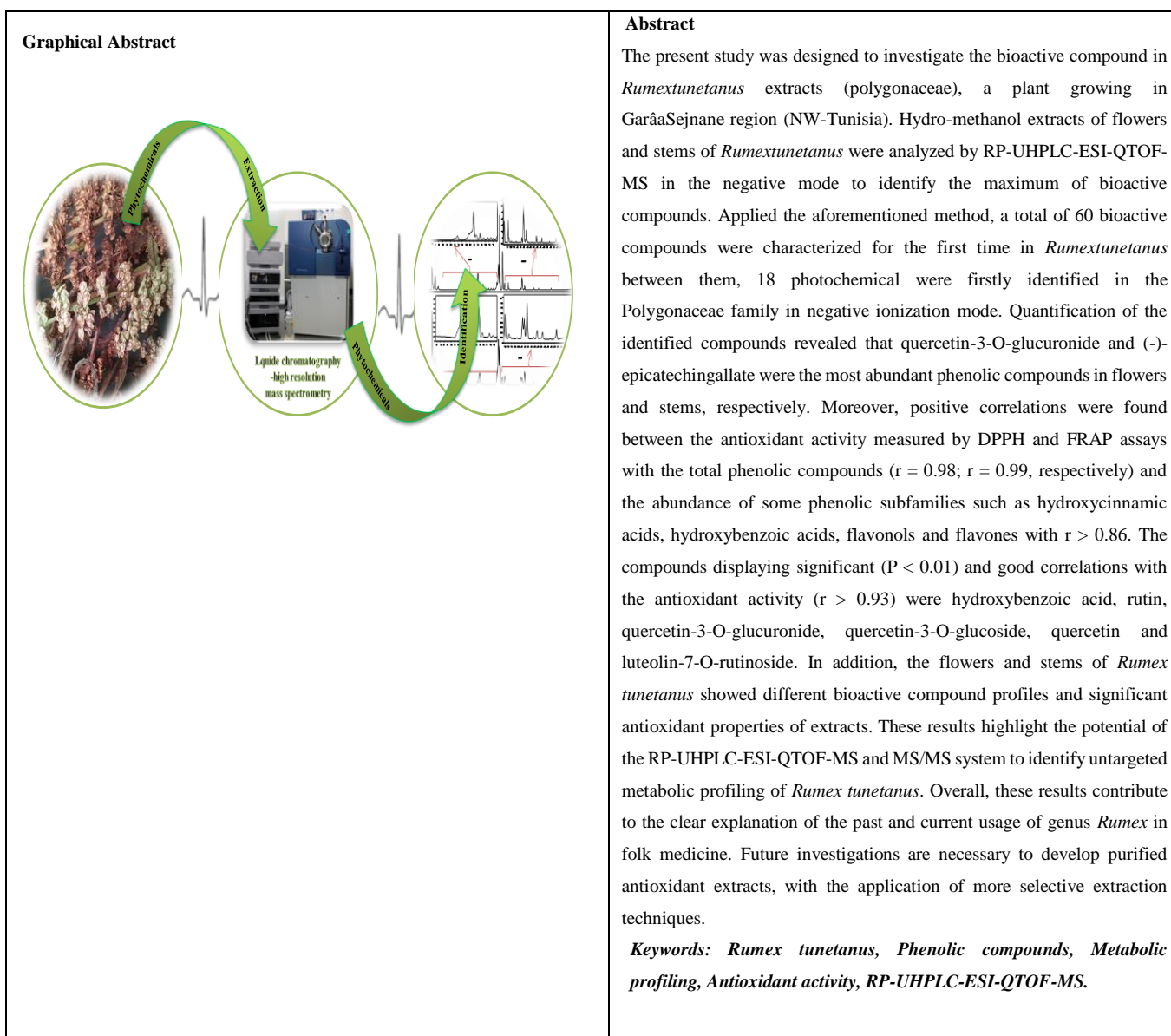


Determination of phenolic composition and antioxidant activity in flowers and stems of *Rumex tunetanus* using RP-UHPLC-ESI-QTOF-MS

JoudaAbidi^a, SondaAmmar^a, Mohamed Bouaziz^{a*}.

^aLaboratoire d'Electrochimie et Environnement, Ecole Nationale d'Ingénieurs de Sfax, Université de Sfax, BP1173, 3038 Sfax, Tunisia.



Corresponding author:

*Prof. Mohamed BOUAZIZ, Tel: +216 98 667 581 / Fax: +216 74 674 364.

E-mail: mohamed.bouaziz@fsg.rnu.tn

Introduction

In response to the increased popularity and greater demand for medicinal plants, a recent study by world health organization claimed 80% dependency of word population on ethnomedicines[1].

In fact, the importance of medicinal plants in solving the health care problems of the world is gaining increasing attention and it is growing phenomenally at the international level. However, the pharmacological evaluation of substances from plants is an established method for the identification of new compounds which can lead to the development of novel and safe medicinal agents in order to increase the dangerous side effects of synthetic molecules [2]. The genus *Rumex* represents one of the most important genera of Polygonaceae family, including approximately 200 species widely distributed in North American, European, African and Asian countries [3]. Since very old times, different species of *Rumex* genus have been used for relief of symptoms of diseases. Much interest in *Rumex* species emanates from their long use in folk medicines as well as their pharmacological properties pharmacological activities such as anti-inflammatory, antidiuretic, antitumor, analgesic, antifungal and anti-viral activities [1]. Large number of medicinal plants have been investigated for their antioxidant properties. Polyphenols are considered among the most important antioxidants in human diet, and their presence in plant can protect consumers against oxidative stress, cardiovascular and chronic diseases [4]. In fact, *Rumex* genus emerged as a good source of natural antioxidants. The major types of phenolic antioxidants found in *Rumex* genus include phenolic acids and their derivatives, namely gallic acid, dihydroxybenzoic acid, hydroxybenzoic acid and vanillic acid, and flavonoids, namely flavan-3-ols (catechin, epicatechin and epicatechin gallate), flavonols (rutin, quercetin-3-O-glucoside, quercetin-3-O-glucuronide, quercetin and isorhamnetin), as well as condensed tannins as B-type procyanidin dimer and A-type procyanidin trimer [5]. Tunisia flora is known for its diversity of medicinal plant among them *Rumex tunetanus*. This plant was growing in the wet marshes of the Sejenane plain. *Rumex tunetanus* was never seen after its discovery in 1888, and more than 120 years after the discovery of this *Rumex*, we find a large population of *Rumex tunetanus* in December 2009 [6]. Therefore, as potential bioactive markers, the total phenolic content (TPC) and antioxidant capacity of flowers and stems of *Rumex tunetanus* were firstly evaluated. Secondly, their phenolic profiles were extensively studied by ultra-high performance liquid chromatography (UHPLC) coupled with two detection systems, DAD and quadrupole time-of-flight (QTOF)-MS using electrospray ionization in negative ionization mode.

Materials and Methods

Solvents and standards

The solvents used for extraction were ultrapure water and methanol. All solvents used for HPLC-MS analysis were delivered by J.T Baker (Phillipsburg -USA). The reagents used to measure the TPC (total phenol content) and the antioxidant capacity were: Folin-Ciocalteu; sodium carbonate (Na_2CO_3); 2,2-diphenyl 1-picrylhydrazyl (DPPH); 2,4,6-tri(2-pyridyl)-1,3,5-triazine (TPTZ) and ferric chloride ($\text{FeCl}_3 \cdot 6\text{H}_2\text{O}$); gallic acid, acetic acid ($\text{C}_2\text{H}_4\text{O}_2$); Ferric sulfate (FeSO_4); hydrochloric acid (HCl); trihydrate sodium acetate ($\text{C}_2\text{H}_3\text{NaO}_2 \cdot 3\text{H}_2\text{O}$); sodium acetate; ascorbic acid. The phenolic standards used in our work were bought from Sigma-Aldrich (Saint-Louis -Missouri). The degree of purity of the standards was around 95% (w/w).

Flowers, stems and leaves from the *Rumex tunetanus* were collected from the Garâa Sejenane region (NW–Tunisia) in May 2014. The samples were harvested and transferred to the laboratory where they were dried in the dark at room temperature 30 °C, and then they were finely ground prior to extraction.

Flowers and stems were extracted using methanol/water 80:20 (v/v) as described elsewhere [7]. The residue was filtered with a syringe filter (regenerated cellulose, 0.45 µm pore size) and stored at –20 °C until future analysis. The extractions were repeated twice for each studied *rumex tunetanus* part.

Total phenol content and antioxidant capacity assays

The TPC of the extracts was determined in triplicate by the colorimetric assay using the Folin–Ciocalteu reagent as reported by Ammar et al. [8]. The DPPH assay was based on the method described by Gargouri et al. [9]. The FRAP assay was conducted following the method described by Ammar et al. [10].

Mass spectrometry-based analyses

The analyses were made with an Agilent 1200 series rapid resolution (Palo Alto, CA, USA). The system was coupled with a DAD and a 6540 Agilent ultra-high-definition (UHD) accurate-mass Q-TOF LC/MS, which was equipped with Agilent Dual Jet Stream electrospray ionization (Dual AJS ESI) interface. The detail of negative ionization modes was described by Abidi et al [7].

Databases

In addition to consulting literature, the following databases were used to retrieve chemical information: PubChem (<http://pubchem.ncbi.nlm.nih.gov>), ChemSpider (<http://www.chemspider.com>), SciFinder Scholar (<https://scifinder.cas.org>), Reaxys (<http://www.reaxys.com>), Phenol-Explorer (www.phenol-explorer.eu) and KNApSAcK Core System (http://kanaya.naist.jp/knapsack_jsp/top.html). MassBank and Metlin Metabolite Database were used to check fragmentation patterns in some cases.

Results and Discussion

Total phenolic content (TPC) and antioxidant activity of the *Rumex tunetanus* flowers and stems

The determination of TPC of the extracts was performed by the Folin-Ciocalteu assay and calculated as gallic acid equivalents and is listed in Table 1. *Rumex tunetanus* flowers extract was found to contain higher total phenolic contents (146.20 mg GAE/g extract) as compared to *Rumex tunetanus* stems extract (118.88 mg GAE/g extract).

The extracts were evaluated for their antioxidant /radical scavenging activity by DPPH and FRAP. The results of percentage scavenging are depicted in Table 1. In general, both of these two analyses showed that the hydro-methanolic extract of *Rumex tunetanus* exhibited good antioxidant activities. According to the aforementioned results for TPC, the flowers showed higher antioxidant activity values than stems by the two assayed methods. In fact, our results have indicated that the DPPH value of *Rumex tunetanus* flowers and stems extracts are in accordance with those obtained by Yousef et al. [11] for *Rumex tuberosus* flowers (0,65mg/ml of extract) and stems (0,69 mg/ml of extract).

Characterization strategy and fragmentation pattern study

The metabolic profiling of the aqueous methanolic extracts of flowers and stems was performed by using RP-HPLC-DAD-QTOF-MS using ESI in the negative mode. This is the commonest ionization source and mode used to identify the phenolic and non-phenolic compounds in *Rumex tunetanus* flowers and stems. Using this methodology, our characterization steps could be basically summarized by a targeted searching of previously, an untargeted analysis and a predictive study of unreported phenolic structures based on all the spectrometric data obtained by the detection techniques applied (Figure 1).

Thanks to the methodology proposed in the present study, a total of 60 metabolites (60 in flowers and 51 in stems (Table 2) have been tentatively identified, including sugars (1), organic acid (3), hydroxybenzoic acids (8), hydroxycinnamic acids (1), flavonoids (28) as well as tannins (19). These results exemplify that RP-HPLC-DAD-QTOF-MS is useful to detect and characterize novel chemical structures.

Comparison between flowers and stems

Table 1 shows the qualitative differences between flowers and stems: presence (+) or absence (-) of the characterized compounds. In brief, more than 90% of the compounds were in common in both extracts. Moreover, as summary Figure 2 depicts these differences in terms of number of compounds found in each phenolic class. Interestingly, flavonoids and their glycoside derivatives were characterized in the negative ionization mode as a more widely spread phenolic group of *Rumex tunetanus* samples with 28 compounds. To the best of our knowledge, this is the first time that dihydroxybenzoic acid hexoside (8), syringic acid hexoside (12) and tri-*O*-methoxyellagic acid (51) have been identified in Polygonaceae family.

Conclusions

Globally, these results exemplify the usefulness of RP-HPLC-DAD-QTOF-MS to perform characterization studies, such as that described here. In this way, 60 phenolic compounds were characterized in flowers and 51 in stems of *rumex tunetanus* which were classified into twelve compound families that possess potent antioxidant activities. Flavonols, flavanols, flavones, hydroxybenzoic acids and condensed tannins were the most representative groups in both samples by using the relative amounts of the phenolic compounds. In addition, the antioxidant properties of extracts were evaluated in terms of single electron transfer as DPPH and FRAP assay and *Rumex tunetanus* flowers exhibited significantly higher antioxidant activities than the stems. Besides, the isolation of pure compounds with pharmacological activities holds significance in contemporary and future research. Recently, the plant extract was also being used by the researchers to produce nanoparticles [2]. Therefore, there is huge room for research in these directions.

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Extract	TPC	DPPH	FRAP
Flowers	146.2 ± 1.86	0.72 ± 0.230	528.24 ± 32.36
Stems	118 ± 1.05	0.68 ± 0.230	307.04 ± 54.36

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Table 1 Total phenolic content (TPC) (mg of GAE/g of the extract) determination and antioxidant capacities of the studied *Rumex tunetanus* flowers and stems parts as measured by DPPH (mg/ml of extract) and FRAP (mmol eq. FeSO₄/g of the extract) antioxidant assays.

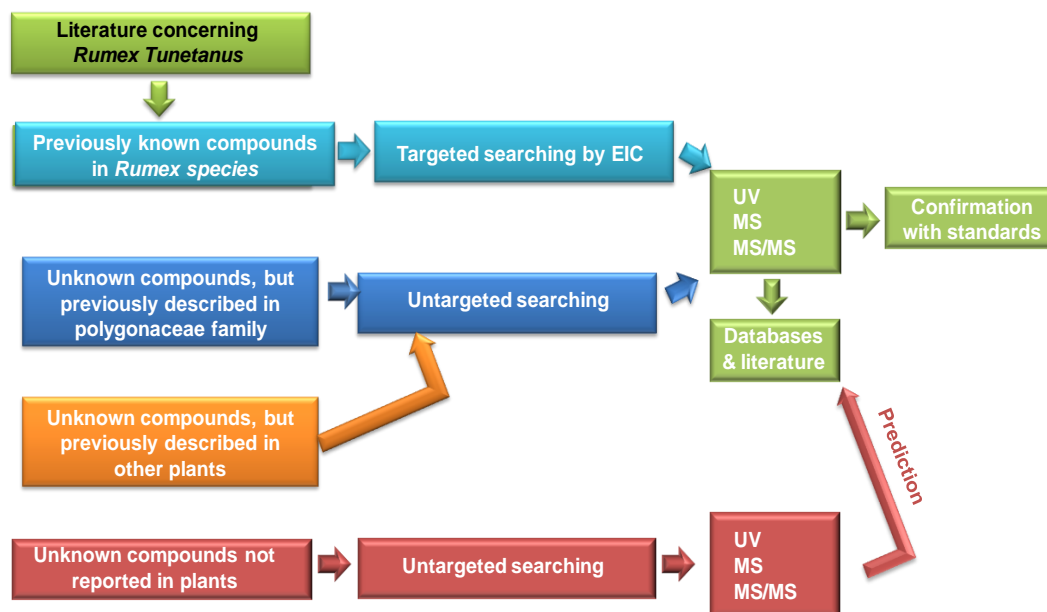


Figure 1: The strategy for deducing of the most possible structures of *Rumex tunetanus* bioactive compounds

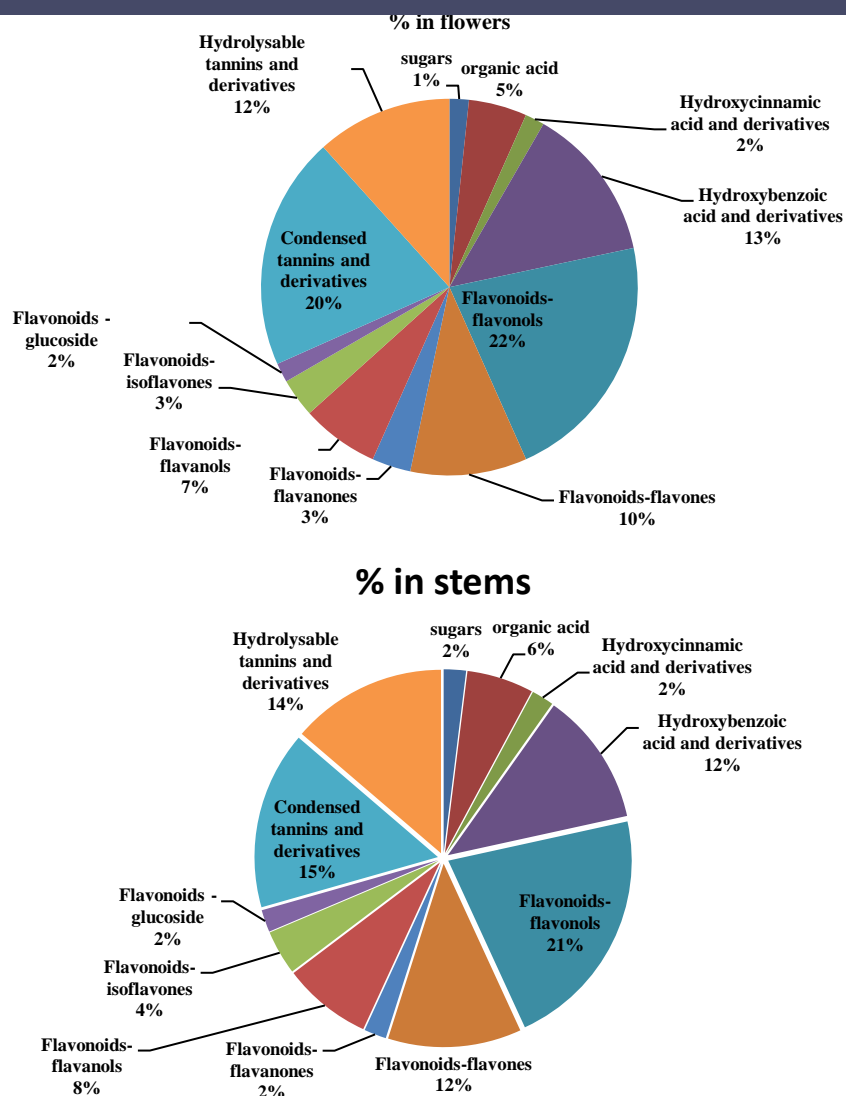


Figure 2. Various compounds found in each phenolic class of flowers and stems of *Rumex tunetanus*

Table 2. Compounds characterized by HPLC-DAD-QTOF-MS in *Rumex tunetanus* flowers and stems

N°	RT ^a (min)	Exp. ^a m/z for [M-H] ⁻	Proposed compound	Presence in		N°	RT ^a (min)	Exp. ^a m/z for [M-H] ⁻	Proposed compound	Presence in	
				Flowers	stems					Flowers	Stems
Sugars						58	36.81	329.0673	Tricin II	+	+
1	1.52	341.1116	Sucrose	+	+	60	39.70	329.0643	Tricin III	+	+
Organic Acid						Flavonoids-flavanones					
2	1.59	191.0542	Quinic acid I*	+	+	50	30.69	287.0562	Eriodictyol*	+	-
4	1.82	191.0538	Quinic acid II	+	+	55	34.39	271.0605	Naringenin*	+	+
5	1.97	191.0183	Citric acid*	+	+	Flavonoids-flavanoles					
Phenolic acid (Hydroxycinnamic acid and derivatives)						16	17.03	289.0727	(+)-Catechin* ^c	+	+
19	17.94	355.1028	Ferulic acid hexoside	+	+	23	19.53	289.0727	(-)-Epicatechin* ^c	+	+
Phenolic acid (Hydroxybenzoic acids and derivatives)						33	23.96	441.0824	(-)-Epicatechingallate ^c	+	+
7	3.75	169.0137	Gallic acid*	+	+	47	27.37	287.0556	Fustin ^b	+	+
8	4.43	315.0725	Dihydroxybenzoic acid hexoside ^b	+	-	Flavonoids-isoflavones					
9	8.88	153.0186	Dihydroxybenzoic acid hexoside ^b	+	+	57	36.57	299.0568	7-Methoxy 2'-hydroxy genistein (cajanin) ^b	+	+
10	12.14	137.024	Hydroxybenzoic acid I*	+	+	59	37.38	359.078	5,7,4-trihydroxy-6,3,5-trimethoxyisoflavone (Irigenin) ^b	+	+
11	12.96	137.0236	Hydroxybenzoic acid II	+	+	Flavonoids-glucoside					
12	13.94	359.0976	Syringic acid hexoside ^b	+	+	46	27.25	591.1357	Flavonoid glucoside-HMG conjugate ^b	+	+
25	20.43	167.0347	Vanillic acid*	+	-	Condensed tannins and derivatives					
51	31.38	343.0429	Tri-O-methoxyellagic acid ^b	+	+	13	15.82	577.1334	B-type procyanidin dimer (I) ^c	+	+
Flavonoids-flavonols						14	16.34	577.1334	B-type procyanidin dimer (II) ^c	+	-
31	23.39	615.0990	Quercetin-O-galloyl-hexoside ^b	+	+	22	18.71	577.1349	B-type procyanidin dimer (III) ^c	+	+
32	23.60	609.1465	Quercetin-3-O-rutinoside (rutin)* ^c	+	+	24	19.81	577.136	B-type procyanidin dimer (IV) ^c	+	-
34	24.11	463.0881	Quercetin-3-O-glucoside I* ^c	+	+	26	20.48	729.1472	B-type procyanidin dimer gallate I	+	-
35	24.33	477.0656	Quercetin-3-O-glucuronide*	+	+	27	20.72	863.1813	A-type procyanidin trimer	+	+

36	24.44	463.0884	Quercetin-3-O-glucoside Iic	+	-	28	21.09	729.1465	B-type procyanidin dimer gallate II	+	-
39	25.31	607.1306	Quercetin-3-[6''-(3-hydroxy-3-methylglutaryl)]β-hexosideb	+	+	29	21.47	577.1363	B-type procyanidin dimer (V)c	+	+
40	25.56	599.1050	Quercetin dihydroxybenzoylhexosideb	+	+	30	21.63	729.1455	B-type procyanidin dimer gallate III	+	+
43	26.44	505.099	Quercetin-3-O-hexosyl-6''-acetate	+	-	37	24.8	729.1464	B-type procyanidin dimer gallate IV	+	-
45	27.71	625.1183	Quercetin-O-dihexoside	+	+	44	26.61	729.1473	B-type procyanidin dimer gallate V	+	+
49	29.67	639.1339	Quercetin-O-feruloylhexosideb	+	+	48	28.13	881.1566	B-type procyanidin dimer digallateb	+	+
52	31.88		Quercetin*c	+	+					+	+
						Hydrolysable tannins and derivatives					
53	32.46	345.0608	Quercetin-3,6-dimethyl etherb	+	+	3	1.70	331.0648	Galloyl glucose I	+	+
54	32.71	315.0514	Isorhamnetin*	+	+	6	2.72	331.0661	Galloyl glucose II	+	+
Flavonoids-flavones						15	16.97	483.0816	Di-O-galloyl-glucose I	+	+
38	25.12	593.1532	Luteolin-7-O-rutinoside (Scolymoside)*b	+	+	17	17.63	453.1036	Hydroxy-methoxyphenyl-O-(O-galloyl)-hexose I ^b	+	+
41	26.00	447.0943	Luteolin-7-O-glucoside (Cynaroside)*c	+	+	18	17.80	483.1122	Hydroxy-dimethoxyphenolgalloyl-glucose ^b	+	+
42	26.18	329.0671	Tricin I	+	+	20	17.94	453.1031	Hydroxy-methoxyphenyl-O-(O-galloyl)-hexose II ^b	+	+
56	35.75	285.0405	Luteolin*c	+	+	21	18.35	483.0763	Di-O-galloyl-glucose II	+	+

^bCompounds described here for first time in family Polygonaceae. Several saccharide combinations and conjugation positions are reported in different plant families (see KnapSack, Reaxys or SciFinder databases)

^ccompounds were previously reported in several Rumex species.

^aRT, retention time;; Exp, experimental

*Identification confirmed by comparison with standards