1 Article

Frequency of CYP3A5 genetic polymorphisms and Tacrolimus pharmacokinetics in Pediatric Liver Transplantation

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18 Abstract: The body of evidence available in paediatrics population is limited for making clinical 19 decisions regarding pharmacotherapy optimization of tacrolimus. The objective of this study was 20 to estimate the frequency of CYP3A5 genetic polymorphisms and their relationship with 21 tacrolimus requirements in paediatric population. This was a longitudinal cohort study, with two-22 year follow-up of 77 patients under 18 who had liver transplant over the period 2009-2012 at the 23 Paediatric Hospital J. P Garrahan. Tacrolimus levels from day 5 to 2-year post-transplant were 24 obtained from hospital records of routine therapeutic drug monitoring. The genotyping of CYP3A5 25 (CYP3A5*1/*3 or *3/*3) were performed in liver biopsies of both the donor and the recipient. 26 Recipients frequency of CYP3A5 *1 expression was 37.1% and 32.2% for Donors. Patient who 27 received an organ expresser showed lower Co/dose especially after 90 days post-surgery. The 28 role of each polymorphism is different according to days after transplantation proceeds and it 29 must be taken into account to optimize the benefits of TAC therapy during the post-transplant 30 induction and maintenance phase.

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32 Keywords: tacrolimus, CYP3A5, liver transplant, pharmacokinetics

34 1. Introduction

35 Tacrolimus (TAC) is a calcineurin inhibitor widely used in solid organ transplantation. TAC has a narrow therapeutic margin and a large intra- and inter -individual variability (1, 2). Incidence 36 37 of rejection and adverse effects remains as problems despite therapeutic drug monitoring of TAC 38 (3). There is growing interest in developing markers those will allow to individualize treatment of 39 TAC. Within this group of potential biomarkers a remarkable example are single nucleotide 40 polymorphisms of CYP3A5 (3-5). This enzyme has a highly polymorphic expression with at least 11 single nucleotide polymorphisms (SNPs) documented (3). The SNP most studied is the 41 42 transition from adenine to guanine at position 6986- intron 3 - CYP3A5 gene (rs 776746), also 43 called CYP3A5*1. This allele is associated with high levels of CYP3A5-mRNA and full functional 44 CYP3A5-protein (6, 7). Caucasian population expresses CYP3A5*1 between 10-40% while Asian 45 population expresses between 50-70 % (8). The CYP3A5*1 (homozygotes and heterozygotes) expressers require much higher daily doses of TAC as well as more time to reach desired serum 46

47 levels of TAC. What is more, expressers have three times the risk of acute rejection within the48 first month after transplant than no-expressers (9).

After liver transplant , simultaneous expression of CYP3A5*1 in both the intestine and the 49 50 implanted liver may occur (3). In a previous study of adult population we showed the interaction 51 does occur: Expression of CYP3A5*1 present in liver donor has great impact on TAC levels 52 adjusted by dose in long-term concentrations; while also the expression of this SNP in the receiver 53 has a greater impact but in time just after transplantation (8). However, kinetic and 54 pharmacodynamic are very different comparing paediatric to adult populations. This can be 55 explained by the greater variability of specific enzymes, which are acquired by the child during 56 growth and altering the clinical response to TAC (3). The body of evidence available in paediatrics 57 population is limited for making clinical decisions regarding the therapeutic optimization of TAC. 58 Thus it is essential to generate more information to optimize and customize monitoring strategies 59 to liver transplant in this population. The objective of this study was to estimate the frequency of 60 CYP3A5 genetic polymorphisms and their relationship with pharmacokinetics in Pediatric Liver 61 Transplantation

62 2. Materials and Methods

A longitudinal study was conducted in f 77 patients under 18 who after liver transplantation
 over the period 2009-2012 at the Paediatric Hospital J. P Garrahan (PHJPG)

Were included patients with full or partial liver graft, from either living donor or cadaveric donor. All patient were receiving tacrolimus with or without steroids and with or without mofetil mycophenolate. Were excluded HIV infected patients, who suffered early death before receiving immunosuppressive regimen with TAC in the immediate postsurgical and patients with partial or total loss of medical records.

70 2.1. Dosage and treatment scheme:

71 Patient information was collected immediately after liver transplantation. Below it is 72 described the scheme of immunosuppression performed in patients according to the Clinical 73 Practice Guidelines of PHJPG for patients after liver transplantation. In induction phase all 74 patients received basiliximab. Patient under 30 kg received 10 mg/dose and over 30 kg received 75 20 mg/dose. Both doses were administered as an intravenous bolus; the former within 8 hours 76 after reperfusion of the graft and the later the fourth post-surgery day. TAC was dispensed in the 77 maintenance phase which started 24 hours after reperfusion. The initial oral regimen was 0.1 78 mg/kg/day every 12 hours. Afterwards the dose of TAC was adjusted to tacrolimus blood levels, 79 liver parameters, kidney function and the viral load of Epstein Barr Virus (10). In patients without 80 infectious activity (viral load less than 4000 copies/ ug DNA) and creatinine clearance less than 81 the expected range for your age, the initial desired TAC blood levels were 8-12 ng/ml during the 82 first month after transplantation (11). It was proceeded a quick immunosuppression reduction in 83 patients with viral load above 4000 copies / ug DNA in 2 consecutive samples or clinical evidence 84 of EBV infection. No antiviral therapy was implemented. In patients who developed renal toxicity, 85 regardless viral load, monitoring of TAC was decreased a 25%. In those cases mycophenolate mofetil (MMF) was added as rescue therapy with an initial dose of 20 mg/kg/day and then it was 86 87 increased up to 40 mg/kg/day after a week of treatment.

88 2.2. Monitoring and quantification of tacrolimus blood levels.

TAC levels from day 5 to 2-year post-transplant were obtained from hospital records of routine therapeutic drug monitoring. The values recorded correlate to monitoring blood levels from samples drawn prior to the morning dose (Co) (C0:is a concentration measured in t=0, before the first dose of the drug).

93 Quantification of TAC was performed by chemiluminescence immunoassay by Architect i1000 of

94 Abbott according to the manufacturer's instructions. The low quantification limit was 2.0 ng/ml.

The linearity was observed from 2-30 ng/ml. The variation coefficient for Quality Control Sampleswas below 6%.

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98 2.3. Information collected.

99 Demographic information (date of birth, gender), anthropometric data (weight, height), 100 indication of transplant, post- transplantation follow-up time; current medication and doses, 101 concomitant medications (The effect of drug drug interaction was analyzed in a previously 102 published article) (12-13), amount of transplanted graft; , amount of postsurgical days, data 103 related to donor type were collected. We registered clinical laboratory results including 104 hematology (hemoglobin, hematocrit, red blod cells, white cells and platelets, RIN) and clinical 105 chemistry results (creatinine, urea nitrogen, total and direct bilirubin, alkaline phosphatase, 106 alanine aminotransferase- GPT or ALT- aspartate aminotransferase- GOT or AST- gamma 107 glutamyl transpeptidase-GGT- and albumin).

108 2.4. DNA isolation and genotyping.

109 The genotyping of CYP3A5 were performed in liver biopsies of both the donor and the 110 recipient. The donor's DNA was obtained from liver biopsies or surgical specimens obtained from 111 the Pathology Service of PHJPG. Each of them were tissue fixed in formalin-buffer, paraffin 112 embedded and sectioned by 10 microns thick.

DNA extraction was performed using commercial kits QIAamp DNA Blood Kit and QIAamp DNA FFPE Tissue following the manufacturer's instructions. We obtained from 20 to 100 ng of DNA in each case. The CYP3A5*3 (rs776746) polymorphism was detected by PCR and directly sequenced. Patients with variants (CYP3A5*1/*1 or CYP3A5*1/*3) were called 'expressers' while those with variants CYP3A5*3/*3 were called 'not expressers'.

118 2.5. Ethical aspects.

119 The proper Informed Consent was signed by a parent or legal guardian before starting any 120 specific evaluations. The study was approved by the office of Teaching and Research of 121 PHJPGand by the Ethics Committee of the Faculty of Pharmacy and Biochemistry, University of 122 Buenos Aires.

123 2.6. Statistical analysis.

We compared daily doses of TAC, Co (TAC levels prior to the morning dose) and Co/dose (concentration adjusted by dose) according to CYP3A5 *1 allele expression between donors and recipients. All values were expressed as mean ± standard deviation. U Mann -Whitney test was used to determine differences between continuous variables among groups. The chi- square test was used to analyze differences between discrete variables. Analysis were performed using STATA 11.0 ©.

130 3. Results

We evaluated 77 paediatric patients medicated with TAC during the first 2 years after transplantation. **Table 1** shows the characteristics of the population studied. We observed 45 patients (58.44 %) with adverse events associated with tacrolimus, 51 patients (66.23 %) had at least one acute cellular rejection episode and 8 patients died (10.39%) during follow-up.

CYP3A5 *1 expression was 37.1% in recipients and 32.2% for Donors. There were not shown
 statistically significant deviations in the distribution of polymorphisms according to the Hardy Weinberg principle (p > 0.05).

A total of 3670 blood concentrations of TAC were analysed during the study period, with a mean of 47.8 samples per patient. We observed a greater difference in expressers recipients

regarding not expressers, especially in the first two weeks postoperative, and tend to reduce thosedifferences over time, see Figure 1.

When adjusted dose by concentrations according to the genotype of the donor, those who
 received an organ expresser showed lower Co/dose especially after 90 days post-surgery. See
 Figure 2. A statistically significant reduction in the Co / dose of 0.00063 ng/ml mg/kg/day was
 observed in comparison with those receiving an organ not expresser (p=0.001).

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Table 1 Characteristics of the studied population (n=77).

Feature	n(%)
Female	46 (59.74)
Age at transplantation (years, ± DE)	5.32 (5.42)
Weight (Kg , ± DE)	21.84(17.89)
Origin	
Argentina	64(83.11)
Bolivia	2(2.60)
Paraguay	9(11.69)
Other	2(2.59)
Primary illness	
Biliary atresia	32(41.55)
Fulminant hepatitis	16(20.77)
Autoimmune hepatitis	11(14.28)
Hepatoblastoma	8(10.38)
Others	10(12.98)
Kind of Donor	
Cadaveric	55(71.42)
Alive	22(28.57)
Kind of Graft	
Full	26(33.76)
Technical variant	51(66.23)

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Figure 1. Temporal behaviour of Co / dose of TAC according recipient genotype.



Co / dose of TAC according donor genotype.



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Figure 2. Temporal behaviour of Co / dose of TAC according donor genotype.

156 4. Discussion

The CYP3A5 polymorphisms has a differential impact in the pharmacokinetics of tacrolimus according it expression in donors and recipients . In contrast to previous studies this is the first study in the paediatric patients that evaluates the effect of polymorphisms on TAC pharmacokinetics at long term. Other works considered shorter periods and generally not Hispanic population.

162 Patients with CYP3A5 allele A (CYP3A5 * 1 or wild type) have a normal splicing of the whole 163 13 exons in this gene. This results in a normal transcript and producing high levels of mRNA, thus 164 expressing the enzyme metabolizing TAC. Patients with allele G (CYP3A5 * 3) have a point 165 mutation (A / G) resulting in the insertion of an inappropriate 'exon' 3B within the transcript. This 166 new exon introduces an early termination codon, leading to a non-functional protein fragment 167 (14). The frequency of expressers (CYP3A5 * 1) in our study was reported to be intermediate 168 between Asian frequency (33% to 66%) and Caucasian (9% to 15%) populations. These 169 estimates are consistent with previous results in studies in Argentine renal transplant patients 170 ,which reported values ranging from 9% to 27%(15) (16) (17). These differences between the 171 caucasian and asian frequencies, reveal the genetic diversity present in latin america as a result 172 from the colonial stage, African (slaves to the 19th century), and post-independence immigrants (the majority of Spain, Italy, France, Europe from the east) (18). 173

Similar results have been found in studies focused on the frequency of variations in other genes related to antineoplastic metabolism (19). Continue to building this pharmacogenetic map in latin america improve the understanding of the variations in the metabolism and the effect of the different medicines, without the need to extrapolate results obtained from other populations.

178 In liver transplanted patients, both donor and recipient carrying the CYP3A5 polymorphisms 179 are associated with changes in the pharmacokinetics of TAC. However, the role of each 180 polymorphism is different according to days after transplantation. We have shown that the recipient CYP3A5 genotype plays a more important role than the donor genotype. Recipients 181 182 CYP3A5 * 1 achieved lower blood concentrations of TAC and lower dose-adjusted with 183 concentrations despite the medical pharmacotherapeutic follow-up (based on adjusting the blood 184 concentrations to the reference therapeutic margins). These findings are consistent with a recent 185 study of 64 post-transplant children with 1 year follow-up (20). It was shown that lower dose-

adjusted (p <0.05) concentrations are required in patients who are expressers, without correlation
 with donor genotype, especially in the first 7 days after transplantation (20).

188 To recognize the role played by the recipient CYP3A5 genotype in the first weeks after 189 transplantation is essential to avoid excessive dose increases to patients who are expressers of 190 this genotype (21).

191 In contrast to receipt, the donor genotype alter significantly kinetics of TAC increasing along 192 with time after transplantation. The effect of CYP3A5 expression on the recipient is an augmented 193 hepatic clearance of the liver implanted with the polymorphism. This tendency was evidenced in 194 our study: a reduction of dose-adjusted concentrations was documented statistically significant 195 after 60 to 90 postoperative days. Such observation might be attached to the time needed by the 196 organ to recover from the ischemia and reperfusion injury, regeneration and graft growth as the 197 months after transplantation occur (22). Our results indicate the importance of know the genotype 198 present in the organ previously to be implanted. This is a priority during the ambulatory follow-up 199 to select patients with greater hepatic clearance, who will get a lower concentration and which 200 may require different medical follow-up to avoid sub-immunosuppression.

201 Our study has limitations mainly given the retrospective nature. Among which are biases 202 due to misclassification of patients either by memory bias or problems to record information in 203 clinical histories- by omitting information or incorrectly record said documents-. There biases 204 could be minimized obtaining always information from primary registers (physical or electronic 205 medical history) and checking with other clinical records (nursing records, pharmacy Hospital). 206 We only analyse concentration at time 0 (C0) per patient due to we use data hospital therapeutic 207 monitoring, and for TAC this concentration is used to clinical monitoring for this drugs. Also effect 208 of other variables in the pharmacokinetics of TAC such as age, drugs interaction and length of 209 the event related to dose-adjusted concentrations were not evaluated , and should be analyse 210 by other studies.

In conclusion, patients after liver transplantation, both donor and recipient carrying CYP3A5 polymorphisms are susceptible to suffer changes in TAC pharmacokinetics. However, the role of each polymorphism is different according to days after transplantation proceeds and it must be taken into account to optimize the benefits of TAC therapy during the post-transplant induction and maintenance phase.

- Acknowledgement: University of Antioquia, Fundación Investigar, Buenos Aires Argentina,
 Paula Schaiquevich and pharmacokinetics laboratory of PHJPG.
- 218 Conflict of interests: None
- Author contributions: All authors contributed equally in the revision, drafting and writing of thearticle.

221 References.

- Undre N. Pharmacokinetics of Tacrolimus: Clinically Relevant Aspects. . Transplantation
 Proceedings, . 1999;31 ((Suppl 7A), 21S-24S).
- Wallemacq P, Armstrong VW, Brunet M, Haufroid V, Holt DW, Johnston A, et al.
 Opportunities to optimize tacrolimus therapy in solid organ transplantation: report of the European
 consensus conference. Therapeutic drug monitoring. 2009;31(2):139-52.
- Staatz CE TS. Clinical pharmacokinetics and pharmacodynamics of tacrolimus in solid
 organ transplantation. Clinical pharmacokinetics. 2004;43:623-53.
- 4. Wang L, McLeod HL, Weinshilboum RM. Genomics and drug response. The New
 England journal of medicine. 2011;364(12):1144-53.

Hesselink DA vSR, van der Heiden IP, et al. . Genetic polymorphisms of the CYP3A4,
 CYP3A5, and MDR-1 genes and pharmacokinetics of the calcineurin inhibitors cyclosporine and
 tacrolimus. . Clin Pharmacol Ther 2003;;74:245-54.

6. Kuehl P ZJ, Lin Y, et al. Sequence diversity in CYP3A promoters and characterization of the genetic basis of polymorphic CYP3A5 expression. Nat Genet 2001;27(4):383-91.

Z36 7. Lamba JK LY, Schuetz EG, et al. . Genetic contribution to variable human CYP3A mediated metabolism. Adv Drug Deliv Rev 2002;54 (10):1271-94.

Buendia JA, Bramuglia G, Staatz CE. Effects of combinational CYP3A5 6986A>G
 polymorphism in graft liver and native intestine on the pharmacokinetics of tacrolimus in liver
 transplant patients: a meta-analysis. Therapeutic drug monitoring. 2014;36(4):442-7.

Rojas LE, Herrero MJ, Boso V, Garcia-Eliz M, Poveda JL, Librero J, et al. Meta-analysis
 and systematic review of the effect of the donor and recipient CYP3A5 6986A>G genotype on
 tacrolimus dose requirements in liver transplantation. Pharmacogenet Genomics.
 2013;23(10):509-17.

Kelly DA, Bucuvalas JC, Alonso EM, Karpen SJ, Allen U, Green M, et al. Long-term
medical management of the pediatric patient after liver transplantation: 2013 practice guideline
by the American Association for the Study of Liver Diseases and the American Society of
Transplantation. Liver transplantation : official publication of the American Association for the
Study of Liver Diseases and the International Liver Transplantation Society. 2013;19(8):798-825.

Buendía JA, Otamendi E, Kravetz MC, Cairo F, Ruf A, de Davila M, et al. Combinational
 Effect of CYP3A5 and MDR-1 Polymorphisms on Tacrolimus Pharmacokinetics in Liver
 Transplant Patients. Exp Clin Transplant. 2015 Oct;13(5):441-8

Riva N, Dip M, Halac E, Cáceres Guido P, Woillard JB, Licciardone N, et al. Survival Time
 to Biopsy-Proven Acute Rejection and Tacrolimus Adverse Drug Reactions in Pediatric Liver
 Transplantation. Ther Drug Monit. 2018 Aug;40(4):401-410>

256 13. Buendía Rodríguez, Jefferson Antonio (2015-02-24). Desarrollo de modelos 257 farmacocinéticos de Tacrolimus en distintas poblaciones de pacientes trasplantados hepáticos: 258 caracterización de variables clínicas y genéticas asociadas a la variabilidad farmacocinética/farmacodinámica (tesis doctoral). Universidad de Buenos Aires. Facultad de 259 260 Farmacia y Bioquímica. [consultado: 8/2/2019] Available in : Repositorio Digital Institucional de 261 Universidad <http://repositoriouba.sisbi.uba.ar/qsdl/cqila de Buenos Aires: 262 bin/library.cgi?a=d&c=posgrauba&cl=CL1&d=HWA 828>.

14. Kuehl P, Zhang J, Lin Y, Lamba J, Assem M, Schuetz J, et al. Sequence diversity in
CYP3A promoters and characterization of the genetic basis of polymorphic CYP3A5 expression.
Nat Genet. 2001;27(4):383-91.

Lavandera J MM, Parera V, Rossetti MV, Batlle A, Buzaleh AM. Identificación de
polimorfismos del CYP3A5 y CYP2B6 en infección por VIH asociada a Porfiria Cutánea Tardía
en la población Argentina. Reunion Cienfitica anual de la Sociedad Argentina de Investigacion
Clinica (SAIC): Sociedad Argentina de Investigacion Clinica (SAIC); 2010.

Larriba J, Imperiali N, Groppa R, Giordani C, Algranatti S, Redal MA. Pharmacogenetics
 of immunosuppressant polymorphism of CYP3A5 in renal transplant recipients. Transplant Proc.
 2010;42(1):257-9.

17. Ferraris JR, Argibay PF, Costa L, Jimenez G, Coccia PA, Ghezzi LF, et al. Influence of
CYP3A5 polymorphism on tacrolimus maintenance doses and serum levels after renal
transplantation: age dependency and pharmacological interaction with steroids. Pediatric
transplantation. 2011;15(5):525-32.

18. Arrieta O, Cardona AF, Federico Bramuglia G, Gallo A, Campos-Parra AD, Serrano S, et
al. Genotyping non-small cell lung cancer (NSCLC) in Latin America. Journal of thoracic oncology
cofficial publication of the International Association for the Study of Lung Cancer.
2011;6(11):1955-9.

19. Roco A, Quiñones L, Agundez JA, Garcia-Marti E, Squicciarini V, Miranda C, et al.
 Frequencies of 23 functionally significant variant alleles related with metabolism of antineoplastic
 drugs in the Chilean population: comparison with Caucasian and Asian populations. Frontiers in
 Genetics. 2012;3(229):1-9.

285 20. Xue F, Han L, Chen Y, Xi Z, Li Q, Xu N, et al. CYP3A5 genotypes affect tacrolimus
286 pharmacokinetics and infectious complications in Chinese pediatric liver transplant patients.
287 Pediatric transplantation. 2014;18(2):166-76.

288 21. Chen SY, Li JL, Meng FH, Wang XD, Liu T, Li J, et al. Individualization of tacrolimus
289 dosage basing on cytochrome P450 3A5 polymorphism - a prospective, randomized, controlled
290 study. Clin Transplant. 2013;2013(24):12101.

291 22. Starkel P, Laurent S, Petit M, Van Den Berge V, Lambotte L, Horsmans Y. Early down-292 regulation of cytochrome P450 3A and 2E1 in the regenerating rat liver is not related to the loss 293 of liver mass or the process of cellular proliferation. Liver. 2000;20(5):405-10.

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295