

**MOLECULAR CHARACTERIZATION OF PHOSPHOLAMBAN AND  
RENINANGIOTENSIN SYSTEM GENES MUTATIONS AND CLINICAL  
EPIDEMIOLOGY IN HUMAN CARDIOMYOPATHY**

1

2 Gnana Veera Subhashini<sup>1\*</sup>, Beesetti Swarna latha<sup>1</sup>, Mavuluri Jayadev<sup>1</sup>, Emmanuel C<sup>2</sup>,  
3 K M Cherian<sup>2</sup>

4 1. Baba Clinical and Genomic Research Centre, Chennai 600113, Tamilnadu, India.

5 2. International Center for Cardio Thoracic and Vascular Diseases, Frontier Lifeline,  
6 Chennai, India.

7

8 \* Correspondence: Dr. N. Gnana Veera Subhashini

9 Email Id: dr.ngvsubhashini@bcgrc.com

10

11 **ABSTRACT**

12 **Background**

13 Cardiomyopathy is commonly observed disease that may occurs due to mutations in  
14 either susceptible genes or modifier gene. People with broad age group are affected  
15 either attributable to spontaneous or inherited mutations of these genes. Various gene  
16 mutations are reported so far but only few of them were studied in detail.

17 **Methods**

18 In the current study, we evaluated epidemiological variables like age, sex, familial status,  
19 parental consanguinity. We also described specific clinical symptoms associated with the  
20 cardiomyopathy condition in Indian population.

## 21 **Results**

22 Our studies on mutation screening of phospholamban gene revealed two transitions  
23 (4880 C/T, 4887 T/G) in 5' flanking region which might cause inherited dilated  
24 cardiomyopathy with refractory congestive heart failure are We further deliberated the  
25 gene polymorphism of renin angiotensin system gene angiotensin-1-converting enzyme  
26 as an associated marker/ modifier in cardiomyopathy patients and their family members.

## 27 **Conclusions**

28 Information on epidemiological, clinical statistics, phospholamban gene mutation analysis  
29 and angiotensin-1-converting enzyme gene polymorphism is essential to guide the  
30 successful execution for future therapies and benefits us to identify those patients at risk  
31 for faster disease progression, congestive heart failure, and arrhythmia.

32

33 **Keywords:** Cardiomyopathy, Hemodynamic and Biochemical Parameters,  
34 Epidemiological and clinical Parameters, Phospholamban Angiotensin-1-converting  
35 enzyme.

36

## 37 **INTRODUCTION**

38 In the present scenario, the debilitating nature of cardiovascular disorder is alarming and  
39 needs a constant watch on the premature morbidity and mortality status. Cardiomyopathy  
40 is the heart muscle disease and most common genetic disease of the heart, characterized

41 by heterogeneous morphologic expression and clinical condition (1). It can manifest  
42 negligible to extreme hypertrophy, minimal to extensive fibrosis, myocyte disarray, absent  
43 to severe left ventricular outflow tract obstruction, distinct septal morphologies, or  
44 hypocontractile (2). Cardiomyopathy can cause heart failure (HF), which in most cases  
45 leads to sudden cardiac death (SCD). Hypertrophic Cardiomyopathy (HCM), Dilated  
46 Cardiomyopathy (DCM), Restrictive Cardiomyopathy (RCM) and Arrhythmogenic Right  
47 Ventricular Cardiomyopathy (ARVC) are four types of cardiomyopathies reported (3).  
48 These types can be primary myocardial disorders or at times develop as a secondary  
49 consequence of a variety of conditions viz., myocardial ischemia, inflammation, viral  
50 infection, increased myocardial pressure or volume load, and toxic agents (4). The  
51 etiology of both HCM and DCM involve cardiac energy imbalances and the clinical  
52 expressions of them are based on the additive factors that are involved in it.

53

54 The prevalence of the dilated cardiomyopathy has formerly been estimated  
55 at 36.5/100,000, with an incidence of 4-8/100,000 persons-year (5). An  
56 echocardiographic analysis of 4111 subjects in CARDIA study by (6) revealed  
57 hypertrophic cardiomyopathy affects 1 in 500 people, a prevalence similar to familial  
58 hypercholesterolemia. Recent study estimates prevalence of dilated and hypertrophic  
59 Cardiomyopathy as 36 cases per 100,000 people and 10–20 cases per 100,000 people  
60 respectively (7). Most of the dilated Cardiomyopathy cases are sporadic; although 20–  
61 35% of them are familial (8). The incidence of dilated cardiomyopathy varies in men and  
62 women. However, in general, heart failure is more common in men (9). The treatment  
63 arm of the Studies on Left Ventricular Dysfunction (SOLVD), in which only 15% of the

64 patients were women, reported no sex-related difference in survival in either the placebo  
65 group or the Enalapril group (10, 11). Age distribution depends on prognosis, diagnosis  
66 or onset of any underlying disease. However, advancing age is reported as an  
67 independent risk factor for mortality in several studies (12).

68 Phospholamban (PLN) is a small transmembrane phosphoprotein of 52 amino acids that  
69 plays an important role in cardiac contraction and relaxation. Phospholamban, expressed  
70 in the sarcoplasmic reticulum membrane controls cellular calcium levels by a mechanism  
71 that depends on its phosphorylation (13). The human ventricle and quadriceps displayed  
72 high levels of phospholamban transcripts and proteins (14), whereas lower expression in  
73 smooth muscles and right atrium. DCM patients with a phospholamban gene mutation  
74 have a chronically inhibited  $\text{Ca}^{2+}$ -ATPases pump, which leads to DCM in their teenage  
75 (15, 16).

76 Susceptibility genes have a role in the development of cardiomyopathies, whereas  
77 modifier genes have a role in the evolution or prognosis of the disease. In most studies  
78 due to limited sample sizes, the role of susceptibility and modifier genes have been only  
79 suggestive (17). Various genes underlying cardiomyopathy have been identified from  
80 linkage as well as candidate gene studies, and include those coding for proteins involved  
81 in the cytoskeleton, the Z-disk, the nuclear envelop, ion conduction and calcium handling  
82 proteins (18). This variability in expression of these causal genes is also seen among  
83 family members sharing the same mutation. This variable expressivity, which confuses  
84 genotype/phenotype correlations, could be partially explained by both environmental  
85 influences and genetic modifiers (19). The angiotensin-1-converting enzyme (ACE)  
86 polymorphism has been commonly studied with variable results. It's noteworthy to study

87 and understand the importance of modifier genes. The genetic polymorphisms of the  
88 renin-angiotensin-aldosterone system (RAAS) are found to influence the progression to  
89 cardiac disorders (20). Angiotensin-1-converting enzyme (ACE), a modifier gene was  
90 perceived to have insertion/Insertion (I/I) genotype associated with low serum ACE  
91 activity levels, Insertion/Deletion (I/D) genotype with intermediate levels and  
92 Deletion/Deletion (D/D) high serum ACE activity levels. The effect of ACE polymorphism  
93 on survival in patients with DCM with a DD genotype had poorer prognosis than other  
94 genotypes (5-year survival rate 49 vs.72%,  $p=0.001$ ) (21). The DD genotype was also  
95 associated with an increase in left ventricular mass (22).

96

97 The present study investigates the association of spectrum of clinical symptoms, the  
98 epidemiological variables like age, sex, familial status, parental consanguinity and the  
99 mutations in the gene encoding phospholamban cardiac protein and to establish the  
100 genotype – phenotype correlations, to identify the modes of inheritance and the risk  
101 stratification in a group of clinically well characterized patients and their relatives  
102 associated with the cardiomyopathy condition in Indian population. We further studied the  
103 role of angiotensin-1-converting enzyme gene polymorphism as an associated marker/  
104 modifier in cardiomyopathy patients and their family members.

105

## 106 **MATERIALS AND METHODS**

107

### 108 **STUDY SUBJECTS**

109 A total of 109 unrelated index cardiomyopathy patients and their family members who  
110 were consecutively enrolled in cardiology units of the Government General Hospital,  
111 Chennai and International Center for Cardiothoracic and Vascular Diseases (ICCTVD),  
112 Dr. K.M. Cherian Heart Foundation, A unit of Frontier lifeline, Chennai, South India, were  
113 included in the present study. A panel of 100 age matched healthy control subjects with  
114 no history of heart disease was obtained. The study protocol was approved by the  
115 Institutional Review Board and a Written consent was obtained from all subjects in  
116 accordance with the Institutional Ethical committee human subject guidelines.

117

## 118 **CLINICAL EVALUATION**

119 The index cases were subjected to standard physical examinations, clinical  
120 evaluations, electrocardiographic and echocardiograph tests to confirm cardiomyopathy.  
121 Cardiomyopathy was diagnosed based on the presence of the following; dilated  
122 cardiomyopathy with Left ventricular ejection fraction (LVEF) <45% and left ventricular  
123 end diastolic diameter >27mm/m<sup>2</sup>. However, patients who had secondary cause were  
124 not included in the present study. Further, hypertrophic cardiomyopathy was confirmed  
125 by LV wall thickness  $\geq 15$ mm and blood pressure  $\leq 160/100$ mmHg. Demographic and  
126 clinical information (Echocardiographic and Electrocardiographic findings) have been  
127 taken from the case sheets of respective hospitals. Family history and parental  
128 consanguinity were collected from the patients. Patients were classified according to New  
129 York Heart Association (NYHA) and exercise capacity has also been taken.

130

## 131 **BLOOD SAMPLE COLLECTION**

132 Five milliliters of peripheral blood were drawn in EDTA coated vacutainer  
133 from patients, family members and control subjects and stored at 4°C until DNA  
134 extraction.

135

#### 136 **DATA COLLECTION**

137 Epidemiology parameters such as height, weight, sex, age at onset, dietary  
138 habits, addictions to smoking and alcohol were collected during personal and clinical  
139 history. Each of the subjects met the Clinical diagnostic criteria *viz.*, 12 lead  
140 electrocardiograms, echocardiogram, clinical symptoms, risk factors, medication and  
141 outcome of the disease etc.

142

#### 143 **CLINICAL CHEMISTRY PARAMETERS:**

144

145 Blood parameters like sugar, urea, creatinine, creatinine phosphokinase (CpK), atrial  
146 natriuretic peptide (ANP), brain natriuretic peptide (BNP), serum aspartate transaminase  
147 (SGOT) and Serum alanine transaminase (SGPT) were analyzed using DaytonaRM  
148 Randox auto analyzer according to the manufacturers' instructions.

149

#### 150 **GENETIC ANALYSIS**

151 In the present study, the index cases are categorized into two groups *viz.*, familial case  
152 showing the incidence of the disease in first and second-degree relatives and in sporadic  
153 cases lack of any familial incidence, presumably non- genetic in origin. The present study

154 examined a comprehensive screening of Phospholamban and a key determinant/modifier  
155 angiotensin-1-converting enzyme gene polymorphism.

156

#### 157 **DNA ISOLATION and QUANTIFICATION**

158 Genomic DNA was extracted from the peripheral blood following the  
159 protocol of (23). Briefly, the collected blood was mixed with an equal volume of TKM1  
160 buffer (10mM Tris-HCl, 10mM KCl, 10mM MgCl<sub>2</sub> and 2mM EDTA) and 100 µl Triton X.  
161 The contents were centrifuged and the pellet was washed with TKM1 repeatedly until the  
162 cell debris is washed out. The pellet is suspended in 800 µl of TKM2 solution (10mM Tris-  
163 HCl, 10mM KCl, 10mM MgCl<sub>2</sub>, 2mM EDTA and 0.4M NaCl) followed by centrifugation and  
164 precipitation of the supernatant in ethanol. The DNA samples were then stored at -20°C  
165 for subsequent analyses. The DNA was quantified using spectrophotometer. The DNA  
166 was diluted in TE to yield 50ng/µl concentration.

167

#### 168 **PHOSPHOLAMBAN (PLN) POLYMERASE CHAIN REACTION**

169 The primers of the hotspots exons of Phospholamban (PLN) forward primer 5'-  
170 tatttttctcataattaaattcctgc-3' and reverse primer 5'-aaagtaagaattaccaaagtcagcg-3' for  
171 Exon 1 and forward primer 5'-aacaatagtgctgaggaagatgaa-3' and reverse primer 5'-  
172 ttgtttcctgtctgcatgg-3' for Exon 2 were used (24).

173

#### 174 **ANGIOTENSIN-1-CONVERTING ENZYME GENOTYPING**

175 The primers of the hotspots exons of the Angiotensin-1-Converting Enzyme (ACE)  
176 forward primer 5'-tatttttctcataattaaattcctgc-3' and reverse primer 5'-



177 aaagtaagaattaccaaagtcagcg-3' were used (25). Angitensin-1-converting enzyme  
178 genotyping was based on amplification of genomic DNA by PCR and the products were  
179 detected on 2% agarose gel. Amplification products 490 bp and 190 bp corresponding to  
180 the I and D alleles respectively (26).

181

## 182 **DNA SEQUENCING**

183 Genomic DNA from individuals with different SSCP patterns was amplified and  
184 sequenced with using Applied Biosystems 3730xl DNA Analyzer. The sequencing PCR  
185 was carried on 96 well micro-titer plates in a 5 µl reaction volume containing nuclease  
186 free water, the amplified template, "BigDye" (fluorescently labeled ddNTPs, dNTPs) and  
187 primers. The amplified DNA was precipitated by incubating at room temperature with 25µl  
188 of 3M-sodium acetate in ethanol (120 µl of 3M-sodium acetate in 3 ml of 100% ethanol)  
189 for 15min. The DNA was made single stranded by adding Hi-Di Form amide and  
190 sequenced on ABI3730xl automated DNA analyzer. The chromatograms obtained were  
191 analyzed on "Auto assembler" chromatogram analyzer on a Macintosh operating system.  
192 Complete sequencing work was carried out at the "Center for Cellular and Molecular  
193 Biology" Hyderabad.

194

## 195 **3.3. BIOINFORMATICS**

196 Multiple Sequence Analysis (MSA) was used for sequence comparison between  
197 generated sequences of the present study and to the corresponding reference sequences  
198 obtained from GenBank. The generated gene sequences were translated as protein  
199 sequences for future analysis (ExpASY Tool). Using Conserved Domains Database and

200 Motif Finder Database alteration in the conserved domains and motifs as a consequence  
201 of mutation were identified for the generated sequences.

202

### 203 **3.4. STATISTICAL ANALYSES**

204 Statistical analyses were carried out using Statistical Analysis Solutions (**SAS 9.2**). The  
205 mean and standard deviation were computed for various quantitative parameters and  
206 calculated. P-value <0.05 was considered significant. Association and relative risk  
207 estimates were carried out using Chi square test for the qualitative parameters at 1% and  
208 5% levels of significance. The Allele frequencies were computed by gene counting  
209 method. Departure from Hardy- Weinberg equilibrium was tested by chi-square test. The  
210 odds ratio was computed using SAS.

211

212

213

## 214 **RESULTS**

### 215 **EPIDEMIOLOGICAL STUDY**

#### 216 **Patients and Baseline Characteristics**

217 A total of 109 unrelated index patients who have echocardiographically and  
218 electrocardiographically assessed for cardiomyopathy and 100 aged matched control  
219 subjects were studied. About 73 percent of the patients had dilated cardiomyopathy and  
220 27 percent hypertrophic cardiomyopathy. Details on age, body mass index, blood  
221 pressure, blood biochemical profile, associated clinical and non-clinical features of the  
222 patients, control subjects were given in (Table 1). Males were significantly higher among

223 the patients (73%). The mean age of all patients was  $37.22 \pm 12.43$  years. Occurrence  
224 was familial in 24% of cases, but sporadic in the other 76% of patients. In patients group  
225 about 30% of them had family history of sudden cardiac death and 14% of them showed  
226 parental consanguinity. Mean value of body mass index and body surface area was same  
227 in patients and control ( $p=0.085$ ,  $p=0.515$ ).

228

### 229 **Hemodynamic and Biochemical Parameters**

230 Diastolic blood pressure, heart rate and systolic blood pressure of cardiomyopathy  
231 patients was extremely significant when compared with the control subjects ( $p<0.05$ ).  
232 Cardiomyopathy patients were characterized with high atrial natriuretic peptide (ANP) and  
233 brain natriuretic peptide (BNP) level ( $p<0.001$ ). Contrastingly creatinine phosphokinase  
234 (CpK), serum aspartate transaminase (SGOT), serum alanine transaminase (SGPT),  
235 glucose, urea and creatinine levels did not show much variation between patients and  
236 control group (Table 1). Similarly, co-morbid factors such as diabetes, obesity, smoking  
237 and alcoholism did not show any influence on patient population.

238

### 239 **CLINICAL CHARACTERISTICS**

240 The electrocardiographic and echocardiographic characteristics of the cardiomyopathic  
241 patients and control group were given in Table 2. All comparisons were significant  
242 between cardiomyopathic patients and control ( $p<0.001$ ) except fractional shortening, left  
243 ventricular ejection fraction, outflow tract gradient, early to late (E/A ratio) trans mitral flow  
244 velocity, and left ventricular systolic and diastolic volume.

245

246

247

## 248 **MOLECULAR ASPECTS**

### 249 **Phospholamban Gene Mutation**

250 Phospholamban gene is located on chromosome 6q22.1. Genomic sequence of the gene  
251 is 12146 bps nucleotides long which encodes 1742 bps mRNA coding region comprises  
252 159 bps which encodes 2exons. The protein is a pentamer and is a major substrate for  
253 cAMP-dependent protein kinase in the cardiac muscle. The protein inhibits Ca<sup>2+</sup>-ATPase  
254 in unphosphorylated state. The protein is a key regulator of cardiac diastolic function.

255 Table 3 shows the detailed description of the mutations in phospholamban gene. Seven  
256 mutations –two in the 5' flanking region, two in exon 1 and three in the intron1 region  
257 (Figure 1 (a-f)). Transition T/G in 4887 nucleotide regions of 5' flanking region was  
258 observed in two HCM probands with four affected family members and also in one dilated  
259 cardiomyopathic proband with no affected relatives. 15511-15512 insertion T was  
260 observed in one dilated cardiomyopathic proband with 2 affected family members and  
261 one dilated cardiomyopathic proband with no affected family members.

262

### 263 ***Description of Familial Condition***

#### 264 **4887(T/G) mutation:**

265 In the pedigree of the H9 proband (male, 21years) (Figure 2), the proband's father and  
266 his two uncles died young of sudden cardiac arrest. The younger brother of the proband's  
267 father has an affected son. The mutation was in heterozygotic state and it is not observed  
268 in unaffected.

269 In the pedigree of H10 proband (Figure 3), the T to G transition was observed in two  
270 family members The H10 proband (Male, age: 41years) had an affected offspring who  
271 was deceased subsequently to this study. The proband's parents are consanguineous  
272 and his mother died of cardiac arrest. The proband's brother had also carried this mutation  
273 and his daughter died unexpectedly at the age of 18 years.

#### 274 **15512 ins T mutation:**

275 The intronic mutation in PLN gene was observed in one dilated cardiomyopathy patient  
276 and in his family members (Figure 4). The D15 proband (Male, age: 37 years) who carried  
277 this mutation has an affected father (66 years). The proband's father's brother died of  
278 sudden cardiac arrest at the age of 22 years. The mutation in their family members were  
279 in heterozygotic state and not found in unaffected.

280

#### 281 **Angiotensin-1-Converting Enzyme Gene Polymorphism**

282 The present study reports on the molecular screening of a population of 109 unrelated  
283 index patients and 100 controls. Mutational screening was performed in one modifier  
284 gene Angiotensin-1-converting enzyme (ACE) i.e., zinc metallopeptidase widely  
285 distributed on the surface of endothelial and epithelial cells (27). When in demand, renin  
286 activity leads to the conversion of angiotensinogen to angiotensin I. ACE then converts  
287 angiotensin I to angiotensin II, which have been implicated in the modulation of cardiac  
288 growth (28). The human ACE gene is located on chromosome 17q23 and includes 26  
289 exons. Polymorphism involving the presence (insertion I/I) or absence (deletion D/D) of a  
290 287-basepair sequence of DNA in intron 16 of this modifier gene, encoding for ACE with  
291 differences in plasma ACE levels is examined in Cardiomyopathy patients (29). Table 4

292 illustrates variation in the genotype and allele frequency of the patient and control. Most  
293 notably the deletion homozygotes found to be higher in the patients. Also, the percent of  
294 heterozygote varied significantly among the two groups. Similarly, the deletion allele  
295 frequency was considerably higher in patients about 60% than controls (44%). The chi  
296 square value in patients and controls was 0.109 and 1.148 respectively.

297 The genotypes DD vs ID, DD vs II, ID vs II, DD + ID vs II, DD vs ID + II and D vs I was  
298 taken for the odds ratio comparison. All the odds ratio values were higher than the  
299 threshold value of 1. The odds ratio was highest for the genotype DD vs II (3.82) with a  
300  $\chi^2$  value of 10.59. This was followed by DD + ID vs II and ID vs II genotypes having an  
301 odds ratio of 2.79 ( $\chi^2 = 8.60$ ) and 2.27 ( $\chi^2 = 4.48$ ) respectively. The DD vs ID genotype  
302 comparison had an odds ratio of 0.60 ( $\chi^2 = 1.94$ ). It should be noted that the  $\chi^2$  value is  
303 taken to evaluate the significance of the genotypes. Patients with D allele tend to be a  
304 higher risk and modify the disease pattern when compared with I allele. The odds ratio  
305 was found out to be 1.14 with a  $\chi^2$  value 12.09. In Table 5 the chi square and the odds  
306 ratio values were calculated to prove the significance of the genotype of patients and  
307 reference to the control.

308

309

310

## 311 **DISCUSSION**

312 Cardiomyopathies are diseases of the heart muscle and a cause of concern. They exhibit  
313 a wide spectrum in disease onset, manifestation as well as in progression (30). Significant  
314 differences were observed for age and sex ratio between control and patients with

315 cardiomyopathy. Mean age of diagnosis was higher in the patient data of the present  
316 study than in studies of HCM and DCM previously reported (31,32). This may be due to  
317 mutation carriers screened in a predictive setting represent an asymptomatic subgroup  
318 within the total population of affected. Besides age, gender was an important cofactor in  
319 the clinical manifestation of HCM and DCM (33). The cardiomyopathy patients of the  
320 present study were characterized with significant increase in atrial natriuretic peptide and  
321 brain natriuretic peptide, which were correlated with left ventricular ejection fraction, mean  
322 pulmonary arterial pressure and pulmonary artery wedge pressure. Present data provides  
323 evidence that ANP and BNP are the best indicators for heart failure in cardiomyopathy  
324 patients. An elevated mean diastolic blood pressure ( $81.41 \pm 4.76$ ) and systolic blood  
325 pressure ( $125.0 \pm 9.17$ ) were observed in cardiomyopathy patient's data of the present  
326 study. A higher diastolic and systolic blood pressure has been observed in Caucasians  
327 (34), Chinese (35) and Japanese (36) origin.

328 Phospholamban is an inhibitor of endogenous sarcoplasmic reticulum calcium ATPase in  
329 dephosphorylated condition. It plays a regulatory role in the calcium handling during the  
330 process of cardiac contraction/relaxation. Mutations in this gene shown to associate with  
331 elevated cytosol calcium concentration. Phospholamban is phosphorylated by protein  
332 kinase A to increase the reuptake of calcium into sarcoplasmic reticulum (37,38,39).

333 Besides dilated cardiomyopathy though this is the first report to show Phospholamban  
334 gene mutation associated with hypertrophic cardiomyopathy, none of the all identified  
335 mutations falls neither within the coding region nor at conserved domain. Similar to this  
336 study there are few other studies had shown flanking regions and promoter variants of  
337 PLN associated with DCM/heart failure and HCM (40, 41). Alternatively, there are reports

338 that show no associations (42,43). It is possible that PLN gene mutations can express at  
339 a lower level leading to a smaller pathogenic effect during sixth decade of life in these  
340 individuals. Contrastingly two familial cases showed mutation of PLN (4887 T/G) possibly  
341 decrease the transcriptional activity of promoter and associated with hypertrophic  
342 cardiomyopathy.

343 The mutation that is located near the promoter region has been defined as the fragment  
344 with maximal transcriptional activity (44). The increase or the decrease in phospholamban  
345 activity due to the disruption around the promoter region can lead to cardiomyopathy (41).  
346 Some carriers do not exhibit clinical conditions and mutation may be due to other genetic  
347 and environmental factors. This study concludes that PLN gene mutations are not  
348 frequent cause of hypertrophic or dilated cardiomyopathy in Indian population.

349 Although similar data were lacking, it was observed that Indians show a greater variation  
350 and this may be due to the polymorphism in ACE gene (modifier gene). Further, role of a  
351 dual peptide system *viz.*, brain, and atrial natriuretic peptides in sodium balance and blood  
352 pressure regulation in those patients cannot be ruled out (45). Angiotensin-1-converting  
353 enzyme gene insertion/deletion (I/D) polymorphism is considered as an important  
354 modifier gene, which may influence the clinical phenotype of the cardiovascular disorders  
355 (46). The present data revealed that the frequencies of the DD genotype and D allele  
356 were significantly higher in patients compared with controls, and were associated with  
357 increased risk of HCM and DCM. Furthermore, regression analysis revealed that the  
358 genotypes DD and D allele were independent risk factors for these cardiomyopathies in  
359 Indian population. Patients with the DD genotype had the highest odds ratio of disease  
360 susceptibility and the subjects with II genotype have a lower risk of developing



361 cardiomyopathies that may possibly through a cardio protective effect. The D allele  
362 compared with I allele has more than 25 percent increased risk to cardiomyopathy. On  
363 comparison, the prevalence of D allele in the present study (Indian population) was  
364 slightly higher (about 60%) than Tunisian and Turkish populations (47,48). Association of  
365 DD genotype / D allele with HCM and DCM have been reporting in many studies (49,50)  
366 and the polymorphism considered as a modifier gene marker to cardiomyopathy. The DD  
367 genotype was also found to be associated with hypertension, restenosis, diabetes and  
368 myocardial infarction (51,52). However, absences of association (53,54) or influencing  
369 the cardiac phenotype (55) were reported in few other studies. These variations were  
370 partly accounted for ethnicity of the patients and to the sampling of the patients (56, 57).

371

## 372 **CONCLUSION:**

373 Among the two types of cardiomyopathies; dilated cardiomyopathy (73%) was found to  
374 be most predominant, whereas hypertrophic cardiomyopathy accounts for 27%. In  
375 general, increased male predominance (73%) was observed in both the types of  
376 cardiomyopathies. Familial occurrence was in 24% of the patients, parental consanguinity  
377 was 14% and 30% had family history of sudden cardiac death. Diastolic blood pressure  
378 of the cardiomyopathy patients was significant ( $p<0.05$ ) than the systolic blood pressure  
379 and heart rate. Also, they were characterized with higher Atrial Natriuretic Peptide and  
380 Brain Natriuretic Peptide ( $p<0.001$ ). Contrastingly co-morbid factors and other enzymes  
381 did not show much influence between patients and control subjects. Mutation screening  
382 of phospholamban gene revealed two transitions (4880 C/T, 4887 T/G) in 5' flanking  
383 region. Among them 4887 T/G transition was inherited in a hypertrophic cardiomyopathy

384 patient's family. Two transitions (5073 T/G, 5076 A/T) in exon 1 and two transitions (15507  
385 C/G, 15516 T/C) and an insertion (15512 T ins) in intron 1 were observed. Mutations  
386 located near to the promoter region have been defined as the fragment with maximal  
387 transcriptional activity. Mutations of this gene cause inherited dilated cardiomyopathy with  
388 refractory congestive heart failure. The protein is involved in blood circulation, and  
389 calcium ion transportation. No mutations occurred in the coding region of this gene so  
390 there is no change in the protein sequence consequently. Mutations located close to the  
391 promoter region have been defined as the fragment with maximal transcriptional activity.  
392 The increase or decrease in phospholamban activity due to the disruption around the  
393 promoter region can lead to cardiomyopathy. No mutations were observed in control  
394 subjects. Further, the deletion allele frequency of angiotensin-1- converting enzyme was  
395 considerably higher in cardiomyopathy patients (61.5%) than the control subjects (44%).

396

#### 397 **Availability of data and materials:**

398 All data relevant to the findings are available in this paper. We are bound by the  
399 participant consent which precludes data deposition in repositories. Further non-  
400 identifiable data is available upon request to **Dr. N. Gnana Veera Subhashini**  
401 **(dr.ngvsubhashini@bcgrc.com)**

402

#### 403 **Funding**

404 This work was Self-funded.

405 All data generated or analyzed during this study are included in this published article.

#### 406 **Authors' contributions**

407 GVS, MJ, BSL, EC and KMC made substantial contributions to conception and design of  
408 the experiments, or to acquisition, analysis, or interpretation of the data; GVS, EC and  
409 KMC were involved in drafting or critically revising the manuscript for important intellectual  
410 content; BSL and GVS gave final approval of the version to be published. All authors  
411 agree to be accountable for all aspects of the work, and will ensure that questions related  
412 to the accuracy or integrity of any part of the work are appropriately investigated and  
413 resolved. All authors read and approved the final manuscript.

#### 414 **Consent for publication**

415 The authors declare that they have received consent from study subjects/Participants for  
416 publication.

#### 417 **Competing interests**

418 The authors declare that they have no competing interests in relation to this manuscript  
419 or this study.

420

#### 421 **REFERENCES:**

422 1 Towbin, Jeffrey A. "Inherited Cardiomyopathies." *Circulation Journal*, vol. 78, no. 10,  
423 2014, pp. 2347–2356., doi:10.1253/circj.cj-14-0893

424 2 Maron, B J. "Contemporary Considerations for Risk Stratification, Sudden Death and  
425 Prevention in Hypertrophic Cardiomyopathy." *Heart*, vol. 89, no. 9, 2003, pp. 977–978.,  
426 doi:10.1136/heart.89.9.977.

427 3 Jacoby, D., and W. J. McKenna. "Genetics of Inherited Cardiomyopathy." *European*  
428 *Heart Journal*, vol. 33, no. 3, 2011, pp. 296–304., doi:10.1093/eurheartj/ehr260.

- 429 4 Vinten-Johansen, Jakob, et al. "Inflammation, Proinflammatory Mediators and  
430 Myocardial Ischemiaâ Reperfusion Injury." *Hematology/Oncology Clinics of North*  
431 *America*, vol. 21, no. 1, 2007, pp. 123–145., doi:10.1016/j.hoc.2006.11.010.
- 432 5 Codd MB, Sugrue DD, Gersh BJ. Epidemiology of idiopathic dilated and hypertrophic  
433 cardiomyopathy. A population-based study in Olmsted County, Minnesota, 1975-1984.  
434 *Circulation*. 1989; 80(3):564-72.
- 435 6 Maron BJ, Wolfson JK, Epstein SE, Roberts WC. Intramural ("small vessel") coronary  
436 artery disease in hypertrophic cardiomyopathy. *J Am Coll Cardiol*. 1986; 8(3):545-57
- 437 7 Schwartz. W.M; Louis M Bell Jr., Peter M Bingham; Esther K Chung  
438 et al., *The 5-Minute Pediatric Consult*, Fifth Edition, Lippincott Williams & Wilkins, March  
439 2008; ISBN 0-7817-7577-9
- 440 8 Hershberger, Ray E, et al. "Clinical and Genetic Issues in Dilated Cardiomyopathy: A  
441 Review for Genetics Professionals." *Genetics in Medicine*, vol. 12, no. 11, 2010, pp. 655–  
442 667., doi:10.1097/gim.0b013e3181f2481f.
- 443 9. Meyer, Sven, et al. "Sex Differences in Cardiomyopathies." *European Journal of Heart*  
444 *Failure*, Wiley-Blackwell, 16 Jan. 2014, onlinelibrary.wiley.com/doi/10.1002/ejhf.15/full.
- 445 10. Maron BJ, Wolfson JK, Epstein SE, Roberts WC. Intramural ("small vessel") coronary  
446 artery disease in hypertrophic cardiomyopathy. *J Am Coll Cardiol*. 1986; 8(3):545-57
- 447 11. McMurray, John J. V., et al. "Baseline Characteristics and Treatment of Patients in  
448 Prospective Comparison of ARNI with ACEI to Determine Impact on Global Mortality and  
449 Morbidity in Heart Failure Trial (PARADIGM-HF)." *European Journal of Heart Failure*,  
450 Wiley-Blackwell, 3 June 2014, onlinelibrary.wiley.com/doi/10.1002/ejhf.115/abstract.

- 451 12. Bui, A L, et al. "Epidemiology and Risk Profile of Heart Failure." *Nature Reviews.*  
452 *Cardiology.*, U.S. National Library of Medicine, Jan. 2011,  
453 [www.ncbi.nlm.nih.gov/pubmed/21060326](http://www.ncbi.nlm.nih.gov/pubmed/21060326).
- 454 13. Oxenoid, K., Chou, J. J. The structure of phospholamban pentamer reveals a channel-  
455 like architecture in membranes. *Proc. Nat. Acad. Sci.* 2005; 102: 10870-10875
- 456 14. McTiernan, C. F., Frye, C. S., Lemster, B. H., Kinder, E. A., Ogletree-Hughes, M. L.,  
457 Moravec, C. S., Feldman, A. M. The human phospholamban gene: structure and  
458 expression. *J. Molec. Cell Cardiol.* 1999; 31: 679-692
- 459 15. MacLennan, D.H., Kranias, E.G. Phospholamban: a crucial regulator of cardiac  
460 contractility. *Nat Rev Mol Cell Biol* 2003; 4: 566-577.
- 461 16. Schmitt JP, Kamisago M, Asahi M, Li GH, Ahmad F, Mende U, Kranias EG,  
462 MacLennan DH, Seidman JG, Seidman CE. Dilated cardiomyopathy and heart failure  
463 caused by a mutation in phospholamban. *Science.* 2003; 299:1410-1413.
- 464 17. Eric. "Susceptibility Genes and Modifiers for Cardiac Arrhythmias | Cardiovascular  
465 Research | Oxford Academic." *OUP Academic*, Oxford University Press, 15 Aug. 2005,  
466 [academic.oup.com/cardiovasces/article/67/3/397/505703/Susceptibility-genes-and-](http://academic.oup.com/cardiovasces/article/67/3/397/505703/Susceptibility-genes-and-modifiers-for-cardiac)  
467 [modifiers-for-cardiac](http://academic.oup.com/cardiovasces/article/67/3/397/505703/Susceptibility-genes-and-modifiers-for-cardiac).
- 468 18. Fatkin<sup>1</sup>, Diane, and Christine E. Seidman<sup>4</sup>. "Diane Fatkin." *Cold Spring Harbor*  
469 *Perspectives in Medicine*, 1 Jan. 1970,  
470 [perspectivesinmedicine.cshlp.org/lookup/doi/10.1101/cshperspect.a021063](http://perspectivesinmedicine.cshlp.org/lookup/doi/10.1101/cshperspect.a021063).
- 471 19. Marian, AJ. "Modifier Genes for Hypertrophic Cardiomyopathy." *Current Opinion in*  
472 *Cardiology*, [europepmc.org/articles/PMC2775140](http://europepmc.org/articles/PMC2775140).

- 473 20. Rudnicki, Michael, and Gert Mayer. "Significance of Genetic Polymorphisms of the  
474 Reninâ Angiotensinâ Aldosterone System in Cardiovascular and Renal  
475 Disease." *Pharmacogenomics*, vol. 10, no. 3, 2009, pp. 463–476.,  
476 doi:10.2217/14622416.10.3.463.
- 477 21. Charron P, Komajda M. Are we ready for pharmacogenomics in heart failure? *Eur J*  
478 *Pharmacol.* 2001; 417:1-9
- 479 22. Heribert Schunkert, Hans-Werner Hense, Stephan R et al., "Association between a  
480 Deletion Polymorphism of the Angiotensin-Converting-Enzyme Gene and Left Ventricular  
481 Hypertrophy NEJM." *New England Journal of Medicine*,  
482 [www.nejm.org/doi/10.1056/NEJM199406093302302](http://www.nejm.org/doi/10.1056/NEJM199406093302302).
- 483 23. Lahiri DK, Schnabel B. DNA isolation by a rapid method from human blood samples:  
484 effects of MgCl<sub>2</sub>, EDTA, storage time, and temperature on DNA yield and quality.  
485 *Biochem Genet.* 1993; 31: 321–328.
- 486 24. Susumu Minamisawa, Yoji Sato, Yuriko Tatsuguchi, Tomofumi Fujino, Shinichiro  
487 Imamura, Yoshio Uetsuka, Makoto Nakazawa and Rumiko Matsuoka. Mutation of the  
488 phospholamban promoter associated with hypertrophic cardiomyopathy *Biochemical and*  
489 *Biophysical Research Communications.* 2003; 304.
- 490 25. Rigat, B., Hubert, C., Alhenc-Gelas, F et al., An insertion/deletion polymorphism in  
491 the angiotensin I-converting enzyme gene accounting for half the variance of serum  
492 enzyme levels. *J. Clin. Invest.*, 1990; 86, 1343–1346.
- 493 26. Zhang, Ling, et al. "Interaction of Angiotensin I-Converting Enzyme Insertion-Deletion  
494 Polymorphism and Daily Salt Intake Influences Hypertension in Japanese

- 495 Men." *Hypertension Research*, vol. 29, no. 10, 2006, pp. 751–758.,  
496 doi:10.1291/hypres.29.751.
- 497 27. RÄcken, Christoph, et al. "The Gene Polymorphism of the Angiotensin I-Converting  
498 Enzyme Correlates with Tumor Size and Patient Survival in Colorectal Cancer  
499 Patients." *Neoplasia*, vol. 9, no. 9, 2007, pp. 716–722., doi:10.1593/neo.07418.
- 500 28. Sparks, Matthew A., et al. "Classical Renin-Angiotensin System in Kidney  
501 Physiology." *Comprehensive Physiology*, 2014, pp. 1201–1228.,  
502 doi:10.1002/cphy.c130040.
- 503 29. Leońska-Duniec, A, et al. *Biology of Sport*, Institute of Sport in Warsaw, Sept. 2016,  
504 [www.ncbi.nlm.nih.gov/pmc/articles/PMC4993135/](http://www.ncbi.nlm.nih.gov/pmc/articles/PMC4993135/).
- 505 30. Sisakian, Hamayak. "Cardiomyopathies: Evolution of Pathogenesis Concepts and  
506 Potential for New Therapies." *World Journal of Cardiology*, Baishideng Publishing Group  
507 Inc., 26 June 2014, [www.wjgnet.com/1949-8462/full/v6/i6/478.htm](http://www.wjgnet.com/1949-8462/full/v6/i6/478.htm).
- 508 31. Elliott PM et al. Sudden death in hypertrophic cardiomyopathy: identification of high  
509 risk patients. *J Am Coll Cardiol*. 2000; 36: 2212–2218
- 510 32. Biagini E et al. Dilated-hypokinetic evolution of hypertrophic cardiomyopathy:  
511 prevalence, incidence, risk factors, and prognostic implications in pediatric and adult  
512 patients. *J Am Coll Cardiol*. 2005; 46: 1543–1550
- 513 33. Christiaans I, Erwin Birnie, Irene M. van Langen et al, The yield of risk stratification  
514 for sudden cardiac death in hypertrophic cardiomyopathy myosin-binding protein C gene  
515 mutation carriers: focus on predictive screening. *Eur Heart J*. 2010; 31 (7): 842-848.
- 516 34. Rossi A, Cicoira M, Golia G, Anselmi M, Zardini P. Mitral regurgitation and left

- 517 ventricular diastolic dysfunction similarly affects mitral and pulmonary vein flow Doppler  
518 parameters: the advantage of end-diastolic markers. *J Am Soc Echocardiogr.* 2001;  
519 14(6):562-8.
- 520 35. Fung DC, Yu B, Littlejohn T, Trent RJ. An online locus-specific mutation database for  
521 familial hypertrophic cardiomyopathy. *Hum Mutat.* 1999; 14: 326-32.
- 522 36. Sugimoto K, Katsuya T, Ohkubo T, Hozawa A et al, Association between angiotensin  
523 II type 1 receptor gene polymorphism and essential hypertension: the Ohasama Study.  
524 *Hypertens Res.* 2004; 27(8):551-6.
- 525 37. Kranias EG, Bers DM. Calcium and cardiomyopathies. *Subcell Biochem,* 2007; 45:  
526 523– 537.
- 527 38. Traaseth NJ, et al. Structural and dynamic basis of phospholamban and sarcolipin  
528 inhibition of  $Ca^{2+}$ -ATPase. *Biochemistry.* 2008; 47:3–13.
- 529 39. Vafiadaki E, Demetrios A. Arvanitis, Stamatis N. Pagakis, Vasiliki Papalouka, Despina  
530 Sanoudou, Aikaterini Kontrogianni-Konstantopoulos and Evangelia G. Kranias. The Anti-  
531 apoptotic Protein HAX-1 Interacts with SERCA2 and Regulates Its Protein Levels to  
532 Promote Cell Survival. *Mol. Biol. Cell.* 2009; vol. 20 no. 1 306-318.
- 533 40. Medin M, Hermida-Prieto M, Monserrat L, Laredo R, Rodriguez-Rey JC, Fernandez  
534 X, Castro-Beiras A. Mutational screening of phospholamban gene in hypertrophic and  
535 idiopathic dilated cardiomyopathy and functional study of the PLN -42 C>G mutation. *Eur*  
536 *J Heart Fail.* 2007; Jan; 9(1): 37-43.
- 537 41. Haghghi K, Chen G, Sato Y, Fan GC, He S, Kolokathis F, Pater L, Paraskevaidis I,  
538 Jones WK, Dorn GW 2nd, Kremastinos DT, Kranias EG. A human phospholamban



- 539 promoter polymorphism in dilated cardiomyopathy alters transcriptional regulation by  
540 glucocorticoids. *Hum Mutat.* 2008; (5):640-7.
- 541 42. Kalemi T, Efthimiadis G, Zioutas D, Lambropoulos A, Mitakidou A, Giannakoulas G,  
542 Vassilikos V, Karvounis H, Kotsis A, Parharidis G and Louridas G. Phospholamban gene  
543 mutations are not associated with hypertrophic cardiomyopathy in a Northern Greek  
544 population. *Biochem Genet.* 2005; 43(11-12):637-42
- 545 43. Santos GB Diogo, Alessandra Medeiros, Patrícia C Brum, José G Mill, Alfredo J  
546 Mansur, José E Krieger and Alexandre C Pereira. No evidence for an association  
547 between the -36A>C phospholamban gene polymorphism and a worse prognosis in heart  
548 failure. *BMC Cardiovasc Disord.* 2009; 9: 33.
- 549 44. Simmerman, H. K. B., and L. R. Jones. Phospholamban: protein structure,  
550 mechanism of action, and role in cardiac function. *Physiol. Rev.* 1998; 78:921–947
- 551 45. Buckley MG, Markandu ND, Sagnella GA, MacGregor GA. Brain and atrial natriuretic  
552 peptides: a dual peptide system of potential importance in sodium balance and blood  
553 pressure regulation in patients with essential hypertension. *J Hypertens.* 1994; 27:809-  
554 13.
- 555 46. Candy GP, Skudicky D, Mueller UK, Woodiwiss AJ et al., Association of left ventricular  
556 systolic performance and cavity size with angiotensin-converting enzyme genotype in  
557 idiopathic dilated cardiomyopathy. *Am J Cardiol.* 1999; 83: 740-4.

- 558 47. Rai TS, Dhandapany PS, Ahluwalia TS et al. ACE I/D polymorphism in Indian patients  
559 with hypertrophic and dilated cardiomyopathy. *Molecular and cellular biochemistry* 2008;  
560 311(1-2): 67-72.
- 561 48. Mahjoub S, Mehri S, Bousaada R, Ouarda F, Zaroui A et al, Association of ACE I/D  
562 polymorphism in Tunisian patients with dilated cardiomyopathy. *Journal of Renin-  
563 Angiotensin-Aldosterone System*. 2010 vol. 11 no. 3 187-191.
- 564 49. Raynolds MV, Bristow MR, Bush EW, Abraham WT et al, Angiotensin-converting  
565 enzyme DD genotype in patients with ischaemic or idiopathic dilated cardiomyopathy.  
566 *Lancet*. 1993; 342:1073–1075.
- 567 50. Kawaguchi H. Angiotensin-converting enzyme and angiotensinogen gene  
568 polymorphism in hypertrophic cardiomyopathy. *Exp Clin Cardiol*. 2003; 8(3): 155–159.
- 569 51. Dzau, V. J. Cell biology and genetics of angiotensin in cardiovascular disease. *J.*  
570 *Hypertens*. 1994; 12, 3-10.
- 571 52. Gardemann, A., Fink, M., Sticker, J., Nguyen, Q. D., Humme, J., Katz, N et al., ACE  
572 I/D polymorphism: presence of the ACE D allele increases the risk of coronary artery  
573 disease in younger individuals. *Atherosclerosis*. 1998; 139, 153-159.
- 574 53. Montgomery HE, Keeling PJ, Goldman JH, Humphries SE, Talmud PJ, McKenna WJ.  
575 Lack of association between the insertion/deletion polymorphism of the angiotensin  
576 converting enzyme gene and idiopathic dilated cardiomyopathy. *J Am Coll Cardiol*. 1995;  
577 25:1627-1631.
- 578 54. Yamada Y, Ichihara S, Fujimura T, Yokota M. Lack of association of polymorphisms  
579 of the angiotensin converting enzyme and angiotensinogen genes with nonfamilial  
580 hypertrophic or dilated cardiomyopathy. *Am J Hypertens* 1997; 10: 921-928.

- 581 55. Mahjoub S, Mehri S, Bousaada R, et al. Association of ACE I/D polymorphism in  
 582 Tunisian patients with dilated cardiomyopathy. Journal of the Renin-Angiotensin-  
 583 Aldosterone system. 2011; 11(3): 187-191.
- 584 56. Wang, P., Zou, Y., Fu, C. Y., Zhou, X., Hui, R. MYBPC3 polymorphism is a modifier  
 585 for expression of cardiac hypertrophy in patients with hypertrophic cardiomyopathy.  
 586 Biochem. Biophys. Res. Commun. 2005; 329: 796-799.
- 587 57. Pfohl, M., Koch, M., Prescod, S., Haase, K. K., Haring, H. U. and Karsch, K. R.  
 588 Angiotensin I- converting enzyme gene polymorphism, coronary artery disease and  
 589 myocardial infarction. An angiographic ally controlled study. Eur. Heart J. 1999; 20, 1318-  
 590 1325.

591

592

593

594

**Table 1. Baseline, Hemodynamic and Biochemical Characteristics**

<b>Characteristics</b>	<b>Cases n=109</b>	<b>Control n=100</b>	<b>Cases vs Control P value</b>
<b>Age, Yrs</b>	37.22 ± 12.43	40 ± 12.65	0.111
<b>Gender</b>			
<b>Male, n</b>	73	52	0.034*
<b>Female, n</b>	36	48	
<b>Parental Consanguinity, n (%)</b>	15(13.76)	9	0.386
<b>BMI (kg/m<sup>2</sup>)</b>	24.65 ± 4.08	23.76 ± 3.33	0.085

<b>BSA (m2)</b>	1.79 ± 0.17	1.84 ± 0.75	0.515
<b>Sys BP (mmHg)</b>	125.0 ± 9.17	120.0 ± 14	0.003*
<b>Dys BP(mmHg)</b>	81.41 ± 4.76	78.0 ± 10.0	0.002*
<b>Heart Rate (beats/min)</b>	74.06 ± 4.08	72.3 ± 4.5	0.004*
<b>Familial status, n (%)</b>	26(23.85)		
<b>F/H of SCD, n (%)</b>	33(30.28)		
<b>NYHA</b>			
<b>Class I, II, n %)</b>	37(33.94)		
<b>Class III, IV, n (%)</b>	72 (66.06)		
<b>ANP (pg/ml))</b>	130.21 ± 24.67	21.9 ± 17.6	<0.001
<b>BNP (pg/ml)</b>	110.36 ± 110.12	8.12 ± 8.08	<0.001
<b>CpK ≥ 37U/L, n (%)</b>	26(23.85)	21	0.74
<b>SGOT ≥ 13U/L, n (%)</b>	35(32.11)	20	0.059
<b>SGPT ≥ 17U/L, n (%)</b>	32(29.36)	30	1
<b>Glucose (mg/dl))</b>	96.16 ± 8.0	95 ± 7	0.265
<b>Urea (mg/dl)</b>	27.36 ± 2.29	27.92 ± 4.12	0.232
<b>Creatinine (mg/dl)</b>	0.90 ± 0.32	0.84 ± 0.19	0.098
<b>HTN, n (%)</b>	31(28.44)	17	0.07
<b>DM, n (%)</b>	39(35.78)	27	0.183
<b>Obesity, n (%)</b>	22(20.18)	21	1

<b>Current and Ex-Smokers, n (%)</b>	64(58.72)	72	0.059
<b>Current and Ex-Alcoholics, n (%)</b>	54(49.54)	56	0.406

595

596

597 Data shown as Mean  $\pm$  Standard deviation or number or number (%) F/H of SCD- Family  
 598 history of sudden cardiac death, BMI- Body mass index, BSA- Body surface area, sys  
 599 BP, Dys BP systolic, diastolic blood pressure, NYHA- New York heart association  
 600 functional class, ANPatrial natriuretic peptide, BNP- Brain natriuretic peptide, CpK-  
 601 Creatine phosphokinase, SGOTSerum Aspartate transaminase, SGPT- Serum alanine  
 602 transaminase, HTN- Hypertension, DM Diabetes Mellitus. P value is probability of chi-  
 603 square with one degree of freedom of genotype frequencies in control and case datasets.

604 \*Pvalue<0.05, \*\* Pvalue< 0.01, \*\*\* Pvalue< 0.001.

605

606 **Table 2 Clinical characteristics of Patients and control**

<b>Characteristics</b>	<b>DCM n=80+47=127</b>	<b>HCM n=29+23=52</b>	<b>Control n=100</b>
<b>LVESD(mm)</b>	46.0 $\pm$ 6***	26 $\pm$ 7***	31 $\pm$ 5
<b>LVEDD(mm)</b>	71.5 $\pm$ 2***	69 $\pm$ 7***	52 $\pm$ 3
<b>LVEF (%)</b>	65 $\pm$ 6***	36 $\pm$ 7***	53 $\pm$ 7
<b>FS (%)</b>	39 $\pm$ 7**	38 $\pm$ 8	36 $\pm$ 7
<b>IVST(mm)</b>	18 $\pm$ 6***	11 $\pm$ 3***	8 $\pm$ 2

<b>PWT(mm)</b>	22 ± 5 <sup>***</sup>	12 ± 2 <sup>***</sup>	8 ± 2
<b>LVMI (gm/m<sup>2</sup>)</b>	106 ± 25 <sup>***</sup>	106 ± 49 <sup>***</sup>	61 ± 30
<b>LV Volume/mass ratio (mL/gm)</b>	0.77 ± 0.6 <sup>***</sup>	0.18 ± 0.1 <sup>***</sup>	0.30 ± 0.07
<b>LVESV (ml)</b>	127 ± 61 <sup>***</sup>	38 ± 17 <sup>*</sup>	32 ± 11
<b>LVEDV (ml)</b>	179 ± 7 <sup>***</sup>	103 ± 37 <sup>*</sup>	86 ± 24
<b>LVOT gradient &gt;20mmHg, n (%)</b>	96 (75.59) <sup>**</sup>	39 (75)	57
<b>E/A ratio</b>	1.7 ± 1.1 <sup>*</sup>	1.4 ± 0.9	1.3 ± 1
<b>IVPG (mmHg)</b>	1.1 ± 0.3 <sup>***</sup>	1.4 ± 0.6 <sup>*</sup>	1.6 ± 0.4
<b>LAVI (ml/m<sup>2</sup>)</b>	38 ± 14 <sup>***</sup>	30 ± 9 <sup>***</sup>	18 ± 7
<b>QRS width (ms)</b>	145 ± 11 <sup>***</sup>	106 ± 14 <sup>***</sup>	91 ± 16
<b>PR (ms)</b>	112 ± 11 <sup>***</sup>	101 ± 9 <sup>***</sup>	126 ± 8
<b>Max QTc (ms)</b>	434 ± 24 <sup>***</sup>	455 ± 26 <sup>***</sup>	400 ± 25

607

608

609 Data shown as Mean ± Standard deviation or number (%) LVESD- Left Ventricular End  
610 systolic diameter, LVEDD-Left ventricular end diastolic diameter, LVEF- Left ventricular  
611 ejection fraction, FS- Fractional shortening, IVST- Inter ventricular septal thickness, PWT-  
612 Posterior wall thickness, LVMI- Left ventricular mass index, LAVI- Left atrial volume index,  
613 LVOT- Left ventricular outflow tract, E/A ratio- early-to-late transmitral flow velocity, IVPG-  
614 Intraventricular pressure gradient. P value is probability of chi-square with one degree of

615 freedom of genotype frequencies in control and case datasets. \*Pvalue $\leq$  0.05, \*\* Pvalue $<$   
 616 0.01, \*\*\* Pvalue $<$  0.001.

617

618 **Table 3: Observed Nucleotide Changes in Phospholamban Gene**

<b>Nucleotide changes</b>	<b>Loci</b>	<b>Disease type</b>	<b>Familial case</b>
4880 C/T	5' flanking region	DCM	No
4887 T/G	5' flanking region	HCM	Yes
5073 T/G	Exon1	DCM	No
5076 A/T	Exon1	DCM	No
15507 C/G	Intron1	DCM	No
15512 ins T	Intron1	DCM	Yes
15516 T/C	Intron1	DCM	No

619

620 **Table 4. Alu ACE1 Genotype and Allele frequencies in control and Patients with**  
 621 **cardiomyopathy**

Subject	Genotype			Allele Frequencies		$\chi^2$
	II	ID	DD	I	D	
<b>Control</b>	34	44	22	0.56	0.44	1.148
<b>Patients</b>	17	50	42	0.385	0.615	0.109

622

623

624 **Table 5. Frequency distribution of Alu ACE1 Genotype and allele frequency in**  
625 **Control and Patients of cardiomyopathy**

Case	Vs	$\chi^2$	P value	ODDs	95 % CI	
				Ratio		
<b>ID vs DD</b>		1.9358	0.1641	0.6	0.309	1.147
<b>DD vs II</b>		10.589	0.0011	3.82	1.754	8.311
<b>ID vs II</b>		4.4768	0.0344	2.27	1.118	4.619
<b>DD + ID vs II</b>		8.6037	0.0034	2.79	1.437	5.408
<b>DD vs ID + II</b>		5.9538	0.0147	2.22	1.207	4.092
<b>D vs I</b>		12.088	0.0005	1.14	0.773	1.687

626

627 The present study investigates the association of spectrum of clinical symptoms and the  
628 epidemiological variables like age, sex, familial status, and parental consanguinity. The  
629 current study also evaluates Angiotensin-1-Converting Enzyme gene polymorphism and  
630 its role as a marker / modifier in Cardiomyopathy.

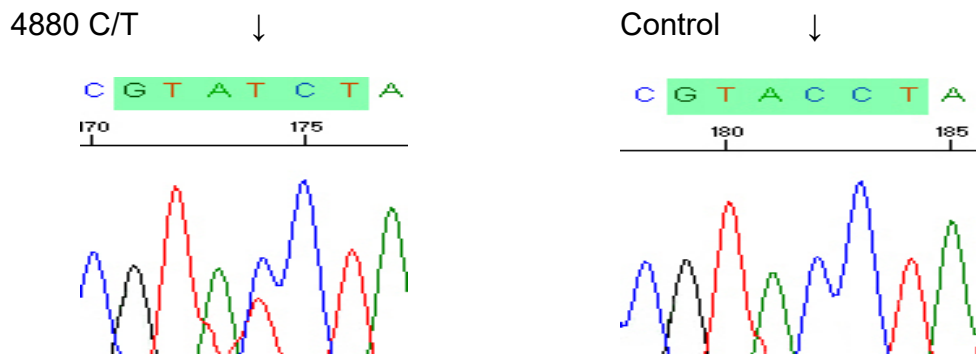
631



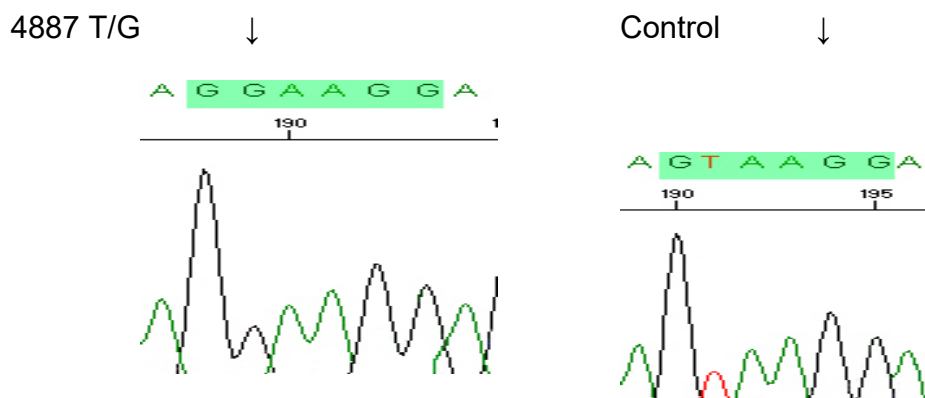
632 **Figure 1: Chromatogram of Phospholamban Gene Mutation**

633

634 (a) Representative Chromatograms showing 4880 C/T

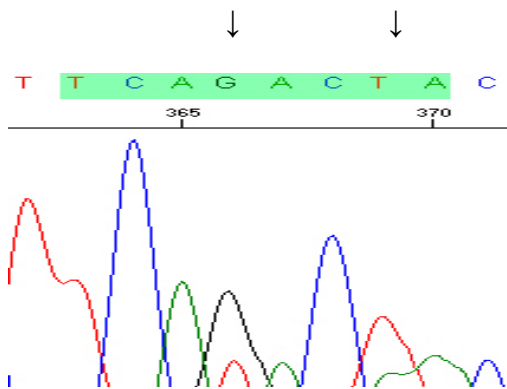


(b) Representative Chromatograms showing 4887 T/G

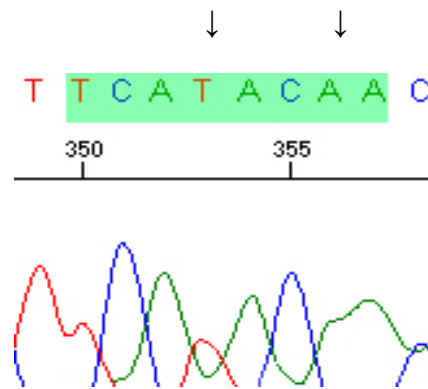


(C) Representative Chromatograms showing 5073 T/G, 5076 A/T

5073 T/G, 5076 A/T

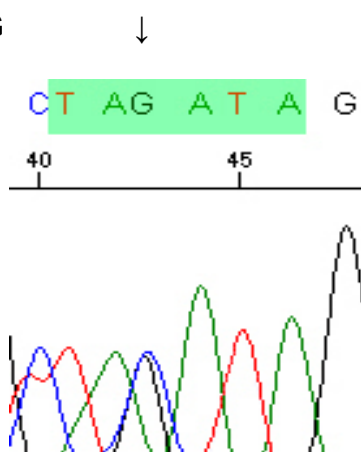


Control

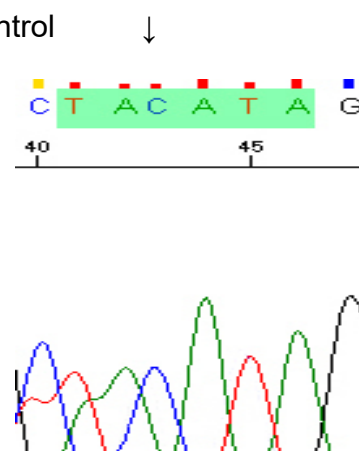


(d) Representative Chromatograms showing 15507 C/G

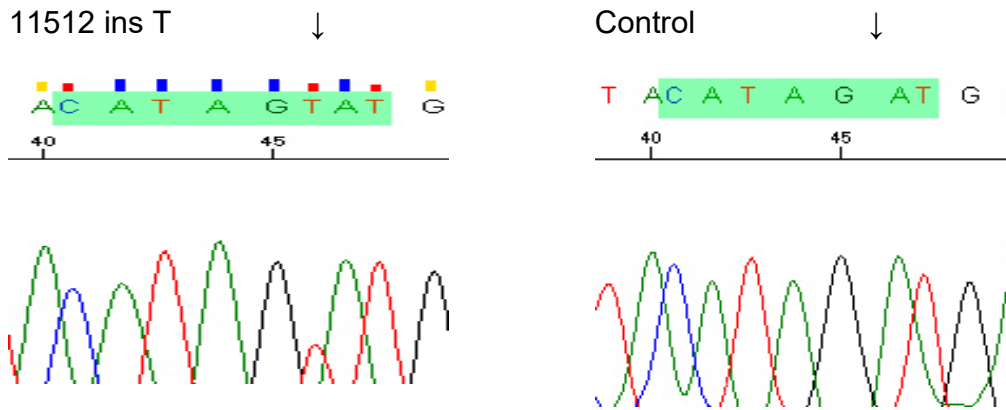
15507 C/G



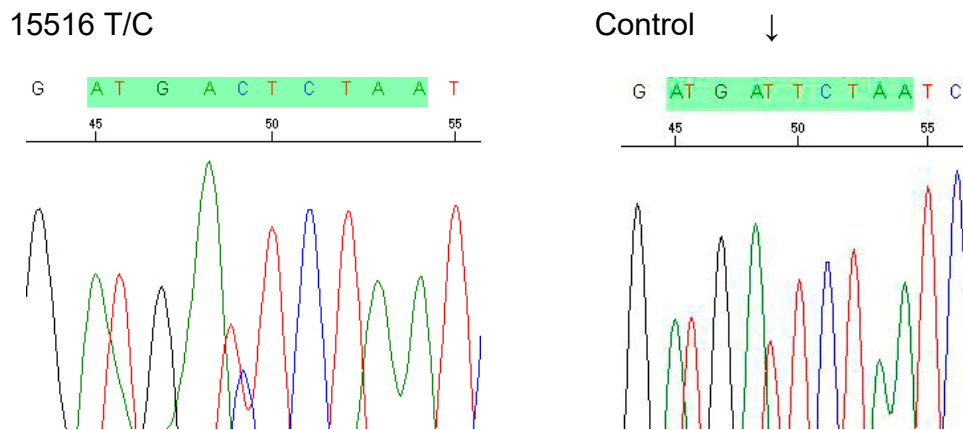
Control



(e) Representative Chromatograms showing 15512 ins T



(f) Representative Chromatograms showing 15516 T/C



635

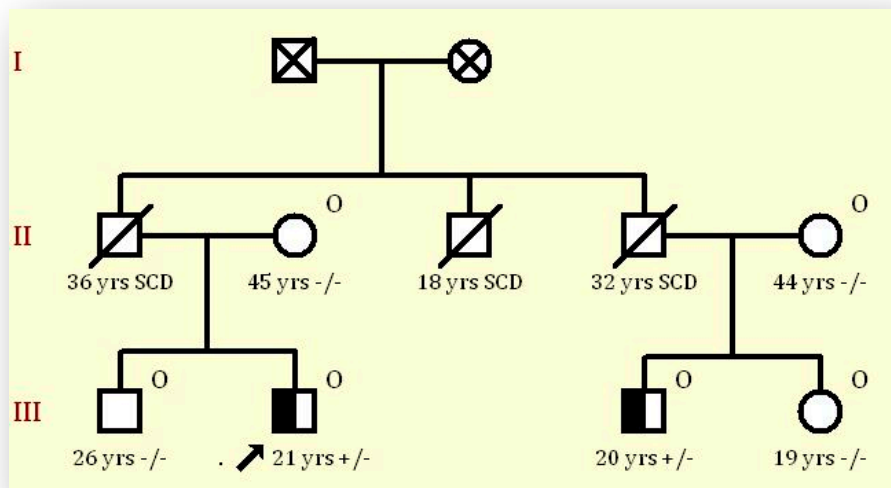
636

637

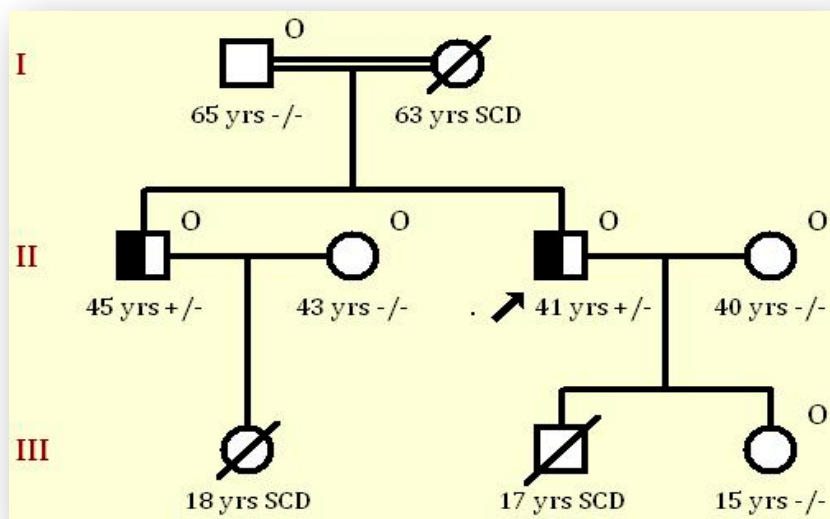
638

639

640

641 **Figure 2: Pedigree of H9 proband with 4887 T→G mutations in 5'flanking region of**642 **PLN gene**

643

644 **Figure 3: Pedigree of H10 proband with 4887 T→G mutations in 5'flanking region**645 **of PLN gene**

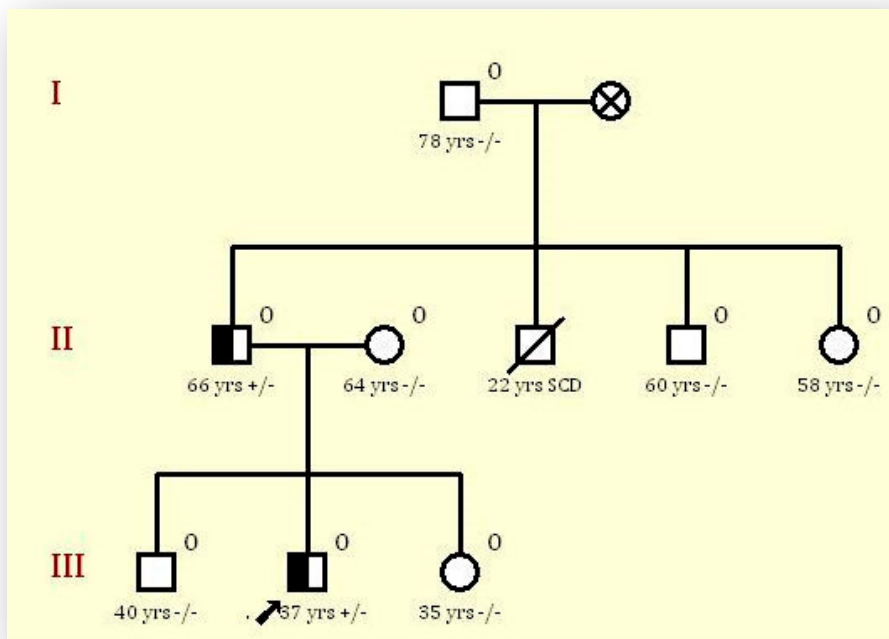
646

647 **Figure 4: Pedigree of D15 proband with 15512 ins T mutations in intron 1 of PLN**

648 **gene**

649

650



651

652 **Note:** +/- presence of mutation in heterozygote state; -/- absence of mutation; ♂ -

653 Probant;

654 o- Gene Sequenced; Age below each indicates age at investigation