

1 *Review*

## 2 **Potentiometric Biosensing Applications of Graphene** 3 **Electrode with Stabilized Polymer Lipid Membranes**

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13 **Abstract:** This review provides informations and details for the fabrication of biosensors that are  
14 composed from lipid membranes and have been utilized and applied to rapidly detect food toxic  
15 compounds, environmental pollutants and analytes of clinical interest. Biosensors based on  
16 polymeric lipid membranes have been used to rapidly detect a wide range of these analytes and  
17 offer several advantages such as fast response, high sensitivity and selectivity, can be portable for  
18 in the field applications, and small size. A description of the construction of these devices and their  
19 applications for the rapid detection of food toxic substance, environmental pollutants and analytes  
20 of clinical interest is provided in this review.

21 **Keywords:** Biosensors; lipid membranes; potentiometry; graphene electrodes  
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### 23 **1. Introduction**

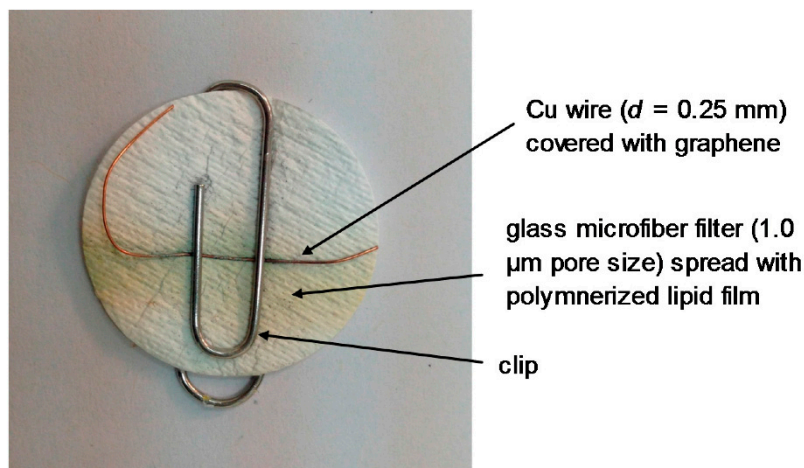
24 A biosensor is an instrument that analyses a sample and provides the chemical concentration  
25 and composition of the sample. A chemical sensor consists by two parts: A biological element that  
26 chemically recognizes the unknown compound and provides this chemical information in a physical  
27 unit that transforms the signal into a measurable electrical, optical, or piezoelectric signal.(eg. voltage,  
28 current, absorbance, etc).

29 The advantages of biosensors as compared to the classical analytical devices (eg. liquid and gas  
30 chromatography, etc) areas follows: faster response times, much higher throughput of samples,  
31 smaller size and can be used for in the field measurements. It is also cheaper and does not require  
32 training of personnel.

33 Chemosensors are similar to biosensors but are composed from a synthetically prepared  
34 molecule instead of a biological element. They recognize small molecules or metal ions by binding to  
35 them. The synthesis, design, and applications of chemosensors are of special interest for the detection  
36 of these analytes. The present review provides applications of chemosensors that have not been well  
37 covered elsewhere, for example the design and synthesis of calixarenes that have been used to rapidly  
38 determine carbamates [1] and **naphthalene acetic acid (NAA)** [2].

39 Nanobiosensors are based on the merging of nanotechnology with biosensors. Such materials  
40 include graphene, carbon nanotubes and nanowires, etc. Metal based nanoparticles are also excellent  
41 hosts for the construction of electrical and optical devices that can be applied to detect nucleic acid  
42 sequences. Various nanomaterials have been explored and their properties were analyzed for  
43 possible applications in biosensors. The research in nanobiosensor technology has prompted the  
44 construction of novel devices and their number increases in an exponential rate.

45 The present review paper deals with the fabrication of biosensing devices that are composed  
46 from lipid membranes on graphene electrodes and have been utilized and applied to rapidly detect  
47 food toxic substances, environmental pollutants and analytes of clinical interest.. These devices are  
48 composed from a polymerized lipid films on graphene electrodes. The construction of these devices  
49 and their applications in the rapid detection of the above analytes are described in the present paper.  
50 Figure 1 provides a schematic of a lipid based biosensor on a graphene electrode that has been used  
51 for the potentiometric detection of urea



52

53 **Figure 1.** Schematic of a lipid membrane based biosensor on graphene electrode. This device was used  
54 for the potentiometric determination of urea (reprinted from reference 3 with permission).

## 55 2. Polymer lipid membranes

56 Stabilized in electrolyte solution lipid film based biosensors were first constructed by Nikolelis  
57 group [4] by using glass fiber filters to support and stabilize the lipid membrane. However, these  
58 membranes were not stable outside an electrolyte solution in the air. Later on, lipid polymer  
59 membranes supported on glass fiber were shown to be stable in air [5]. Recently, the deposition of  
60 these polymer lipid films on graphene electrode [3] has shown to offer many advantages such as  
61 increased sensitivity and selectivity, rapid response times and provided biosensors for urea for  
62 remote sensing.

63 Biosensors have been developed during the last 40 years and a pioneer in this field of science  
64 was Antony Turner, Prof. Emeritus at Linköping University. Lipid membrane biosensors appeared  
65 in 1990 with the pioneer work of Ulrich J. Krull and Dimitrios P. Nikolelis in University of Toronto  
66 and Athens; Prof. Tibor Hianik also pioneer the work of lipid membrane based biosensors focused  
67 mainly on metal supported lipid films. Recently nanotechnology has offered a route to prepare  
68 nanosensors based on lipid membranes by using graphene electrodes.

69 Biosensors that are based on lipid membranes offer an alternative to the standard analytical  
70 methods (i.e., chromatography or mass spectroscopy) such as rapid response times, small size, are  
71 not time consuming, they have a low cost, they can be portable for uses in the field and most  
72 importantly they are biocompatible. Recent advances in the construction of stabilized in the air lipid  
73 films has allowed their applications for the detection of a wide range of compounds that are toxic in  
74 foods or are of environmental or clinical interest.

75 The design, construction and storage of polymer lipid membranes on microporous filtering  
76 media such as glass fibers has been described extensively in one of our previous papers [5]. This  
77 paper has examined the simplicity for membrane construction and how stable are these lipid  
78 membranes after they are stored in the air so that they can be used a large number of times specially  
79 in a repetitive way. The preparation technique of the polymer lipid membranes has a standard route  
80 and is as follows, 5 mg of phosphatidyl choline (PC) is stirred in a small vial with 0.070 ml of  
81 methacrylic acid, 0.8 ml of ethylene glycol dimethacrylate, 8 mg of 2,2'-azobis-(2-methylpropionitrile)

82 and 1.0 ml of acetonitrile. The roles of methacrylic acid, ethylene glycol dimethacrylate and 2,2'-  
83 azobis-(2-methylpropionitrile) (AIBN) were explained in one of our previous paper [5]. Nitrogen  
84 passed through this mixture by sparging for ca. 1 min, then was sonicated for ½ hour and stored in  
85 the refrigerator. In order to prepare stabilized polymer lipid membranes, 0.15 ml of this mixture was  
86 injected on the surface of a microfiber filter and was then irradiated with a UV deuterium lamp for 4  
87 hours. This polymer lipid film can directly be used to construct devices based on lipid membranes.

88 These stabilized polymeric lipid membranes based biosensors can be used as excellent host  
89 matrices for to maintain the activity of a biological compound and used for the transduction of the  
90 activity of a wide number of biological active compounds such as enzymes, antibodies and artificial  
91 or natural receptors [5]. These devices are stable in air for more than one month and provide a  
92 response to a wide range of compound/ analytes [6-8]. The advantages of polymerization by  
93 irradiation by using a UV deuterium lamp instead of heating at 60 °C for 4 hours has been described  
94 in one of our previous papers [9].

95 In this paper [9], the UV irradiation was used instead of heating the mixture at 60 °C because  
96 this process retains the activity of the enzyme, antibody or receptor, heating may deactivate these  
97 biological "receptors". This method to prepare stable biosensors based on lipid membranes by  
98 heating at 60 °C was empirical; on the other hand, the mechanism of polymerization was never  
99 investigated. In this paper [9], the preparation technique of stable lipid films was explored by using  
100 physicochemical methods i.e. Raman and IR spectrophotometry, differential scanning calorimetry  
101 (DSC) and scanning electron microscopy (SEM) experiments. The results of Raman and IR have  
102 shown that the polymerization kinetics finishes after 4 h. The polymerization mechanism and  
103 generation of the signal was further on explored by Raman and IR spectroscopy, micro-Raman  
104 spectroscopy, DSC and SEM experiments [2,3,9,10]. These membranes were stable for storage in air  
105 for a period of longer than two months.

106 Through this route the practical use of biosensors based on lipid membranes was possible,  
107 because the technique has allowed incorporation of natural ion-channels in these devices and  
108 eventually will permit these devices to be commercialized.

109 This procedure by irradiation with a UV light has shown to retain the activity of  
110 acetylcholinesterase for the detection of carbamates [9]; heating at 60 °C may would deactivate the  
111 activity. A very important criteria to for practical applications of lipid membrane based biosensors is  
112 the ability to store these membranes at temperatures of 25 °C for long period of time. A number of  
113 practical characteristics of these lipid membrane based biosensors such as stability at ambient  
114 temperatures, reproducibility and reusability have been shown in their analytical applications in our  
115 recent papers [9-12].

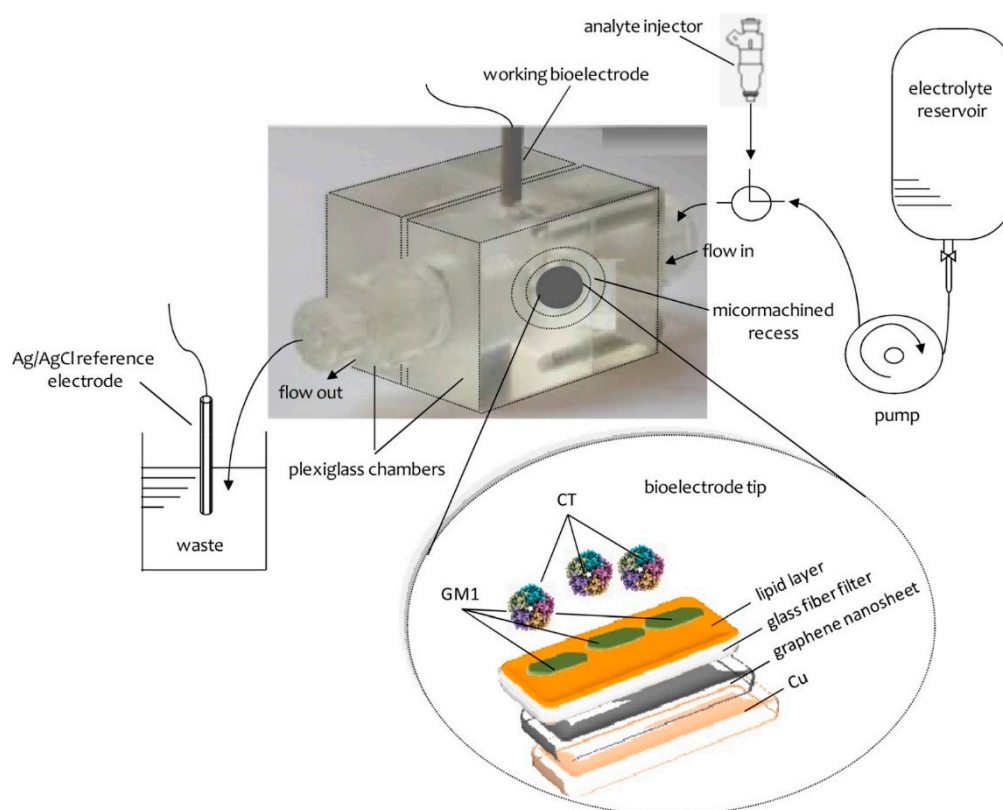
### 116 3. Polymer Lipid membranes on Graphene electrode

117 We recently have reported a method on how to construct devices based on polymeric lipid films  
118 incorporated in a graphene electrode [3]. This technique is as follows: a homogeneous dispersion (ca.  
119 0.4 mg/mL) of graphene in a solvent N-methyl-pyrrolidone (NMP) was sonicated for ca. 180 h and  
120 then was centrifugated at 700 rpm for 2 h [21]. the prolong sonication time is requisite in order to  
121 reduce the size of the flakes which is required in many applications.

122 At this stage, this suspension was deposited on a copper wire (with d=0.25 mm) which was  
123 placed on a glass fiber filter and the organic solvent was evaporated by a fan heater. The use of the  
124 wire was to help the connection in order to provide the voltage as the output of the signal.

125 The "receptor" molecules were incorporated in the lipid films at the stage of polymerization by  
126 spreading 10 mL of a "receptor" suspension onto the polymerization mixture (for instance, in order  
127 to prepare devices for the detection of cholera toxin, 10 mL of the ganglioside (which acts as a receptor  
128 in this case) suspension was mixed with 0.15 mL of the polymerization mixture using a microsyringe  
129 at the surface of the microfiber filter. The graphene electrodes with incorporated lipid membranes  
130 are stored at 4 °C when not in use for periods of more than 3 months..

131 The construction of the potentiometric biosensor concluded after the encapsulation of the filter-  
 132 supported polymer lipid membrane onto the copper wire which contained the graphene nanosheets  
 133 (Figure 2).



134  
 135 **Figure 2.** A simplified scheme of the experimental device and the bioelectrode edge surface  
 136 .(Reprinted from ref. 11).

#### 137 4. Potentiometric applications on biosensors based on lipid membranes on graphene electrodes

138 An article was written in the literature which describes a potentiometric urea lipid membrane  
 139 based nanosensor incorporated on graphene [3]. The investigation of the structure of graphene  
 140 nanosheets was made using atomic force microscopy (AFM) and transmission electron microscopy  
 141 (TEM) measurements. UV-Vis and Fourier transform IR (FTIR) spectrophotometry were used to  
 142 examine the pre- and postconjugated surfaces of graphene nanosheets. This potentiometric urea  
 143 nanosensor had a good reproducibility, reusability, selectivity, fast response times (~4 s), long shelf  
 144 life and high sensitivity having a slope of ca. 70 mV/decade in the concentration range of urea  
 145 between  $1 \times 10^{-6}$  M to  $1 \times 10^{-3}$  M.

146 A cholesterol potentiometric nanosensor has also appeared in the literature by fabrication using  
 147 the immobilization techniques of stable polymer lipid film which was deposited on graphene [12].  
 148 This nanosensor used the enzyme cholesterol oxidase to retain a response towards cholesterol. This  
 149 device has shown a good reproducibility, high selectivity and excellent sensing capability having a  
 150 linear slope curve of ~64 mV per decade. Given that there is a biocompatibility between lipid films  
 151 and human biofluids (eg., serum and urine) this gives the possibility to use this device for human  
 152 blood samples and other biological fluids.

153 An article that provides the construction of a potentiometric D-dimer nanosensor on graphene  
 154 using polymer lipid membranes has appeared in the literature [13]. This graphene biosensor was  
 155 applied to construct a selective and sensitive immunosensor for the determination of D-dimer; this  
 156 was accomplished by using as “receptor” the mouse anti human D-dimer antibody which was  
 157 immobilized on stabilized polymer lipid membranes on graphene bioelectrode. The range of  
 158 response towards D-dimer concentrations was between  $10^{-6}$   $\mu\text{g/L}$  to  $10^{-3}$   $\mu\text{g/L}$  and the response times  
 159 were ca. 15 s. This potentiometric D-dimer device can be constructed very easily and has a good



160 reproducibility, reusability, selectivity, rapid response times, long shelf life and high sensitivity with  
161 a linear logarithmic slope of ca. 59 mV/decade in the D-dimer concentration range from  $10^{-6}$   $\mu\text{g/L}$  to  
162  $10^{-3}$   $\mu\text{g/L}$ .

163 A carbofuran chemosensor based on graphene electrode using lipid membranes has appeared  
164 in the literature for the determination of carbofuran in real samples of fruits and vegetables [1]. The  
165 graphene electrode was used as a basis to developing a chemosensor to determine carbofuran very  
166 sensitive and selectively using the immobilization technique of an artificial receptor deposited onto  
167 stable lipid membranes. The method of preparation of this receptor was described in this paper [1]  
168 and the technique included altering the hydroxyl groups of resorcin[4]arene in phosphoryl groups.  
169 The chemosensor has a response the mM concentration range of carbofuran and has rapid response  
170 times (ca. 20 s). This potentiometric carbofuran chemosensor is constructed easily and is very  
171 reproducible and sensitive; its response times are very short (20 s), exhibits, a long period of stability  
172 having logarithmic concentration response with a slope of. 59 mV/decade in the range between  $10^{-6}$   
173 to  $10^{-3}$  M.

174 A potentiometric miniature cholera toxin device which was based on a graphene electrode with  
175 deposited lipid membranes has appeared in the literature [14]. Ganglioside GM1 (which is the natural  
176 receptor for cholera toxin) was mixed with the lipid mixture prior to polymerization. This device had  
177 a good selectivity and sensitivity with nanomolar detection limits and rapid response times (ca. 5  
178 min). The nanosensor was easily constructed and shown excellent characteristics (i.e., reproducibility,  
179 reusability, selectivity, long shelf life). The slope was ca. 60 mV/decade over a logarithmic cholera  
180 toxin concentration. The method applied for the determination of this toxin in water samples of lakes.

181 A naphthalene acetic acid (NAA) potentiometric device using the described graphene electrode  
182 in which the lipid membrane transducer was deposited has been described in the literature [2]. The  
183 receptor (auxin-binding protein 1) was placed in the lipid mixture prior to polymerization. The  
184 selectivity and sensitivity of response were very good, the detection limits were in the mM range and  
185 the response times were ca. 5 minutes. The device can be constructed very easily and shows excellent  
186 reproducible results and can be reused many times. The shelf life was very long and the slope of the  
187 potentiometric nanosensor was 56 mV/decade of NAA concentration. An evaluation/ validation of  
188 this device was made by using spiked fruits and vegetables.

189 A saxitoxin nanosensor based on graphene electrodes with deposited lipid membrane and in  
190 mixture with anti-STX (the natural saxitoxin receptor) was described in the literature [15]. Saxitoxin  
191 was selectively determined in concentrations between of  $1 \times 10^{-9}$  M to  $1 \times 10^{-6}$  M, the response times  
192 were between. 5–20 min, and the detection limit was 1 nM. This nanosensor can be constructed very  
193 easily and shows a good reproducibility and selectivity. This device can be used many times, it shows  
194 a stability at ambient temperatures and the shelf life is long. The slope of the electrode is. 60  
195 mV/decade of saxitoxin. The method was applied in real samples (i.e., validated in lake waters and  
196 shellfish samples). The present technology can be adapted to detecting other toxins and can be a  
197 weapon against bioterrorism.

## 198 5. Conclusions

199 This review article deals with construction of biosensors based on polymer lipid membranes on  
200 graphene electrode and their potentiometric applications. These applications include the rapid  
201 detection of a wide range of toxic substances in foods, environmental pollutants and compounds of  
202 biomedical attention. These biosensors have shown adequate selectivity, sensitivity, reproducibility,  
203 rapid response times, they are easy to construct. The person who uses these devices can be non-skilled  
204 and these nanosensors can be used for in the field measurements. Research is now directed to the use  
205 of nanotechnological advances to construct nanosensors with even more improved characteristics  
206 that could be portable and used for the rapid determination of these analytes.  
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