1 Article

2 Synthesis of a reactive oxygen species-responsive

doxorubicin derivative

- 4 James B. Delehanty¹, Shivani Das¹, Efram Goldberg², Ajmeeta Sangtani¹ and D. Andrew Knight^{2,*}
- Center for Bio/Molecular Science and Engineering, Code 6900, Naval Research Laboratory, Washington, DC
 20375, USA
 - ² Chemistry Department. Florida Institute of Technology, 150 West University Boulevard, Melbourne, FL
 - * Correspondence: aknight@fit.edu; Tel.: +01-321-674-8175

Abstract: A heterobifunctional reactive oxygen species (ROS)-responsive linker for directed drug assembly onto and delivery from a quantum dot (QD) nanoparticle carrier was synthesized and coupled to doxorubicin using EDC/sulfo-NHS coupling. The doxorubicin conjugate was characterized using ¹H NMR and LC-MS and subsequently reacted under conditions of ROS formation (Cu²⁺/H₂O₂) resulting in successful and rapid thioacetal oxidative cleavage which was monitored using ¹H NMR. The deprotected amine linker is amenable to peptide or protein conjugation prior to QD assembly or to direct conjugation to cognate reactive groups on ligands that cap the QD surface.

Keywords: doxorubicin; heterobifunctional; linker.

1. Introduction

The use of nanoparticles (NPs) for the controlled delivery of drugs to cells and tissues has emerged as an attractive alternative to traditional systemic delivery of drugs [1]. This is driven largely by the advantages afforded by NP-based drug delivery which include (1) large drug payload per dose, (2) longer circulation time and clearance, and (3) specific drug targeting to diseased tissues. One of the more attractive aspects of NP-mediated drug delivery is the potential to control the spatial and temporal release of the NP-appended drug in response to specific triggering stimuli [2]. Semiconductor quantum dots (QDs) are an excellent prototypical NP platform for the delivery of therapeutics given their bright, photostable luminescence (for tracking) and amenability to decoration with drug cargos (for delivery) [3]. Doxorubicin is a potent and widely-used anticancer therapeutic, yet it elicits significant nonspecific cardiotoxicity when its delivery/release is not properly targeted or controlled [4]. Thus, there is wide interest in modulating its cellular targeting and release.

Here we have synthesized a heterobifunctional, cleavable linker for the controlled release of doxorubicin from the surface of a QD carrier. The conjugate bears a reactive oxygen species (ROS)-responsive linkage with doxorubicin at one end and a protected amine at the other end. We demonstrate the ability of the ROS- reactive doxorubicin-linker to actively release the drug moiety in the presence of copper-catalyzed hydrogen peroxide-generated ROS. Acid cleavage of the trifluoracetate protecting group yields a reactive amine that can be conjugated (1) to a peptide backbone prior to peptide self-assembly to the QD surface or (2) directly to the QD surface *via* attachment to reactive "handles" on the termini of QD-capping ligands (*e.g.*, to the carboxyl groups on dihydrolipoic acid (DHLA) ligands (Fig. 1) . When appended onto the surface of a QD, the ROS-reactive conjugate described herein can be integrated into a multifunctional phototriggered ensemble where the light-harvesting central QD excites an appended photosensitizer molecule to generate ROS leading to the combined, multistage generation of ROS coupled with doxorubicin release.

2 of 5

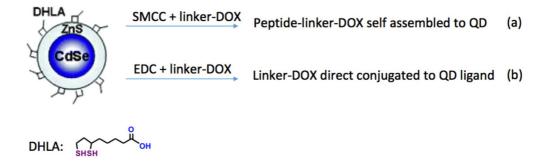


Figure 1. The linker-doxorubicin (DOX) conjugate can be conjugated (a) to a peptide prior to the assembly of the peptide onto the QD surface or (b) directly to a QD-capping ligand such as dihydrolipoic acid (DHLA).

2. Results and Discussion

47 48 49

50

51

52

53

54

55

56

57

58

59

60

61

62

63

64

65

66

67

68

69

70

71

72

73

74

75

76

77

78

79

80

81

The method we chose for the preparation of a ROS responsive doxorubicin conjugate is shown in Figure 2a, and required the synthesis of a doubly protected heterobifunctional linker 3 containing a dithioacetal group which would undergo cleavage when exposed to reactive oxygen species. Protection of the carboxylic acid group of the linker was achieved using a methyl ester and the amine functionality was protected using a trifluoroacetate group. This strategy has been described previously by Ling et al.⁵ Reaction of cysteamine hydrochloride and ethyl trifluoroacetamine under basic conditions gave 2,2,2-trifluoro-N-(2-mercaptoethyl)acetamide 1 in 86 % yield. modification of the reaction conditions gave a modest improvement of yield described previously in the literature (65 %).⁶ The doubly protected *S*, *S*-thioacetal **2** was then synthesized via the reaction of 1 with methyl 3-mercaptiopropioante under Lewis acidic conditions using boron trifluoride etherate. Efficient deprotection of 2 using pig liver esterase in PBS buffer gave the amine protected thioketal 3 with a free COOH group which would allow conjugation to amines via amide coupling. A number of amide coupling methods have previously been reported for attaching doxorubicin to carboxylic acids. For example, dicyclohexylcarbodiimide (DCC) was used to attach doxorubicin to poly(amidoamine) dendrimer encapsulated gold nanoparticles.⁷ Monosubstituted fatty acyl doxorubicin conjugates were prepared via reaction of doxorubicin with fatty acids in the presence of the coupling agent HBTU.8 We chose to use EDC/sulfo-NHS coupling to attach doxorubicin to the ROS-responsive linker 3 as shown in Figure 2b. A ten-fold excess of the linker was used for the reaction with doxorubicin performed in DMF at room tempature. After work-up involving extraction of the conjugate into CH2Cl2, and purification over silica gel, compound 4 was isolated in 50 % yield. Characterization of 4 was achieved using ¹H NMR and mass spectrometry (see Supplementary Materials, Figures S1-2), however our attempts to obtain a ¹³C NMR spectrum with suitable signal/noise were unsuccessful.

To demonstrate the response of the conjugate to reactive oxygen species which would result in cleavage and release of doxorubicin, an in situ ¹H NMR experiment was performed in which reactive ROS were catalytically generated using copper(II) ion and hydrogen peroxide in CD₃OD as described by Ling et al.⁵ The cupric ion-hydrogen peroxide reaction follows typical Fenton behavior, generating superoxide radicals, molecular oxygen and hydroxyl radicals according to equations (i)-(iii)⁹:

3 of 5

$$HS \longrightarrow NH_2HCI + MeO \longrightarrow CF_3$$

$$\downarrow (i)$$

$$HS \longrightarrow NH_2HCI + MeO \longrightarrow CF_3$$

$$\downarrow (iii)$$

$$MeO \longrightarrow S \longrightarrow NH \longrightarrow CF_3$$

$$\downarrow (iii)$$

$$Me \longrightarrow Me \longrightarrow Me \longrightarrow H \longrightarrow CF_3$$

$$\downarrow (iii)$$

$$HO \longrightarrow S \longrightarrow NH \longrightarrow CF_3$$

$$\downarrow (iii)$$

$$HO \longrightarrow Me \longrightarrow Me \longrightarrow H \longrightarrow CF_3$$

$$\downarrow (iii)$$

$$Me \longrightarrow Me \longrightarrow H \longrightarrow CF_3$$

$$\downarrow (iii)$$

$$Me \longrightarrow Me \longrightarrow H \longrightarrow CF_3$$

$$\downarrow (iii)$$

$$Me \longrightarrow Me \longrightarrow H \longrightarrow CF_3$$

$$\downarrow (iii)$$

$$Me \longrightarrow Me \longrightarrow H \longrightarrow CF_3$$

$$\downarrow (iii)$$

$$Me \longrightarrow Me \longrightarrow H \longrightarrow CF_3$$

$$\downarrow (iii)$$

$$Me \longrightarrow Me \longrightarrow H \longrightarrow CF_3$$

$$\downarrow (iii)$$

$$Me \longrightarrow Me \longrightarrow H \longrightarrow CF_3$$

$$\downarrow (iii)$$

$$Me \longrightarrow Me \longrightarrow H \longrightarrow CF_3$$

$$\downarrow (iii)$$

$$Me \longrightarrow Me \longrightarrow H \longrightarrow CF_3$$

$$\downarrow (iiii)$$

$$Me \longrightarrow Me \longrightarrow H \longrightarrow CF_3$$

$$\downarrow (iiii)$$

$$Me \longrightarrow Me \longrightarrow H \longrightarrow CF_3$$

$$\downarrow (iiii)$$

$$Me \longrightarrow Me \longrightarrow H \longrightarrow CF_3$$

$$\downarrow (iiii)$$

$$Me \longrightarrow Me \longrightarrow H \longrightarrow CF_3$$

$$\downarrow (iiii)$$

$$Me \longrightarrow Me \longrightarrow H \longrightarrow CF_3$$

$$\downarrow (iiii)$$

$$Me \longrightarrow Me \longrightarrow H \longrightarrow CF_3$$

$$\downarrow (iiii)$$

$$Me \longrightarrow Me \longrightarrow H \longrightarrow CF_3$$

$$\downarrow (iiii)$$

$$Me \longrightarrow Me \longrightarrow H \longrightarrow CF_3$$

$$\downarrow (iiii)$$

$$Me \longrightarrow Me \longrightarrow H \longrightarrow CF_3$$

$$\downarrow (iiii)$$

$$Me \longrightarrow Me \longrightarrow H \longrightarrow CF_3$$

$$\downarrow (iiii)$$

$$Me \longrightarrow Me \longrightarrow H \longrightarrow CF_3$$

$$\downarrow (iiii)$$

$$Me \longrightarrow Me \longrightarrow H \longrightarrow CF_3$$

$$\downarrow (iiii)$$

$$Me \longrightarrow Me \longrightarrow H \longrightarrow CF_3$$

$$\downarrow (iiii)$$

$$Me \longrightarrow Me \longrightarrow H \longrightarrow CF_3$$

$$\downarrow (iiii)$$

$$Me \longrightarrow Me \longrightarrow H \longrightarrow CF_3$$

$$\downarrow (iiii)$$

$$Me \longrightarrow Me \longrightarrow H \longrightarrow CF_3$$

$$\downarrow (iiii)$$

$$Me \longrightarrow Me \longrightarrow H \longrightarrow CF_3$$

$$\downarrow (iiii)$$

$$Me \longrightarrow Me \longrightarrow H \longrightarrow CF_3$$

$$\downarrow (iiii)$$

$$Me \longrightarrow Me \longrightarrow H \longrightarrow CF_3$$

$$\downarrow (iiii)$$

$$Me \longrightarrow Me \longrightarrow H \longrightarrow CF_3$$

$$\downarrow (iiii)$$

$$Me \longrightarrow Me \longrightarrow H \longrightarrow CF_3$$

$$\downarrow (iiii)$$

$$Me \longrightarrow Me \longrightarrow H \longrightarrow CF_3$$

$$\downarrow (iiii)$$

$$Me \longrightarrow Me \longrightarrow H \longrightarrow CF_3$$

$$\downarrow (iiii)$$

$$Me \longrightarrow Me \longrightarrow H \longrightarrow CF_3$$

$$\downarrow (iiii)$$

$$Me \longrightarrow Me \longrightarrow H \longrightarrow CF_3$$

$$\downarrow (iiii)$$

$$Me \longrightarrow Me \longrightarrow H \longrightarrow CF_3$$

$$\downarrow (iiii)$$

$$Me \longrightarrow Me \longrightarrow H \longrightarrow CF_3$$

$$\downarrow (iiii)$$

$$Me \longrightarrow Me \longrightarrow H \longrightarrow CF_3$$

$$\downarrow (iiii)$$

$$\downarrow (iiiii)$$

$$\downarrow (iiiii)$$

$$\downarrow (iiiii)$$

$$\downarrow (iiiii)$$

$$\downarrow (iiii)$$

$$\downarrow (iiiii)$$

$$\downarrow (iiiii)$$

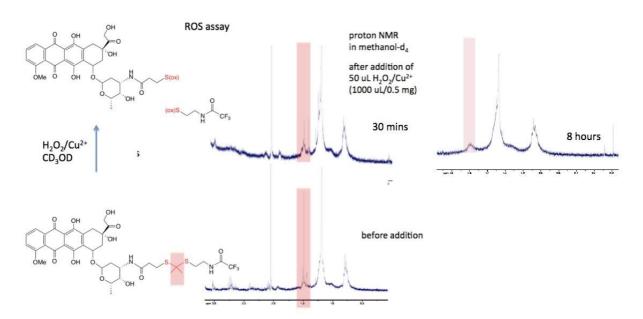
$$\downarrow (iiiii)$$

$$\downarrow ($$

Figure 2. (a) Synthesis of heterobifunctional linker 3. (i) Et₃N, MeOH; (ii) acteone, BF₃•Et₂O, CH₃CN, 0 °C; (iii) porcine liver esterase, acetone, PBS. (b) Synthesis of ROS responsive doxorubicin conjugate 4.

Briefly, a solution of doxorubicin conjugate 4 in CD₃OD was placed in an NMR tube, and a spectrum was recorded which showed the presence of methyl protons at 1.5 ppm due to the dimethyl thioacetal group. On addition of hydrogen peroxide and a catalytic amount of copper chloride, a second ¹H NMR spectrum was recorded after 30 mins. Reduction in intensity of methyl protons indicated successful cleavage of the thioacetyl group (from peak integration, 80% reduction in total amount of CH₃ protons). After 8 hours a final NMR spectrum showed complete disappearance of the dimethyl thioacetal group (Figure 3).

Figure 3. Reactive oxygen species assay: ¹H NMR spectra showing disappearance of SC(CH₃)₂S methyl protons (pink bars).



*peak broadness due to paramagnetic Cu2+

4 of 5

3. Materials and Methods

All reactions were conducted in oven-dried glassware, under air unless otherwise noted. ¹H NMR spectra were recorded on Avance Bruker 400-MHz spectrometers. Mass spectra were recorded on an Agilent LC-Mass Spectrometer in ESI or APCI modes. Chemical shifts for ¹H NMR spectra are reported (in parts per million) relative the residual solvent. Silica gel (200-300 mesh) was used for column chromatography. TLC plates were visualized by exposure to iodine vapor.

Solvents were obtained as follows: dimethylformamide (DMF), dichloromethane, methanol, acetone, acetonitrile (Aldrich) used as received. CDCl₃, CD₂Cl₂, D₂O, CD₃OD (Cambridge Isotopes Laboratory or Aldrich) used as received.

Reagents and chemicals were obtained as follows: Doxorubicin hydrochloride (Aldrich), N-(3-dimethylaminopropyl)-N'-ethylcarbodiimide hydrochloride (Pierce) , N-hydroxysulfosuccinimide sodium salt (Pierce), copper(II) chloride (Aldrich), trimethylsilylpropyl sulfonate (Aldrich), 30% hydrogen peroxide (Aldrich), silica gel (230-400 mesh, Fisher), anhydrous sodium sulfate (Aldrich) used as received. Heterobifunctional dithioacetal $\bf 3$ was prepared according to the literature procedure.

Conjugation of heterobifunctional dithioacetal 3 with doxorubicin: Compound 3 (5.52 mg, (1.00)17.3 doxorubicin hydrochloride 1.73 N-(3-dimethylaminopropyl)-N'-ethylcarbodiimide hydrochloride (EDC) (0.330 mg, 1.73 mol) and N-hydroxysulfosuccinimide sodium salt (sulfo-NHS) (0.380 mg, 1.730 mol) were combined in DMF (2.0 mL) and stirred at room temperature for 8 hours. Then, DI H₂O (1 mL) was added and the aqueous phase was extracted with CH₂Cl₂ (5 x 1 mL). The CH₂Cl₂ extracts were combined and dried over anhydrous Na₂SO₄. The solvent was removed in vacuo and the resulting red residue was dissolved in CH₂Cl₂/MeOH (2 mL, 50/50 v/v) and purified by passing over a silica gel plug (ca. 1 g) and washing the plug with a further portion of CH₂Cl₂/MeOH to give a red-orange solution. The solvent was removed to give 4 as a red solid (0.73 mg, 50% yield). A ¹H NMR spectrum recorded in CDCl₃ is included in the Supplementary Materials. ¹H NMR : 0.80-1.21 (m, 6H), 1.61 (s, 6H), 2.01-2.97 (m, 8H), 3.54-3.82 (m, 11H), 5.23 (m, 2H), 6.81 (s, br, 1H, NH).

Oxidation of 4 using copper(II)-hydrogen peroxide: A 5 mm NMR tube was charged with conjugate **4** (9.45 mg) and CD₃OD (ca. 0.7 mL). A trace of trimethylsilylpropyl sulfonate was added as an internal reference and a ¹H NMR spectrum was immediately recorded. Then, a solution of CuCl₂ in 30% H₂O₂ (15 L, 200 mM) was added and after 30 mins a second ¹H NMR spectrum was recorded. Attempts to isolated oxidative products using TLC were unsuccessful.

4. Conclusions

In summary, we have synthesized a new doxorubicin-conjugated heterobifunctional ROS-responsive linker molecule using EDC/sulfo-NHS amide bond coupling in which a carboxylic acid functional group is coupled to the native primary amine of the anti-cancer drug doxorubicin. The amine group of the linker is protected by an acid labile trifluoracetate moiety which can be readily removed for further conjugation (e.g., to peptides, proteins). The presence of a ROS-responsive dithioacetal group as part of the linker allows the rapid release of doxorubicin to a target when exposed to ROS generated *in situ*. Using ¹H NMR, we have demonstrated that the linker is cleaved readily when exposed to ROS generated from Cu²⁺/H₂O₂.

- Supplementary Materials: The following are available online at www.mdpi.com/xxx/s1, Figure S1: title, Table S1: title, Video S1: title.
- Funding: This work received support from the Base Funding Program at the Naval Research Laboratory.

 D.A.K. is grateful to the Office of Naval Research for a Summer Faculty Fellowship.
- 151 Conflicts of Interest: The authors declare no conflict of interest. The funders had no role in the design of the study; in the collection, analyses, or interpretation of data; in the writing of the manuscript, and in the decision to publish the results.

Peer-reviewed version available at Molecules 2018, 23, 1809; doi:10.3390/molecules23071809

5 of 5

155 References

156

- 1. Swain S.; Sahu, P.K.; Beg, S.; Babu, S. M. Nanoparticles for cancer targeting: Current and future directions. *Curr. Drug Deliv.* **2016**, *13*, 1290-1302.
- Sangtani, A.; Nag, O.K.; Field, L.D.; Breger, J.C.; Delehanty, J.B. Multifunctional nanoparticle composites: progress in the use of soft and hard nanoparticles for drug delivery and imaging. *Wiley Interdiscip. Rev. Nanomed. Nanobiotechnol.* 2017, 9, 1-23.
- 162 3. Delehanty. J.B.; Boeneman, K.; Bradburne, C.E.; Robertson, K.; Medintz, I.L. Quantum dots: a powerful tool for understanding the intricacies of nanoparticle-mediated drug delivery. *Expert. Opin. Drug. Deliv.* 2009, 6, 1091-1112.
- 165 4. Danz, E.D.B.; Skramsted, J.; Henry, N.; Bennett, J.A.; Keller, R.S. Resveratrol prevents doxorubicin cardiotoxicity through mitochondrial stabilization and the Sirt1 pathway. *Free Radical. Bio. Med.* 2009, 46, 1589-1597.
- Ling, X.; Zhang, S.; Shao, P.; Wang, P.; Ma, X.; Bai, M. Synthesis of a reactive oxygen species responsive heterobifunctional thioketal linker. *Tetrahedron Lett.* **2015**, *56*, 5242–5244.
- 170 6. Shim, M, S.; Xia, Y. A reactive oxygen species (ROS)-responsive polymer for safe, efficient, and targeted gene delivery in cancer cells. *Angew. Chem, Int. Ed.* **2013**, *52*, 6926–6929.
- 7. Sagita, E.; Djajadisastra, J.; Mutalib, A. Cytotoxic enhancement of doxorubicin in conjugation with PAMAM G4.5 dendrimer containing gold nanoparticles. *Int. J. PharmTech. Res.* **2016**, *9*, 348-356.
- 174 8. Chhikara, B.S.; St. Jean, N.; Mandal, D.; Kumar, A.; Parang, K. Fatty acyl amide derivatives of doxorubicin: Synthesis and *in vitroi* anticancer activities. *Eur. J. Med. Chem.* **2011**, *46*, 2037-2042.
- 9. Carvalho do Lago, L.C.; Matias, A.C.; Nomura, C.S.; Cerchiaro, G. J. Inorg. Biochem. 2011, 105, 189–194.