

# Social plasticity in the fish brain: Neuroscientific and ethological aspects

Karen Maruska<sup>1\*\*</sup>, Marta C. Soares<sup>2\*</sup>, Monica Lima-Maximino<sup>3,4</sup>, Diógenes Henrique de Siqueira-Silva<sup>5,6</sup>, Caio Maximino<sup>4,5</sup>✦

<sup>1</sup> Department of Biological Sciences, Louisiana State University, Baton Rouge, USA

<sup>2</sup> Centro de Investigação em Biodiversidade e Recursos Genéticos – CIBIO, Universidade do Porto, Vairão, Portugal

<sup>3</sup> Laboratório de Biofísica e Neurofarmacologia, Universidade do Estado do Pará, Campus VIII, Marabá, Brazil

<sup>4</sup> Grupo de Pesquisas em Neuropsicofarmacologia e Psicopatologia Experimental

<sup>5</sup> Laboratório de Neurociências e Comportamento “Frederico Guilherme Graeff”, Universidade Federal do Sul e Sudeste do Pará, Marabá, Brazil

<sup>6</sup> Grupo de Estudos em Reprodução de Peixes Amazônicos, Universidade Federal do Sul e Sudeste do Pará, Marabá, Brazil

✦ Author to whom correspondence should be addressed

Caio Maximino

KM and MCS contributed equally to this paper as co-first authors

22 Laboratório de Neurociências e Comportamento – Instituto de Estudos em Saúde e Biológicas,  
23 Universidade Federal do Sul e Sudeste do Pará, Unidade III  
24 +55-94-2101-7161  
25 Av. dos Ipês, S/N, s/ CEP, Bairro Cidade Jardim, Marabá/PA, Brazil  
26 cmaximino@unifesspa.edu.br

27

28

## Abstract

Social plasticity, defined as the ability to adaptively change the expression of social behavior according to previous experience and to social context, is a key ecological performance trait that should be viewed as crucial for Darwinian fitness. The neural mechanisms for social plasticity are poorly understood, in part due to skewed reliance on rodent models. Fish model organisms are relevant in the field of social plasticity for at least two reasons: first, the diversity of social organization among fish species is staggering, increasing the breadth of evolutionary relevant questions that can be asked. Second, that diversity also suggests translational relevance, since it is more likely that “core” mechanisms of social plasticity are discovered by appealing to a wider variety of social arrangements than relying on a single species. We analyze examples of social plasticity across fish species with different social organizations, concluding that a “core” mechanism is the initiation of behavioral shifts through the modulation of a conserved “social decision-making network”, along with other relevant brain regions, by monoamines, neuropeptides, and steroid hormones, as well as the consolidation of these shifts via neurogenomic adjustments and regulation of the expression of plasticity molecules.

**Keywords:** Brain plasticity; cichlids; cleanerfish; social plasticity; social decision making network; zebrafish.

# 1. Introduction

The ability to adapt to changing social environments is a crucial characteristic of biological systems; social plasticity, defined “as the ability to adaptively change the expression of social behavior according to previous experience and to social context” (Teles et al. 2016) is a key ecological performance trait that should be viewed as crucial for Darwinian fitness (Taborsky and Oliveira 2012). The neural mechanisms for social plasticity are poorly understood, in part due to over-reliance on rodent models and examination of only a single sex within a species. In principle, these mechanisms involve the modulation of the activity of the social decision-making brain network (O’Connell and Hofmann 2012a) and other relevant brain nuclei on the short term - by neuromodulators (e.g., neuropeptides and monoamines) and hormones (eg., sex steroids and glucocorticoids) - and on the long term - by modulating gene expression patterns across the network (Oliveira, 2009; Taborsky and Oliveira 2012; Cardoso et al. 2015).

The present review summarizes ongoing research on social plasticity in the fish brain. Fish model organisms are relevant and an ideal vertebrate group to examine social plasticity due to at least two reasons: first, the diversity of social organization between fish species is staggering, increasing the breadth of evolutionary relevant questions that can be addressed. Second, that diversity also suggests translational relevance, since it is more likely that “core” mechanisms of social plasticity are discovered by appealing to a wider variety of social arrangements than relying on a single species. Both issues are discussed in Sections 2 to 4. We proceed by discussing

research on social and reproductive status as triggers for plasticity, and examining sensory and cognitive aspects of social plasticity in fishes. Finally, the specific example of cleanerfish, which exhibit mutualism but is currently under-studied in behavioral and molecular neuroscience, is analyzed. We hope to demonstrate that, from cichlids to poeciliids, from zebrafish to cleanerfish, the variety of social interaction networks observed among fishes is of interest to both evolutionary neuroscientists and behavioral neuroscientists interested in describing the core mechanisms regulating and driving social plasticity.

## 2. Fish as models in the neurosciences

While a great deal of work on social plasticity of the brain has been made using rodents, this strict focus risks missing opportunities to answer questions on the evolutionary neuroscience of social plasticity (Panksepp et al., 2002; Striedter et al., 2014; Taborsky and Oliveira, 2012) , as well as missing the rich variety of social environments that fish occupy (Keenleyside, 1979) . The use of rodents is certainly relevant due to the phylogenetic position occupied by these animals in relation to humans, but can overlook important variants of social organization that exist in fishes. Indeed, more than 30,000 species of fishes are estimated to occupy both marine and freshwater habitats (<http://www.coml.org/>) (Nelson et al., 2016) . These species occupy different social niches (Section 3), and include single fishes with solitary living and territorial mosaics, animals which establish male-female pairs, animals which live in

92 small groups, animals which form shoals and schools, and many other varieties of  
93 social organizations (Keenleyside, 1979). Identifying how these different social societies  
94 impact brain and behavior, and how changing social environments alter these domains,  
95 is an important question to be exploited in the field of social plasticity (Figure 1).



**Daffodil cichlid (*Neolamprologus pulcher* Trewavas & Poll 1952)**

- Cooperative breeder, with a dominant breeding pair and non-breeding helpers
- Model organism to study cooperative behavior



**Zebrafish (*Danio rerio* Hamilton 1822)**

- Lives in small groups, shoaling preferentially with conspecifics, but also including small heterospecifics
- Model organism in genetics and developmental biology
- Lots of genomic and proteomic tools



**Bluestreak cleaner wrasse (*Labroides dimidiatus* Valenciennes 1839)**

- Lives in small groups in cleaning stations, where it interacts with clients
- Model organism to study cooperative behavior
- High level of cognitive plasticity



**Panuco swordtail (*Xiphophorus nigrens* Rosen 1960)**

- Low sexual conflict in relation to other poeciliids
- Sexual selection by females
- Cognitive and neural plasticity due to sexual conflict



**African cichlid (*Astatotilapia burtoni* Günther 1894)**

- Model organism to study behavior and physiology of cichlids
- Reversible male social roles (dominant/subordinate)
- Social plasticity of sensory modalities

*Figure 1: Selected fish species used in behavioral neuroscience and ethology and their social organizations. Different species occupy different social niches, showing potential to investigate one aspect of social plasticity.*

It has been suggested (Gerlai, 2014; Kas et al., 2011) that increasing the breadth of species used in biomedical research “can robustly enhance our ability to identify biological features and mechanisms that are relevant to the studied behavioral phenomena” (Gerlai, 2014, p. 55) . The choice of species and model organisms, in the case of the neurosciences, is usually guided by practical advantages (including fertility, throughput, and developmental speed), the existence of well-established research communities and data availability (including genomic and transcriptomic data), and the amenability to genetic manipulations and relative simplicity of the nervous system (Maximino et al., 2015) . In addition to using well-established model organisms, behavioral neuroscience can benefit from focusing on other, carefully chosen species to amplify the field of discovery and increase translational relevance (Gerlai, 2014; Hall et al., 2014; Maximino et al., 2015) . In the context of evolutionary neuroscience, Striedter et al. (2014) used the term “reference species” to signify “carefully selected species from phylogenetically widely spaced vertebrate and invertebrate groups”. These reference species are not “models for some other species, but [...] a basis for comparisons that may reveal both similarities and differences” (Striedter et al., 2014, p. 5) , ultimately increasing translational relevance by “allowing one to identify common features across species [that are likely to be] shared not just among the studied laboratory organisms but also with humans” (Gerlai, 2014, p. 55) . In fact, historically, many of the most significant discoveries allowing the field of basic neuroscience to advance were made in diverse taxa ranging from invertebrates (e.g. squid, *Aplysia*, crustaceans) to vertebrates (e.g. fishes, frogs, mammals). A similar approach to other fields of neuroscience can benefit the field by comparing taxa to infer how variations in



one domain (e.g., gene expression, connectivity, activation patterns) relates to variation in behavior; of relevance to the question of social plasticity, fish species can be used to better understand how variations in social environment impact relates to variations in behavior and brain structure and function (Soares et al., 2018) .

One advantage of using fishes to better understand social plasticity of the vertebrate brain is the ability to study species in a naturalistic context; something rarely achieved in other social vertebrate models such as rodents. Studying fish social behavior in the wild or in laboratory settings that include salient sensory, environmental, and social factors is crucial for discovering accurate neurobiological mechanisms as well as the selective pressures leading to evolutionary adaptations. Because many aspects of neural function are conserved, investigations in amenable systems like fishes have and will continue to provide valuable insights for biomedical applications. Recent advances in genomic and genetic tools for different fish species also increases their utility in social neuroscience. Thus, by abiding to Krogh's principle that for most biological problems, there exists a species that is ideally suited (Krogh, 1929), the diversity of fish species becomes a cornucopia of possibilities for knowledge advances. This includes promise for better understanding mechanisms of and treatments for reproductive, endocrine, neuroendocrine, and neurological disorders in humans that can be influenced by the social environment.

Fish are also excellent reference species to study social plasticity because their brains are admirably plastic. Neural plasticity can involve structural changes, such as alterations in cell population size or connectivity between different nuclei by changing axonal growth and survival or dendritic synaptic connections (Cline, 2001). In contrast to

mammals, in which neurogenesis is very limited in adulthood, the formation of new neurons continues throughout the fish's entire life (Zupanc and Sîrbulescu, 2011). Continuing expression of growth-associated protein-43 (GAP-43), a marker of axonogenesis, in the brains of coho salmon *Oncorhynchus kisutch* (Ebbeson and Braithwaite, 2012), suggests that connectivity also changes throughout adulthood. Moreover, heightened plasticity is observed at critical periods of development, such as the preparations associated with leaving fresh water and migrating to the ocean in salmon such as *Salmo salar* and *Onchorhynchus* sp. In these species, important behavioral preparations in this transition period include olfactory imprinting on their natal stream and switching from territorial to schooling behavior; these changes are accompanied by sequential changes in structure reorganization, and increased neuronal differentiation, neurogenesis, axonogenesis, and synaptogenesis (reviewed in Ebbeson and Braithwaite, 2012). Finally, the high adult neurogenesis also begets increased neuronal regeneration: after injury of nervous tissue, a massive surge of apoptotic cell death occurs at the lesion site, followed by a marked increase in cell proliferation and neurogenesis. There is also evidence for structural reorganization and neurogenesis related to the social environment, including social isolation, in several fish species (Sorensen et al., 2007; Maruska et al., 2012; Dunlap et al., 2013). The exquisite plasticity of the fish brain suggests that this group could represent interesting reference species and/or model organisms in the study of the social plasticity of the brain.

Further, fishes are becoming valuable models to study impacts of anthropogenic noise, pollution, and climate change on sensory and brain function (Ashur et al., 2017; Braun, 2015; Fisher and Oleksiak, 2007; Lai et al., 2017), with important consequences

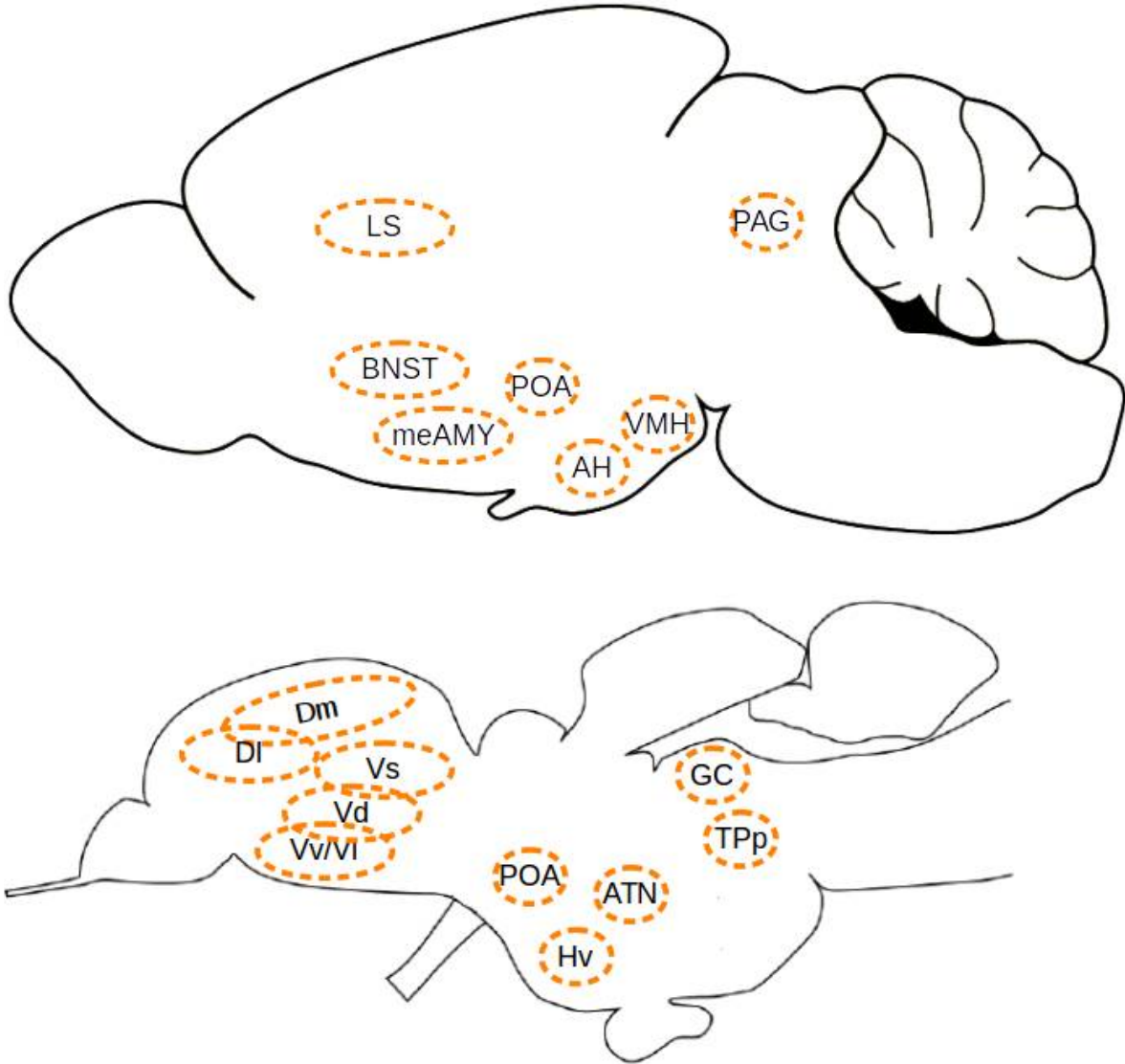
for management and conservation. Sensory cues play an integral role in the daily lives and survival of marine and freshwater organisms, including influences on homing, settlement, predator detection and evasion, foraging, conspecific social interactions, and reproductive interactions. Ocean acidification, as a consequence of climate change and pollution, directly affects the performance of sensory systems of marine organisms, including chemosensation, acoustic detection, and vision (review in Ashur et al., 2017). Sound pollution (anthropogenic noise) disrupts both the octavolateralis systems of fishes (which include the vestibular, auditory, lateral line and electrosensory systems) and the sonic environment that provide ecological and ethological cues for fish behavior (review in Braun, 2015). These impacts on systems which show critical social plasticity are beginning to be understood, and studying these in diverse fish species have important ecological and economic consequences, particularly in identifying how fishes may or may not be able to adapt to a changing world.

In addition to increasing possibilities to understand social plasticity, fish models can also increase translational relevance for research in social behavior (Oliveira, 2009; Soares et al., 2018) . This is relevant, because human sociality is crucial for mental health, and social stressors represent a very important source of suffering that can lead to mental disorders (Soares et al., 2018) . Moreover, alterations in social behavior are also observed in different disorders, including social anxiety disorder, autism, Williams syndrome, reactive attachment disorder, and disinhibited attachment disorder (Kennedy and Adolphs, 2012) . The use of relevant model organisms and reference species, including fish, is important to understand the value and expression of social behavior and the role of genes X (social) environment interactions in shaping mental disorders

(Huhman, 2006; Lim et al., 2005; McOmish et al., 2014; Soares et al., 2018) .

### 3. The social brain of fishes

Across vertebrates, social behavior is controlled by a survival circuit involved in both reward and sociality, termed the “social decision making network” (SDMN; Figure 2). This network involves a series of brain regions that regulate and integrate responses to salient stimuli (including social and non-social stimuli) (O’Connell and Hofmann 2011). The SDMN involves, in mammals, the lateral septum, extended medial amygdala, preoptic area/paraventricular nucleus (POA/PVN), anterior hypothalamus, ventromedial hypothalamus, and periaqueductal gray area (Figure 2). This network is involved in multiple forms of social behavior, including sexual behavior and courtship, aggression, and parental care, and its nodes are reciprocally and massively connected (Goodson 2005; O’Connell and Hofmann, 2011). By definition, these nodes also express sex hormone receptors (Forlano and Bass, 2011; O’Connell and Hofmann, 2011, 2012).



*Figure 2: The social decision-making network (SDMN) of rodents (top) and fish (bottom). Abbreviations: LS – lateral septum; BNST – bed nucleus of the stria terminalis; meAMY – medial amygdala; VMH – ventromedial hypothalamus; AH – anterior hypothalamus; PAG – periaqueductal gray area; Dm – medial zone of the dorsal telencephalon; DI – lateral zone of the dorsal telencephalon; Vs – supracommissural zone of the ventral telencephalon; Vd – dorsal zone of the ventral telencephalon; Vv – ventral zone of the ventral telencephalon; VI – lateral zone of the ventral telencephalon; POA – preoptic area; Hv – ventral hypothalamus; ATN – anterior tuberal nucleus; TPp – posterior tuberculum*

Nodes of the SDMN have been identified in teleost fish, although for many of them only partial homologies have been established and many remain undefined, debated, or controversial (cf. Soares et al. 2018 for a review; Goodson and Kingsbury, 2013). This is primarily due to the differences in forebrain development between teleosts (eversion) and tetrapods (inversion). While the SDMN represents a starting framework to study neural mechanisms of social behaviors, it is becoming increasingly clear that many other brain regions (e.g. raphe nuclei, habenula, reticular nuclei, and many others) are also involved in both receiving sensory inputs critical for decisions, as well as controlling the context-dependent behavioral outputs. Based on topology, hodology, and expression of receptors for sexual hormones, O'Connell and Hofmann (2011) originally proposed direct one-to-one homologs of the periaqueductal gray area (= griseum centrale of teleosts) and preoptic area (POA) in fishes and tetrapods, and the anterior (ATN) and ventral tuberal (VTN) nuclei as homologues in part for ventromedial hypothalamus (VMH) and anterior hypothalamus (AH), respectively. Homologues also have been proposed for the basolateral (pallial) amygdala (= dorsomedial telencephalon in part, Dm); and hippocampus (= dorsolateral telencephalon, DI). Subpallial structures in the SDMN probably represent partial homologies, with the ventral telencephalon (Vd, Vv, Vc) presenting a pallidal portion and a striatal portion that are equivalent to the pallidum and striatum (including a subpallial amygdala). Several more recent studies in zebrafish and electric fish have refined and redefined some of these homologies (Ganz et al., 2012; Harvey-Girard et al., 2013; Elliott et al., 2017, and we should recognize that the neuroanatomical homologies between fishes and tetrapods will continue to be updated as more functional studies are

performed. As we will see, functional and structural changes in these socially-relevant regions, as well as in the connectivity among them, is an important consequence of social plasticity in fish.

## 4. Social organization in fishes

Fishes form a unique group, being the most numerous and diverse of the vertebrates, dominating the aquatic environment by showing a remarkable panoply of behavioral characteristics and adaptations (Moyle and Cech, 2000). Fishes are still erroneously viewed as primitive and inflexible, controlled by fixed behavioral predispositions; however, the reality is very different. In the last three decades, evidence has contributed to repair these misconceptions, and reintroduced fishes as an “equal” group of vertebrates, rich in all sort of behaviors, even those considered as sophisticated or cognitively complex (Brown et al., 2006).

Fish occupy all aquatic niches. The level of diversification and adaptation of fishes is remarkable. Behavioral traits maximize adaptation to the environment, giving access to food, reproductive opportunities, or social aggregation (which includes the whole socialization domain). The social environment of fish may have fitness consequences, as it is both a source of wellbeing as well as of conflict. In general, we may organize fish social systems into 3 categories: i) solitary, ii) individualized social units, and iii) collective social assemblages (Keenleyside, 1979, Bshary et al., 2002). Fish species living in individualized social units, may be found as pairs, harems, or in territorial neighbouring mosaics (which is the case of many pomacentrids). In the case

of collective social systems, these may vary in size, from smaller group units to large schools (Keenleyside, 1979). These social structures or networks may be based on distinct arrays of associations or behaviors; for instance, some are related to feeding, others to defense against predators or conspecifics, to mating, or even to cooperation (which may aggregate several of these functions) (Krause et al., 2008).

The variable functions of these social networks challenge individuals in multiple ways, with variable associated costs and benefits for different types of social and non-social information. The structure of each network will determine the value of each information source, which will modulate the animals' capabilities in return. For example, the ability to recognize individuals within and outside the network and to gather information on relationships between group members affects the ability to make alliances, to participate in group activities, and to compete for access to food and mates (Croft et al., 2005).

There are some examples in the literature of solitary fishes, including some butterflyfishes, and pikes (*Esox lucius* and *Esox masquinongy*) - which are considered to be solitary and relatively sedentary carnivores showing little social interaction besides reproduction (Keenleyside, 1979). Most solitary fish studied so far are home-ranging, and not particularly territorial, but there are always exceptions (Keenleyside 1979). One interesting exception is *Betta splendens*, which show a marked territorial and aggressive behavior, especially in males (Simpson, 1968). Thus, even in solitary fish, social behaviors (agonistic encounters) are sometimes unavoidable.

Two marine families (Blenniidae and Pomacentridae) and one predominantly freshwater family (Salmonidae) are best representatives of fish living in territorial



mosaics - that is, a system in which the home range is subdivided in a mosaic of contiguous territories (Keenleyside, 1979). The mosaic of contiguous territory occupied by these animals is a flat, two-dimensional system, with territory occupancy being driven mainly by requirements of food and shelter: "Long-term utilization of benthic food can be assured by the mosaic system, each individual guarding its own resources. At the same time, thorough familiarity with escape routes and shelters within the territory reduces vulnerability to predators." (Keenleyside, 1979, p. 162). As a result, fish living in territorial mosaics have a highly fluctuating social environment as they are challenged for their territories by younger conspecifics.

Stable, long-term male-female pairs are rare among fishes (Keenleyside, 1979). Some cichlid species, such the species of *Cichla* genre (Kullander and Ferreira, 2006), appear to form pair bonds for up to several weeks, with both parents collaborating in raising the brood until the young fish disperse (Gross and Sargent, 1985), and many species of butterflyfishes (*Chaetodon*) have been observed to form monogamous male-female pairs for up to three years (Fricke, 1973). Interestingly, in monogamous convict cichlids (*Amatitlania nigrofasciata*), pair formation depends on arginine vasotocin (AVT) and isotocin (IT) (homologues of the mammalian vasopressin and oxytocin systems) (Oldfield and Hofmann, 2011).

Several species live in small groups with moderate to high complexity. In the wild, the model organism zebrafish (*Danio rerio*) live in small groups that form shoals, which usually include small heterospecifics (Suriyampola et al., 2015). Shoaling behavior has been exploited as a tool to study the neurobiology of social behavior in zebrafish (Soares et al., 2018). Within the shoal, dominant-subordinate relationships are

established, a model of social plasticity that has also been exploited successfully (cf. section 4.1, below). However, many other important grouping schemes are observed across fish species that form small groups. For example, the freshwater African cichlid *Neolamprologus pulcher* lives on the rocky substrata of Lake Tanganyika, where it forms small groups made up of a dominant breeding pair and 0-20 smaller non-breeding subordinates called helpers (Wong and Balshine, 2011). These helpers are organized into size-based dominant-subordinate hierarchies, reflecting queues for breeding status: when the dominant female dies, helper females take its place; helper males are more likely to disperse and take over a dominant position in other groups (Wong and Balshine, 2011). Importantly, both breeders and helpers defend the territory, do maintenance work (digging and removing debris), and care for the brood (Taborsky and Limberger, 1981). This system has been used to study the neural bases of cooperative behavior (e.g., Taborsky et al., 2013; Nyman et al., 2017; Kasper et al., 2018a, 2018b).

To navigate these complex social contexts, animals need a wide array of social skills (Soares, 2017; Soares et al., 2018a). For example, living in a territorial mosaic will demand of an individual high investments in defense and competitive skills, while being a part of extended family groups will also demand defense capabilities but mostly in fine synchrony with others in the group (Bshary et al., 2002). Thus, social organization in fish take many forms, some of which exhibit complex social strategies and tactics, demands of sharp social learning capacities, elevated levels of communication, and even deception (Krause and Ruxton, 2002). Among these, cooperation and the expression of cooperative strategies among fishes stands out in terms of sociality (Soares et al., 2018b). And fish do cooperate, perhaps in lower frequencies when

compared to birds and mammals (Balshine and Buston, 2008) but even so, this takes many forms: from the best-known conditional approach during predator inspection (Pitcher et al., 1986), to cooperative hunting (Bshary et al., 2006), cooperative breeding (Wong and Balshine, 2011), and finally to the notable cleaning mutualisms (Côté, 2000).

## 5. Social status and plasticity

### 5.1 Social status and plasticity of the brain and behavior

Dominance hierarchies are an integral part of the social structure in many animal societies. As such, an individual's position or rank in the population has profound effects on their reproductive potential, access to food and other resources, overall health, and survival (Sapolsky, 2005; Wingfield and Sapolsky, 2003). Fishes are champions of diverse social societies, with examples ranging from solitary living species to group-living species that exist in either constant or ephemeral (e.g. during breeding season) hierarchies. Because of their great diversity in social, reproductive, and parental strategies, fishes are powerful taxa to examine interactions between the brain and behavior.

Establishment of dominance hierarchies often leads to specialized social interactions and behaviors, resulting in each individual of the group occupying a specific rank in the population. This social position is constantly evaluated and reinforced by aggressive and reproductive interactions with other individuals of both higher and lower

rank. In zebrafish (*Danio rerio*), dominant–subordinate relationships occur both between males and between females (Paull et al., 2010), and dominant males and females are more aggressive and bolder (Paull et al., 2010; Dahlbom et al., 2011). Moreover, dominance is associated with higher gonadosomatic indices and higher mRNA levels of the steroid receptors AR (in males) and ESR1 (in females) in the gonads (Filby et al., 2010). These gonad differences result in a greater total reproductive success in males but not in females, but dominant females sire more offspring with the dominant male (Paull et al., 2010).

In zebrafish, much of the work on social status has been done targeting stress and arousal pathways. The establishment of a hierarchy increases cortisol levels in both dominants and subordinates, but no differences are found between dominant and subordinate fish after dominance (Pavlidis et al., 2011). Pavlidis et al. (2011) found that dominant zebrafish show increased mRNA levels of the mineralocorticoid receptor in the brain, but no changes were observed in either corticotropin-releasing factor (CRF) or the glucocorticoid receptor (GR). Filby et al. (2010), however, found that subordinate zebrafish had higher cortisol levels in the first day of a dominance interaction, an effect which, consistent with Pavlidis et al. (2011), disappears in the fifth day. Lower telencephalic mRNA levels of neuropeptide Y (NPY), CRF, and GR are also observed, in the first day of a dominance interaction, in subordinates (Filby et al., 2010). As a result of this hypocortisolemic profile, male subordinate zebrafish had higher splenic levels of interleukin 1 $\beta$  and interferon gamma, while female subordinates had lower levels of these cytokines, as well as of tumor necrosis factor  $\alpha$  (TNF- $\alpha$ ) and interferon 1 (Filby et al., 2010). Dominant zebrafish showed higher expression of tyrosine

hydroxylase 2 (TH2) and histidine decarboxylase (HDC), while subordinates had higher mRNA levels of AVT in SUB, while catechol-O-methyltransferase (COMT) mRNA levels were down-regulated in both DOM and SUB compared to control fish. In addition, mRNA levels of hypocretin/orexin (Hcr) were up-regulated in dominant compared to subordinate and control males (Pavlidis et al 2010). Thus, a participation of catecholaminergic and peptide systems associated with arousal is associated with social status in zebrafish. However, social status was not associated with dopamine levels in the forebrain or hindbrain in zebrafish, although serotonin activity was higher in the hindbrain of subordinate animals (Dahlbom et al. 2012). Finally, differences in the AVT system have also been observed, as dominant zebrafish express AVT in 1-3 pairs of large cells in the magnocellular preoptic area, while subordinate individuals express AVT in 7–11 pairs of small cells in the parvocellular preoptic area (Larson et al. 2006).

Cichlid fishes of the African (e.g. Haplochromines, *Oreochromis* spp.) and Neotropical (e.g. *Cichlasoma* spp.) clades are ideally-suited to address questions related to the interactions between neural function and social status for several reasons. They are socially diverse, easy to manipulate, show remarkable plasticity in behavior, physiology, and brain function, and have evolved the ability to not only assess their social environment via multiple sensory channels, but also to be deceptive and determine both their own position and that of others within the hierarchy (Grosenick et al., 2007; Maruska and Fernald, 2018). Thus, cichlids display a level of social plasticity that rivals or exceeds that of many other vertebrates. In fact, they have already revealed many important discoveries about how the brain regulates and is influenced by social interactions at levels from behavior to molecular mechanisms.

There has been considerable focus on social status-specific differences in the brain of *Astatotilapia burtoni* (formerly *Haplochromis*). For example, cell proliferation (Maruska et al., 2012), neural activation in decision centers (revealed by markers such as immediate early genes), distribution, abundance, or activation of cells expressing neuromodulatory substances or their receptors (Loveland et al., 2014; Maruska et al., 2013b; O'Connell and Hofmann, 2012b; Renn et al., 2008), and aspects of the stress response system (Carpenter et al., 2014; Chen and Fernald, 2008) can all differ between dominant and subordinate males. These factors are also well known to modulate the expression of diverse social behaviors in fishes as well as other vertebrates, including species like rodents and primates that are closer in phylogenetic position to humans. Similar types of social status differences are also observed in other fish species [e.g. zebrafish, other cichlids, electric fishes, salmonids, and others; (Gilmour et al., 2005; Maruska, 2014; Miller et al., 2017; Perrone and Silva, 2018; Teles et al., 2016)], and in some cases result in a change in an individuals' sex (male to female, or vice versa) that is accompanied by plasticity in neural circuits (Black et al., 2005; Semsar et al., 2001; Todd et al., 2018). These examples support the broad relevance of fishes for addressing questions related to how an animal's social rank impacts their brain and reproductive fitness, health, and survival. Thus, by taking advantage of controlled status transitions and natural dominance hierarchies, fishes have and will continue to reveal insights on neural plasticity that can inform studies in other taxa.

One interesting advance in that field comes from the induction of changes in social status in zebrafish (Teles et al. 2016). Four social phenotypes were

experimentally induced: Winners and Losers of a real-opponent interaction; Mirror-fighters, that fight their own image in a mirror and thus do not experience a change in social status despite the expression of aggressive behavior; and non-interacting fish. By analyzing the expression of plasticity genes (*wnt3*, *neurod*, *npas4*, *bdnf*, and *nlgn1* and *nlgn2*) in regions of the SDMN, authors were able to identify markers of social plasticity that are associated with social status changes: winners were characterized by an increase of the expression of neurogenesis genes (*wnt3* and *neurod*) in Dm, and of neuroligin genes in Vv and Vs; and losers were characterized by an increase of the expression of *bdnf* in DI and of *wnt3* in Vv, and by a decrease in the expression of *nlgn2* in Vs (Teles et al. 2016). These results suggest the participation of mechanisms of neural plasticity in the establishment of social hierarchies in zebrafish.

In *A. burtoni*, establishment of dominant-subordinate hierarchy in females induce neurotranscriptomic differences across ranks. Using cDNA microarray, Renn et al. (2016) found several hormonal and neuropeptide genes showed increased expression in dominant females, including prolactin, AVT, brain aromatase, and glycoprotein alpha polypeptide subunit, a necessary precursor step in the production of active gonadotropin-releasing hormone (GnRH), luteinizing hormone, follicle-stimulating hormone, and thyrotropin. Interestingly, prolactin, AVT, and glycoprotein alpha polypeptide subunit are also upregulated in the brains of male dominant *A. burtoni* (Renn et al. 2008). Again, neuroplasticity genes were also found to be differentially regulated by social plasticity: FK506-binding protein 1, cell cycle associated protein 1, neuromodulin, and dynamin-1 were up-regulated in dominant females, while voltage-dependent N-type calcium channel subunit alpha-1B was expressed at higher levels in

subordinate females (Renn et al. 2016). In males, similarly, neuromodulin, synaptophysin, Programmed cell death protein 4, and Schwannomin interacting protein 1 are upregulated in the brain of dominants (Renn et al. 2008).

## 5.2. Social status and sensory plasticity

Studies in fishes have also revealed important neural mechanisms involved in sensory plasticity. Because the ability to assess the social environment is critical, particularly for species living in dominance hierarchies, conspecific communication and sensory perception is of paramount importance. For example, social rank or alternative reproductive phenotypes (e.g. nesting vs satellite/sneaker males) within a species are often associated with differences in the ability to detect auditory, olfactory, and visual information that is necessary for their status-specific behaviors such as reproduction, territory defense, feeding, and growth. In *A. burtoni*, dominant males have higher levels of modulatory receptors (e.g. steroid receptors) in the ear (Maruska and Fernald, 2010b) and olfactory bulbs (Maruska and Fernald, 2010c), as well as a greater response of the olfactory epithelium to food-related odorants (Nikonov et al., 2017) compared to subordinate males. This may facilitate detection of acoustic social signals and prey/food to support their territorial status, which comes with an associated trade-off of reduced feeding time and growth (Hofmann et al., 1999). Further, conspicuous dominant males show an increased startle response probability compared to less conspicuous subordinate males, possibly mediated by serotonin at the Mauthner



neurons, that may allow them to better escape from predators (Neumeister et al., 2010; Whitaker et al., 2011). In plainfin midshipman fish, in which males are either large nesting type I or small satellite type II, there are status-dependent differences in both the vocal and auditory systems that are crucial to the reproductive fitness of this species that relies heavily on acoustic signaling (Forlano et al., 2016; Maruska and Sisneros, 2015). Sensory plasticity can be modulated both at peripheral sensory structures (i.e. ear, olfactory epithelium, retina), as well as centrally in the brain, and in most cases is controlled by plasticity in signaling of neuromodulatory molecules such as steroids, neuropeptides, and biogenic amines. Social communication in fishes is often multisensory, but also depends on the reproductive strategies employed by the species, and the habitat and environmental conditions where they live, which can dictate the transmission properties and effectiveness of information sent via different sensory channels. Because modulation of sensory systems is common across vertebrates, including in humans, the diversity of sensory dependence seen in fishes (i.e. species that rely on multisensory information vs those relying primarily on a single sense to reproduce) provides unique opportunities to uncover basic mechanisms of sensory function. The conservation of sensory structures across taxa allows research in fishes to also reveal how perception of social information can be modulated by neurochemicals that change with their social rank. As discussed above, fishes are becoming valuable models to study impacts of anthropogenic noise, pollution, and climate change on sensory and brain function, opening opportunities to study the interaction of those factors with ecologically relevant sensory plasticity.

473

474 

### 5.3. Is the establishment of social rank inherently stressful?

475 Social stressors are powerful activators of the hypothalamus-pituitary-adrenal  
476 (interrenal) axis in many species, and in rodents is even used as a model for depression  
477 (Beery and Kaufer 2015). Initial studies on rodents increased the comprehension  
478 regarding how stressor type, timing, and other factors affect physiology and behavior.  
479 The animal's social status affects its access to feeding, mates, and shelter, and different  
480 studies showed the consequences of rank-related stress to the physiology and health of  
481 dominant and subordinate individuals (Francis et al., 1993; Qvarnström and Forsgren,  
482 1998; Valdimarsson & Metcalfe, 2001; Iwata et al., 2008). Models for social stress in  
483 zebrafish revealed that dominant males and females are more aggressive and show  
484 higher growth than subordinate animals, which show a higher brain expression of genes  
485 associated with the activation of the hypothalamus-pituitary-interrenal (HPI) axis  
486 (including *crh*, *npv*, and *gr*), higher cortisol levels, peripheral expression of pro-  
487 inflammatory cytokines, and compromised reproductive activity immediately after  
488 hierarchy establishment (Filby et al. 2010)

489 Higher cortisol levels are not always observed in subordinates in relation to  
490 dominants, however. Pavlidis and collaborators (2011) also established a model for  
491 social rank stress in zebrafish, forcing a dyadic interaction for five days - after which  
492 dominance was successfully established. After the establishment of dominance, both  
493 subordinates and dominant males showed higher whole-trunk cortisol concentrations

than control animals; no differences were observed, however, between dominant and subordinate animals. Teles et al. (2016) showed a significant increase in cortisol in winner animals (that is, animals which win a 30 min fight against a conspecific) and in animals fighting against a mirror (that is, animals which display aggression without establishing dominance), but not in loser animals. These results suggest that, at the initiation of a social rank interaction, cortisol is more associated with aggression levels than with classical stress responses (i.e., behavioral inhibition, hypoaggressiveness, etc). However, these relationships between cortisol and stress or other behaviors like aggression are not always consistent across species. For example, the circulating cortisol levels in the cichlid *A. burtoni* are quite sensitive to experimental paradigms, with some studies showing higher levels in subordinate males, while others detect no differences between dominant and subordinates (Fox et al., 1997; Maruska 2015).

In rainbow trout subjected to stressful social interactions, dominant animals show higher aggressive behavior immediately after rank establishment, but only subordinate animals present elevated plasma cortisol levels associated with other signs of chronic stress, such as reduced feeding and reduced serotonergic activity in the brainstem (Sørensen et al. 2012). Moreover, subordinates had reduced proliferation of adult neural cells than controls, and neural cell proliferation was negatively correlated with the intensity of aggression received at the end of the social rank establishment (Sørensen et al. 2012). A study in the cichlid *A. burtoni* examined the behavioral consequences and neural activation patterns of repeated social defeat from the same aggressor and showed that individual males will switch between proactive and reactive coping behaviors over time, and each coping behavior is associated with distinct activation

patterns in the brain (e.g. reactive had greater activation in raphe nuclei, while proactive had greater activation in Dm, Vs, Vc, Vd, Vp, Vv, TPp, ATn) (Butler et al., 2018).

These fish data reaffirm the literature that was built on rodents, which show links between the neurobiology of stress and behavior, with social interactions sometimes acting as a stressor. In rodents, social stressors are used as models for depression, taking into consideration how social plasticity - including social subordination, crowding, social isolation, and social instability - influences stress responses (Beery and Kaufer 2015). Fishes are increasingly used as models for better understanding human mental health disorders, many of which are associated with stress responses, such as anxiety, depression, and PTSD. For this translational approach to be effective and informative, studies in fishes are necessary because it is important to understand the evolution and conservation of neural networks underlying the behaviours typically displayed in these neurological conditions.

#### 5.4. Reproductive status and plasticity

One of the most important consequences of an individual's social status is their reproductive potential. Within a population, dominant individuals typically have an up-regulated reproductive axis and more mating opportunities compared to subordinate individuals. Studies in the African cichlid *Astatotilapia burtoni* in particular have revealed important insights on how social rank impacts the reproductive brain [see (Fernald and Maruska, 2012; Maruska, 2014; Maruska and Fernald, 2014; Maruska and Fernald,

2018) for reviews]. In this species, males form hierarchies in which a small percentage of dominant individuals hold territories that they aggressively defend from rivals and use to court and spawn with females. Subordinate males comprise the majority of the population, but lack territories and therefore have minimal chances to reproduce. Their shallow habitats along the shores of Lake Tanganyika, however, are dynamic and there are frequent opportunities for males to rise or fall in social rank, thereby switching between subordinate and dominant status. These social transitions are associated with many dramatic changes in the brain and sensory structures that allow them to better adapt to their status-specific lifestyles.

The brain controls both the activity of the reproductive axis, as well as the expression of behaviors and must be inherently plastic to accommodate social change. In *A. burtoni*, dominant males have larger gonadotropin-releasing hormone (GnRH1) neurons in the preoptic area with distinct cell and firing properties (Davis and Fernald, 1990; Ma et al., 2015; Maruska and Fernald, 2013) compared to subordinate males, which leads to increased pituitary and testes activity to promote greater reproductive potential, higher circulating sex steroids, and increased territorial and reproductive behaviors [see (Maruska and Fernald, 2013; Maruska and Fernald, 2014; Maruska and Fernald, 2018) for reviews]. When subordinate males perceive an opportunity to gain a territory and rise in social rank, within minutes, they begin looking and behaving like dominant males (Burmeister et al., 2005; Maruska and Fernald, 2010a). This social ascent is also associated with numerous rapid (minutes to days) cellular and molecular changes from the brain to the testes (Maruska and Fernald, 2014; Maruska and Fernald, 2018), and similar changes occur on a slower timescale (days to weeks) when

males fall in rank (Maruska et al., 2013a; Maruska, 2015). Because changes in social position occur across invertebrate and vertebrate taxa, this rapid neural and behavioral plasticity in cichlids provides unique opportunities to test functional, mechanistic, and evolutionary hypotheses. The melanocortin system, for example, is a neuroendocrine system that links body pigmentation to fish behavior, being able to cause an impact on the evolution of the phenotypic and genetic diversity in fish (Seehausen & Schluter, 2004). In *A. burtoni*, yellow males were confirmed to be more aggressive than the blue counterparts. However, when they were submitted to the neuropeptide  $\alpha$ -melanocyte-stimulating hormone, only the blue morph-type individuals increased their aggressiveness (Dijkstra et al., 2018).

Important work on sexual conflict and social plasticity has been done in poeciliids (see Cummings, 2018, for a review), due to the variety of reproductive strategies and levels of sexual conflict found across species. Conflict levels vary across poeciliid species mainly as a result of variation in male mating systems, given that females of most species have long gestation periods and, being able to store sperm, are rarely sperm-limited; males, on the other hand, vary from systems dominated by male coercion (about half of the species) to systems that include both courtship and coercive tactics. The result is a sex ratio that is male-biased, with the optimal mating rate for males being several times a day, while for females less than one mating episode per month is optimal. Interesting species differences are observed, with guppy (*Poecilia reticulata* Peter 1859) and mosquitofish (*Gambusia affinis* Baird & Girard 1853) males attempting mating more than once per minute, and Panuco swordtail (*Xiphophorus nigrensis* Rosen 1960) males attempting mating between 0.25 and 5 times per minute

(Magurran and Maciás Garcia, 2000). Thus, the relative investment in a single reproductive event varies between males and females across poeciliid species, but females always allocate more resources towards foraging and avoiding male harassment (Houde 1997; Magurran 2011), while males allocate most of their resources towards mating attempts (Magurran and Seghers 1994).

The resulting selective pressures produced species in which males and females display sexual dimorphism in emotional behavior and cognition (Cummings, 2018). In guppies and mosquitofish, females that experience a high degree of sexual coercion exhibited a greater tendency to shoaling in the presence of male conspecifics relative to being presented with conspecific females, while females from the *X. hellerii* or *X. mayae* do not shoal in response to male conspecifics (Dadda, 2015). Moreover, females from species with high sexual conflict also occupy different habitats, inhabiting areas with higher predation risks to avoid male harassment (Croft et al. 2006; Darden and Croft 2008); as a result, females from species with high sexual conflict show less exploratory behavior and increased anxiety-like behavior (Cummings, 2018). Interestingly, in *G. affinis*, a species with high sexual conflict, males that showed less neophobia and anxiety performed better on a numerosity discrimination task, whereas females showed no relationship between exploration and learning performance (Etheredge et al., 2018). No differences were found between *G. affinis* males and females in learning performance, but high-performance learner males exhibited different behavioral attributes than high-performance learner females: while high-performance females showed higher mate choice, activity, and anxiety (key responses to social conflict), high-performance males do not show higher levels of any behavioral trait (Etheredge et al.,

2018).

In an interesting approach, Cummings and colleagues analyzed gene expression in whole brains or in the dorsolateral telencephalon (DI, a putative homologue of the hippocampus in teleosts) of females species with low conflict (*X. nigrensis*) or high conflict (*G. affinis*) that varied in levels of preference for courting vs. coercive male conspecifics (Cummings et al., 2008; Lynch et al., 2012; Wong et al., 2012; Wang et al., 2014). In *X. nigrensis*, the expression of *neuroligin-3* and *neuroserpin*, genes associated with synaptic plasticity, is positively associated with preference for courting males in both the whole-brain and DI samples; however, in *G. affinis*, these genes are negatively associated with preference for courting males in whole-brain samples. When *G. affinis* females are exposed to a courting heterospecific male (a large *Poecilia latipinna*), a positive correlation is observed between preference and *neuroserpin* brain levels, while a negative correlation is observed when females are exposed to a coercing *P. latipinna* (Cummings et al., 2008; Lynch et al., 2012; Wong et al., 2012; Wang et al., 2014). These results suggest that different expression of neuroplasticity genes across species is not due to species differences *per se*, but that females are responding more to male reproductive tactics than species identity. Finally, in *X. nigrensis*, females exposed to two courting males show a high correlation in the expression of *neuroligin-3* in regions that are associated with social decision making than females exposed to a courting and a coercive male, and little correlation is observed across these regions when females are exposed to two coercive males (Wong and Cummings, 2014), suggesting that interaction with courting phenotypes, more than coercive phenotypes, demand engagement of brain plasticity (Figure 3).



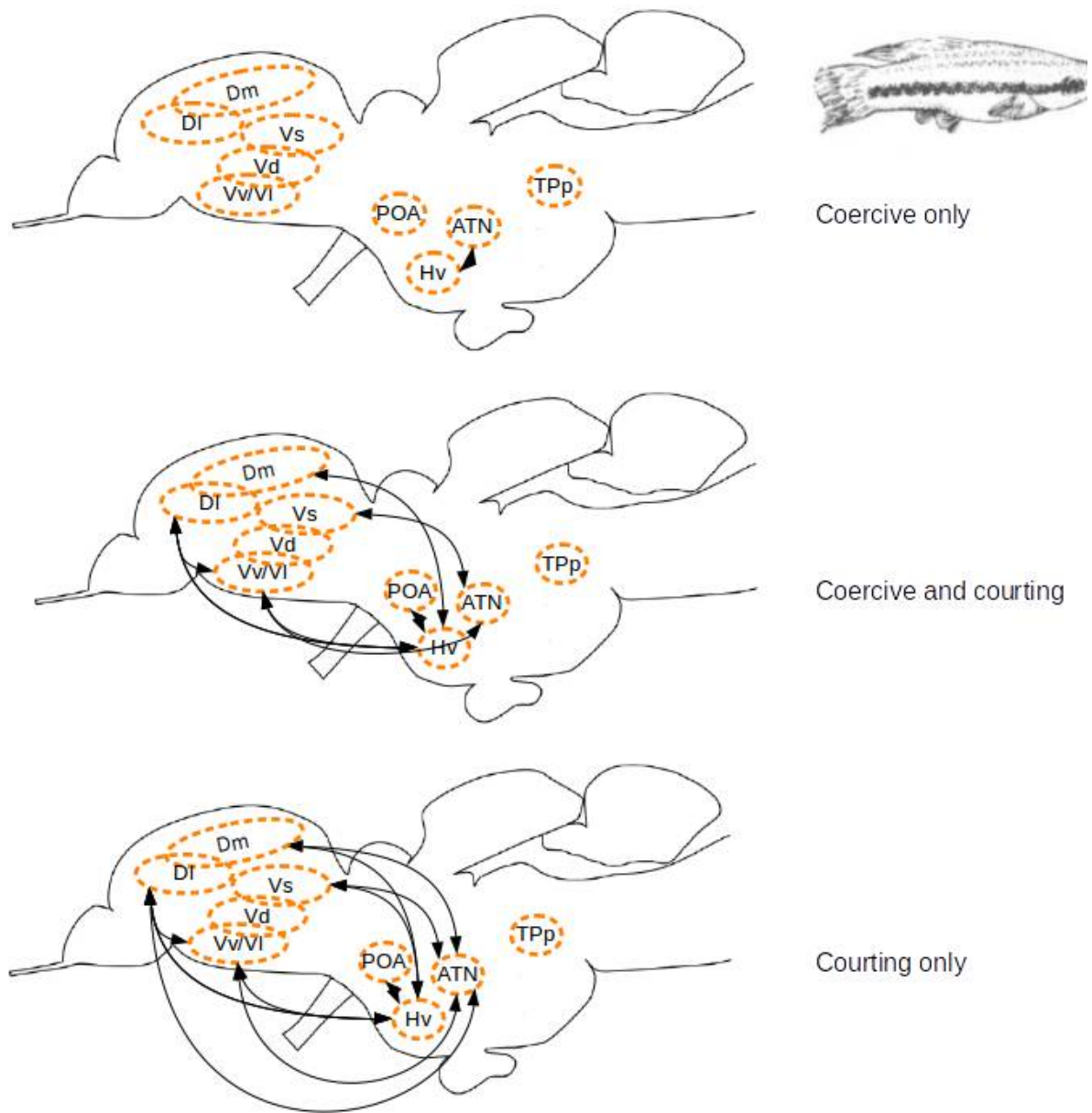


Figure 3: Sexual conflict and mate choice induce neuroplasticity between areas of the SDM in female *X. nigrensis*. Adapted from Cummings (2018)  
Abbreviations: Dm – medial zone of the dorsal telencephalon; DI – lateral zone of the dorsal telencephalon; Vs – supracommissural zone of the ventral telencephalon; Vd – dorsal zone of the ventral telencephalon; Vv – ventral zone of the ventral telencephalon; VI – lateral zone of the ventral telencephalon; POA – preoptic area; Hv – ventral hypothalamus; ATN – anterior tuberal nucleus; TPp – posterior tuberculum

630

The most remarkable expression of social plasticity, however, is the sex change, an adaptive strategy that has already been observed for 27 teleost families (Sadovy de Mitcheson and Liu, 2008). Most of them are marine species, such the Epinephelidae groupers *Epinephelus akaara* and *E. awoara*, both functional protogynous hermaphrodites (Liu et al., 2016), in which individuals first function as female and then males, and in the protandrous hermaphrodites Amphiprioninae, as in *Amphiprion melanopus* (Choi et al., 2016) and *A. ocellaris* (Khoo et al., 2018), in which animals are first males and then females. Theoretically, these strategies evolved to ensure a high mating success. In protogynous hermaphrodites, for example, larger males tend to be dominant, commonly monopolizing mating, either by defending spawning sites that females visit or by controlling a harem of females. Thus, if an individual acts as a female when small and as male after achieving a large size, it would have a greater offspring in comparison to a gonochoristic individual (Warner, 1984). Moreover, usually a protogynous male also controls the emergence of other males by aggressive dominance over females. The former tendency was demonstrated by Lo Nostro and Guerrero (1996) in the swamp eel *Synbranchus marmoratus*, in which the so-called primary male, which directly develops as male, were smaller (13 cm), while the larger ones (91 cm) were found to be secondary males, which develop from functional females. Contrary to the general findings in protogynous hermaphrodite species, in which female individuals are the majority, in this study the authors showed that most of the population (80%) was composed of secondary males. That occurrence might be a result of the survival strategy adopted by the species, since it inhabits streams and

swamps that periodically drought leaving the individual isolated. Thus, protogynous specimens are hypothetically more efficient to establish new colonies (Lo Nostro and Guerrero, 1996). The authors suggested that sex differentiation in this sex-changing species can be triggered by an initiating event in the brain, rather than directly on the gonad. Indeed, chronic administration of salmon GnRH analogue or the dopamine receptor antagonist domperidone induced sex change in female *S. marmoratus* and spermiation in males (Ravaglia et al 1997).

In the bluehead wrasse (*Thalassoma bifasciatum*), a diandric protogynous labrid, removal of a terminal phase male from a patch reef induces aggressive behavior in the largest female, targeted towards other females, as well as courtship towards smaller females, suggesting that one important control of sex change is dominance hierarchies (Warner and Swearer 1991). In this species, sex change is accompanied by increases in the expression of GnRH in the POA (Grober et al. 1991), and terminal phase individuals show higher GnRH expression than intermediate phase individuals (Grober and Bass 1991). The closely-related *T. duperrey* is also diandric, but females have a larger home range and do not always mate with the same male; the presence of smaller individuals promotes sex reversal and the presence of larger individuals inhibits sex reversal (Ross 1986). In this species, norepinephrine appears to stimulate gonadal sex reversal, while dopamine exerts inhibitory action on the initiation of sex reversal and serotonin inhibits both initiation and completion of sex reversal (Larson et al. 2003a). During sex change in females, monoamine metabolism changes in the SDMN, as well as in the locus coeruleus and raphe nucleus (Larson et al. 2003b); during the first week of sex change, when animals undergo behavioral changes, serotonergic activity in the

Dm is increased, while in the POA it is decreased (Larson et al. 2003b). Norepinephrinergic activity is decreased, and dopaminergic activity is increased, in the VMH; in the locus coeruleus, norepinephrinergic activity is increased, while in the raphe nucleus, there is a decrease in serotonergic activity at the time of behavioral sex reversal (Larson et al 2003b). Both studies suggest that behavioral sex reversal is under the control of serotonin in the raphe, while gonadal sex change is mediated by serotonergic effects on norepinephrine in the POA. In addition to sex-change, there are numerous other examples of alternative reproductive phenotypes across fish species, all showing plasticity in behavior, the brain, and physiology that can be useful for revealing core neural mechanisms across taxa (for reviews see Maruska et al, 2018; Oliveira et al., 2008).

## 6. Cognitive plasticity and social interactions

By definition, cognitive social plasticity refers to the ability to change patterns of cognition and behavior in the context of social engagement. This field, as with many others, was initially developed to tackle “human questions”, as part of social psychology, and later applied to other vertebrates including fish. Indeed, the diversity of fish species, social systems and cognitive abilities, together with the variety of adaptive contexts in which these species evolved, stirred the interest to generate new behavioral and cognitive paradigms with higher ecological validity (Hall et al., 2014). Recently, the use of fish in neurosciences, mostly zebrafish, and the development of new molecular tools, has created new possibilities and consequently, taken many more model fish species

into the spotlight. However, when thinking about social behavior and cognition, not many fish species or families come to mind, except for a few well studied examples as the cichlids (Grosnick and Fernald, 2007; Greenwood et al., 2008; Oldfield and Hofmann, 2011; Winberg et al., 2008) and the cleaner wrasses (Bshary and Côté, 2008; Soares 2017); with a few others coming as runner-up candidates, such as zebrafish (Oliveira 2013), poeciliids (Dugatkin, 1988, 1991), or cleaning gobies (Côté and Soares, 2011).

In this section, we will focus on the cleanerfish example, as it is possibly the best studied fish system in terms of complex social behavior, and includes both interspecific and conspecific components (Soares 2017). Cleaners are suitable candidates as model organisms to study the role of social plasticity in contextual social cognition and decision making; moreover, they are also putative models for more “organizational” questions, as those focusing on the evolutionary emergence of cleaning behavior (between species, Soares et al 2018a), ontogenetic changes in cleaning behavior (the case of facultative cleaners, Soares et al 2018a), and of alternative strategies (adaptive behavioral ecotypes, as with the Caribbean cleaning goby *Elacatinus prochilos*) (Côté and Soares 2011). Neuroendocrine trade-offs are assumed to have a crucial role on cleaners’ highly plastic social performance, enabling a successful navigation within and across challenges posed by the social environment (Soares 2017, Soares et al 2018ab). Recent research, while still at its start, has provided us with a few good potential candidates regarding the proximate mechanisms, such as the nonapeptide AVT, the stress steroid cortisol, and the monoamines dopamine and serotonin.

## 6.1. Social plasticity and the special case of the cleaner fish system

Living with others is a struggle. Fish, as other vertebrates, need to use previously acquired information and combine it with the current social environment, if they are to avoid dangerous costs. These include being eaten by a predator while inspecting it, or being expelled from the social group (Oliveira 2009). According to Oliveira (2012), the ability to appropriately adjust social behavior relies on social plasticity mechanisms that occur in different temporal scales: the first is associated with *changes* to life-history, with those being reversible (breeding vs non-breeding) or irreversible (juvenile cleaners vs adult non-cleaners); the second occurs *during* the same life-history stage, described as punctual or short term behavioral fluctuations (behavioral flexibility). Here, we prefer to envision these categories in a less fixed manner, using them to virtually differentiate the multiple domains of social plasticity, some working between irreversible life stages, others seasonally, and some punctually; the first two being mostly modulated by non-social factors (like diet, reproduction), and the latter by contextual changes of the social environment.

Fish are remarkable examples of social plasticity and behavioral flexibility. A notable case of a highly social and cooperative species, the indo-pacific bluestreak cleaner wrasse *Labroides dimidiatus*, has been exploited as a model for sociality and cooperation. Individuals of this species are known to exhibit advanced cognitive skills (Soares 2017), making them good examples of strategic sophistication in decision making in teleosts, which ultimately translates into a unique demonstration of fish social

plasticity (Bshary 2001, Bshary and Côté 2008, Soares 2017). Naturally, the cleaning system had long raised interest, as the very idea that fishes could repeatedly move away from all their activities just to visit a specific and territorial, smaller and colourful other, was at best puzzling (Côté 2000).

The discovery of conflict in what seemed as harmless ectoparasite gleaning behavior, was a massive step forward in understanding the social complexity of this system: Grutter and Bshary (2003) showed that these cleaners, while foraging on ectoparasites, preferred to feed directly on client mucus, which is energetically costly for the client fish to produce and constitutes cheating. In our view, this was the starting point that enabled the accumulation of evidence on the social and cooperative building blocks of this system. Cleaners and clients are involved in a challenging network of interactions, based on behavioral and physiological costs and benefits (Soares 2017). Clients were discovered to make use of partner control mechanisms aiming to reduce the frequency of cheating events; these mechanisms include close monitoring of cleaner behavior, for instance, in choosing beforehand (eavesdropping) and reacting with termination, switching to other cleaners, or retaliating when bites occur during the cleaning (Bshary and Côté 2008). Thus, clients not only gain from successful parasite removal, but also with a whole neuroendocrine cascade of wellbeing derived from physical and visual contact (Ros et al 2011, Soares et al 2011, Soares et al 2017a, Abreu et al 2018). As a dynamic biological “market”, cleaners are challenged to respond in flexible way, changes that may occur in relation to clients’ species and identity (familiarity – previous recognition, Tebbich et al 2002, Soares et al 2017b); these cognitive modules and physiological mechanisms will be further discussed below.



Adding to all this behavioral plasticity, most cleaner species undergo seasonal variations, mostly related to reproduction, which have significant dietary physiological impact (Soares et al 2014), and hence impact on cleaners' cooperative levels (Bshary 2002). Temperature and other environmental fluctuations also influence cleaners' social plasticity, especially when these tend to be extreme. This was firstly documented in a recent study by Tricki and colleagues (2017), which found that, following the extreme weather events affecting the Great Barrier Reef (consecutive cyclones and the 2016 El Nino event), cleaners failed to display the previously documented strategic abilities (lower ability to manage their reputation and to learn to prioritize food sources as to maximize food intake). These changes were mostly due to a change in clientele densities, which means that the contextual social environment is crucial to the development of cleaners high-end social and interspecific abilities, cognitive differences that had been previously noted by Wismer et al. (2014) in cleaner wrasses from different reefs, and in other species of dedicated (full time) cleaners (the Caribbean cleaning gobies *Elacatinus* spp., Soares et al 2008a).

It's also relevant to note that cleaners are found in many different fish families and most of which only do so in a certain life stage (during the juvenile phase; Côté 2000, Vaughn et al 2016). These transitions are of significant social relevance, as these facultative cleaner species stop interacting with heterospecifics to focus exclusively on their conspecific networks (frequently during adulthood). Finally, some species of cleaning gobies (most notably the case of *Elacatinus prochilus*, Côté and Soares 2011) show alternative mixed strategies, which seem to adaptably express cleaning behavior, with the absence of cleaning being associated with the sponge-dwelling habitat and the



presence of cleaning associated with living on substrata other than sponge (Rüber et al., 2003; Taylor & Hellberg, 2005). Whether this is an overall privilege of *Elacatinus* spp cleaning gobies or something observed in certain socio-environmental conditions or species, is yet to be disclosed. But even in systems other than *L. dimidiatus*, variations of behavioral output are cumulatively being documented: for instance, with the effects of competition in cleaning gobies (Soares et al 2008b), or the effects of uneven habitat use to cleaner-client familiarity and to cleaners' cheating levels (Oates et al 2010). At this point much have been done in terms of behavior, ecology, and physiology of cooperative behavior in cleaner fish, however, the cleaner-client mutualism is still a promising system, most especially in matters of social plasticity and the underlying brain mechanisms.

## 6.2. Nonapeptides in social plasticity of cleaner fish

Arginine-Vasotocin has been found to have tremendous effects on the cleaning predisposition of cleaner wrasses. Intramuscular injection of AVT made cleaners cease inspecting clients and instead turned their focus to conspecific activities, while those injected with V1a receptor antagonists were stimulated to clean (Soares et al. 2012a). Similar learning and cooperative deficits were subsequently found after exogenous AVT injections (Cardoso et al., 2015a, 2015b). AVT appears to work as a switch, turning "on" or "off" the expression of interspecific cooperative behavior of cleaner wrasses, indicating that lower levels of AVT could be a prerequisite for approaching and

interacting with clients while higher AVT activity could predispose cleaners into mating activities (Cardoso et al., 2015a). Interestingly, examination of brain active nonapeptide levels of mixed sex pairs of cleaner wrasse *L. dimidiatus* demonstrated that in males, forebrain isotocin (IT) levels increased with the level of pair association, but no relationship was found for females (Figure 3B; Cardoso et al. 2015c). These males were found to receive more tactile stimulation from female partners, but seem to contribute to an overall decrease of cleaning service quality given to clients (i.e. cheated more often; Cardoso et al 2015c). Recently, male cleaner forebrain IT levels have been found to increase when introduced to clients (visual-only or full contact), compared to conspecifics (Abreu et al 2018b; Fig 3B).

This evidence pointed towards the need for comparative studies. Kulczykowska and colleagues (2015) looked at nonapeptide levels in different brain regions of four species of closely related Labrid fish, and found that in the cerebellum of the obligate cleaners *L. dimidiatus* and *L. bicolor*, AVT and IT levels were higher than those of facultative cleaner *L. australis* (in which juveniles are cleaners and adults are corallivorous) and of a non-cleaner species (the corallivorous *Labrichthys unilineatus*), suggesting that AVT levels at the cerebellum may be associated with the expression of cleaning behavior (Figure 4A). Thus at this point, it's safe to say that the nonapeptides AVT and IT are implicated in decision-making in cleaner wrasse, but with AVT strongly mediating structural and perhaps life-history changes between cleaning and non-cleaning, however much is yet to be discovered.

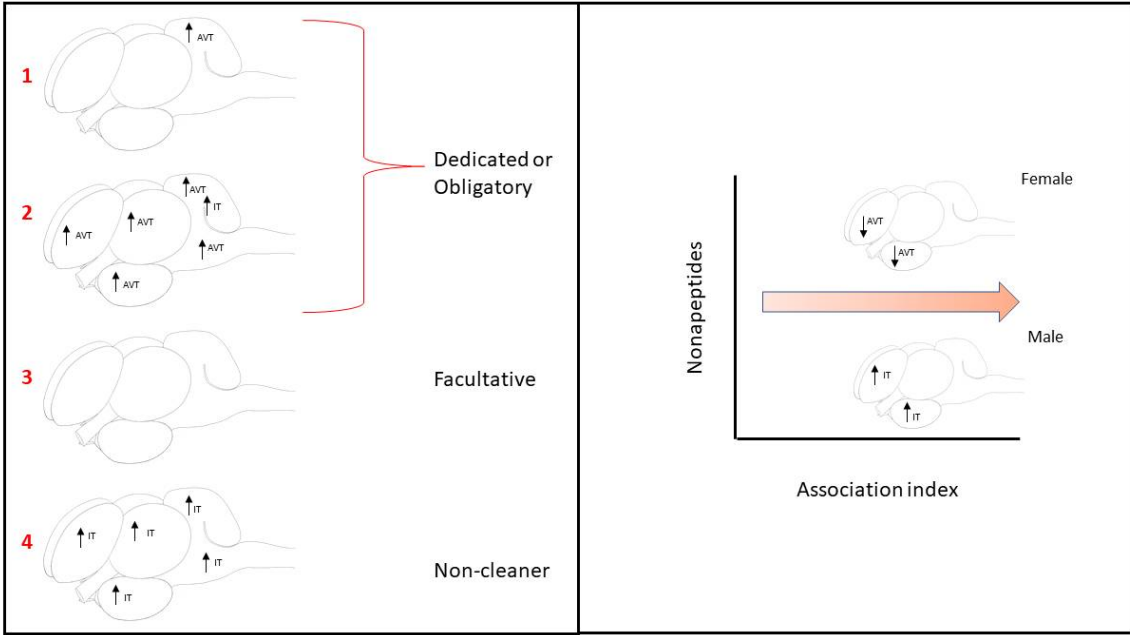


Figure 4: (Left) Differences in nonapeptide levels between closely-related cleanerfish species (adapted from Kulczykowska et al., 2015). (Right) Differences between male and female *L. dimidiatus* (adapted from Cardoso et al. 2015)

### 6.3. Cortisol in social plasticity in cleaner fish

Cortisol was elected as one of the strongest candidate modulators of cleaners and clients' behavioral decisions (Soares et al 2014, Binning et al 2017, Soares 2017). The involvement of cortisol was more obvious to clients, as these visit cleaners to have their parasites removed, and these parasites may cause discomfort, itching, disease, and immune dysfunctions (Côté 2000). On the other hand, for cleaners, cortisol should have a role as a mediator of stress and antipredator responses, since some clients are dangerous piscivores, which cleaners graze around and inside their mouths and leave unharmed (Soares et al 2007a). Interaction of cleaners and clients increase cortisol

840 levels in the first, causing them to attend these clients faster and during longer bouts of  
841 time (Soares et al 2007a, 2012b). Indeed, it has been hypothesized that variations in  
842 cortisol levels work as finer modulators of cleaners' behavioral profiles by contributing to  
843 behavioral shifts (Soares 2017).

844 Bshary (2002) proposed a description of two non-fixed cleaner behavioral  
845 strategies: the first is adopted by the great majority of cleaners, which show low interest  
846 in small clients and focus on the best treatment of larger clients, and the second -  
847 known as "biting", in which cleaners behave in the opposite manner and bite ("cheat")  
848 the larger and most valuable non-predatory clients. Cortisol plays a decisive role in  
849 these behavioral shifts, which occur in stages of higher metabolic demands (and as a  
850 consequence, increased glucocorticoid expenditure), such as during stages of high  
851 reproductive effort, high growth effort, or during stages of change in female social rank  
852 (see Soares et al 2014). However, the scope of influence by cortisol variations seems to  
853 be set by the social environment inhabited by cleaners. For instance, only cleaner  
854 wrasses inhabiting highly complex social environments seem to respond to exogenous  
855 cortisol injections with strategy shifts (e.g. tactical deception of clients: more tactile  
856 stimulation to small clients and more bites to large clients; Binning et al 2017). Another  
857 good example is the case of ecotype differences in cleaning gobies (Soares and Côté  
858 2011), as the absence of cleaning is related to sponge-dwelling habitat and the  
859 presence of cleaning is associated with living on substrata rather than sponge (Rüber et  
860 al., 2003; Taylor & Hellberg, 2005). Stress mechanisms are involved in this system, with  
861 species that show both cleaner and non-cleaner strategies (*E. prochilos*) responding  
862 more to stressful events and in this way enabling the necessary robustness that allows

for them to develop in one of these social environments: one more hierarchical and aggressive (sponges), and another more tolerant and riskier (substratum) (White et al 2007, Soares et al 2018a). There is also a high degree of behavioral flexibility; for instance, sponge dwellers have been observed to inspect clients (Côté and Soares, 2011). However, species of cleaning gobies that are specialized in cleaning (such as *E. evelynae*), depending exclusively on the client-derived food and inspecting dangerous clients often, seem to depend on more precise cortisol responses that enable them to react to smaller trophic differences between clients (Soares et al 2012b, Soares et al 2018a).

#### 6.4. Monoamines in social plasticity in cleaner fish

Cleaner fish (and other teleosts) cognitive function requires flexible coordination of multiple specialized areas of the brain; some of these regions are part of the SDMN. A participation of monoamines in learning and cognition has already been demonstrated in *L. dimidiatus*, as dopamine D1 receptor agonists improves learning of both a cue discrimination task and a side discrimination task (Messias et al. 2016), while serotonin 1A receptor antagonists delays learning of a cue discrimination task (Soares et al. 2016). Recently, efforts have been made to understand how the main brain areas respond, in terms of serotonergic and dopaminergic signalling, during the processing of complex social and mutualistic information (Abreu et al 2018c, Maximino et al 2018 [<https://doi.org/10.1101/326843>]). Abreu and colleagues (2018c) demonstrated that

diencephalon serotonergic activity is particularly responsive in a situation of visual stimulation even in absence of actual physical contact (Figure 4), suggesting that this region processes the cleaner's intrinsic motivation to interact regardless of the outcome (but see Paula et al 2015). While the decrease of dopaminergic activation was expected in the case of social reward omission, this was seemingly only observed when cleaners were prevented to interact with novel conspecifics rather than clients (Figure 4). The importance of cleaners' conspecific relationships, the value of couple dynamics to the overall expression of a series of "bilateral" behaviors between pair partners (conspecifics) and clients, and how these are modulated by cleaners' brain mechanisms, leaves plenty to look forward to in future studies. Interestingly, it was the actual cleaning engagement that resulted in significant change of dopaminergic activity, this time at the cerebellum (Abreu et al 2018c; Figure 4). As is the case with reproductive plasticity and choice in poeciliids, then, the cerebellum rises as a main area for processing mutualistic information (Kulczykowska et al 2015), as it is strongly implicated in cognitive and emotional functions, namely in those linked to associative learning and memory processes (Rodriguez-Ortiz, 2005).

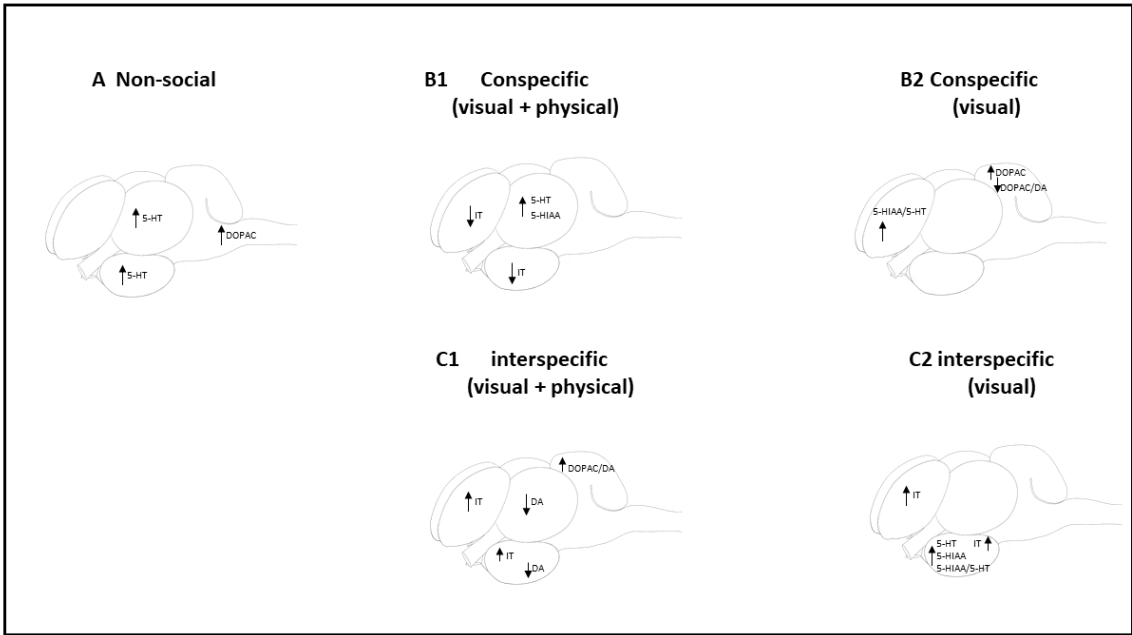


Figure 5: Cleaner brains presents different neuroendocrine shifts that are related to contextual treatments at 3 levels: A) non-social, B) conspecific, and C) interspecific. At different macro-areas: forebrain, diencephalon, optic tectum, cerebellum, and brainstem. Experimental setup, individual cleaner *Labroides dimidiatus* exposed to A) a ball, B) another conspecific, and C) a client. Adapted from Abreu et al. 2018a,b

## 7. Conclusions

Social plasticity represents a challenging, yet evolutionarily and translationally relevant field of investigation across species. The over-reliance on a few “choice species”, under the assumption that they more closely model the human organism, not only does not make evolutionary sense, but also risks missing translationally relevant mechanisms (Gerlai 2014) of social plasticity. The wide range of social organizations across fish species makes them ideal model organisms to study the relationship between social plasticity and brain plasticity.

Patterns of modulation emerge when species and situations are compared. For example, a role for nonapeptides is observed during the establishment of dominance hierarchies; given the relationships between AVT and aggression in fishes, this is not surprising, as behavioral shifts towards more aggressive phenotypes are advantageous when establishing dominance. A limitation of using fish, in this sense, is that brain contents cannot typically be analyzed without killing the animal, and therefore it is not yet possible to know whether nonapeptide levels are already high before the establishment of hierarchies (and therefore predict dominance), or whether they increase to shift behavior. Interestingly, however, in Bluestreak cleaners nonapeptides are implicated in decision-making in cleaner wrasse, with AVT acting as an ontogenetic and punctual switch between cleaning and non-cleaning; since cleaning is not expected to involve aggression, these results underline the role of nonapeptides not only on aggression-based social dynamics, but also on cooperation and mutualism.

Monoamines are also implicated in neural plasticity, especially in the contexts of motivation, arousal, and emotional behavior (Forlano and Bess, 2011). Serotonin is a “pleiotropic” neurotransmitter, being involved in aggression, antipredator defense, fear and anxiety, and social behavior (Herculano and Maximino 2014). In fishes, changes in the serotonergic system were associated with behavioral sex reversal, with serotonin mediating decreased aggression and dominance. Social status changes also mediate sensory plasticity by modulating the serotonergic system, and diencephalic serotonin is important for the establishment of interspecific cooperation in cleaner wrasse. Thus, social plasticity of the serotonergic system appears to be important to induce behavioral shifts associated with aggression and antipredator behavior. Also, the raphe nucleus



932 which contains serotonergic neurons was implicated in regulating reactive coping  
933 behaviors associated with repeated social defeat in an African cichlid (Butler et al.,  
934 2018). Peptides and monoamines, as well as cortisol, appear to act as “initiators” of  
935 brain plasticity in a context of social plasticity.

936 Not surprisingly, changing social status, reproduction, or behavioral strategy  
937 begets structural and functional changes in the brain. The work reviewed here showed  
938 that markers of neuroplasticity and cell proliferation are stimulated by social status  
939 changes, as well as by reproductive status changes (including sex reversal and mate  
940 choice). Interestingly, these effects appear to be restricted to dominants, in the case of  
941 social status. Whether the absence of effects in subordinates results from technical  
942 limitations (e.g., lack of sensitivity), stress effects (which can impair memory, for  
943 example), or other reason is still unknown. However, the breadth of species and  
944 contexts in which these plastic changes take place points to these molecules as “core”  
945 mechanisms in consolidating brain plasticity in a context of social plasticity.

946 Overall, fishes represent an under-studied but promising taxon in the field of  
947 social plasticity. In addition to zebrafish, which have been introduced in the  
948 neuroscience literature, many other species have vibrant communities, important “base”  
949 knowledge to ground neurobehavioral studies on neuroethology, and species-specific  
950 social organizations which beget the need to understand commonalities and differences  
951 in terms of social plasticity in the brain. The future awaits more neuroscientists working  
952 with these animals to increase both evolutionary neuroscience and translational studies.

953

## Acknowledgments

KPM acknowledges support from the National Science Foundation (IOS-1456004; IOS-1456558). MCS is currently supported by SFRH/BPD/109433/2015 (research focusing on cleanerfish financed by the Foundation for Science and Technology-FCT, grant PTDC/MAR/105276/2008 given to MCS). MLM is a recipient of a CNPq grant (Edital Universal 2016; Processo 423735/2016). CM is the recipient of a CNPq grant (Edital Universtal 2016; 400726/2016-5).

## References

- Abreu, M.S., Kulczykowska, E., Cardoso, S.C., André, I.G., Morais, M., Gozdowska, M., Soares, M.C., 2018. Nonapeptide levels in cleaner fish brain during interactions with unfamiliar intra and interspecific partners. *Behav Ecol Sociobiol.* 72, 122
- Abreu, M.S., Messias, J.P.M., Thörnqvist, P.O., Winberg, S., Soares, M.C., 2018. Monoaminergic changes at the forebrain and diencephalon signal the occurrence of mutualistic and conspecific trade-offs in fish. *Scientific Reports* 8:7346.
- Abreu, M.S., Messias, J.P.M., Thörnqvist, P.O., Winberg, S., Soares, M.C., 2018. The variable monoaminergic outcomes of cleaner fish' brains when facing different social and mutualistic contexts. *Peerj* 6:e4830.
- Ashur, M.M., Johnston, N.K., Dixon, D.L., 2017. Impacts of Ocean Acidification on Sensory Function in Marine Organisms. *Integr Comp Biol.* 57, 63-80.
- Beery, A.K., Kaufer, D., 2015. Stress, social behavior, and resilience: Insights from rodents.

- 975 Neurobiol Stress. 1, 116-127.
- 976 Binning, S.A., Rey, O., Triki, Z., Wismer, S., Soares, M.C., Bshary, R., 2017. Reputation  
977 management promotes strategic adjustment of service quality in cleaner wrasse. Scientific  
978 Reports 7: 8425
- 979 Black, M.P., et al., 2005. Socially induced and rapid increases in aggression are inversely  
980 related to brain aromatase activity in a sex-changing fish, *Lythrypnus dalli*. Proc Biol Sci. 272,  
981 2435-40.
- 982 Braun, C.B., 2015. Signals and noise in the octavolateralis systems: what is the impact of  
983 human activities on fish sensory function? Integr Biol. 10, 4-14.
- 984 Brown, C., Laland, K., Krause, J., 2006. Fish cognition and behaviour, edited by Culum Brown,  
985 Kevin Laland, Jens. Krause, Blackwell Publishing.
- 986 Bshary, R. 2001: The cleaner fish market. In: Economics in Nature (Noë, R., van Hooff, J. A. R.  
987 A. M. & Hammerstein, P., eds). Cambridge Univ. Press, Cambridge, pp. 146—172.
- 988 Bshary, R., Côté, I. M., 2008. 'New perspectives on marine cleaning mutualism. In: Fish  
989 Behaviour, eds C. Magnhagen, V. A. Braithwaite, E. Forsgren and B. G. Kapoor (New  
990 Hampshire, NH: Science Publishers), 563–592.
- 991 Bshary, R., Hohner, A., Ait-el-Djoudi, K., Fricke, H., 2006. Interspecific Communicative and  
992 Coordinated Hunting between Groupers and Giant Moray Eels in the Red Sea. PLoS Biol. 4,  
993 e431.
- 994 Bshary, R., Wickler, W., Fricke, H., 2002. Fish cognition: A primate eye's view. Anim Cogn. 5, 1-  
995 13.
- 996 Burmeister, S.S., Jarvis, E.D., Fernald, R.D., 2005. Rapid behavioral and genomic responses to  
997 social opportunity. PLoS Biol. 3, e363.
- 998 Butler, J. M., Whitlow, S. M., Roberts, D. A., Maruska, K. P., 2018. Neural and behavioral  
999 correlates of repeated social defeat. Scientific Reports. 8, 6818.

- 1000 Cardoso, S. A., Teles, M. C., Oliveira, R. F., 2015. Neurogenomic mechanisms of social  
1001 plasticity. J Exp Biol. 218, 140-149.
- 1002 Cardoso, S.C., Bshary, R., Mazzei, R., Paitio, J., Oliveira, R., Soares, M.C., 2015a. Arginine  
1003 vasotocin modulates learning in the mutualistic cleanerfish *Labroides dimidiatus*. Behav Ecol  
1004 Sociobiol. 69, 1173–1181
- 1005 Cardoso, S.C., Paitio, J.R., Oliveira, R.F., Bshary, R., Soares, M.C., 2015b. Arginine vasotocin  
1006 reduces levels of cooperative behaviour in a cleaner fish. Physiol Behav. 139, 314-320
- 1007 Cardoso, S.C., Paula, J.R., André, G.I., Messias, J.P., Gozdowska, M., Kulczykowska, E.,  
1008 Soares, M.C., 2015c. Forebrain neuropeptide regulation of pair association and behavior in  
1009 cooperating cleaner fish. Physiol Behav. 145, 1-7.
- 1010 Carpenter, R.E., et al., 2014. Social opportunity rapidly regulates expression of CRF and CRF  
1011 receptors in the brain during social ascent of a teleost fish, *Astatotilapia burtoni*. PLoS ONE.  
1012 9(5): e96632.
- 1013 Chen, C.C., Fernald, R.D., 2008. Sequences, expression patterns and regulation of the  
1014 corticotropin-releasing factor system in a teleost. Gen Comp Endocrinol. 157, 148-55
- 1015 Cline, H. T., 2001. Dendritic arbor development and synaptogenesis. Curr Opin Neurobiol 11,  
1016 118 – 26.
- 1017 Côté, I.M., 2000. Evolution and ecology of cleaning symbioses in the sea. Oceanogr. Mar.  
1018 Biol. 38, 311–355.
- 1019 Côté, I.M. & Soares, M.C., 2011. Gobies as cleaners. In: The Biology of gobies (ed: R.A.  
1020 Patzner, J.L. Van Tassell, M. Kovacic & B.G. Kapoor), Science Publishers Inc
- 1021 Croft, D.P., James, R., Ward, A.J.W., Botham, M.S., Mawdsley, D., Krause, J., 2005.  
1022 Assortative interactions and social networks in fish. Oecologia. 143, 211-219..
- 1023 Croft, D.P., Morrell, L.J., Wade, A.S., Piyapong, C., Ioannou, C.C., Dyer, J.R.G., Champman,  
1024 B.B., Yan, W., Krause, J., 2006. Predation risk as a driving force for sexual segregation: a

- 1025 cross-population comparison. *Am Nat.* 167, 867–878.
- 1026 Cummings, M., 2018. Sexual conflict and sexually dimorphic cognition—reviewing their  
1027 relationship in poeciliid fishes. *Behav Biol Sociobiol.* 72, 73.
- 1028 Cummings, M.E., Larkins-Ford, J., Reilly, C.R.L., Wong, R.Y., Ramsey, M.E., Hofmann, H.A.,  
1029 2008. Sexual and social stimuli elicit rapid and contrasting genomic responses. *Proc R Soc*  
1030 *Lond B.* 275,393–402.
- 1031 Dadda, M., 2015. Female social response to male sexual harassment in poeciliid fish: a  
1032 comparison of six species. *Front Psychol.* 6, 1453.
- 1033 Dahlbom, S.J., Lagman, D., Lundstedt-Enkel, K., Sundström, L.F., Winberg, S., 2011. Boldness  
1034 Predicts Social Status in Zebrafish (*Danio rerio*). *PLoS ONE.* 6, e23565.
- 1035 Dahlbom, S.J., Backström, T., Lundstedt-Enkel, K., Winberg, S., 2012. Aggression and  
1036 monoamines: Effects of sex and social rank in zebrafish (*Danio rerio*). *Behav Brain Res.* 228,  
1037 333-338.
- 1038 Darden, S.K., Croft, D.P., 2008. Male harassment drives females to alter habitat use and leads  
1039 to segregation of the sexes. *Biol Lett.* 4, 449– 451
- 1040 Davis, M.R., Fernald, R.D., 1990. Social control of neuronal soma size. *J Neurobiol.* 21, 1180-8.
- 1041 Dunlap, K. D., Chung, M., Castellano, J. F., 2013. Influence of long-term social interaction on  
1042 chirping behavior, steroid levels, and neurogenesis in weakly electric fish. *J Exp Biol.* 216, 2434-  
1043 2441.
- 1044 Ebbeson, L. O. E., Braithwaite, V. A., 2012. Environmental effects on fish neural plasticity and  
1045 cognition. *J Fish Biol.* 81, 2151-74.
- 1046 Elliott, S. B., Harvey-Girard, E., Giassi, A. C. C., Maler, L., 2017. Hippocampal-like circuitry in  
1047 the pallium of an electric fish: possible substrates for recursive pattern separation and  
1048 completion. *J Comp Neurol.*, 525, 8-46.
- 1049 Etheredge, R.I., Avenas, C., Armstrong, M.J., Cummings, M.E., 2018. Sex specific cognitive-

- behavioural profiles emerging from individual variation in numerosity discrimination in *Gambusia affinis*. Anim Cogn. 21, 37–53.
- Fernald, R.D., Maruska, K.P., 2012. Social information changes the brain. Proc Natl Acad Sci U S A. 109 Suppl 2, 17194-9.
- Filby, A.L., Paul, G. C., Bartlett, E.J., van Look, K.J.W., Tyler, C.R., 2010. Physiological and health consequences of social status in zebrafish (*Danio rerio*). Physiol Behav. 101, 576-587.
- Fisher, M.A., Oleksiak, M.F., 2007. Convergence and divergence in gene expression among natural populations exposed to pollution. BMC Genomics. 8, 108.
- Forlano, P. M., Bass, A. H., 2011. Neural and Hormonal Mechanisms of Reproductive-Related Arousal in Fishes. Horm Behav. 59, 616–629.
- Forlano, P.M., et al., 2016. Hormone-dependent plasticity of auditory systems in fishes. In: Hearing and Hormones. Springer Handbook of Auditory Research, Vol. 57. Vol., A.H. Bass, J.A. Sisneros, A.N. Popper, R. Fay, eds. Springer Science, New York, pp. 15-51.
- Fox, H. E., White, S. A., Kao, M., Fernald, R. D., 1997. Stress and dominance in a social fish. J Neurosci. 17, 6463-6469.
- Francis, R.C., Soma, K., Fernald, R.D., 1993. Social regulation of the brain-pituitary-gonadal axis. Proc Natl Acad Sci USA. 90, 7794–7798.
- Fricke, H.W. 1973. Eine Fische-Seeigel-Partnerschaft. Untersuchungen optischer Reizparameter beim Formenerkennen. Mar Bio. 119, 290-297.
- Ganz, J., Kaslin, J., Freudenreich, D., Machate, A., Geffarth, M., Brand, M., 2012. Subdivisions of the adult zebrafish pallium based on molecular marker analysis. J Comp Neurol. 520, 633-655.
- Gerlai, R., 2014. Fish in behavior research: Unique tools with a great promise! J Neurosci Methods. 234, 54-58.
- Gilmour, K.M., Dibattista, J.D., Thomas, J.B., 2005. Physiological causes and consequences of

- 1075 social status in salmonid fish. *Integr Comp Biol.* 45, 263-73.
- 1076 Goodson, J. L., 2005. The Vertebrate Social Behaviour Network: Evolutionary Themes and  
1077 Variations. *Horm Behav.* 48, 11–22.
- 1078 Goodson, J. L., Kingsbury, M. A., 2013. What's in a name? Considerations of homologies and  
1079 nomenclature for vertebrate social behavior networks. *Horm Behav.* 64, 103-112.
- 1080 Greenwood, A.K., Wark, A.R., Fernald, R.D., Hofmann, H.A., 2008. Expression of arginine  
1081 vasotocin in distinct preoptic regions is associated with dominant and subordinate  
1082 behaviour in an African cichlid fish. *Proc. R. Soc. B Biol. Sci.* 275, 2393–2402.
- 1083 Grober, M. S., Bass, A. H., 1991. Neuronal correlates of sex/role change in labrid fishes: LHRH-  
1084 like immunoreactivity. *Brain Behav Evol.* 38, 302-312.
- 1085 Grober, M. S., Jackson, I. M. D., Bass, A. H., 1991. Gonadal steroids affect LHRH preoptic cell  
1086 number in a sex/role changing fish. *J Neurobiol.* 22, 734-741.
- 1087 Grosenick, L., Clement, T.S., Fernald, R.D., 2007. Fish can infer social rank by observation  
1088 alone. *Nature.* 445, 429-32.
- 1089 Gross, M.R., Sargent, R.C., 1985. The evolution of male and female parental care in fishes. *Am*  
1090 *Zool.* 25, 807-822.
- 1091 Grutter, A. S., Bshary, R., 2003. Cleaner Wrasse Prefer Client Mucus: Support for Partner  
1092 Control Mechanisms in Cleaning Interactions. *Proc Royal Soc B.* 270, S242–S244.
- 1093 Hall, Z.J., de Serrano, A.R., Rodd, F.H., Tropepe, V., 2014. Casting a wider fish net on  
1094 animal models in neuropsychiatric research. *Prog. Neuro-Psychopharmacol. Biol.*  
1095 *Psychiatry* 55, 7–15.
- 1096 Harvey-Girard, E., Giassi, A. C. C., Ellis, W., Maler, L., 2013. Expression of the cannabinoid  
1097 CB1 receptor in the gymnotiform fish brain and its implications for the organization of the teleost  
1098 pallium. *J Comp Neurol.* 521, 949-975.
- 1099 Herculano, A.M., Maximino, C., 2014. Serotonergic modulation of zebrafish behavior: towards a

- 1100 paradox. Prog Neuropsychopharmacol Biol Psychiatry. 55, 50-66.
- 1101 Hofmann, H.A., Benson, M.E., Fernald, R.D., 1999. Social status regulates growth rate:  
1102 consequences for life-history strategies. Proc Natl Acad Sci U S A. 96, 14171-6.
- 1103 Iwata, E., Nagai, Y., Hyoudou, M., Sasaki, H., 2008 Social environment and sex differentiation  
1104 in the false anemonefish, *Amphiprion ocellaris*. Zool Sci. 25, 123–128.
- 1105 Kasper, C., Colombo, M., Aubin-Horth, N., Taborsky, B., 2018a. Brain activation patterns  
1106 following a cooperation opportunity in a highly social cichlid fish. Physiol Behav. 195, 37-47.
- 1107 Kasper, C., Hebert, F. O., Aubin-Horth, N., Taborsky, B., 2018b. Divergent brain gene  
1108 expression profiles between alternative behavioural helper types in a cooperative breeder. Mol  
1109 Ecol.
- 1110 Keenlyside, M.H.A. 1979. Diversity and Adaptation in Fish Behaviour. Springer-Verlag, Berlin.
- 1111 Khoo, M. L., Das, S. K., & Ghaffar, M. A. 2018. Growth pattern, diet and reproductive biology of  
1112 the clownfish *Amphiprion ocellaris* in waters of Pulau Tioman, Malaysia. Egypt J Aquat Res.
- 1113 Krause, J and GD Ruxton 2002. Living in groups. Oxford series in Ecology and Evolution.  
1114 Oxford University press. Oxford.
- 1115 Krogh, A., 1929. The Progress of Physiology. Science. 70, 200-4.
- 1116 Kulczykowska, E., Cardoso, S.C., Gozdowska, M., André, G.I., Paula, J.R., Slebiada, M.,  
1117 Oliveira, R.F., Soares, M.C., 2015. Brain Levels of nonapeptides in four labrid fish with different  
1118 levels of mutualistic behaviour. General Comp Endocrinol. 222, 99-105.
- 1119 Kullander, S. O., & Ferreira, E. J. 2006. A review of the South American cichlid genus *Cichla*,  
1120 with descriptions of nine new species (Teleostei: Cichlidae). Ichthyol Explor Freshw. 17(4), 289-  
1121 398.
- 1122 Lai, F., et al., 2017. Responses of neurogenesis and neuroplasticity related genes to elevated  
1123 CO<sub>2</sub> levels in the brain of three teleost species. Biol Lett. 13.
- 1124 Larson, E. T., Norris, D. O., Grau, E. G., Summers, C. H., 2003a. Monoamines stimulate sex



- 1125 reversal in the saddleback wrasse. *Gen Comp Endocrinol.* 130, 289-298.
- 1126 Larson, E. T., Norris, D. O., Summers, C. H., 2003b. Monoaminergic changes associated with  
1127 socially induced sex reversal in the saddle-back wrasse. *Neuroscience.* 119, 251-263.
- 1128 Larson, E.T., O'Malley, D.M., Melloni Jr, R.H., 2006. Aggression and vasotocin are associated  
1129 with dominant-subordinate relationships in zebrafish. *Behav Brain Res.* 167, 94-102.
- 1130 Liu, M., Wang, Y. Y., Shan, X. J., Kang, B., & Ding, S. X. 2016. Primary male development of  
1131 two sequentially hermaphroditic groupers, *Epinephelus akaara* and *Epinephelus awoara*  
1132 (Perciformes: Epinephelidae). *J. Fish Biol.* 88(4), 1598-1613.
- 1133 Lo Nostro, F. L., & Guerrero, G. A. 1996. Presence of primary and secondary males in a  
1134 population of the protogynous *Synbranchus marmoratus*. *J. Fish Biol.* 49(5), 788-800.
- 1135 Loveland, J.L., et al., 2014. Social status differences regulate the serotonergic system of a  
1136 cichlid fish, *Astatotilapia burtoni*. *J Exp Biol.* 217, 2680-90.
- 1137 Lynch, K.S., Ramsey, M.E., Cummings, M.E., 2012. The mate choice brain: Comparing gene  
1138 profiles between female choice and male coercive poeciliids. *Genes Brain Behav.* 11, 222–229.
- 1139 Ma, Y., et al., 2015. Electrical synapses connect a network of gonadotropin releasing hormone  
1140 neurons in a cichlid fish. *Proc Natl Acad Sci U S A.* 112, 3805-10.
- 1141 Maruska, K.P., Butler, J.M., Field, K.E., 2018. Alternative Reproductive Phenotypes within  
1142 Species. *In: Encyclopedia of Reproduction*, 2<sup>nd</sup> edition, Volume 6, pp. 41-49. (Skinner, M.K, ed).  
1143 Academic Press: Elsevier.
- 1144 Maruska, K.P., Fernald, R.D., 2010a. Behavioral and physiological plasticity: Rapid changes  
1145 during social ascent in an African cichlid fish. *Horm Behav.* 58, 230-240.
- 1146 Maruska, K.P., Fernald, R.D., 2010b. Steroid receptor expression in the fish inner ear varies  
1147 with sex, social status, and reproductive state. *BMC Neurosci.* 11:58.
- 1148 Maruska, K.P., Fernald, R.D., 2010c. Reproductive status regulates expression of sex steroid  
1149 and GnRH receptors in the olfactory bulb. *Behav Brain Res.* 213, 208-217.

- 1150 Maruska, K.P., Carpenter, R.E., Fernald, R.D., 2012. Characterization of cell proliferation  
1151 throughout the brain of the African cichlid fish *Astatotilapia burtoni* and its regulation by social  
1152 status. J Comp Neurol. 520, 3471-91.
- 1153 Maruska, K.P., et al., 2013a. Social descent with territory loss causes rapid behavioral,  
1154 endocrine, and transcriptional changes in the brain. J Exp Biol.
- 1155 Maruska, K.P., Fernald, R.D., 2013. Social Regulation of Male Reproductive Plasticity in an  
1156 African Cichlid Fish. Integr Comp Biol. 207, 2-12.
- 1157 Maruska, K.P., et al., 2013b. Social opportunity causes rapid transcriptional changes in the  
1158 social behavior network of the brain in an African cichlid fish. J Neuroendocrinol. 25, 145-157.
- 1159 Maruska, K.P., 2014. Social regulation of reproduction in male cichlid fishes. Gen Comp  
1160 Endocrinol. 207, 2-12.
- 1161 Maruska, K.P., Fernald, R.D., 2014. Social regulation of gene expression in the African cichlid  
1162 fish *Astatotilapia burtoni*. In: Handbook of Molecular Psychology. Vol., T. Canli, ed.^eds. Oxford  
1163 University Press, New York, N.Y., pp. 52-78.
- 1164 Maruska, K.P., 2015. Social Transitions Cause Rapid Behavioral and Neuroendocrine Changes.  
1165 Integr Comp Biol. 55: 294-306.
- 1166 Maruska, K.P., Sisneros, J.A., 2015. Sex steroid-dependent modulation of acoustic  
1167 communication systems in fishes. In: Sound Communication in Fishes. Vol., F. Ladich, ed.^eds.  
1168 Springer-Vienna, pp. 207-233.
- 1169 Maruska, K.P., Fernald, R.D., 2018. *Astatotilapia burtoni*: A Model System for Analyzing the  
1170 Neurobiology of Behavior. ACS Chem Neurosci.
- 1171 Maximino, C., Gomes, A.C., de Abreu, M.S., Cardoso, S., Lima-Maximino, M., Winberg, S.,  
1172 Soares, M.C., 2018. Profiles of cooperative brains: A discriminant analysis of cleaner and client  
1173 fish monoaminergic responses to different social contexts. bioRxiv preprint. 326843.
- 1174 Messias, J.P.M., Santos, T.P., Pinto, M., Soares, M.C., 2016. Stimulation of dopamine D1

- 1175 receptor improves learning capacity in cooperating cleaner fish. *Proc Royal Soc B.* 283,  
1176 20152272.
- 1177 Miller, T.H., et al., 2017. Social Status-Dependent Shift in Neural Circuit Activation Affects  
1178 Decision Making. *J Neurosci.* 37, 2137-2148.
- 1179 Moyle PB, Cech JJ 2000 *Fishes an introduction to ichthyology*, 4<sup>th</sup> edition. Prentice Hall.
- 1180 Neumeister, H., et al., 2010. Social and ecological regulation of a decision-making circuit.  
1181 *Journal of neurophysiology.* 104, 3180-3188.
- 1182 Nikonov, A.A., et al., 2017. Reproductive and metabolic state differences in olfactory responses  
1183 to amino acids in a mouth brooding African cichlid fish. *J Exp Biol.* 220, 2980-2992.
- 1184 Nyman, C., Fischer, S., Aubin-Horth, N., Taborsky, B., 2017. Effect of the early social  
1185 environment on behavioural and genomic responses to a social challenge in a cooperatively  
1186 breeding vertebrate. *Mol Ecol.* 26, 3186-3203.
- 1187 Oates, J., Manica, A., and Bshary, R., 2010. Roving decreases service quality in the cleaner  
1188 wrasse *Labroides bicolor*. *Ethology.* 116, 309–315.
- 1189 O'Connell, L. A., Hofmann, H. A., 2012a. Evolution of a Vertebrate Social Decision-Making  
1190 Network. *Science.* 336, 1154-1157.
- 1191 O'Connell, L.A., Hofmann, H.A., 2012b. Social status predicts how sex steroid receptors  
1192 regulate complex behavior across levels of biological organization. *Endocrinology.* 153, 1341-  
1193 1351.
- 1194 Oliveira, R.F., Taborsky, M., Brockmann, H.J., 2008. *Alternative Reproductive Tactics: An*  
1195 *Integrative Approach.* Cambridge University Press, Cambridge,  
1196 MA
- 1197 Oliveira, R. F., 2009. Social behavior in context: Hormonal modulation of behavioral plasticity  
1198 and social competence. *Int Comp Biol.* 49, 423-440.
- 1199 Oldfield, R.G., Hofmann, H.A., 2011. Neuropeptide regulation of social behavior in a

- 1200 monogamous cichlid fish. *Physiol. Behav.* 102, 296–303.
- 1201 Paula, J.R., Messias, J.P., Grutter, A.S., Bshary, R., Soares, M.C., 2015. The role of serotonin  
1202 in the modulation of cooperative behaviour. *Behav Ecol.* 26, 1005-1012.
- 1203 Paull, G.C., Filby, A.L., Giddins, H.G., Coe, T.S., Hamilton, P.B., Tyler, C.R., 2010. Dominance  
1204 Hierarchies in Zebrafish (*Danio rerio*) and Their Relationship with Reproductive Success.  
1205 *Zebrafish.* 7, 109-117.
- 1206 Perrone, R., Silva, A.C., 2018. Status-Dependent Vasotocin Modulation of Dominance and  
1207 Subordination in the Weakly Electric Fish *Gymnotus omarorum*. *Front Behav Neurosci.* 12, 1.
- 1208 Qvamström A, Forsgren E, 1998. Should females prefer dominant males? *Trends Ecol Evol.* 13,  
1209 498–501.
- 1210 Ravaglia, M.A., Lo Nostro, F.L., Maggese, M.C., Guerrero, G.A., Somoza, G.M., 1997.  
1211 Characterization of molecular variants of GnRH, induction of spermiation and sex reversal using  
1212 salmon GnRH-A and domperidone in the protogynous diandric fish, *Synbranchus marmoratus*  
1213 Bloch (Teleostei, Synbranchidae). *Fish Physiol Biochem.* 16, 425-436.
- 1214 Renn, S.C., Aubin-Horth, N., Hofmann, H.A., 2008. Fish and chips: Functional genomics of  
1215 social plasticity in an African cichlid fish. *J Exp Biol.* 211, 3041-56.
- 1216 Renn, S.C., O'Rourke, C.F., Aubin-Horth, N., Fraser, E.J., Hofmann, H.A., 2016. Dissecting the  
1217 Transcriptional Patterns of Social Dominance across Teleosts. *Int Comp Biol.* 56, 1250-1265.
- 1218 Ros, A.F.H., Lusa, J., Meyer, M., Soares, M., Oliveira, R.F., Brossard, M., Bshary, R., 2011.  
1219 Does access to the bluestreak cleaner wrasse *Labroides dimidiatus* affect indicators of stress  
1220 and health in resident reef fishes in the Red Sea? *Horm Behav.* 59, 151-158.
- 1221 Ross, R. M., 1986. Social organization and mating system of the Hawaiian reef fish *Thalassoma*  
1222 *duperrey* (Labridae). *Proc Second Intl Conf Indo-pacific Fish.* 1986, 794-802.
- 1223 Rüber, L., van Tassell, J. L., Zardoya, R., 2003. Rapid Speciation and Ecological Divergence in  
1224 the American Seven-Spined Gobies (Gobiidae, Gobiosomatini) Inferred from a Molecular

- 1225 Phylogeny. *Evolution*. 57, 1584–1598
- 1226 Sadovy de Mitcheson, Y., Liu, M., 2008. Functional hermaphroditism in teleosts. *Fish and*
- 1227 *Fisheries*. 9, 1–43.
- 1228 Sapolsky, R.M., 2005. The influence of social hierarchy on primate health. *Science*. 308, 648-
- 1229 52.
- 1230 Seehausen, O. & Schluter, D., 2004. Male-male competition and nuptial-colour displacement as
- 1231 a diversifying force in Lake Victoria cichlid fishes. *Proc. R. Soc. Lond. B* 271, 1345–1353
- 1232 Semsar, K., Kandel, F.L., Godwin, J., 2001. Manipulations of the AVT system shift social status
- 1233 and related courtship and aggressive behavior in the bluehead wrasse. *Hormones Behav.* 40,
- 1234 21-31.
- 1235 Simpson, M. J. A., 1968. The Display of the Siamese Fighting Fish, *Betta splendens*. *Anim*
- 1236 *Behav Monographs*. 1, 1-73.
- 1237 Soares, M.C., 2017. The neurobiology of cooperation: The cleanerfish swims into the spotlight.
- 1238 *Frontiers Behav Neurosci*. 11, 191.
- 1239 Soares, M.C., Cardoso, S.C., Côté, I.M., 2007a. Client preferences by caribbean cleaning
- 1240 gobies: Food, safety or something else? *Behav Ecol Sociobiol*. 61, 1015-1022.
- 1241 Soares, M.C., Bshary, R., Cardoso, S.C., Côté, I.M, 2008. Does competition for clients increase
- 1242 service quality in cleaning gobies. *Ethology*. 114, 625–632
- 1243 Soares, M.C., Oliveira, R., Ros, A.F.H., Grutter, A., Bshary, R., 2011. Tactile stimulation lower
- 1244 stress in fish. *Nat Commun*. 2, 534.
- 1245 Soares, M.C., Bshary, R., Mendonça, R., Grutter, A., Oliveira, R., 2012. Neuropeptide
- 1246 modulation of cooperative behaviour: Arginine vasotocin decreases prosocial behaviour in
- 1247 cleanerfish. *PLoS ONE*. 7, e39583.
- 1248 Soares, M.C., Bshary, R., Cardoso S.C., Côté, I.M., Oliveira, R.F., 2012. Face your fears:
- 1249 Cleanerfish inspect predators despite being stressed by them. *PLoS ONE*. 7, e39781.

- 1250 Soares, MC., Cardoso, S.C., Grutter, A., Oliveira, R., Bshary, R., 2014. Cortisol mediates  
1251 cleaner wrasse switch from cooperation to cheating and tactical deception. *Horm Behav.* 66,  
1252 346-345.
- 1253 Soares, M.C., de Paula, J.R., Bshary, R., 2016. Serotonin blockade delays learning  
1254 performance in a cooperative fish. *Anim Cogn.* 19, 1027–1030.
- 1255 Soares, M.C., Cardoso, S.C., André, I.G., Mazzei, R., Morais, M., Gozdowska, M., Kalamarz-  
1256 Kubiak, H., Kulczykowska, E., 2017a. Region specific changes in nonapeptide levels during  
1257 client fish interactions with allopatric and sympatric cleaner fish. *PLoS ONE.* 12, e0180290.
- 1258 Soares, M.C., Santos, T.P., Santos, M.J.P.M., 2017b. Dopamine disruption increases  
1259 cleanerfish cooperative investment to novel client partners. *Royal Soc Open Sci.* 4, 160609.
- 1260 Soares, M.C., Gerlai, R., Maximino, C., 2018a. The integration of sociality, monoamines and  
1261 stress neuroendocrinology in fish models: Applications in the neurosciences. *J Fish Biol.*
- 1262 Soares, M.C., Cardoso, S.C., Carvalho, T.S., Maximino, C., 2018b. The use of model fish as  
1263 tools for research the biological mechanisms of cooperative behaviour: a future for translational  
1264 research concerning social anxiety disorders? *Progr Neuro-Psychopharmacol Biol Psychiatry.*  
1265 82, 205-215.
- 1266 Sørensen, C., Nilsson, G.E., Summers, C.H., Øverli, Ø, 2012. Social stress reduces forebrain  
1267 cell proliferation in rainbow trout (*Oncorhynchus mykiss*). *Behav Brain Res.* 227, 311-318.
- 1268 Sorensen, C., Overli, O., Summers, C. H., Nilsson, G. E., 2007. Social regulation of  
1269 neurogenesis in teleosts. *Brain Behav Evol.* 70, 239-246.
- 1270 Suriyampola, P., Shelton, D., Shukla, R., Roy, T., Bhat, A., Martins, E. P., 2015. Zebrafish  
1271 Social Behavior in the Wild. *Zebrafish.* 13, 1-8.
- 1272 Taborsky, M., Limberger, D., 1981. Helpers in fish. *Behav Ecol Sociobiol.* 8, 143–14.
- 1273 Taborsky, B., Oliveira, R. F., 2012. Social Competence: An Evolutionary Approach. *Trends Ecol*  
1274 *Evol.* 27, 670-688.

- 1275 Taborsky, B., Tschirren, L., Meunier, C., Aubin-Horth, N. 2013. Stable reprogramming of brain  
1276 transcription profiles by the early social environment in a cooperatively breeding fish. Proc Royal  
1277 Soc B. 280, 20122605.
- 1278 Taylor, M. S., Hellberg, M. E., 2005. Marine Radiations at Small Geographic Scales: Speciation  
1279 in Neotropical Reef Gobies (Elacatinus). Evolution. 59, 374–385.
- 1280 Tebbich, S., Bshary, R., Grutter, A. S., 2002. Cleaner Fish *Labroides dimidiatus* Recognise  
1281 Familiar Clients. Animal Cogn. 5, 139–145.
- 1282 Teles, M.C., Cardoso, S.D., Oliveira, R.F., 2016. Social Plasticity Relies on Different  
1283 Neuroplasticity Mechanisms across the Brain Social Decision-Making Network in Zebrafish.  
1284 Front Behav Neurosci. 10, 16.
- 1285 Todd, E.V., et al., 2018. Female Mimicry by Sneaker Males Has a Transcriptomic Signature in  
1286 Both the Brain and the Gonad in a Sex-Changing Fish. Mol Biol Evol. 35, 225-241.
- 1287 Triki, Z., Wismer, S., Levorato, E., Bshary, R., 2018. A decrease in the abundance and strategic  
1288 sophistication of cleaner fish after environmental perturbations. 24, 481-489.
- 1289 Valdimarsson, S.K., Metcalfe, N.B., 2001. Is the level of aggression and dispersion in territorial  
1290 fish dependent on light intensity? Anim Behav. 61, 1143–1149.
- 1291 Vaughan, D. B., Grutter, A. S., Costello, M. J., Hutson, K. S., 2016. Cleaner fishes and shrimp  
1292 diversity and a re-evaluation of cleaning symbioses. Fish Fish. 18, 698–716.
- 1293 Wang, S., Ramsey, M.E., Cummings, M.E., 2014. Plasticity of the mate choice mind: Evoking  
1294 choice-like brain responses in coercive mating systems. Genes Brain Behav. 13, 365–375.
- 1295 Warner, R. R., Swearer, S. E., 1991. Social control of sex change in the bluehead wrasse,  
1296 *Thalassoma bifasciatum* (Pisces: Labridae). Biol Bull. 181, 199-204.
- 1297 Whitaker, K.W., et al., 2011. Serotonergic modulation of startle-escape plasticity in an African  
1298 cichlid fish: a single-cell molecular and physiological analysis of a vital neural circuit. J  
1299 Neurophysiol. 106, 127-37.

- 1300 White, J. W., Grigsby, C. J., Warner, R. R., 2007 Cleaning behaviour Is Riskier and Less  
 1301 Profitable than an Alternative Strategy for a Facultative Cleaner Fish. Coral Reefs. 26, 87– 94.
- 1302 Wingfield, J.C., Sapolsky, R.M., 2003. Reproduction and resistance to stress: when and how. J  
 1303 Neuroendocrinol. 15, 711-24.
- 1304 Wismer, S., Pinto, A. I., Vail, A. L., Grutter, A. S., Bshary, R., 2014. Variation in cleaner wrasse  
 1305 cooperation and cognition: potentially caused by developmental environment? Ethology. 120,  
 1306 519–531.
- 1307 Wong M, Balshine S. 2011. The evolution of cooperative breeding in the African cichlid fish,  
 1308 *Neolamprologus pulcher*. Biol Rev Camb Philos Soc. 86, 511-530.
- 1309 Wong, R.Y., Cummings, M.E., 2014. Expression patterns of *neuroligin-3* and *tyrosine*  
 1310 *hydroxylase* across the brain in mate choice contexts in female swordtails. Brain Behav Evol.  
 1311 83, 231–243.
- 1312 Wong, R.Y., Ramsey, M.E., Cummings, M.E., 2012. Localizing brain regions associated with  
 1313 female mate preference behavior in a swordtail. PLoS One. 7, e50355.
- 1314 Zupanc, G. K., Sirbulescu, R. F., 2011. Adult neurogenesis and neuronal regeneration in the  
 1315 central nervous system of teleost fish. Eur J Neurosci. 34, 917-29.