

1 Article

# 2 NiFe alloy reduced on graphene oxide for 3 electrochemical non-enzymatic glucose sensing

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13

14 **Abstract:** NiFe alloy nanoparticles/graphene oxide hybrid (NiFe/GO) was prepared for  
15 electrochemical glucose sensing. The as-prepared NiFe/GO hybrid was characterized by  
16 transmission electron microscopy (TEM) and X-ray diffraction (XRD). The results indicated that  
17 NiFe alloy nanoparticles can be successfully deposited on GO. The electrochemical glucose sensing  
18 performance of the as-prepared NiFe/GO was studied by cyclic voltammetry and amperometric  
19 measurement. Results showed that NiFe/GO modified glassy carbon electrode had sensitivity of  
20 173  $\mu\text{A mM}^{-1}\text{cm}^{-2}$  for glucose sensing with a linear range up to 5 mM, which was superior to  
21 commonly used Ni nanoparticles. Furthermore, high selectivity for glucose detection can be  
22 achieved by NiFe/GO. All the results demonstrated that NiFe/GO hybrid was promising for using  
23 in electrochemical glucose sensing.

24 **Keywords:** NiFe alloy; graphene oxide; glucose; non-enzymatic sensor

25

## 26 1. Introduction

27 Glucose sensing is important in many fields, such as medical diagnostics and food industry [1].  
28 The earliest glucose sensing is reported by Clark and Lyons in 1962 using biological enzyme [2].  
29 Since then, biological enzymes such as glucose oxidase and glucose dehydrogenase have been  
30 widely used for glucose sensing. However, the biological enzymes are susceptible to external  
31 condition such as temperature, humidity, pH, and so on, which lead to unstability of biological  
32 enzymes [3–5]. The expensive price and complicated mobilizing methods of biological enzymes also  
33 restrict its application [6]. Therefore, to address these problems, the research focus has been  
34 transferred to develop electrochemical non-enzymatic glucose sensors.

35 Due to its high stability and sensitivity, non-enzymatic electrochemical technology is a good  
36 choice for glucose sensing [7]. Various nanoparticles have been reported for constructing  
37 electrochemical non-enzymatic biosensors, such as noble metal Pt nanoparticles (Pt NPs) [3,8,9], Au  
38 NPs [10,11], Pd NPs [12,13], etc. However, the expensive price of these noble metals restricts their  
39 practical application. Furthermore, noble metals constructed electrochemical glucose sensors are  
40 usually poisoned by chloride ion containing widely in body's blood [14]. In order to exploit cheap  
41 electrochemical non-enzymatic glucose biosensors, considerable attention has been focusing on  
42 non-noble metal materials, such as Ni NPs [15–17],  $\text{Co(OH)}_2$  nanotubes [18], Cu NPs [19,20], etc.  
43 Recent studies reveal that bimetallic materials [21], especially bimetal alloys, exhibit better catalytic  
44 performance than that of monometallic counterparts [22]. Xu et al. and Chen et al. both found that  
45 Pt-Ni alloy exhibited enhanced sensitivity for glucose sensing [23,24]. Besides, PtRu alloy [25], PtAu  
46 alloy [26], PtCo alloy [27], CoCu alloy [28], and NiCo alloy [29] have been used in glucose sensing.

47 A support material is also important in electrochemical glucose sensor [30]. Carbon based  
48 materials (especially carbon nanotube and graphene) have extensively used to load the  
49 electrochemical catalyst due to its excellent conductivity, such as loading cupric oxide [31] or  
50 growing copper nanoparticles [32] on carbon nanotubes (CNTs) for electrochemical glucose sensing.  
51 Graphene and its oxide (graphene oxide, GO) have triggered researchers' interest since 2004 because  
52 of their good electrical conductivity and large specific surface area. Due to these exceptional  
53 properties, Graphene and GO have been chosen as supporting material in electrochemistry [33]. GO  
54 loaded with NiS [34], NiCo alloy [29] and CuS NPs [35] have been applied for electrochemical  
55 glucose sensing.

56 Based on the above considerations, we tried to study the alloying of commonly used Ni NPs in  
57 the scope of glucose detection with Fe element on GO, because NiFe alloy shows high  
58 electrochemical performance in various processes [36–39]. To the best of our knowledge, the NiFe  
59 alloy nanoparticles/graphene oxide hybrid (NiFe/GO) has never been reported for electrochemical  
60 glucose sensing. The NiFe/GO reveals better electrocatalytic performance towards glucose oxidation  
61 than the commonly used Ni NPs. High performance of NiFe alloy nanoparticles and superior  
62 conductivity of GO endow the NiFe/GO biosensor to electrochemically sensing glucose in wide  
63 concentration range.

## 64 2. Materials and Methods

### 65 2.1. Chemicals and reagents

66 All reagents used in experiments were analytical grade. Graphite powder was purchased from  
67 Alfa Aesar. Nickel sulfate hexahydrate, iron sulfate heptahydrate, hydrazine hydrate (80 wt%) and  
68 glucose are procured from Sinopharm Chemical Reagent Co., Ltd. Ascorbic acid (AA), uric acid  
69 (UA), and dopamine (DA) were purchased from ACROS. Deionized water (18.2 MΩ·cm) was used  
70 in all experiments.

### 71 2.2. Preparation of GO

72 GO was prepared by modified Hummers method [40]. 2 g Graphite and 1 g NaNO<sub>3</sub> were mixed  
73 with 50 mL H<sub>2</sub>SO<sub>4</sub> (95%) in a 250 mL flask within an ice bath to keep a low temperature (Caution: ice  
74 bath is important and necessary). Then, 6 g KMnO<sub>4</sub> was added slowly into the above suspension  
75 with vigorous stirring. In this adding process, the reaction temperature should be kept lower than 20  
76 °C. After that, the mixture was stirred at room temperature overnight. At the end, 60 mL H<sub>2</sub>O was  
77 added slowly with vigorous agitation. The reaction temperature was increased rapidly up to 95 °C,  
78 and the color of suspension changed to yellow. Then, 10 mL of 30% H<sub>2</sub>O<sub>2</sub> was added to the mixture.  
79 Finally, the obtained product was washed by rinsing with 5% HCl and then deionized water for  
80 several times until the pH of filtrate reached 7. After dry in vacuum dryer, gray powder GO was  
81 obtained.

### 82 2.3. Preparation of NiFe/GO composites

83 For the preparation of NiFe/GO composite (for example, the ratio of Ni to Fe is 1:1, denoted as  
84 NiFe/GO), 0.1 g of the as-prepared GO, 278.1 mg FeSO<sub>4</sub>·7H<sub>2</sub>O (1 mmol) and 262.9 mg NiSO<sub>4</sub>·6H<sub>2</sub>O (1  
85 mmol) were added to 10 mL deionized water. The mixture was sonicated for 30 min to get an evenly  
86 dispersed solution. Then, 15 mL hydrazine hydrate was dropped slowly into the above solution  
87 followed by refluxing at 100 °C for 3 h under N<sub>2</sub> atmosphere. After reaction, the obtained NiFe/GO  
88 composite was washed using deionized water for several times. Finally, the washed NiFe/GO  
89 composite was dried in vacuum for further use. Other composites (different Ni to Fe mass ratios,  
90 such as Ni<sub>1</sub>Fe<sub>4</sub>/GO, Ni<sub>4</sub>Fe<sub>1</sub>/GO, Ni/GO and Fe/GO) were prepared by adjusting the ratio of Ni and Fe.  
91 NiFe alloy NPs were also prepared for comparison using the same method without adding GO.

92

#### 93 2.4. Preparation of the NiFe/GO hybrid modified electrode

94 5 mg NiFe/GO hybrid was dispersed in 1 mL solution that containing 0.5 mL ethyl alcohol and  
95 0.5 mL deionized water. The above solution was sonicated for 30 minutes to get an evenly dispersed  
96 mixture. After sonicating, a certain amount of the mixture was dropped onto bare glassy carbon  
97 electrode (GCE) followed by adding 2  $\mu$ L Nafion solution (0.5%) to entrap the NiFe/GO. The  
98 prepared electrode was denoted as NiFe/GO/GCE. For comparing, Ni/GO/GCE, Fe/GO/GCE and  
99 NiFe/GCE were fabricated similarly.

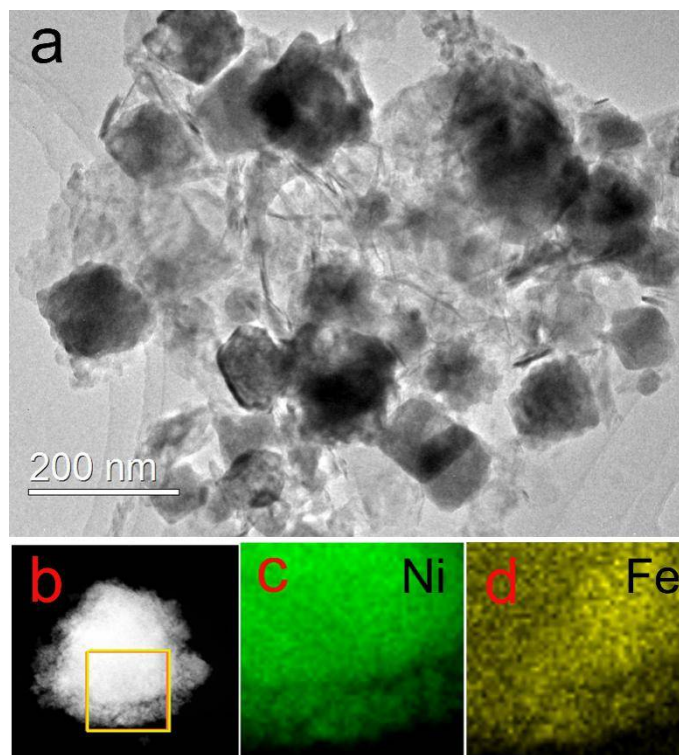
#### 100 2.5. Apparatus and measurements

101 The morphologies of NiFe/GO composite were collected with a Tecnai G<sup>2</sup>F<sup>30</sup> electron  
102 microscope. XRD data were conducted on a Rigaku D/max-2400 diffractometer operating at a  
103 voltage of 40 kV and a current of 40 mA using Cu-K radiation as the X-ray source. Electrochemical  
104 characterization was performed on a CHI 660C electrochemical workstation with the modified GCE  
105 as working electrode, Pt wire as counter electrode and Ag/AgCl (3 M KCl) as reference electrode. All  
106 voltages used in the manuscript were versus to Ag/AgCl (3 M KCl) electrode.

### 107 3. Results and discussion

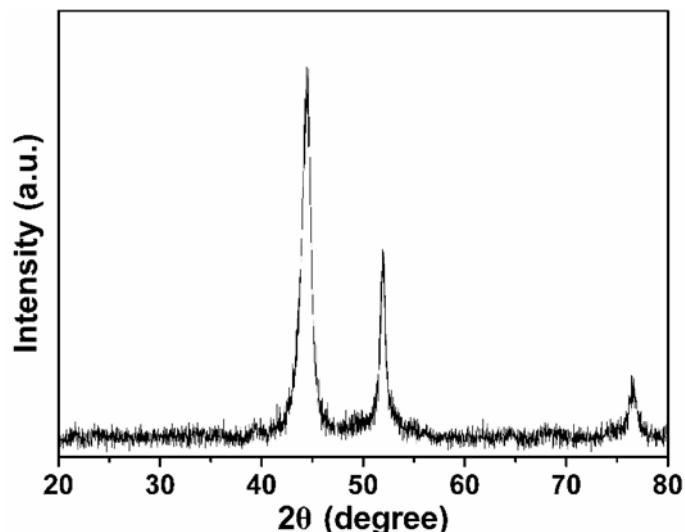
#### 108 3.1. Structural characterization

109 The TEM image of the NiFe/GO is displayed in Figure 1. As shown in Figure 1a that NiFe alloys  
110 with diameter of about 100 nm are loaded on GO. Figure 1b-d show the element mapping of NiFe  
111 alloy. It can be seen clearly that Ni and Fe element are evenly distributed in NiFe alloy. This prove  
112 the successful preparation of NiFe alloy. Figure 3 shows the XRD pattern of NiFe/GO to further  
113 investigate the crystalline structure. The diffraction patterns located 44.5°, 51.9° and 76.4° can be  
114 ascribed to the diffraction of (111), (200), and (220) crystal planes of the NiFe alloy, respectively [41].  
115 No peaks of GO can be found due to its small amount.



116

117 Figure 1. (a) TEM image of the NiFe/GO and (b, c d) element mapping of NiFe alloy.

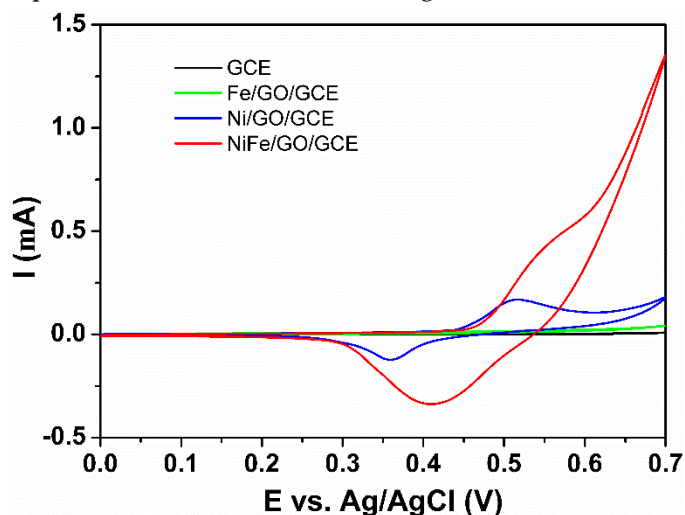


118

119 Figure 2. XRD pattern of NiFe/GO.

## 120 3.2. Electrochemical measurements

121 Figure 3 shows the typical cyclic voltammograms of the as-prepared GCE, Fe/GO/GCE,  
 122 Ni/GO/GCE and NiFe/GO/GCE in 0.1 M NaOH solution in the presence of 1.0 mM glucose. It can be  
 123 seen from Figure 3 that GCE and Fe/GO/GCE show very small current in the whole voltage range.  
 124 Oxidation peaks between 0.50 to 0.55 V can be observed for Ni/GO/GCE and NiFe/GO/GCE, which  
 125 can be ascribed to the oxidation of  $\text{Ni}^{2+}$  to  $\text{Ni}^{3+}$ . The  $\text{Ni}^{3+}$  accounts for the oxidation of glucose [15].  
 126 When Fe was added to Ni to form NiFe alloy, the current increases obviously. This indicates that  
 127 NiFe alloy have higher performance for electrochemical glucose detection.

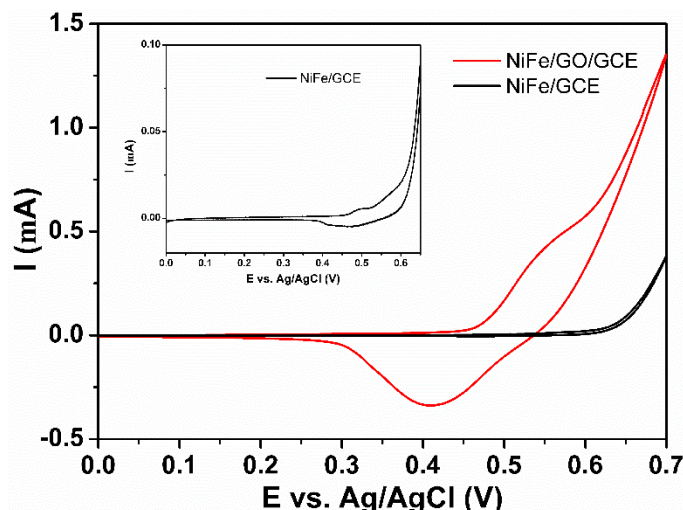


128

129 Figure 3. CVs of GCE, Fe/GO/GCE, Ni/GO/GCE and NiFe/GO/GCE in 0.1 M NaOH solution in the  
 130 presence of 1.0 mM glucose at a scan rate of 10 mV/s.

131 To illustrate the effect of GO, the performance of NiFe/GCE and NiFe/GO/GCE were  
 132 investigated. As shown in Figure 4, NiFe/GCE has only negligible current and the redox peaks can  
 133 be seen in the inset figure. NiFe/GO/GCE shows much larger current than that of NiFe/GCE,  
 134 indicating GO is important in the composite electrode. The GO not only acts as support to load NiFe  
 135 alloy from aggregation, but also plays an important role for electronic transmission.

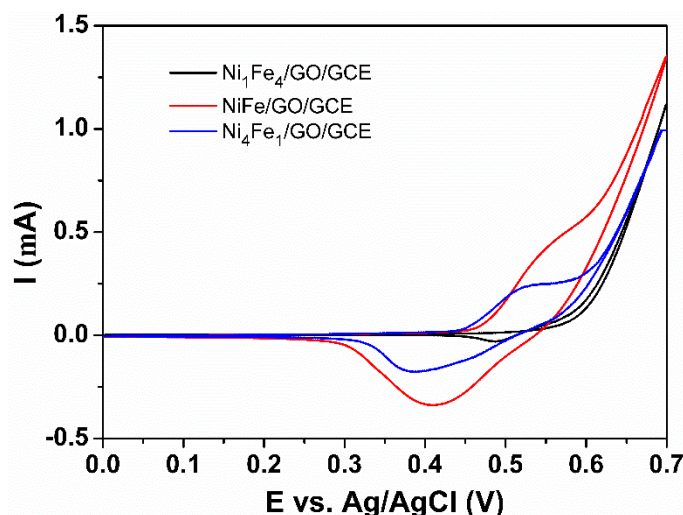




136

137 Figure 4. CVs of NiFe/GCE and NiFe/GO/GCE in 0.1 M NaOH solution in the presence of 1.0 mM  
 138 glucose at a scan rate of 10 mV/s. Inset: CV of NiFe/GCE.

139 Furthermore, the effect of the ratio between Ni and Fe elements were investigated. As shown  
 140 in Figure 5, Ni<sub>1</sub>Fe<sub>4</sub>/GO/GCE shows the smallest current. With the increase of Ni amount,  
 141 NiFe/GO/GCE shows the biggest current. More Ni amount, instead, decrease the current as  
 142 Ni<sub>4</sub>Fe<sub>1</sub>/GO/GCE shows. So, NiFe/GO/GCE was used for further research.



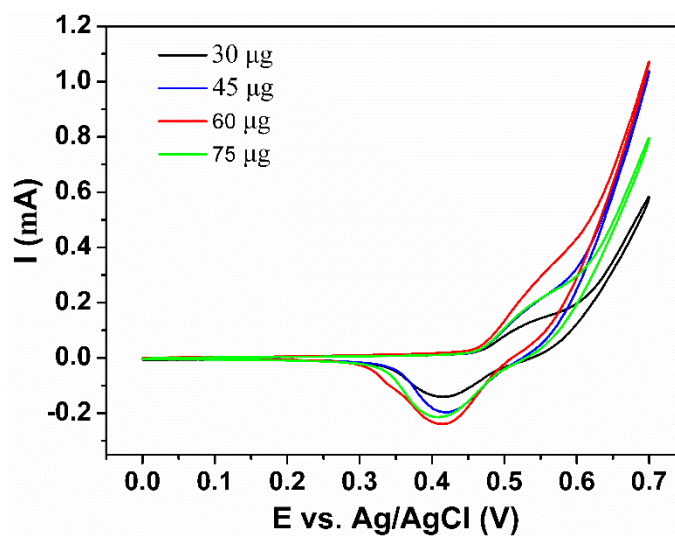
143

144 Figure 5. CVs of Ni<sub>1</sub>Fe<sub>4</sub>/GO/GCE, NiFe/GO/GCE and Ni<sub>4</sub>Fe<sub>1</sub>/GO/GCE in 0.1 M NaOH solution in the  
 145 presence of 1.0 mM glucose at a scan rate of 10 mV/s.

146 In order to improve the electrocatalytic performance of NiFe/GO/GCE, the loading amount of  
 147 NiFe/GO were studied. Figure 6 shows CV curves of NiFe/GO/GCE with different amount of  
 148 NiFe/GO loading on GCE. It can be seen that the current increases gradually from the loading  
 149 amount of 30 to 60  $\mu\text{g}$  NiFe/GO. When the loading amount exceeding 60  $\mu\text{g}$ , the current decreases.  
 150 So, 60  $\mu\text{g}$  is the optima loading amount and used in the later experiments. This phenomenon can be  
 151 explained by the change of catalytic sites. At first, increasing loading amount of the NiFe alloy  
 152 increased catalytic sites. However, overmuch loading amount limited mass transfer process,  
 153 leading to the decrease of current [19].

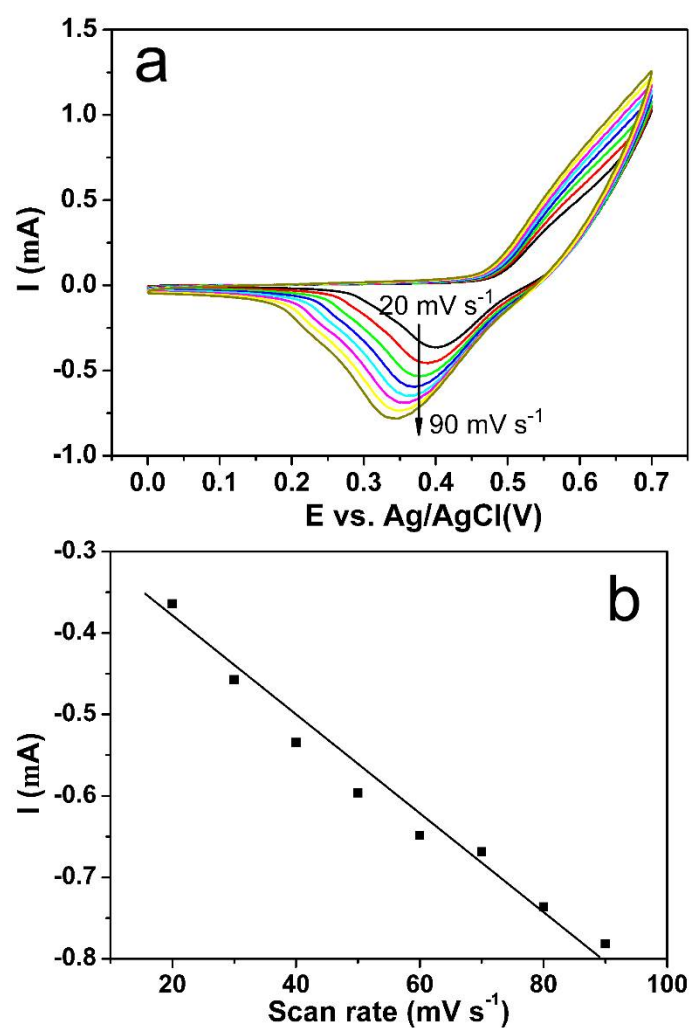
154 Cyclic voltammetric measurements were performed at increased scan rates to better  
 155 understand the electrocatalytic properties of the NiFe/GO/GCE for glucose oxidation. As  
 156 demonstrated in Figure 7a, cathodic peak currents of NiFe/GO/GCE increased with the increasing  
 157 scan rate. Figure 7b give the relationship between cathodic peak current and scan rate. It can be  
 158 seen that cathodic peak currents increases linearly with the used scan rates. This result manifested

159 the rate determining step was surface reaction control rather than diffusion control, which is  
160 favorable for quantitative analysis.



161

162 Figure 6. The effect of modificaion amount on the performance of electrode.

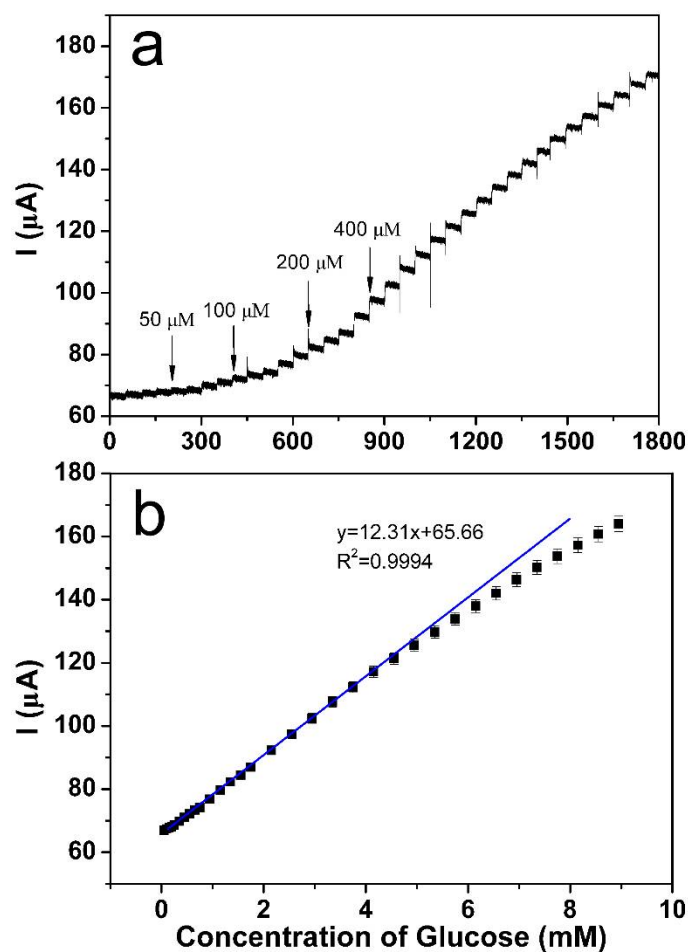


163

164 Figure 7. (a) Cyclic voltammograms of NiFe/GO/GCE at different scan rates from 20 to 90 mV/s in 0.1  
165 M NaOH with 1.0 mM glucose. (b) Plot of cathode current vs the scan rate.

### 166 3.3. Amperometric response towards glucose sensing

167 Amperometric measurement was carried out to show the current response of the NiFe/GO/GCE  
168 towards continuously adding of various concentrations of glucose, which was carried out in  
169 vigorous stirred electrolyte. As shown in figure 8a, the NiFe/GO/GCE generates fast current  
170 response upon the addition of glucose from 50 to 400  $\mu\text{M}$  everytime. According to the amperometric  
171 current, the corresponding calibration curve was plotted in Figure 8b. It can be seen that the  
172 NiFe/GO/GCE sensor showed a wide linear sensing range form 0 to 5 mM ( $R^2 = 0.9994$ ), and a  
173 sensitivity of  $173 \mu\text{A mM}^{-1}\text{cm}^{-2}$ . The high sensitivity can be ascribed to the high electrocatalytic  
174 activity of NiFe alloy and excellent electronic transmission performance of GO.  
175

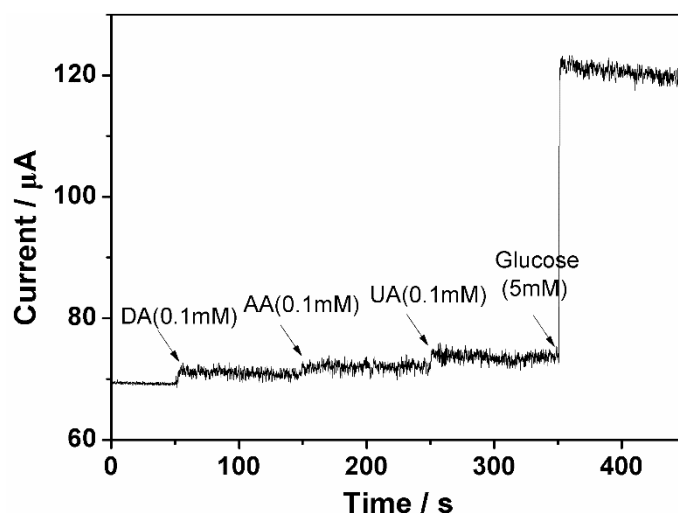


176

177 Figure 8. (a) Amperometric currents measured with continuous addition of glucose from 50 to 400  
178  $\mu\text{M}$  at 0.55 V for NiFe/GO/GCE. (b) The corresponding calibration curve.

### 179 3.4. Specificity and reproducibility of the NiFe/GO/GCE

180 DA, AA and UA are co-exist in human blood, which will influence the electrochemical  
181 non-enzymatic glucose sensing. To investigate the effect of DA, AA and UA on the glucose sensing,  
182 amperometric response test was carried out. Figure 9 shows the amperometric response current with  
183 addition of DA, AA, UA and glucose. It can be seen that the current generated by DA, AA and UA in  
184 normal physiological concentration are only 4.1%, 1.9% and 2.6% compared to that of glucose,  
185 respectively. This research demonstrates that the NiFe/GO/GCE possesses specificity for glucose  
186 sensing and can be used in reality sensing.



187

188 Figure 9. Amperometric response of NiFe/GO/GCE to interferent at physiological concentration.

189 To check the reproducibility of our manufacturing operation, we fabricate two electrodes in  
 190 same condition and test their double layer capacitance (Cdl). Cyclic voltammograms were carried  
 191 out in 0.1 M NaOH solution at scan rate from 10 to 100 mV s<sup>-1</sup> (Figure 10a and b). Then the  
 192 electrochemical active surface area was estimated by testing capacitive current at non-faraday region  
 193 at different scan rates, from which double-layer charging (Cdl) was determined by plotting the  $\Delta J =$   
 194  $(J_a - J_c)$  at 0.10 V vs. Ag/AgCl against the scan rate as shown in Figure 10c and d. The linear slope is  
 195 equivalent to twice of the Cdl, which can be used to represent the electrochemical active surface area  
 196 [42]. From Figure 10c and d, it can be seen that the two fabricated electrodes have almost the same  
 197 linear slope, revealing the same electrochemical active surface area of the two electrodes fabricated  
 198 at same condition. That is, our fabrication process is reliable.

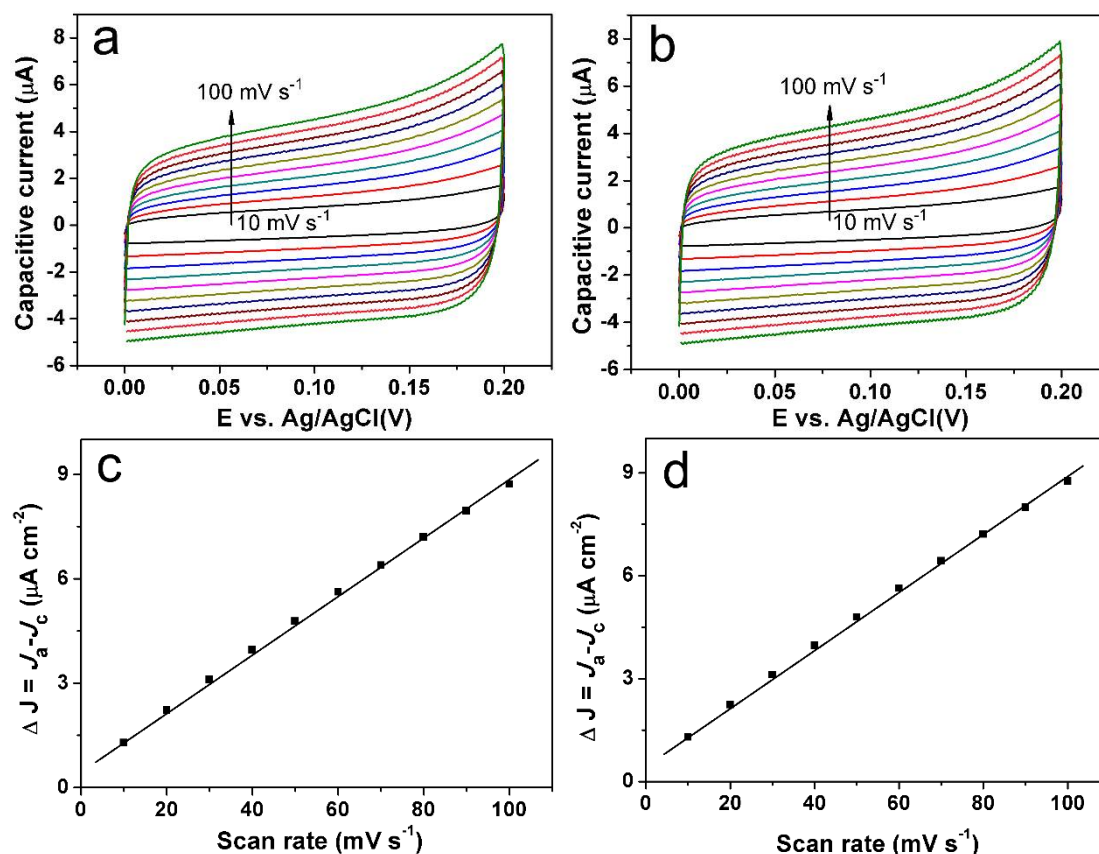
### 199 3.5. Practical applications

200 For practical analysis, the NiFe/GO/GCE was used to detect glucose concentration in human  
 201 serum by amperometric measurement. Briefly, 20  $\mu$ L serum was injected into 15 mL of 0.1 M NaOH  
 202 solution, then the current response at +0.55 V on NiFe/GO/GCE was recorded. The recovery value  
 203 was confirmed by standard injection of glucose with known concentration to the above sample,  
 204 then record current at +0.55 V. As listed in Table 1, the recovery values of all three samples are close  
 205 to 100%, indicating good practical application prospect of NiFe/GO/GCE electrode.

206 Table 1 Amperometric detection of glucose in serum

Sample	Concentration (mmol L <sup>-1</sup> )	RSD (%)	Added (mmol L <sup>-1</sup> )	Found (mmol L <sup>-1</sup> )	Recovery (%)
1	9.3	3.5	1	10.1	98
2	5.8	3.1	1	6.7	99
3	7.1	3.4	1	8.0	99





207

208 Figure 10. Cyclic voltammograms with different scan rates in potential range of 0.0 V to 0.2 V vs.  
 209 Ag/AgCl where no Faradaic processes occur for first (A) and second fabrication (B) of electrode.  
 210 Charging current density differences first (C) and second fabrication (D) of electrode plotted against  
 211 scan rates.

#### 212 4. Conclusions

213 In summary, a NiFe/GO/GCE electrochemical glucose sensor have successfully fabricated. The  
 214 addition of Fe element into Ni nanoparticle to form NiFe alloy nanoparticle improved the  
 215 electrochemical performance of glucose sensor. In addition, the GO not only acts as support to load  
 216 NiFe alloy from aggregation, but also play an important role for electronic transmission. The  
 217 NiFe/GO/GCE electrode exhibited excellent sensitivity (173 μA mM<sup>-1</sup>cm<sup>-2</sup>) and wide detection linear  
 218 range (up to 5 mM). In addition, NiFe/GO/GCE electrode shows high selectivity for glucose  
 219 detection and can be applied to glucose detection in human serum. All results demonstrate that  
 220 NiFe/GO/GCE electrode is a promising candidate in the development of cheap, stable and sensitive  
 221 non-enzymatic glucose sensors.

222

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