

## **Impact of Maternal Air Pollution Exposure on Children's Lung Health: An Indian Perspective**

Pritam Saha<sup>1</sup>, Ebin Johny<sup>2</sup>, Ashish Dangi<sup>1</sup>, Sopan B Shinde<sup>1</sup>, Mathew Suji Eapen<sup>3</sup>,  
Sukhwinder Singh Sohal<sup>3</sup>, VGM Naidu<sup>1</sup>, Pawan Sharma<sup>4,5</sup>

<sup>1</sup>Department of Pharmacology, National Institute of Pharmaceutical Education and Research, Guwahati, Assam; <sup>2</sup>Department of Pharmacy Practice, National Institute of Pharmaceutical Education and Research, Guwahati, Assam; <sup>3</sup>Respiratory Translational Research Group, Department of Laboratory Medicine, School of Health Sciences, University of Tasmania, Launceston, Tasmania, Australia, 7248; <sup>4</sup>Medical Sciences, School of Life Sciences, Faculty of Science, University of Technology Sydney, NSW, Australia, 2007; and <sup>5</sup>Woolcock Emphysema Centre, Woolcock Institute of Medical Research, The University of Sydney, Sydney, NSW, Australia 2037.

### **\*Address for Correspondence:**

Pawan Sharma, PhD  
The University of Technology Sydney  
15 Broadway, NSW 2007, Australia  
Tel: + 61 2 9514 3815  
E-mail: Pawan.Sharma@uts.edu.au

## Abstract

Air pollution has become a growing invisible killer in recent years and is major cause of morbidity and mortality globally. India stands 10<sup>th</sup> among the highly polluted countries with an average PM<sub>10</sub> level of 134µg/m<sup>3</sup> per year. It is also reported that 99% of India's population comes across air pollution level that exceed the World Health Organization Air Quality Guideline (AQG), PM<sub>2.5</sub> permissible levels of 10 µg/m<sup>3</sup>. Maternal exposure to air pollution has a serious health outcome to the offspring because it can affect embryonic phases of development during the gestation period. Fetus is more prone to air pollution effect during embryonic developmental phases due to oxidative stress as antioxidant mechanisms are lacking at that stage. Any injury during this vulnerable period (embryonic phase) will have long-term impact on offspring health both in early and later in life. Epidemiological studies have revealed that maternal exposure to air pollution increases the risk of developing airways disease in offspring due to impaired lung development *in utero*. In this review, we discuss cellular mechanisms involved in maternal exposure to air pollution and how it can impact development of airways disease in offspring. Better understanding of these mechanisms in context of maternal exposure to air pollution can offer newer avenue to prevent development of airways disease in offspring.

## Keywords

PM, Air pollution, maternal exposure, Airway disease

## Introduction

In recent years air pollution has become a leading cause of morbidity and death globally (1, 2). Air pollutant particularly particulate matter (PM) pose a major havoc to human health by causing serious respiratory illness such as chronic obstructive Pulmonary Disease (COPD), asthma and act as a trigger to various forms of chronic interstitial lung disease and lung cancer (ref). PM consist of fine and coarse particulate matter; fine PM<sub>2.5</sub> has an aerodynamic diameter of  $<2.5\mu\text{M}$ , while the coarse PM<sub>10</sub> is  $<10\mu\text{M}$ . Both PMs primarily emanate from vehicles, industrial exhausts and household sources (3). It has been well established that both PM<sub>2.5</sub> and PM<sub>10</sub> can cause serious respiratory damage (4). According to the World Health Organization (WHO) report in 2018, air pollution induces adverse health effects and as a result 8 million people undergo premature deaths every year. Air pollution contributes at least 23% of deaths from lung cancer, 43% occur through respiratory disease, 24% from strokes and 23% suffer from ischemic heart diseases related deaths (5). An interesting observation is that 91% of the world's population inhale poor quality air that exceeds WHO guideline of air pollution level. Further, increasing modernization also accounts for the increasing concentrations of airborne PM. Both short, and long-term exposure to air pollutants cause major respiratory associated pathological changes such as reduced pulmonary function, increase in respiratory infections and development of asthma and COPD (6).

One of the more important factors that is of concern is the effect of air pollutants during maternal gestational period. Indeed, increased presence of PM is associated with numerous adverse birth outcomes, such as pre-term delivery (7), low birth weight (8), small gestational age of births (9), congenital heart defects (10), intrauterine growth restriction (11, 12), stillbirth (13), and spontaneous abortion (14). Further, associated morbidities includes neurodevelopmental

impairment leading to attention-deficit hyperactivity disorder (ADHD) and possible autism (15-17), neurological disorders such as impaired neurocognitive ability, nervous system sequelae (18), infant eczema (19). Increased maternal PM exposure is likely to increase the risk of childhood asthma as well (20).

Essentially, PM is a mixture of solid and liquid particles consisting of inorganic and organic matter, dust particles, poly aromatic hydrocarbons (PAH), volatile compounds like sulphates, nitrates, ammonia and minute quantities of metals (TMs), as well as water and unidentified compounds (21). The source of PM can be divided into households (indoor) and commercial (outdoor). Indoor sources includes cigarette smoking, construction work, cooking with kerosene stove or conventional wood burning (chulas), burning of incense or mosquito repellent coils, pesticides and cleaning agents used at homes and room fresheners (aerosols) (22). Outdoor sources include industrial exhaust, diesel vehicles, burning of coals, wood, and road dust. Further sources of PM can be elucidated depending upon mode of emission, such as those from anthropogenic and natural sources (23). Anthropogenic PM source generally would include burning of coal, petroleum products, biomass, industrial processing, agricultural activities, vehicular exhaust, and friction between tires and road, while natural sources would include volcanic eruptions, dust storms, forest fires among others (24, 25). However, the adverse health effects of PM not only depend on the source of PM but also by the chemical composition that adsorb on its surface [27].

India stands among the ten most highly polluted countries with an average PM<sub>10</sub> level of 134µg/m<sup>3</sup> per year. Sadly, out of 100 most polluted cities around the world, 42 Indian cities are listed as highly polluted. Of utmost concern is that 99% of India's population is exposed to air pollution level that exceed the WHO Air Quality Guideline (AQG) PM<sub>2.5</sub> permissible levels of 10 µg/m<sup>3</sup>

(26). Recent evidence also show that on an average PM<sub>2.5</sub> levels in India are found to be in the range of ~10-100 µg/m<sup>3</sup>, a grave situation which needs urgent attention (27, 28). Because PM<sub>2.5</sub> comes under India's 10<sup>th</sup> health associated risk factors which causes premature death of approximately 1.67 million people according to estimates given by Global Burden of Disease (29, 30). In India higher concentrations of PM<sub>2.5</sub> are observed in northern parts rather than in southern region (28), and this is because northern India is landlocked. This geographical feature leads to poor wind circulation, thus causing air pollutants to dwindle in the atmosphere for longer periods. In contrast, southern India is surrounded by coastal region wherein sea breeze/wind plays a crucial role in pushing away the pollutants from the region. Moreover, during winters in North India, the creation of high-pressure zone obstructs wind generation, impacting the movement of the pollutants. Among metropolitan cities in India, New Delhi, Kolkata, Mumbai, and Hyderabad shows maximum PM<sub>2.5</sub> levels with an average concentration: 40-81 µg/m<sup>3</sup> per year, again exceeding the safe levels (40µg/m<sup>3</sup>). Air pollutants in India contribute to the decrease in life expectancy on an average by 3.4 years in the general population (31). According to the reports published by Environmental Performance Index (EPI), approximately 3.5 billion people, i.e. half of the world's population are exposed to air pollutant of which three-quarters are of India's population (32). There are several descriptive studies conducted in India to find the impact of air pollution in neonates and children as represented in the Table 1 (33-35), however, these studies did not explore the underlying pathophysiological mechanisms. This review will provide an overview of all health issues concerning particulate matter exposure both indoor and outdoor with special emphasis on maternal exposure and its implications on children's health in India. It will also address the unwanted health effects and highlight some novel mechanisms that can be future targets for drug therapy.

## Evidence of maternal air pollution-induced health effects in offspring

Studies reveal that exposure to PM has been attributed for a large number of health issues in offspring including reduced lung function, increased chances of lower respiratory infections, cardiovascular diseases, exacerbation of chronic respiratory disease and premature mortality (32, 40). Birth weight is a foremost parameter to check for fetal growth. For example, it is now evident that maternal exposure to PM leads to lower birth weight (<2.5Kg), preterm deliveries and IUGR, which are likely factors associated with increased morbidity and mortality in offspring as well as heightened risk of other health complications later in life (41). PM exposure results in the generation of reactive oxygen species (ROS) and reactive nitrogen species (RNS), both cause oxidative stress that triggers DNA damage leading to the development of placental DNA adducts formation (42). In addition, one of the PM components i.e. PAH binds to placental growth factor receptors resulting in an inadequate transplacental-nutrient exchange. Transplacental nutrient and oxygen transport are crucial during gestation period for regulating normal fetal growth and development (43). Moreover, PM also contain some inorganic metals like chromium, aluminum, silicon, titanium, iron, and copper that results in up-regulation of pro-inflammatory mediators contributing to pulmonary inflammation (44). Inadequate placental perfusion may also cause inflammation resulting in growth restriction *in utero* due to interference with the nutrient-oxygen transfer of the fetus leading to deoxygenation of maternal blood or both (41). It has been reported that maternal exposure to PM alters hematological parameters such as blood coagulation capacity, change in viscosity, levels of hemoglobin, platelets, and white blood cells, which further correlates to PM toxicity, leading to adverse fetal growth (45). In addition to the changes in hematological parameters, there is also endothelial dysfunction, which is attributed to maternal PM exposure during gestational period (46). It has been reported that systemic inflammation and oxidative stress

caused by PM exposure can increase plasma dimethyl arginine concentration (47). As dimethyl arginine is an endogenous nitric oxide (NO) synthase inhibitor (48) which decrease NO levels, as a consequence lead to endothelial dysfunction which is linked to impaired vascular function and increased risk of cardiovascular diseases. Studies also suggest that maternal exposure to PM leads to changes in hemodynamic parameter like increase in blood pressure, and this may be due to stimulation of sympathetic nerve and vasoconstriction (49). Pollutants derived oxidative stress in pregnant mothers can block the NO-mediated vasodilatation leading to increased blood pressure, which adversely affects offspring and leads to pre-term delivery and IUGR (50). Further, maternal PM exposure also results in inadequate fetal development (51) mainly due to abnormal blood flow to fetus. This decrease in blood flow triggers inflammatory signals to release parturition associated cytokines which results in membrane rupture, cervical ripening, myometrial contraction and ultimately contributes to premature delivery (52), plausible mechanism for PM exposure-induced complications in offspring.

## **Maternal air pollution and airways disease**

### *Development of COPD and asthma*

Though smoking is the greatest risk factor for the development of COPD, other exposures such as air pollution has emerged on the top as per WHO estimates, 14% premature deaths were a result of air pollution-induced COPD globally (53). COPD symptoms includes wheezing, breathlessness, chronic cough and further severe exacerbations, leading to reduced quality of life, with rapid decline in lung function (54-57) and risk of early mortality (58). Asthma is one of the most prevalent pediatric chronic inflammatory respiratory disease characterized by reduced airflow, airway hyperresponsiveness and airway remodeling (59). However, in case of asthma air

flow obstruction is reversible while in COPD airflow obstruction is irreversible due to the extensive loss of the small airways and this has major implication on mortality in the later stages of the disease (60). Epidemiological studies have found strong association between air pollution and incidence of asthma in children (61, 62), though, many studies also indicate that these associations begin in *in utero* itself (63-66). There is a strong evidence to believe that the maternal immunity plays a pivotal role in the fetal-infant immune response development (67). Maternal allergies may delay the onset of transformation to a non-allergic immune response to inhaled allergens in children, thereby increasing the chances for the development of allergic sensitization and/or asthma to the offspring (67). Generally, lungs begin to develop during the canalicular phase i.e. 17<sup>th</sup> –26<sup>th</sup> week of embryonic development. During gestation period, as antioxidant defense functions are subdued, thus lungs are indeed more prone to oxidative stress. Further evidence of oxidative stress was observed in mouse model on day 16, which resembles canalicular stage of human pulmonary development wherein a decrease in number of peripheral airway branching, and alveoli formation was observed (67).

It has been established that air pollutants cause oxidative stress by impairing cellular endogenous antioxidant production capabilities and thus increasing in ROS generation (68). This increase in ROS leads to mitochondrial dysfunction and carbonyl stress which can further damage the tissue by modifying the function of vital proteins such as NADH-ubiquinone reductase, succinate-ubiquinone reductase, cytochrome c oxidase (69). Increases in ROS is also known to causes ER stress leading to the upregulation of inflammatory genes as well as impairment of autophagic process thus further aggravating the pathogenesis. Autophagy is a preserved protective catabolic process which facilitate lysosomal degradation or clearance of misfolded or unwanted proteins.



Oxidative stress is the leading driver in the development of airway diseases in offspring. Because oxidative stress causes ER stress and lipid peroxidation which results in mitochondrial dysfunction, pulmonary and placental inflammation and thereby negatively affecting nutrient and fetal oxygen transport system, ultimately leading to impairment in fetal lung development (46, 70-73). Similarly, maternal exposure to air pollution causes oxidative stress and mitochondrial impairment in fetus which triggers pulmonary and placental inflammation and thereby negatively affecting nutrient and oxygen transport (73). Moreover air pollutants contains PM in which transitions metals get adsorbed which further aggravate ROS generation by the process of Fenton reaction (69). Fenton reaction lead to the subsequent activation of cellular signaling cascades, transcription and activation of inflammatory cells and release of pro-inflammatory cytokines and cause pulmonary and placental inflammation (46, 74). Rise in the ROS level can lead to oxidative modification of lipids, proteins and DNA thus causing further damage. Further, carbonyl stress also results in enhanced polyunsaturated fatty acids (PUFAs) and metabolites peroxidation (75). Collectively, ROS induce deficiency in pulmonary development in offspring as shown in Figure 1, increasing their susceptibility to acquire asthma and COPD in later in life.

### ***In utero cellular mechanisms involved in the development of airway disease***

Maternal exposure to air pollution affects the developing foetus which can be deleterious in terms of impacting offspring's lung health and consequential development of airways disease later in life. These cellular mechanisms/pathways involved are shown in Figure 2, and some of the key pathways are described below.

#### ***ER Stress***

The endoplasmic reticulum (ER) is one of the key cellular organelles which play pivotal role in biosynthetic and signaling functions in the cell. ER is associated with  $\text{Ca}^{2+}$  homeostasis as well as

Ca<sup>2+</sup> mediated signaling cascades. ER also provides the platform for the synthesis, folding, and modification of biomolecules which are to be secreted in the plasma membrane (76, 77). These processes are assisted by various intrinsic chaperones and Ca<sup>2+</sup>-binding glucose-regulated proteins 78 (GRP78) or BiP (immunoglobulin heavy chain binding protein), calreticulin, calnexin, and protein folding enzymes, such as the thioredoxin-like protein disulfide isomerase (PDI) (78). Misfolded proteins are retro-translocated to the cytosol through a process known as ER-associated protein degradation (ERAD), followed by 26S proteasomal degradation. The imbalance between ER protein folding load and capacity, leads to the aggregation of misfolded proteins in the lumen, a condition that causes ER stress. ER having defensive or adaptive response called Unfolded Protein Response (UPR) elements that are mainly responsible to decrease the overloaded protein synthesis or to maintain ER homeostasis. The UPR elements regulate ER homeostasis by harmonizing the sensual shutdown in translation process of protein along with a programmed gene transcriptional process to increase ER folding capacity. If this co-ordinated gene transcriptional fails to provide proper ER homeostasis, then the consequential stress can induce intrinsic cellular apoptotic pathways (79). Proteasomes are essentially responsible to clear the accumulated misfolded protein aggregates in ER, however, when proteasome systems are dysfunctional, the UPR actively induces autophagy (79, 80). ER stress also induce the upregulation of inflammatory genes like NFkB and secretion of cytokines such as IL-23 and type I interferon (81, 82). It has been reported that GRP78 attune UPR activation and plays a pivotal role in embryogenesis (83). As evident from animal data that mouse pulmonary growth is initiated at Embryonic Day (E) 9.5 by the process of cell proliferation and differentiation from anterior foregut endoderm. However, lung maturation stage is at E16.5 to postnatal Day 5, characterised by decrease in cell proliferation and formation of newly differentiated cell types, including alveolar epithelial type-1 (AT-1) and

alveolar epithelial type-2 (AT-2) cells in distal part of the lung (84). AT-2 cells are surfactant producing cells cuboidal in shape, having a lamellar bodies act as a precursor cells for AT-1 cells (which has the extensive surface area for gas exchange) during developmental stage (85, 86). During the saccular stage of embryonic development, enhancement of surfactant protein secretions by AT-2 cells allow lung to make it ready for postnatal respiration (87). As surfactant plays a crucial role in the reduction of surface tension, which allow the lung to inflate with maximum capacity by increasing the compliance, thus decreasing the work for breathing. However, AT-2 cells are regulated by ER homeostasis. So increase in ER stress results in apoptosis of AT-2 cells thereby, reduction in surfactant protein secretion. Thus, ER stress can lead to respiratory dysfunction in offspring (83). It has been reported that PERK and IREs regulate crosstalk between protective and apoptotic UPRs signalling (83). However, modulation of PERK upon ER stress regulates cell survival signalling by reduction of translational process *via* phosphorylation of Eukaryotic translation Initiation factor 2a (eIF-2a) and encourage apoptosis *via* the PERK-eIF-2a–ATF4–CHOP pathway (88). It has been reported that transforming growth factor (TGF)- $\beta$  having a pivotal role in the regulation of apoptosis in lung epithelial cells apart from its role in bifurcation and septae formation during pulmonary development (89). It is also reported that ER Stress and TGF- $\beta$ /Smad signalling pathways are tightly interlinked and accounts for apoptosis of AT-2 cells leading to decrease in surfactant secretion which in turn affect the alveolar epithelium formation in developing lung of offspring (83).

### *Autophagy*

Autophagy means “self-eating”. It is a fundamental and highly conserved process of lysosomal degradation and recycling of misfolded proteins or damaged cytoplasmic cellular organelles to maintain cellular homeostasis (90). Autophagic process includes engulfment of cell organelles

within the vesicular membrane referred to as autophagosome followed by lysosomal degradation with the help of lysosomal degradative enzymes (91). It plays a crucial role in both pathological and physiological conditions such as cell survival, cellular energy production and metabolism as well as the innate and adaptive immune system (92, 93). Several studies demonstrate that hereditary association of genes involved in autophagic process such as ULK1, SQSTM1, MAP1, LC3B, beclin-1, and Atg5 in asthmatics, and also found an elevated number of double membrane autophagosomes in fibroblast and epithelial cells in asthma patients in comparison with healthy population (94, 95). It has been reported that an elevated number of autophagosome in lung sample derived from COPD patients in comparison with healthy volunteers (96). Under normal physiological conditions autophagy plays a protective role in the cell survival and clearance of damaged proteins whereas in stress conditions its regulation is impaired leading to development of various diseases. It is now known that an altered cellular autophagic process is seen in stress conditions like asthma and chronic obstructive pulmonary disease (90). However, dysregulation of autophagy cause breakdown of intracellular components to generate an energy source for extracellular matrix (ECM) production, and generation of inflammatory mediators and release of pro-fibrotic signalling molecules, consequently results in airway remodelling. Furthermore, it has also been reported that selective autophagy such as mitophagy (elimination of damaged or dysfunctional mitochondria) and ciliophagy (elimination of damaged cilia or components of cilia) also comes into play in airways disease (97, 98). The transmembrane proteins Atg9 and VMP-1 (99, 100) are pivotal for autophagosome formation and the assembly formed by Atg1, Atg13, and Atg17/FIP200 proteins are essential for relocation of protein Atg9 to the autophagosome leading to double membrane autophagosome formation, a key indicator for autophagy determination. However, the mammalian target of rapamycin complex1 (mTORC1) regulate autophagy by

repressing the activity of assembly proteins Atg1-Atg13-Atg17/FIP200 and thus, inhibition of mTORC1 induces the initiation of autophagic process. Furthermore, rapamycin which blocks mTORC1 results in induction of autophagy. Bafilomycin A1 a molecule inhibits the fusion with autophagosomes as well as inhibits vacuolar type H<sup>+</sup>-ATPase leads to impairment of lysosomal degradation capability (101). The kinase Akt, which is the upstream of regulators of mTORC1 phosphorylates and inactivates TSC2 and thereby Akt activation stimulates mTORC1 and inhibits autophagy. Increased autophagy in the context of airways disease act as a pro-survival factor required for the normal recycling process of damaged proteins or organelles and there is an increase in autophagic marker in sputum and peripheral blood cells in asthma (102).

Autophagy plays a pivotal role in embryogenesis as autophagic process initiates during the fertilization phase from two cell to eight cells division while defect in beclin-1 protein lead to embryonic death (E10–E14) with neural tube formation impairment (103). As evident from clinical data, during the early gestation period i.e 7–11 weeks interstitial extra-villous trophoblast (EVT) cells occupy placental side, and trophoblastic plug restricts the maternal blood flow, resulting in hypoxic condition. Moreover, during 12–16 weeks endovascular EVT dilates the blood vessels in spiral arteries leads to elevation in intervillous space blood flow (104). In normal physiology there is increase in autophagic markers such as autophagosomes as evident from lung tissue specimens whereas under pathological conditions decrease in autophagosomes results in impairment of lysosomal degradation. However, autophagic inhibition in trophoblasts induces inadequate placental blood supply in initial phase of gestation period lead to hypoxic condition in embryo. Furthermore, releases of anti-angiogenic factors from villi due to hypoxic conditions aggravate the dysregulation of trophoblasts by inhibiting autophagic process.

Thus, autophagic inhibition causes impairment of homeostasis in trophoblast cells. However, controlled regulation of autophagic process is necessary for reproduction. On the contrary, dysregulation of functional autophagy negatively impact offspring health because inadequate placental blood flow affect lung development in offspring (46, 104). In this way, dysregulation of autophagy lead to impaired lung development, ultimately leading to development of airways disease.

### *Mitochondrial damage*

Mitochondria are the leading subcellular organelle from which ROS is generated under physiological conditions during catalyzation of ATP production, which is linked to electron transport chain that occurs in the inner mitochondrial membrane. The mitochondrial respiratory chain has four complexes in the inner membrane where electrons move in a synchronous manner i.e. move from a high to a low redox potential resulting in the generation of mitochondrial membrane potential (105-107). Air pollution which has both organic and inorganic compounds is capable of generating ROS which interferes in the electron transfer chain system by disrupting the Q cycle that operates between complexes I and III resulting in mitochondrial superoxide production (108, 109). Air pollutants also have the ability to disrupt mitochondrial permeability transition pore (110), a crucial molecule that regulates mitochondrial respiration and regulate the cellular apoptosis in a controlled manner (111, 112). However, disruption of the mitochondrial outer membrane by air pollutants, lead to release of several pro-apoptotic proteins such as cytochrome c, second mitochondria-derived activator of caspases (SMAC) and apoptosis inducing factor (AIF) into the cytosol where they activate apoptotic signalling pathways ultimately leading to programmed cell death (112, 113). Besides direct effects of air pollutants on mitochondria, the physiology of this organelle can also be altered indirectly by increasing ROS generation and  $\text{Ca}^{2+}$

flux in the cell. Even free intracellular calcium  $[Ca^{2+}]_i$  plays an important role in regulating the opening and closure of the mitochondrial permeability transition pore (114). Moreover, ROS such as  $H_2O_2$  can induce rise of  $[Ca^{2+}]_i$  in a various cells by either inhibiting sarco/endoplasmic reticulum  $Ca^{2+}$ -ATPase (SERCA) or plasma membrane  $Ca^{2+}$ -ATPase (PMCA) leading to the activation of 1, 3, 5-trisphosphate (IP3) receptors (115, 116). Various constituents in air pollution can directly target mitochondrial membrane and cause structural damage (69). The process in which non-functional or damaged mitochondria are cleared *via* mitochondria-selective autophagy are known as 'mitophagy' and has been reported as pivotal mechanism in the development of airways disease (117).

## Conclusion

Air pollution has become a growing concern in recent years in India and globally causing enormous burden to the society as well as loss of life. It has been well established that air pollutant particularly PM penetrate deep into the lungs causes pathological changes leading to impaired pulmonary function. Epidemiological studies demonstrate that exposure to PM increases oxidative stress leading to serious health effects. Similar mechanisms are involved in maternal air pollution exposure-induced health effects on offspring that is a result of increased oxidative stress leading to oxidative damage of biomolecules, placental inflammation and induction of autophagy. Moreover, oxidative stress also lead to mitochondrial damage, causing release of pro-apoptotic factors leading to cell death. Thus, a better therapeutic approach would be to target air pollution induced oxidative stress by enhancing endogenous antioxidant mechanisms in the cell. Further, targeting ER stress by autophagy inhibitors could also be a vital approach for future research into preventing the development of pollutant induced airways disease. Finally, air pollution induced

adverse health outcomes could be prevented by comprehensive approached that eliminates or reduces the burden of PM both indoors and outdoors.



**Funding**

This work was supported by the Rebecca L. Cooper Foundation and the UTS Chancellors Fellowship (to PS), the Clifford Craig Foundation (to SSS).

**Competing interests**

The authors declare that there are no competing interests associated with the manuscript.

## References

1. Kelly FJ, Fussell JC. Air pollution and public health: emerging hazards and improved understanding of risk. *Environmental geochemistry and health*. 2015;37(4):631-49.
2. Forouzanfar MH, Afshin A, Alexander LT, Anderson HR, Bhutta ZA, Biryukov S, et al. Global, regional, and national comparative risk assessment of 79 behavioural, environmental and occupational, and metabolic risks or clusters of risks, 1990–2015: a systematic analysis for the Global Burden of Disease Study 2015. *The Lancet*. 2016;388(10053):1659-724.
3. Kim K-H, Kabir E, Kabir S. A review on the human health impact of airborne particulate matter. *Environment international*. 2015;74:136-43.
4. Parker JD, Woodruff TJ, Basu R, Schoendorf KC. Air pollution and birth weight among term infants in California. *Pediatrics*. 2005;115(1):121-8.
5. Organization WH. World Health Organization. 2018.
6. Arbex MA, Santos UdP, Martins LC, Saldiva PHN, Pereira LAA, Braga ALF. Air pollution and the respiratory system. *Jornal Brasileiro de Pneumologia*. 2012;38(5):643-55.
7. Dadvand P, Parker J, Bell ML, Bonzini M, Brauer M, Darrow LA, et al. Maternal exposure to particulate air pollution and term birth weight: a multi-country evaluation of effect and heterogeneity. *Environmental health perspectives*. 2013;121(3):267.
8. Shah PS, Balkhair T, births KSGoDoPL. Air pollution and birth outcomes: a systematic review. *Environment international*. 2011;37(2):498-516.
9. Stieb DM, Chen L, Eshoul M, Judek S. Ambient air pollution, birth weight and preterm birth: a systematic review and meta-analysis. *Environmental research*. 2012;117:100-11.
10. Zhang B, Liang S, Zhao J, Qian Z, Bassig BA, Yang R, et al. Maternal exposure to air pollutant PM 2.5 and PM 10 during pregnancy and risk of congenital heart defects. *Journal of Exposure Science and Environmental Epidemiology*. 2016;26(4):422.
11. Rich DQ, Demissie K, Lu S-E, Kamat L, Wartenberg D, Rhoads GG. Ambient air pollutant concentrations during pregnancy and the risk of fetal growth restriction. *Journal of Epidemiology & Community Health*. 2009;jech. 2008.082792.
12. Choi H, Rauh V, Garfinkel R, Tu Y, Perera FP. Prenatal exposure to airborne polycyclic aromatic hydrocarbons and risk of intrauterine growth restriction. *Environmental health perspectives*. 2008;116(5):658.
13. Hyland A, Piazza KM, Hovey KM, Ockene JK, Andrews CA, Rivard C, et al. Associations of lifetime active and passive smoking with spontaneous abortion, stillbirth and tubal ectopic pregnancy: a cross-sectional analysis of historical data from the Women's Health Initiative. *Tobacco control*. 2015;24(4):328-35.
14. Wu T, Hu Y, Chen C, Yang F, Li Z, Fang Z, et al. Passive smoking, metabolic gene polymorphisms, and infant birth weight in a prospective cohort study of Chinese women. *American journal of epidemiology*. 2007;166(3):313-22.
15. Siddique S, Banerjee M, Ray MR, Lahiri T. Attention-deficit hyperactivity disorder in children chronically exposed to high level of vehicular pollution. *European journal of pediatrics*. 2011;170(7):923-9.
16. Newman NC, Ryan P, LeMasters G, Levin L, Bernstein D, Hershey GKK, et al. Traffic-related air pollution exposure in the first year of life and behavioral scores at 7 years of age. *Environmental health perspectives*. 2013;121(6):731.

17. Gong T, Almqvist C, Bölte S, Lichtenstein P, Anckarsäter H, Lind T, et al. Exposure to air pollution from traffic and neurodevelopmental disorders in Swedish twins. *Twin Research and Human Genetics*. 2014;17(6):553-62.
18. Xu X, Ha SU, Basnet R. A review of epidemiological research on adverse neurological effects of exposure to ambient air pollution. *Frontiers in public health*. 2016;4:157.
19. Jedrychowski W, Perera F, Maugeri U, Mrozek-Budzyn D, Miller RL, Flak E, et al. Effects of prenatal and perinatal exposure to fine air pollutants and maternal fish consumption on the occurrence of infantile eczema. *International archives of allergy and immunology*. 2011;155(3):275-81.
20. Lavigne É, Bélair M-A, Duque DR, Do MT, Stieb DM, Hystad P, et al. Effect modification of perinatal exposure to air pollution and childhood asthma incidence. *European Respiratory Journal*. 2018:1701884.
21. Juda-Rezler K, Reizer M, Oudinet J-P. Determination and analysis of PM10 source apportionment during episodes of air pollution in Central Eastern European urban areas: The case of wintertime 2006. *Atmospheric Environment*. 2011;45(36):6557-66.
22. Apte K, Salvi S. Household air pollution and its effects on health. *F1000Research*. 2016;5.
23. Atkinson RW, Fuller GW, Anderson HR, Harrison RM, Armstrong B. Urban ambient particle metrics and health: a time-series analysis. *Epidemiology*. 2010:501-11.
24. Srimuruganandam B, Nagendra SS. Source characterization of PM10 and PM2. 5 mass using a chemical mass balance model at urban roadside. *Science of the total environment*. 2012;433:8-19.
25. Misra C, Geller MD, Shah P, Sioutas C, Solomon PA. Development and Evaluation of a Continuous Coarse (PM10–PM25) Particle Monitor. *Journal of the Air & Waste Management Association*. 2001;51(9):1309-17.
26. Brauer M, Freedman G, Frostad J, Van Donkelaar A, Martin RV, Dentener F, et al. Ambient air pollution exposure estimation for the global burden of disease 2013. *Environmental science & technology*. 2015;50(1):79-88.
27. Pant P, Guttikunda SK, Peltier RE. Exposure to particulate matter in India: A synthesis of findings and future directions. *Environmental research*. 2016;147:480-96.
28. Dey S, Di Girolamo L, van Donkelaar A, Tripathi S, Gupta T, Mohan M. Variability of outdoor fine particulate (PM2. 5) concentration in the Indian Subcontinent: A remote sensing approach. *Remote Sensing of Environment*. 2012;127:153-61.
29. Lim SS, Vos T, Flaxman AD, Danaei G, Shibuya K, Adair-Rohani H, et al. A comparative risk assessment of burden of disease and injury attributable to 67 risk factors and risk factor clusters in 21 regions, 1990–2010: a systematic analysis for the Global Burden of Disease Study 2010. *The Lancet*. 2012;380(9859):2224-60.
30. Balakrishnan K, Ramaswamy P, Sambandam S, Thangavel G, Ghosh S, Johnson P, et al. Air pollution from household solid fuel combustion in India: an overview of exposure and health related information to inform health research priorities. *Global health action*. 2011;4(1):5638.
31. Barbier EB. *Natural resources and economic development*: Cambridge University Press; 2007.
32. Guaita R, Pichiule M, Maté T, Linares C, Díaz J. Short-term impact of particulate matter (PM2. 5) on respiratory mortality in Madrid. *International journal of environmental health research*. 2011;21(4):260-74.
33. Joseph A, Sawant A, Srivastava A. PM10 and its impacts on health-a case study in Mumbai. *International journal of environmental health research*. 2003;13(2):207-14.

34. Balakrishnan K, Sambandam S, Ramaswamy P, Ghosh S, Venkatesan V, Thangavel G, et al. Establishing integrated rural–urban cohorts to assess air pollution-related health effects in pregnant women, children and adults in Southern India: an overview of objectives, design and methods in the Tamil Nadu Air Pollution and Health Effects (TAPHE) study. *BMJ open*. 2015;5(6):e008090.
35. Tobollik M, Razum O, Wintermeyer D, Plass D. Burden of Outdoor Air Pollution in Kerala, India—A First Health Risk Assessment at State Level. *International journal of environmental research and public health*. 2015;12(9):10602-19.
36. Demedts IK, Demoor T, Bracke KR, Joos GF, Brusselle GG. Role of apoptosis in the pathogenesis of COPD and pulmonary emphysema. *Respiratory research*. 2006;7(1):53.
37. Siddique S, Banerjee M, Ray MR, Lahiri T. Air pollution and its impact on lung function of children in Delhi, the capital city of India. *Water, Air, & Soil Pollution*. 2010;212(1-4):89-100.
38. Kumar R, Nagar JK, Goel N, Kumar P, Kushwah AS, Gaur SN. Indoor air pollution and asthma in children at Delhi, India. *Advances in Respiratory Medicine*. 2015;83(4):275-82.
39. Murlidhar V. Case Report: An 11-year-old boy with silico-tuberculosis attributable to secondary exposure to sandstone mining in central India. *BMJ case reports*. 2015;2015.
40. Perez L, Tobías A, Querol X, Pey J, Alastuey A, Díaz J, et al. Saharan dust, particulate matter and cause-specific mortality: a case–crossover study in Barcelona (Spain). *Environment international*. 2012;48:150-5.
41. Sun X, Luo X, Zhao C, Zhang B, Tao J, Yang Z, et al. The associations between birth weight and exposure to fine particulate matter (PM<sub>2.5</sub>) and its chemical constituents during pregnancy: A meta-analysis. *Environmental pollution*. 2016;211:38-47.
42. Perera FP, Whyatt RM, Jedrychowski W, Rauh V, Manchester D, Santella RM, et al. Recent developments in molecular epidemiology: a study of the effects of environmental polycyclic aromatic hydrocarbons on birth outcomes in Poland. *American journal of epidemiology*. 1998;147(3):309-14.
43. Dejmeek J, Solanský I, Benes I, Lenícek J, Srám RJ. The impact of polycyclic aromatic hydrocarbons and fine particles on pregnancy outcome. *Environmental health perspectives*. 2000;108(12):1159.
44. Risom L, Møller P, Loft S. Oxidative stress-induced DNA damage by particulate air pollution. *Mutation Research/Fundamental and Molecular Mechanisms of Mutagenesis*. 2005;592(1):119-37.
45. Bobak M. Outdoor air pollution, low birth weight, and prematurity. *Environmental health perspectives*. 2000;108(2):173.
46. Kannan S, Misra DP, Dvonch JT, Krishnakumar A. Exposures to airborne particulate matter and adverse perinatal outcomes: a biologically plausible mechanistic framework for exploring potential. *Ciencia & saude coletiva*. 2007;12:1591-602.
47. Valkonen V-P, Päivä H, Salonen JT, Lakka TA, Lehtimäki T, Laakso J, et al. Risk of acute coronary events and serum concentration of asymmetrical dimethylarginine. *The Lancet*. 2001;358(9299):2127-8.
48. Leone A, Moncada S, Vallance P, Calver A, Collier J. Accumulation of an endogenous inhibitor of nitric oxide synthesis in chronic renal failure. *The Lancet*. 1992;339(8793):572-5.
49. Ibalá-Mulli A, Stieber J, Wichmann H-E, Koenig W, Peters A. Effects of air pollution on blood pressure: a population-based approach. *American Journal of Public Health*. 2001;91(4):571.
50. Misra DP. The effect of the pregnancy-induced hypertension on fetal growth: a review of the literature. *Paediatric and perinatal epidemiology*. 1996;10(3):244-63.

51. Duvekot JJ, Cheriex EC, Pieters FA, Peeters LL. Severely impaired fetal growth is preceded by maternal hemodynamic maladaptation in very early pregnancy. *Acta obstetrica et gynecologica Scandinavica*. 1995;74(9):693-7.
52. Keelan J, Blumenstein M, Helliwell R, Sato T, Marvin K, Mitchell M. Cytokines, prostaglandins and parturition—a review. *Placenta*. 2003;24:S33-S46.
53. Hansel NN, McCormack MC, Kim V. The effects of air pollution and temperature on COPD. *COPD: Journal of Chronic Obstructive Pulmonary Disease*. 2016;13(3):372-9.
54. Donaldson G, Seemungal T, Bhowmik A, Wedzicha J. Relationship between exacerbation frequency and lung function decline in chronic obstructive pulmonary disease. *Thorax*. 2002;57(10):847-52.
55. Seemungal TA, Donaldson GC, Paul EA, Bestall JC, Jeffries DJ, Wedzicha JA. Effect of exacerbation on quality of life in patients with chronic obstructive pulmonary disease. *American journal of respiratory and critical care medicine*. 1998;157(5):1418-22.
56. Eapen MS, Myers S, Walters EH, Sohal SS. Airway inflammation in chronic obstructive pulmonary disease (COPD): a true paradox. *Expert review of respiratory medicine*. 2017;11(10):827-39. Epub 2017/07/27.
57. Sohal S, Ward C, Danial W, Wood-Baker R, Walters E. Recent advances in understanding inflammation and remodeling in the airways in chronic obstructive pulmonary disease. *Expert Rev Respir Med*. 2013;7(3):275 - 88.
58. Connors Jr AF, Dawson NV, Thomas C, Harrell Jr FE, Desbiens N, Fulkerson WJ, et al. Outcomes following acute exacerbation of severe chronic obstructive lung disease. The SUPPORT investigators (Study to Understand Prognoses and Preferences for Outcomes and Risks of Treatments). *American journal of respiratory and critical care medicine*. 1996;154(4):959-67.
59. Sahiner UM, Birben E, Erzurum S, Sackesen C, Kalayci O. Oxidative stress in asthma. *World Allergy Organization Journal*. 2011;4(10):151.
60. Hogg JC, Pare PD, Hackett TL. The Contribution of Small Airway Obstruction to the Pathogenesis of Chronic Obstructive Pulmonary Disease. *Physiological reviews*. 2017;97(2):529-52. Epub 2017/02/06.
61. Khreis H, Kelly C, Tate J, Parslow R, Lucas K, Nieuwenhuijsen M. Exposure to traffic-related air pollution and risk of development of childhood asthma: a systematic review and meta-analysis. *Environment international*. 2017;100:1-31.
62. Gasana J, Dillikar D, Mendy A, Forno E, Vieira ER. Motor vehicle air pollution and asthma in children: a meta-analysis. *Environmental research*. 2012;117:36-45.
63. Leon Hsu H-H, Mathilda Chiu Y-H, Coull BA, Kloog I, Schwartz J, Lee A, et al. Prenatal particulate air pollution and asthma onset in urban children. Identifying sensitive windows and sex differences. *American journal of respiratory and critical care medicine*. 2015;192(9):1052-9.
64. Clark NA, Demers PA, Karr CJ, Koehoorn M, Lencar C, Tamburic L, et al. Effect of early life exposure to air pollution on development of childhood asthma. *Environmental health perspectives*. 2010;118(2):284.
65. Sbihi H, Tamburic L, Koehoorn M, Brauer M. Perinatal air pollution exposure and development of asthma from birth to age 10 years. *European Respiratory Journal*. 2016;ERJ-00746-2015.
66. Deng Q, Lu C, Li Y, Sundell J, Norbäck D. Exposure to outdoor air pollution during trimesters of pregnancy and childhood asthma, allergic rhinitis, and eczema. *Environmental research*. 2016;150:119-27.

67. Barrett EG. Maternal influence in the transmission of asthma susceptibility. *Pulmonary pharmacology & therapeutics*. 2008;21(3):474-84.
68. Kirkham PA, Barnes PJ. Oxidative stress in COPD. *Chest*. 2013;144(1):266-73.
69. Xia T, Kovochich M, Nel AE. Impairment of mitochondrial function by particulate matter (PM) and their toxic components: implications for PM-induced cardiovascular and lung disease. *Front Biosci*. 2007;12(1):1238.
70. van der Toorn M, Rezayat D, Kauffman HF, Bakker SJ, Gans RO, Koëter GH, et al. Lipid-soluble components in cigarette smoke induce mitochondrial production of reactive oxygen species in lung epithelial cells. *American Journal of Physiology-Lung Cellular and Molecular Physiology*. 2009;297(1):L109-L14.
71. Aguilera-Aguirre L, Bacsí A, Saavedra-Molina A, Kurosky A, Sur S, Boldogh I. Mitochondrial dysfunction increases allergic airway inflammation. *The Journal of Immunology*. 2009;jimmunol. 0900228.
72. Pinamonti S, Leis M, Barbieri A, Leoni D, Muzzoli M, Sostero S, et al. Detection of xanthine oxidase activity products by EPR and HPLC in bronchoalveolar lavage fluid from patients with chronic obstructive pulmonary disease. *Free Radical Biology and Medicine*. 1998;25(7):771-9.
73. Aaron SD, Angel JB, Lunau M, Wright K, Fex C, Le Saux N, et al. Granulocyte inflammatory markers and airway infection during acute exacerbation of chronic obstructive pulmonary disease. *American journal of respiratory and critical care medicine*. 2001;163(2):349-55.
74. Wood L, Gibson P, Garg M. Biomarkers of lipid peroxidation, airway inflammation and asthma. *European Respiratory Journal*. 2003;21(1):177-86.
75. Huang S-K, Zhang Q, Qiu Z, Chung KF. Mechanistic impact of outdoor air pollution on asthma and allergic diseases. *Journal of thoracic disease*. 2015;7(1):23.
76. Berridge MJ. The endoplasmic reticulum: a multifunctional signaling organelle. *Cell calcium*. 2002;32(5-6):235-49.
77. Görlach A, Klappa P, Kietzmann DT. The endoplasmic reticulum: folding, calcium homeostasis, signaling, and redox control. *Antioxidants & redox signaling*. 2006;8(9-10):1391-418.
78. Sevier CS, Kaiser CA. Ero1 and redox homeostasis in the endoplasmic reticulum. *Biochimica et Biophysica Acta (BBA)-Molecular Cell Research*. 2008;1783(4):549-56.
79. Szegezdi E, Logue SE, Gorman AM, Samali A. Mediators of endoplasmic reticulum stress-induced apoptosis. *EMBO reports*. 2006;7(9):880-5.
80. Kouroku Y, Fujita E, Tanida I, Ueno T, Isoai A, Kumagai H, et al. ER stress (PERK/eIF2 $\alpha$  phosphorylation) mediates the polyglutamine-induced LC3 conversion, an essential step for autophagy formation. *Cell death and differentiation*. 2007;14(2):230.
81. Smith JA, Turner MJ, DeLay ML, Klenk EI, Sowders DP, Colbert RA. Endoplasmic reticulum stress and the unfolded protein response are linked to synergistic IFN- $\beta$  induction via X-box binding protein 1. *European journal of immunology*. 2008;38(5):1194-203.
82. Kim SR, Im Kim D, Kang MR, Lee KS, Park SY, Jeong JS, et al. Endoplasmic reticulum stress influences bronchial asthma pathogenesis by modulating nuclear factor  $\kappa$ B activation. *Journal of Allergy and Clinical Immunology*. 2013;132(6):1397-408. e11.
83. Flodby P, Li C, Liu Y, Wang H, Marconett CN, Laird-Offringa IA, et al. The 78-kD glucose-regulated protein regulates endoplasmic reticulum homeostasis and distal epithelial cell



survival during lung development. *American journal of respiratory cell and molecular biology*. 2016;55(1):135-49.

84. Morrissey EE, Hogan BL. Preparing for the first breath: genetic and cellular mechanisms in lung development. *Developmental cell*. 2010;18(1):8-23.

85. Adamson I, Bowden D. Derivation of type 1 epithelium from type 2 cells in the developing rat lung. *Laboratory investigation; a journal of technical methods and pathology*. 1975;32(6):736-45.

86. Barkauskas CE, Counce MJ, Rackley CR, Bowie EJ, Keene DR, Stripp BR, et al. Type 2 alveolar cells are stem cells in adult lung. *The Journal of clinical investigation*. 2013;123(7):3025-36.

87. Mimura N, Hamada H, Kashio M, Jin H, Toyama Y, Kimura K, et al. Aberrant quality control in the endoplasmic reticulum impairs the biosynthesis of pulmonary surfactant in mice expressing mutant BiP. *Cell death and differentiation*. 2007;14(8):1475.

88. Tabas I, Ron D. Integrating the mechanisms of apoptosis induced by endoplasmic reticulum stress. *Nature cell biology*. 2011;13(3):184.

89. Sureshbabu A, Syed MA, Boddupalli CS, Dhodapkar MV, Homer RJ, Minoo P, et al. Conditional overexpression of TGF $\beta$ 1 promotes pulmonary inflammation, apoptosis and mortality via TGF $\beta$ 2 in the developing mouse lung. *Respiratory research*. 2015;16(1):4.

90. Kota A, Deshpande D, Haghi M, Oliver B, Sharma P. Autophagy and airway fibrosis: Is there a link? *F1000Research*. 2017;6.

91. Mizushima N, Levine B, Cuervo AM, Klionsky DJ. Autophagy fights disease through cellular self-digestion. *Nature*. 2008;451(7182):1069.

92. Levine B, Klionsky DJ. Development by self-digestion: molecular mechanisms and biological functions of autophagy. *Developmental cell*. 2004;6(4):463-77.

93. Zeki A, Yeganeh B, Kenyon N, Post M, Ghavami S. Autophagy in airway diseases: a new frontier in human asthma? *Allergy*. 2016;71(1):5-14.

94. Poon AH, Chouiali F, Tse SM, Litonjua AA, Hussain SN, Bagloli CJ, et al. Genetic and histologic evidence for autophagy in asthma pathogenesis. *Journal of Allergy and Clinical Immunology*. 2012;129(2):569-71.

95. Poon A, Eidelman D, Laprise C, Hamid Q. ATG5, autophagy and lung function in asthma. *Autophagy*. 2012;8(4):694-5.

96. Chen Z-H, Kim HP, Sciurba FC, Lee S-J, Feghali-Bostwick C, Stolz DB, et al. Egr-1 regulates autophagy in cigarette smoke-induced chronic obstructive pulmonary disease. *PloS one*. 2008;3(10):e3316.

97. Mizumura K, Cloonan SM, Nakahira K, Bhashyam AR, Cervo M, Kitada T, et al. Mitophagy-dependent necroptosis contributes to the pathogenesis of COPD. *The Journal of clinical investigation*. 2014;124(9):3987-4003.

98. Lam HC, Cloonan SM, Bhashyam AR, Haspel JA, Singh A, Sathirapongsasuti JF, et al. Histone deacetylase 6-mediated selective autophagy regulates COPD-associated cilia dysfunction. *The Journal of clinical investigation*. 2013;123(12):5212-30.

99. Webber JL, Young AR, Tooze SA. Atg9 trafficking in mammalian cells. *Autophagy*. 2007;3(1):54-6.

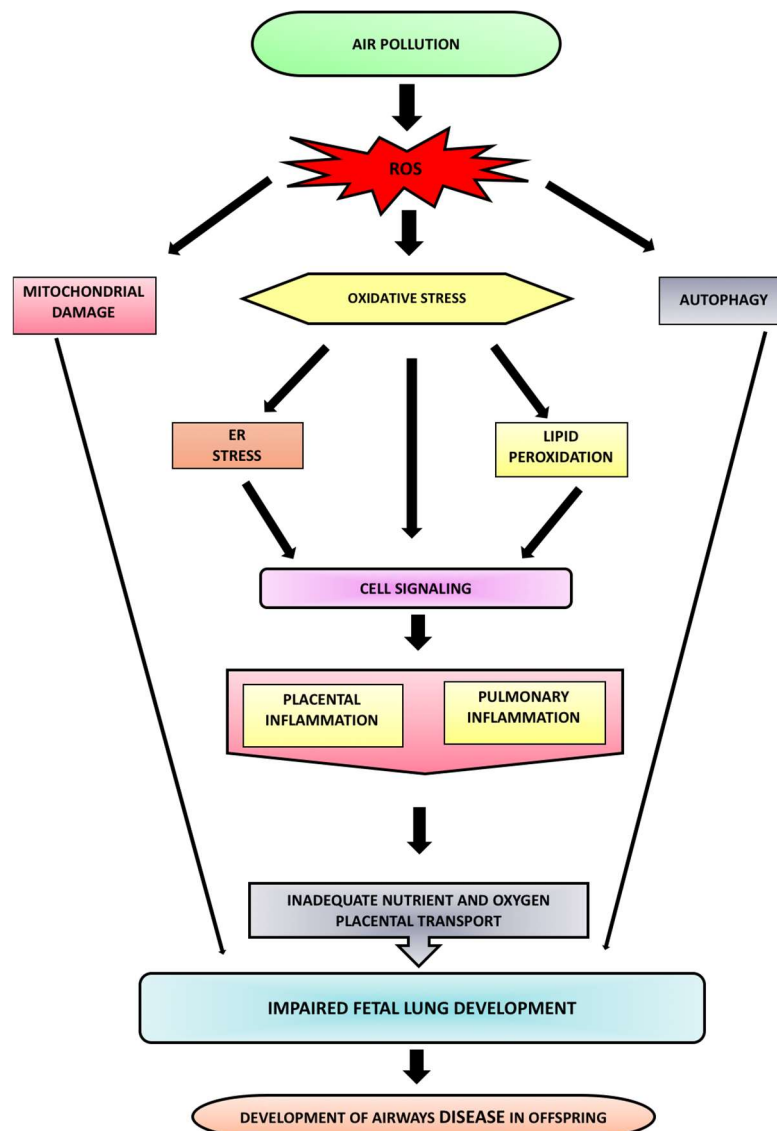
100. Ropolo A, Grasso D, Pardo R, Sacchetti ML, Archange C, Re AL, et al. The pancreatitis-induced vacuole membrane protein 1 triggers autophagy in mammalian cells. *Journal of Biological Chemistry*. 2007.

101. Yang Y-p, Hu L-f, Zheng H-f, Mao C-j, Hu W-d, Xiong K-p, et al. Application and interpretation of current autophagy inhibitors and activators. *Acta Pharmacologica Sinica*. 2013;34(5):625.
102. Ban GY, Pham D, Trinh T, Lee SI, Suh DH, Yang EM, et al. Autophagy mechanisms in sputum and peripheral blood cells of patients with severe asthma: a new therapeutic target. *Clinical & Experimental Allergy*. 2016;46(1):48-59.
103. Fimia GM, Stoykova A, Romagnoli A, Giunta L, Di Bartolomeo S, Nardacci R, et al. Ambra1 regulates autophagy and development of the nervous system. *Nature*. 2007;447(7148):1121.
104. Nakashima A, Aoki A, Kusabiraki T, Shima T, Yoshino O, Cheng SB, et al. Role of autophagy in oocytogenesis, embryogenesis, implantation, and pathophysiology of pre-eclampsia. *Journal of Obstetrics and Gynaecology Research*. 2017;43(4):633-43.
105. Raha S, Robinson BH. Mitochondria, oxygen free radicals, disease and ageing. *Trends in biochemical sciences*. 2000;25(10):502-8.
106. Andreyev AY, Kushnareva YE, Starkov A. Mitochondrial metabolism of reactive oxygen species. *Biochemistry (Moscow)*. 2005;70(2):200-14.
107. Papa S, Skulachev V. Reactive oxygen species, mitochondria, apoptosis and aging. *Detection of Mitochondrial Diseases*: Springer; 1997. p. 305-19.
108. Walter L, Nogueira V, Leverve X, Heitz M-P, Bernardi P, Fontaine E. Three classes of ubiquinone analogs regulate the mitochondrial permeability transition pore through a common site. *Journal of Biological Chemistry*. 2000;275(38):29521-7.
109. Fontaine E, Ichas F, Bernardi P. A ubiquinone-binding site regulates the mitochondrial permeability transition pore. *Journal of Biological Chemistry*. 1998;273(40):25734-40.
110. Xia T, Korge P, Weiss JN, Li N, Venkatesen MI, Sioutas C, et al. Quinones and aromatic chemical compounds in particulate matter induce mitochondrial dysfunction: implications for ultrafine particle toxicity. *Environmental health perspectives*. 2004;112(14):1347.
111. Bernardi P, Petronilli V, Di Lisa F, Forte M. A mitochondrial perspective on cell death. *Trends in biochemical sciences*. 2001;26(2):112-7.
112. Zamzami N, Kroemer G. The mitochondrion in apoptosis: how Pandora's box opens. *Nature Reviews Molecular Cell Biology*. 2001;2(1):67.
113. Jiang X, Wang X. Cytochrome C-mediated apoptosis. *Annual review of biochemistry*. 2004;73(1):87-106.
114. Ichas F, Mazat J-P. From calcium signaling to cell death: two conformations for the mitochondrial permeability transition pore. Switching from low-to high-conductance state. *Biochimica et Biophysica Acta (BBA)-Bioenergetics*. 1998;1366(1-2):33-50.
115. Redondo PC, Salido GM, Rosado JA, Pariente JA. Effect of hydrogen peroxide on Ca<sup>2+</sup> mobilisation in human platelets through sulphhydryl oxidation dependent and independent mechanisms. *Biochemical pharmacology*. 2004;67(3):491-502.
116. Brookes PS, Yoon Y, Robotham JL, Anders M, Sheu S-S. Calcium, ATP, and ROS: a mitochondrial love-hate triangle. *American Journal of Physiology-Cell Physiology*. 2004;287(4):C817-C33.
117. Rubinsztein DC, Mariño G, Kroemer G. Autophagy and aging. *Cell*. 2011;146(5):682-95.

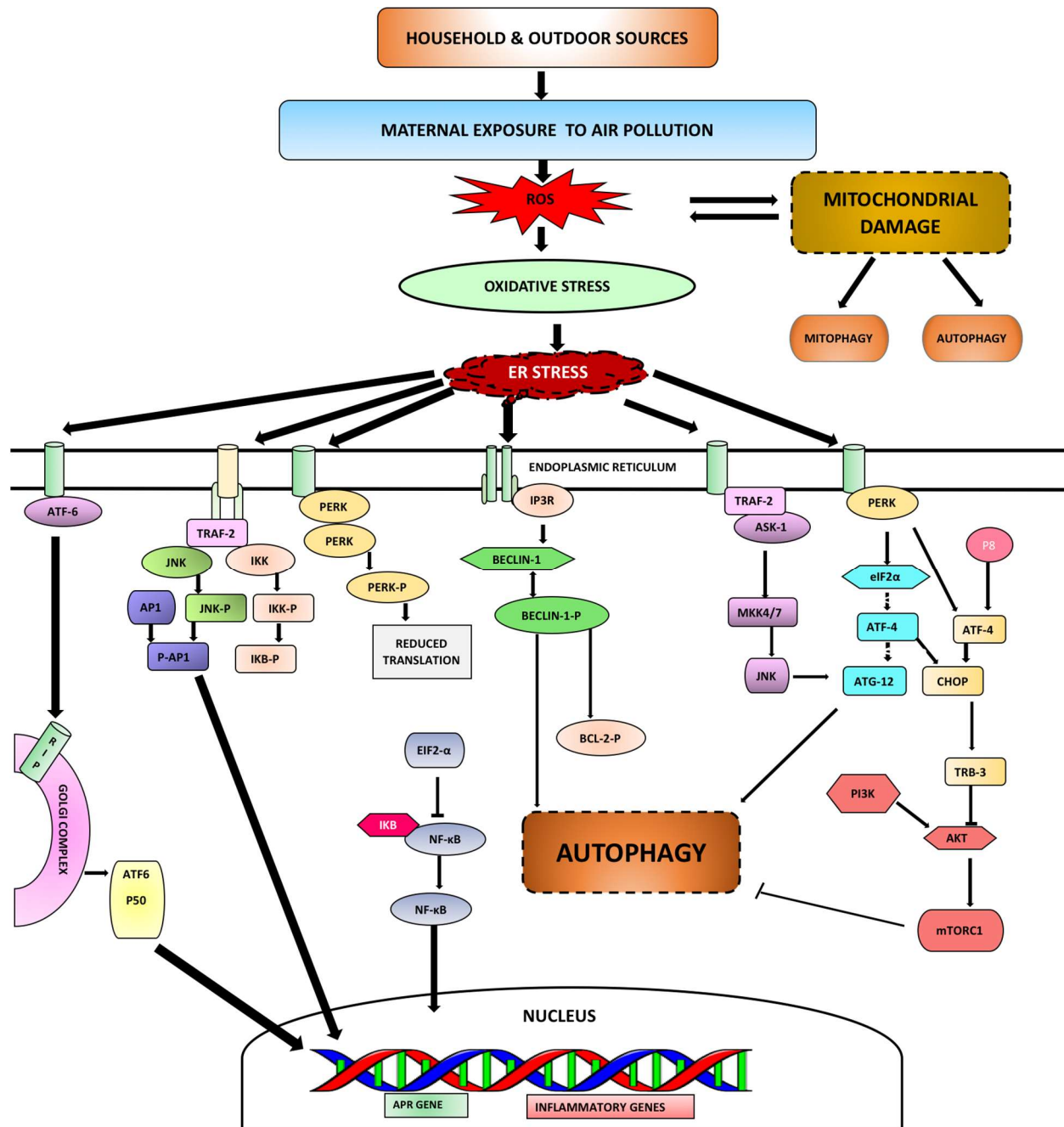


S. No.	Author	Study Design	Sample Size	Exposure	Parameter studied	Comments and association
1	Padhy et. al 2009 (36)	Case control	Control (n = 105) Biomass user (n = 115)	Biomass combustion smoke	Wheeze	OD=2.15 (1.80–4.75)
					Sneezing	OD=2.47 (1.92–3.50)**
						Control
						Case
					Haemoglobin (g/dl)	13.5 ± 0.8
						11.0 ± 0.5*
					RBC (× 10 <sup>6</sup> /μl)	4.3 ± 0.6
						3.8 ± 0.4*
					WBC (× 10 <sup>3</sup> /μl)	6.2 ± 0.2
						9.1 ± 0.2***
					Eosinophil (×10 <sup>3</sup> /μl)	0.2 ± 0.05
						0.9 ± 0.1**
2	Siddique et.al 2010 (37)	Case Control	Case=969 (school children of Delhi), Control=850 (students from rural area)	PM10	ADHD	OD=2.066 (1.079–3.958)
3	Kumar et al 2015 (38)	Cohort	3104 children	Indoor SPM levels	Asthma	Indoor SPM level was significantly ( $p < 0.001$ ) higher in the asthmatic children's houses.
4	Murlidhar et. al 2015 (39)	Case report	11-year-old boy malnourished	Secondary exposure to sandstone mining	Silico-tuberculosis	Mother started working in the mines soon after her marriage and the family lives close to the mines

**Table 1.** List of studies that has been carried out in Indian Population on air pollution



**Figure 1. Plausible mechanisms for the development airways disease in offspring.** Air pollution induces ROS generation which leads to oxidative stress followed by ER stress and lipid peroxidation resulting in up regulation of inflammatory genes. This causes pulmonary and placental inflammation and thereby negatively affect nutrient and fetal oxygen transport system. Excessive ROS generation also results in mitochondrial damage and induction of autophagy. Thus, oxidative stress, ER stress, autophagy and mitochondrial damage causes impaired fetal lung development leading to development of airways disease later in life.



**Figure 2. Cellular mechanisms involved upon maternal exposure to air pollution.** Air pollutants enter into the lungs while breathing which increases ROS generation resulting in oxidative stress. Increases in ROS cause mitochondrial damage which further initiates cell death either by apoptosis or mitophagy. Oxidative stress induces ER stress leading to the stimulation of IRE1α kinase which interact with tumor-necrosis factor-α (TNF-α) and receptor-associated factor 2 (TRAF2) followed by interaction with Jun N-terminal kinase (JNK) thus stimulating several transcription factors and apoptotic factors. JNK also stimulate the inflammatory genes expression

by phosphorylating the transcription factor activator protein-1 (AP-1) which enhances inflammation by regulating the transcription of cytokines and chemokines. Similarly, EIF2 $\alpha$  interaction with IKK activates NF $\kappa$ B resulting in its transport to the nucleus, where it activates the inflammatory gene transcription. ATF-6 along with p50 protein activate the transcription of APR genes resulting in systemic inflammation. PERK protein upon phosphorylation lead to reduced translation. Inositol 1,4,5-trisphosphate receptor (IP3R) interaction with Beclin-1 protein upon phosphorylation inhibit autophagy. However, inhibition of IP3R initiates detachment with Beclin-1 leading to autophagy in absence of Ca<sup>2+</sup>. Interaction of TRAF2 with ASK-1 protein lead to JNK activation which promotes Bcl-2 phosphorylation, leading to detachment with Beclin-1. PERK interaction to phosphorylated eIF2 $\alpha$  can lead to autophagy thorough an interaction of ATF4 dependent Atg12 protein expression. Alternatively, P8 protein interaction with ATF4 stimulates the up-regulation of pseudokinase TRB3 which further leads to activation of autophagy by inhibition of the Akt/mTORC1 complex.