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2 **Dental adhesion enhancement on zirconia inspired** 3 **by mussel's priming strategy using catechol**

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10

11 **Abstract:** Zirconia has recently become one of the most popular dental materials
12 in prosthodontics being used in crowns, bridges, and to implants. However, weak
13 bonding strength of dental adhesives and resins to zirconia surface has been a
14 grand challenge in dentistry, thus finding a better adhesion to zirconia is urgently
15 required. Marine sessile organisms such as mussels use a unique priming strategy
16 to produce a strong bonding to wet mineral surfaces; one of the distinctive
17 chemical features in the mussel's adhesive primer proteins is high catechol
18 contents among others. In this study, we pursued a bioinspired adhesion strategy,
19 using a synthetic catechol primer applied to dental zirconia surfaces to study the
20 effect of catecholic priming to shear bonding strength. Catechol priming provided
21 a statistically significant enhancement ($P < 0.05$) in shear bonding strength
22 compared to the bonding strength without priming, and relatively stronger
23 bonding than commercially available zirconia priming techniques. This new
24 bioinspired dental priming approach can be an excellent addition to the
25 practitioner's toolkit to improve dental bonding to zirconia.

26 **Keywords:** Zirconia; primer; priming; bonding; catechol; dental; prosthodontics

27

28 **1. Introduction**

29 Ceramics have been widely used for dental and biomaterials, e.g., for
30 prostheses. Zirconia has recently become one of the most popular ceramic dental
31 and biomaterials with its superior mechanical properties: highly tough and strong,
32 wear resistant, shock resistant, while both chemically and dimensionally stable [1].
33 Its superior mechanical properties are due to a martensitic type phase
34 transformation that prevents crack propagation [2]. However, owing to its high
35 chemical resistance, traditional dental ceramic bonding techniques (e.g., chemical
36 ceramic etching) are not compatible with zirconia [2]. Traditional mechanical
37 grinding or sand blasting methods on dental ceramics can be an alternative, but it
38 may cause the phase transformation and hydrothermal fatigue degradation [3].

39 Priming (or adhesive priming) is a surface treatment that can promote
40 adhesion of coatings or adhesives to the substrate of interest. Silane-based primers
41 (known as silane coupling agents) are commonly used to prime dental surfaces,
42 including ceramics. Despite many attempts to improve the bonding strength to
43 zirconia using the silane-based coupling agents, no significant improvement has
44 been reported yet [3].

45 In the past decades, the National Institute of Dental and Craniofacial Research
46 (NIDCR) has supported interdisciplinary studies such as marine mussel adhesion
47 as the oral environment has much in common with the intertidal environment;
48 both oral and marine adhesions are subjected to opportunistic microbes, cyclic
49 stress, constant fluid flow with variations in salinity, temperature and pH [4].
50 Figure 1 shows marine sessile organisms adhere to rocks at the seashore. The
51 NIDCR's long-term investment to the bioadhesion [4,5] has recently begun to show
52 potential for practical dental [6] and biomedical [7,8] applications. Several dental
53 applications of catechol have been reported using catechol-containing polymers
54 such as tannin as a dental sealant [9] and catecholic methacrylamide for a dental
55 adhesive [10] via catechol-ferric iron complexation. Despite the inherent aesthetic
56 limitations of Fe-catechol complexation, (complexes are highly colored and darken
57 over time) the previous work exposed the potential of catechol chemistry for dental
58 materials applications. In contrast to these previous approaches which mimic the
59 mussels use of catecholic complexation and crosslinking chemistry for
60 enhancement of *cohesive* strength within the byssus, we, in this study instead aimed
61 to mimic the mussel's means of priming surfaces with catecholic bondings that
62 increase the *adhesive* strength of mussel plaque to mineral surfaces [11].
63



64

65 **Figure 1.** Mussels and other marine sessile organisms such as mussels anchored to mineral surfaces
66 at seashore in Santa Barbara, California, U. S.

67 One of the useful chemical features of mussel's adhesive mechanism is the use
68 of catechol- and phospho-rich proteins as surface primers [11-13]. Similarly, in
69 dentistry phosphorous-based primers such as 10-methacryloyloxydecyl
70 dihydrogen phosphate (MDP) are commonly used based in part on the strength of
71 phosphate binding to Calcium ions in hydroxyapatite (the main mineral in human

72 teeth — higher than 90 wt. % in enamel and 70 wt. % in dentin — and in human
 73 bone — up to 70 wt. %). By bridging the surface minerals to a polymerizable
 74 methacrylate, MDP helps to achieve bonding performance of methacrylate-based
 75 dental resins.

76 MDP has also been shown to improve adhesion of dental resins to zirconia
 77 surface by ionic phosphate-Zr binding [3,14], yet further increases to the strength
 78 of such resins bound to zirconia is urgently required and many researchers are
 79 looking for alternative chemistry that can fulfill this demand. Very recently, our
 80 group has reported adhesion enhancement of dental adhesion to silica, steel and
 81 tooth enamel using the catechol-containing primers [6,15]. In contrast to the large
 82 body of literature regarding phosphate-primers, and despite over 10,000
 83 peer-reviewed papers published within the past decade on catechol-mediated
 84 biological adhesion [11], the use of catechol chemistry for strength enhancement
 85 and surface priming in dental settings remains largely unexplored.

86 In this study, we investigated if one of the most pressing issues in
 87 prosthodontics, poor adhesion to zirconia, could be overcome with catecholic
 88 bioinspired surface priming. Using a previously reported bifunctional
 89 catechol-methacrylate to prime zirconia, we were able to demonstrate significant
 90 improvement in the knife shear bonding strength compared to untreated surfaces.
 91 Moreover, the bonding performance is superior to commercial dental primers
 92 reliant on acidic phosphate and carboxylate functionality. Our results highlight the
 93 potential of catechol-mediated surface priming to solve bonding issues with
 94 zirconia for dental and biomedical applications. In addition, we were able to
 95 correlate bonding performance with surface coverage as measured by SEM and
 96 offer an improved synthetic method to access the catechol methacrylate primer in
 97 higher purity compared to the previously reported method [6,16,17].

98 2. Materials and Methods

99 All chemicals were purchased from Sigma Aldrich. All dental resins and
 100 primers were purchased as shown in table 1.

101 **Table 1.** Commercial zirconia dental primers.

Primers	Composition ¹	Manufacturer
Zirconia liner (ZL)	4-META	SunMedical Co. Ltd (Japan)
Alloy primer (AP)	10-MDP, VBATDT	Kuraray Dental, Inc. (Japan)
Universal primer (UP)	MAC-10, MTU-6	Tokuyama Dental, Corp. (Japan)

102 ¹ 4-META: 4-methacryloyloxy ethyl trimellitate anhydride; 10-MDP; 10-Methacryloyloxydecyl dihydrogen
 103 phosphate, VBATDT: 6-(4-Vinylbenzyl-n-propyl)amino-1,3,5-triazine-2,4-dithiol; MAC-10:
 104 11-Methacryloylundecane-1,1- dicarboxylic acid; MTU-6:6-methacryloyloxyhexyl-2-thiouracil-5-carboxylate

105
 106

107 2.1. Synthesis of Catechol Methacrylate Primer

108 Triethyl silylane (TES)-protected (or silylated) catechol-methacrylate was
109 synthesized from eugenol by the previously reported method [16] and generously
110 provided to us by Osaka Organic Chemical Industry LTD (Japan). By modifying
111 the previously reported conditions for TES-deprotection, the Catecholic
112 Methacrylate could be obtained more economically, with a higher degree of purity
113 according to the following procedure. 411 mg of TES-protected catechol
114 methacrylate (0.855 mmol, 1 equiv.) and 229 mg of benzoic acid (1.88 mmol, 2.2
115 equiv., 1.1 equiv. per TES group) were dissolved in ca. 3-5 ml of Tetrahydrofuran
116 (THF), (Note: Distilled/anhydrous THF was not used), and stirred at ambient
117 temperature, whereupon a 1 M solution of Tetrabutylammonium fluoride (TBAF)
118 in THF (1.88 ml, 1.88 mmol, 2.2 equiv.) was added dropwise to the stirred mixture.
119 The mixture was stirred at ambient temperature until Thin-layer chromatography
120 (TLC) indicates complete conversion (30-120 min). Once judged complete, the stir
121 bar was removed, and THF was removed by rotary evaporation. The crude residue
122 was then suspended in 150 ml of ether (Et₂O), and washed twice with DI water,
123 once with brine, dried over sodium sulfate. Once dry, the solution was filtered and
124 evaporated under reduced pressure, whereupon the residue was dissolved in a
125 minimum amount of DCM, loaded on top of a silica column, and purified further
126 by flash chromatography. (Note: Catechols adhere strongly to silica, and are
127 subject to decomposition during chromatography, thus chromatography was
128 performed quickly, and with a minimum amount of silica gel. Oxidative
129 decomposition during chromatography could be mitigated by addition of a small
130 amount (0.1 % v/v) of AcOH to the eluent, although this required prolonged and
131 undesirable amounts of time on high-vacuum to remove traces of acid. The use of
132 silica gel impregnated with ascorbic acid was also found to reduce decomposition)
133 [18]. After eluting sequentially with 0%, 25%, then 60% EtOAc/Hexanes, fractions
134 containing desired material were then pooled in a round bottom flask, and a very
135 small (ca. 0.1-0.4 mg) crystal of BHT as inhibitor was added, then the solution
136 evaporated. A small quantity of DCM was used to transfer the residue to a small,
137 tared vial, and residual volatiles were removed by repeated coevaporation of the
138 residue with pentanes, followed by high vacuum, to afford 147 mg (68% of
139 theoretical) of the title compound as a clear viscous oil. The material was stored
140 in the freezer in glass vials tightly wrapped with parafilm, and protected from
141 light.

142 The Catechol-Methacrylate primer solution was prepared as follows. First,
143 Methanol was degassed by sparging with Argon for 15-30 minutes. The primer
144 was removed by gently scraping some of the compound with a spatula, into a
145 tared vial, which was then weighed, fitted with a rubber septa, and purged with
146 argon. Sufficient degassed MeOH was then added via syringe to make a 1 mg/ml
147 solution, the septa was then replaced with a screw cap, and the vial was vigorously
148 vortexed for 10-20 minutes until no further primer could be observed adhering to
149 the sides of the vial. Solutions were sealed tightly, protected from light, and stored
150 in the freezer.

151

152 *2.2. Zirconia specimen*

153 A zirconia block for dental crown was cut using a water cooled diamond saw
154 (Buehler Isomet Low Speed saw, model #11-1180, Buehler Ltd., Lake Bluff, IL) with
155 a 0.15 mm thick diamond blade to 2 mm thickness and 5 mm x 5 mm width. All
156 specimens were polished with sandpaper #1200 in order to have the same degree
157 of roughness.

158

159 *2.3. Surface treatment*

160 We divided specimens into 5 groups according to the types of used primers.
161 Each group is ZL using Zirconia Liner, AP using Alloy Primer, UP using Universal
162 Primer, CP using fresh Catechol primer (1 mg/mL methanol solution) [15], and
163 Control (non-treated specimen). The primers were applied according to the
164 manufacturer's instructions.

165

166 *2.4. Surface morphology*

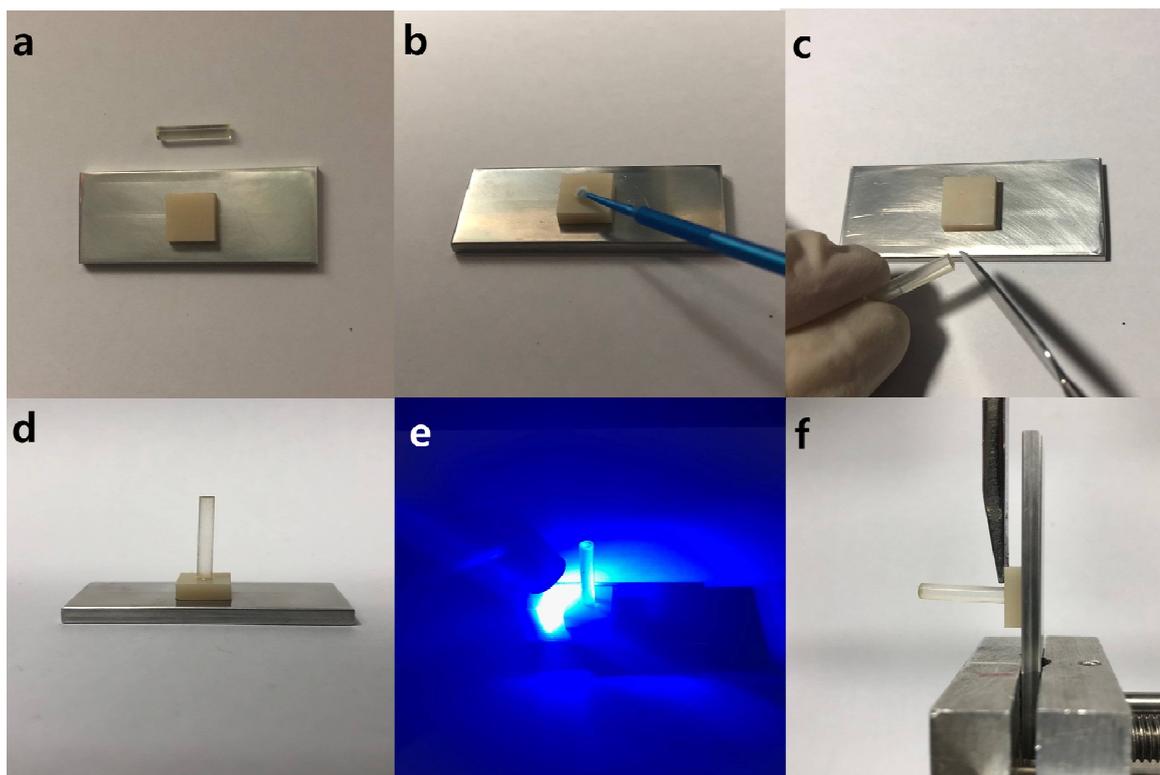
167 SEM analysis was conducted for the primed (surface-coated with primers) and
168 the non-primed zirconia surfaces. To prepare the primed samples, the commercial
169 primers were applied on the surfaces as directed in the manuals provided by the
170 manufacturers. Catechol primer solution was applied according to the previous
171 report [15]. The surface images were collected using field emission scanning
172 electron microscope (FESEM, Hitachi S-4700, Hitachi Co., Tokyo, Japan) at two
173 different magnifications (x250, x1000).

174

175 *2.5. Knife-edge shear bond strength*

176 First, a cured dental resin rod 2.5 mm in diameter shown in Figure 2 was
177 prepared using a common dental monomer mixture (49.5 wt.% of Bis-GMA, 49.5
178 wt. % of TEGDMA, and 1.0 wt. % of DMAEMA and CQ mixture at 1:2 molar ratio).
179 The bottom of the rod was polished and leveled flat with #800 sandpaper. Dental
180 resin cement (Bisco, IL, USA) was then applied onto the bottom surface of the rod
181 (Figure 2 (c)) as to simulate the bottom of the resin. The rod was then placed on the
182 primer-treated zirconia surface at orthogonal to the surface and lightly pushed
183 (Figure 2 (d)). Subsequently, blue light (Demi™ Ultra Dental Curing Lights, Kavo
184 Kerr, CA, USA) was applied all around surface for the complete cure for 20-40
185 seconds (Figure 2 (e)). All specimens were stored at room temperature for 1 day
186 prior to strength testing. Knife-edge shear bond test was performed at crosshead
187 down speed of 1 mm/min (Figure 2 (f)) by a universal testing machine (UTM 4465,
188 Instron, MA, USA). Each test repeated 10 times (n = 10); average and standard
189 deviation were calculated.

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Figure 2. Images taken during sample preparation : (a) A zirconia sample adhered to a stainless-steel plate; (b) Apply primer solutions on the zirconia surface; (c) Dental resin cement applied to the bottom of resin rod; (d) The resin rod placed on the surface; (e) Light curing; (f) knife-edge shear bonding test.

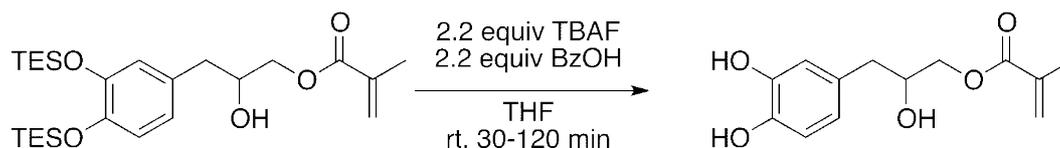
196

197 3. Results and Discussion

198 3.1. Improved synthesis of Catechol methacrylate primer

199 We had some concerns and encountered some difficulties with the previously
200 reported TES-deprotection method [6]. The prior method involved the use of
201 tetrabutylammonium fluoride (TBAF) as limiting reagent, and an excess of the
202 more valuable TES-protected intermediate, the reaction apparently being driven to
203 completion by reaction with adventitious water. Since the prior procedure
204 required that TBAF be used without a buffer, phenolic groups are liable to stay
205 ionized and in the aqueous phase without an acidic workup. Additionally, since
206 the previous conditions are basic due to the use of fluoride anion; catechol moieties
207 are susceptible to autooxidation, resulting in lower yields, yellow sample
208 coloration, and baseline impurities. These basic conditions also apparently are
209 the origin of the observed migration of some of the methacrylates to the internal
210 site visible in the proton NMR. In this study, we have increased the yield, purity,
211 and reproducibility of this silyl-deprotection by buffering the TBAF in the reaction
212 mixture with an equimolar amount of benzoic acid (Scheme 1).

213



Scheme 1. Synthetic scheme of catechol methacrylate from silylated eugenol.

3.2. SEM imaging of primed zirconia surface

217 Surface morphology of zirconia surfaces with and without primer treatments
 218 was studied with a SEM (Figure 3). In the SEM study, CP containing catechol
 219 methacrylate shows a complete coverage of zirconia surface, whereas the
 220 commercial zirconia primers (ZL, AP and UP) containing phospho- and/or
 221 carboxyl- methacrylates show partial coating on zirconia surfaces. We predicted
 222 that higher surface coverage would be positively correlated with the adhesive
 223 performance of dental resins to zirconia surface, by allowing for a greater number
 224 of contacts between zirconia and resin, which was subsequently supported by the
 225 results of the shear bonding tests (*vide infra*).
 226

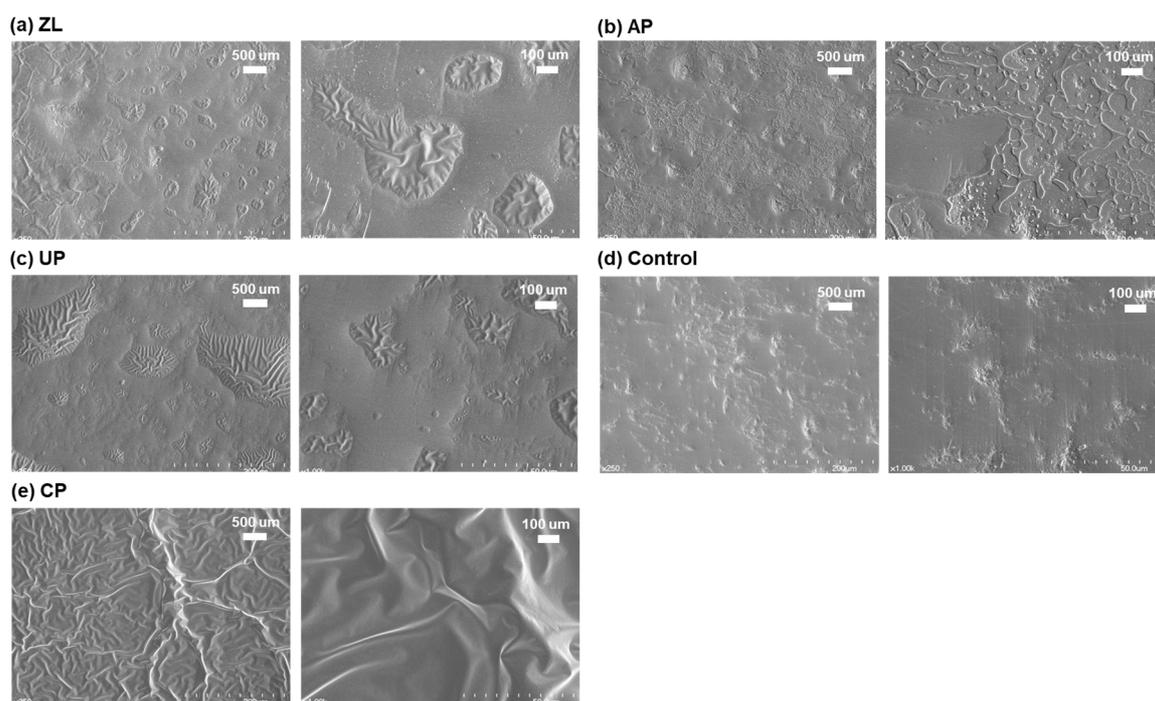
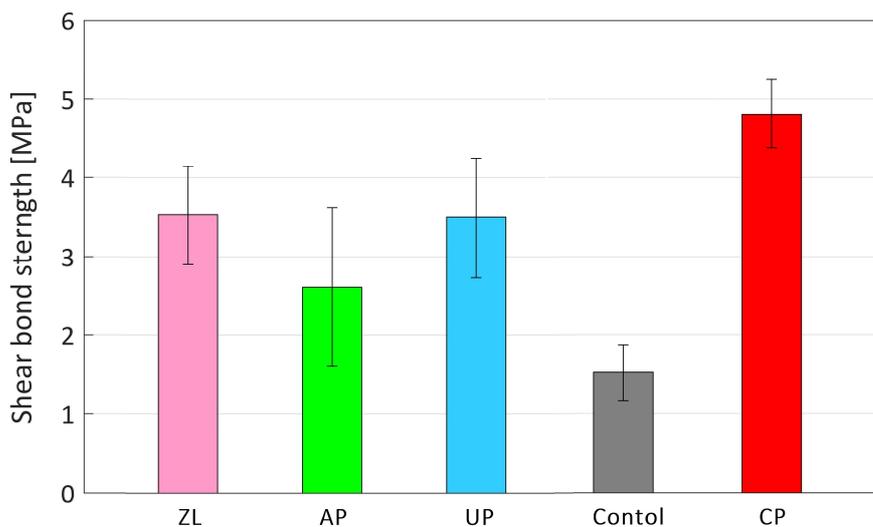


Figure 3. SEM images of zirconia surfaces primed with: (a) ZL: Zirconia Liner, (b) AP: Alloy Primer, (c) UP: Universal Primer, (d) Control: No primer, (e) CP: Catechol primer.

3.3. Shear bonding strength of dental adhesives on bioinspired catechol primed zirconia surface

232 Knife shear bonding test was conducted to measure bonding performance of
 233 dental adhesives on zirconia surface. The means, standard deviations of the
 234 knife-edge shear bond strengths are shown in Figure 4. With the complete surface
 235 coverage of CP on zirconia surface shown in SEM analysis in Figure 3, the shear

236 bonding of CP ($4.81 \text{ MPa} \pm 0.43$, $n = 10$; the mean of maximum shear strength, \pm as
237 standard deviation, and the n is number of experiments) was higher than
238 commercial zirconia primers containing phosphate and carboxylate methacrylates,
239 i.e., ZL ($3.52 \text{ MPa} \pm 1.01$, $n = 10$), AP ($2.61 \text{ MPa} \pm 0.76$, $n = 10$) and UP ($3.49 \text{ MPa} \pm$
240 1.03 , $n = 10$), and three times stronger than the control without any primer (1.52
241 $\text{MPa} \pm 0.35$, $n = 10$). The statistical certainty of the comparison ($p < 0.05$) was
242 confirmed by the Student's t -test.
243



244

245 **Figure 4.** Knife shear bonding strength of ZL, AP, UP, Control (No primer) and CP.

246

247 4. Conclusions

248 In this study, we were able to replicate one of the marine mussel's mechanisms
249 of priming mineral surfaces by using a synthetic small molecule to prime zirconia
250 surfaces. Our strategy of tethering of a polymerizable methacrylate to a catechol
251 moiety was validated by superior bonding performance of our bioinspired primer
252 to zirconia, relative to several popular commercially available formulations. SEM
253 imaging shows that in contrast to those utilizing acidic monomers, the catecholic
254 primer provides a much greater degree of surface coverage, which is correlated
255 with the observed enhancement in dental bonding performance. Considering the
256 catecholic primers' higher strength and ease of application, this priming strategy is
257 well poised for further development in dental applications requiring bonding to
258 zirconia.

259

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263

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265 experiments; R.L. synthesized the catechol methacrylate; K.A. wrote the manuscript; all authors have
266 contributed to analyzing the data and revising the manuscript.”

267

268 **Conflicts of Interest:** The authors declare no conflict of interest.

269

270 References

- 271 1. Piconi, C.; Maccauro, G. Zirconia as a ceramic biomaterial. *Biomaterials* **1999**, *20*, 1-25.
- 272 2. Paes, P.N.G.; Bastian, F.L.; Jardim, P.M. The influence of y-tzp surface treatment on topography and
273 ceramic/resin cement interfacial fracture toughness. *Dent. Mater.* **33**, 976-989.
- 274 3. Chuang, S.-F.; Kang, L.-L.; Liu, Y.-C.; Lin, J.-C.; Wang, C.-C.; Chen, H.-M.; Tai, C.-K. Effects of silane- and
275 mdp-based primers application orders on zirconia–resin adhesion—a tof-sims study. *Dent.*
276 *Mater.* **33**, 923-933.
- 277 4. Holten-Andersen, N.; Waite, J.H. Mussel-designed protective coatings for compliant substrates. *Journal of*
278 *Dental Research* **2008**, *87*, 701-709.
- 279 5. Martinez Rodriguez, N.R.; Das, S.; Kaufman, Y.; Wei, W.; Israelachvili, J.N.; Waite, J.H. Mussel adhesive
280 protein provides cohesive matrix for collagen type-1 α . *Biomaterials* **2015**, *51*, 51-57.
- 281 6. Seo, S.; Lee, D.W.; Ahn, J.S.; Cunha, K.; Filippidi, E.; Ju, S.W.; Shin, E.; Kim, B.S.; Levine, Z.A.; Lins, R.D., *et*
282 *al.* Significant performance enhancement of polymer resins by bioinspired dynamic bonding. *Adv. Mater.*
283 **2017**, *29*.
- 284 7. Kastrup, C.J.; Nahrendorf, M.; Figueiredo, J.L.; Lee, H.; Kambhampati, S.; Lee, T.; Cho, S.W.; Gorbатов, R.;
285 Iwamoto, Y.; Dang, T.T., *et al.* Painting blood vessels and atherosclerotic plaques with an adhesive drug
286 depot. *Proc. Natl. Acad. Sci. U.S.A.* **2012**, *109*, 21444-21449.
- 287 8. Kivelio, A.; Dekoninck, P.; Perrini, M.; Brubaker, C.E.; Messersmith, P.B.; Mazza, E.; Deprest, J.;
288 Zimmermann, R.; Ehrbar, M.; Ochsenein-Koelble, N. Mussel mimetic tissue adhesive for fetal membrane
289 repair: Initial in vivo investigation in rabbits. *European Journal of Obstetrics & Gynecology and Reproductive*
290 *Biology* **2013**, *171*, 240-245.
- 291 9. Oh, D.X.; Prajatelista, E.; Ju, S.-W.; Jeong Kim, H.; Baek, S.-J.; Joon Cha, H.; Ho Jun, S.; Ahn, J.-S.; Soo
292 Hwang, D. A rapid, efficient, and facile solution for dental hypersensitivity: The tannin–iron complex.
293 *Scientific Reports* **2015**, *5*, 10884.
- 294 10. Lee, S.-B.; González-Cabezas, C.; Kim, K.-M.; Kim, K.-N.; Kuroda, K. Catechol-functionalized synthetic
295 polymer as a dental adhesive to contaminated dentin surface for a composite restoration.
296 *Biomacromolecules* **2015**, *16*, 2265-2275.
- 297 11. Ahn, B.K. Perspectives on mussel-inspired wet adhesion. *J. Am. Chem. Soc.* **2017**, *139*, 10166-10171.
- 298 12. Danner, E.W.; Kan, Y.J.; Hammer, M.U.; Israelachvili, J.N.; Waite, J.H. Adhesion of mussel foot protein
299 mfp-5 to mica: An underwater superglue. *Biochemistry* **2012**, *51*, 6511-6518.
- 300 13. Ahn, B.K.; Das, S.; Linstadt, R.; Kaufman, Y.; Martinez-Rodriguez, N.R.; Mirshafian, R.; Kesselman, E.;
301 Talmon, Y.; Lipshutz, B.H.; Israelachvili, J.N., *et al.* High-performance mussel-inspired adhesives of
302 reduced complexity. *Nat Commun* **2015**, *6*.
- 303 14. Rickman, R.D.; Verkhoturov, S.V.; Balderas, S.; Bestaoui, N.; Clearfield, A.; Schweikert, E.A.
304 Characterization of surface structure by cluster coincidental ion mass spectrometry. *Appl. Surf. Sci.* **2004**,
305 *231-232*, 106-112.
- 306 15. Shin, E.; Ju, S.W.; An, L.; Ahn, E.; Ahn, J.-S.; Kim, B.-S.; Ahn, B.K. Bioinspired catecholic primers for rigid
307 and ductile dental resin composites. *ACS Applied Materials & Interfaces* **2018**, *10*, 1520-1527.
- 308 16. Ahn, B.K.; Lee, D.W.; Israelachvili, J.N.; Waite, J.H. Surface-initiated self-healing of polymers in aqueous
309 media. *Nat Mater* **2014**, *13*, 867-872.
- 310 17. Seo, S.; Das, S.; Zalicki, P.J.; Mirshafian, R.; Eisenbach, C.D.; Israelachvili, J.N.; Waite, J.H.; Ahn, B.K.
311 Microphase behavior and enhanced wet-cohesion of synthetic copolyampholytes inspired by a mussel
312 foot protein. *J. Am. Chem. Soc.* **2015**, *137*, 9214-9217.
- 313 18. Gelbke, H.P.; Knuppen, R. A new method for preventing oxidative decomposition of catechol estrogens
314 during chromatography. *J. Chromatogr. A* **1972**, *72*, 465-471.
- 315