Can mannose-binding lectin activation help in fighting bacterial pathogen in poultry production systems? – A review

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Simple Summary: In the quest to combat poultry related diseases, poultry farmer adopts different methods of which are not economical and less effective in long term. In recent years, level of production in poultry has been greatly affected by bacterial pathogens which in turn lead to great loss at the end of production cycle. Poultry animals possess an innate immunity called mannose binding lectin which could be activated at exposure to poultry related diseases. Nevertheless, most poultry farmers’ fails to take advantage of MBL activation based on some management practices adopted. Study on MBL has gained some consideration in human but less attention in poultry production. This review aims at revealing how mannose binding lectin activation can help in fighting bacterial pathogens across different poultry production systems. This includes different methods of quantifying and detecting MBL levels, management practices adopted as it affects MBL levels across different age. This knowledge will benefit poultry farmers in curtailing production cost and ensuring poultry animals’ welfare without incessant use of antibiotics.

Abstract: Bacterial pathogens have been attributed to poultry housing structure, financial strength, and incessant use of antibiotics, variable seasons and management systems practiced. Variant forms of bacterial pathogens can be detected by recognizing the molecular pattern of the pathogens through an innate immune mechanism such as mannose-binding lectin. Mannose-binding lectin (MBL) possesses an innate pattern recognition molecule that easily sequestered to region of infections and inflammations. This works by attaching itself to antigen surface thus hinders proliferation and disease activity in the host organism. Baker’s method, nephelometric assays technique, Enzyme-Linked Immunosorbent Assay technique, Polymerase Chain Reaction, Deoxyribonucleic Acid typing and other biotechnology related methods are techniques used in detecting and quantifying MBL. Mannose-binding lectin levels in serum can be influenced by age, management systems, feed formulation strategies and seasons. Therefore, knowledge of MBL should be encouraged in all aspect of poultry production, in order to discourage incessant use of drugs at a slight exposure to prevailing bacterial which can help in maximizing cost.

Keywords: mannose-binding lectin, poultry, production system, pathogens, innate immune response
1. Introduction

Poultry production success recently has been categorized based on nutrition, breeding techniques and management practices adopted [1]. The poultry management systems are based on the poultry production size and can be simplified according to free range or backyard [2], improved backyard [3], semi-intensive or smallholder production and intensive systems or commercial production [4-5]. Out of all the aforementioned management systems, commercial poultry production is known for its role in fulfilling the ever-increasing demand of poultry products. Farmers are confronted with various issues in cultivating operations and management strategies, out of which diseases thrives a lot as a major challenge [6]. Despite the development of commercial poultry production in different countries, smallholder farming system takes a large percentage in meeting the demand of the country at large [7].

Smallholder farming system has a long dated history that makes it prevalent with the rustic and certain urban populaces. Among other poultry production types, smallholder farming system account for about 90% of the national production [7-8]. Backyard production system (BPS) is the easily adopted type of poultry production methods in most part of rural areas [9]; most farmers embraced this practice as a major source of livelihood and a means to penury reduction [10-11]. Poultry birds are kept in a low put-in yield structure, with little rummaging feed rarely supplemented with edibles leftovers and grains. Poultry and their by-products are normally devoured by their owners, sold locally and utilized as donations [9]. In BPS, chicken becomes more prone to infectious and contagious diseases due to close contact with each other; because such practices expose healthy animals to infected animals [13].

2. Disease mostly affecting poultry production systems

The diseases mostly predominant in poultry production are Newcastle diseases, Fowl Pox, Fowl Cholera, Infectious Bursal Disease (IBD), Mycoplasmosis, Salmonellosis, Colibacillosis, Escherichia coli and Campylobacter spp. among others. These diseases are combated with vaccination and for some reasons unknown, vaccination becomes expensive and ineffective in some cases hereby, putting farmers in a state of great loss [14-15]. Bacterial infection in chicken is one of the major threats regarding feed efficiency, animal welfare and food security. They are key vector of food-borne disease mostly prevailing in poultry production systems [16]. Bacterial pathogens prevalence in poultry production is caused by variability in housing structure and inadequate dietary supplementation [16; 81]. Diverse forms of bacterial pathogens can be detected by recognizing the molecular pattern of the pathogens through an innate immune mechanism [17], identification of specificities of antigens and nature of response [18]. However, the ideal improvement of poultry bird’s immune response is of great concern to poultry farmers [19-20]. When health is affected, assimilation and retention of nutrients is prejudiced which affects animals’ health. When poultry health is affected, it could results to complement system activities such as receptor specificities in gut and activation of mannose-binding lectin [21-22]. Poultry has been recorded as one of the major source of human infection due to its availability and cost effectiveness [29]. Bacterial infection among other infection has in long term poised a major threat in chicken production, its prevention and treatment seems expensive. In recent years, several studies have featured the significance of innate immunity. In this review, we will focus on the major role of MBL, its mode of protection, its detection and quantifying mechanism and its impact in fighting bacterial pathogens within poultry production system.
3. Mannose-binding lectin structure and function

There is no generalized nomenclature under International Union of Immunological Societies (IUIS) for mannose-binding lectin. [35, 23] which explain the structure of MBL. Mannose-binding lectin has a bunch like structure with different oligomeric structures which has been predicted in different forms such as: dimers, trimers, tetramers, hexamers. Mannose-binding lectin expresses its orientation mostly in tetramers shape with its readiness to activates complement system thereafter knit itself to the surface of microbes [36, 37]. [Fig1]

Mannose-binding lectin also has four exons with different functions. Three indistinguishable peptide chains of 32 kilo dalton (kDa) made up the subunits, which accumulation of many subunits forms the higher order oligomers of mannose-binding lectin [27,28, 38]. Three of such affixes associate to give a classical collagenous triple helix [39]. Each chain has a lectin domain, wound loop hydrophobic neck region (containing 30 amino acids, collagen-like space and CRD area), a cysteine-rich N-terminal region and a collagenous region(115 amino acids)[40].Cysteine deposits in the cysteine-rich region forms inter-chain disulfide link forming an oligomerization.

The cysteine-rich region has ultimate function to detect carbohydrates on pathogens [41]; with potency to bind to wide range of sugar such as N-acetyl-D-glucosamine, mannose, N-acetyl-mannosamine, fructose and glucose. Such binding can be expressed as lock (antigen) and key (innate antibody) pattern [42] with great affinity to interact to pathogens such as virus, fungi and bacterial [42]. Mannose-binding lectin is a functioning member of collectin family possessing an antibody affinity with regards to its molecular structure, which helps in attacking antigens with little or no variation at binding region [41]. Binding of mannose-binding lectin can be activated by prompting complement fixation on bacterial surface, obstruction of viral infectivity and opsonizing fungi growth before activation of appropriate A\(^b\) response [19]. Mannose-binding lectin interfaces with variation of bacterial and viral irresistible pathogens through its Carbohydrate Region Domain (CRD) [19].

4. Process of binding of MBL to bacterial pathogens in poultry

Mannose-binding lectin is synthesized by the liver but can be found in nasopharyngeal secretions, inflamed joints, amniotic fluid, and serum. Immune response possesses an innate pattern recognition molecule that easily sequestered to region of infections and inflammations [23-26]. Mannose-binding lectin attaches itself to microbial surfaces by opsonophagocytosis in two different ways. The two ways are: through lectin pathway (bridging gap between innate and adaptive immunity) and through foreign cell lysis [152, 158]. Mannose-binding lectin serves as first line of protection against foreign invaders, such as bacterial, virus, fungi [158]. Also, it eliminates altered self cells such as necrotic or apoptotic cells, amyloids (protein aggregates) in poultry [158]

The most vital actuation of the supplement activities is by lectin pathway [19]. Lectin pathway is interceded by the mannose-binding lectin associated serine protease (MASP-2) prompting the formation of the C3 convertase C4b2a. This then takes charge of the cleavage of C3 and the age of various C3b parts which tie covalently to the surface of the antigens. Such regions are taken by the CR1 (CD35) receptor of the phagocyte. Progressively, some C3b is changed over to iC3b, captured by CR3 receptors. Coordinate action of mannose-binding lectin by phagocytes has been reported in various studies [153, 153, 19, and 41]. But the exact collectin receptor integrated has
not yet been distinguished [41]. Mannose-binding lectin is equally ready to advance action by a
dosage subordinate balance of cytokine discharge from monocytes. The interplay between MBL,
MASPs and ficolins are solely dependent on immune status of poultry animal, stage and line of
infection or presence of secondary agents of infection [158]

Mannose-binding lectin in apoptosis has been proposed to bind especially to apoptotic T cells
and polymorphonuclear neutrophils through the globular CRD region. [44]. Reproducing
ingestion of this ligation on the phagocyte surface of the multi-functional protein, calreticulin
has been the mode of action of mannose-binding lectin. Calreticulin (otherwise called the
cC1qR) bound to the endocytic receptor protein CD91, otherwise called the α-2-macroglobulin
receptor. Utilization of these proteins gives another case of apoptotic cell autonomy which is
catalyzed by mode of activation of the affected and self-cell structure.

Mannose-binding lectin fights bacterial by a mechanism called “Ante-antibody” [158]. Ante-
antibody is disease modifier molecules which performs a function both at early stage and at later
stage before adaptive system develops antibody production mechanism [158-159]. Protein nature
of mannose-binding lectin permits provision of a non-specific antigen against bacterial
pathogens. After decline of maternal antibodies at stage of exposure, mannose-binding lectin
helps in fighting infections before the full active state of antibody repertoire. [44, 152, 153].

Mannose-binding lectin does not only bind bacterial but also binds viral diseases such as avian
flu virus (AIV). It straight forwardly represses hemagglutinin action and infectivity of a few
strains of AIV. Moreover, MBL goes about as an opsonin; upgrading neutrophil reactivity
against AIV [45]. Mannose-binding lectin provides a pre-immune response against bacterial
pathogens. Therefore, mannose-binding lectin can bind various strains of bacterial especially in poultry
animals by its presence and suitable level of mannose-binding lectin by hindering the growth,
development and augmentation of bacterial, viral and fungi pathogens in poultry [47].

The list of some of the bacterial that can mannose-binding lectin can bind are shown on table 1.

5. Past and present methods of detecting and quantifying mannose-binding lectin in
Poultry

Bakers Methods

At early stage, mannose-binding lectin was first detected by its capacity to opsonize heat killed
baker’s yeast. Its deficiency was generally associated with failure to opsonize yeast bacterial
[54]. This was a conventional way of detection known to be economically reasonable. Baker’s
methods have ability to uncover both quantitative and qualitative data of bacterial present and its
nature (pathogenic or non-pathogenic) in sampled organism’s analyzed [55-56].Techniques for
recognition are as follows: pre enhancement, sample improvement, specific plating, and
biochemical screening in relation to serological affirmation. Distinguishing proof of the strains
of bacterial consumes time. Hence, demonstrates the requirement for further less time consuming
tests [57-58]. Baker’s method success relies basically on multiplication ability of organism to
reveal visible strains [59-60]. This method is quite labour intensive due to many processes
involving expertise to prepare culture medium, inoculating plates and strains counting. It takes
time for just identification of viable pathogens [56]. The challenges mentioned leads to reasons
to invent such a rapid method. Bakers Methods have this limitation that it can only show the
quantify activity rather than showing the concentration (gravimetric) of amount of mannose-binding lectin present in the serum [61]. The challenges listed above leads to invention of rapid methods such as nephelometric assay among other rapid assays [62].

**Nephelometric Assay Technique**

Nephelometric assay measure permits high-touchy C-reactive protein detection in serum and plasma with specie self-reliability features. It is mostly utilized for pediatric patients, children and can test volume up to 44ul. It has a simple methodology contrasted with Enzyme Linked Immunosorbent Assay (ELISA) and short examining time compared to baker’s method yet less dependable than ELISA. It has a short identification scope of 0.09 – 5.6ul/ml, because of its short scope of recognition. It can be utilized for some specimen in brief time (Table 2). It gives off an impression of being unsatisfactory after a few years. [61]. It has short range of detection. [62]

**Enzyme Linked Immunosorbent Assay Technique**

Enzyme Linked Immunosorbent Assay (ELISA) is a fast, delicate and particular serological test. It is recently known and richly utilized method for measuring MBL protein in serum. Enzyme Linked Immunosorbent Assay reveals the incidence of hormones, peptides, antigens, antibodies, and pathogens utilizing enzyme connected antibodies present in the blood, urine, swab and any other fluids [63-64]. Such strategies include a few brooding strides to paint antibodies, antigens and additionally blocking techniques, track by a progression of washing steps. Protein covering steps may require a brooding period between four hours to twelve hours; consequently, using an ELISA test to identify an analyte may take from a few hours to as long as 24 hours [65]. There are different types of antigens which can be used to coat ELISA plate such as: whole cell antigen, sonicated antigen, potassium thiocyanate extract, glycoprotein antigens, lipoprotein antigens, sodium salicylate extract, and heat extract antigen [66-68]. Antigen to be used can be grouped based on their individual properties such as: specificities, preparation duration, antibodies to be detected, ability to distinguish between infected or immune resistant animals [68]. Its target DNA ranges from 0 to 10um concentration, it shows visible colour from red to blue or grey. The colour changes as the DNA concentration increases [72]. ELISA can be used for detecting other avian species and human.

**DNA Typing and Polymerase Chain Reaction (PCR)**

Use of biotechnology techniques such as DNA typing, Genotype typing, PCR techniques: This has really helped in precision and easy detection of bacterial pathogens in liver, blood, swab of chickens [73]. Biotechnology leads to brisk method of an assay, which reveals result within a short time. It helps in early detection and shows variation of microorganism at varying length and numbers [75]. Selection of a particular method to be adopted are based on some factors such as choice of testing needed, range of detection, quantity of sample to be analyzed, field of expertise of technologist [75], capital available for purchase of equipment and nature of tools [74]. It is an expensive tool compared to other methods (Table 3). Even though PCR appears to be the latest means of detecting mannose-binding lectin in serum and most powerful, it has the following limitations which are, usage is quite complicating, need a trained personnel. It also require hygienic, high sanitary environment for test procedure.
Mannose-binding lectin deficiency in poultry

Poultry wellbeing can be ascertain, provided the immune system produces sufficient amount of mannose-binding lectin (MBL). Mannose-binding lectin produced can then be transfer to the blood hereby, up-regulating the production amid intense phases of infection diseases [86-88]. Mannose-binding lectin deficiency in other word is immunodeficiency, such deficiency results in malfunctioning of innate immunity response. At this point, poultry animal’s susceptibility to diseases increases to great number of associated infections [23]. Mannose-binding lectin ties to a scope of clinically significant bacterial pathogens demonstrating a variable example of dependability [43]. It possesses opsonin and lectin pathway activator, which ties across numerous lectin spaces to the rehashing sugar molecules shown on the surface of varied clinically applicable microbial species. [23, 79, 89] reported that liver and serum are the two major areas for isolation of chicken mannose-binding lectin (cmbl) using cmbl-specific monoclonal antibodies. The possibility of comparative absence of mannose-binding lectin may predispose the host to disease [88]. Basically, mannose-binding lectin-subordinate opsonin insufficiency in serum corresponds with a phenotype of intermittent infectivity [89]. Absence of one out of three amino acid single nucleotide polymorphisms (SNPs) in exon 1 of the mannose-binding lectin quality disturbs the collagen helix [90]. Such absence creates the impression that scattered collagen chain acts like a predominant harmful reactions. Scattered collagen chains results in a decline pathway levels of mannose-binding lectin with great likelihood of initiating complement activities, which can lead to its deficiency [21]. Mannose-binding lectin insufficiency and increased bacterial infection in chicken are positively correlated, hereby signifies that mannose-binding lectin sufficiency fights different predisposing infections in poultry animals [91]

6. Mannose-binding lectin activation: A tool for combating bacterial pathogens in poultry production systems

Food-borne infections incorporate general classes of ailments with divergent degrees of severity, extending from mild illness to severe ailment caused by either poisons discharged by the vector, or by the organism itself [92, 93]. [94] has a global view of intensifying the aptitude of global and local facilities in the supervision of food borne pathogens such as campylobacter, escherichia, listeria, salmonella among other bacterial pathogens.

Campylobacter spp is known to be pattern less in nature forming wide variation in structure and difficult to trace hereby, forming a store house for diseases [95]. It is one of the main sources of zoonotic enteric diseases in most developing and developed nations [97]. Most of the causes of campylobacter are widely attributed to poultry [98] cattle, pigs, sheep and ostriches, cats and dogs are also carrier of campylobacter [99]. Campylobacter spp is known as the normal human gastroenteritis, which accounts for one of four key worldwide reasons for diarrhea sicknesses [94].Campylobacter affects meat production [95, 101]. On-field inspecting of poultry can be performed by gathering or slaughtering chickens before butcher and taking fecal droppings or swabs from the cloaca [100, 99]. Study by [101] reveals organically raised flocks of three farms confirmed after strict analysis of seven consecutive crop cycles turnout that colonization of campylobacter in the intestine of chicken is strongly influenced by management practices and not on effect of rearing period duration. In many developing nations like South Africa, the revealed frequency has expanded consistently accounting as the most recent in twenty (20) years, among other enteric infection with constant increase every year [102]. Study by [103] recorded
Parallel and consistent access to water and soil was linked as a major route for *Campylobacter* infection in poultry [104]. *Campylobacter* positive flocks were observed to be relatively high among smallholder farming system compared to commercial farming system accounting to about 49.2% and 36.7% respectively [104]. This could be resulted to incessantly exposure to free grazing, along these lines expanding the quantity of conceivable pathogenic sources [104,105]. Chickens should be restricted from roaming around and picking unscreened feed in order to manage bacterial within the chicken house most especially at 4th to 6th week[79], strict hand hygiene practices by consumers, sanitary measures of both animal feed and housing [106], quarantine test before introducing birds from hatchery, children below five years old are mostly prone thus, there is need for conscious cleaning of hands, genetically immune advantageous breed should be considered for production [100]. Mannose-binding lectin appears to be sufficient from day old to beginning of 4th week and from 6th week to adult stage under proper feeding and good management systems [79, 82, and 23].

*Escherichia coli* (E. coli) are microscopic organisms mostly lives in human and avian digestive organs [107]. [96] reported that *E. coli* disease results to about 48 million sick patients, 128,000 hospitalized, 3000 death cases from food borne related cases every year in United States, which makes it one of the prevailing bacterial pathogen. *E. coli* O157:H7, Extra intestinal Pathogenic *E.coli* (ExPEC) and different strains like *E. coli* O145 and *E. coli* O121:H19 deliver a poison called Shiga poison, which causes ailment in people and are in charge of most prevailing related *E.coli* infections [107, 108]. *E. coli* microscopic organisms in poultry are not responsible for causing diseases on vector but on any human who consume such infested animals and mostly the digestion tracts serves as shelter for such pathogen [109]. *Escherichia coli* causes series of illness disorders in poultry, including yolk sac disease, respiratory tract contamination, swollen head disorder, septicemia, airsaculitis and pericarditis [110]. Infections most cases are triggered or activated by predisposing agents, such as bacterial infection, environmentally influenced agents and interaction between bacterial and environment [109]. *E. coli* confines from chicken feacal samples and the poultry house condition attacked the inside organs of tested chickens [111]. Contrary to most findings, destructive related genotypes and phenotypes, incorporating into vivo harmfulness, of fecal exudes from healthy chickens thus, infect both human and poultry by contact with chicken intestine, carcasses, urine and some meat products [112]. *E.coli* has been a major challenge to poultry production systems, but can be managed under proper management systems, quarantine techniques, avoiding infected animals [113] and crossing of genetically resistant poultry [83].

*Salmonella spp* has been known to withstand and procreate under low temperature and high thermal condition. In a study reviewed by [114], diverse food-borne related pathogens have been related with the nearness of *Salmonella spp*. Such items includes meat, chicken, pork and their side-effects, fish, eggs, dairy items, chocolate, breakfast oats, snacks and almonds, shelled nut margarine, newborn child recipe, and pet treats. *Salmonella* is transmitted by means of the fecal-oral course [115], through contact with contaminated poultry birds from various sources, vertical transmission from affected hens and debased feed [116,117].*Salmonella* infected poultry normally seem healthy, but can shed microscopic organism discontinuously thus, making infected bird (s) which shed microbes hard to be distinguished [117]. *Salmonella* acknowledges the resistant structure amid attack of intestinal epithelial cells, by distinguishing the pathogenic microorganisms through the enlistment of different phagocytic cell lineages [115]. Range of
innate immune response is in charge of the early identification, intense control of *Salmonella* such as neutrophils depletion and mannose-binding lectin [118]. It has been shown that *Salmonella* attacks the intestinal epithelial layer at the antigen-sampling microfold cells. Subsequently, *Salmonella* encounter dendrite cells and macrophages, followed by an influx of neutrophils, monocytes and more macrophages [119]. Moreover, historically, coccidiosis disease leaves chickens more defenseless to different contaminations, for example, *salmonellosis* and necrotic enteritis [120,121]. *Salmonellosis* is affects nourishment and well-being concern of poultry animals, which can cause serious sustenance borne sickness in people, and shows a significant worldwide weight of difficulty in developing and developed countries [122]. Control measures for *salmonella* in chicken production system are hygienic strategies and proper management practices [123], vaccination using dead animal as inoculums [123], feeding birds with specific antibodies from egg gotten from hyper-immunized hens [124], use of genetically resistant chicken germline [109] and building of innate immune system such as mannose-binding lectin [123].

*Listeria spp.* is widely seen in temperate region especially in soil, manure/sewage, animal feed, water, poultry feaces or excreta, plant extract and poultry walls [125]. *Listeria* is generally observed in birds such as poultry, geese, turkey serving as primary host and carrier of *listeria*, playing a critical function in infecting both the animals and the poultry production system at large [77,127]. Its ability to simulate other bacterial symptoms such as coccidiosis, staphylococcus makes it harder to be detectable [127]. [77] reviewed *listeriosis* (*silage diseases*) as to be infrequent but severe food borne disease, of which its causal agent is bacterium *listeria monocytogenes*. It can also be found in both prepared and processed foods particles and has been greatly linked to high morbidity and mortality most especially in poultry production [129]. *Listeriosis* symptoms seem not specific with intricate diagnosis. In extreme cases it expresses itself as septicemia and encephalitis [129]. Age has a great influence in susceptibility to listeriosis. Young birds are susceptible with mortality rate up to 40% revealing chronic infection and adult seldom has meningocerephalitis and death in some cases. Alteration in immune response, damp/moist conditions, cold and moist litters are factors which lead to easy infection of *listeriosis*[130-132]. Asian countries recorded 48 cases of *listeriosis* from 1996 to 2008 in Taiwan [133], In China, 479 isolates were accounted from 1964 to 2010 [128]. Recently, South Africa had 365 cases of *listeriosis* which were detected in Gauteng, resulting in 28 deaths, which led to closure of an abattoir in Gauteng after some meat samples were tested positive to *listeria* pathogen [134]. *Listeria* can be treated by use of systemic antibiotics [135], bacteriophage therapy [136,137], essential oils from plant origin such as thyme, rosemary [138], use of Probiotic [137], use of herbal remedy such as plant extracts [138-140], use of nanotechnology [127], building of innate immune response among which are toll like receptors, cytokines, avian egg antibodies, immunotherapy, mannose-binding lectin [127]. It can be easily prevented and controlled by proper hygiene, proper disposal of rotten food products, water treatment and constant cleaning of water tanks and proper disposal of contaminated silage and not fed to animals [127]

7. Mannose-binding lectin association with management practices in poultry production systems

**Commercial poultry System:** Although, adopting serene environment in commercial production system in combating bacterial pathogens do creates some variability. Such as
chickens are reared differently from day old to maturity and development of micro flora varies in different poultry animals. Some poultry are hatched under serene environment which inhibit growth of micro flora [142]. At slight exposure to bacterial diseases most of them are affected because their immunity was not developed from hatchery to maturity stage [142]. Commercial system production system mostly put measures against predators by fencing the chicken houses and proper vaccination at different age this act mostly do not allow growth of immune response such as mannose-binding lectin against infection [144]. There is need to formulate feeds that will boost mannose-binding lectin in the production system aiming at sequential increment of productivity level.

**Smallholder poultry production systems:** plays a major role in socio economic activities [144], gifts and token [145], strengthen in-laws relationship [146], provision of manure [144], with short life cycle and quick turnovers [144] It has low input and output mechanism; it also maximizes low quality available feed for maintenance and high productivity [146]. Smallholder farmers have adopted ethno veterinary method with no appropriate measurement techniques [145] due to limited veterinary extension services and this method exposes chicken to bacterial infections among other infections [81]. In smallholder farming systems in their little way, strict consideration are considered in rearing chicken such like consistency in cleaning of chicken house in order to discourage growth of bacterial pathogens [143]. Predation and diseases are major factors affecting smallholder farmers in most rural areas because of land topography, establishing healthy management system which build innate immunity such as mannose binding lectin as against incessant use of vaccination and ethno-veterinary practices in cases of low income earner farmers in managing diseases in this region are to be embraced in order to reduce infections.

**Age:** Mannose-binding lectin level varies at different week in chickens. Indigenous chicken should be discouraged or restricted from grazing at age week 4 to week 7 because the mannose-binding lectin level in chicken at this age reduces and the chicken are prone to be infected with bacterial diseases among other diseases at a slight exposure to unclean food materials [86,79,23].

**Season:** Study by [143] reveals that season has impact on prevalence of bacterial diseases and summer predisposes chicken to more bacterial diseases, farmers should take some strict management practices at this time of the year in order to discourage bacterial infection within and among the chicken house. Whereas in commercial production system, vaccination and treatment are done at slight exposure to infection, which reduces the immune response to infection at entry hereby discourage the activation of mannose-binding lectin against infections [143]. Quarantine measures, general cleaning and proper management system are still the best ways to hinder proliferation of bacterial in poultry production [145].

**Feed Formulation Strategies:** Present day dietary and farming procedures have been intended to create range with greater potential for development, yield and bolster effectiveness that have brought about compromising health status [66]. Non starchy polysaccharides are available in plant based feeds which contains organic acids that are easily fermented in the cecum of chickens and such reaction are harmful to growth of pathogens, such fermentation discourages binding of MBL in the serum of chicken hereby predisposing chicken to bacterial infection [148]. Presence of sufficient amount of amino acid in the feed leads to right orientation and binding strength of mannose-binding lectin in the chicken. Amino acids also produce micronutrients such as iron which helps in natural resistance to colonization of pathogens in the
body system through activation of MBL [92]. Mannose-binding lectin as an associate of Ca\(^{2+}\)-
dependent animal lectin binds through its CRD and such binding occurs in the presence of
calcium; which depicts that feed formulated with calcium supplements deficient inhibits
activation of mannose-binding lectin [149]. Immune systems response to pathogenic bacterial
diseases was linked to dietary zinc insufficiency [66]. Such as Zinc- Methionine (Zn-Met) as a
major component of cellular poultry immunity, Zinc-Sulphur (Zn-Su) and Zinc-Oxygen (Zn-O)
in chicks feed in corn- soybean diets [150]. Zn-Met and Zn-O responds to antibody in relation to
*salmonella pullorum*, *salmonella eneteritis* and *Escherichia coli* among other bacterial [66]. In-
vitro macrophage phagocytosis is boosted by Zn-Met supplemnetations which augment
macrophage phagocytosis reaction in young poultry against *salmonella* and survival of adults
against *E.coli* [152, 153].Zinc is a major component of diverse metallo-enzymes such as the one
involve in gene replication. DeoxyribonucleicAcid (DNA) and Ribonucleic acid (RNA)
polymerase deficient in zinc concentrations reduces the activity of deoxythymidine kinase (zinc-
dependent enzyme). Such effect results in decreasing protein and collagen synthesis [152,153].

Mannose-binding lectin level can be significantly affected by supplementing animal feeds with
essential oil such as thymol and carvacrol to poultry diet. As the uses of essential oil gets a rapid
attention of poultry farmers, its usage can help in providing an environment for easy binding of
lectin to infectious diseases in poultry production [31]. Aromatic compounds separated from
plant source are called essential oils. Some studies have been investigated about importance of
MBL in channel catfish [31] and rainbow trout *Oncorhynchus mykiss* [82] and come about show
enhancements in weight gain, feed conversion ratio, and insusceptibility to infectious bacterial
disease.

According to [79] there is a positive correlation between mice (rat family) and chicken mannose-
bindig lectin level. Reduction in zinc concentration can affect binding of Mannose-binding
lectin in poultry. Broad analysis has been done in observing the connection of nutrition and
immunology in human especially in relation to zinc inadequacy [54]. But such impacts in poultry
vulnerability and irresistibility to bacterial infections have been vaguely studied [55].

### 7. Conclusion and Application

It is well known that poultry products are generally acceptable due to its availability and cost.
Findings from this study elucidates that bacterial is a major threat for poultry production. While
the knowledge about mannose-binding lectin can help most poultry farmers to manage bacterial
infection throughout divergent rearing season with little or no loss in production. Mannose-
binding lectin recognize and bind the surface of bacterial, viruses and fungi by agglutinating
microbial surfaces, lysis of Gram-negative bacterial and also opsonize a wide range of potential
pathogens for phagocytosis. Mannose-binding lectin also influences phagocytosis in the absence
of complement activation through an interaction with one or more collectin receptors. In
addition, MBL can alter the function of microbial structures to prevent infection. Mannose-
binding lectin levels can be influenced with age, management system, and season and feed
formulation strategies. Therefore, it can be said that mannose-binding lectin is a way of
maximizing the innate potential of poultry animals against bacterial disease at point of entry
before the activation of adaptive immunity. However, Mannose-binding lectin insufficiency has
been known to expose chicken to different infections such as virus, fungi and bacterial.
Consequently, mannose-binding lectin should be extensively study in poultry production as it is
studied in human; such study will help poultry farmers to maximize the immunosuppressant
ability of poultry animals in order to maintain poultry health across different age and season to
enhance food security from farm to fork and to improve productivity and profit. Therefore,
knowledge of MBL should be encouraged in all aspect of poultry production, which will leads to
cost effectiveness in usage of drugs thus, discourage incessant use of drugs, consideration of
exposure to prevailing bacterial.

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Table 1: Shows bacterial, organisms and site of binding mannose-binding lectin

<table>
<thead>
<tr>
<th>Bacterial</th>
<th>Organisms</th>
<th>Site of binding</th>
<th>References</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>Bifidobacterium bifidum</em></td>
<td>Chicken</td>
<td>Serum</td>
<td>[48]</td>
</tr>
<tr>
<td><em>Burkholderiacepacia</em></td>
<td>Chicken</td>
<td>Carbohydrates structure</td>
<td>[48]</td>
</tr>
<tr>
<td><em>Chlamydia pneumonia</em></td>
<td>Chicken</td>
<td>Serum</td>
<td>[49]</td>
</tr>
<tr>
<td><em>Escherichia coli</em></td>
<td>Chicken</td>
<td>Carbohydrates Structure</td>
<td>[50]</td>
</tr>
<tr>
<td><em>Fusobacterium spp</em></td>
<td>Ruminant/monogastric</td>
<td>Serum</td>
<td>[51]</td>
</tr>
<tr>
<td><em>Haemophilus influenza</em></td>
<td>Monogastric</td>
<td>Serum</td>
<td>[48]</td>
</tr>
<tr>
<td><em>Leptotrichi abucaulis</em></td>
<td>Chicken and Avian species</td>
<td>Serum</td>
<td>[52]</td>
</tr>
<tr>
<td><em>Listeria monocytogenes</em></td>
<td>Chicken</td>
<td>Serum</td>
<td>[48]</td>
</tr>
<tr>
<td><em>Mycobacterium avium</em></td>
<td>Avian species</td>
<td>Serum</td>
<td>[52]</td>
</tr>
<tr>
<td><em>Neisseria meningitides</em></td>
<td>Livestock</td>
<td>MASP</td>
<td>[53]</td>
</tr>
<tr>
<td><em>Propionibacterium acnes</em></td>
<td>Avian species</td>
<td>MASP 1 and 2</td>
<td>[51]</td>
</tr>
<tr>
<td><em>Staphylococcus aureus</em></td>
<td>Avian species</td>
<td>MASP 1 and 2</td>
<td>[48]</td>
</tr>
<tr>
<td><em>Streptococcus pneumoniae</em></td>
<td>Avian species</td>
<td>MASP 1 and 2</td>
<td>[52]</td>
</tr>
<tr>
<td><em>Campylobacter spp</em></td>
<td>Avian species</td>
<td>MASP 1 and 2</td>
<td>[51]</td>
</tr>
</tbody>
</table>
Table 2: past methods of detecting mannose binding lectin

<table>
<thead>
<tr>
<th>Organism</th>
<th>Mannose-binding lectin Assay, detection and sample volume</th>
<th>Cross Reactivity and Interference</th>
<th>Price and Sample Size</th>
<th>References</th>
</tr>
</thead>
<tbody>
<tr>
<td>Chicken</td>
<td>Baker’s Yeast</td>
<td>Less sensitive</td>
<td>Small sample 1-10 sample sizes, consumes time, less reliable and less reproducible</td>
<td>[54]</td>
</tr>
<tr>
<td>Human Adult <em>Homo sapiens</em></td>
<td>Baker’s yeast</td>
<td>Not Sensitive</td>
<td>Small sample size and not expensive</td>
<td>[61]</td>
</tr>
<tr>
<td>Pediatric Patient</td>
<td>Nephelometric Assay 0.09 - 5.6 ug/ml Sample volume of 44ul</td>
<td>Not specific capture and primary detection. Used Rabbit anti- antibody Shorter Assay Time but not reliable and Less- Sensitive</td>
<td>Expensive and Large Sample size</td>
<td>[76]</td>
</tr>
<tr>
<td>Children 0.1 – 15.9 years</td>
<td>Nephelometric Assay 0.09 - 5.6 ug/ml</td>
<td>Less Sensitive</td>
<td>Large Sample Size and Less Expensive</td>
<td>[77]</td>
</tr>
<tr>
<td>Organism</td>
<td>Mannose-binding lectin Assay, detection and sample volume</td>
<td>Cross Reactivity and Interference</td>
<td>Price and Sample Size</td>
<td>References</td>
</tr>
<tr>
<td>----------------------------------</td>
<td>-----------------------------------------------------------</td>
<td>---------------------------------------------------------------------------------------------------------------</td>
<td>------------------------------------------------------------</td>
<td>------------</td>
</tr>
<tr>
<td>Human Adult</td>
<td>ELISA 0.05–2.5 Ag/ml</td>
<td>Specific capture and primary detection antibodies. Short Assay Time and very sensitive</td>
<td>Very Expensive and Medium Sample Size</td>
<td>[78]</td>
</tr>
<tr>
<td>Pedriatric patients</td>
<td>ELISA</td>
<td>Not Specific monoclonal and polyclonal detection. Short Assay time, sensitive and reliable</td>
<td>Expensive</td>
<td>[76]</td>
</tr>
<tr>
<td>Chicken <em>Gallus gallus domestica</em></td>
<td>ELISA Combine use of ELISA and PCR</td>
<td>Specific monoclonal and polyclonal detection. Very reliable</td>
<td>Expensive and Medium sample size</td>
<td>[79,23,80]</td>
</tr>
<tr>
<td>Mice <em>Mus</em></td>
<td>ELISA</td>
<td>Very Reliable</td>
<td>Expensive and medium sample size</td>
<td>[29]</td>
</tr>
<tr>
<td><em>Ictalurus punctatus</em> and <em>Ictalurus furcatus</em> Blue and Channel catfish</td>
<td>DNA Typing Real Time PCR</td>
<td>Extremely Reliable NCBI Blast through Open reading frame methods</td>
<td>Extremely Expensive and Reliable</td>
<td>[31]</td>
</tr>
<tr>
<td>Chicken <em>Gallus gallus domestica</em></td>
<td>Genomic DNA Real Time PCR Multiplex PCR</td>
<td>Two-step real-time reverse transcription PCR (real-time RT-PCR) method.</td>
<td>Extremely Expensive, highly reproducible and highly reliable</td>
<td>[83,84]</td>
</tr>
<tr>
<td>Human</td>
<td>DNA Typing Real Time PCR Multiplex PCR</td>
<td>Detection of allele, Haplotyping, sequence-specific oligonucleotide probes, amplification of the variants</td>
<td>Economical and renders rapid: reliable results without ambiguities. Hardy-Weinberg correlated,</td>
<td></td>
</tr>
</tbody>
</table>

Table 3: Present and Future methods of detecting mannose-binding lectin
Fig 1: Diagrammatic structure of mannose-binding lectin attaching itself to bacterial pathogens [41].