol. Med.* **2005**, 11:316-321.

200. Shachaf, C. M.; Gentles, A. J.; Elchuri, S.; Sahoo, D.; Soen, Y.; Sharpe, O.; Perez, O. D.; Chang, M.; Mitchel, D.; Robinson, W. H.; Dill, D.; Nolan, G. P.; Plevritis, S. K.; Felsher, D. W. Genomic and proteomic analysis reveals a threshold level of MYC required for tumor maintenance. *Cancer Res.* **2008**, 68:5132-5142.
201. Nguyen, A. T.; Emelyanov, A.; Koh, C. H.; Spitsbergen, J. M.; Parinov, S.; Gong, Z. An inducible kras(V12) transgenic zebrafish model for liver tumorigenesis and chemical drug screening. *Dis. Model. Mech.* **2012**, 5:63-72.
202. Uchiyama, K.; Watanabe, D.; Hayasaka, M.; Hanaoka, K. A novel imprinted transgene located near a repetitive element that exhibits allelic imbalance in DNA methylation during early development. *Dev. Growth Differ.* **2014**, 56:653-668.
203. Zheng, W.; Li, Z.; Nguyen, A. T.; Li, C.; Emelyanov, A.; Gong, Z. Xmrk, kras and myc transgenic zebrafish liver cancer models share molecular signatures with subsets of human hepatocellular carcinoma. *PLoS. One* **2014**, 9:e91179-doi: 10.1371/journal.pone.0091179.
204. Sun, L.; Nguyen, A. T.; Spitsbergen, J. M.; Gong, Z. Myc-induced liver tumors in transgenic zebrafish can regress in tp53 null mutation. *PLoS. One* **2015**, 10:e0117249.
205. Anders, K.; Kershaw, O.; Larue, L.; Gruber, A. D.; Blankenstein, T. The immune system prevents recurrence of transplanted but not autochthonous antigenic tumors after oncogene inactivation therapy. *Int. J. Cancer* **2017**, 141:2551-2561.
206. Dolezal, J. M.; Wang, H.; Kulkarni, S.; Jackson, L.; Lu, J.; Ranganathan, S.; Goetzman, E. S.; Bharathi, S. S.; Beezhold, K.; Byersdorfer, C. A.; Prochownik, E. V. Sequential adaptive changes in a c-Myc-driven model of hepatocellular carcinoma. *J. Biol. Chem.* **2017**, 292:10068-10086.
207. Tonelli, C.; Morelli, M. J.; Sabo, A.; Verrecchia, A.; Rotta, L.; Capra, T.; Bianchi, S.; Campaner, S.; Amati, B. Genome-wide analysis of p53-regulated transcription in Myc-driven lymphomas. *Oncogene* **2017**, 36:2921-2929.
208. Martins, C. P.; Brown-Swigart, L.; Evan, G. I. Modeling the therapeutic efficacy of p53 restoration in tumors. *Cell* **2006**, 127:1323-1334.
209. Ventura, A.; Kirsch, D. G.; McLaughlin, M. E.; Tuveson, D. A.; Grimm, J.; Lintault, L.; Newman, J.; Reczek, E. E.; Weissleder, R.; Jacks, T. Restoration of p53 function leads to tumour regression in vivo. *Nature* **2007**, 445:661-665.
210. Xue, W.; Zender, L.; Miething, C.; Dickins, R. A.; Hernando, E.; Krizhanovsky, V.; Cordon-Cardo, C.; Lowe, S. W. Senescence and tumour clearance is triggered by p53 restoration in murine liver carcinomas. *Nature* **2007**, 445:656-660.
211. Wang, Y.; Suh, Y. A.; Fuller, M. Y.; Jackson, J. G.; Xiong, S.; Terzian, T.; Quintas-Cardama, A.; Bankson, J. A.; El-Naggar, A. K.; Lozano, G. Restoring expression of wild-type p53 suppresses tumor growth but does not cause tumor regression in mice with a p53 missense mutation. *J. Clin. Invest.* **2011**, 121:893-904.
212. Weinstein, I. B. Cancer. Addiction to oncogenes--the Achilles heel of cancer. *Science* **2002**, 297:63-64.
213. Weinstein, I. B.; Joe, A. K. Mechanisms of disease: Oncogene addiction--a rationale for molecular targeting in cancer therapy. *Nat. Clin. Pract. Oncol.* **2006**, 3:448-457.
214. Weinstein, I. B.; Joe, A. Oncogene addiction. *Cancer Res.* **2008**, 68:3077-3080.
215. Felsher, D. W. Tumor dormancy: death and resurrection of cancer as seen through transgenic mouse models. *Cell Cycle* **2006**, 5:1808-1811.
216. Felsher, D. W. Tumor dormancy and oncogene addiction. *APMIS* **2008**, 116:629-637.
217. Felsher, D. W. MYC Inactivation Elicits Oncogene Addiction through Both Tumor Cell-Intrinsic and Host-Dependent Mechanisms. *Genes Cancer* **2010**, 1:597-604.
218. Shachaf, C. M.; Felsher, D. W. Tumor dormancy and MYC inactivation: pushing cancer to the brink of normalcy. *Cancer Res.* **2005**, 65:4471-4474.

219. Tsujiuchi, T.; Nakae, D.; Konishi, Y. Multi-step lung carcinogenesis model induced by oral administration of N-nitrosobis(2-hydroxypropyl)amine in rats. *Exp. Toxicol. Pathol.* **2014**, *66*:81-88.
220. Challis, G. B.; Stam, H. J. The spontaneous regression of cancer. A review of cases from 1900 to 1987. *Acta Oncol.* **1990**, *29*:545-550.
221. Everson, T. C. SPONTANEOUS REGRESSION OF CANCER. *Ann. N. Y. Acad. Sci.* **1964**, *114*:721-735.
222. Everson, T. C. Spontaneous regression of cancer. *Prog. Clin. Cancer* 1967, *3*:79-95.
223. Kleef, R.; Jonas, W. B.; Knogler, W.; Stenzinger, W. Fever, cancer incidence and spontaneous remissions. *Neuroimmunomodulation* **2001**, *9*:55-64.
224. Stephenson, H. E., Jr. Spontaneous regression of cancer evaluated by computerized data. *Natl. Cancer Inst. Monogr.* 1976, *44*:43-47.
225. Thomas, J. A.; Badini, M. The role of innate immunity in spontaneous regression of cancer. *Indian J. Cancer* **2011**, *48*:246-251.
226. Bodey, B.; Bodey, B., Jr.; Siegel, S. E.; Kaiser, H. E. The spontaneous regression of neoplasms in mammals: possible mechanisms and their application in immunotherapy. *In Vivo* **1998**, *12*:107-122.
227. Kaiser, H. E.; Bodey, B., Jr.; Siegel, S. E.; Groger, A. M.; Bodey, B. Spontaneous neoplastic regression: the significance of apoptosis. *In Vivo* **2000**, *14*:773-788.
228. Nagorsen, D.; Marincola, F. M.; Kaiser, H. E. Bacteria-related spontaneous and therapeutic remission of human malignancies. *In Vivo* 2002, *16*:551-556.
229. Jessy, T. Immunity over inability: The spontaneous regression of cancer. *J. Nat. Sci. Biol. Med.* **2011**, *2*:43-49.
230. Ahmadi, M. P.; Cornejo, K. M.; Hutchinson, L.; Tomaszewicz, K.; Dresser, K.; Deng, A.; O'Donnell, P. Complete Spontaneous Regression of Merkel Cell Carcinoma After Biopsy: A Case Report and Review of the Literature. *Am. J. Dermatopathol.* **2016**, *38*:e154-e158.
231. Brodeur, G. M. Spontaneous regression of neuroblastoma. *Cell Tissue Res.* **2018**: doi: 10.1007/s00441-017-2761-2.
232. Buhler, H.; Pirovino, M.; Akobiantz, A.; Altorfer, J.; Weitzel, M.; Maranta, E.; Schmid, M. Regression of liver cell adenoma. A follow-up study of three consecutive patients after discontinuation of oral contraceptive use. *Gastroenterology* **1982**, *82*:775-782.
233. Sakamaki, A.; Kamimura, K.; Abe, S.; Tsuchiya, A.; Takamura, M.; Kawai, H.; Yamagiwa, S.; Terai, S. Spontaneous regression of hepatocellular carcinoma: A mini-review. *World J. Gastroenterol.* **2017**, *23*:3797-3804.
234. BLOOM, H. J.; RICHARDSON, W. W.; HARRIES, E. J. Natural history of untreated breast cancer (1805-1933). Comparison of untreated and treated cases according to histological grade of malignancy. *Br. Med J.* **1962**, *2*:213-221.
235. Beller, U.; Beckman, E. M.; TWOMBLY, G. H. Spontaneous regression of advanced endometrial carcinoma. *Gynecol. Oncol.* **1984**, *17*:381-385.
236. Hobohm, U. Fever and cancer in perspective. *Cancer Immunol. Immunother.* **2001**, *50*:391-396.
237. Jerry, LM.; Challis; EB. Oncology. In: Rakel and RE. *Textbook of family practice*. 3rd edn. ed; **1952**: p1061-1081.
238. Smith, J. L., Jr.; Stehlin, J. S., Jr. Spontaneous regression of primary malignant melanomas with regional metastases. *Cancer* **1965**, *18*:1399-1415.
239. Ribero, S.; Gualano, M. R.; Osella-Abate, S.; Scaioli, G.; Bert, F.; Sanlorenzo, M.; Balagna, E.; Fierro, M. T.; Macripo, G.; Sapino, A.; Siliquini, R.; Quaglino, P. Association of Histologic Regression in Primary Melanoma With Sentinel Lymph Node Status: A Systematic Review and Meta-analysis. *JAMA Dermatol.* **2015**, *151*:1301-1307.
240. Pique-Duran, E.; Palacios-Llopis, S.; Martinez-Martin, M.; Perez-Cejudo, J. A. Complete regression of melanoma associated with vitiligo. *Dermatol. Online J.* **2011**, *17*:4-
<https://escholarship.org/uc/item/7sn7h2j7#main>.

241. Ong, S. F.; Harden, M.; Irandoust, S.; Lee, R. W. Spontaneous regression of pulmonary metastatic melanoma. *Respirol. Case Rep.* **2016**, 4:7-9.
242. Khosravi, H.; Akabane, A. L.; Alloo, A.; Nazarian, R. M.; Boland, G. M. Metastatic melanoma with spontaneous complete regression of a thick primary lesion. *JAAD Case Rep.* **2016**, 2:439-441.
243. Kang, S.; Barnhill, R. L.; Mihm, M. C., Jr.; Sober, A. J. Histologic regression in malignant melanoma: an interobserver concordance study. *J. Cutan. Pathol.* **1993**, 20:126-129.
244. Emanuel, P. O.; Mannion, M.; Phelps, R. G. Complete regression of primary malignant melanoma. *Am. J. Dermatopathol.* **2008**, 30:178-181.
245. Cervinkova, M.; Kucerova, P.; Cizkova, J. Spontaneous regression of malignant melanoma - is it based on the interplay between host immune system and melanoma antigens? *Anticancer Drugs* **2017**, 28:819-830.
246. Ribero, S.; Moscarella, E.; Ferrara, G.; Piana, S.; Argenziano, G.; Longo, C. Regression in cutaneous melanoma: a comprehensive review from diagnosis to prognosis. *J. Eur. Acad. Dermatol. Venereol.* **2016**, 30:2030-2037.
247. Bruns, P. Die Heilwirkung des Erysipels auf Geschwulste. *Beiträge zur Klinischen Chirurgie* **1887**, 3:443-446.
248. Busch, W. Über den Einfluss welche heftigere Erysipeln zuweilig auf organisierte Neubildungenausüben. *Verhandlungen des Naturhistorischen Vereines der Preussischen Rheinlande und Westphalens* **1866**, 23:28-30.
249. Busch, W. Aus der sitzung der medicinischen section. *Berliner Klinische Wochenschrift* **1868**, 5:137-138.
250. Fehleisen, F. Ueber die Züchtung der Erysipelkokken auf künstlichem Nährboden und ihre Übertragbarkeit auf den Menschen. *Dtsch Med. Wochenschr.* **1882**, 8:553-554.
251. Hobohm, U. Fever therapy revisited. *Br. J. Cancer* **2005**, 92:421-425.
252. Hobohm, U. Healing heat: Harnessing infection to flight cancer. *Am. Sci.* **2009**, 97:34-41.
253. Hobohm, U. Toward general prophylactic cancer vaccination. *Bioessays* **2009**, 31:1071-1079.
254. Hopton Cann, S. A.; van Netten, J. P.; van, N. C. Dr William Coley and tumour regression: a place in history or in the future. *Postgrad. Med. J.* **2003**, 79:672-680.
255. Hopton Cann, S. A.; van Netten, J. P.; van, N. C. Acute infections as a means of cancer prevention: opposing effects to chronic infections? *Cancer Detect. Prev.* **2006**, 30:83-93.
256. Hopton Cann, S. A. Peak fever: helpful or harmful? *Heart Lung* **2011**, 40:585-586.
257. Kienle, G. S. Fever in Cancer Treatment: Coley's Therapy and Epidemiologic Observations. *Glob. Adv. Health Med.* **2012**, 1:92-100.
258. Nauts, H. C.; FOWLER, G. A.; BOGATKO, F. H. A review of the influence of bacterial infection and of bacterial products (Coley's toxins) on malignant tumors in man; a critical analysis of 30 inoperable cases treated by Coley's mixed toxins, in which diagnosis was confirmed by microscopic examination selected for special study. *Acta Med. Scand. Suppl.* **1953**, 276:1-103.
259. Nauts, H. C. The beneficial effects of bacterial infections on host resistance to cancer. End results in 449 cases. A study and abstracts of reports in the world medical literature (1775-1980) and personal communication. *Cancer Res. Inst. Monog.* **1980**, 8:1-225.
260. Nauts, H. C. Bacterial pyrogens: beneficial effects on cancer patients. *Prog. Clin. Biol. Res.* **1982**, 107:687-696.
261. Nauts, H. C. Bacteria and cancer--antagonisms and benefits. *Cancer Surv.* **1989**, 8:713-723.
262. Nauts, H. C.; McLaren, J. R. Coley toxins--the first century. *Adv. Exp. Med. Biol.* **1990**, 267:483-500.
263. Repasky, E. A.; Evans, S. S.; Dewhirst, M. W. Temperature Matters! And Why it Should Matter to Tumor Immunologists. *Cancer Immunol. Res.* **2013**, 1:210-216.

264. Tang, Z. Y.; Zhou, H. Y.; Zhao, G.; Chai, L. M.; Zhou, M.; Lu, J. Z.; Liu, K. D.; Havas, H. F.; Nauts, H. C. Preliminary result of mixed bacterial vaccine as adjuvant treatment of hepatocellular carcinoma. *Med. Oncol. Tumor Pharmacother.* **1991**, 8:23-28.
265. Wiemann, B.; Starnes, C. O. Coley's toxins, tumor necrosis factor and cancer research: a historical perspective. *Pharmacol. Ther.* **1994**, 64:529-564.
266. Orange, M.; Reuter, U.; Hobohm, U. Coley's Lessons Remembered: Augmenting Mistletoe Therapy. *Integr. Cancer Ther.* **2016**, 15:502-511.
267. Liu, B.; Ezeogu, L.; Zellmer, L.; Yu, B.; Xu, N.; Liao, DJ. Protecting the normal in order to better kill the cancer. *Cancer Med.* **2015**, 4:1394-1403.
268. Ewing, J. The General Pathological Conception of Cancer. *Can. Med. Assoc. J.* **1935**, 33:125-135.
269. Rubin, H. Cell damage, aging and transformation: a multilevel analysis of carcinogenesis. *Anticancer Res.* **1999**, 19:4877-4886.
270. Chow, M.; Rubin, H. Relation of the slow growth phenotype to neoplastic transformation: possible significance for human cancer. *In Vitro Cell Dev. Biol. Anim.* **1999**, 35:449-458.
271. Teotonio, H.; Rose, M. R. Perspective: reverse evolution. *Evolution* **2001**, 55:653-660.
272. Teotonio, H.; Chelo, I. M.; Bradic, M.; Rose, M. R.; Long, A. D. Experimental evolution reveals natural selection on standing genetic variation. *Nat. Genet.* 2009, 41:251-257.
273. Hirschhorn, R. In vivo reversion to normal of inherited mutations in humans. *J. Med. Genet.* **2003**, 40:721-728.
274. Lai-Cheong, J. E.; McGrath, J. A.; Uitto, J. Revertant mosaicism in skin: natural gene therapy. *Trends Mol. Med.* **2011**, 17:140-148.
275. Pasmooij, A. M.; Jonkman, M. F.; Uitto, J. Revertant mosaicism in heritable skin diseases: mechanisms of natural gene therapy. *Discov. Med.* **2012**, 14:167-179.
276. van, D. E.; Pretorius, P. J. Point mutation instability (PIN) mutator phenotype as model for true back mutations seen in hereditary tyrosinemia type 1 - a hypothesis. *J. Inherit. Metab. Dis.* **2012**, 35:407-411.
277. Ashworth, A. Drug resistance caused by reversion mutation. *Cancer Res.* **2008**, 68:10021-10023.
278. Bouwman, P.; Jonkers, J. Molecular Pathways: How Can BRCA-Mutated Tumors Become Resistant to PARP Inhibitors? *Clin. Cancer Res.* 2014, 20:540-547.
279. Dhillon, K. K.; Swisher, E. M.; Taniguchi, T. Secondary mutations of BRCA1/2 and drug resistance. *Cancer Sci.* **2011**, 102:663-669.
280. Baum, J. K.; Bookstein, J. J.; Holtz, F.; Klein, E. W. Possible association between benign hepatomas and oral contraceptives. *Lancet* **1973**, 2:926-929.
281. Horvath, E.; Kovacs, K.; Ross, R. C. Letter: Benign hepatoma in a young woman on contraceptive steroids. *Lancet* **1974**, 1:357-358.
282. Knapp, W. A.; Ruebner, B. H. Letter: Hepatomas and oral contraceptives. *Lancet* **1974**, 1:270-271.
283. Oral contraceptives and cancer. *Lancet* **1972**, 2:911-DOI: [https://doi.org/10.1016/S0140-6736\(72\)92541-X](https://doi.org/10.1016/S0140-6736(72)92541-X).
284. Lingeman, C. H. Letter: Liver-cell neoplasms and oral contraceptives. *Lancet* 1974, 1:64.
285. Thalassinou, N. C.; Lymberatos, C.; Hadjioannou, J.; Gardikas, C. Letter: Liver-cell carcinoma after long-term oestrogen-like drugs. *Lancet* **1974**, 1:270.
286. Wendel, H. A. Oral contraceptives and cancer. *Lancet* **1972**, 2:1139.
287. AGNEW, L. R.; GARDNER, W. U. The incidence of spontaneous hepatomas in C3H, C3H (low milk factor), and CBA mice and the effect of estrogen and androgen on the occurrence of these tumors in C3H mice. *Cancer Res.* **1952**, 12:757-761.
288. PULLINGER, B. D.; HEAD, M. A. HEPATOMA IN INTACT C3HF MALE AND VIRGIN FEMALE MICE AND AFTER GONADECOTOMY ALONE OR SEBSEQUENT TREATMENT WITH OESTROGEN. *Br. J. Cancer* **1964**, 13:521-527.

289. ANDERVONT, H. B.; DUNN, T. B. Transplantation of spontaneous and induced hepatomas in inbred mice. *J. Natl. Cancer Inst.* **1952**, 13:455-503.
290. Wotherspoon, A. C.; Doglioni, C.; Diss, T. C.; Pan, L.; Moschini, A.; de, B. M.; Isaacson, P. G. Regression of primary low-grade B-cell gastric lymphoma of mucosa-associated lymphoid tissue type after eradication of *Helicobacter pylori*. *Lancet* **1993**, 342:575-577.
291. Wundisch, T.; Thiede, C.; Morgner, A.; Dempfle, A.; Gunther, A.; Liu, H.; Ye, H.; Du, M. Q.; Kim, T. D.; Bayerdorffer, E.; Stolte, M.; Neubauer, A. Long-term follow-up of gastric MALT lymphoma after *Helicobacter pylori* eradication. *J. Clin. Oncol.* **2005**, 23:8018-8024.
292. Morgner, A.; Thiede, C.; Bayerdorffer, E.; Alpen, B.; Wundisch, T.; Neubauer, A.; Stolte, M. Long-term follow-up of gastric MALT lymphoma after H. pylori eradication. *Curr. Gastroenterol. Rep.* **2001**, 3:516-522.
293. Morgner, A.; Miehke, S.; Fischbach, W.; Schmitt, W.; Muller-Hermelink, H.; Greiner, A.; Thiede, C.; Schetelig, J.; Neubauer, A.; Stolte, M.; Ehninger, G.; Bayerdorffer, E. Complete remission of primary high-grade B-cell gastric lymphoma after cure of *Helicobacter pylori* infection. *J. Clin. Oncol.* **2001**, 19:2041-2048.
294. Wundisch, T.; Dieckhoff, P.; Greene, B.; Thiede, C.; Wilhelm, C.; Stolte, M.; Neubauer, A. Second cancers and residual disease in patients treated for gastric mucosa-associated lymphoid tissue lymphoma by *Helicobacter pylori* eradication and followed for 10 years. *Gastroenterology* **2012**, 143:936-942.
295. Bertoni, F.; Conconi, A.; Capella, C.; Motta, T.; Giardini, R.; Ponzoni, M.; Pedrinis, E.; Novero, D.; Rinaldi, P.; Cazzaniga, G.; Biondi, A.; Wotherspoon, A.; Hancock, B. W.; Smith, P.; Souhami, R.; Cotter, F. E.; Cavalli, F.; Zucca, E. Molecular follow-up in gastric mucosa-associated lymphoid tissue lymphomas: early analysis of the LY03 cooperative trial. *Blood* **2002**, 99:2541-2544.
296. Park, J. B.; Koo, J. S. *Helicobacter pylori* infection in gastric mucosa-associated lymphoid tissue lymphoma. *World J. Gastroenterol.* **2014**, 20:2751-2759.
297. El, H. H.; El-Sabban, M.; Hasegawa, H.; Zaatari, G.; Ablain, J.; Saab, S. T.; Janin, A.; Mahfouz, R.; Nasr, R.; Kfoury, Y.; Nicot, C.; Hermine, O.; Hall, W.; de, T. H.; Bazarbachi, A. Therapy-induced selective loss of leukemia-initiating activity in murine adult T cell leukemia. *J. Exp. Med.* **2010**, 207:2785-2792.
298. Gill, P. S.; Harrington, W., Jr.; Kaplan, M. H.; Ribeiro, R. C.; Bennett, J. M.; Liebman, H. A.; Bernstein-Singer, M.; Espina, B. M.; Cabral, L.; Allen, S.; . Treatment of adult T-cell leukemia-lymphoma with a combination of interferon alfa and zidovudine. *N. Engl. J. Med.* **1995**, 332:1744-1748.
299. Hermine, O.; Bouscary, D.; Gessain, A.; Turlure, P.; Leblond, V.; Franck, N.; Buzyn-Veil, A.; Rio, B.; Macintyre, E.; Dreyfus, F.; . Brief report: treatment of adult T-cell leukemia-lymphoma with zidovudine and interferon alfa. *N. Engl. J. Med.* **1995**, 332:1749-1751.
300. Axelrod, R. Launching "the evolution of cooperation". *J. Theor. Biol.* **2012**, 299:21-24.
301. Axelrod, R.; Axelrod, D. E.; Pienta, K. J. Evolution of cooperation among tumor cells. *Proc. Natl. Acad. Sci. U. S. A* **2006**, 103:13474-13479.
302. Wang, G.; Chen, L.; Yu, B.; Zellmer, L.; Xu, N.; Liao, D. J. Learning about the Importance of Mutation Prevention from Curable Cancers and Benign Tumors. *J. Cancer* **2016**, 7:436-445.
303. Jia QW.; Chen XH.; Jia YP.; Dou XX.; Ezeogu L.; Xu NZ.; Liao DJ. is type 2 diabetes one of such aging phenomena that lack an irreversible structural change? *J. Diabetes Metab.* **2105**, 6:543-doi: 10.4172/2155-6156.1000543.
304. Holmquist, G. P. Cell-selfish modes of evolution and mutations directed after transcriptional bypass. *Mutat. Res.* **2002**, 510:141-152.
305. Liu, B.; Xu, N.; Man, Y.; Shen, H.; Avital, I.; Stojadinovic, A.; Liao, D. J. Apoptosis in Living Animals Is Assisted by Scavenger Cells and Thus May Not Mainly Go through the Cytochrome C-Caspase Pathway. *J. Cancer* **2013**, 4:716-723.

306. Liao, D. J. The scavenger cell hypothesis of apoptosis: apoptosis redefined as a process by which a cell in living tissue is destroyed by phagocytosis. *Med. Hypotheses* **2005**, 65:23-28.
307. Liu, X. D.; Yang, W. X.; Guan, Z. Z.; Yu, W. F.; Fan, B.; Xu, N. Z.; Liao, D. J. There are only four basic modes of cell death, although there are many ad-hoc variants adapted to different situations. *Cell & Bioscience* **2018**, 8:doi.org/10.1186/s13578-018-0206-6.
308. Dou, X.; Chen, L.; Lei, M.; Zellmer, L.; Jia, Q.; Ling, P.; He, Y.; Yang, W.; Liao, D. J. Evaluating the Remote Control of Programmed Cell Death, with or without a Compensatory Cell Proliferation. *Int. J Biol. Sci.* **2018**, 14:1800-1812.
309. REVIEWS. *Br. Med. J.* 1903, 1:376-378.
310. Solt, D. B.; Farber, E. A new principle for the analysis of chemical carcinogenesis. *Nature* **1976**, 263:702-703.
311. Solt, D. B.; Medline, A.; Farber, E. Rapid emergence of carcinogen-induced hyperplastic lesions in a new model for the sequential analysis of liver carcinogenesis. *Am. J. Pathol.* **1977**, 88:595-618.
312. Liao, D.; Porsch-Hallstrom, I.; Gustafsson, J. A.; Blanck, A. Sex differences at the initiation stage of rat liver carcinogenesis--influence of growth hormone. *Carcinogenesis* **1993**, 14:2045-2049.
313. Liao, D. Z.; Blanck, A.; Gustafsson, J. A.; Hallstrom, I. P. Expression of the c-jun, jun-B, ets-2 and liver regeneration factor-1 (LRF-1) genes during promotion and progression of rat liver carcinogenesis in the resistant hepatocyte model. *Cancer Lett.* **1996**, 100:215-221.
314. Blanck, A.; Liao, D.; Gustafsson, J. A.; Hallstrom, I. P. Hormonal regulation of sex differentiated parameters in liver nodules from rats treated in the resistant hepatocyte model. *Carcinogenesis* **1995**, 16:231-235.
315. Flodby, P.; Liao, D. Z.; Blanck, A.; Xanthopoulos, K. G.; Hallstrom, I. P. Expression of the liver-enriched transcription factors C/EBP alpha, C/EBP beta, HNF-1, and HNF-4 in preneoplastic nodules and hepatocellular carcinoma in rat liver. *Mol. Carcinog.* **1995**, 12:103-109.
316. Farber, E. The Biology of Carcinogen-Induced Hepatocyte Nodules and Related Liver Lesions in the Rats. *Toxicol. Pathol.* **1982**, 10:197-201.
317. Farber, E. The step-by-step development of epithelial cancer: from phenotype to genotype. *Adv. Cancer Res.* **1996**, 70:21-48.
318. Herceg, Z.; Lambert, M. P.; van, V. K.; Demetriou, C.; Vineis, P.; Smith, M. T.; Straif, K.; Wild, C. P. Towards incorporating epigenetic mechanisms into carcinogen identification and evaluation. *Carcinogenesis* **2013**, 34:1955-1967.
319. Herceg, Z.; Vaissiere, T. Epigenetic mechanisms and cancer: an interface between the environment and the genome. *Epigenetics.* **2011**, 6:804-819.
320. Herceg, Z. Epigenetic Mechanisms as an Interface Between the Environment and Genome. *Adv. Exp. Med. Biol.* **2016**, 903:3-15.
321. Patel, S.; Shah, K.; Mirza, S.; Daga, A.; Rawal, R. Epigenetic regulators governing cancer stem cells and epithelial-mesenchymal transition in oral squamous cell carcinoma. *Curr. Stem Cell Res. Ther.* **2015**, 10:140-152.
322. Salemi, R.; Marconi, A.; Di, S., V; Franco, S.; Rapisarda, V.; Libra, M. Epigenetic alterations and occupational exposure to benzene, fibers, and heavy metals associated with tumor development (Review). *Mol. Med. Rep.* **2017**, 15:3366-3371.
323. Thomson, J. P.; Moggs, J. G.; Wolf, C. R.; Meehan, R. R. Epigenetic profiles as defined signatures of xenobiotic exposure. *Mutat. Res. Genet. Toxicol. Environ. Mutagen.* **2014**, 764-765:3-9.
324. Berenblum, I. A re-evaluation of the concept of cocarcinogenesis. *Prog. Exp. Tumor Res.* **1969**, 11:21-30.
325. Berenblum, I. Challenging problems in cocarcinogenesis. *Cancer Res.* **1985**, 45:1917-1921.
326. Cairns, J. Cocarcinogenesis and biological effects of tumor promoters. Conclusions and perspectives. *Carcinog. Compr. Surv.* **1982**, 7:647-651.

327. Brash, D.; Cairns, J. The mysterious steps in carcinogenesis: addendum. *Br. J Cancer* **2009**, *101*:1490.
328. Cairns, J. Mutation and cancer: the antecedents to our studies of adaptive mutation. *Genetics* **1998**, *148*:1433-1440.
329. Cairns, J.; Overbaugh, J.; Miller, S. The origin of mutants. *Nature* **1988**, *335*:142-145.
330. Kennedy, A. R. Is a mutagenic event involved in radiation induced malignant transformation? *Mutat. Res.* **1996**, *350*:81-91.
331. Kennedy, A. R. Is there a critical target gene for the first step in carcinogenesis? *Environ. Health Perspect.* **1991**, *93*:199-203.
332. Kennedy, A. R.; Cairns, J.; Little, J. B. Timing of the steps in transformation of C3H 10T 1/2 cells by X-irradiation. *Nature* **1984**, *307*:85-86.
333. Kennedy, A. R.; Fox, M.; Murphy, G.; Little, J. B. Relationship between x-ray exposure and malignant transformation in C3H 10T1/2 cells. *Proc. Natl. Acad. Sci. U. S. A* **1980**, *77*:7262-7266.
334. Rous, P. Recent Advances in Cancer Research. *Bull. N. Y. Acad. Med.* **1947**, *23*:65-78.
335. Zellmer, L.; Han, Y. P.; Chen, L. C.; Xu, N. Z.; Liao, D. J. Does the cytochrome c-caspase pathway of cell death occur physiologically in animals? *J. Tumor Med. Prev.* **2017**, *1*:JTMP.MS.ID.555557.pdf.
336. Zhang, J.; Lou, X.; Zellmer, L.; Liu, S.; Xu, N.; Liao, D. J. Just like the rest of evolution in Mother Nature, the evolution of cancers may be driven by natural selection, and not by haphazard mutations. *Oncoscience* **2014**, *1*:580-590.
337. Burrow, G. N.; Wortzman, G.; Rewcastle, N. B.; Holgate, R. C.; Kovacs, K. Microadenomas of the pituitary and abnormal sellar tomograms in an unselected autopsy series. *N. Engl. J. Med.* **1981**, *304*:156-158.
338. Chambers, E. F.; Turski, P. A.; LaMasters, D.; Newton, T. H. Regions of low density in the contrast-enhanced pituitary gland: normal and pathologic processes. *Radiology* **1982**, *144*:109-113.
339. Costello, R. T. Subclinical Adenoma of the Pituitary Gland. *Am. J. Pathol.* **1936**, *12*:205-216.
340. Muhr, C.; Bergstrom, K.; Grimelius, L.; Larsson, S. G. A parallel study of the roentgen anatomy of the sella turcica and the histopathology of the pituitary gland in 205 autopsy specimens. *Neuroradiology* **1981**, *21*:55-65.
341. Parent, A. D.; Bebin, J.; Smith, R. R. Incidental pituitary adenomas. *J. Neurosurg.* **1981**, *54*:228-231.
342. Hall, W. A.; Luciano, M. G.; Doppman, J. L.; Patronas, N. J.; Oldfield, E. H. Pituitary magnetic resonance imaging in normal human volunteers: occult adenomas in the general population. *Ann. Intern. Med.* **1994**, *120*:817-820.
343. Rich, A. R. On the frequency of occurrence of occult carcinoma of the prostate. *J Urol* **1935**, *33*:3-7.
344. Rich, A. R. Classics in oncology. On the frequency of occurrence of occult carcinoma of the prostate: Arnold Rice Rich, M.D., *Journal of Urology* *33*:3, 1935. *CA Cancer J. Clin.* **1979**, *29*:115-119.
345. Rich, A. R. On the frequency of occurrence of occult carcinoma of the prostate. 1934. *Int. J. Epidemiol.* **2007**, *36*:274-277.
346. Cairns, J. Mutation selection and the natural history of cancer. *Nature* **1975**, *255*:197-200.
347. Cairns, J. Cancer and the immortal strand hypothesis. *Genetics* **2006**, *174*:1069-1072.
348. FISHER, J. C. Multiple-mutation theory of carcinogenesis. *Nature* **1958**, *181*:651-652.
349. Beckman, R. A.; Loeb, L. A. Evolutionary dynamics and significance of multiple subclonal mutations in cancer. *DNA Repair (Amst)* **2017**, *56*:7-15.
350. Fox, E. J.; Loeb, L. A. Lethal mutagenesis: targeting the mutator phenotype in cancer. *Semin. Cancer Biol.* **2010**, *20*:353-359.
351. Prindle, M. J.; Fox, E. J.; Loeb, L. A. The mutator phenotype in cancer: molecular mechanisms and targeting strategies. *Curr. Drug Targets.* **2010**, *11*:1296-1303.
352. Venkatesan, R. N.; Loeb, L. A. The multiplicity of mutations in human cancers. *Adv. Exp. Med. Biol.* **2005**, *570*:3-17.

353. Venkatesan, R. N.; Bielas, J. H.; Loeb, L. A. Generation of mutator mutants during carcinogenesis. *DNA Repair (Amst)* **2006**, 5:294-302.
354. Bodmer, W. F.; Cottrell, S.; Frischauf, A. M.; Kerr, I. B.; Murday, V. A.; Rowan, A. J.; Smith, M. F.; Solomon, E.; Thomas, H.; Varesco, L. Genetic analysis of colorectal cancer. *Princess Takamatsu Symp.* **1989**, 20:49-59.
355. Koorey, D. J.; McCaughan, G. W. Tumour suppressor genes and colorectal neoplasia. *J. Gastroenterol. Hepatol.* **1993**, 8:174-184.
356. Nagase, H.; Nakamura, Y. Mutations of the APC (adenomatous polyposis coli) gene. *Hum. Mutat.* **1993**, 2:425-434.
357. Nakamura, Y.; Nishisho, I.; Kinzler, K. W.; Vogelstein, B.; Miyoshi, Y.; Miki, Y.; Ando, H.; Horii, A.; Nagase, H. Mutations of the adenomatous polyposis coli gene in familial polyposis coli patients and sporadic colorectal tumors. *Princess Takamatsu Symp.* **1991**, 22:285-292.
358. Williams, A. C.; Browne, S. J.; Manning, A. M.; Hague, A.; van der Stappen, J. W.; Paraskeva, C. Biological consequences of the genetic changes which occur during human colorectal carcinogenesis. *Semin. Cancer Biol.* **1993**, 4:153-159.
359. Rubin, H. Fields and field cancerization: the preneoplastic origins of cancer: asymptomatic hyperplastic fields are precursors of neoplasia, and their progression to tumors can be tracked by saturation density in culture. *Bioessays* **2011**, 33:224-231.
360. Farber, E. Putative precursor lesions: summary and some analytical considerations. *Cancer Res.* **1976**, 36:2703-2705.
361. Williams, D. Thyroid Growth and Cancer. *Eur. Thyroid J.* **2015**, 4:164-173.
362. Loeb, L. On Transplantation of tumors. *J. Med. Res.* **1901**, 6:28-38.
363. Loeb, L. ON SOME CONDITIONS OF TISSUE GROWTH, ESPECIALLY IN CULTURE MEDIA. *Science* **1911**, 34:414-415.
364. TWOMBLY, G. H.; MEISEL, D. The growth of mammalian tumors in fertile eggs; is a filterable cancer virus produced? *Cancer Res.* **1946**, 6:82-91.
365. Heilman, F. R.; Bittner, J. J. Observations on mouse tumors cultivated in the yolk sac of the embryonic chick. *Cancer Res.* **1912**, 4:578-582.
366. MCDUFFIE, N. G., Jr.; GIBSON, B. S.; TAYLOR, A. Study of toxic factors associated with mouse mammary carcinomas in egg cultures. *Cancer Res.* **1960**, 20:1631-1635.
367. TAYLOR, A.; CARMICHAEL, N. Toxic factor associated with egg-cultivated tumors. *Cancer Res.* **1960**, 20:1636-1639.
368. DALAL, U. C.; TAYLOR, A.; MCKENNA, G. F. The effect of plan extracts on egg cultivated tumor tissue. *Tex. Rep. Biol. Med.* **1958**, 16:439-442.
369. Murphy, J. B.; Rous, P. THE BEHAVIOR OF CHICKEN SARCOMA IMPLANTED IN THE DEVELOPING EMBRYO. *J. Exp. Med.* **1912**, 15:119-132.
370. TAYLOR, A.; CARMICHAEL, N. Egg cultivated tumor protects embryo against vaccinia virus. *Proc. Soc. Exp. Biol. Med.* **1953**, 83:676-678.
371. TAYLOR, A.; CARMICHAEL, N.; MCKENNA, G. F.; BURLAGE, H. M. Inhibition of the growth of egg cultivated tumor tissue by extracts of *Cooperia pedunculata* Herb. *Proc. Soc. Exp. Biol. Med.* **1951**, 77:841-843.
372. TAYLOR, A.; CARMICHAEL, N.; NORRIS, T. Temperature level and the growth of embryo and tumor of tumor-bearing eggs. *Proc. Soc. Exp. Biol. Med.* **1947**, 66:165-171.
373. TAYLOR, A.; CARMICHAEL, N. Stromal malignancy in mouse-grown transplants of egg-cultivated mouse mammary carcinoma. *Cancer Res.* **1947**, 7:78-87.
374. Murphy, J. B. Transplantability of malignant tumors to the embryos of a foreign species. *J. A. M. A.* **1912**, 59:874-875.

375. Kalirai, H.; Shahidipour, H.; Coupland, S. E.; Luyten, G. Use of the Chick Embryo Model in Uveal Melanoma. *Ocul. Oncol. Pathol.* **2015**, 1:133-140.
376. Ribatti, D. The chick embryo chorioallantoic membrane as a model for tumor biology. *Exp. Cell Res.* **2014**, 328:314-324.
377. Kain, K. H.; Miller, J. W.; Jones-Paris, C. R.; Thomason, R. T.; Lewis, J. D.; Bader, D. M.; Barnett, J. V.; Zijlstra, A. The chick embryo as an expanding experimental model for cancer and cardiovascular research. *Dev. Dyn.* **2014**, 243:216-228.
378. Jefferies, B.; Lenze, F.; Sathe, A.; Truong, N.; Anton, M.; von Eisenhart-Rothe, R.; Nawroth, R.; Mayer-Kuckuk, P. Non-invasive imaging of engineered human tumors in the living chicken embryo. *Sci. Rep.* **2017**, 7:4991-doi: 10.1038/s41598-017-04572-1.
379. Ribatti, D. The chick embryo chorioallantoic membrane (CAM) assay. *Reprod. Toxicol.* **2017**, 70:97-101.
380. Rovithi, M.; Avan, A.; Funel, N.; Leon, L. G.; Gomez, V. E.; Wurdinger, T.; Griffioen, A. W.; Verheul, H. M.; Giovannetti, E. Development of bioluminescent chick chorioallantoic membrane (CAM) models for primary pancreatic cancer cells: a platform for drug testing. *Sci. Rep.* **2017**, 7:44686-doi: 10.1038/srep44686.
381. Beedie, S. L.; Rore, H. M.; Barnett, S.; Chau, C. H.; Luo, W.; Greig, N. H.; Figg, W. D.; Vargesson, N. In vivo screening and discovery of novel candidate thalidomide analogs in the zebrafish embryo and chicken embryo model systems. *Oncotarget* **2016**, 7:33237-33245.
382. Kim, Y.; Williams, K. C.; Gavin, C. T.; Jardine, E.; Chambers, A. F.; Leong, H. S. Quantification of cancer cell extravasation in vivo. *Nat. Protoc.* **2016**, 11:937-948.
383. Haraguchi, S.; Matsubara, Y.; Hosoe, M. Chick embryos can form teratomas from microinjected mouse embryonic stem cells. *Dev. Growth Differ.* **2016**, 58:194-204.
384. Herrmann, A.; Moss, D.; See, V. The Chorioallantoic Membrane of the Chick Embryo to Assess Tumor Formation and Metastasis. *Methods Mol. Biol.* **2016**, 1464:97-105.
385. Deryugina, E. I. Chorioallantoic Membrane Microtumor Model to Study the Mechanisms of Tumor Angiogenesis, Vascular Permeability, and Tumor Cell Intravasation. *Methods Mol. Biol.* **2016**, 1430:283-298.
386. Ames, J. J.; Henderson, T.; Liaw, L.; Brooks, P. C. Methods for Analyzing Tumor Angiogenesis in the Chick Chorioallantoic Membrane Model. *Methods Mol. Biol.* **2016**, 1406:255-269.
387. LEIGHTON, J. INVASION AND METASTASIS OF HETEROLOGOUS TUMORS IN THE CHICK EMBRYO. *Prog. Exp. Tumor Res.* **1964**, 4:98-125.
388. Bracke, M. E.; Roman, B. I.; Stevens, C. V.; Mus, L. M.; Parmar, V. S.; De, W. O.; Mareel, M. M. Chick Heart Invasion Assay for Testing the Invasiveness of Cancer Cells and the Activity of Potentially Anti-invasive Compounds. *J. Vis. Exp.* **2015**:e52792-doi: 10.3791/52792.
389. Bracke, M. E.; Parmar, V. S.; Depass, A. L.; Stevens, C. V.; Vanhoecke, B. W.; Mareel, M. M. Chick heart invasion assay. *Methods Mol. Biol.* **2014**, 1070:93-106.
390. Bracke, M. E.; Boterberg, T.; Mareel, M. M. Chick heart invasion assay. *Methods Mol. Med.* **2001**, 58:91-102.
391. Engers, R.; Gerharz, C. D.; Donner, A.; Mrzyk, S.; Krause-Paulus, R.; Petek, O.; Gabbert, H. E. In vitro invasiveness of human epithelioid-sarcoma cell lines: association with cell motility and inverse correlation with the expression of tissue inhibitor of metalloproteinases. *Int. J. Cancer* **1999**, 80:406-412.
392. EASTY, G. C.; EASTY, D. M. AN ORGAN CULTURE SYSTEM FOR THE EXAMINATION OF TUMOR INVASION. *Nature* **1963**, 199:1104-1105.
393. Mareel, M.; Kint, J.; Meyvisch, C. Methods of study of the invasion of malignant C3H-mouse fibroblasts into embryonic chick heart in vitro. *Virchows Arch. B Cell Pathol. Incl. Mol. Pathol.* **1979**, 30:95-111.

394. SCHERER, W. F.; SYVERTON, J. T.; GEY, G. O. Studies on the propagation in vitro of poliomyelitis viruses. IV. Viral multiplication in a stable strain of human malignant epithelial cells (strain HeLa) derived from an epidermoid carcinoma of the cervix. *J. Exp. Med.* **1953**, 97:695-710.
395. Rubin, H.; Rubin, A. L. Phenotypic selection as the biological mode of epigenetic conversion and reversion in cell transformation. *Proc. Natl. Acad. Sci. U. S. A* **2018**, 115:E725-E732.
396. BILLINGHAM, R. E. TRANSPLANTATION: PAST, PRESENT AND FUTURE. *J. Invest Dermatol.* **1963**, 41:165-180.
397. Klein, G. The usefulness and limitations of tumor transplantation in cancer research: a review. *Cancer Res.* **1959**, 19:343-358.
398. KALISS, N. Immunological enhancement of tumor homografts in mice: a review. *Cancer Res.* **1958**, 18:992-1003.
399. HAY, L. J. A review of mammary carcinoma in mice and of transplantation of carcinoma. *Surg. Staff. Semin. U. S. Veterans. Adm Hosp. Minneap.* **1947**, 3:126-134.
400. BARRETT, M. K. Some immunogenetic influences upon transplanted tumors. *Cancer Res.* **1952**, 12:535-542.
401. BARRETT, M. K.; DERINGER, M. K.; HANSEN, W. H. Induced adaptation in a tumor: specificity of the change. *J. Natl. Cancer Inst.* **1953**, 14:381-394.
402. BARRETT, M. K.; DERINGER, M. K. An induced adaptation in a transplantable tumor of mice. *J. Natl. Cancer Inst.* **1950**, 11:51-59.
403. DUNHAM, L. J.; STEWART, H. L. A survey of transplantable and transmissible animal tumors. *J. Natl. Cancer Inst.* **1953**, 13:1299-1377.
404. GORER, P. A. Some recent work on tumor immunity. *Adv. Cancer Res.* **1956**, 4:149-186.
405. HAUSCHKA, T. S. Tissue genetics of neoplastic cell populations. *Proc. Can. Cancer Conf.* **1957**, 2:305-345.
406. HAUSCHKA, T. S. Methods of conditioning the graft in tumor transplantation. *J. Natl. Cancer Inst.* **1953**, 14:723-739.
407. HIRSCH, H. M. Some aspects of the problem of immunity against transplanted and spontaneous tumors. *Bacteriol. Rev.* 1962, 26:336-353.
408. HIRSCH, H. M. Tumor immunity and tissue transplantation. *J. Lancet* **1959**, 79:340-347.
409. HIRSCH, H. M. Tumor isoimmunity. *Experientia* **1958**, 14:269-271.
410. KALISS, N. The transplanted tumor as a research tool in cancer immunology. *Cancer Res.* **1961**, 21:1203-1208.
411. LAW, L. W. Genetic studies in experimental cancer. *Adv. Cancer Res.* **1954**, 2:281-352.
412. GREENE, H. S.; NEWTON, B. L. Evolution of cancer of the uterine fundus in the rabbit. *Cancer* **1948**, 1:82-99.
413. GREENE, H. S. On the development of cancer. *Sci. Am.* **1948**, 179:40-43.
414. GREENE, H. S. Heterologous transplantation of the Brown-Pearce tumors. *Cancer Res.* **1949**, 9:728-35.
415. GREENE, H. S. Attributes of embryonic tissues after growth and development in heterologous hosts. *Cancer Res.* **1955**, 15:170-172.
416. GREENE, H. S. Pathology in fields collateral to tissue culture. *J. Natl. Cancer Inst.* **1957**, 19:711-721.
417. GREENE, H. S. The significance of transplantability. *Trans. Stud. Coll. Physicians Phila.* **1957**, 24:101-104.
418. GREENE, H. S. Heterotransplantation of tumors. *Ann. N. Y. Acad. Sci.* **1957**, 69:818-829.
419. GREENE, H. S.; HARVEY, E. K. METASTASIS OF HETEROLOGOUSLY TRANSPLANTED TUMORS. *Cancer Res.* **1964**, 24:1678-1687.
420. Vivarelli, S.; Wagstaff, L.; Piddini, E. Cell wars: regulation of cell survival and proliferation by cell competition. *Essays Biochem.* **2012**, 53:69-82.

421. Wagstaff, L.; Kolahgar, G.; Piddini, E. Competitive cell interactions in cancer: a cellular tug of war. *Trends Cell Biol.* **2013**, 23:160-167.
422. Allard, D.; Stoker, M.; Gherardi, E. A G2/M cell cycle block in transformed cells by contact with normal neighbors. *Cell Cycle* **2003**, 2:484-487.
423. Flaberg, E.; Markasz, L.; Petranji, G.; Stuber, G.; Dicso, F.; Alchihabi, N.; Olah, E.; Csizy, I.; Jozsa, T.; Andren, O.; Johansson, J. E.; Andersson, S. O.; Klein, G.; Szekely, L. High-throughput live-cell imaging reveals differential inhibition of tumor cell proliferation by human fibroblasts. *Int. J. Cancer* **2011**, 128:2793-2802.
424. Furuta, S.; Jiang, X.; Gu, B.; Cheng, E.; Chen, P. L.; Lee, W. H. Depletion of BRCA1 impairs differentiation but enhances proliferation of mammary epithelial cells. *Proc. Natl. Acad. Sci. U. S. A* **2005**, 102:9176-9181.
425. Furuta, S.; Jeng, Y. M.; Zhou, L.; Huang, L.; Kuhn, I.; Bissell, M. J.; Lee, W. H. IL-25 causes apoptosis of IL-25R-expressing breast cancer cells without toxicity to nonmalignant cells. *Sci. Transl. Med.* **2011**, 3:78ra31-doi: 10.1126/scitranslmed.3001374.
426. Kosaka, N.; Iguchi, H.; Yoshioka, Y.; Hagiwara, K.; Takeshita, F.; Ochiya, T. Competitive interactions of cancer cells and normal cells via secretory microRNAs. *J. Biol. Chem.* **2012**, 287:1397-1405.
427. Stoker, M. REGULATION OF GROWTH AND ORIENTATION IN HAMSTER CELLS TRANSFORMED BY POLYOMA VIRUS. *Virology* **1964**, 24:165-174.
428. Stoker, M.; Gherardi, E.; Perryman, M.; Gray, J. Scatter factor is a fibroblast-derived modulator of epithelial cell mobility. *Nature* **1987**, 327:239-242.
429. Weaver, V. M.; Petersen, O. W.; Wang, F.; Larabell, C. A.; Briand, P.; Damsky, C.; Bissell, M. J. Reversion of the malignant phenotype of human breast cells in three-dimensional culture and in vivo by integrin blocking antibodies. *J. Cell Biol.* **1997**, 137:231-245.
430. Rubin, H. Rethinking "cancer as a dynamic developmental disorder" a quarter century later. *Cancer Res.* **2009**, 69:2171-2175.
431. Rubin, H. Microenvironmental regulation of the initiated cell. *Adv. Cancer Res.* **2003**, 90:1-62.
432. Rubin, H. Contact interactions between cells that suppress neoplastic development: can they also explain metastatic dormancy? *Adv. Cancer Res.* **2008**, 100:159-202.
433. Rubin, H. Ordered heterogeneity and its decline in cancer and aging. *Adv. Cancer Res.* **2007**, 98:117-147.
434. Aktipis, C. A.; Nesse, R. M. Evolutionary foundations for cancer biology. *Evol. Appl.* **2013**, 6:144-159.
435. Aktipis, C. A.; Boddy, A. M.; Jansen, G.; Hibner, U.; Hochberg, M. E.; Maley, C. C.; Wilkinson, G. S. Cancer across the tree of life: cooperation and cheating in multicellularity. *Philos. Trans. R. Soc. Lond B Biol. Sci.* **2015**, 370:-pii: 20140219. doi: 10.1098/rstb.2014.0219.
436. Thomas, F.; Kareva, I.; Raven, N.; Hamede, R.; Pujol, P.; Roche, B.; Ujvari, B. Evolved Dependence in Response to Cancer. *Trends Ecol. Evol.* **2018**, 33:269-276.
437. Thomas, F.; Vavre, F.; Tissot, T.; Vittecoq, M.; Giraudeau, M.; Bernex, F.; Misse, D.; Renaud, F.; Raven, N.; Beckmann, C.; Hamede, R.; Biro, P. A.; Ujvari, B. Cancer Is Not (Only) a Senescence Problem. *Trends Cancer* **2018**, 4:169-172.
438. Thomas, F.; Jacqueline, C.; Tissot, T.; Henard, M.; Blanchet, S.; Loot, G.; Dawson, E.; Mery, F.; Renaud, F.; Montagne, J.; Beckmann, C.; Biro, P. A.; Hamede, R.; Ujvari, B. The importance of cancer cells for animal evolutionary ecology. *Nat. Ecol. Evol.* **2017**, 1:1592-1595.
439. Hennings, H.; Robinson, V. A.; Michael, D. M.; Pettit, G. R.; Jung, R.; Yuspa, S. H. Development of an in vitro analogue of initiated mouse epidermis to study tumor promoters and antipromoters. *Cancer Res.* **1990**, 50:4794-4800.
440. Strickland, J. E.; Ueda, M.; Hennings, H.; Yuspa, S. H. A model for initiated mouse skin: suppression of papilloma but not carcinoma formation by normal epidermal cells in grafts on athymic nude mice. *Cancer Res.* **1992**, 52:1439-1444.

441. Lazebnik, Y. What are the hallmarks of cancer? *Nat. Rev. Cancer* **2010**, 10:232-233.
442. Tyzzer, E. E. A Series of spontaneous tumors in Mice with Observations on the Influence of Heredity on the Frequency of their Occurrence. *J. Med. Res.* **1909**, 21:479-518.
443. Tyzzer, E. E. A Study of Heredity in Relation to the Development of tumors in Mice. *J. Med. Res.* 1907, 17:199-211.
444. Farber, E.; Solt, D.; Cameron, R.; Laishes, B.; Ogawa, K.; Medline, A. Newer insights into the pathogenesis of liver cancer. *Am. J. Pathol.* **1977**, 89:477-482.
445. Farber, E. Pre-cancerous steps in carcinogenesis. Their physiological adaptive nature. *Biochim. Biophys. Acta* **1984**, 738:171-180.
446. Farber, E.; Rubin, H. Cellular adaptation in the origin and development of cancer. *Cancer Res.* **1991**, 51:2751-2761.
447. Farber, E. Cell proliferation as a major risk factor for cancer: a concept of doubtful validity. *Cancer Res.* **1995**, 55:3759-3762.
448. Farber, E. Cell proliferation is not a major risk factor for cancer. *Mod. Pathol.* **1996**, 9:606.
449. Farber, E. Risk assessment for possible carcinogens: a critical look. *Drug Metab. Rev.* **2000**, 32:143-151.
450. Chao, M. P.; Majeti, R.; Weissman, I. L. Programmed cell removal: a new obstacle in the road to developing cancer. *Nat. Rev. Cancer* **2012**, 12:58-67.
451. Sapienza, P.; Mallette, F. A. Cellular Senescence in Postmitotic Cells: Beyond Growth Arrest. *Trends Cell Biol.* **2018**:pii: S0962-8924(18)30059-X. doi: 10.1016/j.tcb.2018.03.003.
452. Childs, B. G.; Baker, D. J.; Kirkland, J. L.; Campisi, J.; van Deursen, J. M. Senescence and apoptosis: dueling or complementary cell fates? *EMBO Rep.* **2014**, 15:1139-1153.
453. Lopez-Otin, C.; Blasco, M. A.; Partridge, L.; Serrano, M.; Kroemer, G. The hallmarks of aging. *Cell* **2013**, 153:1194-1217.
454. van Deursen, J. M. The role of senescent cells in ageing. *Nature* **2014**, 509:439-446.
455. Vicencio, J. M.; Galluzzi, L.; Tajeddine, N.; Ortiz, C.; Criollo, A.; Tasdemir, E.; Morselli, E.; Ben, Y. A.; Maiuri, M. C.; Lavandro, S.; Kroemer, G. Senescence, apoptosis or autophagy? When a damaged cell must decide its path--a mini-review. *Gerontology* **2008**, 54:92-99.
456. Coppe, J. P.; Patil, C. K.; Rodier, F.; Sun, Y.; Munoz, D. P.; Goldstein, J.; Nelson, P. S.; Desprez, P. Y.; Campisi, J. Senescence-associated secretory phenotypes reveal cell-nonautonomous functions of oncogenic RAS and the p53 tumor suppressor. *PLoS. Biol.* **2008**, 6:2853-2868.
457. Krtolica, A.; Parrinello, S.; Lockett, S.; Desprez, P. Y.; Campisi, J. Senescent fibroblasts promote epithelial cell growth and tumorigenesis: a link between cancer and aging. *Proc. Natl. Acad. Sci. U. S. A* **2001**, 98:12072-12077.
458. Liu, D.; Hornsby, P. J. Senescent human fibroblasts increase the early growth of xenograft tumors via matrix metalloproteinase secretion. *Cancer Res.* **2007**, 67:3117-3126.
459. Childs, B. G.; Durik, M.; Baker, D. J.; van Deursen, J. M. Cellular senescence in aging and age-related disease: from mechanisms to therapy. *Nat. Med.* **2015**, 21:1424-1435.
460. Canino, C.; Mori, F.; Cambria, A.; Diamantini, A.; Germoni, S.; Alessandrini, G.; Borsellino, G.; Galati, R.; Battistini, L.; Blandino, R.; Facciolo, F.; Citro, G.; Strano, S.; Muti, P.; Blandino, G.; Ciocce, M. SASP mediates chemoresistance and tumor-initiating-activity of mesothelioma cells. *Oncogene* **2012**, 31:3148-3163.
461. Coppe, J. P.; Desprez, P. Y.; Krtolica, A.; Campisi, J. The senescence-associated secretory phenotype: the dark side of tumor suppression. *Annu. Rev. Pathol.* **2010**, 5:99-118.
462. Davalos, A. R.; Coppe, J. P.; Campisi, J.; Desprez, P. Y. Senescent cells as a source of inflammatory factors for tumor progression. *Cancer Metastasis Rev.* **2010**, 29:273-283.
463. Ghosh, K.; Capell, B. C. The Senescence-Associated Secretory Phenotype: Critical Effector in Skin Cancer and Aging. *J. Invest. Dermatol.* **2016**, 136:2133-2139.

464. Greten, T. F.; Eggert, T. Cellular senescence associated immune responses in liver cancer. *Hepat. Oncol.* **2017**, 4:123-127.
465. Lecot, P.; Alimirah, F.; Desprez, P. Y.; Campisi, J.; Wiley, C. Context-dependent effects of cellular senescence in cancer development. *Br. J. Cancer* **2016**, 114:1180-1184.
466. Pare, R.; Yang, T.; Shin, J. S.; Lee, C. S. The significance of the senescence pathway in breast cancer progression. *J. Clin. Pathol.* **2013**, 66:491-495.
467. Valenzuela, C. A.; Quintanilla, R.; Moore-Carrasco, R.; Brown, N. E. The Potential Role of Senescence As a Modulator of Platelets and Tumorigenesis. *Front. Oncol.* 2017, 7:188- doi: 10.3389/fonc.2017.00188.
468. Eriksson, L. C.; Blanck, A.; Bock, K. W.; Mannervik, B. Metabolism of xenobiotics in hepatocyte nodules. *Toxicol. Pathol.* **1987**, 15:27-42.
469. Carnero, A.; Blanco-Aparicio, C.; Kondoh, H.; Lleonart, M. E.; Martinez-Leal, J. F.; Mondello, C.; Scovassi, A. I.; Bisson, W. H.; Amedei, A.; Roy, R.; Woodrick, J.; Colacci, A.; Vaccari, M.; Raju, J.; Al-Mulla, F.; Al-Temaimi, R.; Salem, H. K.; Memeo, L.; Forte, S.; Singh, N.; Hamid, R. A.; Ryan, E. P.; Brown, D. G.; Wise, J. P., Sr.; Wise, S. S.; Yasaei, H. Disruptive chemicals, senescence and immortality. *Carcinogenesis* **2015**, 36 Suppl 1:S19-S37.
470. Maqsood, M. I.; Matin, M. M.; Bahrami, A. R.; Ghasroldasht, M. M. Immortality of cell lines: challenges and advantages of establishment. *Cell Biol. Int.* **2013**, 37:1038-1045.
471. Gunes, C.; Avila, A. I.; Rudolph, K. L. Telomeres in cancer. *Differentiation* **2017**, 99:41-50.
472. Russo, I.; Silver, A. R.; Cuthbert, A. P.; Griffin, D. K.; Trott, D. A.; Newbold, R. F. A telomere-independent senescence mechanism is the sole barrier to Syrian hamster cell immortalization. *Oncogene* **1998**, 17:3417-3426.
473. Garbe, J. C.; Vrba, L.; Sputova, K.; Fuchs, L.; Novak, P.; Brothman, A. R.; Jackson, M.; Chin, K.; LaBarge, M. A.; Watts, G.; Futscher, B. W.; Stampfer, M. R. Immortalization of normal human mammary epithelial cells in two steps by direct targeting of senescence barriers does not require gross genomic alterations. *Cell Cycle* **2014**, 13:3423-3435.
474. Klocke, R.; Gomez-Lechon, M. J.; Ehrhardt, A.; Mendoza-Figueroa, T.; Donato, M. T.; Lopez-Revilla, R.; Castell, J. V.; Paul, D. Establishment and characterization of immortal hepatocytes derived from various transgenic mouse lines. *Biochem. Biophys. Res. Commun.* **2002**, 294:864-871.
475. Vogelstein, B.; Kinzler, K. W. The multistep nature of cancer. *Trends Genet.* **1993**, 9:138-141.
476. Land, H.; Parada, L. F.; Weinberg, R. A. Tumorigenic conversion of primary embryo fibroblasts requires at least two cooperating oncogenes. *Nature* **1983**, 304:596-602.
477. Lazarov, M.; Kubo, Y.; Cai, T.; Dajee, M.; Tarutani, M.; Lin, Q.; Fang, M.; Tao, S.; Green, C. L.; Khavari, P. A. CDK4 coexpression with Ras generates malignant human epidermal tumorigenesis. *Nat. Med.* **2002**, 8:1105-1114.
478. Creton, S.; Aardema, M. J.; Carmichael, P. L.; Harvey, J. S.; Martin, F. L.; Newbold, R. F.; O'Donovan, M. R.; Pant, K.; Poth, A.; Sakai, A.; Sasaki, K.; Scott, A. D.; Schechtman, L. M.; Shen, R. R.; Tanaka, N.; Yasaei, H. Cell transformation assays for prediction of carcinogenic potential: state of the science and future research needs. *Mutagenesis* **2012**, 27:93-101.
479. Pickles, J. C.; Pant, K.; Mcginty, L. A.; Yasaei, H.; Roberts, T.; Scott, A. D.; Newbold, R. F. A mechanistic evaluation of the Syrian hamster embryo cell transformation assay (pH 6.7) and molecular events leading to senescence bypass in SHE cells. *Mutat. Res. Genet. Toxicol. Environ. Mutagen.* **2016**, 802:50-58.
480. Newbold, R. F.; Overell, R. W.; Connell, J. R. Induction of immortality is an early event in malignant transformation of mammalian cells by carcinogens. *Nature* **1982**, 299:633-635.
481. Newbold, R. F.; Overell, R. W. Fibroblast immortality is a prerequisite for transformation by EJ c-Ha-ras oncogene. *Nature* **1983**, 304:648-651.

482. Newbold, R. F. Multistep malignant transformation of mammalian cells by carcinogens: induction of immortality as a key event. *Carcinog. Compr. Surv.* **1985**, 9:17-28.
483. Newbold, R. F. Malignant transformation of mammalian cells in culture: delineation of stages and role of cellular oncogene activation. *IARC Sci. Publ.* **1985**, 67:31-53.
484. Newbold, R. F.; Cuthbert, A. P.; Themis, M.; Trott, D. A.; Blair, A. L.; Li, W. Cell immortalization as a key, rate-limiting event in malignant transformation: approaches toward a molecular genetic analysis. *Toxicol. Lett.* **1993**, 67:211-230.
485. Freedman, V. H.; Shin, S. I. Cellular tumorigenicity in nude mice: correlation with cell growth in semi-solid medium. *Cell* **1974**, 3:355-359.
486. Vogelstein, B.; Kinzler, K. W. Cancer genes and the pathways they control. *Nat. Med.* **2004**, 10:789-799.
487. Cobaleda, C.; Sanchez-Garcia, I. Stem cell aging and cancer: immortal but vulnerable. *Cell Cycle* **2011**, 10:2823-2824.
488. Alspach, E.; Fu, Y.; Stewart, S. A. Senescence and the pro-tumorigenic stroma. *Crit. Rev. Oncog.* **2013**, 18:549-558.
489. Pazolli, E.; Stewart, S. A. Senescence: the good the bad and the dysfunctional. *Curr. Opin. Genet. Dev.* **2008**, 18:42-47.
490. Ruhland, M. K.; Coussens, L. M.; Stewart, S. A. Senescence and cancer: An evolving inflammatory paradox. *Biochim. Biophys. Acta* **2016**, 1865:14-22.
491. Stewart, S. A.; Weinberg, R. A. Telomeres: cancer to human aging. *Annu. Rev. Cell Dev. Biol.* **2006**, 22:531-557.
492. Collado, M.; Serrano, M. Senescence in tumours: evidence from mice and humans. *Nat. Rev. Cancer* **2010**, 10:51-57.
493. Collado, M.; Blasco, M. A.; Serrano, M. Cellular senescence in cancer and aging. *Cell* **2007**, 130:223-233.
494. Collado, M.; Serrano, M. The power and the promise of oncogene-induced senescence markers. *Nat. Rev. Cancer* **2006**, 6:472-476.
495. Collado, M.; Serrano, M. The senescent side of tumor suppression. *Cell Cycle* **2005**, 4:1722-1724.
496. Collado, M.; Gil, J.; Efeyan, A.; Guerra, C.; Schuhmacher, A. J.; Barradas, M.; Benguria, A.; Zaballos, A.; Flores, J. M.; Barbacid, M.; Beach, D.; Serrano, M. Tumour biology: senescence in premalignant tumours. *Nature* **2005**, 436:642-doi:10.1038/436642a.
497. Soo, J. K.; Mackenzie Ross, A. D.; Kallenberg, D. M.; Milagre, C.; Heung, C. W.; Chow, J.; Hill, L.; Hoare, S.; Collinson, R. S.; Hossain, M.; Keith, W. N.; Marais, R.; Bennett, D. C. Malignancy without immortality? Cellular immortalization as a possible late event in melanoma progression. *Pigment Cell Melanoma Res.* **2011**, 24:490-503.
498. Thomas, F.; Nesse, R. M.; Gatenby, R.; Gidoin, C.; Renaud, F.; Roche, B.; Ujvari, B. Evolutionary Ecology of Organs: A Missing Link in Cancer Development? *Trends Cancer* **2016**, 2:409-415.
499. HAYFLICK, L. THE LIMITED IN VITRO LIFETIME OF HUMAN DIPLOID CELL STRAINS. *Exp. Cell Res.* **1965**, 37:614-636.
500. Polymenis, M.; Kennedy, B. K. Unbalanced Growth, Senescence and Aging. *Adv. Exp. Med. Biol.* **2017**, 1002:189-208.
501. Passaro, F.; Testa, G. Implications of Cellular Aging in Cardiac Reprogramming. *Front. Cardiovasc. Med.* **2018**, 5:43-doi: 10.3389/fcvm.2018.00043.
502. Shakeri, H.; Lemmens, K.; Gevaert, A. B.; De Meyer, G. R. Y.; Segers, V. Cellular senescence links aging and diabetes in cardiovascular disease. *Am. J Physiol Heart Circ. Physiol.* **2018**:-doi: 10.1152/ajpheart.00287.2018.
503. Falandry, C.; Bonnefoy, M.; Freyer, G.; Gilson, E. Biology of cancer and aging: a complex association with cellular senescence. *J. Clin. Oncol.* **2014**, 32:2604-2610.

504. Sikora, E.; Bielak-Zmijewska, A.; Mosieniak, G. Cellular senescence in ageing, age-related disease and longevity. *Curr. Vasc. Pharmacol.* **2014**, *12*:698-706.
505. Tan, F. C.; Hutchison, E. R.; Eitan, E.; Mattson, M. P. Are there roles for brain cell senescence in aging and neurodegenerative disorders? *Biogerontology* **2014**, *15*:643-660.
506. de Magalhaes, J. P.; Passos, J. F. Stress, cell senescence and organismal ageing. *Mech. Ageing Dev.* **2018**, *170*:2-9.
507. Faragher, R. G.; McArdle, A.; Willows, A.; Ostler, E. L. Senescence in the aging process. *F1000Res.* **2017**, *6*:1219-doi: 10.12688/f1000research.10903.1.
508. Rufini, A.; Tucci, P.; Celardo, I.; Melino, G. Senescence and aging: the critical roles of p53. *Oncogene* **2013**, *32*:5129-5143.
509. Bhatia-Dey, N.; Kanherkar, R. R.; Stair, S. E.; Makarev, E. O.; Csoka, A. B. Cellular Senescence as the Causal Nexus of Aging. *Front. Genet.* **2016**, *7*:13-doi: 10.3389/fgene.2016.00013.
510. Antoniou, A.; Hebrant, A.; Dom, G.; Dumont, J. E.; Maenhaut, C. Cancer stem cells, a fuzzy evolving concept: a cell population or a cell property? *Cell Cycle* **2013**, *12*:3743-3748.
511. Maenhaut, C.; Dumont, J. E.; Roger, P. P.; van Staveren, W. C. Cancer stem cells: a reality, a myth, a fuzzy concept or a misnomer? An analysis. *Carcinogenesis* **2010**, *31*:149-158.
512. Floor, S.; van Staveren, W. C.; Larsimont, D.; Dumont, J. E.; Maenhaut, C. Cancer cells in epithelial-to-mesenchymal transition and tumor-propagating-cancer stem cells: distinct, overlapping or same populations. *Oncogene* **2011**, *30*:4609-4621.
513. Dalerba, P.; Cho, R. W.; Clarke, M. F. Cancer stem cells: models and concepts. *Annu. Rev. Med.* **2007**, *58*:267-284.
514. Lapidot, T.; Sirard, C.; Vormoor, J.; Murdoch, B.; Hoang, T.; Caceres-Cortes, J.; Minden, M.; Paterson, B.; Caligiuri, M. A.; Dick, J. E. A cell initiating human acute myeloid leukaemia after transplantation into SCID mice. *Nature* **1994**, *367*:645-648.
515. Clarke, M. F.; Fuller, M. Stem cells and cancer: two faces of eve. *Cell* **2006**, *124*:1111-1115.
516. O'Connor, M. L.; Xiang, D.; Shigdar, S.; Macdonald, J.; Li, Y.; Wang, T.; Pu, C.; Wang, Z.; Qiao, L.; Duan, W. Cancer stem cells: A contentious hypothesis now moving forward. *Cancer Lett.* **2014**, *344*:180-187.
517. Smalley, M.; Ashworth, A. Stem cells and breast cancer: A field in transit. *Nat. Rev. Cancer* **2003**, *3*:832-844.
518. van Staveren, W. C.; Solis, D. Y.; Hebrant, A.; Detours, V.; Dumont, J. E.; Maenhaut, C. Human cancer cell lines: Experimental models for cancer cells in situ? For cancer stem cells? *Biochim. Biophys. Acta* **2009**, *1795*:92-103.
519. Brinckerhoff, C. E. Cancer Stem Cells (CSCs) in melanoma: There's smoke, but is there fire? *J. Cell Physiol* **2017**, *232*:2674-2678.
520. Waring, R. H.; Harris, R. M.; Mitchell, S. C. In utero exposure to carcinogens: Epigenetics, developmental disruption and consequences in later life. *Maturitas* **2016**, *86*:59-63.
521. Lemercier, C.; To, R. Q.; Swanson, B. J.; Lyons, G. E.; Konieczny, S. F. Mist1: a novel basic helix-loop-helix transcription factor exhibits a developmentally regulated expression pattern. *Dev. Biol.* **1997**, *182*:101-113.
522. Tuveson, D. A.; Zhu, L.; Gopinathan, A.; Willis, N. A.; Kachatrian, L.; Grochow, R.; Pin, C. L.; Mitin, N. Y.; Taparowsky, E. J.; Gimotty, P. A.; Hruban, R. H.; Jacks, T.; Konieczny, S. F. Mist1-KrasG12D knock-in mice develop mixed differentiation metastatic exocrine pancreatic carcinoma and hepatocellular carcinoma. *Cancer Res.* **2006**, *66*:242-247.
523. Chiodi, I.; Belgiovine, C.; Dona, F.; Scovassi, A. I.; Mondello, C. Drug treatment of cancer cell lines: a way to select for cancer stem cells? *Cancers (Basel)* **2011**, *3*:1111-1128.
524. Pattabiraman, D. R.; Weinberg, R. A. Tackling the cancer stem cells - what challenges do they pose? *Nat. Rev. Drug Discov.* **2014**, *13*:497-512.

525. Bakhshinyan, D.; Adile, A. A.; Qazi, M. A.; Singh, M.; Kameda-Smith, M. M.; Yelle, N.; Chokshi, C.; Venugopal, C.; Singh, S. K. Introduction to Cancer Stem Cells: Past, Present, and Future. *Methods Mol. Biol.* **2018**, 1692:1-16.
526. Mertins, S. D. Cancer stem cells: a systems biology view of their role in prognosis and therapy. *Anticancer Drugs* **2014**, 25:353-367.
527. Muinao, T.; Deka Boruah, H. P.; Pal, M. Diagnostic and Prognostic Biomarkers in ovarian cancer and the potential roles of cancer stem cells - An updated review. *Exp. Cell Res.* **2018**, 362:1-10.
528. Schwitalla, S. Tumor cell plasticity: the challenge to catch a moving target. *J. Gastroenterol.* **2014**, 49:618-627.
529. Valle, S.; Martin-Hijano, L.; Alcalá, S.; Alonso-Nocelo, M.; Sainz, B., Jr. The Ever-Evolving Concept of the Cancer Stem Cell in Pancreatic Cancer. *Cancers (Basel)* **2018**, 10:pii: E33. doi: 10.3390/cancers10020033.
530. Chaffer, C. L.; Weinberg, R. A. How does multistep tumorigenesis really proceed? *Cancer Discov.* **2015**, 5:22-24.
531. Ksiazek, K. Let's stop overlooking bacterial aging. *Biogerontology* **2010**, 11:717-723.
532. Gomez, J. M. Aging in bacteria, immortality or not-a critical review. *Curr. Aging Sci.* 2010, 3:198-218.
533. Florea, M. Aging and immortality in unicellular species. *Mech. Ageing Dev.* **2017**, 167:5-15.
534. Hallsworth, J. E. Stress-free microbes lack vitality. *Fungal. Biol.* **2018**, 122:379-385.
535. da Silva-Diz, V.; Lorenzo-Sanz, L.; Bernat-Peguera, A.; Lopez-Cerda, M.; Munoz, P. Cancer cell plasticity: Impact on tumor progression and therapy response. *Semin. Cancer Biol.* **2018**, 53:48-58.
536. Zhang, Y.; Toy, K. A.; Kleer, C. G. Metaplastic breast carcinomas are enriched in markers of tumor-initiating cells and epithelial to mesenchymal transition. *Mod. Pathol.* **2012**, 25:178-184.
537. Barnes, P. J.; Boutilier, R.; Chiasson, D.; Rayson, D. Metaplastic breast carcinoma: clinical-pathologic characteristics and HER2/neu expression. *Breast Cancer Res. Treat.* **2005**, 91:173-178.
538. Catroppo, J. F.; Lara, J. F. Metastatic metaplastic carcinoma of the breast (MCB): an uncharacteristic pattern of presentation with clinicopathologic correlation. *Diagn. Cytopathol.* **2001**, 25:285-291.
539. Chhieng, C.; Cranor, M.; Lesser, M. E.; Rosen, P. P. Metaplastic carcinoma of the breast with osteocartilaginous heterologous elements. *Am. J. Surg. Pathol.* **1998**, 22:188-194.
540. Sasano, H.; Shizawa, S.; Nagura, H.; Yamaki, T. Mucinous adenocarcinoma arising in a giant urachal cyst associated with pseudomyxoma peritonei and stromal osseous metaplasia. *Pathol. Int.* **1997**, 47:502-505.
541. Eble, J. N.; Young, R. H. Carcinoma of the urinary bladder: a review of its diverse morphology. *Semin. Diagn. Pathol.* **1997**, 14:98-108.
542. Haque, S.; Eisen, R. N.; West, A. B. Heterotopic bone formation in the gastrointestinal tract. *Arch. Pathol. Lab. Med.* **1996**, 120:666-670.
543. Kirchhof, N.; Steinhauer, D.; Fey, K. Equine adenocarcinomas of the large intestine with osseous metaplasia. *J. Comp. Pathol.* **1996**, 114:451-456.
544. Bennett, J. H.; Jones, J.; Speight, P. M. Odontogenic squamous cell carcinoma with osseous metaplasia. *J. Oral Pathol. Med.* **1993**, 22:286-288.
545. Hanada, M.; Nakano, K.; Ii, Y.; Yamashita, H. Carcinosarcoma of the esophagus with osseous and cartilagenous production. A combined study of keratin immunohistochemistry and electron microscopy. *Acta Pathol. Jpn.* **1984**, 34:669-678.
546. Caluori, D.; Gallo, P. Case report of heterotopic bone formation in metastatic carcinoma of the colon. *Tumori.* **1979**, 65:345-351.
547. Groisman, G. M.; Benkov, K. J.; Adsay, V.; Dische, M. R. Osseous metaplasia in benign colorectal polyps. *Arch. Pathol. Lab Med.* **1994**, 118:64-65.
548. Fox, E. J.; Prindle, M. J.; Loeb, L. A. Do mutator mutations fuel tumorigenesis? *Cancer Metastasis Rev.* **2013**, 32:353-361.

549. FURTH, J.; Kahn, M. C. The transmission of leukemia of mice with a single cell. *Am. J. Cancer* **1937**, 31:276-282.
550. KLEINSMITH, L. J.; PIERCE, G. B., Jr. MULTIPOTENTIALITY OF SINGLE EMBRYONAL CARCINOMA CELLS. *Cancer Res.* **1964**, 24:1544-1551.
551. van den Brenk, H. A. Effect of immunological attenuation on cell dosage required to establish single or double tumour homografts. *Br. J. Cancer* **1961**, 15:798-803.
552. ISHIBASHI, K. Studies on the number of cells necessary for the transplantation of Yoshida sarcoma; transmission of the tumor with a single cell. *Gan* **1950**, 41:1-14.
553. HEWITT, H. B. Transplantation of mouse sarcoma with small numbers of single cells. *Nature* **1952**, 170:622-623.
554. HEWITT, H. B. Studies of the quantitative transplantation of mouse sarcoma. *Br. J. Cancer* **1953**, 7:367-383.
555. Miyahira, A. K.; Den, R. B.; Carlo, M. I.; de, L. R.; Hope, T. A.; Karzai, F.; McKay, R. R.; Salami, S. S.; Simons, J. W.; Pienta, K. J.; Soule, H. R. Tumor cell heterogeneity and resistance; report from the 2018 Coffey-Holden Prostate Cancer Academy Meeting. *Prostate* **2019**, 79:244-258.
556. Lawson, D. A.; Kessenbrock, K.; Davis, R. T.; Pervolarakis, N.; Werb, Z. Tumour heterogeneity and metastasis at single-cell resolution. *Nat. Cell Biol.* **2018**, 20:1349-1360.
557. Kusoglu, A.; Biray, A. C. Cancer stem cells: A brief review of the current status. *Gene* **2019**, 681:80-85.
558. Ben-David, U.; Beroukhim, R.; Golub, T. R. Genomic evolution of cancer models: perils and opportunities. *Nat. Rev. Cancer* **2019**, 19:97-109.
559. O'Connor, C. J.; Chen, T.; Gonzalez, I.; Cao, D.; Peng, Y. Cancer stem cells in triple-negative breast cancer: a potential target and prognostic marker. *Biomark. Med.* **2018**, 17:813-820.
560. Condiotti, R.; Guo, W.; Ben-Porath, I. Evolving views of breast cancer stem cells and their differentiation States. *Crit. Rev. Oncog.* **2014**, 19:337-348.
561. Nakshatri, H.; Srour, E. F.; Badve, S. Breast cancer stem cells and intrinsic subtypes: controversies rage on. *Curr. Stem Cell Res. Ther.* **2009**, 4:50-60.
562. Lapinska, K.; Faria, G.; McGonagle, S.; Macumber, K. M.; Heerboth, S.; Sarkar, S. Cancer Progenitor Cells: The Result of an Epigenetic Event? *Anticancer Res.* **2018**, 38:1-6.
563. Hanahan, D.; Weinberg, R. A. Hallmarks of cancer: the next generation. *Cell* **2011**, 144:646-674.
564. Weinberg, R. A. Coming full circle-from endless complexity to simplicity and back again. *Cell* **2014**, 157:267-271.
565. Blagosklonny, M. V. Cell immortality and hallmarks of cancer. *Cell Cycle* 2003, 2:296-299.
566. Durgan, J.; Florey, O. Cancer cell cannibalism: Multiple triggers emerge for entosis. *Biochim. Biophys. Acta* **2018**, 1865:831-841.
567. Krajcovic, M.; Overholtzer, M. Mechanisms of ploidy increase in human cancers: a new role for cell cannibalism. *Cancer Res.* **2012**, 72:1596-1601.
568. Lozupone, F.; Fais, S. Cancer Cell Cannibalism: A Primeval Option to Survive. *Curr. Mol. Med.* **2015**, 15:836-841.
569. Matarrese, P.; Ciarlo, L.; Tinari, A.; Piacentini, M.; Malorni, W. Xeno-cannibalism as an exacerbation of self-cannibalism: a possible fruitful survival strategy for cancer cells. *Curr. Pharm. Des.* **2008**, 14:245-252.
570. Sharma, N.; Dey, P. Cell cannibalism and cancer. *Diagn. Cytopathol.* 2011, 39:229-233.
571. Blagosklonny, M. V. NCI's provocative questions on cancer: some answers to ignite discussion. *Oncotarget* **2011**, 2:1352-1367.
572. Mareel, M. M.; Van Roy, F. M.; De, B. P. The invasive phenotypes. *Cancer Metastasis Rev.* **1990**, 9:45-62.

573. Rhim, A. D.; Mirek, E. T.; Aiello, N. M.; Maitra, A.; Bailey, J. M.; McAllister, F.; Reichert, M.; Beatty, G. L.; Rustgi, A. K.; Vonderheide, R. H.; Leach, S. D.; Stanger, B. Z. EMT and dissemination precede pancreatic tumor formation. *Cell* **2012**, 148:349-361.
574. Deryugina, E. I.; Kiosses, W. B. Intratumoral Cancer Cell Intravasation Can Occur Independent of Invasion into the Adjacent Stroma. *Cell Rep.* **2017**, 19:601-616.
575. Podsypanina, K.; Du, Y. C.; Jechlinger, M.; Beverly, L. J.; Hambardzumyan, D.; Varmus, H. Seeding and propagation of untransformed mouse mammary cells in the lung. *Science* **2008**, 321:1841-1844.
576. Weinberg, R. A. Leaving home early: reexamination of the canonical models of tumor progression. *Cancer Cell* **2008**, 14:283-284.
577. Liao, D. Z.; Porsch-Hallstrom, I.; Gustafsson, J. A.; Blanck, A. Persistent sex differences in growth control of early rat liver lesions are programmed during promotion in the resistant hepatocyte model. *Hepatology* **1996**, 23:835-839.

Fig 1: The Solt-Farber's "resistant hepatocyte" model of liver carcinogenesis in the rat. A toxic dose of diethylnitrosamine (DEN) will 1) cause liver necrosis and 2) create initiated hepatocytes. Two weeks later, when the liver has recovered from the necrosis, the rat will be given a low dose of 2-acetylaminofluorene (AAF) for two weeks, function of which is to inhibit proliferation, so-called mitoinhibition, of hepatocytes, but initiated cells are resistant to this inhibition. In the middle of AAF treatment, hepatectomy will be performed to remove two-thirds of the liver, which provides a strong impetus for regeneration. Because normal hepatocytes are mitoinhibited, all regeneration pressure is imposed onto the initiated cells, driving them to proliferate robustly and form nodules. The image at the left shows these nodules immunohistochemically stained for the P form of glutathione S transferase, a marker for the nodular cells, in the three remaining lobes of the liver four weeks post cessation of AAF treatment [187, 577]. These nodules will regress afterwards but some new focal cells, which can proliferate spontaneously and are coined by Farber as "phenotype 4", will later develop at some of the sites of these nodules and progress to overt cancers [310, 311].

Fig 2: Illustration of a speculative difference at the time point for the establishment of immortality and autonomy between tumorigenesis in most animal models and that in most human situations. In humans, immortality (Immort.) and autonomy (Auto.) may occur at a very early time point, thus establishing small lesions as genuinely benign or malignant neoplasms. In contrast, tumorigenesis in most animal models is a stepwise procedure of initiation, promotion and, in some cases, progression as well. Initiated cells are still mortal and thus are not neoplastic. Immortality and autonomy in animal models occur at late promotion or at the progression.

Fig 3: Illustration of our three-hit hypothesis. To couple the traditional two-hit principle with the initiation-promotion theory, we meditate that the first genetic hit establishes initiated cells that are still mortal and non-autonomous, whereas the second hit creates immortality and autonomy that establish neoplastic cells, either benign or malignant. Since formation of benign neoplasms also requires two genetic hits, we extrapolate that, in some animal models and probably also in many human situations, establishment of malignant morphologies and behaviors requires a third hit on the relevant gene(s).

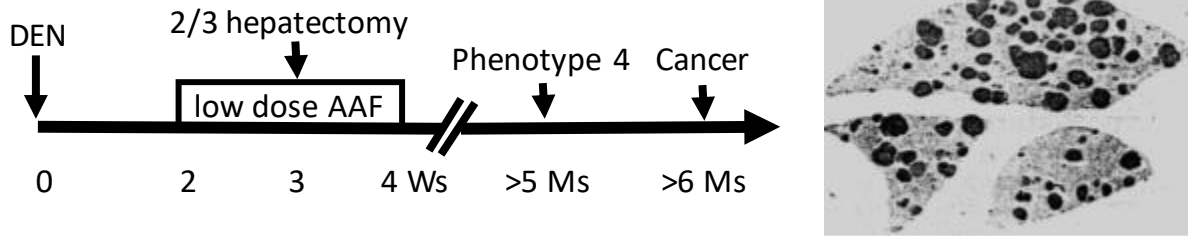


Figure 1

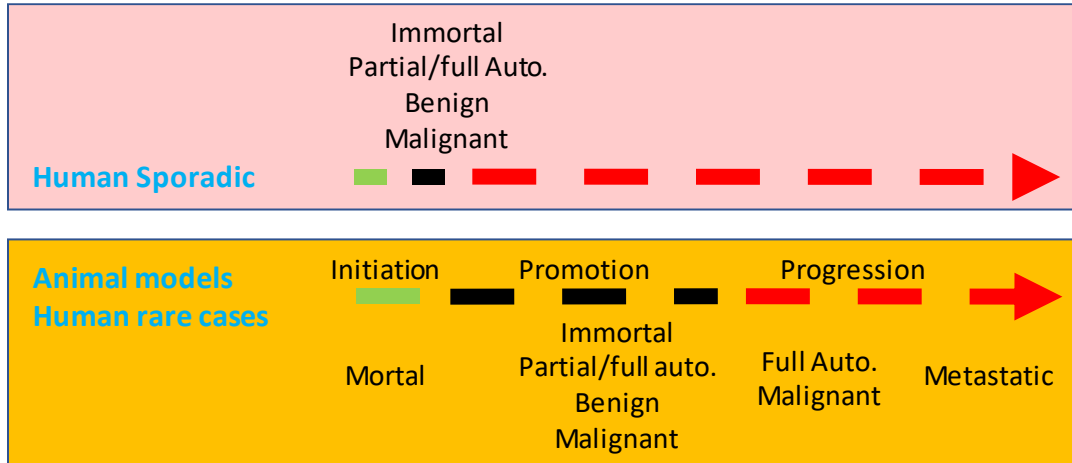


Figure 2

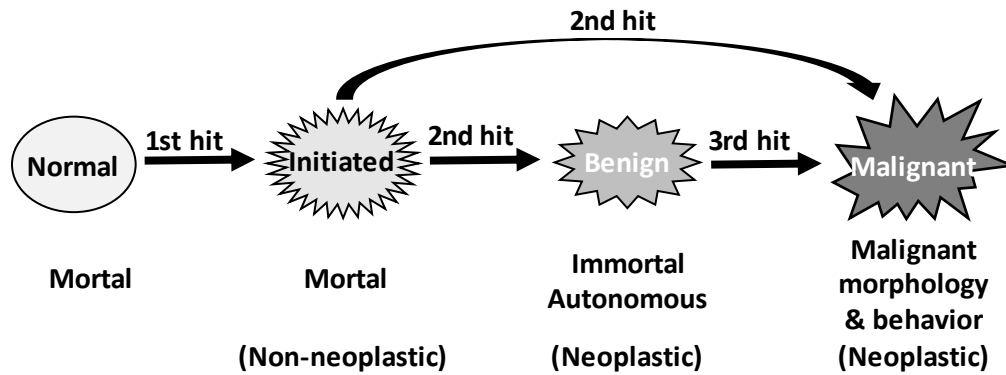


Figure 3