

Development of a transformation method for *Metschnikowia borealis* and other CUG-serine yeasts.

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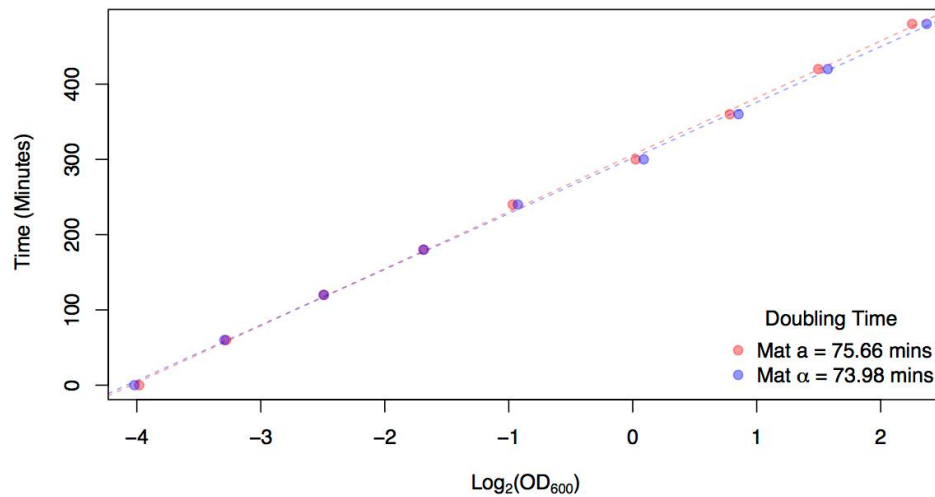
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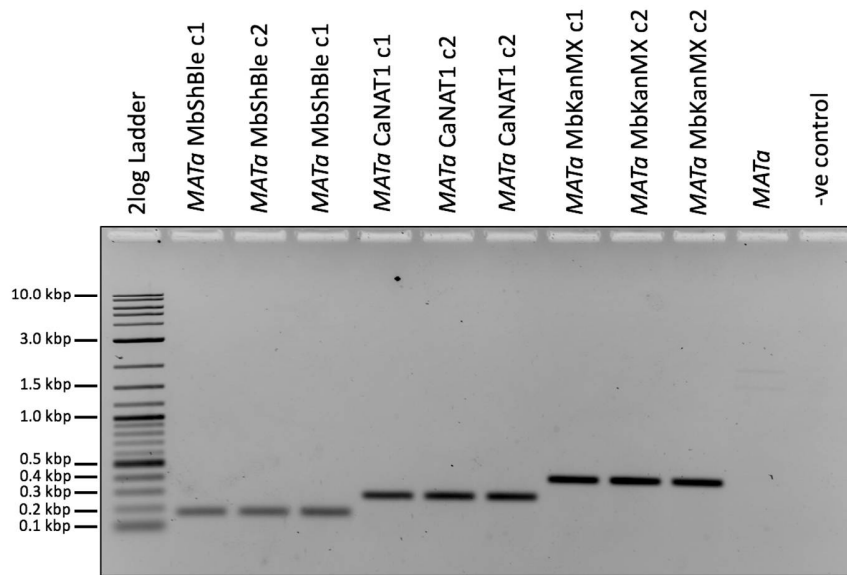
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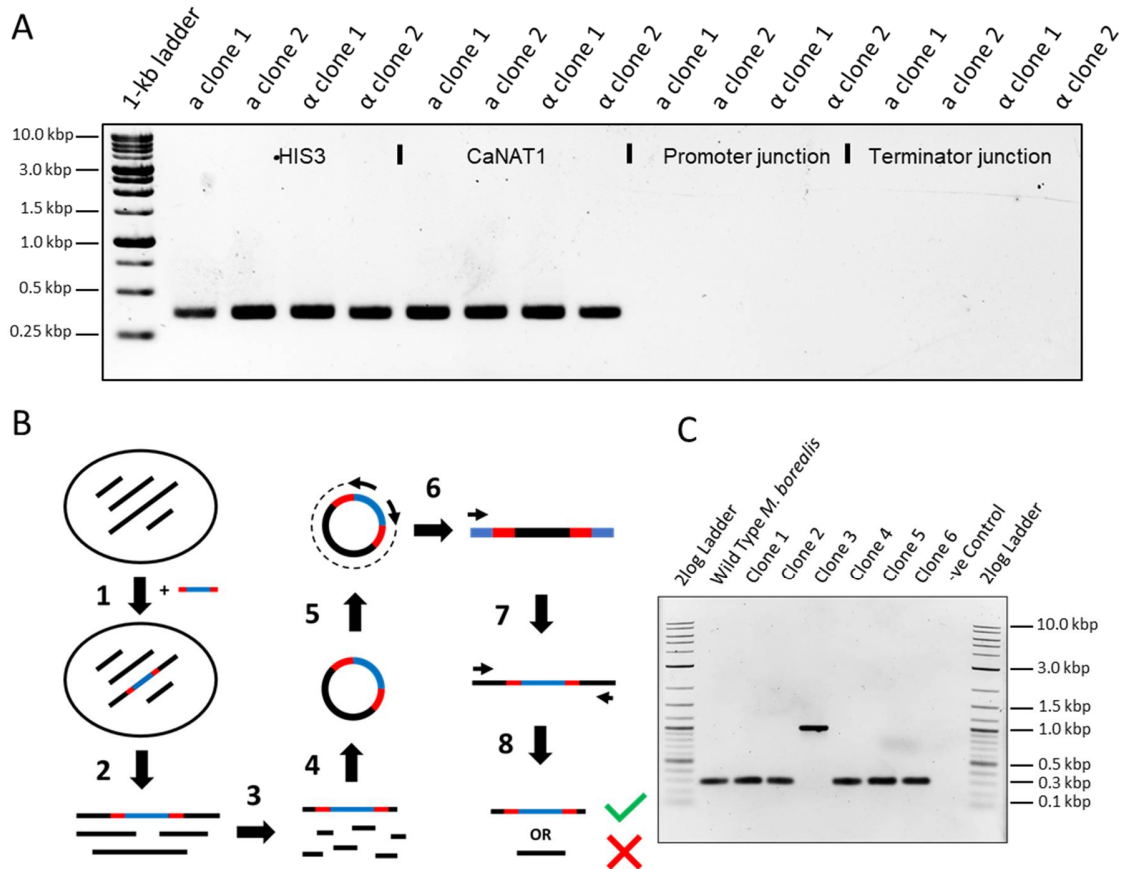
Supplementary Figures and Tables:



Supplementary Figure S1. Growth rate of *M. borealis* MATa and MATα. Three cultures of each mating type of MATa and MATα were grown to mid-log phase ($OD_{600} = 1.0$), and diluted in 50 mL of YPAD to an optical density of 0.065. Each culture was then grown at 30°C with shaking at 225 rpm, and OD_{600} was recorded at 1-hr time points for 8 hours. The average OD_{600} of the three cultures for each mating type was recorded at each time point, and the doubling times were calculated as the slopes of the lines of best fit of $\text{Log}_2(OD_{600})$ versus each time point in minutes.



Supplementary Figure S2. Genotyping transformants. Three colonies of *M. borealis* *MATa* that were transformed with MbShBle, CaNAT1, and MbKanMX (Figure 2) were genotyped by Multiplex PCR (Quiagen) using primers that amplify each selectable marker. Expected sizes were 185 base-pairs (MbShBle), 283 base-pairs (CaNAT1), and 404 base-pairs (MbKanMX). Two negative controls include PCR performed with DNA isolated with untransformed *MATa* strain as well as without any DNA (-ve control). The PCR was run for 28 cycles with all three sets of primers present in each reaction.



Supplementary Figure S3. Identification of insertion sites. *M. borealis* mating types a and α were transformed with a PCR-linearized CaNAT1 in yeast alternative nuclear code, flanked by 60 base-pair sequences of the *M. borealis* *HIS3* promoter and terminator by electroporation. (A) Two colonies of a and α mating types were screened by PCR to look for a targeted knockout of *HIS3*. Lanes 2-5 used primers that bind within the *M. borealis* *HIS3* gene (expected size 416 base pairs), lanes 6-9 used primers that bind within the CaNAT1 marker (expected size 388 base pairs), lanes 10-14 used primers to amplify across the *HIS3* promoter-CaNAT1 insertion junction (expected size 682 base-pairs), and lanes 14-17 used primers to amplify across the CaNAT1-*HIS3* terminator junction (expected size 779 base-pairs). (B) Schematic of the protocol used to identify the insertion site: 1) Lithium acetate/electroporation to transform *M. borealis* with the marker DNA for insertion; 2) Alkaline lysis to isolate *M. borealis* DNA; 3) Restriction digest with *Cfo*I; 4) Ligate sticky ends with T4 ligase; 5) PCR amplify the adjacent genomic DNA; 6) Sequence PCR product; 7) Design primers ~150 base-pairs upstream and downstream of the insertion site; 8) PCR amplify expected insertion site. (C) Confirmation of one insertion site. Primers were designed to amplify across the insertion site identified in clone 3, and the site was PCR-amplified in wildtype *M. borealis*, as well as transformants 1-6. Expected size of the site is ~300 base-pairs (wildtype) and ~1000 base-pairs (with the CaNAT1 vector insertion).

Supplementary Table S1. Identification of antibiotic sensitivity. Cultures of *M. borealis* MAT α and MAT α were grown to OD₆₀₀ of 1.5, concentrated to OD₆₀₀ = 3.0, and three dilutions were plated onto YPAD with various concentrations of zeocin or nourseothricin or combination of these two, or geneticin (G418). Plates were incubated for 2-4 days, and colonies were counted. Zeo = zeocin; NTC = nourseothricin; G418 = geneticin; NG = no growth; C = confluent growth.

Growth Time	MAT α						MAT α					
	2 days			4 days			2 days			4 days		
	10 ⁰	10 ¹	10 ²	10 ⁰	10 ¹	10 ²	10 ⁰	10 ¹	10 ²	10 ⁰	10 ¹	10 ²
Zeo 50 mg L ⁻¹	1	NG	NG	7	2	NG	2	NG	NG	8	3	1
Zeo 75 mg L ⁻¹	NG	NG	NG	1	NG	NG	NG	NG	NG	1	1	NG
Zeo 100 mg L ⁻¹	NG	NG	NG	1	NG	NG	NG	NG	NG	NG	NG	NG
Zeo 125 mg L ⁻¹	NG	NG	NG	NG	NG	NG	NG	NG	NG	NG	NG	NG
NTC 50 mg L ⁻¹	NG	NG	NG	NG	NG	NG	NG	NG	NG	NG	NG	NG
NTC 75 mg L ⁻¹	NG	NG	NG	NG	NG	NG	NG	NG	NG	NG	NG	NG
NTC 100 mg L ⁻¹	NG	NG	NG	NG	NG	NG	NG	NG	NG	NG	NG	NG
NTC 125 mg L ⁻¹	NG	NG	NG	NG	NG	NG	NG	NG	NG	NG	NG	NG
Zeo 25 mg L ⁻¹ NTC 25 mg L ⁻¹	NG	NG	NG	NG	NG	NG	NG	NG	NG	NG	NG	NG
Zeo 25 mg L ⁻¹ NTC 50 mg L ⁻¹	NG	NG	NG	NG	NG	NG	NG	NG	NG	NG	NG	NG
Zeo 50 mg L ⁻¹ NTC 25 mg L ⁻¹	NG	NG	NG	NG	NG	NG	NG	NG	NG	NG	NG	NG
Zeo 50 mg L ⁻¹ NTC 50 mg L ⁻¹	NG	NG	NG	NG	NG	NG	NG	NG	NG	NG	NG	NG
G418 200 mg L ⁻¹	C	C	NG	C	C	C	C	C	NG	C	C	C
G418 300 mg L ⁻¹	NG	NG	NG	8	1	NG	NG	NG	NG	12	1	NG
G418 400 mg L ⁻¹	NG	NG	NG	2	NG	NG	NG	NG	NG	1	NG	NG

Supplementary Table S2. Transformation efficiencies for *M. borealis*. Cultures of *MAT α* and *MAT α* were transformed with CaNAT1 by electroporation and lithium acetate methods alongside negative controls in triplicate, plated on YPAD with 75 mg L⁻¹ nourseothricin, and incubated for 48 hrs at 30°C for colonies to appear. No colonies grew on any of the control plates. Efficiency is given as colony forming units per μ g of DNA for 10⁸ cells. Av = average efficiency for each strain; Electro = electroporation; LiOAc = lithium acetate.

Strain	Method	# Cells	Vector DNA (μ g)	Plated	Colonies	Efficiency	Av.
<i>MATα</i>	Electro.	10 ⁸	1	2%	45	2250	1600
	Electro.	10 ⁸	1	2%	13	650	
	Electro.	10 ⁸	1	2%	38	1900	
<i>MATα</i>	Electro.	10 ⁸	1	2%	9	450	817
	Electro.	10 ⁸	1	2%	19	950	
	Electro.	10 ⁸	1	2%	21	1050	
<i>MATα</i>	LiOAc	10 ⁸	1	100%	13	13	9
	LiOAc	10 ⁸	1	100%	4	4	
	LiOAc	10 ⁸	1	100%	9	9	
<i>MATα</i>	LiOAc	10 ⁸	1	100%	5	5	4
	LiOAc	10 ⁸	1	100%	1	1	
	LiOAc	10 ⁸	1	100%	5	5	

Supplementary Table S3. Transformation results for additional yeast strains. An additional 19 yeast strains were transformed by electroporation using the PCR-linearized CaNAT1 gene (in standard code or yeast alternative nuclear code) flanked by 60-base-pair *ADH1* promoter and terminator sequences from *M. borealis*. Transformants were plated on YPAD with 75-200 mg L⁻¹ nourseothricin and incubated at 30°C for 2 days until colonies appeared. *M. bicuspidata* and *M. orientalis* were incubated at 27°C for 4 days until colonies appeared. NTC = nourseothricin; Alt. = CaNAT1 in yeast alternative nuclear code; Std. = CaNAT1 in standard code; Ctrl. = negative control.

Strain	NTC (mg L ⁻¹)	Plated	Number of Colonies		
			Alt.	Std.	Ctrl.
<i>Candida aff bentonensis</i>	75	40%	27	30	0
<i>Candida bromeliacearum</i>	100	10%	54	0	0
<i>Candida intermedia</i>	100	100%	15	0	0
<i>Candida saopaulonensis</i>	200	10%	42	0	0
<i>Candida tolerans</i>	100	10%	25	0	0
<i>Candida ubatubensis</i>	100	20%	41	0	0
<i>Clavispora lusitaniae</i>	75	40%	16	0	0
<i>Metschnikowia agaves</i>	100	10%	25	0	0
<i>Metschnikowia bicuspidata</i>	75	40%	4	0	0
<i>Metschnikowia caudate</i>	100	20%	9	0	0
<i>Metschnikowia drosophilae</i>	100	40%	4	0	0
<i>Metschnikowia gelsemii</i>	100	10%	93	0	0
<i>Metschnikowia gruessii</i>	100	10%	37	0	0
<i>Metschnikowia lunata</i>	100	10%	64	0	0
<i>Metschnikowia orientalis</i>	100	100%	13	0	0
<i>Metschnikowia pulcherrima</i>	100	10%	112	0	0
<i>Metschnikowia rancensis</i>	100	10%	27	0	0
<i>Metschnikowia reukaufii</i>	200	10%	121	0	0
<i>Saccharomyces cerevisiae</i>	100	10%	32	29	0

Primers used in this study:

Primers to amplify CaNAT1 (with <i>ADH1</i> promoter and terminator elements)	
BK420F	TCTTTCTTCACTATTCAAACATACATTGAATACAACCAAGCATCAATTAAGAAAA ATGTCTACTACTTTGGATGATACTG
BK420R	AGAAATGCAATGAACGATGAACTATTTATTGTGTATTGGGAGGGGGTCAAAGAG TTTATGGACATGGCATAGACATATAC
Primers to amplify CaNAT1 (with <i>HIS3</i> promoter and terminator elements)	
BK398F	ATTATGACTCTTCCCAATCTTTTCTCACTCATTACCAATACTAACAGATCAACCC CAAAATGTCTACTACTTTGGATGA
BK398R	CTTCGTATCTATGATTCTACACCGCATATAGTGGTCACTTAAATAATTCTATATG TCGCTTATGGACATGGCATAGACA
Primers to genotype attempted <i>HIS3</i> knock-outs – amplifying <i>HIS3</i>	
BK402F	CATGCTCTAGCCAAGCACTCGGGCT
BK402R	CTGATTGCCTCCTTTATGGCGATAG
Primers to genotype attempted <i>HIS3</i> knock-outs – amplifying CaNAT1	
BK364F	TGTTCCAGGTGATGCTGAAG
BK364R	CAACCACAAATGACCAGCAC
Primers to genotype attempted <i>HIS3</i> knock-outs – promoter junction	
BK402F	CATGCTCTAGCCAAGCACTCGGGCT
BK364R	CAACCACAAATGACCAGCAC
Primers to genotype attempted <i>HIS3</i> knock-outs – terminator junction	
BK364F	TGTTCCAGGTGATGCTGAAG
BK402R	CTGATTGCCTCCTTTATGGCGATAG
Primers to genotype <i>M. borealis</i> <i>MATα</i> clones – amplifying insertion sequences	
BK476F	GTGCTGGTCATTTGTGGTTG
BK478R	TCAATGGTGGATCAACTGGA
Primers to genotype <i>M. borealis</i> <i>MATα</i> clones – confirming insertion site in gDNA	
BK523F	AGAGCTGGGCCAATAAGGAG
BK523RA	TTGTCACATCAAGTTTCCTTGG
Multiplex genotyping primers for presence of MbShBle gene	
BK578_F	TTGCTGGTGTCTGTTGAGTTC
BK578_R	CTCAGCGTACAACCTCGTCCA
Multiplex genotyping primers for presence of CaNAT1 gene	
BK579_F	TCCAGTTGATCCACCATTGA
BK579_R	CAACCACAAATGACCAGCAC
Multiplex genotyping primers for presence of MbKanMX gene	
BK580_F	GACGTTACCGACGAGATGGT
BK580_R	TCACCGTGGGTAACAACAGA

Gene sequences:

Wild-type NAT:

ATGACCACTCTTGACGACACGGCTTACCGGTACCGCACCAGTGTCCCGGGGGACGCCGAGGCCA
TCGAGGCACTGGATGGGTCTTACCACCGACACCGTCTTCCGCGTCACCGCCACCGGGGACGG
CTTACCCTGCGGGAGGTGCCGGTGGACCCGCCCTGACCAAGGTGTTCCCGACGACGAATCG
GACGACGAATCGGACGCCGGGGAGGACGGCGACCCGGACTCCCGGACGTTTCGTCGCGTACGGG
GACGACGGCGACCTGGCGGGCTTCGTGGTCTGTCGTAACCGGCTGGAACCGCCGGCTGACCGT
CGAGGACATCGAGGTCCGCCCCGGAGCACCGGGGGCACGGGGTCCGGCGCGGCTTATGGGGCT
CGCGACGGAGTTCGCCCCGCGAGCGGGGGCGCCGGGCACCTCTGGCTGGAGGTACCAACGTCAAC
GCACCGGCGATCCACGCGTACCGGCGGATGGGGTTCACCTCTGCGGCCTGGACACCGCCCTGT
ACGACGGCACCGCCTCGGACGGCGAGCAGGCGCTCTACATGAGCATGCCCTGCCCTGA

CaNAT1:

ATGTCTACTACTTTGGATGATACTGCTTATAGATACAGAACTTCTGTTCCAGGTGATGCTGAAGCT
ATTGAAGCTTTGGATGGTCTTTCACTACCGATACTGTTTTAGAGTTACTGCTACTGGTATGGTT
TCACTTTGAGAGAAGTTCAGTTGATCCACCATTGACTAAGGTTTTCCAGATGATGAATCCGAT
GATGAATCCGATGCTGGTGAAGATGGTATCCAGATTCTAGAACTTTGTTGCTTATGGTGATGA
TGGTGATTTGGCTGGTTTTGTTGTTTCTTATTCTGGTTGGAACAGAAGATTGACTGTTGAAGAT
ATTGAAGTTGCTCCAGAACATAGAGGTCATGGTGTGGTAGAGCTTTGATGGGTTTGGCTACTGA
ATTGCCAGAGAAAGAGGTGCTGGTCATTTGTGGTTGGAAGTTACCAATGTTAATGCTCCAGCTA
TTCATGCTTATAGAAGAATGGGTTTCACTTTGTGTGGTTTGGATACTGCTTTATACGATGGTACTGC
TTCCGATGGTGAACAAGCTTTGTATATGTCTATGCCATGTCCATAA

CaNAT1 with CUG codons:

ATGTCTACTACTTTGGATGATACTGCTTATAGATACAGAACTTCTGTTCCAGGTGATGCTGAAGCT
ATTGAAGCTCTGGATGGTCTTTCACTACCGATACTGTTTTAGAGTTACTGCTACTGGTATGGTT
TCACTCTGAGAGAAGTTCAGTTGATCCACCATTGACTAAGGTTTTCCAGATGATGAATCCGAT
GATGAATCCGATGCTGGTGAAGATGGTATCCAGATTCTAGAACTTTGTTGCTTATGGTGATGA
TGGTGATCTGGCTGGTTTTGTTGTTTCTTATTCTGGTTGGAACAGAAGACTGACTGTTGAAGA
TATTGAAGTTGCTCCAGAACATAGAGGTCATGGTGTGGTAGAGCTTTGATGGGTTTGGCTACTG
AATTCGCCAGAGAAAGAGGTGCTGGTCATTTGTGGCTGGAAGTTACCAATGTTAATGCTCCAGCT
ATTCATGCTTATAGAAGAATGGGTTTCACTTTGTGTGGTCTGGATACTGCTCTGTACGATGGTACT
GCTTCCGATGGTGAACAAGCTTTGTATATGTCTATGCCATGTCCATAA

Wild-type Sh ble:

ATGGCCAAGTTGACCAGTGCCGTTCCGGTGTCCACCGCGCGACGTCGCCGGAGCGGTTCGAGTT
CTGGACCGACCGGCTCGGGTCTCCCGGACTTCGTGGAGGACGACTTCGCCGGTGTGGTCCGGG
ACGACGTGACCCTGTTCATCAGCGGGTCCAGGACCAGGTGGTGCCGGACAACACCTGGCCTG
GGTGTGGGTGCGCGGCCTGGACGAGCTGTACGCCGAGTGGTCCGGAGGTCGTGTCCACGAACCTC
CGGGACGCCTCCGGGCCGCCATGACCGAGATCGGGCAGCAGCCGTGGGGGCGGGAGTTCGCC
CTGCGCAGCCCGGCCGCAACTGCGTGCCTTCGTGGCCGAGGAGCAGGACTGA

MbShBle:

ATGGCTAAGTTGACCTCTGCTGTTCCAGTTTTGACCGCTAGAGACGTTGCTGGTGTCTGTTGAGTTC
TGGACCGACAGATTGGGTTTCTCTAGAGACTTCGTTGAGGACGACTTCGCTGGTGTGTTAGAGA
CGACGTTACCTTGTTCAATTTCTGCTGTTCAAGACCAAGTTGTTCCAGACAACACCTTGGCTGGGT
TTGGGTTAGAGGTTTGGACGAGTTGTACGCTGAGTGGTCTGAGGTTGTTTCTACCAACTTCAGAGA
CGCTTCTGGTCCAGCTATGACCGAGATTGGTGAGCAACCATGGGGTAGAGAGTTCGCTTTGAGAG
ACCCAGCTGGTAACTGTGTTCACTTCGTTGCTGAGGAGCAAGACTGA

MbShBle with CUG codons:

ATGGCTAAGTTGACCTCTGCTGTTCCAGTTTTGACCGCTAGAGACGTTGCTGGTGTCTGTTGAGTTC
TGGACCGACAGATTGGGTTTCTCTAGAGACTTCGTTGAGGACGACTTCGCTGGTGTGTTAGAGA
CGACGTTACCTTGTTCAATTTCTGCTGTTCAAGACCAAGTTGTTCCAGACAACACCTTGGCTGGGT
TTGGGTTAGAGGTTTGGACGAGCTGTACGCTGAGTGGTCTGAGGTTGTTTCTACCAACTTCAGAG
ACGCTTCTGGTCCAGCTATGACCGAGATTGGTGAGCAACCATGGGGTAGAGAGTTCGCTCTGAG
AGACCCAGCTGGTAACTGTGTTCACTTCGTTGCTGAGGAGCAAGACTGA

Wild-type KanMX:

ATGGGTAAGGAAAAGACTCACGTTTCGAGGCCGCGATTAATCCAACATGGATGCTGATTTATA
TGGGTATAAATGGGCTCGCGATAATGTCGGGCAATCAGGTGCGACAATCTATCGATTGTATGGGA
AGCCCGATGCGCCAGAGTTGTTTCTGAAACATGGCAAAGGTAGCGTTGCCAATGATGTTACAGAT
GAGATGGTCAGACTAACTGGCTGACGGAATTTATGCCTCTCCGACCATCAAGCATTTTATCCG
TACTCTGATGATGCATGGTACTCACCCTGCGATCCCCGGCAAAACAGCATTCCAGGTATTAG
AAGAATATCCTGATTGAGGTGAAAATATTGTTGATGCGCTGGCAGTGTTCTCGCGCCGGTTGCATT
CGATTCTGTTGTAATTGTCCTTTTAAACAGCGATCGCGTATTTCGTCTCGCTCAGGCGCAATCAC
GAATGAATAACGGTTTGGTTGATGCGAGTGATTTTATGACGAGCGTAATGGCTGGCCTGTTGAA
CAAGTCTGGAAAGAAATGCATAAGCTTTTCCATTCTCACCGGATTCAGTCGTCACTCATGGTGA
TTTCTCACTTGATAACCTTATTTTTGACGAGGGGAAATTAATAGGTTGTATTGATGTTGGACGAGT
CGGAATCGCAGACCGATAACCAGGATCTTGCCATCCTATGGAACCTGCCTCGGTGAGTTTTCTCCTTC
ATTACAGAAACGGCTTTTTCAAAAATATGGTATTGATAATCCTGATATGAATAAATTGCAGTTTC
ATTTGATGCTCGATGAGTTTTTCTAA

MbKanMX:

ATGGGTAAGGAGAAGACCCACGTTTCTAGACCAAGATTGAACTCTAACATGGACGCTGACTTGT
ACGGTTACAAGTGGGCTAGAGACAACGTTGGTCAATCTGGTGTCTACCATCTACAGATTGTACGGT
AAGCCAGACGCTCCAGAGTTGTTCTTGAAGCACGGTAAGGGTTCTGTTGCTAACGACGTTACCGA
CGAGATGGTTAGATTGAACTGGTTGACCGAGTTCATGCCATTGCCAACCATCAAGCACTTCATCA
GAACCCAGACGACGCTTGGTTGTTGACCACCGCTATCCCAGGTAAGACCGCTTTCCAAGTTTTG
GAGGAGTACCCAGACTCTGGTGAGAACATCGTTGACGCTTTGGCTGTTTTCTTGAGAAGATTGCA
CTCTATCCCAGTTTGTAACCTGTCCATTCAACTCTGACAGAGTTTTTCAAGATTGGCTCAAGCTCAATC
TAGAATGAACAACGGTTTGGTTGACGCTTCTGACTTCGACGACGAGAGAAACGGTTGGCCAGTTG
AGCAAGTTTGAAGGAGATGCACAAGTTGTTGCCATTCTCTCCAGACTCTGTTGTTACCCACGGT
GACTTCTCTTTGGACAACCTGATCTTCGACGAGGGTAAGTTGATCGGTTGTATCGACGTTGGTGA
GTTGGTATCGCTGACAGATACCAAGACTTGGCTATCTTGTGGAACCTGTTTGGGTGAGTTCTCTCCA

TCTTTGCAAAAGAGATTGTTCCAAAAGTACGGTATCGACAACCCAGACATGAACAAGTTGCAAT
TCCACTTGATGTTGGACGAGTTCTTCTAA

MbKanMX with CUG codons:

ATGGGTAAGGAGAAGACCCACGTTTCTAGACCAAGATTGAACTCTAACATGGACGCTGACTTGT
ACGGTTACAAGTGGGCTAGAGACAACGTTGGTCAATCTGGTGCTACCATCTACAGATTGTACGGT
AAGCCAGACGCTCCAGAGTTGTTCTGAAGCACGGTAAGGGTCTGTTGCTAACGACGTTACCGA
CGAGATGGTTAGATTGAACTGGCTGACCGAGTTCATGCCATTGCCAACCATCAAGCACTTCATCA
GAACCCAGACGACGCTTGGTTGTTGACCACCGCTATCCCAGGTAAGACCGCTTCCAAAGTTTTG
GAGGAGTACCCAGACTCTGGTGAGAACATCGTTGACGCTCTGGCTGTTTTCTGAGAAGATTGCA
CTCTATCCCAGTTTGTAACTGTCCATTCAACTCTGACAGAGTTTTTCAGATTGGCTCAAGCTCAATC
TAGAATGAACAACGGTTTTGGTTGACGCTTCTGACTTCGACGACGAGAGAAACGGTTGGCCAGTTG
AGCAAGTTTTGGAAGGAGATGCACAAGTTGTTGCCATTCTCTCCAGACTCTGTTGTTACCCACGGT
GACTTCTCTTTGGACAACCTTGATCTTCGACGAGGGTAAGTTGATCGGTTGTATCGACGTTGGTAGA
GTTGGTATCGCTGACAGATACCAAGACTTGGCTATCTTGTGAACTGTTGGGTGAGTTCTCTCCA
TCTTTGCAAAAGAGATTGTTCCAAAAGTACGGTATCGACAACCCAGACATGAACAAGTTGCAAT
TCCACTTGATGTTGGACGAGTTCTTCTAA

***M. borealis* ADH1 promoter:**

TTGTCATTGTCAGCAAAGTACATTGATCTGTATTCTTCCAAACCCAAGACATTCTTGACGAAAGCT
TGGATTCTTATAAACTAGCACTTCATGCTTCCACTGCATGCTTCTTACCAGAATCAATGAAAGT
GGAGAGATCCAAGTAATTGAGCACCTCAAAAATGATGAGACGGTGAAATCCATAGGATTCTTA
CTCCGTGCCATTCAACACCATGGCATAAGTATGTTGCATCGTCCTGAAATCGTTGACTCATCATGGT
TTTTGACATGTGAACCACCATTCGATTTGACCTTTGGTAATATCAACAATGTCAAAGAGTACCGA
GACGAACTGGATCGCTTCTTCGACAATCCCGTGGAGTTGTACGAAACTTGCCGTATGAGTGGCC
CTCACATCTTGTGTGTTGGAACCAATGGAGTACTTAGTGACACAGGAATTGCCTCAGTACCATG
AATGCCACAGGTATTTCAATAGCTACTTTCATTGGGACTCACGTCGTCAAGGAGATTTGATTGTGT
TCTGCAAAACAATCTGAGGTGTTGCTTCGTAATAAATTTACGTTGGAGAAGCGACTTATCTAAA
CGCATAAATGTGTGCGCATTGAAACACGCATCAAAATGCGCTCGTCGGCTAATGTCGGAAAGGC
CGCTCTCGCTCTTCTAAATCTTGTAACTATCGGGAAATAACTGATATCAAATCATGCCACCCGAC
AATTGCAGCAGATCTGAGACCTGCATAATTATGAGTCAAGAAATATCATAAAATGCGTGCATTGT
ACTTAACTTTAAAGTCTACTCTTTCCTAAAACCTTAGCATCCTCCTCTGCATGAGTATGCGCTT
AAGTGTGCAACAAAGCCAGAAATCACACCACGCACATAGAAGCAGCAAACATTTCGTGACTAT
AAATATGATGCTTCGCCGACTCCAGCAATTCTTCTTCTTCACTATTCAAACATACATTGAATAC
AACCAAGCATCAATTAAGAAAA

***M. borealis* ADH1 Terminator:**

ACTCTTTGACCCCCTCCCAATACACAATAAATAGTTCATCGTTCATTGCATTTCTTCTCCAACCTCTG
CTATGCGTTCCTCATCACGTTCTGCTGGATCTGGGTGATCAATTGCTCATCATTCAACACGTCAA
AAAGCTTATTTTCTCTGCCAACGCATCGCTGATGTTGCTAGTTTCGAATTCAACCTTTTCATTT
CATATCCTTCTCTGCCAAGGCCCTTTGTCAACTCGTTCACCCGGAGGTTGGCATTCTCCAACCTTCAT
GGTTTGTGATTGAGCCGGTAATAAGCGATTCCGTTTCTTCATGCAAATGCGAATTTTCTTTAAT
AAGCCGTTTCGTTATCAGCATGGCCTGCTGATGTTTTTACATATGCTAATTGCTCAAAGCCTTCAC
GAAGTCTGCGTCTCGTCTCTATCTGGTCAGTTTGTTCAACTGGTCTAGTATTGCATTCCACGGC
TCATTCATTTTGGTTAAAAATGTGAGTCTCCAAGCCCCAATATGGTAATTTGGATGTGTGTTTCAA

GTTTCCTATGTGTCAAATCAGGCTCAGAATGACACATTGCAAGTATCTGGGGTCTTAAATTGCCA
GTGCACAGTGCCACGGTTGGCGCCTTGATAAGCTTACGACAGAGCATATAACCTCAAACCGAA
ATACACCATCCTAGAATATACTGTCCTATAATATTGACCATTTGAAGAGTCAGTTTTGCTTCTTCA
CTGGTCTAAGAATTGCATTGCACTGCTGCACTAATTGTCCTTAACTCACTTAATTGCCTGTGTATTG
ACATCTGCAAACATCTTCCCCTGTGATTAGCAAAGAAGCAGGTGAACCGATCGTCTCCTTAGAA
TGGCTGAGCCTTCTTCTTCATGGTGGTCTCTGTTATATCGCTCCGAATCTACCAAACAAGCTGATA
TTTCTTCAAAGGCCACTTCCCCTGGGACACTACCTCCTGCCAACTCTAAAAAGGAAACGAATACT
CAAAAGGCCTC

***M. borealis* HIS3 promoter:**

TGTCTAGCTCTAACACCTTGATGGCCACAACCTTTTCAGTTTTCTTGTGATACCCTTTGTAGACGAC
ACCGAACTTGCCTTGCCGATGACCTTTGTTCTTTGGTAAGAAGAAGTACTTAGCATGGTGAAAG
TGGAGGAGTGTCTCAAGGCCAAGGACCTAGGGGAATTTGAGATCGAGTACTTTCTTTCATGG
AAAGAGAGTTCAGAAGAACGCCTTTGGAACTTCTGCCAATGTTTATAAGCGCTTGAAAATAAGT
ATGGCGTACAGCGATATCCTGCAGAGTTGATCCGATGGATTATGGTGAAGGCCAGCACGTTGGG
CGTCCAAAACAGAGATCGGGTCGTGAAGGAAAGTGTACCAGTATGAAAATTGAAGGCGCGGTA
GCCAGGATCTGCAGCTGAATACGGTGGGTCGCGCCTGAGAAACAAGATGCACCGAATGCGGAG
GAAGCTTCAAGTAGAGAGCAGAAAGCAAAGATGCTAATTTACGACGAAGGGGTTTTCGAAGGGT
TCGGAGCGTTTCGATGCGTTTTGCCAGCCAAATTTGCCAACTAATTCGAAAGCCACCATTGTTAAT
CACCAAAGGTCAGCTGCCTGAATGGTCAGGTGTACATGTGCAGTCAGTAGACAATAGTGGTGG
ACTTTTGGATTTTCAGGCAAAGAAGTGCGAAGTAAGCTATCAAATGACCAGTGTGCCAACACAC
CATGCAAGAAATTCGGCCACAAGCAAAAAGAGTTGAATTTTCGCCGGCGATGCAAAATAAAAAA
AAAAAAGACCCAAAAAACTTAGCCTTCTGTTCCAAAATCAGAAATTGAAAGGTCAGTCT
TTTGTGCTCGAATCGGGTACTTCCGAGTTTGTGCTGCACTGGTCCGGCCATTTTGGTGTGGATG
GAACTTTGGTGAACGGAATTTGCCAGAGATATGCGCATTATGACTCTTCCCAATCTTTTCTCACT
CATTACCAATACTAACAGATCAACCCCAA

***M. borealis* HIS3 terminator:**

GCGACATATAGAATTATTTAAGTGACCACTATATGCGGTGTAGGAATCATAGATACGAAGCGAA
AAGTCAGAGGTGCGCATTACAACCTTCGGGGCACCGCTTTCGATGCGCATCTTCTACTACTACC
ATGAGAAGGTTCCACACTAGTGGTATCCGCCAAGTCATCAAGCCAGTGTCAAACCTGCATGATCT
GAAGAAAGGACTCAAGAAATTTGAAGATTCCTTCAATGCAGGTAGTAACCGGAAGCTGGAGCA
GAAAATATGGGACAAGTTGAATATCTCCAAGCACGAGTTTTTCATACGGAAATATGGCAACATTT
CGCCCGAAAAACGAAAACAGTTGGATGACAAAGTCAACCCGACAAAGGTCGCTCCGCGAGCAGA
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