Neurologic and inflammatory manifestations in Sjögren’s syndrome: the role of tryptophan/kynurenine pathway

Fabíola Reis de Oliveira¹, Marina Zilio Fantucci¹, Leidiane Adriano¹,
Valéria Valim², Thiago Mattar Cunha¹, Paulo Louzada Junior¹,
Eduardo Melani Rocha¹

¹Ribeirao Preto Medical School, Ribeirao Preto, University of Sao Paulo, Ribeirao Preto, SP, Brazil.
²Espírito Santo Federal University, Vitoria-ES, Brazil.

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Corresponding Author:
Eduardo Melani Rocha
Department of Ophthalmology, Otorhinolaryngology and Head & Neck Surgery, RibeirãoPreto Medical School, University of São Paulo.
Av. Bandeirantes, 3900, 14049-900 – Ribeirão Preto SP, Brazil.
emrocha@fmrp.usp.br
Phone/fax: 55-16-3602-0593

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Abstract

For decades, neurologic, psychological and cognitive alterations, and other extra glandular manifestations have been described and are being considered part of the Sjögren’s syndrome (SS). The lacrimal glands (LG), the ocular surface (OS), salivary glands (SG) and the central nervous system (CNS) are integrated to modulate the autonomic functions and the hippocampus, which is linked to the autonomic nervous system, modulate behavior responses compromised in the SS.

Recent studies confirm that the tryptophan/kynurenine pathway (TKP) can be stimulated by interferon-γ (IFN-γ) and other cytokines, activating the indoleamine-pyrrole 2,3-dioxygenase (IDO) in SS. This pathway interferes on serotonergic and glutamatergic neurotransmission, mostly in the hippocampus, and other structures of the CNS. Therefore, it is plausible that TKP induces neurological manifestations, and contributes to the discrepancy between symptoms and signs, including manifestations of hyperalgesia and depression in patients with SS, for example. Observations from clinical studies in AIDS, graft versus host disease, lupus and SS, but also from experimental studies support this hypothesis. Therapeutic strategies are reexamined and new options designed and tested to regulate the TKP. In the future, the confirmation and application of this concept may offer a clue to the mosaic of manifestations of SS.

Key words: IDO, kynurenine, pain, Sjögren’s syndrome, tryptophan
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1. Introduction

Sjögren’s syndrome (SS) is defined as an exocrinopathy of salivary and lacrimal glands (SG and LG) mediated by autoimmune mechanisms that can manifest neurologic dysfunctions, and those neurologic dysfunctions may take part in the physiopathology of the disease [1-5]. However, the neurological manifestations are not considered in the definition or the diagnosis, despite those manifestations are present in the disease progress evaluation and are being reported in association with SS with more attention in recent years [6-9]. Of interest, 60-80% of patients develop neurological manifestations before or at SS diagnosis (early systemic presentation) indicating that neurological inflammatory damage is precocious and could have a role in disease mechanism [10].

The tryptophan/kynurenine signaling pathway (TKP) intermediates the serotonergic and glutamatergic neurotransmission. It is also known to take part in the inflammatory mechanisms of the neurogenic manifestations of autoimmune diseases through the action of indoleamine 2,3-dioxygenase (IDO), the rate-limiting enzyme in the tryptophan degradation [11-15].

This review summarizes the actual status of the knowledge of the neurological manifestations in SS and presents the hypothesis of the association between these neurological manifestations and the TKP (Box 1).

Box 1. Summary of evidences linking Sjögren’s syndrome (SS) and the tryptophan/kynurenine signaling pathway (TKP)
• Association among chronic inflammation, pain and neuropathic disorders
• Triggered by Interferon
• Modulated by sex hormones
• Tryptophan deprivation induces dry eye
• Sjögren’s syndrome and salivary gland aggression leads to increased expression of kynurenine

2. Autoimmunity, neuropathy and chronic pain

Autoimmunity is linked to chronic neuropathy in at least three different forms. The first and major outcome observed is the pain, in its different dysfunctional manifestations, as alldynia, dysesthesia, hypo or hyperestesia and hyperalgesia [9,16,17]. Second, chronic inflammation in the target organs generates a noxious stimulus that may persist in a further phase where the inflammatory process is already resolved [18,19]. Third, the central nervous system (CNS), mostly the autonomic nervous system dysfunction can induce or perpetuate an unbalanced inflammatory response in the target-innervated organs [3,20]. Therefore, an unified theory of the relationship between the neural and immune mechanisms of the SS manifestations take in account that lesions of the autonomic and peripheral neural system reduce the threshold for inflammatory and noxious events in their connected organs, and disturb the balance between pro and anti-inflammatory mediators [21-23] (Figure 1).
Figure 1. A schematic model, showing the interrelationships among the autonomic nervous system dysfunction, pathological pain and chronic inflammation in Sjögren’s syndrome (SS).

The chronic pain and persistent inflammation mediated by humoral factors and neurotransmitters associate autoimmunity and neuropathy in several target organs, including the hippocampus and the lacrimal functional unit (LFU) on SS [20,24-28]. The dissociation between signs and symptoms of dry eye and dry mouth delays and makes the diagnosis more difficult in SS but paradoxically, those individuals present a high association between pain sensation in different organs and dry eye and dry mouth [5,29-35].

Our hypothesis consider the TKP in the mechanism of SS neuropathy, since compensatory crosstalk pathways can drive the dissociation between signs and symptoms among the immune, the endocrine and the neural systems [36-41].
This hypothesis predicts a spectrum of manifestations with two different poles of the SS disease. In one pole, the major characteristic is the chronic non-inflammatory pain with neuropathic features. In the opposite pole, primary SS present with inflammatory activity, including extra-glandular manifestations (EGM). However, these profiles can only be documented in long-term cohort studies [42]. In other words, we hypothesized that SS patients with Interferon-γ (IFN-γ)-inducible TKP activation could develop chronic pain and low EGM disease activity because the TKP promotes an immunosuppressive and neuro sensitive effect. Therefore IFN-γ-inducible TKP could be the missing link between disease activity and neural manifestations in SS [43] (Figure 1).

2.a. SS and Neurological manifestations

Neurological signs and symptoms have been described in SS. Peripheral neuropathy ranges from 8-49%, depending on selecting bias related to different classification criteria, and whether neurologic manifestations were diagnosed based on clinical marked symptoms versus asymptomatic, or detected by electrophysiological studies [2,44-48]. Distal sensory and sensorimotor neuropathies are the most common manifestations of peripheral nerve disease in pSS. Sensory neuropathies include painful nonataxic sensory polyneuropathy, small fiber neuropathy, dorsal root ganglionitis, and trigeminal neuropathy. Other forms are also described including multiplex mononeuritis, acute or chronic inflammatory demyelinating polyradiculoneuropathy (CIDP), multiple cranial neuropathy specially III, V, VI, VII, IX, X and
XII and last but not least, dysautonomia, which is very common in SS, and can reach a frequency of 40% of SS patients, in combination or not with different neurological manifestations [49-52]. Moreover, CNS manifestations may cause focal syndromes (such as multiple-sclerosis-like, epilepsy, movement disorders, neuromyelitis optica, and pseudotumor) and diffuse syndromes (encephalitis, meningitis, cognitive dysfunction, psychiatric disorders). Acute or chronic myelopathy and inferior motor neuron disease may also occur [44,45,49,53-58] (Table 1).
<table>
<thead>
<tr>
<th>Description of the disorder</th>
<th>Study model</th>
<th>N</th>
<th>Author, year</th>
</tr>
</thead>
<tbody>
<tr>
<td>Major depressive and panic disorder</td>
<td>Case series</td>
<td>2</td>
<td>Pelizza et al, 2010 [59]</td>
</tr>
<tr>
<td>Visual hallucinations</td>
<td>Case report</td>
<td>1</td>
<td>Wong et al, 2014 [60]</td>
</tr>
<tr>
<td>Dementia</td>
<td>Case report</td>
<td>1</td>
<td>Caselli et al, 1991 [61]</td>
</tr>
<tr>
<td>Migraine, neuropsychiatric disease, and focal acute neurological deficits</td>
<td>Prospective</td>
<td>48</td>
<td>Escudero et al, 1995 [62]</td>
</tr>
<tr>
<td>Cerebral manifestations (focal and diffuse) and spinal cord disease</td>
<td>Case series</td>
<td>16</td>
<td>Alexander et al, 1982 [63]</td>
</tr>
<tr>
<td>Peripheral nervous system abnormalities</td>
<td>Cross-sectional</td>
<td>39</td>
<td>Barendregt et al, 2001 [46]</td>
</tr>
<tr>
<td>Sensorimotor/sensory axonal polyneuropathy, spinal cord disease, and cognitive dysfunction</td>
<td>Prospective</td>
<td>25</td>
<td>Lafitte et al, 2001 [64]</td>
</tr>
<tr>
<td>Peripheral neuropathy</td>
<td>Cross-sectional</td>
<td>46</td>
<td>Gemignani et al, 1994 [65]</td>
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<tr>
<td>CNS disease, mostly non-focal dysfunction; PNS disease, mostly mild or severe sensory or sensory-motor polyneuropathies</td>
<td>Cohort</td>
<td>87</td>
<td>Govoni et al, 1999 [66]</td>
</tr>
<tr>
<td>Sensory ataxic neuropathy, painful sensory neuropathy without sensory ataxia, multiple mononeuropathy, multiple cranial neuropathy, trigeminal neuropathy, autonomic neuropathy, and radiculoneuropathy</td>
<td>Cross-sectional</td>
<td>92</td>
<td>Mori et al, 2005 [44]</td>
</tr>
<tr>
<td>Motor neuropathy, sensory neuropathy, sensorimotor neuropathy, and small-diameter nerve fiber neuropathy</td>
<td>Cross-sectional</td>
<td>62</td>
<td>Goransson et al, 2006 [48]</td>
</tr>
<tr>
<td>Complaints of painful distal paresthesias in the feet, abnormal sweating, and decreased pinprick sensation. Small-fiber neuropathy</td>
<td>Cohort</td>
<td>32</td>
<td>Lopate et al, 2006 [67]</td>
</tr>
<tr>
<td>Subcortical dementia</td>
<td>Case report</td>
<td>1</td>
<td>Kawashima et al, 1993 [68]</td>
</tr>
<tr>
<td>PNS involvement: small fiber neuropathy; trigeminal, facial, or trochlear nerves involvement; multiple mononeuropathy; sensorimotor polyneuropathy; autonomic neuropathy; and myasthenia gravis. CNS involvement: headache; spinal cord involvement; seizures; motor and sensory deficit; movement disorders; neuromyelitis optica; aseptic meningitis. Cognitive dysfunction</td>
<td>Retrospective cross-sectional case-control</td>
<td>93</td>
<td>Teixeira et al, 2013 [69]</td>
</tr>
<tr>
<td>Atypical neurologic manifestations: pseudotumoral lesion; multiple mononeuropathy; progressive multiple sclerosis; and myelitis along with progressive cognitive disorders</td>
<td>Case series</td>
<td>4</td>
<td>Michel et al, 2011 [70]</td>
</tr>
<tr>
<td>Neuromyelitis optica</td>
<td>Case series</td>
<td>2</td>
<td>Nitescu et al, 2011 [71]</td>
</tr>
<tr>
<td>Peripheral neuropathy: axonal sensorimotor polyneuropathy, pure sensory neuronopathy, mononeuropathy multiplex, and demyelinating polyradiculoneuropathy</td>
<td>Cross-sectional</td>
<td>102</td>
<td>Brito-Zeron et al, 2013 [72]</td>
</tr>
<tr>
<td>Multiple sclerosis</td>
<td>Case report</td>
<td>1</td>
<td>Liu et al, 2014 [73]</td>
</tr>
<tr>
<td>Neuropathy axonal (pure sensory or sensorimotor)</td>
<td>Cross-sectional</td>
<td>44</td>
<td>Pavlakis et al, 2011 [74]</td>
</tr>
<tr>
<td>CNS vasculitic involvement</td>
<td>Case report</td>
<td>1</td>
<td>Hasiloglu et al, 2012 [75]</td>
</tr>
<tr>
<td>Neuromyelitis optica</td>
<td>Retrospective</td>
<td>43</td>
<td>Qiao et al, 2015 [55]</td>
</tr>
<tr>
<td>Autonomic dysfunction</td>
<td>Cohort</td>
<td>154</td>
<td>Koh et al, 2017 [76]</td>
</tr>
</tbody>
</table>

Table 1. List of the main neurocognitive changes in Sjögren syndrome described in several study models
N: Number of study cases; CSN: Central Nervous System; PNS: Peripheral Nervous System
This high variability of clinical manifestations of SS with different responses to treatment should be related to diverse and complex mechanisms of injury such as vasculitis and lymphocytic infiltration. Changes in the structure of the dorsal spinal cord and the dorsal ganglion roots, association with autonomic dysfunctions, decrease in the white matter, and loss of the gray matter of the hippocampal area were observed in SS patients [77-79].

2.b. SS and the mechanisms of neurological manifestations

The mechanisms triggering the neurological manifestations in SS are unclear. They involve genetic predisposition, environmental agents, trauma and posttraumatic stress, autoimmunity against the CNS and peripheral nervous systems (PNS), in addition to the neuroimmunendocrine network disruption [47,80-85]. More specifically, unknown causes trigger DNA demethylation, microRNA abnormal expression, unbalance of Interferon I (α and β) and II (γ), and anti-neuron autoantibodies production [55,83,86-88]. The 2-5 oligo-adenylate synthetase 1 (OAS1) gene defect leads to lower responsiveness to IFN-γ, and induces higher production of IFN-γ causing severe complications as neuropathy in SS [85].

The increase in corneal nerve thickness and higher number of antigen presenting cells in the cornea, nerve vasculitis, nonvascular encephalitis, neuromyelitis, axonal and CNS degeneration are hypothesized to be responsible for those sensorial, autonomic, cognitive or behavioral neurological manifestations of SS [24,47,81,89].

It is interesting to notice that the same structures that are altered in image exams of SS patients compared to control individuals are those where the TKP has shown to be more sensitive to handle in experimental studies, as synapses of the
sensorial fibers, at the dorsal ganglion root, hippocampus, thalamus and LFU [2,12,13,27,39,77,90-95] (Table 2).

Table 2. Changes in the Nervous system and in the exocrine glands in Sjögren’s Syndrome (SS) and after the modulation of the tryptophan/kynurenine pathway (TKP).

<table>
<thead>
<tr>
<th>Structure</th>
<th>Sjögren’s Syndrome</th>
<th>Tryptophan/kynurenine pathway</th>
</tr>
</thead>
<tbody>
<tr>
<td>Dorsal ganglion root</td>
<td>Dorsal ganglion root alterations in the MRI, associated with increased intradermal nerve fiber density in skin biopsy [96]</td>
<td>Sciatic injury increase kynurenine monooxygenase (KMO) in the dorsal ganglion root of rats [91]</td>
</tr>
<tr>
<td>Hippocampus</td>
<td>Hippocampal atrophy in SS patients [27]</td>
<td>IDO and kynurenine-3-hydroxylase increase in the hippocampus after day 2 after CNS ischemia [97].</td>
</tr>
<tr>
<td>Exocrine Glands and LFU</td>
<td>Changes in the LG and the SG in the MRI, nerve changes in the cornea of SS patients [94,98]</td>
<td>Increase in kynurenine in salivary gland after ductal ligation, LG atrophy due to tryptophan deprivation [90,99]</td>
</tr>
</tbody>
</table>

MRI: magnetic resonance image, SS: Sjögren’s syndrome, LG: lacrimal gland, SG: salivary gland, CNS: central nervous system, IDO: indoleamine 2,3-dioxygenase

2.c. The immune and the endocrine modulation of neurological findings in SS
The concept implicit in the neuroimmunendocrine network predicts that cellular and molecular communication among those three systems are responsible for the homeostasis of the body organs; and a disruption in this network take part in the mechanism of the diseases [100-103]. Acetyl-choline (Ach), dopamine, glutamatergic and other neurotransmitters, are also secreted by lymphocytes [104]. In the other hand, the autonomic nervous system is capable of modulating lymphocytes proliferation in target organs like spleen, liver, kidney, and brain [23,105,106]. In addition, the sensory neurons are capable to secrete peptides with immunomodulatory properties as galanin, netrin-1, and somatostatin and promote or attenuate inflammatory responses [107,108].

Hormones, in particular the sex hormones, can modulate the inflammation and the pain, sensitizing the ionic receptors expressed in the neurons and epithelial cells, called transient receptor of potential (TRPs), and stimulating growth factors and cytokines expression in the target tissues as lacrimal and salivary glands (LG and SG), hippocampus and trigeminal ganglion of the CNS, and also on other target tissues[109-116]. Those mechanisms explain the sex hormones mediated amplification and perpetuation of the inflammatory process, the pain hypersensitivity and exocrine glands dryness manifestations in SS, where estrogen potentiate the pain and pro-inflammatory mediators and androgens work oppositely [80,111,117-119]. Although the trigger of the first event in SS and the steps to the chronic phase are poorly understood, there are strong clinical association between female sex hormones, and the inflammatory mediators involved in innate, and also adaptive immunity [4,44,111,113,120,121] (Figure 1).
These observations taken together indicate that hormones and neural pathways are involved in immune responses, resulting in variability in the pain sensation, inflammatory reaction, tissue integrity and functional disruption [5,23]. Those observations are in agreement with recent reports describing multiple comorbidities and extra glandular manifestations (EGM) in SS patients [2,92,122].

The loss or damage of the nerve fibers, dorsal root ganglionitis, nerve vasculitis and reduction of the CNS matter can be observed in image exams, as magnetic resonance image (MRI) or skin biopsies in SS; they are also implicated in the mechanisms of the diseases, and work as diagnostic markers of SS [24,77,78,96]. For example, the cognitive impairment on SS patients are associated with antibodies against NR2 subtype of the N-metyl Aspartate receptor (anti-NR2 antibodies) in the cerebrospinal fluid (CSF) mediating hippocampal gray matter atrophy, observed by MRI [27]. In summary, the SS inflammatory activity in the PNS and CNS causes the signs and symptoms described above, and are modulated by hormones, neurotransmitters and cytokines susceptible to the TKP interference [13,14,102].

3. The tryptophan/kynurenine pathway (TKP)

The metabolism of L-tryptophan (LTF) leads to the generation of several neuroactive compounds by the serotoninergic and the kynurenine pathways. In the serotoninergic pathway, L-tryptophan is metabolized to serotonin (5-HT), and in some cells, to melatonin. Serotonin acts as a neurotransmitter with a variety of functions on behavior and psychiatric symptoms (depression and anxiety), platelet aggregation, gastrointestinal
tract control (satiety, secretion, and peristalsis) and tumor resistance.

Melatonin acts as a neurohormone in the circadian rhythm with also an anti-inflammatory, anti-angiogenic and anti-tumoral immunomodulatory effects [123] (Figure 2).

In the catabolic TKP, the tryptophan 2,3-dioxygenase (TDO) specifically metabolizes tryptophan in the liver and responds to hormonal regulations, such as cortisol and glucagon, and to the tryptophan concentration [124] (Figure 2).

The IDO enzyme regulates both innate and adaptive immune responses through the degradation of the essential amino acid tryptophan into kynurenine and other metabolites, which suppress effector T-cell function and promote the differentiation of regulatory T cells. The IDO also metabolizes serotonin, melatonin, and tryptophan until N-formil kynurenine and kynurenine in the lung, the brain, and blood, producing quinolinic acid and nicotinamide adenine nucleotide (NAD+) [125-127]. The TKP is responsible for 95-99% of the entire tryptophan catabolism [128] (Figure 2).

**Figure 2.** The tryptophan/kynurenine pathway (TKP) and their resulting metabolites.
IDO is found mainly in immune cells, and has enzymatic activity in the cytoplasm and also transcriptional activity in the nucleus, playing a unique role as a signaling molecule, modulating immune responses [129-131]. The
nuclear effect contributes to an enzyme “self-amplification” in an IFN-dependent loop that may account for the phenotype of tolerance, attenuating or preventing immune reactions and mediating persistent pain in several conditions, including SS \cite{129,130,132-134}. The IDO activity is induced by macrophages and by cytokines, like IFN-γ and TNF-α, by prostaglandins, by viral and bacterial infections and by lipopolysaccharides, by dendritic cells and the sub-products of kynurenine \cite{135-139}. IDO overexpression has been documented in patients with systemic lupus erythematosus (SLE) and SS, as well as in sepsis \cite{15,36,140}. In patients who are positive for the IFN gene expression signature, Treg cell levels are elevated in combination with increased IDO activity, with tolerance and immune modulation\cite{132}. Those regulatory T lymphocytes cells represent a diverse subclass of T cells, a protagonist in the maintenance of self-tolerance and immune modulation\cite{141}.

The TKP is catalyzed by the IDO enzyme and it can be induced by the cytokine IFN-γ, as observed in studies with mice where IFN-γ or IFN-γ receptor knockout prevent kynurenine production \cite{142}. The phenomenon is dependent on antigen-presenting cells (APCs) mentioned above, as observed in mice models with graft-versus host disease (GVHD) and induction of this TKP can extend the life-span and reduce the inflammation of the gut of the wild-type mice, but not of those IFN-γ receptor knockout mice \cite{142}.

The balance between INF-α and IFN-γ activation in the inflammatory processes, either in response to an exogenous aggression or as part of an autoimmune disease as SS can interfere in the activation of the TKP and in the resulting intensity of this inflammatory process \cite{143-145}. The production of the IFN-γ and TKP activation is also dependent on the genetic background.
and on the interaction between the T cells and other cells in the target tissues of inflammatory diseases, where the pretreatment IFN-γ can suppress the presentation of the auto antigens by those target cells [146-148].

The isoform IDO2, and also TDO1 are enzymes involved in the catabolism of the amino acid tryptophan and operate in similar ways in the immunomodulation, with variations in the target tissues, cells involved in the metabolic pathways in several conditions. However, further studies are necessary to clarify their diversity in diseases, including SS [149,150].

4. The TKP and the neurologic manifestations

The relationship between tryptophan, serotonin, and depression has an extended history in psychiatry. The development of depressive symptoms is correlated with high levels of tryptophan metabolites in urine [37] and with a decrease of tryptophan in the blood and cerebrospinal fluid [151,152]. The tryptophan transport across the blood-brain barrier, specific inflammation and damage caused by brain-reactive autoantibodies and immune complexes play a critical role in the regulation of tryptophan metabolism in the brain [123]. There is substantial evidence to suggest that in addition to serotonergic neurons, other cells such as astrocytes, dendritic cells, microglia, and macrophages also synthesize multiple neuroactive metabolites via the enzyme IDO and the TKP at CNS [153]. Evidences underlying the mechanisms have come from clinical studies that examined the effects of IFN-α on the mood of cancer and hepatitis C virus-infected patients. In both, the development of depressive symptoms was associated with decreased circulating tryptophan levels and enhanced formation of kynurenine, indicating that IDO pathway is active [154-156].
Tryptophan metabolites generated in the TKP have been associated with neurodegenerative diseases such as acquired immunodeficiency syndrome (AIDS)-related dementia, Alzheimer’s and Huntington’s diseases, and neuropsychiatric diseases like bipolar disorder and schizophrenia [157,158]. The synthesis of kynurenine in the CNS is affected by dietary intake of tryptophan and by the gut microbiota, and deprivation induces depression, cognitive dysfunction (attention, memory, and execution), among other neurologic dysfunctions through the glutamatergic receptors [14,40,159-162].

Tryptophan metabolism in astrocytes leads to the production of the kynurenic acid reported to take part in neuroprotective actions. On the other hand, microglia cells give rise to metabolites with reactive oxidative properties including hydroxykynurenine and 3-hydroxyanthranilic acid and quinolinic acid which also acts as an agonist at the glutamate N-methyl-D-aspartate (NMDA) receptor subtype and may contribute to excitotoxicity and neurotoxicity [163-165].

In healthy subjects, there is a well-adjusted system in the TKP by the action of kynurenines aminotransferases (KATs) towards kynurenic acid or to quinolinic acid by (KMO). Kynurenic acid, which is reported to promote neuroprotective and immunosuppressive actions in CNS, and plays a role as an NMDA antagonist, blocking this glutamatergic receptor. An enhanced IDO activity and a deviation to KMO downstream are observed by stimulation of cytokines such as IFN-γ [166]. KMO has a clear inflammatory and pro-apoptotic drive. Its intermediate compound, quinolinic acid, acts as an agonist at NMDA receptor, modulating excitatory amino acid transmission, and may serve as neurotoxic agent implicated in the pathogenesis of several.
neurologic diseases\cite{123,157}. Likewise, 3-hidroxikynurenine has also neurotoxic effect, probably associated with the conversion of reactive oxygen species and apoptosis \cite{167-170,78}.

The NR2 subtype NMDA receptor is ubiquitously distributed through the brain with a unusually high density in the hippocampus \cite{171}. Hippocampus is a brain structure, linked to the autonomous nervous system, with critical importance for memory formation and learning, and is also affected in mood disorders and in SS \cite{77,78,172,173}. Likewise, the NR2B subunit of NMDA receptor is widespread in dorsal root ganglion and may mediate peripheral sensitization and visceral pain \cite{174}. Those receptors are critically involved in the initiation and maintenance of neuronal hyperexcitability after noxious events, by C-fiber stimulation \cite{175}.

In the rodent model of peripheral nerve injury, tibial and peroneal nerves of one leg were sectioned, but the sural branch nerve was left intact. After seven days, it was observed that IDO1 was activated and the kynurenine rose in the bloodstream, accompanied by the depressive behavior, confirmed by the extended time of immobility in the forced swim test and allodynia in the paw withdraw in response to mechanical stimulation with von Frey hair. Those findings were followed by an increase in the levels of KMO, quinolinic acid, and reduce the kynurenic acid in the contralateral hippocampus, confirming the role of the hippocampal neurons as the site in the CNS for the perpetuation of the symptoms. The inhibitory effect of interleukin-1β (IL-1β) by injection of IL-1β receptor antagonist in the CNS ventricular space reduced the depressive behavior and KMO mRNA levels but did not change the allodynia \cite{39}. Depression was associated with decreased
levels of tryptophan in patients with cancer receiving therapy with IL-2 and IFN-α, suggesting that those cytokines impact the levels of serotonin [155].

Studies observing the triggering of inflammation of the macaque CNS with poliovirus inoculation revealed that quinolinic acid, kynurenine, and other metabolites of the TKP accumulate in the spinal cord, CSF, but not as much in the bloodstream, in levels that were corresponding to the clinical manifestations of functional damage [12,135]. Moreover, the *in vitro* conversion of L-tryptophan in kynurenine by fetal neuronal cells was dependent upon IFN-γ stimulation in the presence of macrophages in the culture [12].

Confirming the hippocampus as a target regulatory CNS organ in pain and depression, it was shown that chronic pain in rats exposed to social stress or paw arthritis increase the levels of IDO, and also increase the levels of kynurenine and decrease the levels of serotonin in the hippocampus [176]. This situation is similar to the observed in human plasma levels of IDO, kynurenine/tryptophan and serotonin/tryptophan levels in patients with back pain and depression, where the first two rise and the third one decreases, as revealed in the same paper. Moreover, the IDO1 knockout mice presented lower nociceptive and depressive behavior compared to the wild type, and this behavior was not attenuated in the wild type that received an intraperitoneal injection of the NSAID acetaminophen, suggesting that this behavior is not dependent of the inflammatory mechanisms alone. Furthermore, the authors found that IL-6 is overexpressed in rats with arthritis and the Jak2/Stat3 signaling pathway activated in the blood and the hippocampus of rats with depressive and nocioceptive behavior. The injection of IL-6 anti-serum attenuated the allodynia and hyperestesia in those animal models. In Neuro2a
cells (a mice neuroblastoma cell line) in culture, the incubation with IL-6 induced increase in IDO1 mRNA and protein [176].

Therefore, persistent pain with alldynia and hyperalgesia has a central component (spinal and supraspinal cord), supporting the involvement of glutamatergic neurotransmission, associated with TKP signaling in the clinical manifestations and the role of the hippocampus as a critical organ in this process and the TKP in their physiopathology [9,16,28,176,177]. Those facts allow us to exam the relationship among TKP, neurologic manifestations and autoimmune diseases like SS.

4.a. The role of the hippocampus in the TKP of the neurologic manifestations

Despite the agreement among clinical studies on changes in the TKP and SS, it is admissible that the lack of association between the symptoms of depression and fatigue in SS and the changes in the TKP is due to difficulties to monitor the changes in the CNS, more specifically in the hippocampus [43,132,178,179]

Not only SS patients but also individuals exposed to chronic stress present signs of hippocampal structure and NMDA signaling changes [27,180]. Animal models studies reveal that the hippocampus initially adapts to early, high and frequent stress, but the persistence of the aggression rise the levels of glutamate and disrupt the hormonal and neurotransmitters control, leading to a NMDA driven neuronal death and hippocampus atrophy [181,182].

Moreover, hypothalamus-pituitary-adrenal activity has a modulatory effect on those events and the female sex hormone estrogen increase the
synaptic connections and the expression of Nerve Growth Factor (NGF) in the hippocampus, supporting the hormone influence and the increased symptoms and pain sensitivity in the females with SS [111,114,181,183].

The cytokines, like IFN-γ, IL-1, TNF-α increase the kynurenine expression, from the tryptophan by the IDO enzyme, deviating this amino acid form the production of serotonin in the CNS [13]. The resulted imbalance between the kynurenine metabolites and serotonin production in the hippocampus induce depression, slow reactions and other cognitive disorders [184]. The target cells are microglial cells, astrocytes and other inflammatory cells present in the hippocampus and other CNS areas [19]. Once impacted by those cytokines, the cells reduce the glutamate re-uptake, increase the glutamatergic signaling, reduce the capacity to produce serotonin, trigger the nociceptive and depressive behavior, induce cells death (mostly the astrocytes) prolonging the inflammatory effect [13,19,28,39,185].

It is interesting to note that serotonin works as a modulator of the glutamate actions. The PNS sensory transmission has silent glutamate synapses that are activated by serotonin. Once those silent synapses are serotonin-activated, they amplify the peripheral nociceptive glutamate signaling through the NMDA or AMPA receptors from the spinal dorsal horn to the CNS [186,187]. In the CNS, the glutamate-serotonin co-neurotransmission are extensively studied, including in the hippocampus. The five sub-types of serotonergic receptors are expressed in different combinations among several cells, antagonizing the glutamatergic NMDA receptors at different levels, from preventing cells glutamate release to competing for the same intracellular signaling pathways in the hippocampus but not in other brain tissues [188-193].
Also, to demonstrate the differences among the TKP activity in the CNS and in other parts of the body, the systemic treatment with dexamethasone to reduce the inflammation induced by lipopolysaccharide intraperitoneal injection (LPS) promoted a decrease of IDO enzymes in peripheral tissues (lung, spleen and liver) but an increase in the brain microglial cells and astrocytes [185,194]. Moreover, the use of systemic subcutaneous slow-release corticosteroid pellets in rats increased the levels of NR2 NMDA glutamatergic receptors mRNA in the hippocampus [195]. Those observations suggest that corticosteroid treatment, used to reduce chronic inflammation may potentiate the nociceptive and depressive behavior in the long-term.

Also, it was shown in HIV-infected patients, that quinolinic acid (the metabolite of the kynurenine, that mimics the glutamate in NMDA receptors), is several times higher in the brain, than in the cerebrospinal fluid or in the blood [196].

Taken together, these information locate the hippocampus as a primary responsible for nociception and mood control, also a site where the rise in TKP activity increases in response to inflammation, and its metabolites (i.e., quinolinic acid and glutamate) induce manifestation of pain and depressive behavior, which can be amplified by serotonin waking up of extra peripheral silent glutamatergic synapses. Moreover, in the chronic inflammations, like the SS, the levels of serotonin in the CNS are diminished by tryptophan consumption throughout the TKP. The attempt to revert the inflammation with corticosteroids, is not achieved in the CNS as it occurs in other target organs and ends up with the death of microglial cells, astrocytes and neurons, mostly
the hippocampus and the dorsal ganglion root [185,194,197]. These observations support the possible mechanisms of SS neurologic manifestations and the TKP (Figure 3).

**Figure 3.** The tryptophan/kynurenine pathway (TKP) and its implications in the Sjögren’s syndrome physiopathology.

5. **The TKP and the neuropathy in SS**

   Decades ago, reports revealed that the ingestion of L-5-Hydroxytryptophan induced signs and symptoms similar to scleroderma, with high plasma levels of kynurenine [198]. Moreover, excessive doses of tryptophan (upper than 1.2 g/day) triggered eosinophilia, severe muscular weakness and pain, and oral ulcers, with rise of the hepatic enzymes aspartate and alanine aminotransferase (ASA and ALA), in addition to inflammatory infiltrate in various organs [161]. Those events were associated with high plasma levels of kynurenine as observed in cases with scleroderma and SS [140,198].
Salivary gland (SG) ductal ligation in rats induces tissue suffering and atrophy and increase in the systemic levels of kynurenine (as measured in the hair) and was associated not just with salivary hypofunction but also with body weight loss, along of the six months of the experimental period. It indicates that higher levels of plasma kynurenine can reflect a peripheral organ suffering but also that SG damage is sufficient to impact the whole body metabolism as shown by the body weight loss compared to controls [99].

Those observations sound otherwise to the hypothesis for the neuropathic pain combined with less EGM profile for a subgroup of SS patients driven by the TKP, presented in session 2; and the anti-inflammatory and immune tolerance effects of the TKP introduced in session 3b, in models of GVHD, viral infection, tumors or chemical challenges. However, it is necessary to understand the role of different triggers on each target organ, the individual capacity to metabolize and eliminate the side products of the TKP and those events along of the time course of the disease, including the influence of distinct therapeutic strategies.

As mentioned above, the tryptophan levels in the body are dependent on dietary intake, however, the influence in the serotonin conversion is limited to about 10% of the total intake, due to the hemato-encephalic barrier and other environmental stimuli, but also the individual metabolic conditions given by the genetic background [37,159-161,199-201]. For example, when female C57 ovariectomized mice ingest bisphenol A (BPA), an environmental contaminant with endocrine disruption capacity, it causes bowel inflammation and reduction in the levels of tryptophan and serotonin, indicating that
environmental contaminants and the intestinal microbiota affect the TKP in the chronic inflammatory diseases [160].

The flow cytometric analysis of peripheral blood revealed a higher expression of IDO in dendritic cells of pSS patients, and in each of the subgroups, classified either by the presence of clinical or serological activity or none of those, compared to dendritic cells of healthy controls, and those findings were associated with immune regulation by IFN-α [202].

The measurement of the median expression of IDO in antigen presenting cells (APCs) and in T cells were higher in peripheral blood cells of pSS patients than in controls, matched by age and sex, using also specific antibodies and flow cytometry, despite of the heterogeneity of the groups and the high internal variability of the results [203]. Therefore, the T cell mediated autoimmune activity present in autoimmune diseases, including SLE and SS have been associated with higher activity of the TKP, although their effects on auto antigen stimulation and the IFN-γ activity is unknown [202,203].

Different profiles regarding IFN-γ activity were identified in the pSS population, in addition to 55 genes and 19 metabolic pathways were distinctly identified in a subset of pSS patients with fatigue [204,205]. Higher IDO activity was observed in IFN-γ positive pSS patients, with higher levels of IDO mRNA and IFN-γ mRNA in circulating monocytes, and those observations were associated with the up-regulation of apoptotic and neurotoxic downstream steps in the TKP [132]. The levels of the serum tryptophan are higher in healthy than in primary SS women (pSS). On the other hand, the kynurenine levels and the kynurenine/tryptophan ratio are more elevated in pSS than in healthy women and patients with sicca non-SS. Those observations were also
present in pSS men, confirming a higher activity of the IDO enzyme in the TKP [140,206]. Moreover, the higher levels of kynurenic acid were associated with higher levels of inflammatory markers in the serum, as erythrocyte sedimentation rate, c-reactive protein, creatinine, IgA, β-2 microglobulin and positivity to anti-nuclear antibody. Higher levels of kynurenic acid was also associated with a lower proportion of individuals on corticosteroid usage but not with the frequency of neurologic manifestations in the pSS group [140,206].

In another recent study, polyneuropathy combines with more frequent positivity for the autoantibodies anti-Ro (SSa) and anti-La (SSb) in pSS [42]. Taken together, these studies suggest that TKP higher activity is related to clinical and laboratory signs of systemic inflammation, which may disagree with our hypothesis but also can indicate that those studies documented a midway between the pain/neuropathic and the inflammatory poles of the disease [140,206,207]. Although the association of neurological or laboratory findings and kynuriec acid metabolites is evident in those studies, the cause/effect relationship between the metabolites of the TKP and these manifestations is still unclear.

In another study, an association of fibromyalgia, and other psychological symptoms, as anxiety, depression, insomnia, psychoticism and neuroticism with fatigue were observed in a large series of pSS patients with 106 cases, where 32 were fatigued and 74 non-fatigued, identified by the Functional Assessment of Chronic Illness Therapy-Fatigue (FACIT-F) scale, with a cut-off of 30 on a scale ranging from 0 to 52. However the measurement of the IDO mRNA in peripheral blood leukocytes was not different between pSS patients with and without fatigue [178]. In addition, no
other clinical or laboratorial association was identified, except for the number of individuals using hydro chloroquine, which were 50% among the pSS fatigue group and 28% in the pSS non-fatigue group. On the other hand, it was observed a higher expression of IDO-1 mRNA levels associated with plasma levels of IFN-γ [178].

Fatigue was associated with higher levels of TKP activity in SLE patients, but only on those with clinical activity of the disease measured by the Systemic Lupus Erythematous Disease Activity Index (SLEDAI) with the score above five, which may confirms the possibility that the TKP is overexpressed when challenged by higher inflammatory activity [36]. Moreover, the levels of serum tryptophan are lower in SLE than in controls, scleroderma and pSS patients, who may reflect the higher activity and broad manifestations of the disease, compared to the other two conditions (i.e., pSS and SSc) that are more tissue-specific than SLE [179]. Experimental studies mimic the clinical findings of cognitive impairment, but not of depression, associated with microglia and astrocyte activation in the hippocampus of SLE mice models induced by the injection of anti-ribosomal antibodies in the CNS, compared to controls [208]. Unfortunately, the TKP was not investigated in that study.

The failure to present a more conclusive association between the TKP, as a marker of the neurologic manifestations in SS may be due to: a) the heterogeneity of the cases, b) the lack of a healthy control group in some studies, c) the possible dissociation between the levels TKP activity in the blood and the biochemical changes in the CNS tissues, specially the hippocampus, d) a higher TKP activity takes place in response to the inflammatory activity but persists only on those target organs (e.g.;
hippocampus and dorsal ganglion root) after the inflammatory control. Those
pitfalls must be taken into consideration in future studies addressing the
present hypothesis of the association between SS neurologic features and
TKP.

6. Therapy to modulate the TKP

The overexpression of IFN-γ, the induction of pro-inflammatory genes,
as TNF-α, interleukins, BAFF, promoting the activation of B cells and the rise
of autoantibodies in the blood are involved in the physiopathology of SS
[84,143,209-213]. Therefore, the therapeutic strategies to treat SS include
immune modulators and biological therapy to refrain B and T cells activity and
proliferation [214-217]. The limitations of those strategies open opportunities for
new procedures and complementary therapies. Among the several
possibilities, the potential modulation of the TKP has been explored in SS, as
in other autoimmune diseases, including rheumatoid arthritis, SLE, systemic
sclerosis, despite of the uncertainties in the influence of this pathway in the
mechanisms, as pointed in session 5 [15,218] (Table 3).
### Table 3. List of studies showing potential therapeutic strategies to modulate pain and inflammation targeting the therapeutic interventions in the kynurenine pathway

<table>
<thead>
<tr>
<th>Description of the therapeutic interventions in the kynurenine pathway</th>
<th>Study model</th>
<th>Author, year</th>
</tr>
</thead>
<tbody>
<tr>
<td>Administration of inhibitor of kynureninase (mNBA and oMBA) increased the content of kynurenine and KYNA in the brain, blood, liver, and kidney following. There was a decrease in locomotor activity and protection of audiogenic seizures.</td>
<td>Experimental study in rats</td>
<td>Chiarugi et al, 1995</td>
</tr>
<tr>
<td>m-NBA administration significantly increased the concentration of kynurenine and kynurenate in the brain, blood, and in the liver. This increase is associated with sedative and anticonvulsant actions, suggesting a functional antagonism of the excitatory amino acid receptors.</td>
<td>Experimental study in rats</td>
<td>Carpenedo et al, 1994</td>
</tr>
<tr>
<td>Administration of sulfur-containing amino acids, L-cysteine sulphinate, L-cysteate, L-homocysteine sulphinate and L-homocysteate reduced KYNA production and inhibited the activity of KAT I and/or KAT II.</td>
<td>Experimental study in rats</td>
<td>Kocki et al, 2003</td>
</tr>
<tr>
<td>N(G)-nitro-L-arginine and its methyl ester impair brain synthesis of kynurenic acid, probably via NO-independent mechanism, what could contribute, to the enhancement of neurotoxicity or seizures observed in some experimental designs based on their use.</td>
<td>Experimental study in rats</td>
<td>Luchowski et al, 2001</td>
</tr>
<tr>
<td>Activation of the PGC-1α1-PPARα/δ pathway increases skeletal muscle expression of kynurenine aminotransferases, thus enhancing the conversion of kynurenine into kynurenic acid. Reducing plasma kynurenic acid protects the brain from stress-induced changes associated with depression.</td>
<td>Experimental study in rats</td>
<td>Agudelo et al, 2014</td>
</tr>
<tr>
<td>KYNA was an important early mediator of leukocyte recruitment in an in vitro vascular flow model.</td>
<td>In vitro human study</td>
<td>Barth et al, 2009</td>
</tr>
<tr>
<td>In an EAE, IDO activity was increased in the spleen during the preclinical phase, and within the brain and spinal cord at the onset of symptoms. Macrophages/activated microglia expressing IDO during EAE and IDO induction in microglia upon IFN-gamma treatment with synergistic effects of TNF-alpha. Inhibition of IDO by systemic administration of 1-Mt at clinical onset significantly exacerbated disease scores.</td>
<td>Experimental study in rats</td>
<td>Kwidzinski et al, 2005</td>
</tr>
<tr>
<td>The expression and activity of KMO significantly increased in the spinal cord in an EAE. The spinal cord content of 3-HK and quinolinic acid reached neurotoxic levels. Systemic administration of Ro 61-8048, a selective KMO inhibitor, reduced the increase of both 3-HK and quinolinic acid, and caused accumulation of KYNA.</td>
<td>Experimental study in rats</td>
<td>Chiarugi et al, 2001</td>
</tr>
<tr>
<td>QUIN levels were significantly elevated in the more caudal regions of the spinal cords of animals with EAE. The initial elevation in QUIN occurred before the appearance of behavioral signs. Last, treatment with the glucocorticoid dexamethasone prevented both the signs of EAE and the elevation in spinal cord QUIN.</td>
<td>Experimental study in rats</td>
<td>Flanagan et al, 1995</td>
</tr>
<tr>
<td>The NMDA receptor antagonist memantine to modify the neurological course of EAE. Significantly restored BBB integrity, reduced symptoms, and limited inflammatory lesions.</td>
<td>Experimental study in rats</td>
<td>Paul et al, 2002</td>
</tr>
<tr>
<td>In neuropathy, after CCI had an increase in the KMO mRNA levels in the spinal cord and the DRG that were reduced by chronic administration of the microglial inhibitor minocycline. There was a decrease in the intensity of neuropathy. KMO inhibitors (Ro61-6048) administration potentiated the analgesic properties of morphine.</td>
<td>Experimental study in rats</td>
<td>Rojewska et al, 2016</td>
</tr>
</tbody>
</table>
Chronic pain induced depressive behavior and IDO1 upregulation in the bilateral hippocampus. Upregulation of IDO1 resulted in the increased kynurenine/tryptophan ratio and decreased serotonin/tryptophan ratio in the bilateral hippocampus. IOD1 gene knockout or pharmacological inhibition of hippocampal IDO1 activity attenuated both nociceptive and depressive behavior

Experimental study in rats Kim H et al, 2012

mNBA: m-nitrobenzoylalanine; oMBA: o-methoxybenzoylalanine; KYNA: Kynurenic acid; KAT I/II: kynurenine aminotransferase I/II; NO: nitric oxide IDO: indolamine 2,3-dioxygenase; EAE: experimental autoimmune encephalomyelitis; 1-Mt: 1-methyl-tryptophan; KMO: kynurenine 3-monooxygenase; 3-HK: 3-Hydroxykynurenine; QUIN: Quinolinic acid; NMDA: N-methyl-D-aspartate; BBB: blood-brain barrier; CCI: after chronic constriction injury; DRG: dorsal root ganglia; IDO1: indoleamine 2,3-dioxygenase
Considering the broad spectrum of substrates, target tissues, and alternative ways of the TKP, the therapeutic strategies explore interference in different steps, observing one or more of the following outcomes: attenuate inflammation, reduce chronic pain or improve fatigue and depressive feelings in SS [132,203,218].

The traditional non-steroidal anti-inflammatory drugs (NSAID), acetylsalicylic acid (ASA or aspirin) and sodium diclofenac were investigated [230,231]. In rats, the systemic intraperitoneal injection of tryptophan alone was able to increase the kynurenine levels in the blood and the liver between 20 and 120 min. However, after combined injection of subcutaneous diclofenac with intraperitoneal tryptophan, the concentration of kynurenine increased, not just in the plasma and the liver, but also in the spinal cord and the brain, with a remarkable increase in the kidney, after 60 and 120 min. Therefore, diclofenac disturbs the renal clearance of TKP metabolites, which may amplify the anti-inflammatory and the excitatory stimuli on nociceptive NMDA receptors, independently of the prostaglandins analgesic effects [231]. In an opposite way, a study using human peripheral blood mononuclear cells (PBMCs) revealed that aspirin, at the dose of 5 mM, incubated for three days or 2 hours, reduced the tryptophan metabolism and kynurenine production on those human PBMCs, stimulated by Concanavalin A and pokeweed mitogen (PKM), suggesting an inhibitory effect on IFN-γ, based on those triggers mechanisms of action [230]. These observations indicate that NSAID works aside of their known effects on the cyclooxygenase/prostaglandins pathway, and the impact on the TKP can be diverse depending on the cell type and the specific NSAID [15]. It is also interesting to note that Aspirin, a longer and
broadly used analgesic drug, have well known positive effects on dry eye symptoms and LG dysfunction, critical elements on the manifestations of SS [232-234].

In an *in vitro* study, using samples of T cells from 68 SS patients, the co-culture with mesenchymal stem umbilical cells revealed the suppression of proliferation and activation of these T circulating follicular helper cells, associated with enhanced expression and enzymatic activity of IDO, measured by RT-PCR and HPLC, respectively [235]. Another study using human complementarity determining region 1 (hCDR1), a tolerogenic peptide complementary to the human anti-DNA monoclonal antibody has shown to reduce the expression of inflammatory cytokines, downregulate the proliferation and activity of B cells, and increase the expression of anti-inflammatory cytokines in rodent models of SLE [218]. In mature leukocytes of 16 SS individuals, in culture, it was shown that hCDR1 reduces the expression of inflammatory cytokines, including IFN-γ, and increase the expression of anti-inflammatory cytokines, up-regulating the IDO gene expression. However, in the presence of 1mT, an IDO inhibitor, the effect of hCDR1 on the T cell regulator cytokine FOXP3 gene expression is reduced, suggesting that the immune modulatory effect of hCDR1 is partially associated with its action over the IDO [218].

The suppression of the TKP by inhibiting the KMO can reduce the pain triggered by LPS injection in the dorsal ganglion root in rodents. It was demonstrated by systemic administration of the antibiotic minocycline or by the local administration of inhibitors of KMO [91]. They reduced the local levels of pro-inflammatory cytokines in the dorsal ganglion root and the spinal cord
and decreased the pain and the protein expression of the following inflammatory mediators IBA-1, IL-6, IL-1β and NOS2 [91].

Considering the strategy to overload the TKP to modulate the pain sensation, rats were subject to systemic administration of L-4-chlorokynurenine [236]. The experiments revealed that L-4-chlorokynurenine, a NMDA/glutamate receptor antagonist, given by intraperitoneal injection reached the CNS and attenuated the hyperalgesia in four models of pain and behavior response (general behavioral, formalin plantar injection, Carrageenan model and Chung neuropathy) compared to controls, MK-801 and gabapentin [236].

GVHD individuals, non-responders to corticosteroids, presented clinical improvement in the skin inflammation in a proportion of 12 out of the 20 patients with human chorionic gonadotropin (hCG) [237]. They also had a significant increase in the IDO mRNA expression in the PBMCs, and in the IL-10 expression in the blood serum [237]. The thought mechanism of action was to stimulate the IDO mediated immunotolerance, similar to the mother/fetus coexistence [130].

Although useful in the conception and to revert the immune-mediated diseases, the TKP may be deleterious in neoplastic diseases, where it can allow the tumor growth, by inducing IFN secretion and, once activating IDO, suppressing the immune response against the tumor [238]. Following this concept, the first human clinical trial aimed to block the TKP and works as an anti-cancer therapy was recently published [239]. The study investigated whether, Epacadostat, an IDO inhibitor, given orally, would be well tolerated and capable of slowing the growth of tumors in 52 refractory cancer patients,
by removing the immune tolerance against those tumors. The drug, at doses of 200 mg/day, reduced the kynurenine plasma levels, indicating the reduction in the tryptophan degradation. The mean treatment time was 52 (from 7 to 284) days, and the daily doses ranged from 43 to 1400 mg. The side effects included fatigue, nausea, and pain, among others. However, no plasma changes on C-reactive protein or in the levels of the interleukins tested were observed [239]. Further conclusions are limited due to the small number and heterogeneity of the clinical cases.

Therefore, the observations of the present data on interventions in the TKP shows that it has a double direction avenue, where the inhibition at specific steps as KMO activity and quinolinic acid formation has beneficial effects on neuropathic pain and neurodegenerative disorders. On the other hand, enhancing the activity of IDO, what ultimately is capable of inhibiting pro-inflammatory cytokines and reducing the inflammatory process. How, and at which step those events can find a conciliatory mechanism to diminish the chronic pain and neurologic symptoms of SS patients but also prevent the chronic inflammatory reactions will be subject of further investigation.

7. Future perspectives

IFN-γ triggers the deviation of the tryptophan to the TKP pathway in SS, possibly contributing to depression and pain through its action on particular organs, like the hippocampus. However, this signaling pathway is relevant to interrupt the vicious cycle of the inflammation and chronic damage of the involved tissues structure and function, including the PNS, CNS, SG, joints and the LFU. Suppression of the APCs and production of anti-inflammatory cytokines in the exocrine glands and other target tissues are the potential
benefits of the TKP actions. Therefore, IDO and kynurenine metabolites higher expression in SS indicates so far a reactive process to modulate the large, redundant and competitive mechanism of inflammation induced by other pathways. Better ways to access the CNS and PNS organs by imaging analysis and to monitor the local activity of the TKP in SS involved organs, either glandular or not, would allow increase insights about the physiopathology of this signaling pathway in SS, how it interferes with exocrine secretion and the strategies to improve it. More effective treatments and enhanced quality of life for SS patients will be the potential benefits from this knowledge.

8. Conclusions

The glandular and EGM of SS are not exclusively inflammatory, but also involves a neuroimmunendocrine network, where the TKP takes part. The activity of the TKP is hard to track because of the delicate methods to trace the metabolites of this pathway in the CNS. A better understanding of this relationship between the SS physiopathology and the TKP in the CNS and the target tissues may help to clarify the discrepancies among signs and symptoms and the neurological manifestations. This knowledge can improve the therapy for SS.

Figure and Tables Legends

**Box 1.** Summary of evidences linking Sjögren’s syndrome (SS) and the tryptophan/kynurenine signaling pathway (TKP).
Table 1. Neurologic manifestations of Sjögren’s syndrome (SS) showing the study design, number of patients involved and clinical changes observed.

Table 2. Central nervous system (CNS) structures altered in SS and sensitive to tryptophan/kynurenine pathway (TKP) modulation

Table 3. Potential therapeutic strategies to modulate pain and inflammation targeting the tryptophan/kynurenine pathway (TKP).

Figure 1. A schematic model, showing the interrelationships among the autonomic nervous system dysfunction, pathological pain and chronic inflammation in Sjögren’s syndrome (SS).

Figure 2. The tryptophan/kynurenine pathway (TKP) and their resulting metabolites.

Figure 3. The tryptophan/kynurenine pathway (TKP) and its implications in the Sjögren’s syndrome physiopathology.
Box 1. Summary of evidences linking Sjögren’s syndrome and the tryptophan/kynurenine pathway (TKP).

- Association among chronic inflammation, pain and neuropathic disorders
- Triggered by Interferon
- Modulated by sex hormones
- Tryptophan deprivation induces dry eye
- Sjögren’s syndrome and salivary gland aggression leads to increased expression of kynurenine
Back Matter

- **Supplementary Materials:** not applicable

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**Conflicts of Interest:** The authors declare no conflict of interest.
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