

1 Article

2 Biodiversity, evolution and ecological specialization 3 of baculoviruses: a treasure trove for future applied 4 research

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15 **Abstract:** The *Baculoviridae*, a family of insect-specific large DNA viruses, is widely used in both
16 biotechnology and biological control. Its applied value stems from millions of years of evolution
17 influenced by interactions with their hosts and the environment. To understand how ecological
18 interactions, have shaped baculovirus diversification, we reconstructed a robust molecular
19 phylogeny using 217 complete genomes and ~580 isolates for which at least one of four
20 lepidopteran core genes was available. We then used a phylogenetic-concept-based approach
21 (mPTP) to delimit 165 baculovirus species, including 38 species derived from new genetic data.
22 Phylogenetic optimization of ecological characters revealed a general pattern of host conservatism
23 punctuated by occasional shifts between closely related hosts and major shifts between
24 lepidopteran superfamilies. Moreover, we found significant phylogenetic conservatism between
25 baculoviruses and the type of plant growth (woody or herbaceous) associated with their insect
26 hosts. In addition, we found that colonization of new ecological niches sometimes led to viral
27 radiation. These macroevolutionary patterns show that besides selection during the infection
28 process, baculovirus diversification was influenced by tritrophic interactions, explained by their
29 persistence on plants and interactions in the midgut during horizontal transmission. This complete
30 eco-evolutionary framework highlights the potential innovations that could still be harnessed from
31 the diversity of baculoviruses.

32 **Keywords:** cophylogeny; granulovirus; host shifts; Lepidoptera; mPTP; multitrophic interactions;
33 niche conservatism; nucleopolyhedrovirus; phylogenetics; resource tracking; species delimitation.
34

35 1. Introduction

36 The use of baculoviruses (BVs) as expression vectors has mainly focused on the development of
37 a single virus, namely *Autographa californica multiple nucleopolyhedrovirus* (AcMNPV) [1], and to a
38 lesser extent, *Bombyx mori nucleopolyhedrovirus* (BmNPV) [2]. The commercial success of AcMNPV
39 for biotechnological application is undeniable. Aside from historical reasons, the fact that it is a
40 generalist virus that can infect cell lines from different hosts probably explains why AcMNPV was
41 first chosen. The family *Baculoviridae*, however, encompasses hundreds of isolates, many of which
42 have been studied in the context of biological control of insect pests, but some of which could in the
43 future prove equally as useful as AcMNPV for biotechnological applications, by providing new
44 molecular and biochemical products with contrasting antigenic properties or by infecting new, more
45 productive cell lines. In this context, it is important to describe the taxonomical diversity of BVs to
46 ensure that new isolates developed for biotechnological products are not examples of commonly

47 used viral species, which could infringe patents. However, to delineate species boundaries requires
48 an understanding of BV evolution based on both historical and ecological perspectives.

49 The delimitation of species is a notoriously difficult question in biology, particularly for those
50 organisms for which the biological species concept cannot be applied. Viral species are defined by
51 the International Committee on Taxonomy of Viruses (ICTV) as “a monophyletic group of viruses
52 whose properties can be distinguished from those of other species by multiple criteria” [3]. This
53 general definition recognizes the importance of shared evolutionary history to define viral lineages
54 but allows for other biological criteria to be used, whether or not they may have an evolutionary
55 meaning. Recent advances in virus discovery (e.g. [4]) have spurred the need to reconsider the
56 method used for virus classifications [5,6]. Genetic distances are commonly used to delimitate viral
57 species based on distance cut-offs [5]. In the case of BVs, the cut-off between species has been
58 defined as between 0.015 and 0.05 based on the Kimura-2-parameter on sequence alignments of the
59 polyhedrin, lef-8 or lef-9 genes [7]. There is no doubt about the usefulness of those distance
60 thresholds to cluster closely related isolates (distance below 0.015) and to separate distantly related
61 viruses. However, there is often a high degree of uncertainty when clustering at intermediate levels
62 of genetic distances (up to 0.05). Intermediate levels of genetic differentiation could be found within
63 widespread, generalist, or fast evolving viral species. In these cases, the sole use of genetic distances
64 is insufficient to infer clusters and the delimitation criteria are left to the judgment of the taxonomist.
65 There is therefore a need to consider alternative analyses for the delimitation of BV species, which
66 could also be applied to other viral families. Recent advances in molecular phylogenetics now allow
67 the clustering of sequences into species groups, taking into account differences in evolutionary rates
68 or based on coalescence theory (e.g. [8,9]), and which do not require prior knowledge of species
69 limits. These automated methods, designed for DNA barcoding (e.g. [10,11]) to group individual
70 animal sequences of typically one gene into species clusters, are based on universal theory and can
71 therefore be applied to other genes and taxa [12], including viruses.

72 As insect-specific, enveloped, rod-shaped viruses with large circular double-stranded DNA
73 genomes, BVs replicate in the nucleus of infected host cells. Their life cycle is characterized by the
74 packaging of virions within a matrix of protein, forming the so-called occlusion body (OB), which is
75 the vehicle of horizontal transmission. Over 600 BVs have been described from the insect orders
76 Lepidoptera, Hymenoptera and Diptera. More than 90% of the known BVs have been isolated from
77 lepidopteran hosts [13]. The current classification [14] divides the *Baculoviridae* family into four
78 genera [15]. Two genera, the *Alphabaculovirus* and *Betabaculovirus*, infect lepidopteran species, and
79 differ by the morphology of their OBs forming the nucleopolyhedroviruses (NPVs) and
80 granuloviruses (GVs) respectively. The OBs of NPVs contain many virions and have been isolated
81 from both lepidopteran and non-lepidopteran hosts; in contrast, the OBs of the lepidopteran GV
82 contain a single virion [14]. Phylogenetic and comparative genomic studies have shown that both
83 Lepidoptera specific genera are sister groups and that they diversified during the Mesozoic Era after
84 the diversification of insect orders [7,16-18].

85 OBs are the between host transmission stage of the virus and allow the virus to survive outside
86 the host [19]. BVs are generally obligate killers as the host must die before the OBs are released into
87 the environment for horizontal transmission. But latent baculovirus infections [20,21] and vertical
88 transmission [22-24] have been reported. Insect larvae become infected by ingesting host plant tissue
89 contaminated with OBs. Thus, plant and virus have a close interaction in the insect gut and plant
90 chemicals can interact with the virus in numerous ways resulting in altered infectivity [25]. There is
91 therefore the potential for plants to play an important role in the evolution of insect-BV interactions
92 [25-28]. A better understanding of these diverse evolutionary interactions might point to new
93 molecular targets that could lead to biotechnological innovations.

94 In this study our main aims were to 1) provide a robust phylogeny for the whole family
95 *Baculoviridae* using all the genetic data generated so far; 2) delineate baculovirus species using a
96 non-a-priori phylogenetic-concept-based approach; 3) study the evolutionary history of host use of
97 BVs, assessing the level of host specialization and role of host shifts in the speciation of BVs; 4)

98 elucidate the role of host plants in the evolution of BVs. Our results shed light on the biodiversity
99 ecological and evolutionary factors that drive *Baculoviridae* diversification.

100 2. Materials and Methods

101 2.1 Virus isolate sequence database

102 We created a DNA sequence database containing sequences of at least one of the four
103 lepidopteran BV core genes, late expression factor 8 (lef-8), late expression factor 9 (lef-9), per os
104 infectivity factor 2 (pif-2) and polyhedrin (polh). We chose those genes because it has been shown
105 that they bear a strong phylogenetic signal [16] and have been abundantly sequenced. The database
106 includes sequences collated from the public databases GenBank, EMBL and DDBJ (version April
107 2018), and we tried to obtain sequences from one hundred historical BV samples originating from
108 the reference collection at the Centre for Ecology and Hydrology (NERC-CEH), Wallingford, United
109 Kingdom. Primers, PCR amplification protocols and Sanger sequencing are previously described
110 [7,15]. Following current taxonomic practice, each BV sample was associated with the host species
111 from which it was isolated and OB morphology (NPV or GV), as well as sampling information about
112 the virus isolate or strain (if provided), and the name of the first author of the study from which BV
113 sequences derived (Table S1).

114 2.2 Host ecology database

115 For each BV isolate studied, we associated host ecological data. We included the taxonomy of
116 each insect host species (superfamily, family, subfamily), and their geographical distribution
117 (ecozone), established from localities where insect hosts were observed in the field. We also included
118 the host plant range of each insect host, from which we determined the associated insect host plant
119 growth type, distinguishing woody perennial (including shrubby and suffrutescent plants) and
120 annual/biennial herbaceous plants. Information was mainly extracted from the database of the
121 world's lepidopteran host plants of the Natural History Museum of London [29], from the Barcode
122 of Life Data System (BOLD) [30] and from the literature [31] (Table S2).

123 2.3 Baculovirus core-genome phylogeny

124 A phylogenomic approach was used to reconstruct the BV core-genome phylogeny. BVs
125 possess 37 core genes [32] identified in all completely sequenced genomes. Amino acid multiple
126 alignments were performed on the 37 BV core gene products using MAFFT program [33] and,
127 alignments were concatenated prior to the phylogenetic reconstruction. A maximum likelihood
128 (ML) phylogenetic inference was performed on the concatenated multiple alignment under the Le
129 and Gascuel amino acid substitution model, with a gamma distributed among site rate variation and
130 a proportion of invariant sites (LG + Γ + I), as determined by ProtTest3 [34]. The ML analysis was
131 performed with RAxMLv.8.2 program [35] and statistical support for nodes in the ML tree was
132 assessed using a bootstrap approach (with 100 replicates).

133 2.4 Baculovirus isolate phylogeny

134 For each of the four BV core genes (lef-8, lef-9, pif-2 and polh) codon-based multiple alignments
135 were performed on all BV isolate sequences. Alignments were then concatenated and completed
136 with gaps for isolates, for which up to three of the four lepidopteran core genes might be missing. A
137 ML phylogenetic inference was performed on the concatenated codon-based multiple alignment
138 with RAxMLv.8.2 using the BV core-genome phylogeny as backbone constraint tree. The BV isolate
139 phylogeny was reconstructed using the best-fitted substitution model and parameters GTR + Γ + I, as
140 determined by jModelTest2 and statistical support for nodes in the ML tree was assessed using a
141 bootstrap approach (with 100 replicates).

142

143 2.5 Species delimitation

144 A species delimitation analysis was performed on the BV isolate phylogeny using a
145 phylogenetic-species-concept-based method, the multi-rate Poisson tree processes (mPTP) model
146 [36]. The mPTP method is an improved version of the PTP model [9], which models speciation or
147 branching events in terms of number of substitutions and uses heuristic algorithms to identify the
148 most likely classification of branches into population and species-level processes. Moreover, mPTP
149 is fast and incorporates different levels of intraspecific genetic diversity deriving from differences in
150 either the evolutionary history or sampling of each species [36]. The mPTP model delimit species on
151 phylogenies without a priori assumptions, instead of the genetic distance cut-off commonly used.
152 The mPTP results were compared to the commonly used genetic-distances-based method. A
153 previous study calculated an intra-species genetic distance of ≤ 0.015 (up to 0.05 with
154 complementary information) as marker to delimit BV species [7]. This distance is commonly used by
155 the BV taxonomists and was evaluated using the Geneious plugin SpDelim [37], which assesses the
156 within and between species genetic distances in a phylogenetic tree. For both mPTP and SpDelim
157 analyses, the BV isolate ML phylogeny was used.

158 2.6 *Baculovirus* species phylogeny

159 From the species delimitation analysis, one isolate per cluster was selected as a representative of
160 a putative BV species (e.g. isolates with a complete genome or isolates with the four core genes
161 sequenced) and singletons were considered as distinct putative BV species. The topology of these
162 species-representative isolates was extracted from the BV isolate phylogeny by pruning the other
163 taxa, using the 'ape' package [38] for R. A new amino acid multiple alignment was created with only
164 the set of the species-representative isolates. A Bayesian phylogenetic inference was performed on
165 this alignment using BEASTv.1.8.4 [39]. The substitution model and parameters LG + Γ + I were used
166 as well as a fixed strict clock of 1.0. The topology of the above species-representative isolate tree was
167 used as target tree for the calculation of the consensus tree and posterior probabilities.

168 2.7 Phylogeny-trait correlation and ancestral state estimations

169 Correlations between phylogenetic tree structure and isolate trait discrete values (see Table S2)
170 were assessed using the methods implemented in BaTS/befi-BaTS [40]. The number of state
171 randomizations was defined to 100 to yield a null distribution. A correlation was considered
172 unambiguously positive if the Association Index (AI), the Parsimony Score (PS), the Phylogenetic
173 Diversity (PD), the Nearest Taxa (NT) and the Net Relatedness (NR) indices, the Unique Fraction
174 (UniFrac) index, and the maximum monophyletic clade (MC) size probabilities were ≤ 0.01 .
175 Phylogenetic uncertainty was taken into account by using the set of tree topologies estimated by the
176 above Bayesian phylogenetic inference. For those traits that obtained significant P-value, a new
177 Bayesian phylogenetic inference was performed on the species-representative isolate alignment with
178 the same set of parameters as above and adding discrete trait partitions to reconstruct ancestral
179 states.

180 2.8 Host-virus cophylogeny

181 To test for the existence of statistically significant topological congruence between BV species
182 and Lepidoptera hosts, we estimated the maximum number of cospeciation events at different
183 taxonomic levels (superfamily, family, subfamily and genus), needed for reconciling BV species and
184 host phylogenies in TreeMap 3b [41] and compared this estimate to the distribution of
185 corresponding values obtained by randomizing different Lepidoptera trees 100 times while keeping
186 BV-host associations unchanged [42]. Tests at superfamily, family and subfamily taxonomic levels
187 compared the whole BV species phylogeny to published Lepidoptera phylogenies [43-45]. Tests at
188 the genus level were performed by reconstructing lepidopteran host species phylogenies based on
189 published phylogenies (Noctuidae [46], Erebidae [47,48], Lasiocampidae [49], *Spodoptera* genus [50])
190 or when not available on CO1 sequence data extracted from BOLD public database [30] and

191 compared to the BV species tree simplified by pruning monophyletic clades of species sharing a
192 given host genus down to a single species. Analyses were performed using 25 starts from random
193 maps and heuristic searches for up to 25 generations.

194 3. Results

195 3.1. *Baculovirus* phylogeny

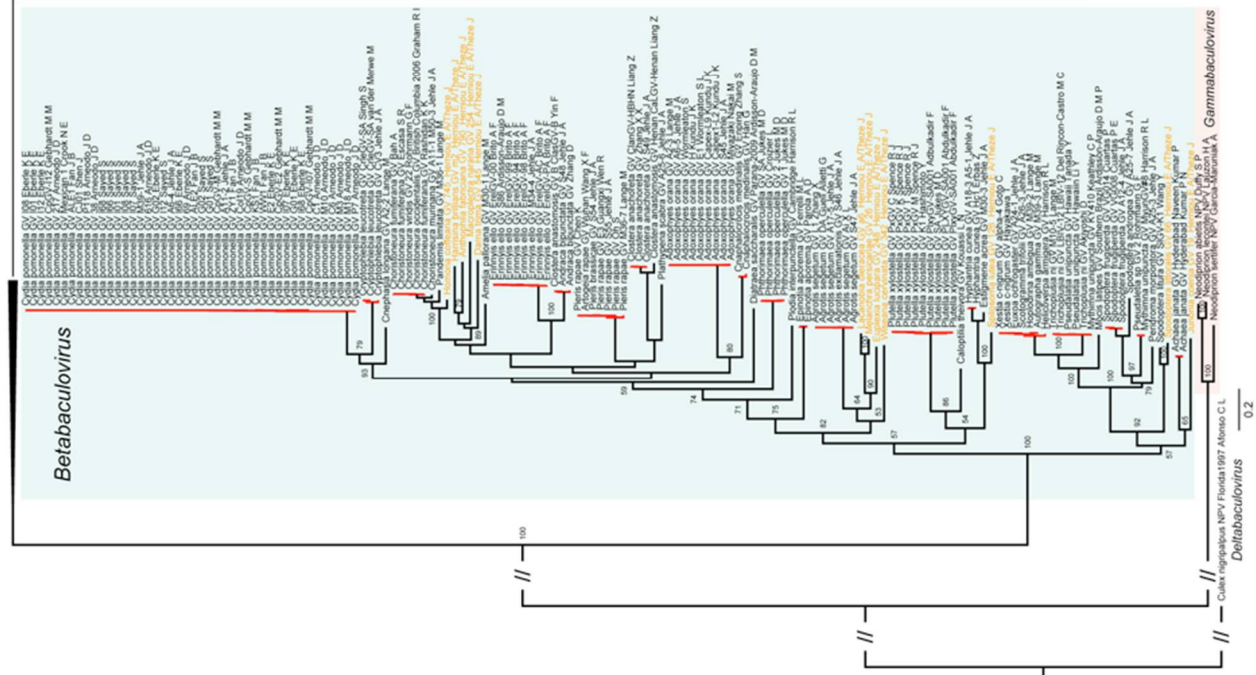
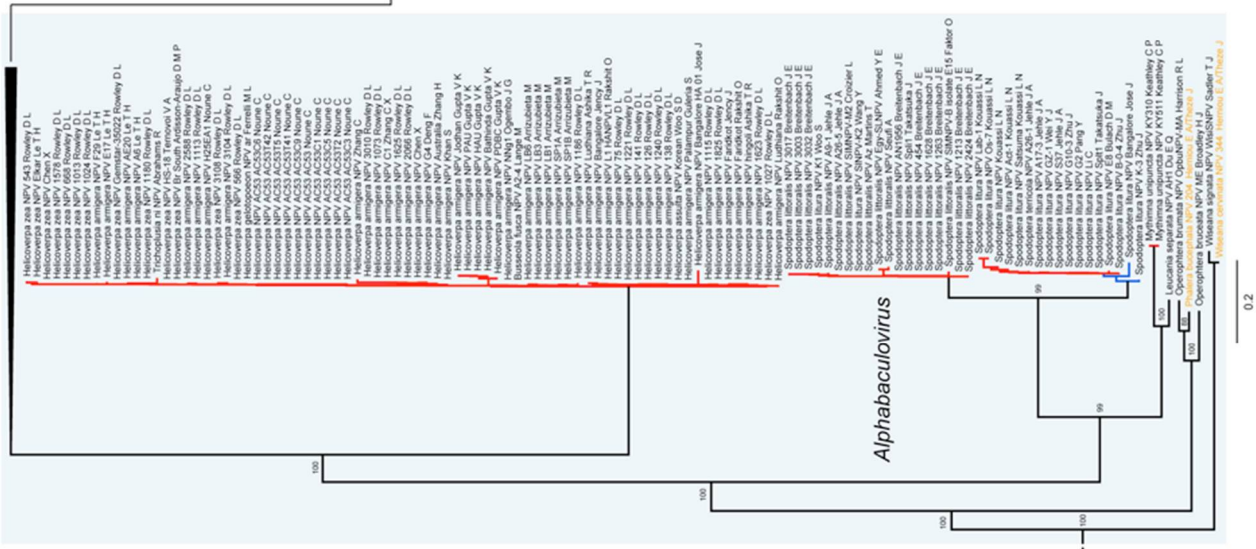
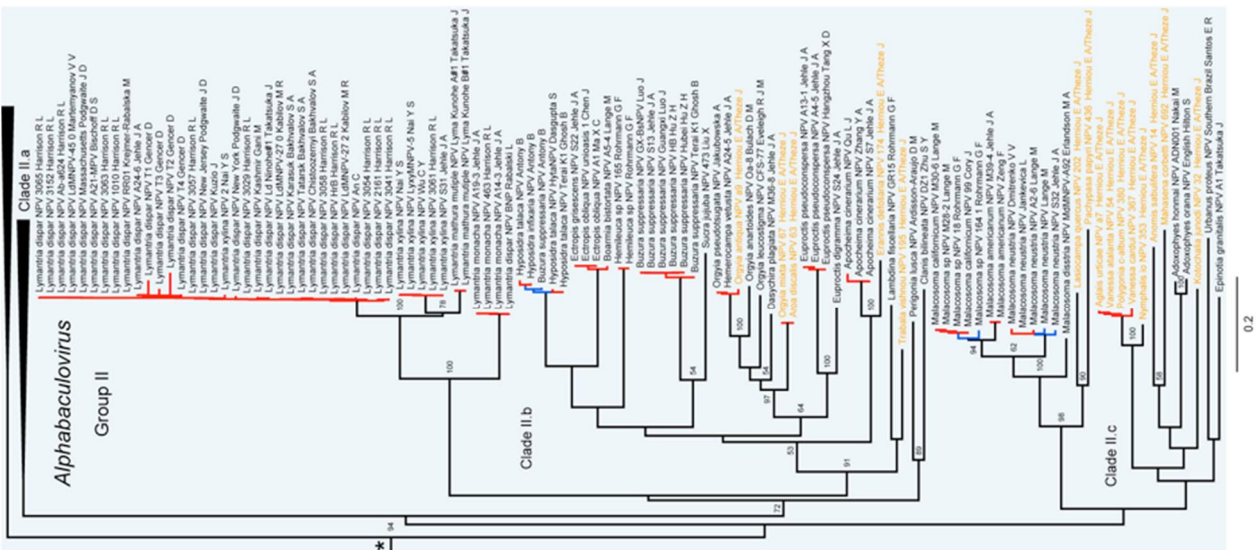
196 Here we present a comprehensive phylogeny of BVs, including most of the relevant BV genetic
197 data available to date. A data-mining analysis was performed on public genetic databases and
198 resulted in the collation of 749 BV isolates properly formatted, containing the nucleotide sequences
199 of at least one of four lepidopteran BV core genes (*lef-8*, *lef-9*, *pif-2* and *polh* genes). We were also
200 able to determine the sequences for 45 historical isolates from our own collection. Overall our
201 working sequence database contains 2053 nucleotide sequences (564 *lef-8* genes, 498 *lef-9* genes, 283
202 *pif-2* genes and 708 *polh* genes) belonging to 794 BV isolates (Table S1). A total of 235 isolates have
203 the sequences of the four genes.

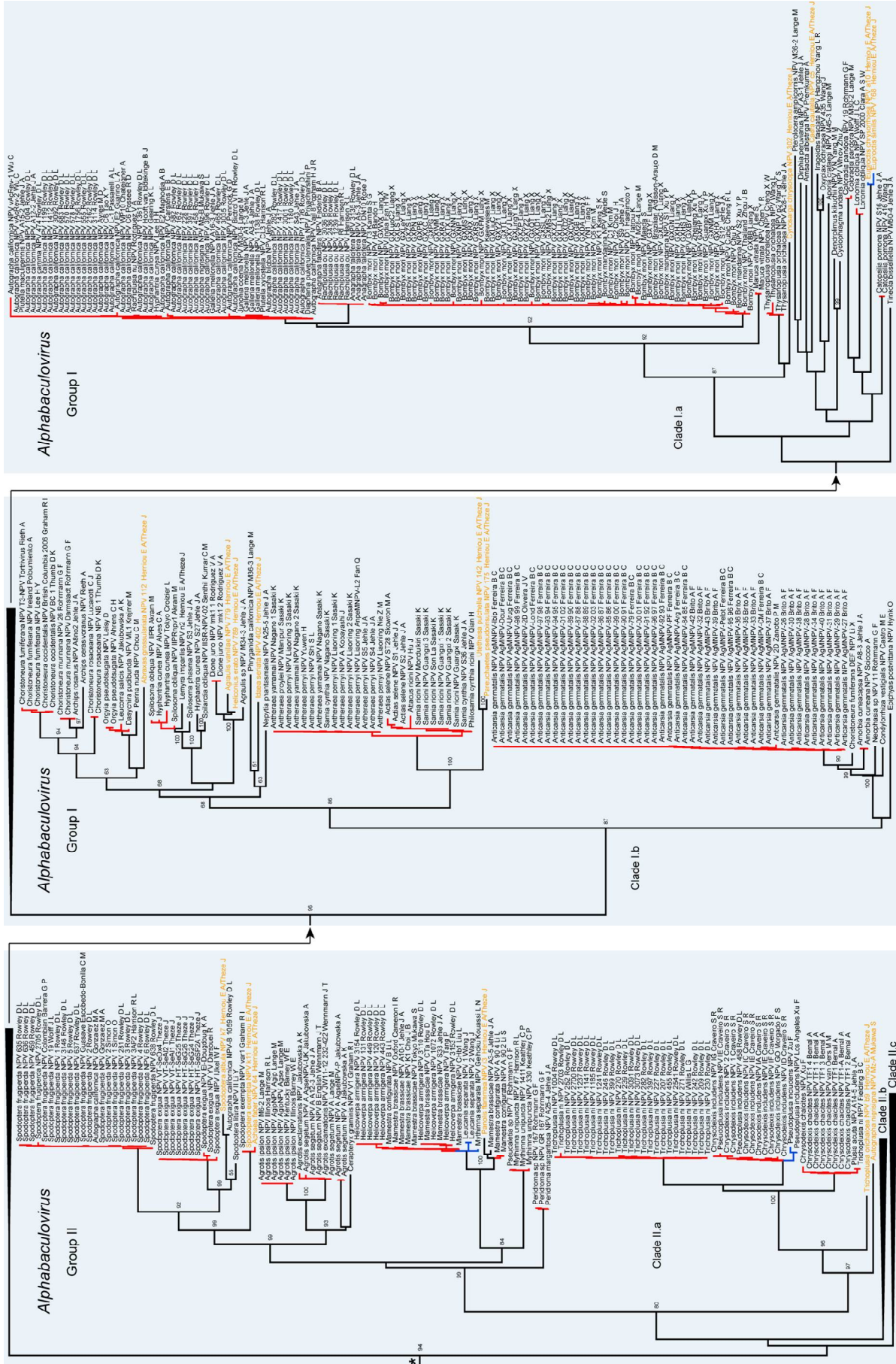
204 Among the BV isolates, 217 have complete genomes and were used to reconstruct the highly
205 supported BV core-genome phylogeny (Figure S1), inferred from the concatenated multiple amino
206 acid alignment of the 37 BV core genes. The tree topology is in accordance with previous studies
207 showing four BV genera infecting three distinct insect orders: the genera *Alphabaculovirus* and
208 *Betabaculovirus* infect lepidopteran species, the genus *Gammabaculovirus* infects hymenopteran
209 species and the genus *Deltabaculovirus* infects dipteran species [14,15]. The topology obtained was
210 used as backbone constraint tree to reconstruct the BV isolate phylogeny. The BV isolate phylogeny
211 (Figure 1; Figure S2) is quite well supported and is in accordance with previous studies showing
212 similar topologies [7,14,17] with notably a separation of two major monophyletic subgroups of
213 alphabaculoviruses (Group I, Group II). In addition, four minor monophyletic subgroups are
214 outgroups to Group I and II. Within Group I we observed a clear division into two distinct
215 monophyletic clades (I.a and I.b) and Group II is divided into three distinct monophyletic clades (II.a,
216 II.b and II.c) (Figure 1; Figure S2).

217 3.2. *Species delimitation*

218 To infer macroevolutionary process, it is necessary to conduct analyses at the species level to
219 avoid interference from intraspecific diversity. A species delimitation analysis was performed on the
220 BV isolate tree (Figure 1; see also Figure S2) using the mPTP method and then compared to the
221 commonly used genetic-distances based method. The mPTP approach delimited 165 distinct
222 putative BV species of which 70 putative BV species have been derived from clusters of two or more
223 isolates and 95 putative species are singletons (based on a unique viral isolate) (Figure 1; see also
224 Figure S2 and Figure S3). The dataset includes 38 new baculovirus species from our historical
225 isolates. The genetic-distances based method delimited 178 putative BV species (72 clusters and 106
226 singletons; Figure 1; see also Figure S2 and Figure S4). The two approaches show 157 species in
227 common, the differences in the genetic-distances based method were mostly observed in
228 alphabaculovirus species showing high levels of sampling and genetic variability (Figure 1; see also
229 Figure S2).

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235 **Figure 1. Baculovirus isolate phylogeny.** The tree was obtained from a maximum likelihood
 236 inference analysis of the concatenated codon-based alignment (794 taxa) of four lepidopteran
 237 baculovirus core genes with the baculovirus core-genome phylogeny used as backbone tree. External
 238 clades colored in red correspond to clusters determined by both the mPTP and SpDelim species
 239 delimitation analysis and in blue the clusters not determined by SpDelim. The two star symbols
 240 points out the same node in the tree. Baculovirus isolates generated in this study are highlighted in
 241 orange. Statistical support for nodes in the tree corresponds to bootstraps (with 100 replicates).

242 Within the 165 distinct species identified by mPTP, 116 putative species are alphabaculoviruses,
 243 45 are betabaculoviruses, three are gammabaculoviruses and one is a deltabaculovirus. The 10th
 244 report of the ICTV (2017, https://talk.ictvonline.org/ictv-reports/ictv_online_report/) only currently
 245 recognizes 68 species in the *Baculoviridae*: 40 in the genus *Alphabaculovirus*, 25 in the genus
 246 *Betabaculovirus*, two in the genus *Gammabaculovirus* and one in the genus *Deltabaculovirus* [14]. The
 247 reduced tree containing only putative BV species (Figure 2) suggests, therefore, 97 new putative BV
 248 species, including 76 alphabaculoviruses and 20 betabaculoviruses.

249 3.3. Phylogenetic conservatism and host shifts

250 Out of the 165 alphabaculovirus and betabaculovirus species identified by mPTP, 161 are
 251 associated with 187 Lepidoptera species from 24 different families. For all those lepidopteran hosts,
 252 we compiled a dataset with the taxonomy (superfamily, family and subfamily), the biogeographical
 253 distribution and the insect host plant range, from which we determined the host plant growth type
 254 (woody versus herbaceous) (Table S2). We performed phylogeny-trait correlation and ancestral state
 255 estimation analyses on the BV species phylogeny to measure whether those traits have evolved
 256 randomly or show phylogenetic conservatism. The different phylogeny-trait correlation tests show
 257 unequivocal significant associations between the taxonomy of insect hosts, the insect host plant
 258 growth type and the BV species phylogeny (AI, PS, PD, NT, NR, UniFrac and MC; $P \leq 0.01$, Table 1).
 259 In contrast, no significant association was found between the insect host biogeography distribution
 260 and the BV species phylogeny, as shown by the high p -values of certain tests (NR, UniFrac and MC;
 261 $p > 0.01$, Table 1).

262 **Table 1.** Phylogeny-trait correlations estimated under different statistical methods

Traits	AI ¹	PS ¹	PD ¹	NT ¹	NR ¹	UniFrac ¹	MC ²
Host superfamily	<0.001	<0.001	<0.001	<0.001	<0.001	<0.001	<0.001 / 21
Host family	<0.001	<0.001	<0.001	<0.001	<0.001	<0.001	<0.001 / 21
Host subfamily	<0.001	<0.001	<0.001	<0.001	<0.001	<0.001	<0.001 / 7
Host biogeography distribution (ecozone)	<0.001	<0.001	<0.001	<0.001	0.58	0.029	0.019 / 4
Insect host plant growth type (Herb/Woody)	<0.001	<0.001	<0.001	<0.001	<0.001	<0.001	<0.001 / 22

264 ¹ p -value of AI: association index; PS: parsimony score; PD: phylogenetic diversity index; NT: nearest taxa
 265 index; NR: net relatedness index; UniFrac: unique fraction index; MC: maximum monophyletic clade size
 266 probability; ² p -value / size of MC: Maximum monophyletic clade size probability and size (number of taxa) of
 267 the largest monophyletic clade associated to one character of the trait

268 Ancestral state estimations on the baculovirus species tree were performed on traits that
 269 showed significant associations such as host taxonomy and insect host plant growth. The
 270 estimations show highly significant levels of phylogenetic conservatism with the different host
 271 taxonomic levels and the insect host plant growth type (Table 1). For the host taxonomy trait we
 272 illustrate only the insect host superfamily optimization as it is the higher taxonomic level and has a
 273 reduced number of character states (Figure 2). The ancestral state estimation of the insect host
 274

275 superfamilies shows that closely related BV species tend to infect closely related lepidopterans (BVs
276 cluster together according to the lepidopteran superfamilies) (Figure 2A). The host use optimization
277 identifies owlet moths (Noctuoidea) as the most likely ancestral hosts of BVs (Figure 2A). Despite
278 significant levels of phylogenetic conservatism several major host shifts across different
279 lepidopteran superfamilies have occurred during BV evolution. Thus, in the *Betabaculovirus* genus
280 we can see a host shift from Noctuoidea to the Tortricoidea superfamily and then colonization of
281 several superfamilies: Papilionoidea, Zygaenoidea, Bombycoidea and a shift back to the Noctuoidea
282 (Figure 2A). The *Alphabaculovirus* genus splits into two large lineages (Group I and II) and four small
283 lineages (Figure 2A). Group I colonizes several Lepidoptera superfamilies, Clade I.a shows no host
284 conservatism harboring a very diverse host range, whereas Clade I.b shows a medium level of host
285 conservatism with a shifting from Noctuoidea to Tortricoidea and Papilionoidea. Group II shows a
286 clear split between Clades II.a, II.b and II.c, with clades II.a and II.b showing a strong conservatism
287 towards Noctuoidea with Clade II.a being specific to Noctuoidea with no apparent host shift. Both
288 clades II.b and II.c show host shifts from Noctuoidea to Geometroidea and Lasiocampoidea and
289 colonization of Bombycoidea, Tortricoidea, Tineoidea and Papilionoidea (Figure 2A).

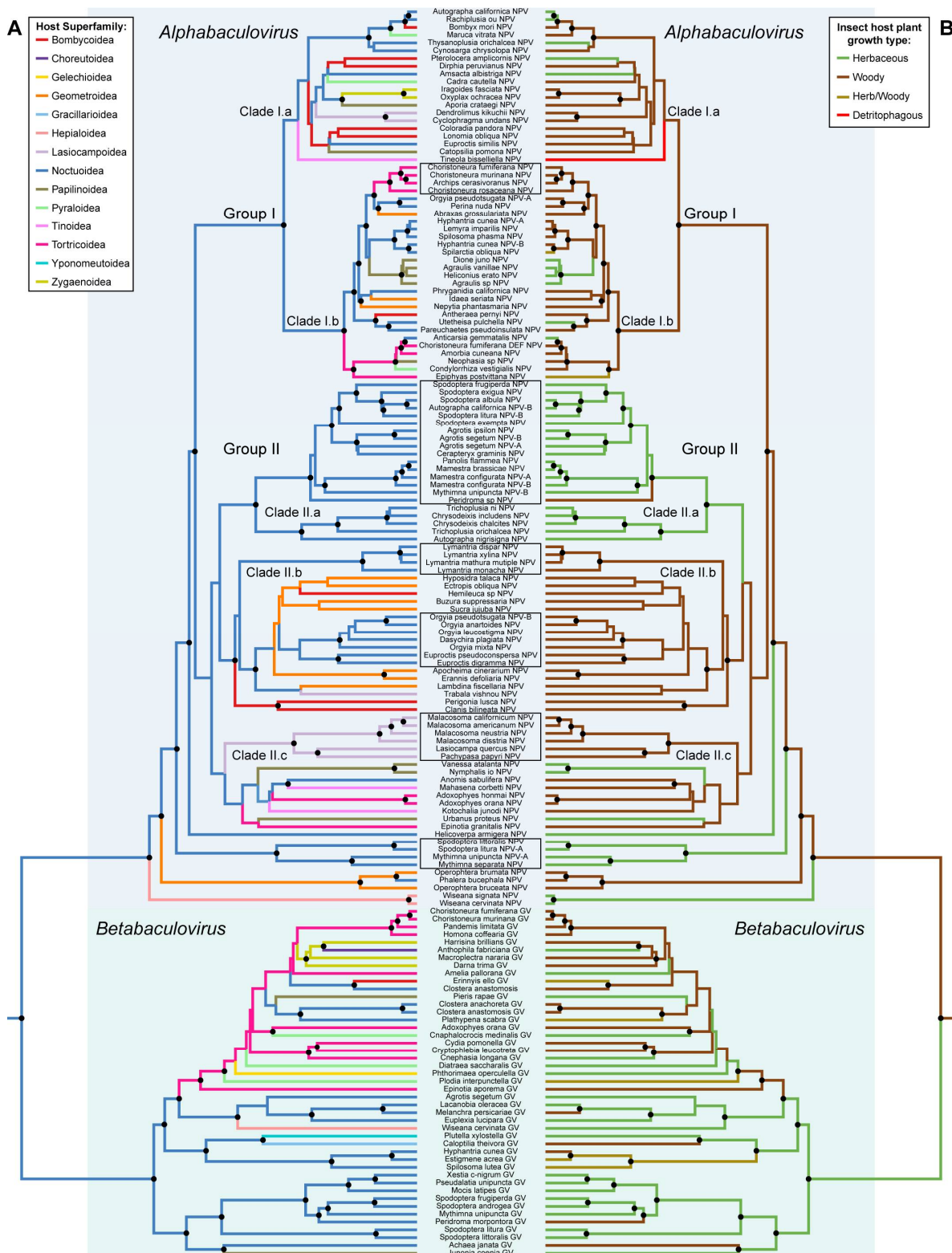
290 Insect host plant growth type (Figure 2B), which can be considered as the local habitat of the
291 virus, shows a strong phylogenetic conservatism (Table 1). Our analyses show woody plants as the
292 most likely ancestral ecological niche of the first lepidopteran BVs. The *Betabaculovirus* genus is
293 ancestrally associated with herbaceous plants before colonizing woody plants. The *Alphabaculovirus*
294 genus is ancestrally associated with woody plants. Group I shows a strong association with woody
295 plants with few shifts to herbaceous plants. The ancestors of group II likely fed on woody plants
296 with colonization of herbaceous plants (Clade II.a). The three small alphabaculovirus clades, that are
297 outgroups to Clade I and II, also show a shift to herbaceous plants. The most notable result on these
298 analyses is the long-term associations with a particular insect host plant growth type. Strikingly, we
299 noticed in the Group II of the *Alphabaculovirus* genus a split of virus infecting Noctuoidea, which
300 seems to be the result of the colonization of herbaceous plant local habitat in Clade II.a (Figure 2).

301 3.4. Cophylogeny between baculoviruses and their lepidopteran hosts

302 The association between the topology of the BV species phylogeny and that of their
303 lepidopteran hosts (at superfamily, family and subfamily levels) is not significantly different from
304 random. In contrast, we found significant topological congruence ($P < 0.5$) for six BV clades at
305 different taxonomic levels (see the six clades highlighted in Figure 2). The *Alphabaculovirus* genus
306 gathers all the clades with significant topological congruence with host topologies. One clade
307 infecting Tortricoidea hosts was detected in Group I, whereas four clades infecting Noctuoidea hosts
308 were identified, and notably three in Group II. One clade infecting Lasiocampoidea was also
309 detected in Group II.

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Figure 2. Phylogenetic optimizations on the lepidopteran baculovirus species tree. Optimizations were obtained by ancestral state estimation of A) host Lepidoptera at superfamily level, and B) insect host plant growth type traits on the ingroup of Lepidoptera infecting baculovirus. The term “Herb/woody” in the legend means that insect hosts feed on herbaceous plants as well as woody plants. Black circles close to phylogenetic nodes refer to posterior probabilities over 0.75. Baculovirus species highlighted with black line rectangles correspond to clades that are significantly topologically congruent ($P < 0.5$) with lepidopteran host clades.

320 4. Discussion

321 4.1. Reconstructing Phylogenies and Delineating Species

322 The *Baculoviridae* is by far the best described insect DNA virus family. For the last 50 years, BV
323 use in biotechnology, either as expression vectors [1] or as microbial control agents for insect pests
324 [51], has led to a vast production of molecular data, especially for the genera infecting the
325 Lepidoptera, on which this study is focused. The first objective of this study was to reconstruct the
326 most accurate and exhaustive BV isolate phylogeny in order to set a solid framework for species
327 delimitation and macroevolutionary inference.

328 The backbone of the tree was built based on 217 whole BV genomes (Figure S1). A secondary
329 ML analysis included an additional 577 isolates, from which we obtained our isolate tree (Figure 1;
330 Figure S2). We chose to cluster these viral isolates into species, which are the basis of biological
331 classification, in order to study baculovirus speciation at a macroevolutionary scale. Baculoviruses
332 like most of large DNA viruses are slow-evolving viruses with an approximate mutation rate of
333 $10^{-6}/10^{-7}$ (expressed as the number of substitutions per nucleotide per generation, defined as a cell
334 infection in viruses) [52]. This mutation rate approaches those observed in Bacteria and lower
335 Eukaryotes. Consequently, we decided to use a phylogenetic-species-concept-based clustering
336 approach, the mPTP model [36], to delimit BV species and to compare the results to the commonly
337 used genetic-distances-based method (intra-species distance ≤ 0.015 ; up to 0.05 with complementary
338 information [7]). Our species delimitation results were consistent results with the current taxonomy
339 proposed by the ICTV [14]. Out of 165 BV species we were able to characterize 97 that are not yet
340 included in the ICTV report. The BV species phylogeny reflects current knowledge on BV diversity
341 and phylogenetic relationships (Figure 1; Figure2). Furthermore, this study is the first to use a
342 phylogenetic clustering approach for species delimitation in viruses, showing that its utility goes
343 beyond vertebrates [53], invertebrates [54,55] and bacteria [12] taxa. Moreover, this approach for
344 species delimitation fully respects the phylogenetic species concept and is less arbitrary than
345 commonly used genetic distance approaches, which do not take into account differences in
346 molecular evolutionary rates or sampling proportions and may thus vary depending on the biology
347 of the lineage studied and on the gene used for phylogenetic reconstruction. Nevertheless, both
348 species delimitation approaches gave relatively consistent results with an overlap of 157 species
349 determined by both methods. The few differences in the genetic distances-based method were
350 mostly observed in heterogeneous species clusters, with isolates infecting one or several hosts and
351 showing high genetic diversity typically resulting in clusters with genetic distances between 0.015
352 and 0.050 [7]. The phylogenetic-based approach does not contradict the results of the
353 genetic-distances based approach but gives additional information to resolve the uncertainties of the
354 genetic-distances based approach.

355 4.2. Evolution of host use and taxa sampling

356 Most of the BV species identified in our study have isolates that infect only one lepidopteran
357 host species. Strikingly, we generally found that closely related hosts belonging to the same genus
358 were infected by different viral species (for example in the genera *Spodoptera*, *Lymantria*, or
359 *Malacosoma*). This leads us to question the ecological reality and biological meaning of some isolates,
360 such as *Trichoplusia ni* NPV and *Busseola fusca* NPV within the *Helicoverpa armigera* NPV clade
361 (Figure 1). However, generalist viruses, capable of infecting different host species, belonging or not
362 to the same genus, exist and the hosts they infect generally have overlapping ecological niches (same
363 host plants). As an example, several isolates from different species of nymphalid butterflies that feed
364 on nettles (*Urtica dioica*) form a single alphabaculovirus species *Vanessa atalanta* NPV.

365 As parasites replicating exclusively in host cells, BVs are involved in durable and intimate
366 obligate interactions with their host, implicating long-term coevolution. The phylogenetic
367 conservation results suggest an ancestral and frequent association with hosts of the Noctuoidea
368 superfamily. This possibly could reflect the actual evolution of lepidopteran BVs and their current
369 host range, as with ~42,000 out of ~157,000 described lepidopteran species the Noctuoidea is the

370 most diversified superfamily (in comparison, the second most abundant superfamily is the
371 Geometroidea with ~23,000 species) [31]. Yet, BV genetic data from public databases is clearly biased
372 towards agro-economically important lepidopteran pests, characterized by large populations,
373 important for sustaining BV populations. The addition of new BV samples from our collection tends
374 to reduce the bias for pests, but BVs from pests still dominate our taxa sampling as these viruses
375 have been isolated and resequenced many times (i.e. large clusters of *Cydia pomonella* GV or
376 *Helicoverpa armigera* NPV; Figure 1) and as at least 62 out of 161 putative lepidopteran BV species
377 are associated with pests (Figure 2A; Table S2). As numerous lepidopteran pests belong to the
378 Noctuoidea superfamily, this probably increases the representation of Noctuoidea infecting BVs in
379 our dataset and could have biased our phylogenetic conservatism results (77 out of 161 putative
380 lepidopteran BV species analyzed attack Noctuoidea, Figure 2A). Only a diverse BV sampling, more
381 representative of the lepidopteran diversity could confirm if Noctuoidea played a key role in BV
382 evolution.

383 4.3. *Cophylogeny and host shifts*

384 Cophylogenetic analyses show no topological congruence between Lepidoptera and the BV
385 species tree, but do show significant cophylogenetic signal between certain internal nodes of the BV
386 species phylogeny and the associated insect host species nodes (Figure 2). This means that present
387 host-use patterns in BVs result mainly from a pattern of host conservatism punctuated with
388 occasional shifting among pre-existing insect lineages [56,57]. This coevolutionary association
389 concurs with a general process of colonization by host tracking [58,59] as previously suggested at the
390 macroevolutionary level of several insect DNA virus families [18]. However, this type of
391 coevolutionary association is only observable for crown groups of viruses infecting hosts belonging
392 to the same family and is lost in more basal nodes. Indeed, over a short timeframe BV speciation
393 remains intimately linked to the speciation of their host, following the biogeography of their hosts
394 and ultimately in certain lineages we observed BV phylogenies that are the mirror images of their
395 host's. Yet on a larger evolutionary scale, the insect-host coevolutionary relationship signal is
396 confused, strongly suggesting that other factors act on BV evolution.

397 4.4. *Ecological Specialization*

398 The distinctive feature of the BV life cycle compared to most viruses is that they produce a
399 transmission stage, which persists outside of the host and has the ability to resist environmental
400 degradation. This feature is also found in other insect viruses such as entomopoxviruses and
401 cypoviruses, which have similar life cycles and OBs. This highlights the importance and the
402 persistence of this dissemination process in insect viruses [60]. As BVs only infect larvae and need to
403 be ingested to initiate infection, they have an intimate association with the plants that their hosts
404 feed on. In addition, there is an increasing body of evidence showing that host plant chemistry can
405 moderate the BV infection process [25,28]. Thus host plant characteristics could define the local
406 biotopes of BVs. We therefore searched for conservatism with the type of plant used by insect hosts,
407 distinguishing woody perennial (including shrubby and suffrutescent plants) and herbaceous
408 plants. Results show ancestral associations to particular plant groups over several tens of million
409 years (Figure 2B), suggesting a pattern of plant-use conservatism punctuated with sporadic shifts
410 between plant growths. This underlines the predominant role of host plant association in BV
411 evolution. As a consequence, BV diversification entangles patterns of host and local biotope
412 conservatism.

413 Virus ecological niches are considered in general as defined only by their hosts,
414 underestimating other factors and notably the influence of the environment. The BV niche combines
415 a set of insect and insect host plant biotic conditions. Viruses consume the resources provided by
416 their hosts in a host tracking fashion and consequently are influenced by their geographical
417 distributions; hosts are therefore the set of biotic conditions where primary speciation takes places
418 through adaptation to insect immunity and competition with other parasites. Host spectrum and
419 hosts shifts are contained within local environments represented by a group of host plants. Host

420 plant-use therefore defines a second ring of biotic environment that BVs experience. The association
421 with particular types of plants persists over millions of years and drives BVs towards particular
422 insect hosts. This is strikingly observable for the group II of the *Alphabaculovirus* genus, where BV
423 infecting Noctuoidea species are split in two distinct clades, in one clade (clade IIa) Noctuoidea
424 species are associated with herbaceous plants and in the other clade (clade IIb) they are associated
425 with woody plants (Figure 2).

426 At the level of the evolutionary history of BVs, our study points out the specialization of BVs
427 highlighted with topological congruence of certain BV-hosts associations. At the macroevolutionary
428 timescale, BVs are specialized to a particular insect order and notably to three orders of
429 holometabolous insects [15]. The BVs infecting Lepidoptera separate into two major groups with
430 different types of OBs, the NPVs and the GVs. GVs (*Betabaculovirus*) seem to be ancestrally
431 associated to herbaceous plants and to have colonized woody plants later. Strikingly, some GVs are
432 associated with internal feeding hosts such as the codling moth, *Cydia pomonella*, the potato tuber
433 moth, *Phthorimaea operculella* or the tea leaf-roller micromoth *Caloptilia theivora* and these types of
434 hosts are not infected by NPVs. The morphology of GV OBs, which are much smaller than those of
435 NPVs, may confer a more targeted dispersal strategy to increase the likelihood of reaching hosts
436 concealed within the plant. Therefore, it is predicted that leaf-mining and/ or stem-boring
437 Lepidoptera should more likely be infected by GVs than by NPVs.

438 NPVs (*Alphabaculovirus*) appear to be ancestrally associated to woody plants and to have
439 colonized herbaceous plants afterwards. NPVs belonging to group II are often very specialized to
440 their hosts according to topological congruence with their associated hosts and the clear split of
441 herbaceous (clade IIa) and woody (clade IIb) clades. By contrast, NPVs belonging to group I show a
442 complex pattern of associations to several lepidopteran superfamilies on herbaceous and woody
443 plants, suggesting more frequent host switches than those observed for group II or GVs.
444 Remarkably, this group includes the well known *Autographa californica* MNPV, which has a large
445 host range, spanning different Lepidoptera species belonging to distantly related families [61]
446 (Figure 1; Figure S2). This BV is a generalist virus, which is an uncommon trait in BVs. Generalism
447 could favor host switching and explain loss of BV-host phylogenetic congruence in certain BV
448 lineages, notably in the clade Ia of the *Alphabaculovirus* genus.

449 The host is by far the most important component of virus ecological niches, but not the only
450 one. Most previous studies focused on microevolutionary patterns of diversification, resulting in
451 host distribution outcomes to explain virus dynamics [62,63]. Here, we discuss the
452 macroevolutionary patterns of an entire virus family based on a reconstruction, as exhaustive as
453 possible, of its history. BVs have a peculiar life cycle where insect hosts and their associated plants
454 are entangled. Viral transmission and fitness are increased with the typical production of OBs for
455 environmental dissemination combined with the modification of host behavior, like the
456 enhancement of climbing behavior [64,65]. Plants are therefore the vessel of viral transmission.
457 Plants attacked by many lepidopteran species could support the evolution of generalist viruses. This
458 in turn could promote host shifts with the subsequent specialization of particular viral lineages.
459 Conversely plants attacked by few specialist Lepidoptera should foster the evolution of specialist
460 viruses. Multitrophic interactions have thus shaped BV evolution as shown by the combined
461 patterns of insect host and insect host plant conservatism, punctuated by occasional shifts among
462 pre-existing insect lineages. However, the direct evolutionary interactions between plants and BVs
463 remain undetermined.

464 5. Conclusion

465 Our current understanding of baculovirus diversity and evolution has been fuelled by decades
466 of research on biological control and biotechnological applications. Here we presented a complete
467 evolutionary framework of the known baculovirus diversity highlighting the complex ecological
468 interactions of these viruses with their hosts. Our study is the first to use a phylogenetic clustering
469 approach for species delimitation in viruses characterizing many new baculovirus species. Our
470 analyses show that host shifts played a major role in the diversification of baculoviruses. It also

471 shows that the colonization of a new ecological niche (herbaceous plants) lead to the radiation of
472 some baculovirus lineages. The species richness resulting from millions of years of evolutionary
473 interactions between the host plant ecology/chemistry and the physiology and ecology of both
474 viruses and their associated insect hosts potentially still hides a treasure trove of genes and
475 molecules that could lead to innovative biotechnology.

476 **Supplementary Materials:** The following are available online at www.mdpi.com/xxx/s1, Figure S1: Baculovirus
477 core-genome phylogeny, Figure S2. Raw baculovirus isolate phylogeny, Table S1. Baculovirus isolate sequence
478 database, Table S2. Host ecology database.

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