

Changes in the Geographic Distribution of the Diana Fritillary (*Speyeria diana*: Nymphalidae) Under Forecasted Predictions of Climate Change

Carrie N. Wells^{1*}, and David Tonkyn²

¹University of North Carolina at Charlotte, Department of Biological Sciences, 9201 University City Blvd, Charlotte, NC 28223, E-mail: Carrie.Wells@uncc.edu

²University of Arkansas at Little Rock, Department of Biology, 2801 South University Ave., Little Rock, AR 72204, Email: dwtonkyn@ualr.edu

*Corresponding author:

Dr. Carrie Wells
Department of Biological Sciences
University of North Carolina at Charlotte
9201 University City Blvd, Charlotte, NC 28223 USA
Phone: 704-687-8513
Fax: 704-687-3128
E-mail: Carrie.Wells@uncc.edu

Abstract

Climate change is predicted to alter the geographic distribution of a wide variety of taxa, including butterfly species. Research has focused primarily on high latitude species in North America, with no known studies examining responses of taxa in the southeastern US. The Diana fritillary (*Speyeria diana*) has experienced a recent range retraction in that region, disappearing from lowland sites and now persisting in two, phylogenetically disjunct mountainous regions. These findings are consistent with the predicted effects of a warming climate on numerous taxa, including other butterfly species in North America and Europe. We used ecological niche modeling to predict future changes to the distribution of *S. diana* under several climate models. To evaluate how climate change might influence the geographic distribution of this butterfly, we developed ecological niche models using Maxent. We used two global circulation models, CCSM and MIROC, under low and high emissions scenarios to predict the future distribution of *S. diana*. Models were evaluated using the Receiver Operating Characteristics Area Under Curve test and the True Skill Statistics (mean AUC = 0.91 ± 0.0028 SE, TSS = 0.87 ± 0.0032 SE for RCP = 4.5, and mean AUC = 0.87 ± 0.0031 SE, TSS = 0.84 ± 0.0032 SE for RCP = 8.5), which both indicate that the models we produced were significantly better than random (0.5). The four modeled climate scenarios resulted in an average loss of 91% of suitable habitat for *S. diana* by 2050. Populations in the Southern Appalachian Mountains were predicted to suffer the most severe fragmentation and reduction in suitable habitat, threatening an important source of genetic diversity for the species. The geographic and genetic isolation of populations in the west suggest that those populations are equally as vulnerable to decline in the future, warranting ongoing conservation of those populations as well. Our results suggest that the Diana fritillary is under threat of decline by 2050 across its entire distribution from climate change, and is likely to be negatively affected by other human-induced factors as well.

Keywords: *Speyeria diana*, butterfly, conservation, fragmentation, global warming, Maxent, WorldClim

Introduction

Understanding how species distributions might shift with changing climate is a critical

component of managing and protecting future biodiversity. Hundreds of species in the US and elsewhere have responded to the warming climate by shifting to higher latitudes or elevations [1-4]. Such range shifts have been documented in a number of taxa [5-7], including alpine plants [8], marine invertebrates [9], marine fishes [10], mosquitoes [11], birds [12, 13], and butterflies [1, 14-18]. A number of species distribution models have been developed to predict the impacts of climate change on species distributions, including bioclimate envelope models, which are useful first estimates of the potential effects of climate change on altering species' ranges [19]. Bioclimate envelope models work by identifying the climatic bounds within which a species currently occurs, and then delineating how those climatic bounds will shift under various future climate projections [20-23].

Most often, researchers are limited to presence-only occurrence data, requiring the use of indirect methods to infer a species' climatic requirements [8, 24-25]. One of the best performing models using presence-only data is maximum entropy modeling, or Maxent [26], which performs well even with low sample sizes typical of rare species [19, 27-28]. Maxent works by comparing climate data from occurrence sites with those from a random sample of sites from the larger landscape to minimize the relative entropy of statistical models' fit to each data set. Species distribution models such as Maxent have been criticized for being overly simplistic, since they do not incorporate external biotic factors such as species interactions [20, 27, 29]. However, such bioclimate envelope models have been used to project with reasonable accuracy whether species ranges will increase or decrease under a changing climate [19, 30-32], which was the primary objective of this study.

Speyeria diana (Nymphalidae) (Cramer 1777) is a butterfly species endemic to the southeastern US and is currently threatened across portions of its range. This species is of particular conservation interest because it has experienced a range collapse in recent decades resulting in an 800 km geographic and genetic disjunction between western populations in the Ouachita and Ozark Mountains and populations in the southern Appalachian Mountains, and has shifted to a higher elevation at an estimated rate of 18m per decade [33]. This range contraction is consistent with the predicted effects of a warming climate, and might represent the first such documented case in the southeastern US, though the region has experienced other environmental changes in recent decades as well [33]. Previous research using coalescent-based population divergence models dated the earliest splitting of the western population from the east at least 20,000 years ago, during the Last Glacial Maximum [34]. In addition, recent geometric morphometric evidence from the wings of *S. diana* further support this long-term spatial and genetic isolation [35]. In light of these pieces of evidence, we used Maxent to model the future distribution of *S. diana* under several future climatic scenarios, to forecast how the range of the butterfly might shift under predicted conditions. Forecasts of large range reductions, or small overlap between current and future ranges, would suggest high vulnerability to climate change. Range reductions of any size in the western distribution would likely threaten those populations, that are genetically isolated and adapted to relatively low dispersal, with the negative effects of genetic drift [34, 35].

Methods

Study species

The Diana fritillary, *Speyeria diana*, is a large and sexually dimorphic nymphalid butterfly, endemic to the southeastern US. Adult males emerge in late May to early June, with females flying several weeks to a month later [36]. Once mated, each female can lay thousands of eggs singly on ground litter during the months of August and September in the vicinity of *Viola* spp., the larval host plant for all *Speyeria* [37]. After hatching, first instar larvae immediately burrow

deep into the leaf litter layer of the forest floor, where they overwinter [38]. In spring, larvae feed on the foliage of freshly emerging violets. Adult Diana butterflies are often found along forest edges or dirt roads containing tall, conspicuous nectar sources such as milkweeds, butterfly bushes or other large summer and fall composites [39-42]. While males begin to die off in late July, females may persist in large numbers, although somewhat cryptically, through October [42].

Distributional dataset

We searched for all known records of *S. diana*, from publications, catalogued and uncatalogued specimens in public and private collections in the United States and Europe, online databases, contemporary field surveys by scientists and amateurs, and our own field surveys. We obtained distributional data from 1,323 pinned *S. diana* specimens from 38 natural history museum collections in the United States and Europe. Four hundred thirty-five additional records (1938–2012) were provided by the Butterfly and Moth Information Network and the participants who contribute to its BAMONA project. Our literature survey produced 153 records (1818–2011) across 54 US counties. We also collected 419 *S. diana* butterflies in our own field surveys. Our dataset represents essentially a complete dataset of all publicly available records for the species, and is as comprehensive as for any taxon in the region [33]. For this reason, our dataset should be especially informative in creating an accurate bioclimate envelope for the species, as collection bias is a major consideration with ecological niche modeling [43, 44].

Species distributional modeling

We developed species distribution models using the popular machine-learning algorithm for ecological modeling, Maxent [26]. Maxent estimates a species' probability distribution that has maximum entropy (closest to uniform), subject to a set of constraints based on the sampling of presence-only data [45]. Because of the difficulty and impracticality of obtaining accurate absence data, presence-only data are most often used in species distribution modeling. In order to offset the lack of absence data, Maxent uses a background sample to compare the distribution of presence data along environmental gradients with the distribution of background points randomly drawn from the study area [46-48]. Locality data and the randomly sampled background points are combined with climatic data to predict the probability of the species' occurrence within each raster grid cell. We used environmental climate data from WorldClim [49] at 30 arc-second resolution or approximately 1 km² grid cells. Bioclimate variables and elevation layers were each clipped to the extent of North America using ESRI ArcMap 10.0, and data extracted to *S. diana* sample localities. Additionally, we collected the same types of locality data for three other species of North American butterflies (*Speyeria cybele*, *Speyeria idalia*, *Battus philenor*), which served as 5,628 random background points for our models. We utilized these background data to minimize spatial bias in our modeling, as data represented by similar butterfly species can be used as pseudo-absence data with the same collection bias as our occurrence data, improving the accuracy of the model [50, 51].

Climatic variables included 19 derived bioclimatic variables that describe annual and seasonal variation in temperature and precipitation averaged for 1950-2000, as well as elevation (Table 1). One concern when modeling species distributions is the strong correlation that occurs between multiple climate variables, which can significantly influence model predictions of species distributions [52]. To test for co-linearity, we performed spatial autocorrelation statistics between all pairs of the 19 bioclimate variables using ESRI ArcMap 10.0. We then selected the biologically most meaningful variable for each group of two or more variables with Pearson correlation coefficients higher than 0.7 (Table 1). This allowed us to reduce the number of bioclimate variables to the 9 most potentially important ones, which were: Minimum

Temperature of Coldest Month, Mean Temperature of Driest Quarter, Precipitation of Wettest Month, Precipitation of Driest Month, Precipitation of Driest Quarter, Isothermality, Mean Diurnal Range (Mean of monthly (maximum temperature - minimum temperature)), Temperature Annual Range, and Annual Precipitation, along with elevation (Table 1). These variables are typically considered to be important determinants of butterfly distributions, as they relate to life history traits. Butterflies are highly sensitive to weather and climate, particularly changes in temperature and rainfall [53]. For example, mean temperature of the coldest month is related to the overwintering survival of first instar larvae, growing degree days above 5°C is regarded as a surrogate for the developmental threshold of the larvae, water balance corresponds to the moisture availability for the larval host and adult nectar plants, and the mean temperature of late summer ensures proper adult emergence and mating [54-59]. Temperature changes affect all aspects of butterfly life history, from their distribution and abundance [14, 54], to their realized fecundity [60-61]. Changes in rainfall levels can influence butterfly larvae indirectly through changes in host plant quality, and generally rainfall is considered to be beneficial because it enhances host plant growth [62].

One concern when modeling species distributions is whether the occurrence records are spatially biased with respect to site accessibility (e.g., towns, roads, trails) [63]. To address this concern, we applied a spatial filter to remove all sampling points that were within 5 km of each other using ESRI ArcMap 10.0. The spatial filter resulted in 254 unique presence points for *S. diana* that were used in the final model. We first modeled the distribution of these 254 occurrences in present-day climate, and then projected the fitted species distribution under two future climate scenarios for the period 2040-2069 (hereafter referred to as 2050). Future climate scenarios were taken from two global circulation models (GCMs) obtained from www.worldclim.org: the Community Climate System Model (CCSM) [64] and the Model for Interdisciplinary Research on Climate (MIROC) [65-66]. These GCMs differ in the reconstruction of several climatic variables and are well known to produce different outcomes for butterfly species [67-68]. For example, in hind-casting Mediterranean butterflies, the CCSM model projects narrower distributions at the last glacial maximum than does MIROC [65-66]. For each of these two GCMs, we considered two different representative concentration pathways (RCPs) [69-73], which are cumulative measures of human emissions of greenhouse gases from all sources expressed in Watts per square meter. These pathways were developed for the Fifth Assessment Report of the Intergovernmental Panel on Climate Change [67] and correspond to a total anthropogenic radiative forcing of RCP = 4.5 W/m² (low) and RCP = 8.5 W/m² (high) [72-73].

We used Maxent's default parameters [26, 50] and a ten-fold cross-validation approach to further reduce bias with respect to locality data. This method divides presence data into ten equal partitions, with nine used to train the model, and the tenth used to test it. These partitions generate ten maps (one map per run), with each raster grid cell containing a value representing the probability of occurrence. These values were used to designate habitat suitability ranging from 0 (unsuitable habitat) to 1 (highly suitable habitat). We averaged the resulting maps for the current climate, and for the two GCMs under RCP = 4.5 and RCP = 8.5. This method resulted in the production of a "low" and "high" average prediction for *S. diana* species distribution in 2050, represented with habitat suitability maps. We measured the goodness of fit for the models using the area under the curve (AUC) of a receiver-operating characteristic (ROC) plot [74]. We used criteria of Swets [75] and considered AUC values higher than 0.7 representative of model predictions significantly better than random values of 0.5 or less [26, 27, 74]. Because AUC has been recognized as a somewhat questionable measure of accuracy, especially when used with background data instead of true absences [74, 77], we also calculated the TSS (true skill statistics), a threshold-dependent evaluation metric [76-77]. The relative importance of each variable's

contribution was assessed by sequential variable removal by Jackknife [26].

Results

Species distributional modeling resulted in “excellent” model fits for *Speyeria diana*, with a mean AUC = 0.91± 0.0028 SE, TSS = 0.87 ± 0.0032 SE for RCP = 4.5, and a mean AUC = 0.87± 0.0031 SE, TSS = 0.84 ± 0.0032 SE for RCP = 8.5 (Table 1). Annual precipitation explained the largest fraction of the distribution of *S. diana* under both RCPs (17.9%, RCP = 4.5; 19.4%, RCP = 8.5). Among the remaining bioclimatic variables, mean temperature of driest quarter had the next highest average percent contribution (10.3%, RCP = 4.5; 25.0%, RCP = 8.5), followed by minimum temperature of coldest month (20.1%, RCP = 4.5; 10.4%, RCP = 8.5), isothermality (7.3%, RCP = 4.5; 7.6%, RCP = 8.5), precipitation of wettest month (3.5%, RCP = 4.5; 3.9%, RCP = 8.5), precipitation of driest month (1.4%, RCP = 4.5; 5.4%, RCP = 8.5), precipitation of driest quarter (3.3%, RCP = 4.5; 2.4%, RCP = 8.5), Elev (1.5%, RCP = 4.5; 3.5%, RCP = 8.5), mean diurnal range (1.8%, RCP = 4.5; 2.8%, RCP = 8.5), and temperature annual range (1.6%, RCP = 4.5; 1.3%, RCP = 8.5) (Table 1).

Modelling with Maxent under the selected climate-change scenarios predicted that habitat suitability would decrease for *S. diana* by 2050 (two-tailed paired t-tests comparing current Maxent values with those of 2050; all P<0.01). The MIROC model resulted in more loss of suitable habitat than CCSM under both RCP scenarios (88.2% versus 92.4% of suitable habitat retained for RCP 4.5, and 90.2% versus 94.3% of suitable habitat retained for RCP 8.5 in CCSM and MIROC, respectively). Both climate models indicate that the loss of core distributional area is modest, with an average of 91.3% of present distributional areas retained. The most drastic reduction in habitat is apparent across the southern Appalachian Mountains (Fig. 2).

Discussion

Our ecological niche models predicted that the amount of suitable habitat for *Speyeria diana* will decline substantially by the year 2050 across its entire distribution. Both CCSM and MIROC climate models predicted severe habitat loss and fragmentation in the Southern Appalachian Mountains by 2050, with some range expansion predicted into higher latitudes in both eastern and western populations. High elevation habitat will be an important refuge for the species across the entire distribution, as the range of *S. diana* is already shifting to higher elevations at an estimated rate of 18m per decade [33]. Recent evidence further suggests that some *S. diana* populations may already be adapting to high elevations, as *S. diana* female forewings from high elevation populations were found to be narrower than low elevation populations, indicating that these females may be more mobile than those from low elevations with wider forewings [35].

Unlike populations in the eastern distribution, the wing shape of western populations of *S. diana* appears to be better adapted for lower dispersal, which is in alignment with findings that western populations of *S. diana* are both spatially and genetically isolated [35]. Our models predicted that the southern edge of highly suitable habitat in the west will recede by 2050; However, as was found in the Southern Appalachian Mountains, suitable habitat was predicted to expand in the higher elevations of the Ozark and Ouachita mountains of Arkansas. The genetic isolation of western populations may ultimately prevent them from adapting to higher elevations as successfully as populations in the eastern distribution of the species. If this is the case, lower elevation populations will be even more vulnerable to climate change than our models predict.

We would like to note that all ecological niche models should be used and interpreted with caution due to various sources of bias and error that result in inaccurate predictions [78]. Some have questioned the applicability of bioclimatic modeling at regional scales due to the

somewhat coarse resolution [79]. However, we are confident that the size of our study area, and our uniquely extensive dataset, provide sufficient data to forecast climate-driven range shifts in *S. diana* with accuracy. Both global circulation models (CCCM and MIROC) were very closely aligned in their outcomes, indicating strong agreement between them. Climate is well understood to play a primary role in shaping the distributions of species [80], and we are confident in our overall findings that suitable habitat for *S. diana* will decline and become increasingly fragmented by 2050.

These results highlight the importance of maintaining connectivity of suitable habitat for *S. diana*, especially in the eastern populations that appear most vulnerable to increased fragmentation and loss of suitable habitat. These populations in the eastern distribution of *S. diana* harbor important genetic diversity that may become lost through genetic drift if these populations become small and isolated. The Ozark and Ouachita Mountains of Arkansas and Missouri appear to be least vulnerable to loss of suitable habitat from climate change, and therefore will be important for the future conservation of *S. diana* after 2050. Due to the geographic and genetic isolation of the western populations, conservation of suitable habitat in the west is equally as important as in the east. Our climate models show that the 800 km disjunction across the center of the range of *S. diana* is not due to complete absence of suitable habitat, but more probably a result of the extensive habitat fragmentation regionally across the Ohio River Valley from agricultural land use change, and other human related factors that were not included in our models. We conclude that maintaining well-connected low and high elevation habitats across the entire distribution of *S. diana*, both now and into the future, will be necessary for this species, even under conservative forecasts of climate change.

Table 1. Elevation plus the 19 bioclimate variables from the WorldClim dataset (Hijmans et al., 2005) collapsed into groups of highly correlated variables (Pearson's correlation coefficient, $r \geq \pm 0.70$), and their corresponding contribution to the Maxent model. The ten variables kept in the final model are bold and highlighted in grey. CCCM and MIROC global circulation models are shown under representative concentration pathways (RCPs) 4.5 (low) and 8.5 (high), as predicted by the IPCC 5th report on climate.

Bioclimate variables	Abbreviation	% Contribution					
		CCCM-45	MIROC-45	AVG	CCCM-85	MIROC-85	AVG
Annual Mean Temperature	Bio 1	4.4	0.7	2.5	0.5	1.4	0.96
Max Temperature of Warmest Month	Bio 5	0.6	1.7	1.2	1.4	0.8	1.1
Min Temperature of Coldest Month	Bio 6	3.9	36.3	20.1	2.6	3.3	10.4
Mean Temperature of Wettest Quarter	Bio 8	14.1	10.2	12.2	4.0	16.8	2.6
Mean Temperature of Driest Quarter	Bio 9	15.5	5.1	10.3	30.2	19.8	25.0
Mean Temperature of Warmest Quarter	Bio 10	0.5	0.8	0.7	0.1	0.3	0.2
Mean Temperature of Coldest Quarter	Bio 11	0.8	12.5	11.9	3.3	1.5	2.4
Precipitation of Wettest Month	Bio 13	3.7	0.2	3.5	2.0	5.8	3.9
Precipitation Seasonality	Bio 15	6.0	3.7	4.9	8.7	2.7	5.6
Precipitation of Wettest Quarter	Bio 16	0.8	0.6	0.7	0.2	0.9	0.6
Precipitation of Warmest Quarter	Bio 18	1.1	0.3	1.0	1.9	1.0	1.5
Precipitation of Driest Month	Bio 14	0.9	1.6	1.4	2.7	8.0	5.4
Precipitation of Driest Quarter	Bio 17	4.2	2.3	3.3	2.2	2.6	2.4
Precipitation of Coldest Quarter	Bio 19	0.1	0.2	0.2	0.2	1.7	0.9
Elevation	Elev	2.0	1.0	1.5	4.9	2.0	3.5
Isothermality (BIO2/BIO7) (* 100)	Bio 3	11.0	3.5	7.3	8.5	6.6	7.6
Temperature Seasonality (standard deviation *100)	Bio 4	6.4	1.0	3.7	0.0	4.2	2.1
Mean Diurnal Range (Mean of monthly (max temp - min temp))	Bio 2	0.6	3.0	1.8	2.0	3.6	2.8
Temperature Annual Range (BIO5-BIO6)	Bio 7	1.2	1.9	1.6	1.5	1.0	1.3
Annual Precipitation	Bio 12	22.3	13.4	17.9	22.9	15.9	19.4
AUC		0.86	0.96	0.91	0.87	0.86	0.87

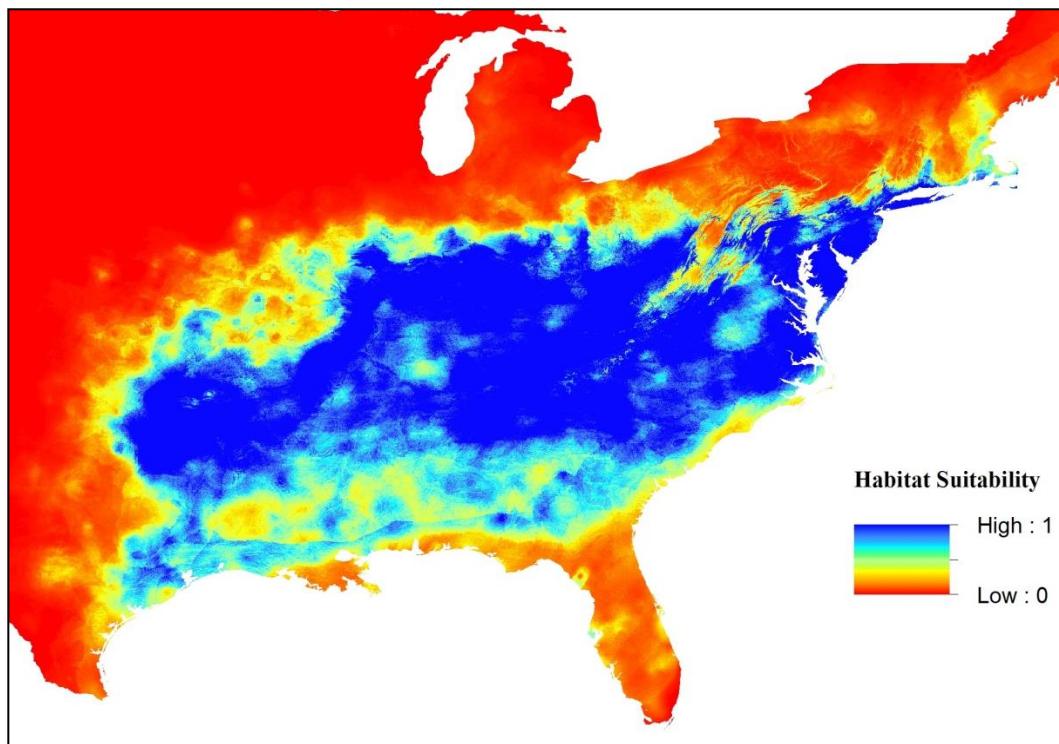


Figure 1 The present-day geographic distribution of *Speyeria diana*, with indices of habitat suitability as predicted by maximum entropy modelling (Maxent) under current climatic conditions (1950-2010).

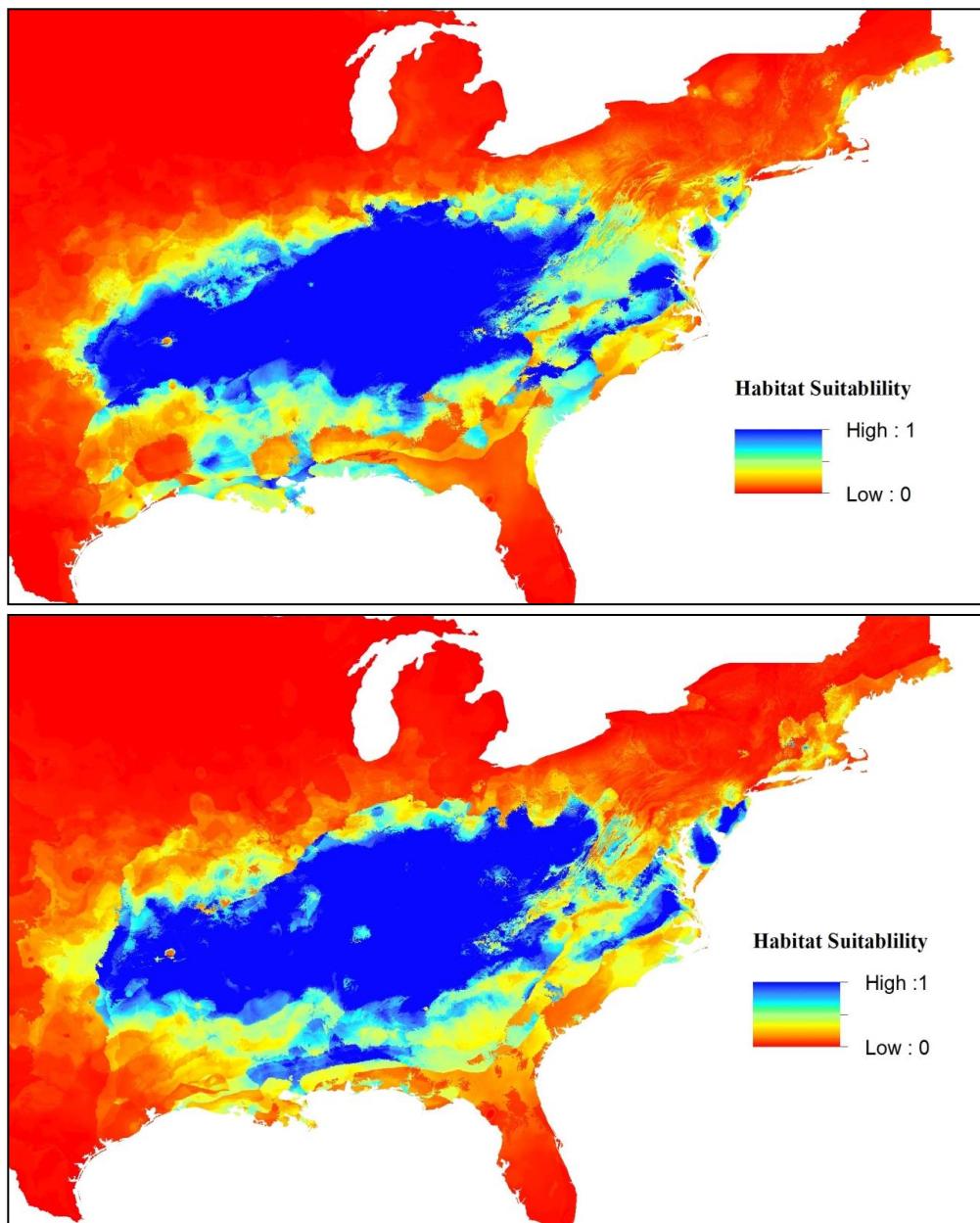


Figure 2 (a) Habitat suitability indices for the projected future distribution of *Speyeria diana* under the CCMA and MIROC RCP 4.5 climate change scenarios; (b) Habitat suitability indices for the projected future distribution of *Speyeria diana* under the CCMA and MIROC RCP 8.5 climate change scenarios.

1. Parmesan, C. Climate and species' ranges. *Nature* **1996**, *382*, 765-766.
2. Parmesan, C.; Yohe, G. A globally coherent fingerprint of climate change impacts across natural systems. *Nature* **2003**, *421*, 37-42.
3. Thomas, C.D.; Franco, A.M.; Hill, J.K. Range retractions and extinction in the face of climate warming. *Trends in Ecology and Evolution* **2004**, *21*, 415-416.
4. Crozier, L.; Dwyer, G. Combining population-dynamic and ecophysiological models to predict climate-induced insect range shifts. *The American Naturalist* **2006**, *167*, 853-866.
5. Walther, G.R.; Post, E.; Convey, P.; Menzel, A.; Parmesan, C.; Beebee, T.; Fromentin, J.M.; Hoegh-Guldberg, O.; Bairlein, F. Ecological Responses to recent climate change. *Nature* **2002**, *416*, 389-395.
6. Root, T.L.; Price, J.T.; Hall, K.R.; Schneider, S.H.; Rosenzweig, C.; Pounds, J.A. Fingerprints of global warming on wild animals and plants. *Nature* **2003**, *421*, 57-60.
7. Parmesan, C. Ecological and evolutionary responses to recent climate change. *Annual Review of Ecology, Evolution, and Systematics* **2006**, *37*, 637-69.
8. Walther, G.R.; Beibner, S.; Conradin, A. Trends in the upward shift of alpine plants. *Journal of Vegetation Science* **2005**, *16*, 541-548.
9. Cheung, W.W.L.; Lam, W.Y.V.; Sarmiento, J.L.; Kearney, K.; Watson, R.; Pauly, D. (2009) Projecting global marine biodiversity impacts under climate change scenarios. *Fish and Fisheries* **2009**, *10*, 235-251.
10. Perry, A.L.; Low, P.J.; Ellis, J.R.; Reynolds, J.D. Climate change and distribution shifts in marine fishes. *Science* **2005**, *308*, 1912-1915.
11. Epstein, P.R.; Diaz, H.; Elias, F.S.; Grabherr, G.; Graham, N.E.; Martens, W.J.M.; Mosley-Thompson, E.; Susskind, E.J. Biological and physical signs of climate change: Focus on mosquito-borne disease. *Bulletin of the American Meteorological Society* **1998**, *78*, 409- 417.
12. Thomas, C.D.; Lennon, J.J. Birds extend their ranges northwards. *Nature* **1999**, *399*, 213.
13. Hitch, A.T.; Leberg, P.L. Breeding distributions of North American bird species moving north as a result of climate change. *Conservation Biology* **2007**, *21*, 534-539.
14. Parmesan, C.; Ryrholm, N.; Stefanescu, C.; Hill, J.; Thomas, C.; Descimon, H.; Huntley, B.; Kaila, L.; Kullberg, J.; Tammaru, T.; Tennent, T.J.; Warren, M. Poleward shifts in geographical ranges of butterfly species associated with regional warming. *Nature* **1999**, *399*, 579-583.
15. Wilson, R.J.; Gutiérrez, D.; Gutiérrez, J.; Martínez, D.; Aguado, R.; Monserrat, V.J. Changes to the elevational limits and extent of species ranges associated with climate change. *Ecology Letters* **2005**, *8*, 1138-1146.

16. Wilson, R.J., Gutiérrez, D.; Gutiérrez, J.; Monserrat, V. An elevational shift in butterfly species richness and composition accompanying recent climate change. *Global Change Biology* **2007**, *13*, 1873– 1887.
17. Asher, J.; Fox, R.; Warren, M.S. British butterfly distributions and the 2010 target. *Journal of Insect Conservation* **2011**, *15*, 291–299.
18. Wilson, R.J.; Maclean, I.M.D. Recent evidence for the climate threat to Lepidoptera and other insects. *Journal of Insect Conservation* **2011**, *15*, 259–268.
19. Garcia, K.; Lasco, R.; Ines, A.; Lyon, B.; Pulhin, F. Predicting geographic distribution and habitat suitability due to climate change of selected threatened forest tree species in the Philippines. *Applied Geography* **2013**, *44*, 12–22.
20. Pearson, R.G.; Dawson, T.P. Predicting the impacts of climate change on the distribution of species: are bioclimate envelope models useful? *Global Ecology and Biogeography* **2003**, *12*, 361–371.
21. Peterson, A.T. Projected climate change effects on Rocky Mountain and Great Plain birds: generalities on biodiversity consequences. *Global Change Biology* **2003**, *9*, 647–55.
22. Elith, J.; Leathwick, J.R. Species distribution models: Ecological explanation and prediction across space and time. *Annual Review of Ecology, Evolution, and Systematics* **2009**, *40*, 677–697.
23. Fordham, D.A.; Resit, A.H.; Araújo, M.B.; Elith, J.; Keith, D.A.; Pearson, R.; Auld, T.D.; Mellin, C.; Morgan, J.W.; Regan, T.J.; Tozer, M.; Watts, M.J.; White, M.; Wintle, B.A.; Yates, C.; Brook, B.W. Plant extinction risk under climate change: Are forecast range shifts alone a good indicator of species vulnerability to global warming? *Global Change Biology* **2012**, *18*, 1357–1371.
24. Thuiller, W.; Lavorel, S.; Araújo, M.B.; Sykes, M.T.; Prentice, I.C. Climate change threats to plant diversity in Europe. *Proceedings of the National Academy of Sciences* **2005**, *102*, 8245–8250.
25. Willis, K.J.; Araújo, M.B.; Bennett, K.D.; Figueroa-Range, B.; Froyd, C.A.; Myers, N. How can a knowledge of the past help to conserve the future? Biodiversity conservation and the relevance of long-term ecological studies. *Philosophical Transactions of the Royal Society of Britain* **2007**, *362*, 175–186.
26. Phillips, S.J.; Anderson, R.; Schapire, R.E. Maximum entropy modeling of species geographic distributions. *Ecological Modelling* **2006**, *190*, 231–259.
27. Elith, J.; Phillips, S.J.; Hastie, T.; Dudík, M.; Chee, Y.E.; Yates, C.J. A statistical explanation of MaxEnt for ecologists. *Diversity and Distributions* **2011**, *17*, 43–57.
28. Weber, T.C. Maximum entropy modeling of mature hardwood forest distribution in four US states. *Forest Ecology and Management* **2011**, *261*, 779–788.

29. Araújo, M.B.; Luoto, M. The importance of biotic interactions for modeling species distributions under climate change. *Global Ecology and Biogeography* **2007**, *16*, 743-753.
30. Araújo, M.B.; Pearson, R.G.; Thuiller, W.; Erhard, M. Validation of species-climate impact models under climate change. *Global Change Biology* **2005**, *11*, 1504-1513.
31. Araújo, M.B.; Whittaker, R.J.; Ladle, R.J.; Erhard, M. Reducing uncertainty in projections of extinction risk from climate change. *Global Ecology and Biogeography* **2005**, *14*, 529-538.
32. Green, R.E.; Collingham, Y.C.; Willis, S.G., Gregory, R.D.; Smith, K.W.; Huntley, B. Performance of climate envelope models in predicting recent changes in bird population size from observed climatic change. *Biology Letters* **2008**, *4*, 599-602.
33. Wells, C.N.; Tonkyn, D.W. Range collapse in the Diana fritillary, *Speyeria diana* (Nymphalidae). *Insect Conservation and Diversity* **2014**, *7*, 365-380.
34. Wells, C.N.; Marko, P.B.; Tonkyn, D.W. The phylogeographic history of the threatened Diana fritillary, *Speyeria diana* (Lepidoptera: Nymphalidae): with implications for conservation. *Conservation Genetics* **2015**, *16*, 703-716.
35. Wells, C.N., Munn, A.; Woodworth, C. Geomorphic Morphometric Differences between Populations of *Speyeria diana* (Lepidoptera: Nymphalidae). *Florida Entomologist* **2018**, *101*(2), 195-202.
36. Opler, P.A.; Krizek, G. *Butterflies East of the Great Plains*. Johns Hopkins University Press: Baltimore, MD., USA, 1984; pp. 294; ISBN: 0801829380
37. Allen, T.J. *The Butterflies of West Virginia and their Caterpillars*. University of Pittsburgh Press, Pittsburgh, PA., USA, 1997. pp. 388; ISBN: 0822939738
38. Cech, R.; Tudor, G. *Butterflies of the East Coast*. Princeton University Press, Princeton, NJ., USA, 2005, pp. 345; ISBN: 069109055
39. Baltosser, W. Flitting with disaster: Humans and habitat are keys to our state butterfly's future. *Arkansas Wildlife*, **2007**, *38*, 6-11.
40. Ross, G.N. What's for dinner? A new look at the role of phytochemicals in butterfly diets. *News of the Lepidopterists' Society* **2003**, *45*, 83-89.
41. Ross, G.N. Diana's Mountain Retreat. *Natural History* **2008**, *72*, 24-28.
42. Adams, J.K.; Finkelstein, I. Late season observations on female Diana fritillary (*Speyeria diana*) aggregating behavior. *News of the Lepidopterists' Society*, **2006**, *48*, 106-107.
43. Araújo, M.B.; Peterson, A.T. Uses and misuses of bioclimatic envelope modeling. *Ecology*, **2012**, *93*, 1527-1539.

44. Loiselle, B.A.; Jørgensen, P.M.; Consiglio, T.; Jiménez, I.; Blake, J.G.; Lohmann, L.G.; Montiel, O.M. Predicting species distributions from herbarium collections: does climate bias in collection sampling influence model outcomes? *Journal of Biogeography* **2007**, *35*, 105–116.
45. Renner, I. W. & Warton, D. I. Equivalence of MAXENT and Poisson Point Process Models for Species Distribution Modeling in Ecology. *Biometrics* **2013**, *69*, 274–281.
46. Gomes, Vitor HF, et al. Species Distribution Modelling: Contrasting presence-only models with plot abundance data. *Scientific reports* **2018**, *8*, 1, 1003. DOI:10.1038/s41598-017-18927-1
47. Elith, J.; Graham, C.H. NCEAS Species Distribution Modelling Group, Novel methods improve prediction of species' distributions from occurrence data. *Ecography* **2006**, *29*, 129–151.
48. Elith, J.; Kearney, M.; Phillips, S. The art of modeling range-shifting species. *Methods in Ecology and Evolution* **2010**, *1*, 330–342.
49. Hijmans, R.J.; Cameron, S.E.; Parra, J.L.; Jones, P.G.; Jarvis, A. Very high resolution interpolated climate surfaces for global land areas. *International Journal of Climatology* **2005**, *25*, 1965–1978.
50. Phillips, S.J.; Dudík, M. Modeling of species distributions with Maxent: new extensions and a comprehensive evaluation. *Ecography* **2008**, *31*, 161–175.
51. Phillips, S.J.; Dudík, M.; Elith, J.; Graham, C.H.; Lehmann, A.; Leathwick, J.; Ferrier, S. Sample selection bias and presence-only distribution models: implications for background and pseudo-absence data. *Ecological applications*, **2009**, *19*, 181–197.
52. Beaumont, L.; Hughes, L.; Poulsen, M. Predicting species distributions: use of climatic parameters in BIOCLIM and its impact on predictions of species' current and future distributions. *Ecological Modelling* **2005**, *186*, 250–269.
53. Dennis, R.L.H. *Butterflies and Climate Change*. Manchester University Press, Manchester, UK., 1993, pp. ISBN:
54. Hill, J.K.; Thomas, C.D.; Huntley, B. Modelling present and potential future ranges of European butterflies using climate response surfaces. In: *Butterflies: Ecology and Evolution Taking Flight* (editors Boggs CL, Watt WB Ehrlich PR). University of Chicago Press, Chicago, 2003, pp. 149–167.
55. Peterson, A.T.; Martínez-Meyer, E.; González-Salazar, C.; Hall, P.W. Modeled climate change effects on distributions of Canadian butterfly species. *Canadian Journal of Zoology* **2004**, *82*, 851–858.
56. Mitikka, V.; Heikkinen, R.K.; Luoto, M.; Araújo, M.B.; Saarinen, K.; Pöyry, J.; Fronzek, S. Predicting range expansion of the map butterfly in Northern Europe using bioclimatic models. *Biodiversity and Conservation* **2008**, *17*, 623–641.

57. Filz, K.J.; Schmitt, T.; Engler, J.O. How fine is fine-scale? Questioning the use of fine-scale bioclimatic data in species distribution models used for forecasting abundance patterns in butterflies. *European Journal of Entomology* **2013**, *110*, 311–317.
58. Zinetti, F.; Dapporto, L.; Vovlas, A.; Chelazzi, G.; Bonelli, S.; Balletto, E.; Ciofi, C. When the rule becomes the exception: No evidence of gene flow between two *Zerynthia* cryptic butterflies suggests the emergence of a new model group. *PLOS ONE* **2013**, *8*, e65746–e65746.
59. Hill, J.K.; Thomas, C.D.; Fox, R.; Telfer, M.G.; Willis, S.G.; Asher, J.; Huntley, B. Responses of butterflies to twentieth century climate warming: implications for future ranges. *Proceedings of the Royal Society of London, Series B-Biological Sciences* **2002**, *269*, 2163–2171.
60. Karlsson, B.; Van Dyck, H. Does habitat fragmentation affect temperature-related life-history traits? A laboratory test with a woodland butterfly. *Proceedings of the Royal Society of London B: Biological Sciences*, **2005**, *272*, 1257–1263.
61. Gibbs, M., Van Dyck, H., & Karlsson, B. Reproductive plasticity, ovarian dynamics and maternal effects in response to temperature and flight in *Pararge aegeria*. *Journal of Insect Physiology*, **2010**, *56*: 1275–1283.
62. Morecroft, M.D.; Bealey, C.E.; Howells, O.; Rennie, S.; Woiwod, I.P. Effects of drought on contrasting insect and plant species in the UK in the mid-1990s. *Global Ecology and Biogeography* **2002**, *11*, 7–22.
63. Kadmon, R.; Farbr, O.; Danin, A. Effect of roadside bias on the accuracy of predictive maps produced by bioclimatic models. *Ecological Applications* **2004**, *14*, 401–413.
64. Gent, P.R.; Danabasoglu, G.; Donner, L.J.; Holland, M.M.; Hunke, E.C.; Jayne, S.R.; Lawrence, D.M.; Neale, R.B.; Rasch, P.J.; Vertenstein, M.; Worley, P.H.; Yang, Z.L.; Zhang, M. The community climate system model, Version 4. *Journal of Climate* **2011**, *24*, 4973–4991.
65. Hasumi, H.; Emori, S. K-1 coupled GCM (MIROC) description, K-1 Tech. Rep. 1, Center for Climate Systems Research, Univ. of Tokyo, Tokyo, 2004, 34pp.
66. Nozawa, T.; Nagashima, T.; Shiogama, H.; Crooks, S.A. Detecting natural influence on surface air temperature change in the early twentieth century. *Geophysical Research Letters*, **2005**, *32*, L20719.
67. Habel, J.C.; Rödder, D.; Scalercio, S.; Meyer, M.; Schmitt, T. Strong genetic cohesiveness between Italy and North Africa in four butterfly species. *Biological Journal of the Linnean Society* **2010**, *99*, 818–830.
68. Habel, J.C.; Husemann, M.; Schmitt, T.; Dapporto, L.; Vandewoestijne, S. A forest butterfly in Sahara desert oases: Isolation does not matter. *Journal of Heredity* **2013**, *104*, 234–247.
69. Moss, R.H.; Edmonds, J.A.; Hibbard, K.A.; Manning, M.R.; Rose, S.K.; van Vuuren, D.P.; Carter, T.R.; Emori, S.; Kainuma, M.; Kram, T.; Meehl, G.A.; Mitchell, J.F.B.; Nakicenovic, N.; Riahi, K.; Smith, S.J.; Stouffer, R.J.; Thomson, A.M.; Weyant, J.P.; Wilbanks, T.J. The next

- generation of scenarios for climate change research and assessment. *Nature* **2010**, *463*, 747–756.
70. van Vuuren, D.P.; Edmonds, J.; Kainuma, M.; Riahi, K.; Thomson, A.; Hibbard, K.; Hurtt, G.C.; Kram, T.; Krey, V.; Lamarque, J.F.; Masui, T.; Meinshausen, M.; Nakicenovic, N.; Smith, S.; Rose, S. The representative concentration pathways: An overview. *Climatic Change* **2011**, *109*, 5–31.
71. Moss, R.H.; Babiker, M.; Brinkman, S.; Calvo, E.; Carter, T.; Edmonds, J.; Elgizouli, I.; Emori, S.; Erda, L.; Hibbard, K.; Jones, R.; Kainuma, M.; Kelleher, J.; Lamarque, J.F.; Manning, M.; Matthews, B.; Meehl, J.; Meyer, L.; Mitchell, J.; Nakicenovic, N.; O'Neill, B.; Pichs, R.; Riahi, K.; Rose, S.; Runci, P.; Stouffer, R.; van Vuuren, D.; Weyant, J.; Wilbanks, T.; van Ypersele, J.P.; Zurek, M. Towards New Scenarios for Analysis of Emissions, Climate Change, Impacts, and Response Strategies. Intergovernmental Panel on Climate Change, Geneva, 2008, 132 pp.
72. Riahi, K.; Rao, S.; Krey, V.; Cho, C.; Chirkov, V.; Fischer, G.; Kindermann, G.; Nakicenovic, N.; Rafaj, P. RCP 8.5—A scenario of comparatively high greenhouse gas emissions. *Climatic Change* **2011**, *109*, 33–57.
73. Thomson, A.; Calvin, K.; Smith, S.; Kyle, P.; Volke, A.; Patel, P.; Delgado-Arias, S.; Bond-Lamberty, B.; Wise, M.; Clarke, L.; Edmonds, J. RCP4.5: a pathway for stabilization of radiative forcing by 2100. *Climatic Change* **2011**, *109*, 77–94.
74. Jiménez-Valverde, A. Insights into the area under the receiver operating characteristic curve (AUC) as a discrimination measure in species distribution modelling. *Global Ecology and Biogeography* **2012**, *21*, 498–507.
75. Swets, J.A. Measuring the accuracy of diagnostic systems. *Science* **1988**, *240*, 1285–1293.
76. Fielding, A.H.; Bell, J.F. A review of methods for the assessment of prediction errors in conservation presence/absence models. *Environmental Conservation*, **1997**, *24*, 38–49.
77. Allouche, O.; Tsoar, A.; Kadmon, R. Assessing the accuracy of species distribution models: prevalence, kappa and the true skill statistic (TSS). *Journal of Applied Ecology* **2006**, *43*, 1223–1232.
78. Beaumont, L.J.; Hughes, L.; Pitman, A.J. Why is the choice of future climate scenarios for species distribution modelling important? *Ecology Letters* **2008**, *11*, 1135–1146.
79. Chen, I. C.; Hill, J.K.; Ohlemuller, R.; Roy, D.B.; Thomas, C. D. Rapid range shifts of species associated with high levels of climate warming. *Science*, **2011**, *333*, 1024–1026
80. Fourcade, Y.; Besnard, A.G.; Secondi, J. Paintings predict the distribution of species, or the challenge of selecting environmental predictors and evaluation statistics. *Global Ecology and Biogeography* **2018**, *27*, 245–256.