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2 Sunlight Assisted Photocatalytic Degradation of

3 Ciprofloxacin in Water Using Fe Doped ZnO

4 Nanoparticles for Potential Public Health

5 Applications

- Sourav Das ¹‡, Soumen Ghosh ¹‡, Ananyo Jyoti Misra¹, Amrita Mishra ¹, Ashok J. Tamhankar ¹²⁵,
 Cecilia Stålsby Lundborg ²⁵, and Suraj K. Tripathy ¹³³ §
- School of Biotechnology, Kalinga Institute of Industrial Technology (KIIT), Bhubaneswar 751024, India;
 sddassourav52@gmail.com (S.D.); soumen.cbel@gmail.com (S.G.); ananyomisra@gmail.com (A.J.M.);
 amritamishrabio@gmail.com (A.M.)
- Department of Public Health Sciences, KarolinskaInstitutet, SE 17177 Stockholm, Sweden;
 cecilia.stalsby.lundborg@ki.se (C.S.L.), ejetee@gmail.com (A. J. T.)
- 3 Schoolof Chemical Technology, Kalinga Institute of Industrial Technology (KIIT), Bhubaneswar 751024,
 India
- * Correspondence: S. K. Tripathy (E-mail: tripathy.suraj@gmail.com, suraj.tripathy@kiitbiotech.ac.in, Tel: +91-9438321810)
- 17 [‡]S.D. and S. G. have equal contribution
- 18 § S. K. T., C. S. L. and A. J. T. have equal contribution

Abstract: Antibiotic residues in aquatic environment have the possibility to induce resistance in environmental bacteria, which ultimately might get transferred to pathogens making treatment of diseases difficultand poses a serious threat to public health. If antibiotic residues in the environment can be eliminated or reduced, it has the possibility to contribute antibiotic resistance. Towards this objective, water containing ciprofloxacin was treated with sunlight assisted photocatalysis using Fe doped ZnOnanoparticles for assessing the degradation potential of this system.Parameters like pH, temperature, catalytic dosage were assessed for the optimum performance of the system. To evaluate degradation of ciprofloxacin, both spectrophotometricas well as microbiological (loss of antibiotic activity)methods were employed. 100 mg/L Fe doped ZnO nanoparticle catalyst and sunlight intensity of 120,000-135,000 lux system gave optimum performance at pH 9 at 30 °C and 40 °C. At these conditions spectrophotometric analysis showed complete degradation of ciprofloxacin (10mg/L) by 210 min. Microbiological studies showed loss of antibacterial activity of the photocatalytically treated ciprofloxacin containingwater against Staphylococcus aureus (108CFU) in 60 min and for Escherichia coli (108CFU) in 75 min. Thedeveloped system, thus possess a potential for treatment of antibiotic contaminated waters for eliminating/reducing antibiotic residues from environment.

Keywords: antibiotic residues; aquatic environment; ciprofloxacin; Fe doped ZnO nanoparticles; photocatalysis; sunlight

1. Introduction

Antibiotic residues in the environment is a major public health challenge [1]. Fluoroquinolones (FQs) are a class of environmentally stable broad spectrum antibiotics, which inhibits the enzymes DNA topoisomerase II (Gyrase) and DNA topoisomerase IV in bacteria thus interfering with their DNA replication machinery [2], [3]. FQs are effective against both gram positive and gram negative

bacteria and are used both in humans and animals Ciprofloxacin is the most commonly used FQ,Studies report the occurrence of FQs including ciprofloxacin in waterbodies world-wide [4]. FQ reach water bodies through excretion after incomplete metabolism within the human/animal gut [5]. Also their presence upto 87 microgram/L and 31mg/L has been demonstrated in waste water discharge [6]. Conventional waste water treatment including biological oxidation and other chemical and physical process leads to only partial removal of these compounds [7]. As a consequence, the presence of broad spectrum antibiotics like FQs even at very minute concentrations, poses a threat to the surrounding ecosystem and human health through the development of antibiotic resistance amongst environmental bacteria [8], which can potentially lead to further spread of resistance to other bacterial populations including human and animal pathogens through processes such as ingestion of untreated or partially purified water or horizontal gene transfer [9].

With the immediate necessity for substantive degradation of such organic environmental pollutants, semiconductor photocatalysis more appropriately, AdvancedOxidation Process (AOP) has proven quite useful [10]. It normally uses a semiconductor metal oxide or one of its doped variants as a photo-oxidant which in presence of light charges up and leads to the generation of highly reactive oxidative species like Hydroxyl radicals (OH·), Superoxide anion (O2·-) and Hydrogen peroxide (H₂O₂) for remediation of organic pollutants. The basic principle behind their action has been shown in the Schematic diagram Figure 1. Till date TiO2and ZnO has been reported to be the best catalyst for photocatalytic application because of optical properties, thus having a much better quantum efficiency in visible light [11]. Moreover owing to its high chemical stability, high oxidation efficiency, low toxicity, less expensive, easy availability and being abundant in nature they are an assured photocatalyst to be employed for mineralization of organic pollutants in both acidic and basic medium [12]. ZnO absorbs a substantial amount in the UV range [12] and UV accounts foronly 3-5% of the sunlight, thus there is insufficient usage of the total sunlight available. So efforts are needed to design catalysts which will show better photocatalytic efficiency in visible region of sunlight [11]. In order to reducesuch problems, modifying the metal oxide semiconductor with transition, alkaline and rare earth metals like Mn, Fe, Co, Ni, Ag, Mg, Pb, N, C, S, P, is done[11]will shift light absorption towards the visible range.

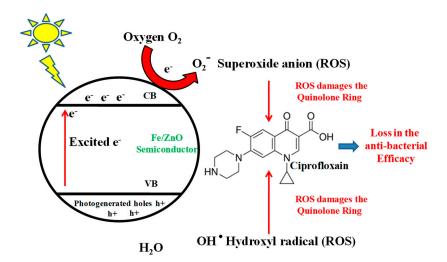


Fig 1. Schematic representation showing generation of reactive oxygen species (ROS) by Fe ZnO nanoparticles on activation with sunlight, and how these ROS attack active components of FQ to degrade them and reduce their anti-bacterial activity

Photocatalysis with ZnO for the degradation of antibiotics like ciprofloxacin, amoxicillin, ampicillin, cloxacillin using different sources of light was earlier performed [13, 14]. Nearly 50%

- degradation of antibiotics was achieved with high rate constant and maximium degradation was reported at pH 10-11. It has been previously reported in one of our studies that, using Fe Doped ZnO
- 82 for photocatalytic applications majorly contributes towards the generation of H₂O₂ in the system,
- 83 which ultimately serves detrimental for the photocatalytic oxidation. Moreover the presence of Fe in
- the system, serves as an added advantage for the photocatalytic oxidation, since it comes in contact
- with H2O2 in the system to generate more of hydroxyl radicals via Fenton process [15] This will
- 86 ultimately exaggerate the oxidation of antibiotic containing water. Thus doping the catalyst with
- 87 iron has some added benefits as far as increasing the photocatalytic efficiency of ZnO are concerned.
- 88 Earlier such Fe Doped ZnO have been used for successful degradation of wastewater bearing dye
- molecules [16]. The aim of this study was to evaluate sun light assisted photocatalytic degradation of
- 90 ciprofloxacin using Fe doped ZnO nanoparticles. Further, residual antibacterial activity of the
- 91 treated water was assessed against a gram positive (Staphylococcus aureus) and gram negative
- 92 (Escherichia coli) bacteria.

93 2. Materials and Methods

- 94 2.1. Materials
- 95 Chemicals used in this study includes Ciprofloxacin Hydrochloride (MP Biomedicals), Zinc Nitrate
- 96 Hexahydrate (Sigma Aldrich, 98%), Trisodium Citrate dihydrate (Sigma Aldrich), Ferric chloride
- 97 (Himedia, India), Luria agar and Luria broth (Himedia). All the chemicals were of molecular grade.
- 98 2.2. Preparation of ciprofloxacin stock solution
- 99 Ciprofloxacin hydrochloride (MP Biomedicals) stock solution (100 mg L-1) was prepared in
- 100 deionized water (NaOH was used to solubilization of ciprofloxacin followed by 5 min of
- ultrasonication), 2 litre at a time and stored in dark at 4°C. Working solutions of 10 mg L-1, (in 300 ml
- deionized water at a time) were prepared for each photocatalysis experiment, as required.
- 103 2.3. Synthesis of nanocrystalline Fe-DopedZnO
- 104 Fe-Doped ZnO was prepared using precipitation route as previously described [15]. Briefly, Zinc
- Nitrate hexahydrate (5.948g), Ferric chloride (0.108g) and Trisodium citrate (5.882) were dissolved in
- 106 500mL distilled water and stirred at 80°C for 60 min. 250mL of NaOH (250mM) was slowly added
- drop by drop into it using a burette until yellowish-white precipitate was formed. The precipitate
- was allowed to come to room temperature and was then centrifuged (10000rpm, 10min), and rinsed
- with distilled water thrice. The precipitate was then dried at 70°C overnight followed by calcination
- at 500°C. The calcined Fe-Doped ZnO powder was characterized as mentioned in our previous
- paper and used for photocatalytic applications.
- 112 2.4.Photocatalytic degradation of ciprofloxacin
- 113 A 300 mL aqueous solution of ciprofloxacin with concentration 10mg L-1was placed in a 500 mL
- borosilicate beaker (with required amount (see below) of Fe-Doped ZnO and was mixed by a
- 115 magnetic stirrer. The set was kept undisturbed in dark for 30 min to allow equilibrium. The
- experiments were performed with different catalyst concentrations 100, 150 and 200mg L-1, at pH
- 2,3,5.5,7,9,10 and 11 (required pH was adjusted with 1NHCl or 1N NaOH), different reaction
- temperature 30° C, 40° C, 50°C and 60°C and different photocatalysts TiO₂ and ZnO at a light
- temperature 50°C, 40°C, 50°C and of C and university photocatalysis 1102 and 2110° at a light
- intensity of 80000 ± 3000 lux, which corresponds to 650W/m². At 15min intervals, upto 210 min,
- $120 \hspace{0.5cm} \text{collected samples were filtered through } 0.45 \hspace{0.5cm} \text{micron membrane filter for spectrophotometric} \\$
- analysis (λ_{max} -280 and 320 nm using Shimadzu UV-1800) and the microbiology experiments for
- assessment of residual antibacterial activity. The time dependent decrease in absorbance values at
- λ_{max} 280 and 320 nm suggests degradation of the antibiotic [14].

2.5. Residual antibacterial activity of the treated water

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Qualitative assay was performed using well diffusion to assess the residual antibacterial activity of the treated water after photocatalytic degradation against the fully susceptible test organisms Staphylococcus aureus (MTCC code 3160) and Escherichia coli (MTCC code 7410) fromMicrobial type culture collection, MTCC Chandigarh, India. Well diffusion method, according to the Clinical and Laboratory StandardsInstitute (CLSI) [17, 18], was employed for the determination of the residual antibacterial activity of the samples. All plates were prepared in 90 mm sterile Petri dishes (Tarsons, India) with 22 mL of Luria Bertani agar, yielding a depth of 4 mm. Test microorganism's 100µL of inoculum suspensions (OD600-0.5, corresponding to 1.0 × 108CFU mL⁻¹) were poured into the agar plates when the temperature reached around 40-45°C using sterile micropipette, and homogenized thoroughly by mixing in a circular motion (pour-plate technique). After solidification, roundwells (6.0 mmin diameter) were punched into the seeded agar plates with a 6mm cork borer. Thewells were filled with 40µL of the treated water samples (collected and filtered after regular time intervals) using a sterile micropipette. These plateswere allowed to stand at 4°C for 2 h and then incubated at 37oC for 24 h. Three sets of simultaneous controls were used. One control was the organism control and consisted of a seeded Petri dish with no photocatalytic treated antibiotic sample. In the second control, samples were introduced in the holes of unseeded Petri dishes to check for sterility. Finally, to ensure the elimination of any solvent effect, wells filled with 40µL of sterile double distilled water were run simultaneously as a third control. The diameters of the inhibition zones (Zone of inhibition/ZOI) were measured in millimeters [19]. Each test was repeated six times and the mean values from the replicates along with standard error of mean (SEM) were calculated.

3. Results and Discussion

3.1 Photocataltytic degradation of ciprofloxacin and process optimization

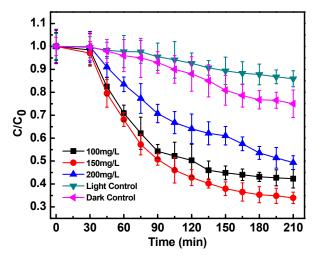


Fig 2. Photocatalytic degradation of antibiotic ciprofloxacin (10 mg/L) in water, in the presence of Fe-ZnO nanoparticles (At different concentrations of 100,150 & 200 mg/L) irradiated with sunlight light intensity of 80000± 3000 lux compared to photolysis (Light control) and degradation in absence of light (Dark control). Co represents initial concentration of ciprofloxacin and C represents concentration of ciprofloxacin at a particular time point. C/Co denotes, time dependent change in ciprofloxacin concentration with respect to initial concentration.

Fig 2 shows the decrease in C/Co(C-Concentration at a particular time, Co - Initial concentration of ciprofloxacin) absorption spectrum of ciprofloxacinat three catalyst concentrations 100,150,200 mg L-1, during sun assisted photocatalysis by Fe Doped ZnO nanoparticles. The values were calculated on the basis of intensity of absorbance peaks at 280 and 320 nm. In both these lambda values, the absorbance showed a decreasing trend at all the three catalyst concentrations 100, 150, and 200 mg L-1. Catalyst concentration of 150mgL⁻¹ caused significant degradation of ciprofloxacin (10mg L⁻¹)up-to 66% in 210 min and was found to be optimum. The other two concentrations were not as effective. The 100mg L⁻¹ catalyst may not have the capability for substantial generation of reactive oxygen species, while the 200 mg L⁻¹ catalyst concentration may be high enough to create catalyst shielding effect. Moreover 200 mg L⁻¹ may possess slow or improper degradation kinetics only 51%. For further experiments, therefore all the degradation experiments were carried out with 150mg/L of Fe-Doped ZnO. There was no significant change in concentration of the ciprofloxacin due to the direct sunlight assisted photolysis (Light Control) which was found to be only 14% [14]. The decrease in C/Co value (upto 25%) of the antibiotic when subjected to dark control reaction (at optimum photocatalyst concentration of 150mg/L), may be attributed to direct adsorption of the antibiotic in presence of doped ZnO nanoparticles [11].

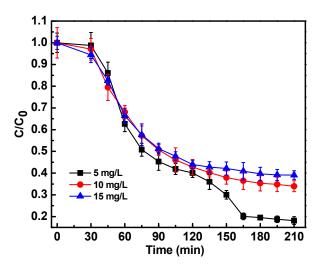
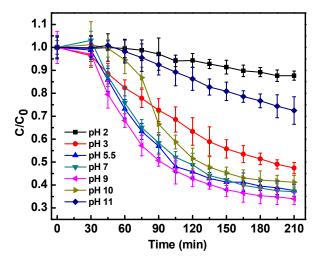


Fig 3. Photocatalytic degradation of antibiotic ciprofloxacin in water at different antibiotic concentration between 5, 10,15 mg/L with optimum Fe doped ZnO nanoparticles concentation of 150 mg/L and irradiated with sunlight intensity of 80000± 3000 lux. Co represents initial concentration of ciprofloxacin and C represents concentration of ciprofloxacin at a particular time point. C/Co denotes, time dependent change in ciprofloxacin concentration with respect to initial concentration.

The concentration of antibiotic in the waste water system is a key parameter to optimize the photocatalytic degradation process. A study was performed with ciprofloxacin concentrations of 5, 10 and 15 mg L⁻¹. Fig. 3 shows the photocatalytic degradation pattern of different concentration of ciprofloxacin with optimized concentrations of Fe doped ZnO nanoparticles. At 10 mg L⁻¹ concentration no peaks at 280 and 320 nm were observed after 210 min of photocatalytic treatment. Suggesting complete degradation of the quinolone ring. Also 5 mg L⁻¹ concentration of ciprofloxacin were completely degraded, since studies with 10 mg L⁻¹ concentrations were previously done and reported, the rest of the photocatalytic study were done with 10 mg L⁻¹ concentration. With 15 mg L⁻¹ ciprofloxacin concentration the degradation kinetics were a bit slower. The possible reasons could be catalyst shielding effect and over occupied catalyst active sites in 15 mg/L concentration [11, 19].

pH modifies the surface charge properties of Fe-Doped ZnO and possibly the chemical structure of the antibiotic, therefore influence of pH on photocatalytic activity of Fe doped ZnO nanoparticles was studied by altering the pH of the reaction mixture in both acidic and basic range. Fig 4 shows the effect of photocatalytic degradation on ciprofloxacin at different pH in presence of Fe-Doped ZnO nanoparticles. The best degradation efficiency of ciprofloxacin with Fe doped ZnO nanoparticles, nearly 65% was seen at pH 9, while the lowest degradation, only 10%, was observed at pH 2 [14]. The maximum ciprofloxacin degradation was obtained at basic pH values between 9 and11 under solar light, where the available hydroxyl ions in the system can react with the valence band holes (h+) to form reactive hydroxyl radicals (OH·), which possesses high oxidation capabilities under photocatalytic conditions, subsequently enhancing the rate of photocatalytic degradation of ciprofloxacin. Similar results for degradation of aromatic compounds were reported earlier [20]. At acidic pH value 2, the solar photocatalytic degradation of ciprofloxacin was hindered due to the high proton concentration, which possesses higher attraction for the hydroxyl anions, quenching the formation of hydroxyl radicals. As free hydroxyl ions in the system are decreased, the formation of hydroxyl radicals becomes limiting. Thus photocatalytic degradation of ciprofloxacin decreased at lower pH. It may also be possibly due to dissolution of Fe doped ZnO at acidic conditions. Similar observations were previously made in the photocatalytic degradation of azo dyes [16].



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Fig 4. Photocatalytic degradation of antibiotic ciprofloxacin (10 mg/L) in water in the presence of Fe-ZnO nanoparticles (150 mg/L) irradiated with sunlight intensity of 80000± 3000 lux at different reaction pH of 2,3,5.5,7,9,10,11. Co represents initial concentration of ciprofloxacin and C represents concentration of ciprofloxacin at a particular time point. C/Co denotes, time dependent change in ciprofloxacin concentration with respect to initial concentration.

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Ciprofloxacin is an ampholytic compound with pKa value of 6.09 for the carboxylic group and 8.74 for the nitrogen on the piperazinyl ring. The isoelectric or zwitterionic point is at pH 7.4. Thus ciprofloxacin seemed to be most sensitive to photocatalytic degradation in a pH closer to its zwitterionic form i.e. at basic pH 9. It has earlier been reported that the maximum stability of the molecule was observed in reaction solution of pH 4.0 [21], where the carboxylic group is un-ionized and basic nitrogen is completely under protonated condition. This adds an advantage to the ciprofloxacin pharmaceutically, because most of the pharmaceutical formulation possess pH between 3.5 and 5.5. This seems good for the pharmaceutical perspective but photocatalytic degradation at such low pH will be a challenge. Interestingly it has been previously reported that,

hospital waste water outfalling to drains have an pH in between 6.7 to 7.7 throughout the year, Moreover the pH of surface water mainly from lakes and rivers in India is between 6.5 to 8.5.[22] The current study thus finds it application for degradation of antibiotics in hospital wastewater and surface water, since at this pH range the photocatalytic degradation was more than 60% as shown in Figure 3.

From experimental observations and previous reports on photocatalytic degradation of organic molecules like dyes [23] and antibiotics [24], we assumed that on irradiation with solar light, within the Fe Doped ZnO nanoparticles, excitation of electrons takes place from valence band into the conduction band. Photo-generated holes in the conduction band on reacting with water molecules in the system generate hydroxyl radicals which possess oxidative nature to get rid of antibiotics adsorbed at Fe-Doped ZnO surface. Moreover the high oxidative prospective of valence band holes can also lead to direct and indirect oxidation of antibiotic. The presence of Fe in the system possesses added advantage to this photocatalytic degradation process. Presence of Fe delays the electron whole recombination, acting as one of the terminal acceptors of electrons, which eventually increases the generation of hydroxyl radicals and reactive species in the system. Also Fe as a Fenton agent is capable of producing reactive oxygen species like OH• radicals through Fenton process, adding more ROS to the system for subsequent degradation of ciprofloxacin[11, 25].

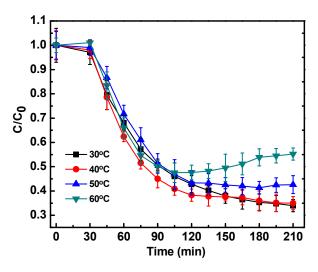


Fig 5. Photocatalytic degradation of antibiotic ciprofloxacin (10 mg/L) in water in the presence of Fe-ZnO nanoparticles (150 mg/L) irradiated with sunlight intensity of 80000± 3000 lux and pH 9 with different reaction temperature. Co represents initial concentration of ciprofloxacin and C represents concentration of ciprofloxacin at a particular time point. C/Co denotes, time dependent change in ciprofloxacin concentration with respect to initial concentration.

Temperature was found to modulate the degradation kinetics (Fig 5). Generally it has been reported that, with increase in temperature the degradation kinetics enhances[11], but in the current study, the opposite trend was observed. With increasing temperature, the degradation kinetics decreased upto 60°C. The possible reason could be the increase in stability of fluoroquinolones on exposure to heat stress. It has been reported by Roca et. al. [26], that FQ can be stable at temperature up to 120°C. In a country like India, where the atmospheric temperature can reach upto 50°C, the technique presented in this paper can be employed for successful degradation of ciprofloxacin and may be other fluoroquinones also, in waste water matrices. The technique presented in this paper may also

find its application for treatment of hospital, pharmaceutical or industrial waste water for degradation of many organic molecules.

3.2 Analysis of residual antibacterial activity of antibiotic after photocatalytic degradation.

Ciprofloxacin, as already discussed, is an antibiotic that belongs to the FQ class of antibiotics. The antibiotics that belong to this group, generally inhibit the growth of several microorganisms via the inhibition of DNA Gyrase, which is a factor is responsible for the division of bacterial cells. ciprofloxacin is active against a wide spectrum of Gram Positive and Gram Negative bacteria and ciprofloxacin and antibiotics of FQ group are widely present in wastewaters such as from hospital, municipal, pharmaceutical industry sources etc [1, 22, 27]. The residues of these antibiotics in the wastewaters generate antibiotic resistant bacteria in the environment, which is a potential major threat to public health.

Table-1 Shows residual antibiotic activity of the antibiotic ciprofloxacin after photocatalytic degradation with Fe Doped ZnOnanaoparticles against *Staphylococcus aureus and Escherichia coli*.

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Test Bacteria	Staphylococcus aureus (10°CFU)			Escherichia coli(10°CFU)		
Time (min)	PCD Zone of inhibition (ZOI) in mm Mean ± SEM	DC Zone of inhibition (ZOI) in mm Mean \pm SEM	PL Zone of inhibition (ZOI) in mm Mean \pm SEM	PCD Zone of inhibition (ZOI) in mm Mean ± SEM	DC Zone of inhibition (ZOI) in mm Mean ± SEM	PL Zone of inhibition (ZOI) in mm Mean ± SEM
0	12 ± 0.3	12.5 ± 0.3	12.5 ± 0.2	15 ± 0.3	15 ± 0.2	14.5 ± 0.3
30	12.5 ± 0.3	12 ± 0.3	12.5 ± 0.2	14.5 ± 0.3	14.5 ± 0.2	14.5 ± 0.2
45	10 ± 0.3	8 ± 0.5	11 ± 0.2	11 ± 0.3	14 ± 0.2	14 ± 0.2
60	7.5 ± 0.2	12 ± 0.5	10 ± 0.2	12 ± 0.4	14 ± 0.2	13.5 ± 0.3
75	5.5 ± 0.2	11 ± 0.3	10.5 ± 0.2	9.5 ± 0.3	14.5 ± 0.2	12 ± 0.5
90	0	11.5 ± 0.5	9 ± 0.3	6 ± 0.2	14.5 ± 0.2	11.5 ± 0.3
105	0	9 ± 0.3	8.5 ± 0.2	0	12 ± 0.3	12 ± 0.3
120	0	10 ± 0.2	7.5 ± 0.3	0	12.5 ± 0.4	10 ± 0.3
135	0	10 ± 0.3	8 ± 0.2	0	14 ± 0.3	9 ± 0.3
150	0	10.5 ± 0.2	7 ± 0.2	0	14 ± 0.2	8.5 ± 0.2

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165	0	9 ± 0.3	6 ± 0.2	0	14.5 ± 0.3	7 ± 0.2
180	0	11 ± 0.3	5.5 ± 0.2	0	14 ± 0.2	0
195	0	11 ± 0.2	0	0	14 ± 0.3	0
210	0	12 ± 0.3	0	0	13 ± 0.5	0

- 272 *SEM stands for Standard Error of Mean, calculated from the standard deviation, PCD-
- 273 Photocatalytic degradation, DC- Dark control, PL- Photolysis, n(number of replicates) = 6
- * Zone of inhibition(ZOI) = Total zone (including the disc) Diameter of the disc (6 mm)
- Note The well diffusion assay have been performed in accordance with the Clinical & Laboratory
- 276 <u>Standards Institute: CLSI Guidelines.</u>

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Note- No ZOI have been observed from the solvent controls i.e with the Distilled water, No contaminating bacteria were found to grow around the treated samples when poured without the test bacteria.

The current work aims to employ photocatalysis for successful degradation of antibiotic ciprofloxacin. After subjecting ciprofloxacin to photocatalytic treatment with Fe doped ZnO nanoparticles, a confirmatory bacterial inhibition experiment was conducted to check whether the antibiotic was completely degraded in the experimental system using test organisms Staphylococcus aureus and Escherichia coli [19]. The results of the experiment (Table 1) showed that for both Staphylococcus aureus and Escherichia coli, ciprofloxacin lost its antibacterial activity after 60 minutes and 75 minutes post irradiation respectively. With an increasing time, a decreasing zone of inhibition in both Staphylococcus aureus and Escherichia coli was evident. The Zone of Inhibition decreased from 12 mm to 5.5 mm and from 15 mm to 6 mm in the case of Staphylococcus aureus and Escherichia coli in 60 min and 75 min post irradiation respectively. It can be seen that Escherichia coli, a gram negative organism shows susceptibility to ciprofloxacin that has been collected 75 minutes post irradiation, which is slightly less than that of Staphylococcus aureus (sample collected 60 minutes post irradiation), before completely showing zero susceptibility in both cases. Escherichia coli is a gram negative microorganism, with a weak cell wall that is made up of Lipopolysaccharide [28, 29]. Therefore it is easy for a disinfecting agent to penetrate its cellular defenses; compared to Staphylococcus aureus, which is Gram positive. In case of light control and dark control, antibacterial activity was not lost even after 120 min for both Escherichia coli and Staphylococcus aureus. There was little decrease in the zone of inhibition parameters and it clearly signified that ciprofloxacin was still present in the case of experimental controls, suggesting that both the photocatalyst (Fe doped ZnO) and sunlight are indispensable in the degradation process.

4. Conclusions and implications

An Fe doped ZnO nanoparticles based sunlight assisted photocatalytic system was developed for degradation of the fluoroquinolone antibiotic- ciprofloxacin in water, also assessing its best performance parameters like pH, temperature, catalytic dosage. The degradation of ciprofloxacin was proved both spectrophotometrically as well as microbiologically by loss of antibiotic activity of the photocatalytically treated water. The developed Fe doped ZnO nanoparticles based photocatalytic system can potentially beusedfor degradation of other fluoroquinolones and other antibiotics as well as other organic contaminants in water. Antibiotic residues in aquatic systems has the potential to induce resistance in bacteria, which has further the potential to infect humans and

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310 thereby become a serious threst to human health. The developed system has therefore potential to 311 contribute to contain antibiotic resistance. 312 313 Acknowledgments: This work was supported by Department of Science & Technology, Government of India (DST/TM/WTI/2K15/75C) and Swedish Research Council (Grant Number (Grant No. 2012-02889 and 314 315 2017-01327). 316 Author Contributions: S.K.T., A.J.T. and C.S.L. created the original study plan. S.D., S.G. and A.J.M designed 317 and executed the experiments under the guidance of S.K.T.A.M. helped in the microbiology experiments. S.D. 318 S.G and A.J.M wrote the manuscript. S.K.T., A.J.T, and C.S.L. reviewed and edited the manuscript. All authors 319 read and approved the manuscript. 320 **Conflicts of Interest:** The authors declare no conflict of interest. 321

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