

1 Article

2 An Experimental Approach To Risk Of Organ 3 Rejection: Demonstration Of False 4 Immunosuppressant Results Due To Radiopaque 5 Agents

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12 Abstract:

13 Background: Immunosuppressant blood levels should be measured at regular periods in order to
14 keep them within the therapeutic index. Although LC-MS/MS is preferred as a reliable method,
15 some molecules like radiopaque agents in blood matrix may lead to false results. The aim of this
16 study is to investigate the effect of seven different radiopaque agents on immunosuppressant drugs.

17 Methods: Seven different radiopaque agents were added into control materials containing
18 tacrolimus, everolimus, sirolimus and cyclosporine A drugs. Measurements were performed by LC-
19 MS/MS instrument. The amount of deviations from target values were calculated.

20 Results: Immunosuppressant blood levels significantly changed after the administration of
21 radiopaque agents. Seven different radiopaque products led to false negative results in tacrolimus
22 and cyclosporine A levels at a rate of 19.77% to 44.45%. The smallest deviations were seen in
23 everolimus levels with administration of RM6 (gadodiamide) and in sirolimus levels with RM1
24 (gadobutrol) at rates of 4.04% and 2.11%, respectively. The highest deviations were observed with
25 RM3 (iohexol) administration in everolimus and sirolimus levels at rates of 153.72% and 171.41%,
26 respectively.

27 Conclusions: False immunosuppressant results associated with radiopaque agents may result in
28 organ rejection. Preferring radiopaque agents that cause the least interference risk is important to
29 reduce the organ rejection risk. However, the least risky method is to obtain samples for drug levels
30 before contrast-enhanced imaging.

31

32 **Keywords:** Organ transplantation, immunosuppressant, radiopaque agents, interference

33

34 1. Introduction

35 Organ transplantation is the only treatment modality for patients with terminal stage organ
36 failure [1]. Immunosuppressants are used to prevent rejection of the organ transplantation [2,3].
37 Blood levels should be measured at regular periods in order to keep these oral drugs within the
38 therapeutic index [3]. LC-MS/MS reference method is used for these drugs since immunoassay
39 methods used commonly may be affected by several endogenous and exogenous molecules [4,5].

40 Although LC-MS/MS is a reference method, it may be influenced by the molecules in the matrix [6].
41 Among these molecules are radiopaque agents used during contrast-enhanced imaging.
42 Macromolecular radiopaque agents that do not permit transmission of X-rays may interfere with
43 immunosuppressant levels. The matrix effect produced by the impact of some molecules found in
44 the blood on ionization phase of the LC-MS/MS measurement method results in false measurement
45 of analytes [7,8]. This was initially demonstrated by Tang and Kebarle (1993) who showed that
46 electrospraying reactions of analytes were reduced by increasing concentrations of organic
47 bases. Although the mechanism underlying the matrix effect is unknown, it is likely to be caused by
48 undetectable serum components that bind to an analyte [9]. False measurement of
49 immunosuppressant concentration may lead to incorrect dose restriction or escalation. In particular,
50 incorrect measurement of drug levels used for immunosuppression in liver or kidney transplantation
51 poses a substantial risk for organ rejection [10,11]. This kind of interference is unpredictable. The
52 focus of this experimental study is to investigate how immunosuppressant drug levels are influenced
53 by radiopaque agents
54

55 2. Methods

56 2.1. Materials:

57 Six different levels of immunosuppressant calibrator (Jasem, Turkey, Lot: CL-3000420150616) and
58 single-level control solution (Lot: CL-3000620150616) were used in this study. Radiopaque agents used
59 for the interference study included iohexol (omnipaque, 755 mg/mL, 100 mL for intravenous injection,
60 GE Healthcare), gadopentetate dimeglumine salt (emaray, 469.01 mg/mL, 15 mL solution for IV
61 injection), gadodiamide (gadotu, 287 mg/mL, 15 mL solution for IV injection), ioversol (optiray, 741
62 mg/mL, 100 mL for intravenous injection), iohexol (kopaq, 755 mg/mL, 100 mL for intravenous
63 injection), gadobutrol (gadovist, 604.72 mg/mL, 15 mL solution for IV injection), gadodiamide
64 (gadodiem, 287 mg/mL, 15 mL IV solution for IV injection). All the solvents and reagents of HPLC were
65 produced by JASEM.
66

67 2.2. Measurement Devices:

68 LC-MS/MS analyzes of the tacrolimus, sirolimus, everolimus, cyclosporine A compounds were
69 performed by using a UHPLC (Nexera, Shimadzu X2, Japan) and a tandem MS instrument (Shimadzu
70 8045, Japan). The liquid chromatography was equipped with LC-40AD binary pumps. The
71 chromatographic separation was performed on a immunosuppressant analytical column (JASEM). The
72 column temperature was fixed at 40 °C. The elution gradient consisted of mobile phase A (water, 5 mM
73 ammonium formate and 0.1% formic acid) and mobile phase B (methanol, 5 mM ammonium formate
74 and 0.1% formic acid).. The solvent flow rate was maintained at 0.5 mL/min and injection volume was
75 settled as 4 µL. For MS detection was carried out by Shimadzu LCMS 8045 model triple quadrupole
76 mass spectrometer equipped with an ESI source operating in negative ionization modes. LC-MS/MS
77 data were collected and processed by Lab Solutions software (Shimadzu, Kyoto, Japan). The multiple
78 reaction monitoring (MRM) modes were used to quantify the analytes: the assay of investigated
79 compounds was performed following two or three transitions per compound, the first one for
80 quantitative aim and the second and/or the third one for confirmation. In this study, various seven
81 radiopaque material which are used widespread in clinic materials were quantified for effect of
82 interference.

83 2.3. Preparation of Samples and Statistic:

84 100 microliters (µL) of control solution was transferred to a centrifuge tube. 200 µL of internal
85 standard and 10 µL of distilled water were added and mixed for 5 seconds in vortex. The resulting
86 mixture was centrifuged at 10,000 rpm for 10 minutes. The supernatant was transferred into a vial and
87 measured in Shimadzu 8045 LC-MS/MS device. For the interference study, 10 µL of radiopaque agent
88 was added into the plasma and measurement was made after mixing with a vortex. Each measurement
89 was repeated 3 times and area values as well as concentration values were calculated by means of the
90 Shimadzu Software. This procedure was repeated individually for each of the 7 different radiopaque

91 agents. Area and concentration values for each of the three measurements were calculated with mean
 92 values for each mixture. Results derived by adding 10 μ L of distilled water into the control material to
 93 exclude interference due to volume expansion were considered as target values. Radiopaque agents
 94 were coded from RM1 to RM7 instead of their commercial names due to copyright issues of trading
 95 companies. No ethics committee approval was required since no blood or tissue samples of human or
 96 animal origin was used in this study. (V2: Concentration of immunosuppressant with radiopaque
 97 material, V1: Concentration of immunosuppressant with distilled water)

$$\text{bias}(\%) = \frac{V2 - V1}{V1} \times 100$$

3. Results

103 Bias values were calculated by interference studies for each of the 7 different radiopaque agents.
 104 A false positive deviation of 153.72% was observed in everolimus level after the administration of
 105 RM3, which contains iohexol. Target level for everolimus was measured as 31.91 ng/mL while it
 106 should have been 12.58 ng/mL. Radiopaque agents coded RM1, RM5 and RM6 led to false negative
 107 results in everolimus levels by -10.06%, -17.51%, -4.04%, while RM2, RM4 and RM7 resulted in false
 108 positive results by 32.49%, 25.89%, and 4.6%, respectively. A false positive result of 171.41% was
 109 measured in sirolimus level after the administration of RM3, which contains iohexol. A false
 110 positivity of 114.01% was measured due to the radiopaque agent RM4, which contains gadopentetate
 111 dimeglumine (table1). False positive results ranging from 2.11% to 87.44% were observed after the
 112 administration of RM1, RM2, RM5, RM6 and RM7. False negativity ranging between 19.77%-29.28%
 113 were observed for tacrolimus levels and between 31.57%-44.45% for Cyclosporine A levels after the
 114 administration of 7 different radiopaque agents (figure1).
 115

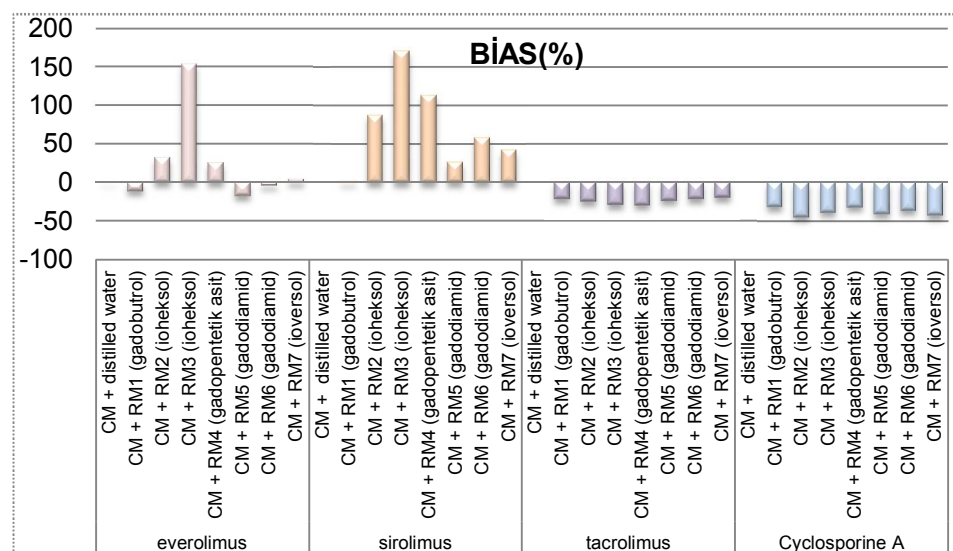


Figure 1. Graphic of percentage of deviation (bias) from target values

Table 1. Immunosuppressant concentrations and percentage of deviation (bias) from target values after radiopaque administration. (CM: Control Material, RM1-7: Radiopaque material)

Drug		Retention Time (sec)	Concentration (ng/mL)	Amount deviation (ng/mL)	of BIAS (%)	Active Substance
Everolimus	CM + Distilled water	3.95	12.58	-	-	-
	CM + RM1	3.94	11.20	-1.38	-10.96	Gadobutrol, 604.72 mg/mL
	CM + RM2	3.94	16.67	4.09	32.49	Iohexol, 755 mg/mL
	CM + RM3	3.95	31.91	19.34	153.72	Iohexol, 755 mg/mL
	CM + RM4	3.95	15.84	3.26	25.89	Gadopentetate dimeglumine salt, 469.01 mg/mL
	CM + RM5	3.95	10.38	-2.20	-17.51	Gadodiamide, 287 mg/mL
	CM + RM6	3.96	12.07	-0.51	-4.04	Gadodiamide, 287 mg/mL
	CM + RM7	3.95	13.16	0.58	4.60	Ioversol, 741 mg/mL
Sirolimus	CM + Distilled water	3.86	13.78	-	-	-
	CM + RM1	3.90	14.07	0.29	2.11	Gadobutrol, 604.72 mg/mL
	CM + RM2	3.89	25.84	12.05	87.44	Iohexol, 755 mg/mL
	CM + RM3	3.91	37.41	23.63	171.41	Iohexol, 755 mg/mL
	CM + RM4	3.89	29.50	15.71	114.01	Gadopentetate dimeglumine salt, 469.01 mg/mL
	CM + RM5	3.88	17.50	3.72	26.96	Gadodiamide, 287 mg/mL
	CM + RM6	3.90	21.92	8.14	59.02	Gadodiamide, 287 mg/mL
	CM + RM7	3.89	19.68	5.90	42.81	Ioversol, 741 mg/mL
Tacrolimus	CM + Distilled water	3.29	12.22	-	-	-
	CM + RM1	3.34	9.57	-2.65	-21.68	Gadobutrol, 604.72 mg/mL
	CM + RM2	3.30	9.19	-3.02	-24.73	Iohexol, 755 mg/mL
	CM + RM3	3.30	8.68	-3.54	-28.96	Iohexol, 755 mg/mL
	CM + RM4	3.33	8.64	-3.58	-29.28	Gadopentetate dimeglumine salt, 469.01 mg/mL
	CM + RM5	3.34	9.34	-2.88	-23.57	Gadodiamide, 287 mg/mL
	CM + RM6	3.32	9.58	-2.64	-21.60	Gadodiamide, 287 mg/mL
	CM + RM7	3.29	9.80	-2.42	-19.77	Ioversol, 741 mg/mL
Cyclosporine A	CM + Distilled water	4.14	204.65			
	CM + RM1	4.15	140.05	-64.61	-31.57	Gadobutrol, 604.72 mg/mL
	CM + RM2	4.14	113.69	-90.96	-44.45	Iohexol, 755 mg/mL
	CM + RM3	4.15	125.82	-78.83	-38.52	Iohexol, 755 mg/mL
	CM + RM4	4.15	139.62	-65.04	-31.78	Gadopentetate dimeglumine salt, 469.01 mg/mL
	CM + RM5	4.14	122.38	-82.28	-40.20	Gadodiamide, 287 mg/mL
	CM + RM6	4.15	130.86	-73.79	-36.06	Gadodiamide, 287 mg/mL
	CM + RM7	4.15	117.73	-86.93	-42.48	Ioversol, 741 mg/mL

122 4. Discussion

123 Radiopaque agents are paramagnetic intravenous diagnostic drugs used in imaging techniques.
 124 Commonly used active substances in routine practice include iohexol, gadobutrol, gadopentetate
 125 dimeglumine salt, gadodiamide and ioversol . These agents may interfere with test results of the
 126 patient when measurements are performed in blood samples collected after imaging techniques
 127 [12,13]. The degree of this interference can change according to the elimination time of these drugs.
 128 In particular, the impact on the results with tacrolimus, sirolimus, cyclosporine A and everolimus,

129 which are used for the immunosuppression of liver transplant patients are important for prognosis
130 of patient [14]. Various studies have been carried out on incorrect measurement of
131 immunosuppressants by immunoassay methods.

132 Elevated blood cyclosporine levels due to the presence of endogenous antibodies were reported by
133 Soldin et al. in their ACMIA immunoassay measurements performed using the Dimension RXL
134 analyzer. De Jonge et al. reported an incorrect cyclosporine level of 492 ng/mL in a 77 year-old patient.
135 However, any cyclosporine molecules could not detect in this patient's blood by measurement of LC-
136 MS. [15]. Sirolimus is also exposed to the interference by metabolites during immunoassay
137 measurements. Morris et al. found a bias of 49.2% with MEIA (microparticle enzyme immunoassay)
138 method compared to the measurements with LC MS/MS [16]. Schmidt et al. evaluated the sirolimus
139 analysis by CMIA (carbonylmetal immunoassay) method and found cross-reaction with sirolimus
140 metabolites. In another study that compared CMIA and LC-MS/MS, deviations of 14% to 39% were
141 observed between mean values. Higher results were found using the CMIA method compared to LC-
142 MS/MS [17].

143 In a study that used 90 samples for everolimus levels with QMS (Quantitative Microsphere System)
144 immunoassay method, everolimus values determined using the QMS everolimus test were found to
145 be approximately 11% higher than those obtained by the LC-MS/MS method [18]. In a study by Hoffer
146 et al. with 169 patient samples, mean everolimus concentration produced by the QMS everolimus test
147 was found to be 31.2% higher than that determined by LC-MS/MS. [19]. Sallustio et al. observed a
148 deviation of 30% between everolimus values measured by FPIA and LC-MS/MS methods [20].

149 Although drug metabolites are the main cause of interference in tacrolimus measurements,
150 incorrect tacrolimus concentrations were reported with low hematocrit values by the MEIA
151 (microparticle enzyme-linked immunoassay) method on AxSYM instrument [21]. Westley et al.
152 found a bias of 33.1% and 20.1% when LC-MS/MS method was compared with CEDIA and MEIA
153 methods, respectively, in renal transplant patients [22]. Bazin et al. evaluated tacrolimus test on the
154 CMIA (Chemiluminescent Microparticle Immunoassay) method and observed an average bias of
155 20% compared to the values found using LC-MS/MS [23]. ACMIA tacrolimus test is affected by
156 rheumatoid factors and endogenous heterophilic antibodies. Altinier et al. described an interference
157 by heterophilic antibodies on ACMIA tacrolimus method. Therapeutic levels of tacrolimus were
158 found in a patient resulting from the presence of heterophilic antibodies even after the treatment was
159 discontinued [24].

160 Despite the fact that superiority of the LC-MS/MS method compared to immunoassay has been
161 demonstrated in several studies, no study has been performed to investigate the impact of
162 radiopaque agents used for organ function imaging in transplant patients on immunosuppressant
163 levels. Analyte results may change by the matrix effect observed as the change in ionization activity
164 in LC-MS/MS measurement in the presence of combustible substances [25]. Although it appears
165 reliable for some clinicians to use this reference method in certain vital tests, it should be kept in mind
166 that false results may occur due to interferences during these measurements. In this interference
167 study performed with the addition of 7 different commercial radiopaque agents, a significant
168 influence was found on the concentrations of tacrolimus, everolimus, sirolimus and cyclosporine A.
169 All of the radiopaque agents included in the present study led to false negative results in tacrolimus
170 and cyclosporine A levels at a rate of 19.77% to 44.45%. False negativity may lead the clinicians to
171 increase drug dose. The smallest deviations were seen in everolimus levels with the administration
172 of RM6 (gadodiamide) and in sirolimus levels with RM1 (gadobutrol) at rates of -4.04% and 2.11%,
173 respectively. RM3 (iohexol) resulted in false positivity of 153.72% and 171.41% in everolimus and
174 sirolimus levels. Incorrectly high measurements of immunosuppressant levels may lead to using
175 insufficient drug doses and increased risk of organ rejection. RM2 and RM3 contain iohexol, RM5
176 and RM6 contain gadodiamide. Different rates of deviation from target levels despite the same active
177 ingredients in commercial products is thought to be caused by different excipients that constitute the
178 polar and apolar structure of these products. This is supported by the study of Bonfiglio et al.
179 reporting that the chemical nature of a component had a significant effect on the degree of the matrix
180 effect. A study including four compounds of different polarities under the same mass

181 spectrophotometric conditions showed that the most polar compound had the highest rate of ion
182 suppression and the least polar compound was less influenced by ion suppression [26]. King et al.
183 showed in a number of experiments that matrix effect is a consequence of the competition between
184 nonvolatile matrix components between analytical ions during the shift to ionization phase [27]. In
185 this study, the competition of molecules differed according to the diversity of radiopaque molecules.
186 The formation efficacy of analyte ions depends on the matrix intensity that enter the electrospray ion
187 source. Some studies have demonstrated that signal suppression is complicated in the manifestation
188 of the matrix effect and involves several many factors. Gas phase proton transfer reactions and the
189 competition at high viscosity are the major factors in the formation of the matrix effect [28].

190 5. Conclusions

191 Although LC-MS/MS is the reference method that provides high specificity, excellent sensitivity
192 and precision for measurements of immunosuppressant drugs, factors of matrix origin should be
193 carefully evaluated. It has been experimentally demonstrated by this study that an interference may
194 occur in blood immunosuppressant levels due to radiopaque agents. False test results due to
195 radiopaque agents may lead to incorrect drug dosing. Choice of radiopaque agents with minimal
196 measurement errors is important to reduce the risk of interference. However, the least risky method
197 is to obtain samples for drug level measurements before contrast-enhanced imaging. Clinicians
198 should interrogate administration of radiopaque agents and the time of sampling in the event that
199 suspicious results are obtained during the measurement of immunosuppressants.

200 REFERENCES

- 201 1. Grinyó, J.M. Why is organ transplantation clinically important? *Cold Spring Harbor perspectives in*
202 *medicine* **2013**, *3*, a014985.
- 203 2. Humar, A.; Ramcharan, T.; Denny, R.; Gillingham, K.J.; Payne, W.D.; Matas, A.J. Are wound
204 complications after a kidney transplant more common with modern immunosuppression?
205 *Transplantation* **2001**, *72*, 1920-1923.
- 206 3. Duncan, M.D.; Wilkes, D.S. Transplant-related immunosuppression: A review of immunosuppression
207 and pulmonary infections. *Proceedings of the American Thoracic Society* **2005**, *2*, 449-455.
- 208 4. Krasowski, M.D.; Drees, D.; Morris, C.S.; Maakestad, J.; Blau, J.L.; Ekins, S. Cross-reactivity of
209 steroid hormone immunoassays: Clinical significance and two-dimensional molecular similarity
210 prediction. *BMC clinical pathology* **2014**, *14*, 33.
- 211 5. Simpson, J.; Zhang, Q.; Ozaeta, P.; Aboleneen, H. A specific method for the measurement of
212 cyclosporin a in human whole blood by liquid chromatography-tandem mass spectrometry.
213 *Therapeutic drug monitoring* **1998**, *20*, 294-300.
- 214 6. Matuszewski, B.; Constanzer, M.; Chavez-Eng, C. Strategies for the assessment of matrix effect in
215 quantitative bioanalytical methods based on hplc- ms/ms. *Analytical chemistry* **2003**, *75*, 3019-3030.
- 216 7. Dams, R.; Huestis, M.A.; Lambert, W.E.; Murphy, C.M. Matrix effect in bio-analysis of illicit drugs
217 with lc-ms/ms: Influence of ionization type, sample preparation, and biofluid. *Journal of the American*
218 *Society for Mass Spectrometry* **2003**, *14*, 1290-1294.
- 219 8. Matuszewski, B.; Constanzer, M.; Chavez-Eng, C. Matrix effect in quantitative lc/ms/ms analyses of
220 biological fluids: A method for determination of finasteride in human plasma at picogram per milliliter
221 concentrations. *Analytical chemistry* **1998**, *70*, 882-889.
- 222 9. Tang, L.; Kebarle, P. Dependence of ion intensity in electrospray mass spectrometry on the
223 concentration of the analytes in the electrosprayed solution. *Analytical chemistry* **1993**, *65*, 3654-3668.

- 224 10. Buchwald, A.; Winkler, K.; Epting, T. Validation of an lc-ms/ms method to determine five
225 immunosuppressants with deuterated internal standards including mpa. *BMC clinical pharmacology*
226 **2012**, *12*, 2.
- 227 11. Srinivas, T.R.; Meier-Kriesche, H.-U. Minimizing immunosuppression, an alternative approach to
228 reducing side effects: Objectives and interim result. *Clinical Journal of the American Society of*
229 *Nephrology* **2008**, *3*, S101-S116.
- 230 12. Otnes, S.; Fogh-Andersen, N.; Rømsing, J.; Thomsen, H.S. Analytical interference by contrast agents
231 in biochemical assays. *Contrast media & molecular imaging* **2017**, 2017.
- 232 13. Xu, C.; Tang, Y.; Ruan, X.; Huang, Q.; Sun, L.; Li, J. The value of gd-bopta-enhanced mris and dwi
233 in the diagnosis of intrahepatic mass-forming cholangiocarcinoma. *Neoplasma* **2017**, *64*, 945-953.
- 234 14. Miloh, T.; Barton, A.; Wheeler, J.; Pham, Y.; Hewitt, W.; Keegan, T.; Sanchez, C.; Bulut, P.; Goss, J.
235 Immunosuppression in pediatric liver transplant recipients: Unique aspects. *Liver Transplantation*
236 **2017**, *23*, 244-256.
- 237 15. de Jonge, H.; Geerts, I.; Declercq, P.; de Loor, H.; Claes, K.; Desmet, K.; Kuypers, D.R. Apparent
238 elevation of cyclosporine whole blood concentrations in a renal allograft recipient. *Therapeutic drug*
239 *monitoring* **2010**, *32*, 529-531.
- 240 16. Morris, R.G.; Salm, P.; Taylor, P.J.; Wicks, F.A.; Theodossi, A. Comparison of the reintroduced meia
241 assay with hplc-ms/ms for the determination of whole-blood sirolimus from transplant recipients.
242 *Therapeutic drug monitoring* **2006**, *28*, 164-168.
- 243 17. Schmid, R.W.; Lotz, J.; Schweigert, R.; Lackner, K.; Aimo, G.; Friese, J.; Rosiere, T.; Dickson, D.;
244 Kenney, D.; Maine, G.T. Multi-site analytical evaluation of a chemiluminescent magnetic
245 microparticle immunoassay (cmia) for sirolimus on the abbott architect analyzer. *Clinical biochemistry*
246 **2009**, *42*, 1543-1548.
- 247 18. Dasgupta, A.; Davis, B.; Chow, L. Evaluation of qms everolimus assay using hitachi 917 analyzer:
248 Comparison with liquid chromatography/mass spectrometry. *Therapeutic drug monitoring* **2011**, *33*,
249 149-154.
- 250 19. Hoffer, E.; Kurnik, D.; Efrati, E.; Scherb, I.; Karasik, M.; Ring, G.; Bentur, Y. Comparison of
251 everolimus qms immunoassay on architect ci4100 and liquid chromatography/mass spectrometry:
252 Lack of agreement in organ-transplanted patients. *Therapeutic drug monitoring* **2015**, *37*, 214-219.
- 253 20. Sallustio, B.C.; Noll, B.D.; Morris, R.G. Comparison of blood sirolimus, tacrolimus and everolimus
254 concentrations measured by lc-ms/ms, hplc-uv and immunoassay methods. *Clinical biochemistry*
255 **2011**, *44*, 231-236.
- 256 21. Armendariz, Y.; Garcia, S.; Lopez, R.M.; Pou, L. Hematocrit influences immunoassay performance
257 for the measurement of tacrolimus in whole blood. *Therapeutic drug monitoring* **2005**, *27*, 766-769.
- 258 22. Westley, I.S.; Taylor, P.J.; Salm, P.; Morris, R.G. Cloned enzyme donor immunoassay tacrolimus
259 assay compared with high-performance liquid chromatography-tandem mass spectrometry and
260 microparticle enzyme immunoassay in liver and renal transplant recipients. *Therapeutic drug*
261 *monitoring* **2007**, *29*, 584-591.
- 262 23. Bazin, C.; Guinedor, A.; Barau, C.; Gozalo, C.; Grimbert, P.; Duvoux, C.; Furlan, V.; Massias, L.;
263 Hulin, A. Evaluation of the architect tacrolimus assay in kidney, liver, and heart transplant recipients.
264 *Journal of pharmaceutical and biomedical analysis* **2010**, *53*, 997-1002.

- 265 24. Altinier, S.; Varagnolo, M.; Zaninotto, M.; Boccagni, P.; Plebani, M. Heterophilic antibody
266 interference in a non-endogenous molecule assay: An apparent elevation in the tacrolimus
267 concentration. *Clinica chimica acta; international journal of clinical chemistry* **2009**, *402*, 193-195.
- 268 25. Taylor, P.J. Matrix effects: The achilles heel of quantitative high-performance liquid chromatography–
269 electrospray–tandem mass spectrometry. *Clinical biochemistry* **2005**, *38*, 328-334.
- 270 26. Bonfiglio, R.; King, R.C.; Olah, T.V.; Merkle, K. The effects of sample preparation methods on the
271 variability of the electrospray ionization response for model drug compounds. *Rapid Communications*
272 *in Mass Spectrometry* **1999**, *13*, 1175-1185.
- 273 27. King, R.; Bonfiglio, R.; Fernandez-Metzler, C.; Miller-Stein, C.; Olah, T. Mechanistic investigation
274 of ionization suppression in electrospray ionization. *Journal of the American Society for Mass*
275 *Spectrometry* **2000**, *11*, 942-950.
- 276 28. Tong, X.S.; Wang, J.; Zheng, S.; Pivnichny, J.V.; Griffin, P.R.; Shen, X.; Donnelly, M.; Vakerich, K.;
277 Nunes, C.; Fenyk-Melody, J. Effect of signal interference from dosing excipients on pharmacokinetic
278 screening of drug candidates by liquid chromatography/mass spectrometry. *Analytical chemistry* **2002**,
279 *74*, 6305-6313.

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