

1 Article

2 Green Synthesis of *Citrus reticulata* Mediated Silver 3 Nanoparticles

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10 **Abstract:** Biosynthesis of nanoparticles for delivery of therapeutic agents has introduced new
11 opportunities in upgrading medical treatment. Plant extracts contains different capping and
12 reducing agents naturally thus provided simpler and less expensive way to synthesize AgNPs. In
13 present work, *Citrus reticulata* mediated stabilised AgNPs was synthesized. Optimum concentration
14 of reactants was achieved by varying the amount of extracts (1-11 ml) and AgNO₃ concentration
15 (0.5-3 mM). Surface Plasmon peak of *Citrus reticulata* mediated AgNPs was determined by
16 UV-visible spectrophotometer and functional groups of capping agents were examined by FTIR
17 analysis. Surface Plasmon peaks of *Citrus reticulata* fresh peel, seed, and juice extracts were observed
18 at 420 nm. But in dry peel extract, absorption peak of AgNPs appeared at 410 nm. Colour of different
19 extracts was changed after the reduction of AgNO₃ to AgNPs by reducing agents present in the
20 extracts. FTIR analysis showed band peaks at 3316 cm⁻¹ correspond to amide (N-H and O-H)
21 stretching vibrations while alkanes peaks was observed at 1638 cm⁻¹ which showed C=C stretching
22 aromatic ring (flavonoids). Furthermore, *Citrus reticulata* fresh peel mediated AgNPs showed
23 impressive stability up-to 112 days. In conclusion, *Citrus reticulata* fresh peel extract provided an
24 excellent source of reducing agents for synthesizing stabilized AgNPs.

25 **Keywords:** Biosynthesis; nanoparticles; plant extracts; *Citrus reticulata*

26 1. Introduction

27 Plants are greatest natural chemists on the earth that start with the transfer of sunlight into chemical
28 energy, a basis of primary products in the biosphere and extend to the astonishingly varied and
29 intricate secondary compounds, which synthesized in their body [1]. Protein based compounds;
30 metallothionins and phytochelators, phytochemicals; polyphenols, tannins and flavonoids are found
31 in naturally occurring compounds, play a key role in different biochemical reaction and for the
32 preparation of nanoparticles [2]. *Citrus reticulata* (Mandarin orange) is one of the famous and most
33 liking fruit of the world which belongs to Rutaceae family. It is reported that *Citrus reticulata* have
34 anti-bacterial [3], anti-fungal [4], anti-diabetic [5], cardio-protective [6], anti-cancer [7], anti-arthritic
35 [8], anti-inflammatory [9], anti-oxidant [10], anti-tubercular, and anti-anxiety medicinal properties
36 [11]. It has diverse and beneficial, capping and reducing phytochemicals like vitamin C, carotenoids
37 and flavonoids and many others [6] which might be involved in the green synthesis of silver

38 nanoparticles [12]. It has been reported that different parts of plants and their extracts have their
39 own antioxidant or reducing properties due to secondary metabolites including flavonoids,
40 polyphenol and tannins which play important function in the biosynthesis of Silver nanoparticles
41 (AgNPs) by the reduction of metals [13].

42 Many physical, electrochemical and chemical methods followed top down and bottom up pathways
43 are being in used for the synthesis of different nanoparticles including AgNPs and nano materials.
44 But the physical and chemical processes are not clean, safe and eco-friendly due to the involvement
45 the of highly technical apparatus, cost [14] and participation of hazards, harmful and explosive
46 chemicals [15,16]. It is therefore, green method was emerged for AgNPs synthesis [13], as it involves
47 low temperature, pressure, and energy as well as it is ecofriendly, radially scalable, safe to handle
48 and cost effective technology with reduced toxicity [17]. Materials having size ranges from 1-100 nm
49 are called nanomaterials which have the greatest importance in the field of nanotechnology.
50 Nanoparticles like gold, silver and copper are prepared by the extract of different parts of plants as
51 ecofriendly product and are being excessively [12] used in the fields of physics, chemistry, biology
52 [18] photography, catalysis, and biological labeling has helped in the development of excessive
53 interest in the field of nanotechnology [19]. Silver nanoparticles are of greatest interest among noble
54 metal nanoparticles due to antibacterial [20], antifungal [21], antitumor [22], antiviral [23] ,
55 anti-diabetes [24] and antioxidant activities [18]. AgNPs are used as catalysts, as optical sensors, in
56 textile engineering, in electronics, in optics, most importantly in the medical field as a therapeutic
57 agent [12] and also in food packaging, preservation, soaps, detergents, shampoos and cosmetics
58 [25]. AgNPs have the greatest antimicrobial activities. At very low concentration these are nontoxic
59 to human and have no serious side effects [26].

60 The aim of the present work was to synthesize the *Citrus reticulata* mediated AgNPs and to screen
61 which part of *Citrus reticulata* fruit was most active for the synthesis of stabilized AgNPs.

62 2. Materials and Methods

63 2.1 Extract preparation

64 Fresh *Citrus reticulatas* were purchased from local market of Lahore and were washed with distilled
65 water. 5 grams of *Citrus reticulata* peels were weighed and cut into small pieces. Resulting pieces of
66 peel were soaked in a beaker containing 100 ml distilled water and mixed for 2 hrs on magnetic
67 stirrer. After mixing, it was boiled on burner for 10 mins, then filtered and stored at 4 °C [12,38].
68 Seeds and dry peel extracts were also prepared by using same said method while fresh juice of *Citrus*
69 *reticulata* was used after centrifugation and filtration.

70 2.2 Preparation of the silver nitrate solution

71 Three dilutions (0.5, 1 and 2 mM) of AgNO₃ were prepared from 3 mM stock solution of AgNO₃
72 which was prepared by dissolving 0.051 g of AgNO₃ in 100 ml distilled water. All these solutions
73 were further used for the synthesis of AgNPs.

74 2.3 Synthesis of silver nanoparticles

75 AgNPs were synthesized by mixing 18 ml AgNO₃ (0.5 mM) in 5 ml fresh peel extract in volumetric
76 flask and then boiled for 30 mins in water bath. AgNPs were also synthesized by using juice, seeds
77 and dry peel extract separately by same above said procedure. The effects of the reaction conditions
78 such as volume of *Citrus reticulata* peel extract and concentration of AgNO₃ solution were evaluated.
79 Different volume of peel extract such as 1 ml, 2 ml, 5 ml and 11 ml were mixed with 18 ml AgNO₃

80 (0.5 mM) in a volumetric flasks separately to synthesize the AgNPs. Similarly, AgNPs biosynthesis
81 was examined by mixing different concentrations of AgNO₃ including 0.5 mM, 1 mM, 2 mM and 3
82 mM in 5 ml of peel extract used respectively while other conditions remains constant. The stability of
83 AgNPs was also checked at 1, 4, 12, 28, 56, 86 and 112 days after the reaction. UV-visible
84 spectrophotometer was used to analyze the formation of AgNPs [35,39].

85 **2.4 Characterization of silver nanoparticles**

86 **2.4.1 Visual analysis**

87 This colour change was noted by visual observation after the mixing of AgNO₃ and respective
88 extract [25].

89 **2.4.2 UV-Visible analysis**

90 Spectral analysis of AgNPs prepared by different reaction conditions were observed at the range
91 between 300 to 600 nm by UV-Visible spectrophotometer (T90+ UV/Vis Spectrometer.PG Instrument
92 LTd) [40].

93 **2.4.3 Fourier transform infrared spectrophotometer (FTIR) analysis**

94 For FTIR analysis, the synthesized AgNPs were centrifuged at 12,000 rpm for 30 mins at 0 °C and
95 then Supernatant was discarded and pallet of AgNPs washed with deionized water. Washing step
96 was repeated for three times [41]. The resulting AgNPs pallet was dissolved in methanol and then
97 evaporated [42]. Powder of AgNPs and fresh peel extract were subjected to FTIR (Agilant Microlab,
98 Carry 630 FTIR) for the identification of functional groups in the compounds present in the peel
99 extract and their involvement in the synthesis of AgNPs was determined [43].

100 **3. Results**

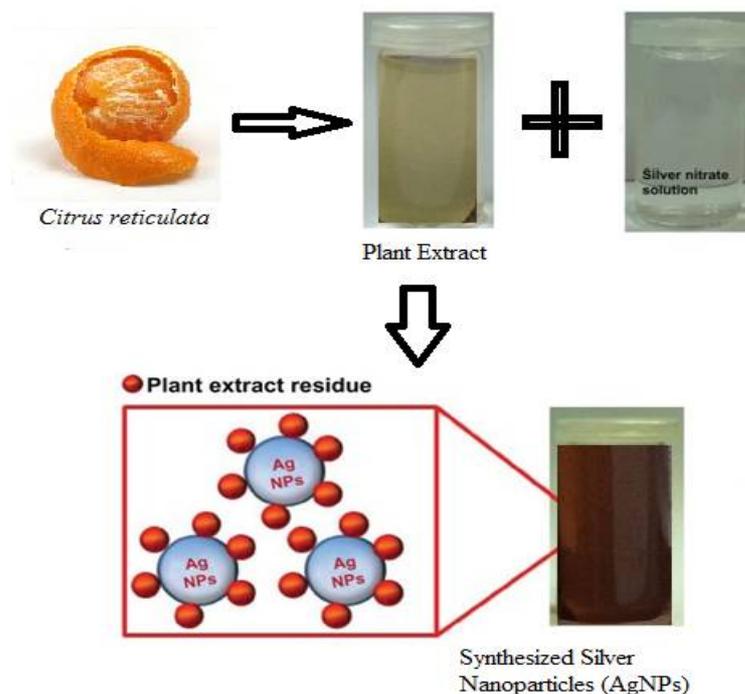
101 **3.1 Visual analysis**

102 Visual examination was done when the *Citrus reticulata* peel extract was mixed with aqueous
103 solution of 0.5 mM silver nitrate and put on water bath. Colour was changed from pale yellow to
104 brown after 30 mints, which was due to reduction of silver ions and indicated that the AgNPs were
105 formed (Fig 1).

106 **3.2 UV-Visible analysis**

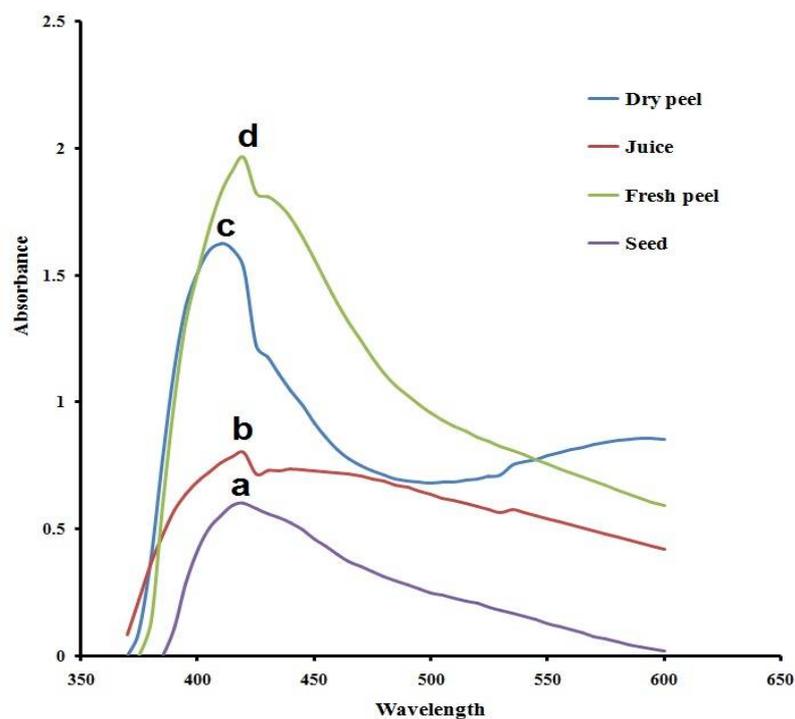
107 Extracts of different parts of *Citrus reticulata* fruit: seed, juice, dry peel and fresh peel were prepared
108 for bio-reduction of AgNO₃. The AgNPs formation was monitored with the help of UV-Visible
109 spectroscopy. UV-visible spectra were recorded immediately after the reaction.

110 A surface plasmon peak was observed at 420 nm for fresh peel, seed, and juice mediated AgNPs,
111 while dry peel mediated AgNPs showed a surface Plasmon peak with a maximum absorption at 410
112 nm. Fresh peel extract was used for further studies because it gave maximum concentration of
113 AgNPs (Fig.2).



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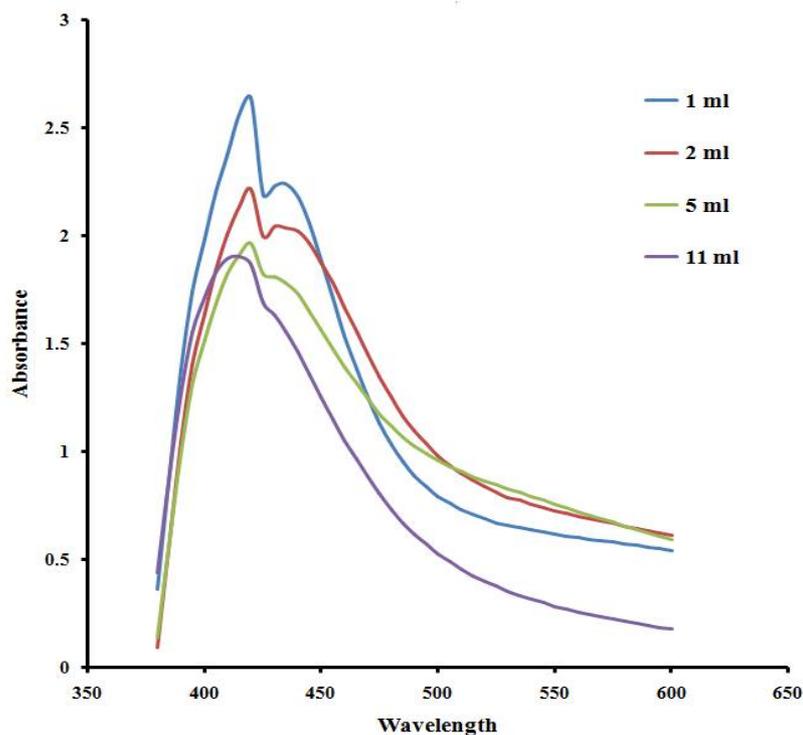
Figure 1: Schematic diagram for the biosynthesis of *Citrus reticulata* mediated silver nanoparticles



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Figure 2: UV-visible absorption spectra of AgNPs synthesized from the extracts of different parts of *Citrus reticulata*. Silver nitrate (AgNO_3) was reduced to silver nanoparticles (AgNPs) with extract isolated from different parts of *Citrus reticulata* fruit and UV-visible absorption spectra were recorded immediately after the reaction. Peak (a) shows seed extract mediated green synthesis of AgNPs absorption spectra, (b) represents juice mediated green synthesis AgNPs absorption spectra while (c) exhibits dry peel extract mediated green synthesis of AgNPs absorption spectra and (d) shows fresh peel mediated green synthesis of AgNPs absorption spectra.

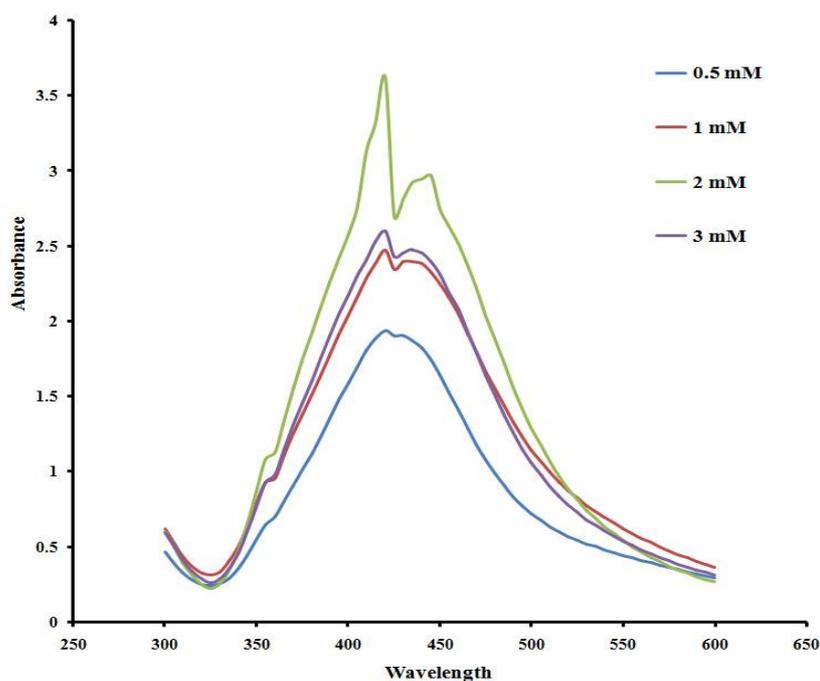
124 The reaction of AgNPs formation was monitored for the varying concentrations of fresh peel extract
125 (1, 2, 5 and 11 ml), keeping AgNO₃ concentration constant. It was observed that colorless solution of
126 AgNO₃ was converted into light brown for 1 ml, brown for 2 ml, radish brown for 5 ml and very
127 light brown for 11 ml . It was also found that concentration of synthesized AgNPs was decreased
128 with increasing volume of fresh peel extract (Fig.3). Furthermore, AgNPs synthesis was examined
129 with varying concentrations of AgNO₃ (0.5, 1, 2 and 3 mM) at constant volume of fresh peel extract
130 (5ml). An increasing trend for AgNPs synthesis was observed with increased concentration of
131 AgNO₃ firstly then it was decreased for 3 mM AgNO₃ (Fig.4).



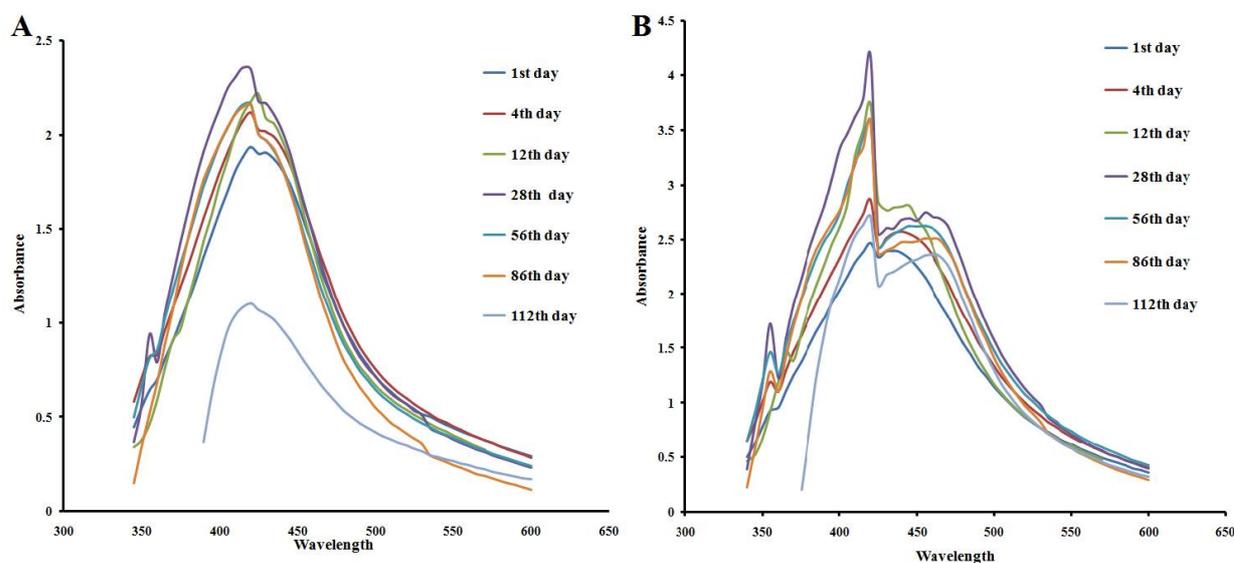
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133 **Figure 3: UV-visible absorption spectra for Fresh peel extract dependent study for AgNPs synthesis.** Aqueous extract of
134 *Citrus reticulata* fresh peel was made by continuous stirring along with boiling and then used as reducing agent for the
135 bio-reduction of AgNO₃ to silver. Concentration of AgNO₃ was kept constant and varying volume of “Fresh peel” extract was
136 used for the reduction of silver and UV-visible spectra were recorded immediately after the reaction.

137 After successful green synthesis of AgNPs, stability of synthesized AgNPs from fresh peel extract as
138 reducing and capping agent was monitored by recording UV-Visible spectra at different time
139 including 1, 4, 12, 28, 56, 86 and 112 days. It was found that, for 0.5 mM silver nitrate solution,
140 concentration of stabilized AgNPs increased until 28 days after the reaction gradually, and then it
141 decreasing as absorption intensity decreased with time. Same trend was observed when we took 1
142 mM AgNO₃ solution instead of 0.5 mM (Fig.5 A-B).

143



144
 145 **Figure 4: Silver nitrate (AgNO_3) concentration dependent UV-visible absorption spectra for fresh *Citrus reticulata* peel**
 146 **mediated AgNPs.** Different dilutions of AgNO_3 solution (0.5 mM, 1 mM, 2 mM and 3 mM) were prepared and mixed with
 147 fresh peel extract (5 ml) to synthesized AgNPs and then immediately their UV-visible spectra were recorded.

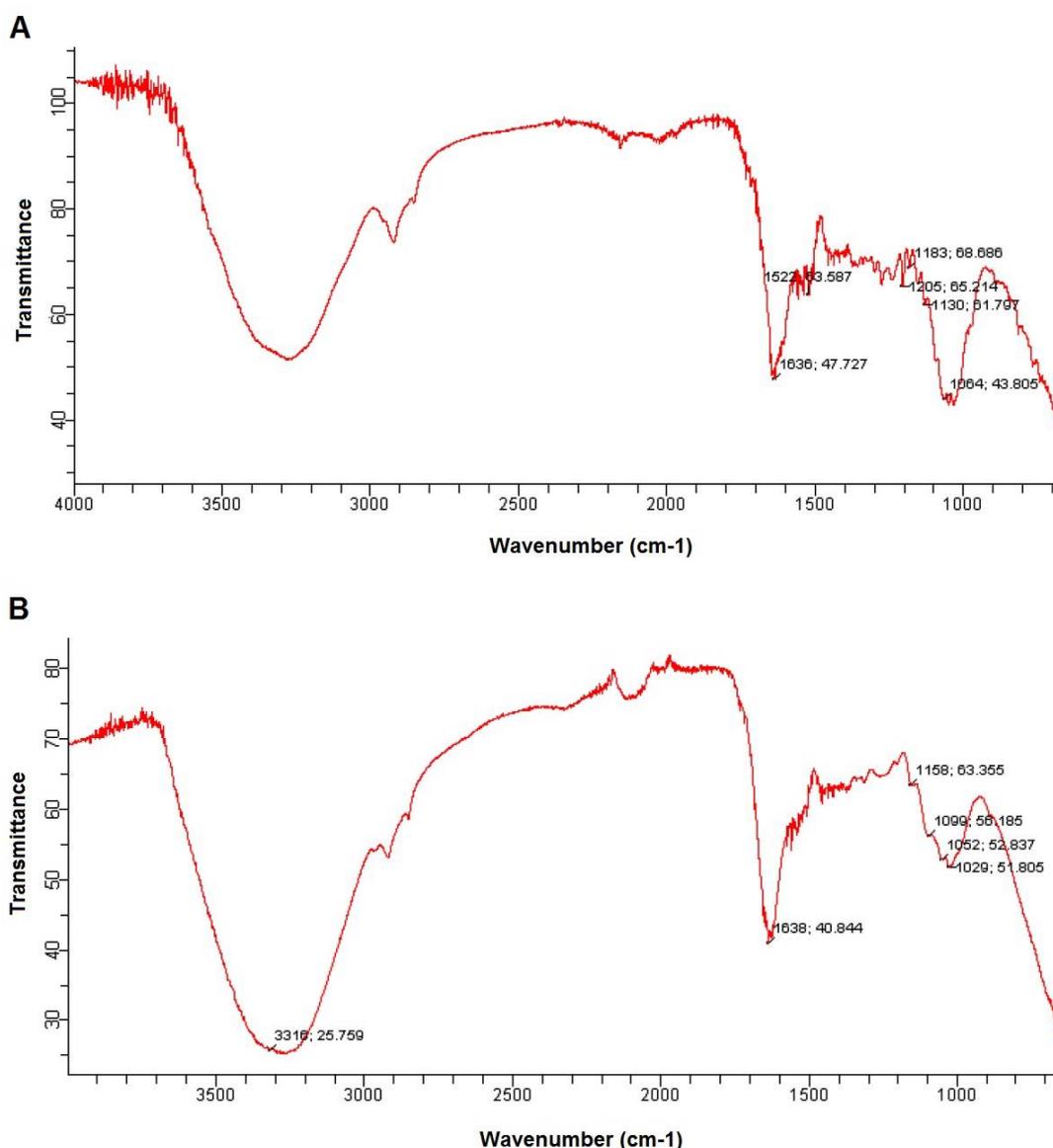


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 149 **Figure 5: UV-visible absorption spectra for the stability of synthesized AgNPs.** (A) AgNPs synthesized by *Citrus reticulata*
 150 fresh peel extract (5 ml) and 0.5 mM AgNO_3 (B) AgNPs synthesized by *Citrus reticulata* fresh peel extract (5 ml) and 1 mM
 151 AgNO_3 and UV-visible spectra were recorded immediately and after different days to determine stability of AgNPs.

152 FTIR Analysis

153 FTIR spectroscopy is useful in probing the chemical composition of the surface of the AgNPs and the
 154 local molecular environment of the capping agents on the nanoparticles. FTIR analysis was used for
 155 the characterization of the analysis of both, plant extracts and the resulting AgNPs. Transmittance
 156 spectra were observed in the region of 1000 to 4000 cm^{-1} . Bands peaks were observed at 3316, 1638,

157 1522, 1205, 1158, 1099, 1052, and 1029 cm^{-1} . These peaks are known to be associated with the
 158 stretching vibrations for N-H and O-H, $\text{C}=\text{C}$ -, C-N stretch, C-O and C-O (polyols) respectively.
 159 The bands below 3000 cm^{-1} is due to C-H stretching and C-N stretching (Fig.6 A-B).



160
 161 **Figure 6: FTIR transmittance spectra for functional group of capping and reducing agents.** (A) Shows FTIR transmittance
 162 spectra for *Citrus reticulata* fresh peel extract and (B) Shows FTIR transmittance spectra of *Citrus reticulata* fresh peel extract
 163 mediated AgNPs.

164 4. Discussion

165 Nanotechnology is one of the growing area of research in the life sciences especially biotechnology
 166 to improve the human health. For the delivery of therapeutic agents and other activities against
 167 different diseases, synthesis of stable and versatile silver nanoparticles (AgNPs) depends on the size
 168 and shape. In the present study different parts of *Citrus reticulata* (*Citrus reticulata*) fruit including
 169 fresh peel, dry peel, seeds and juice were used for the biosynthesis of silver nanoparticles (AgNPs).
 170 Change in color to brown was observed after mixing it with Silver nitrate (AgNO_3) due to the
 171 reduction of silver ions, which indicated the formation of AgNPs as previously observed in several

172 investigations [12,27]. This change in colour is due to phenomena known as surface plasmon
173 resonance (SPR). This phenomenon is because of the oscillatory movement of electrons present in
174 conduction band, in response of electromagnetic radiation [28]. Under Ultraviolet (UV) inspection,
175 nanoparticles give a characteristics absorption band due to excitation of surface plasmons [29].
176 Presently, the confirmation of the AgNPs formation was also made via UV-visible
177 spectrophotometer. Absorption peak from UV-vis spectra are also used as prediction tool for
178 stability and size of particles [30]. According to Mie's theory, the peak shifted to be observed in an
179 absorption spectrum when the mean diameter of the particles changes [31].

180 AgNO₃ was reduced with four extracts (seed, juice, dry peel, wet peel) and UV-vis absorption
181 spectra were recorded immediately after the reaction. We found that the maximum absorption for
182 seed and fresh peel and juice mediated AgNPs was obtained at 420 nm, while in case of dry peel
183 extract it was recorded at 410 nm by UV-vis spectrophotometer. The resulting *Citrus reticulata*
184 mediated AgNPs might be spherical in shape. Previously it was found that gelatin mediated
185 spherical AgNPs showed a characteristic SPR peak at around 400 nm [28]. Peel mediated AgNPs
186 found to be in high concentration when compared with AgNPs synthesized from seed extract and
187 juice. The two main compositional differences between peel and juice components are that the peel
188 contains a higher concentration of ascorbic acid than the juice, and that the peel also contains higher
189 concentrations of active components including d-limonene, hesperidin, naringin, and auraptene
190 than do the juice [4].

191 The maximum concentration of AgNPs was obtained by fresh peel extract as compared to dry peel.
192 It may because of the reason that fresh peel has more essential oil components as compare to dry
193 peel which has less essential oil contents due to evaporation of these antioxidant volatiles during the
194 process of drying [32]. It was also found that the maximum absorbance value (which represents the
195 concentration of nanoparticles) was changed for different samples under study. According to
196 Beer-Lambert law, absorbance relies upon concentration of suspension, thus higher concentration of
197 NPs leads to higher value of absorbance [33]. Previously it was concluded that the absorption
198 wavelength might be represented the estimated nanoparticles size. We supposed the estimated size
199 of *Citrus reticulata* mediated AgNPs size might be in the range of 30-50 nm as presently synthesized
200 AgNPs absorption spectra was recorded at 410 and 420 nm [30].

201 Caroling *et al.*, 2013 reported that the noble metal silver displays characteristic absorbance at around
202 410-430 nm. They suggested that Polyol components, flavonoids and terpenoids are mainly
203 responsible for the reduction of silver ions. Thus the study suggested that aqueous active
204 components like flavonoids, xanthones, and tannins of *G. mangostana* extract might reduce the silver
205 ions [18]. As, it was examined that maximum concentration of AgNPs when fresh peel extract was
206 mixed with AgNO₃. It is therefore, for optimization of reaction parameters, the fresh peel extract was
207 further used in the biosynthesis of AgNPs. We observed that colorless solution of AgNO₃ was
208 converted into light brown for 1 ml, brown for 2 ml, radish brown for 5 ml and very light brown for
209 11 ml. Absorption peak was recorded at 410-420 nm for all volume of fresh peel extracts (1 ml, 2 ml, 5
210 ml and 11 ml). The range of between 410-420 nm of absorption spectra indicated that synthesized
211 AgNPs might have estimated particle size range 30-50 nm [30]. Some literature proved that when the
212 amount of extract is increased the concentration of AgNPs also increased [34]. But in present study
213 we found inverse trend, this might be because of low concentration of AgNO₃ (0.5 mM). In AgNPs
214 dependent study, we examined that absorption of AgNPs increased with increasing concentration of

215 AgNO₃. Depending on their size and shape, nanoparticles are expected to display one or more
216 surface plasmon (SP) bands: Small spherical nanoparticles exhibit a single SP band at small
217 wavelengths, whereas large anisotropic particles reveal two or three SP bands at longer wavelengths
218 [35]. Therefore, the peaks at 440 nm in fig.3 are likely due to larger spherical size of AgNPs. We
219 monitored that fresh peel mediated AgNPs by using both concentration of AgNO₃ solution 0.5 mM
220 and 1 mM separately showed impressive stability of AgNPs at different time including 1, 4, 12, 28,
221 56, 86 and 112 Days. Furthermore, it was noted that from 1 to 28 days of the preparation, intensity of
222 the absorption peak increased and after this peak value started to decrease.

223 The FTIR analysis in present study is showing that the broad band around 3316 cm⁻¹ is related to
224 amide (N-H and OH) stretching vibrations and alkanes. The bands below 3000cm⁻¹ is due to C-H
225 stretching and C-N stretching. The intense absorption observed at 1638cm⁻¹ might be a characteristic
226 of the C=C stretching aromatic ring and this result agree with the result of the Thin layer
227 chromatography (TLC) test of previous study, which referred to the active ingredient in the *Citrus*
228 *reticulata* peel that causes the reduction of Ag⁺ ions and it was found in that the effective group is
229 flavonoids which led to the bioreduction of aqueous silver ions (Ag⁺) [12]. Polyols and phenols
230 reduce silver ions and oxidize unsaturated carbonyl groups, which leads to a broad peak at
231 approximately 1645 cm⁻¹ [36]. This peak at 1638 may also correspond to Ag-O stretching vibration
232 [37]. The bands between 1200cm⁻¹ to 1000cm⁻¹ are attributed to C-O (esters, ethers and polyol) and
233 C-N stretching vibrations (for aliphatic amines), -CH₂, -CH₃ vibrations, and -C-O-H (for alcohols)
234 and -C-O-R (for ethers or esters) vibrations as shown by literature [38]. The similarity in plant
235 extract spectra and their respective AgNPs might be attributed to the coating of the phytochemicals
236 present in extract on the surface of AgNPs [27].

237 5. Conclusion

238 It is concluded that the *Citrus reticulata* fruit extract might be very useful as bio-reductant for the
239 synthesis of AgNPs with impressive stability even after 112 days of the reaction. This indicated that
240 fresh peel extract may have good reducing and capping agents like amide, alkanes, flavonoids,
241 polyols, phenols, esters and ethers (examined in FTIR analysis) which might be involved in the
242 synthesis and stability of AgNPs. To unexposed the hidden properties of fresh peel mediated AgNPs
243 further research may be done to examine the antimicrobial activities.

244 **Author Contributions:** Conceptualization, Rabeea Muzaffar; Data curation, Rabeea Muzaffar;
245 Formal analysis, Muhammad Azam, Malka M. Samra and Muhammad Asim Raza Basra;
246 Investigation, Malka M. Samra; Methodology, Rabeea Muzaffar; Supervision, Muhammad Azam
247 and Muhammad Asim Raza Basra; Visualization, Muhammad Asim Raza Basra; Writing – original
248 draft, Malka M. Samra

249 **Funding:** This research received no external funding.

250 **Acknowledgments:** The work was supported by the Institute of Chemistry, University of The
251 Punjab Lahore, Pakistan.

252 **Conflicts of Interest:** The authors declare no conflict of interests.

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