

Supplementary Information

Sterepinic acids A–C, new carboxylic acids produced by a marine alga-derived Fungus

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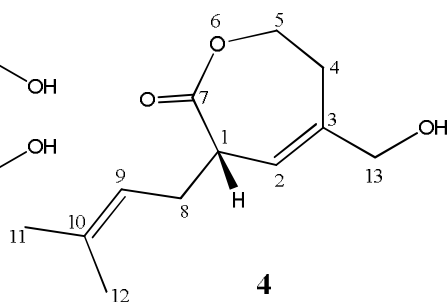
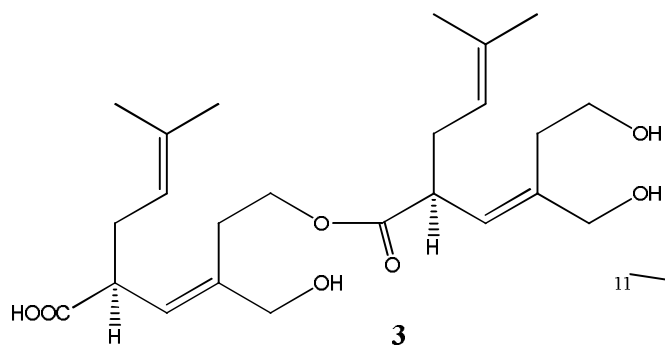
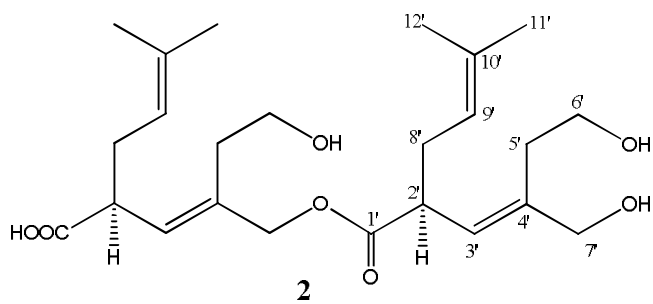
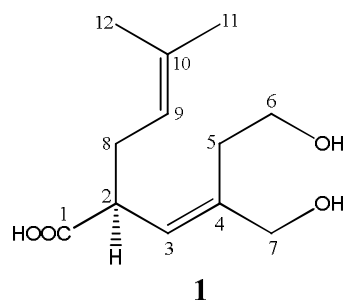


Table S1 NMR spectral data of **1** in CDCl₃

Position	$\delta_{\text{H}}^{\text{a}}$	J/Hz	^1H - ^1H COSY	NOESY ^b	δ_{C}	HMBC (C) ^c
1					177.5 (s)	
2	3.27 m		3, 8A, 8B	9, 11, 12	44.9 (d)	1, 3, 4, 8, 9
3	5.50 d	10.2 (2)	2	8A, 8B, 9, 11, 12	127.0 (d)	1, 2, 5, 7, 8
4					138.7 (s)	
5A	2.25 m		5B, 6		31.9 (t)	3, 4, 6, 7
5B	2.49 m		5A, 6			3, 4, 6, 7
6	3.68 br s		5A, 5B	11, 12	61.0 (t)	4
7	4.03 br s			3, 11, 12	66.8 (t)	3, 4, 5
8A	2.20 m		2, 8B, 9	3, 11, 12	30.9 (t)	1, 2, 3, 9, 10
8B	2.44 m		2, 8A, 9	3, 11, 12		1, 2, 3, 9, 10
9	5.04 dd	7.2 (8), 7.2 (8)	8A, 8B	2, 3, 11, 12	120.2 (d)	2, 8, 11, 12
10					134.1 (s)	
11	1.67 s			2, 3, 6, 7, 8A, 8B, 9	25.7 (q)	9, 10, 12
12	1.60 s			2, 3, 6, 7, 8A, 8B, 9	17.8 (q)	9, 10, 11

a ^1H chemical shift values (δ ppm from SiMe₄) followed by multiplicity and then the coupling constants (J /Hz). Figures in parentheses indicate the proton coupling with that position. b The correlations with geminal and vicinal protons are removed. c Long range ^1H - ^{13}C correlations from H to C observed in the HMBC experiment

Table S2 NMR spectral data of **2** in CDCl₃

Position	$\delta_{\text{H}}^{\text{a}}$		J/Hz	^1H - ^1H COSY	NOESY ^b	δ_{C}	HMBC (C) ^c
1						173.5 (s)	
2	3.28	m		3, 8A, 8B	5, 9	45.4 (d)	1, 3, 4, 8, 9
3	5.49	d	10.8 (2)	2	7A, 7B, 8A, 8B	129.3 (d)	2, 5, 7, 8
4						133.9 (s)	
5A	2.18	m		5B, 6A, 6B	2	32.3 (t)	3, 4, 6, 7
5B	2.54	m		5A, 6A, 6B	2		3, 4, 6, 7
6A	3.65	br s		5A, 5B, 6B		61.4 (t)	
6B	3.72	br s		5A, 5B, 6A			
7A	4.48	d	13.2 (7B)	7B	3	67.9 (t)	1, 3, 4, 5
7B	4.62	d	13.2 (7A)	7A	3		1, 3, 4, 5
8A	2.20	m		2, 8B, 9	3, 11, 12	30.8 ^{d1} (t)	1, 2, 3, 9, 10
8B	2.46	m		2, 8A, 9	3, 11, 12		1, 2, 3, 9, 10
9	5.03	m		8A, 8B	2, 11, 12	120.2 ^{d2} (d)	2, 8, 11, 12
10						134.2 ^{d3} (s)	
11	1.67	s			9	25.7 (q)	9, 10, 12
12	1.60	s			8A, 8B, 9	17.8 (q)	9, 10, 11
1'						173.5 (s)	
2'	3.28	m		3', 8'A, 8'B	5', 9'	45.4 (d)	1', 8', 9'
3'	5.52	d	10.2 (2')	2'	7', 8'A, 8'B	127.2 (d)	5', 7'
4'						139.6 (s)	
5'A	2.29	m		5'B, 6'	2'	32.3 (t)	3', 4', 6', 7'
5'B	2.54	m		5'A, 6'	2'		3', 4', 6', 7'
6'	3.72	br s		5'A, 5'B		60.5 (t)	
7'	4.05	br s			3'	67.5 (t)	3', 4', 5'
8'A	2.20	m		2', 8'B, 9'	3', 11', 12'	30.6 ^{d1} (t)	1', 2', 3', 9', 10'
8'B	2.46	m		2', 8'A, 9'	3', 11', 12'		1', 2', 3', 9', 10'
9'	5.03	m		8'A, 8'B	2', 11', 12'	120.3 ^{d2} (d)	2', 8', 11', 12'
10'						134.3 ^{d3} (s)	
11'	1.67	s			9'	25.7 (q)	9', 10', 12'
12'	1.60	s			8'A, 8'B, 9'	17.8 (q)	9', 10', 11'

a ^1H chemical shift values (d ppm from SiMe₄) followed by multiplicity and then the coupling constants (J/Hz). Figures in parentheses indicate the proton coupling with that position. b The correlations with geminal and vicinal protons are removed. c Long range ^1H - ^{13}C correlations from H to C observed in the HMBC experiment. d1-d3 interchangeable

Table S3 NMR spectral data of **3** in CDCl₃

Position	$\delta_{\text{H}}^{\text{a}}$		J/Hz	^1H - ^1H COSY	NOESY ^b	δ_{C}	HMBC (C) ^c
1						174.3 (s)	
2	3.28	m		3, 8A, 8B	5, 9	44.9 (d)	1, 8, 9
3	5.55	d	9.6 (2)	2	7, 9	129.3 (d)	5, 7
4						137.9 (s)	
5A	2.30	ddd	14.4 (5B), 5.4 (6), 5.4 (6)	5B, 6	2	27.7 (t)	3, 4, 6, 7
5B	2.54	ddd	14.4 (5B), 5.4 (6), 5.4 (6)	5A, 6	2		3, 4, 6, 7
6	4.20	m		5A, 5B		63.5 (t)	1', 4
7	4.07	m			3	66.5 (t)	3, 4
8A	2.20	m		2, 8B, 9	12	31.4 (t)	1, 2, 3, 9, 10
8B	2.44	m		2, 8A, 9	12		1, 2, 3, 9, 10
9	5.02 ^{d1}	dd	7.2 (8A), 7.2 (8B)	8A, 8B	2, 3, 11	120.2 ^{d2} (d)	2, 8, 11, 12
10						134.3 (s)	
11	1.67	s			9	25.7 (q)	9, 10, 12
12	1.59 ^{d3}	s			8A, 8B	17.8 ^{d4} (q)	9, 10, 11
1'						174.3 (s)	
2'	3.28	m		3', 8'A, 8'B	5', 9'	44.9 (d)	1', 3', 4', 8', 9'
3'	5.51	d	9.6 (2')	2'	7', 8'A, 8'B, 9'	127.2 (d)	1', 2', 5', 7', 8'
4'						138.7 (s)	
5'A	2.25	m		5'B, 6'A, 6'B	2'	32.2 (t)	3', 4', 6', 7'
5'B	2.51	m		5'A, 6'A, 6'B	2'		3', 4', 6', 7'
6'A	3.65	br s		5'A, 5'B, 6'B		61.1 (t)	
6'B	3.71	br s		5'A, 5'B, 6'A			
7'	4.02	m			3'	67.4 (t)	3', 4'
8'A	2.20	m		2', 8'B, 9'	12'	31.4 (t)	1', 2', 3', 9', 10'
8'B	2.44	m		2', 8'A, 9'	12'		1', 2', 3', 9', 10'
9'	5.06 ^{d1}	dd	7.2 (8'A), 7.2 (8'B)	8'A, 8'B	2', 3', 11'	120.3 ^{d2} (d)	11', 12'
10'						134.3 (s)	
11'	1.67	s			9'	25.7 (q)	9', 10', 12'
12'	1.61 ^{d3}	s			8'A, 8'B	17.9 ^{d4} (q)	9', 10', 11'

a ^1H chemical shift values (d ppm from SiMe₄) followed by multiplicity and then the coupling constants (J/Hz). Figures in parentheses indicate the proton coupling with that position. b The correlations with geminal and vicinal protons are removed. c Long range ^1H - ^{13}C correlations from H to C observed in the HMBC experiment. d1-d4 interchangeable

Table S4 NMR spectral data of **4** in CDCl₃

Position	$\delta_{\text{H}}^{\text{a}}$		J/Hz	^1H - ^1H COSY	NOESY ^b	δ_{C}	HMBC (C) ^c
1	3.68	m		2, 8A, 8B	9	40.2 (d)	
2	5.36	br s		1	8A, 8B, 9, 13A, 13B	121.2 (d)	4, 7, 8, 13
3						139.2 (s)	
4A	2.45	br d	19.2 (5B)	4B, 5	13A	30.3 (t)	
4B	2.59	m		5A, 5B, 6B	13B		
5 α	4.68	ddd	12.6 (4B), 12.6 (5 β), 1.8 (4A)	4, 5 β		64.4 (t)	3, 4, 7
5 β	4.33	ddd	12.6 (5 α), 4.8 (4B), 2.4 (4A)	4, 5 α			3, 4, 7
6							
7						174.3 (s)	
8A	2.33	ddd	14.4 (8B), 6.6 (1), 6.6 (9)	1, 8B, 9	2, 11	30.1 (t)	1, 2, 7, 9, 10
8B	2.52	ddd	14.4 (8A), 6.6 (1), 6.6 (9)	1, 8A, 9	2, 11		1, 2, 7, 9, 10
9	5.14	dd	6.6 (8A), 6.6 (8B)	8A, 8B	1, 2, 11	120.9 (d)	1, 8, 11, 12
10						134.6 (s)	
11	1.72	s			9	25.8 (q)	9, 10, 12
12	1.67	s			8A, 8B	18.0 (q)	9, 10, 12
13A	3.99	d	13.8 (13B)		2, 4	67.4 (t)	2, 3, 4
13B	4.01	d	13.8 (13A)		2, 4		

a ^1H chemical shift values (δ ppm from SiMe₄) followed by multiplicity and then the coupling constants (J /Hz). Figures in parentheses indicate the proton coupling with that position. b The correlations with geminal and vicinal protons are removed. c Long range ^1H - ^{13}C correlations from H to C observed in the HMBC experiment.

Figure S1 ^1H NMR spectrum of 1

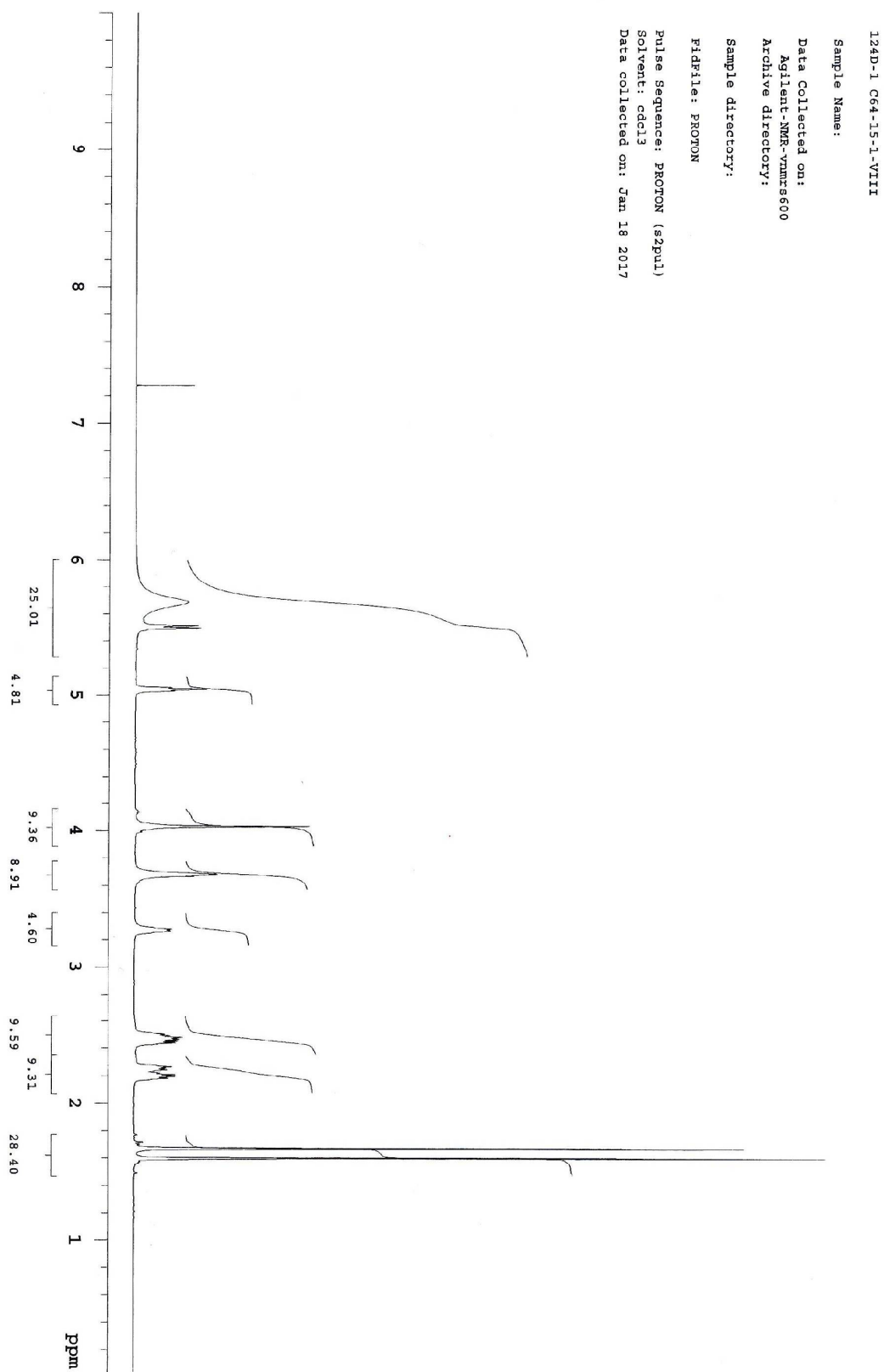


Figure S2 ^{13}C NMR spectrum of 1 in CDCl_3

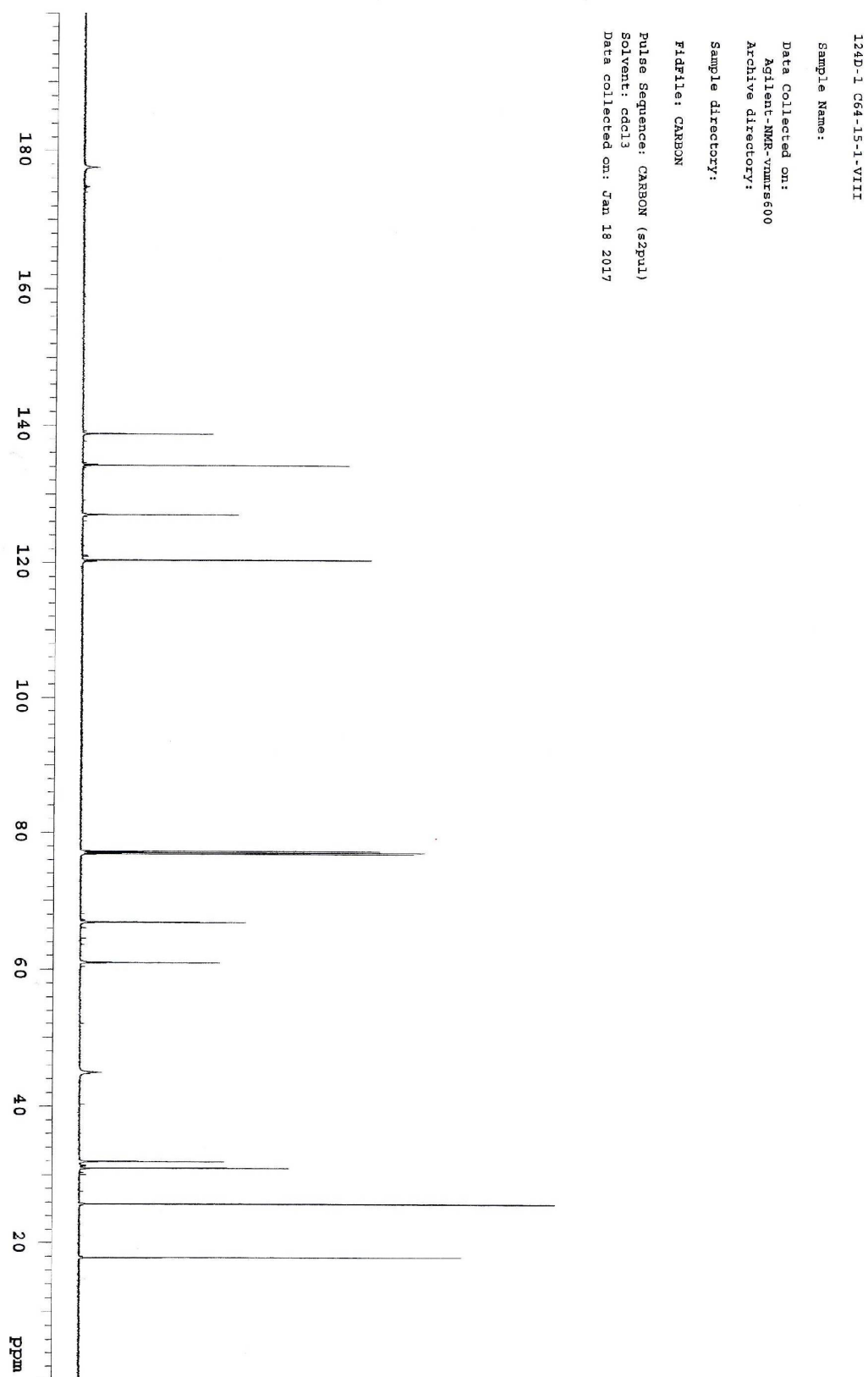


Figure S3 ^1H - ^1H COSY of 1

124D-1 C64-15-1-VIII

exp23 gcosy

SAMPLE		FLAGS	
date	Jan 18 2017	he	in
solvent	cdcl3	spul	y
sample	cdcl3	hagl	6120
ACQUISITION			
sw	3765.1	temp	not used
at	0.150	gain	20
np	4000	F2 PROCESSING	0
ss	32	ab	-0.075
dl	1.000	abs	not used
nt	8	fn	2048
2D ACQUISITION			
sw1	3765.1	ab1	-0.034
ni	256	abs1	not used
d2	0	procl	ip
PRESATURATION			
estcode	n	fn1	DISPLAY
wtcode	n	sp	783.3
tr	TRANSMITTER	hl	3000.3
tr	559.898	wp1	775.9
efrq	-447.1	rfl	2596.6
tpwr	58	rfl	-680.3
pw	9.600	rfl1	-680.3
GRADIENTS			
g1v1e	5102	rfl1	0
g1e	0.001000	wc	200.0
EDratio	1.000	sc	0
g1tab	0.000500	wc2	200.0
DECOUPLER			
dn	C13	vs	139
dm	mn	th	2
a1 cdc av			

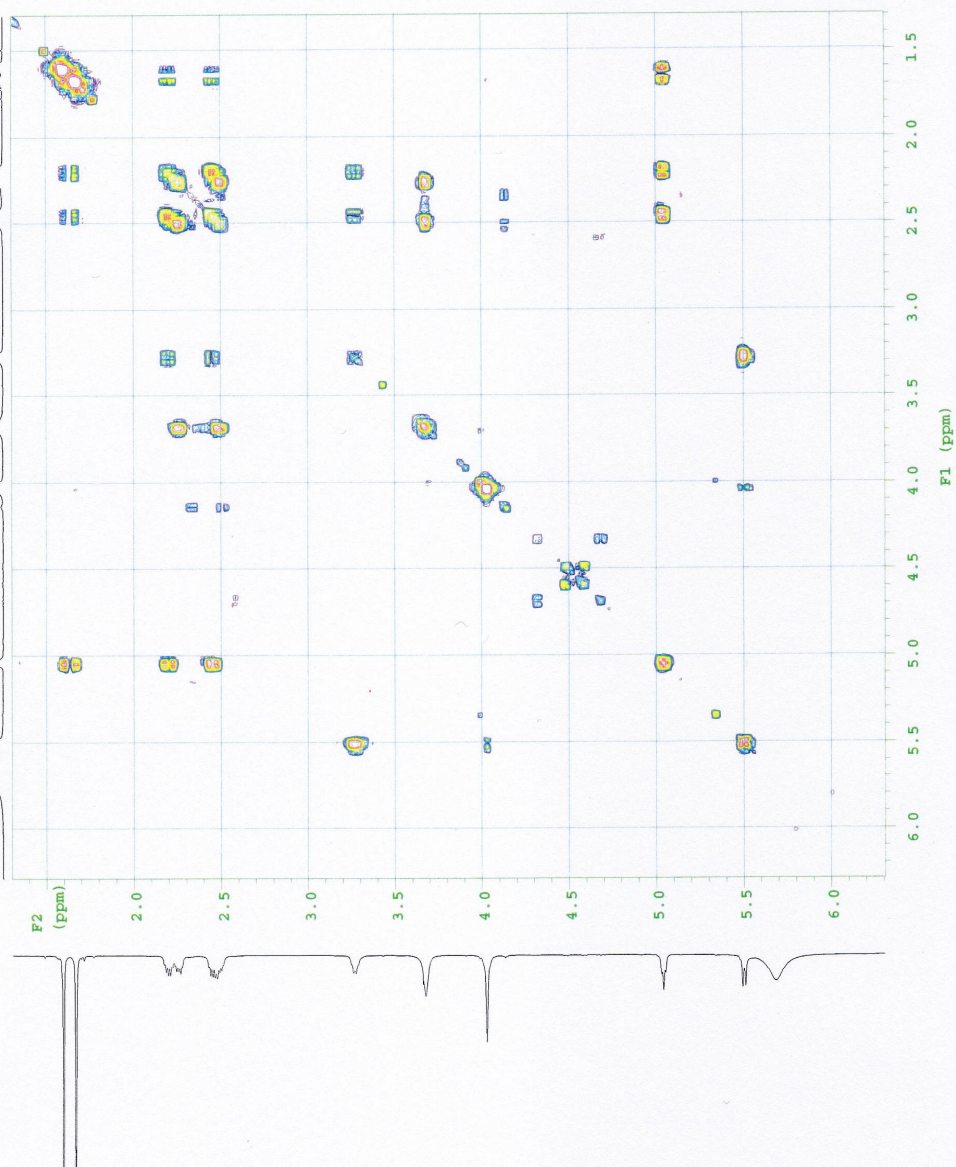


Figure S4 NOESY of 1

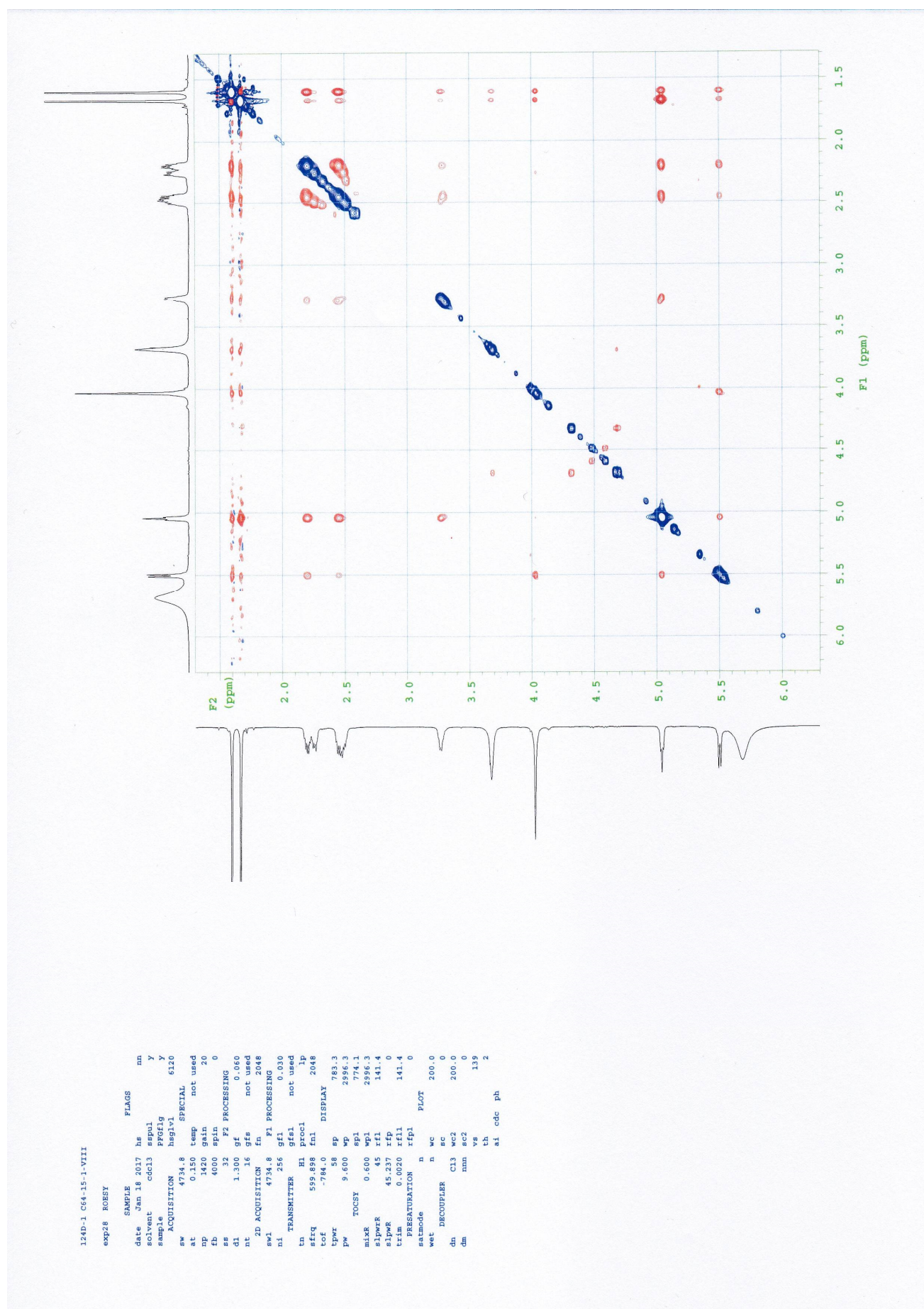


Figure S5 HMQC of **1**

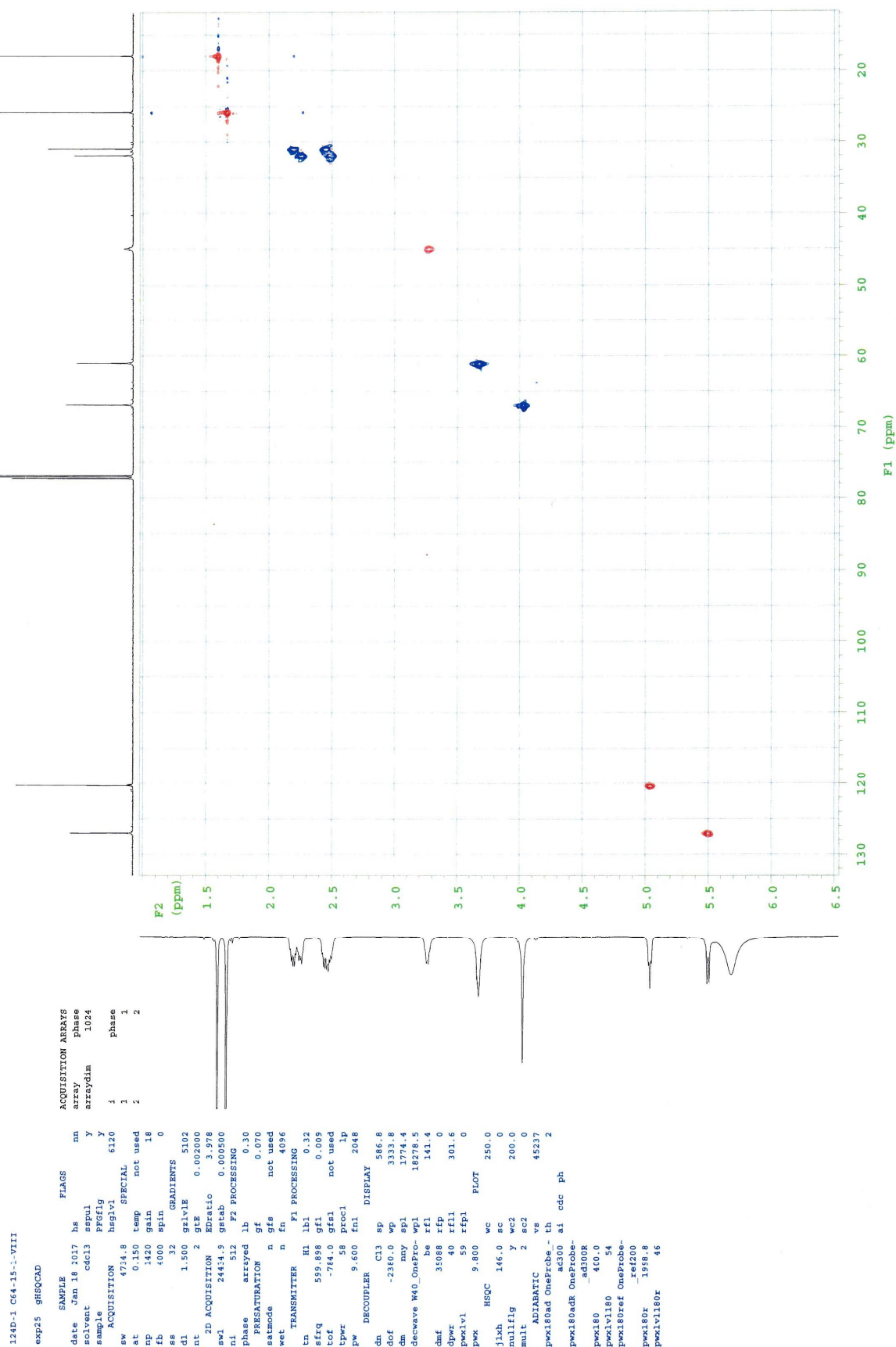


Figure S6 HMBC of 1

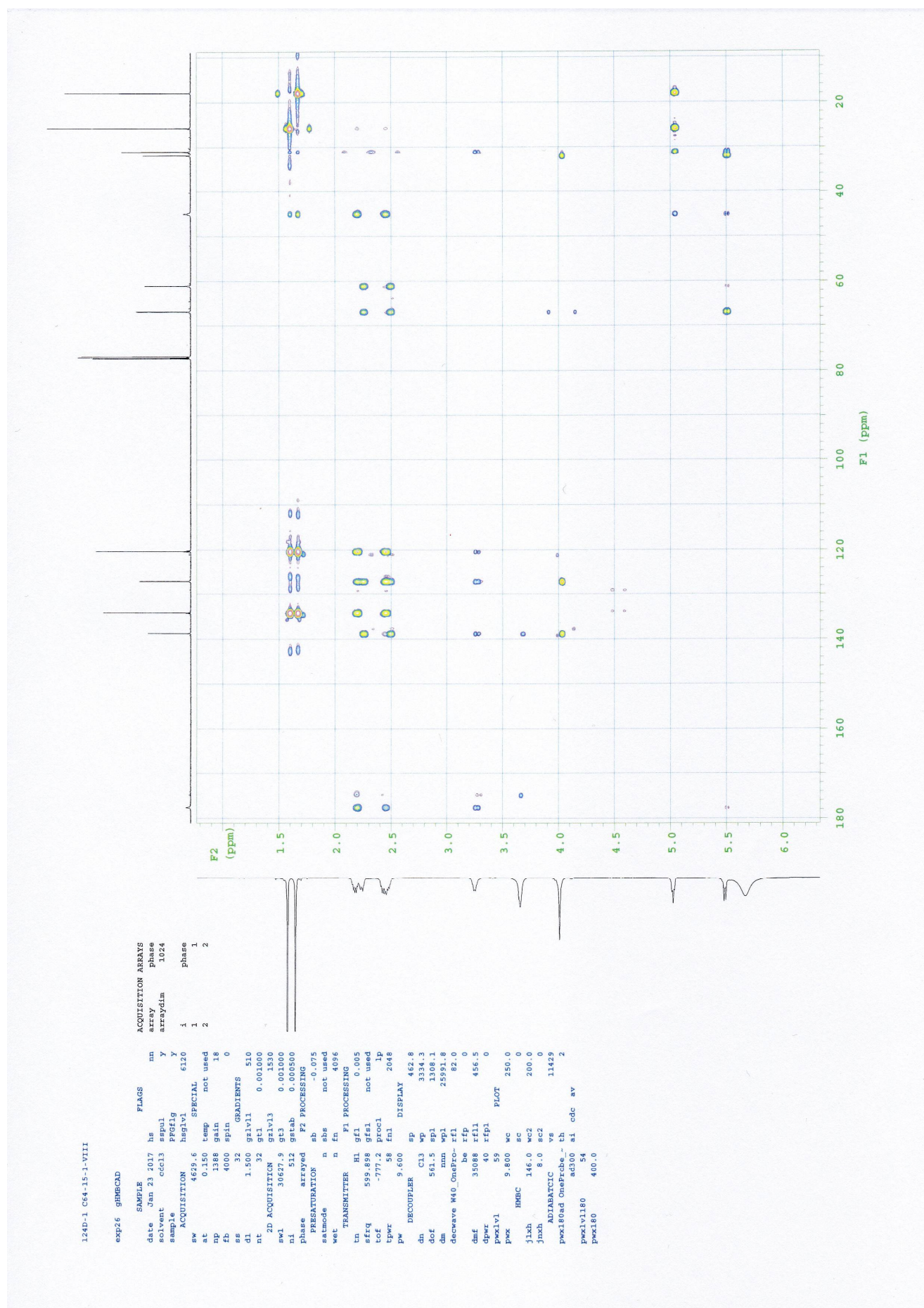


Figure S7 ^1H NMR spectrum of 2 in CDCl_3

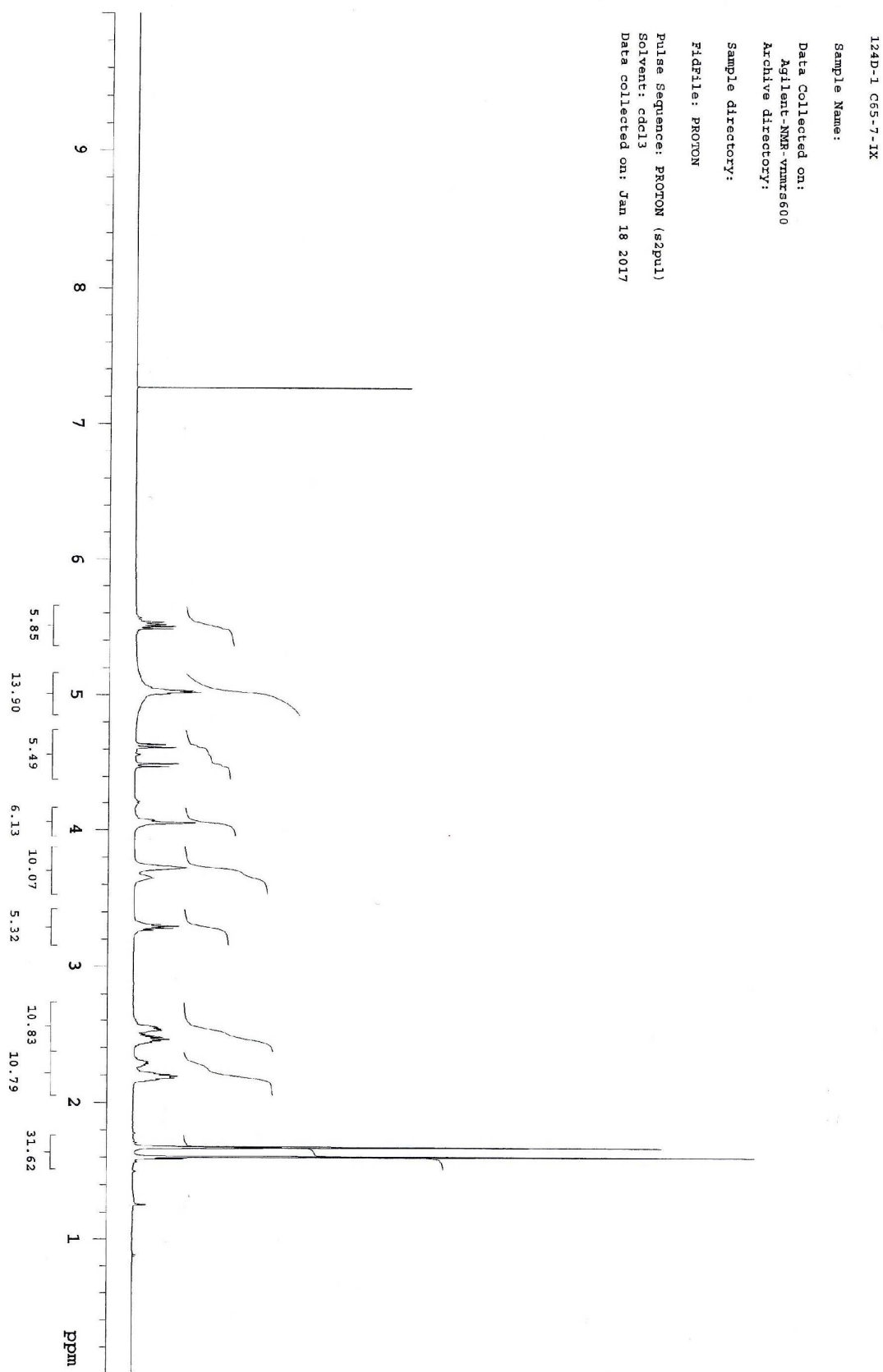


Figure S8 ^{13}C NMR spectrum of 2 in CDCl_3

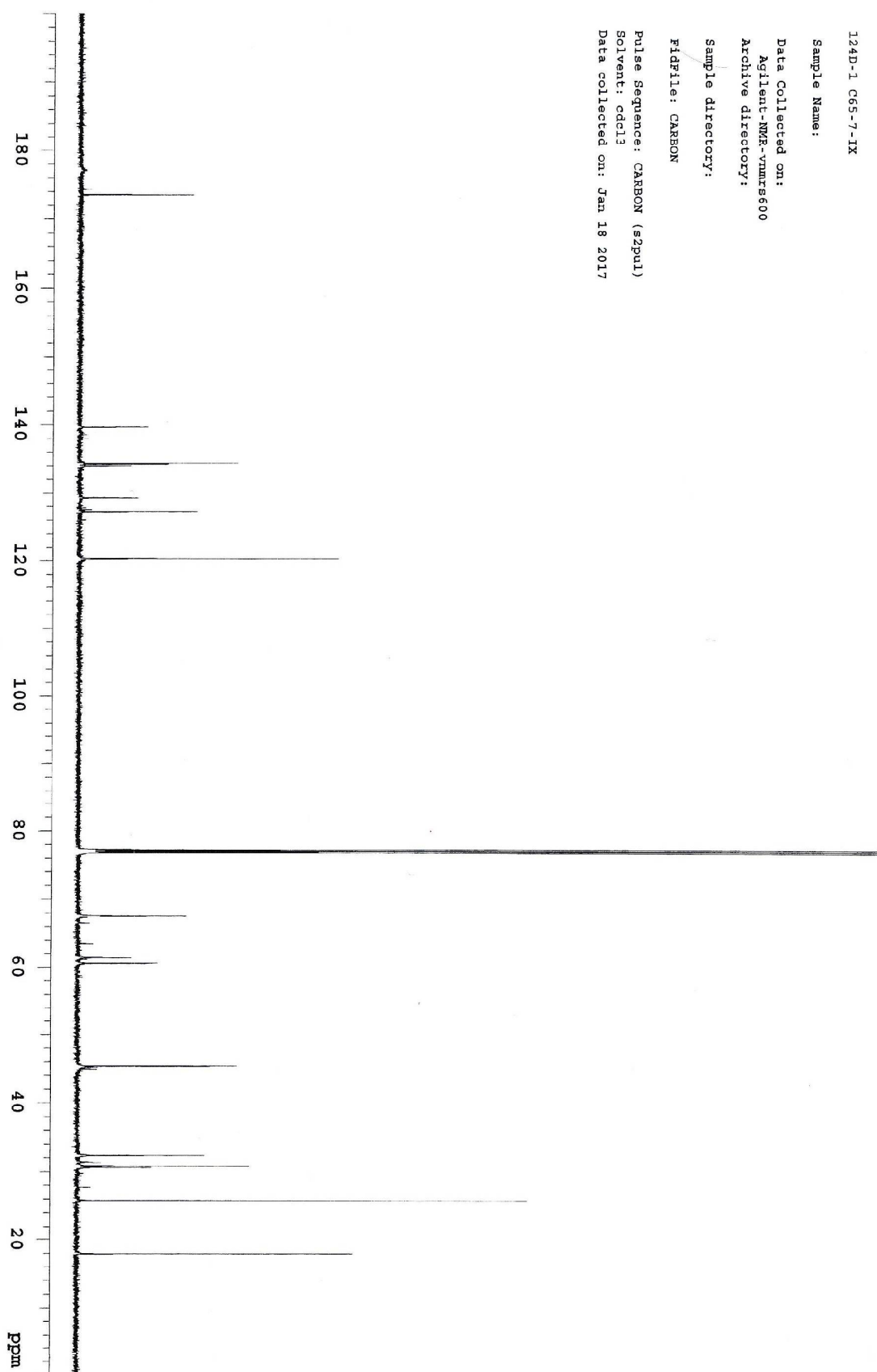


Figure S9 ^1H - ^1H COSY of 2

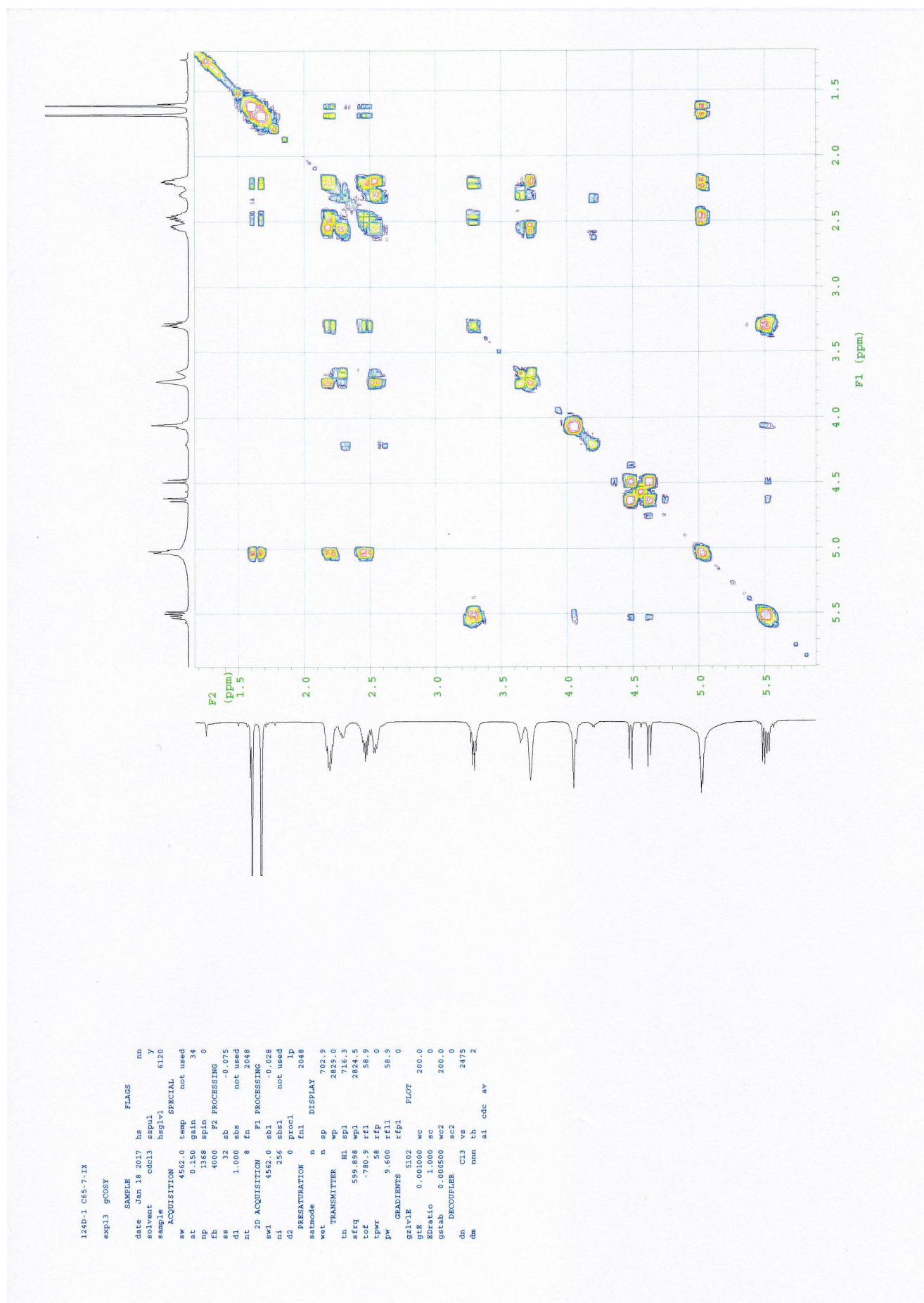


Figure S10 NOESY of 2

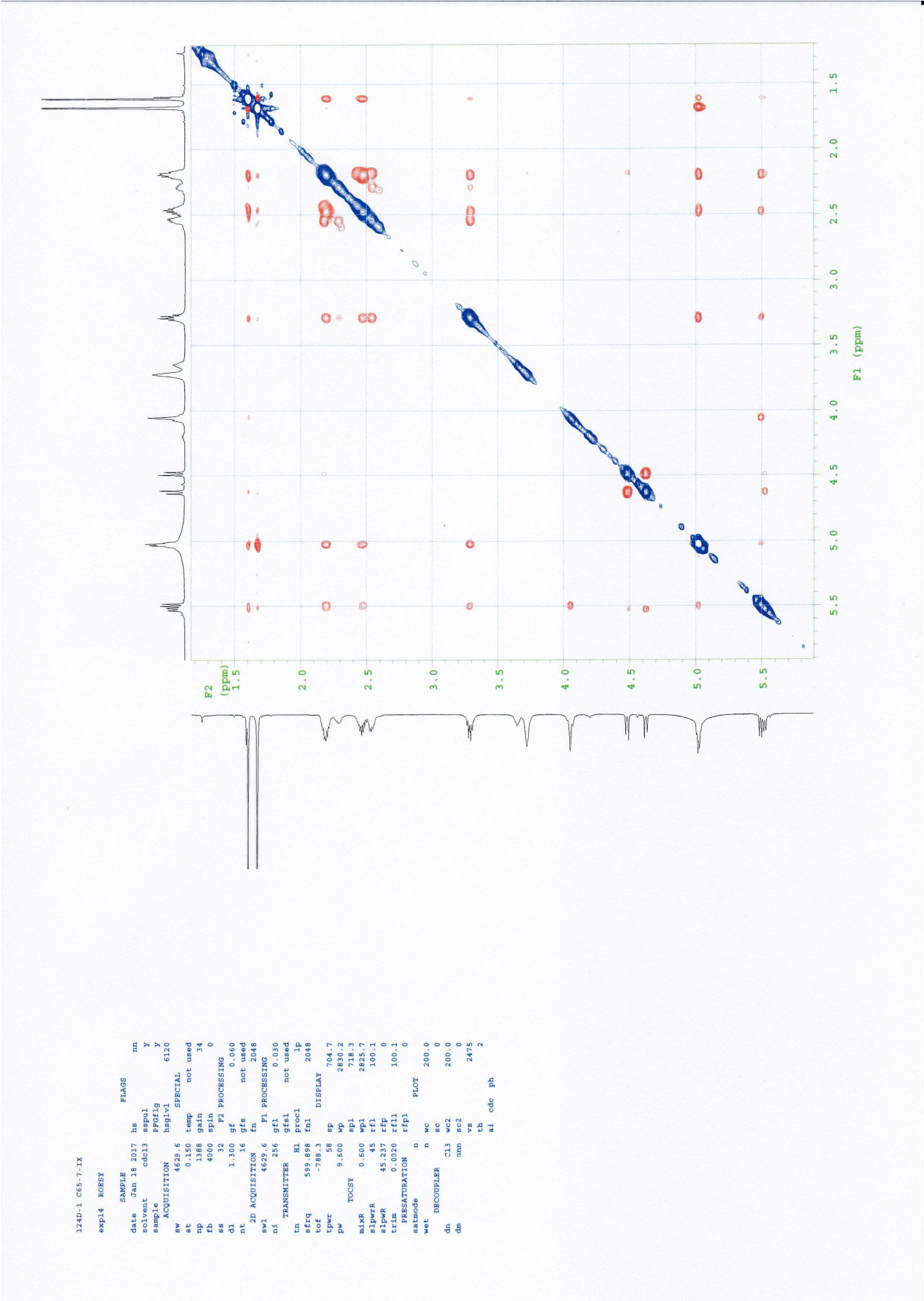


Figure S11 HMQC of 2

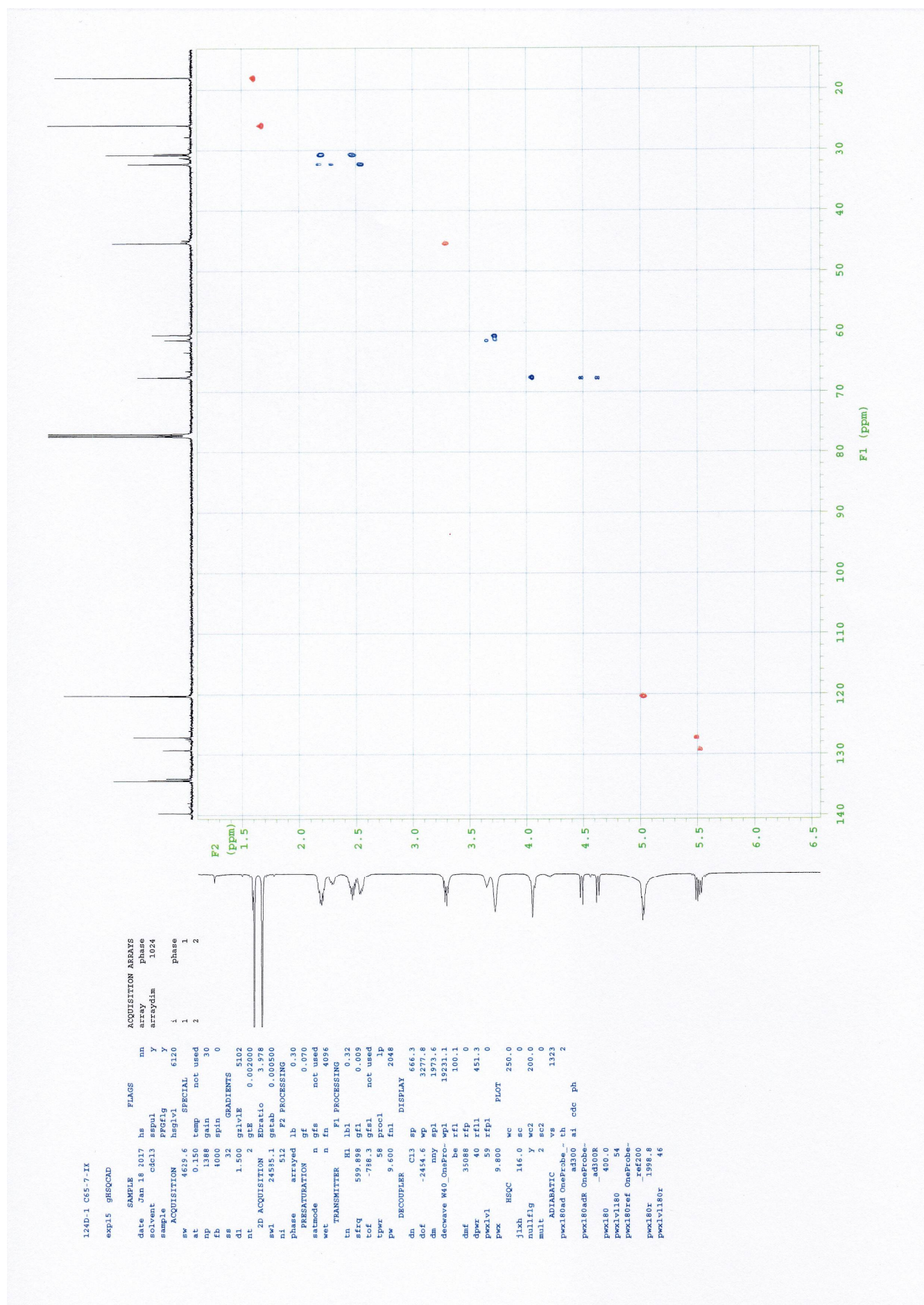


Figure S12 HMBC of 2

124D-1 C65-7-IX

exp16 ghmecad

SAMPLE		FLAGS		ACQUISITION ARRAYS	
date	Jan 18 2017	ha	nm	array	phase
solvent	cdcl3	spul	y	arraydim	1024
sample	cdcl3	pfeflg	y		
acq	4625.6	hsglvi	6120	1	Phase
sw	0.150	temp	not used	2	1
at	1288	gain	30		2
ap	4000	spin	0		
ss	32	GRADIENTS	0		
gt	1.500	g1v11	510		
nt	2D ACQUISITION	g1	0.001000		
ev1	30921.9	g1v13	0.001000		
si	512	g1v1b	0.000500		
Phase	arrayed	eb	F2 PROCESSING		
PRESSATURATION	ch	eb	0.075		
satmode	n	eb	not used		
wet	n	fn	not used		
TRANSMITTER	n	fn	4096		
tn	599.698	gfl	not used		
sfq	-788.3	procl	lp		
tpw	58	fn1	2048		
pw	9.600	DISPLAY			
DECOUPLER	sp	754.4			
dn	C13	wp	2668.7		
dof	411.4	sp1	1359.3		
dm	nmn	wp1	25910.8		
decwave	W40_OnePro-	rfl	100.1		
be	rfl	0			
dmf	35088	rfl1	754.6		
dpwr	40	rfl1	0		
pw1v1	59	PILOT			
pw1	9.800	wc	250.0		
pw1x	HMBC	sc	0		
j1xh	146.0	wc2	200.0		
j1xh	8.0	sc2	0		
ADIAATIC	vs	952			
pw180ad	OneProbe	th	2		
ad300	ai	cdc	av		
pw1v1180	54				
pw180	400.0				

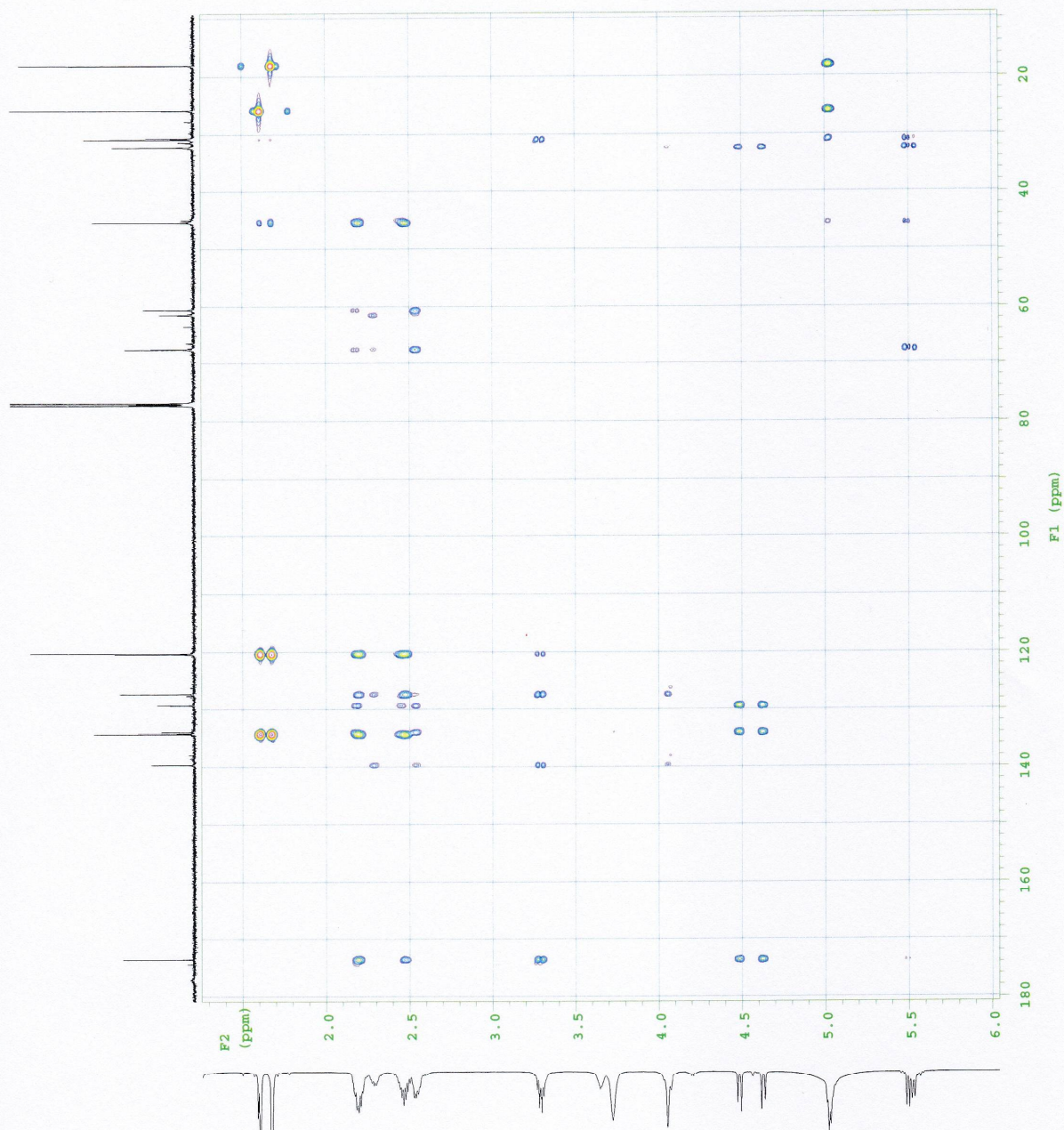


Figure S13 ^1H NMR spectrum of 3 in CDCl_3

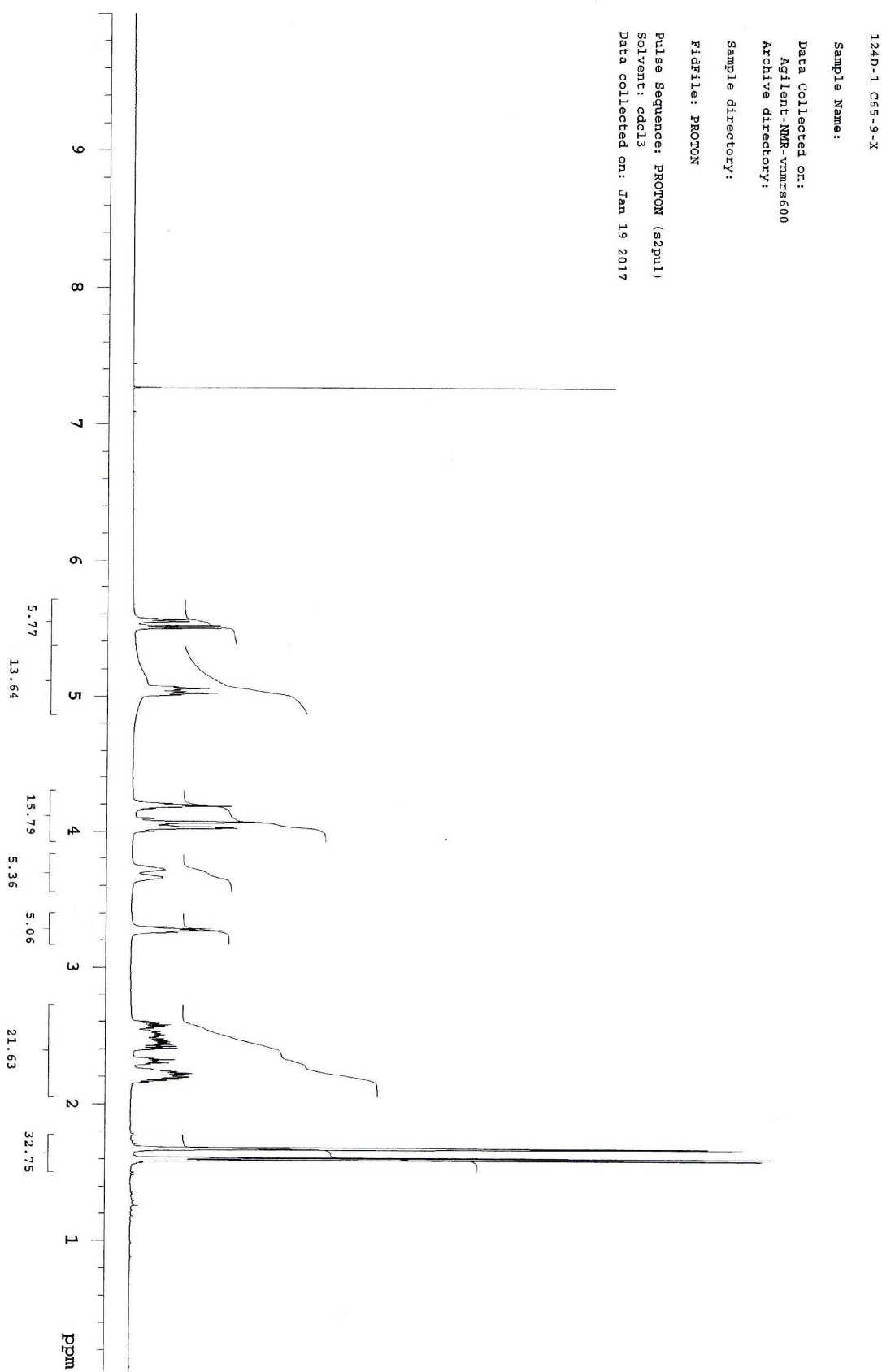


Figure S14 ^{13}C NMR spectrum of 3 in CDCl_3

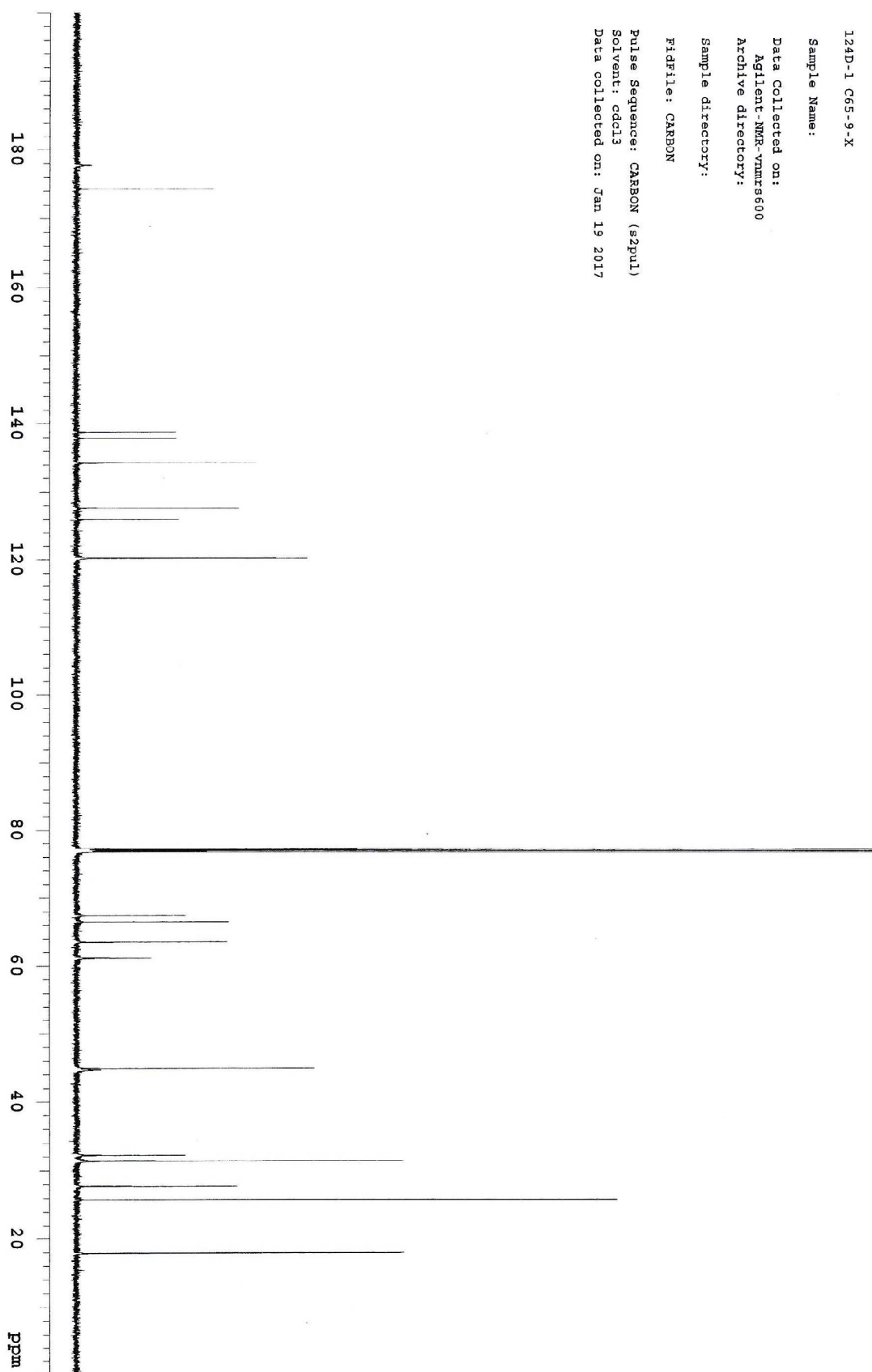


Figure S15 ^1H - ^1H COSY of 3

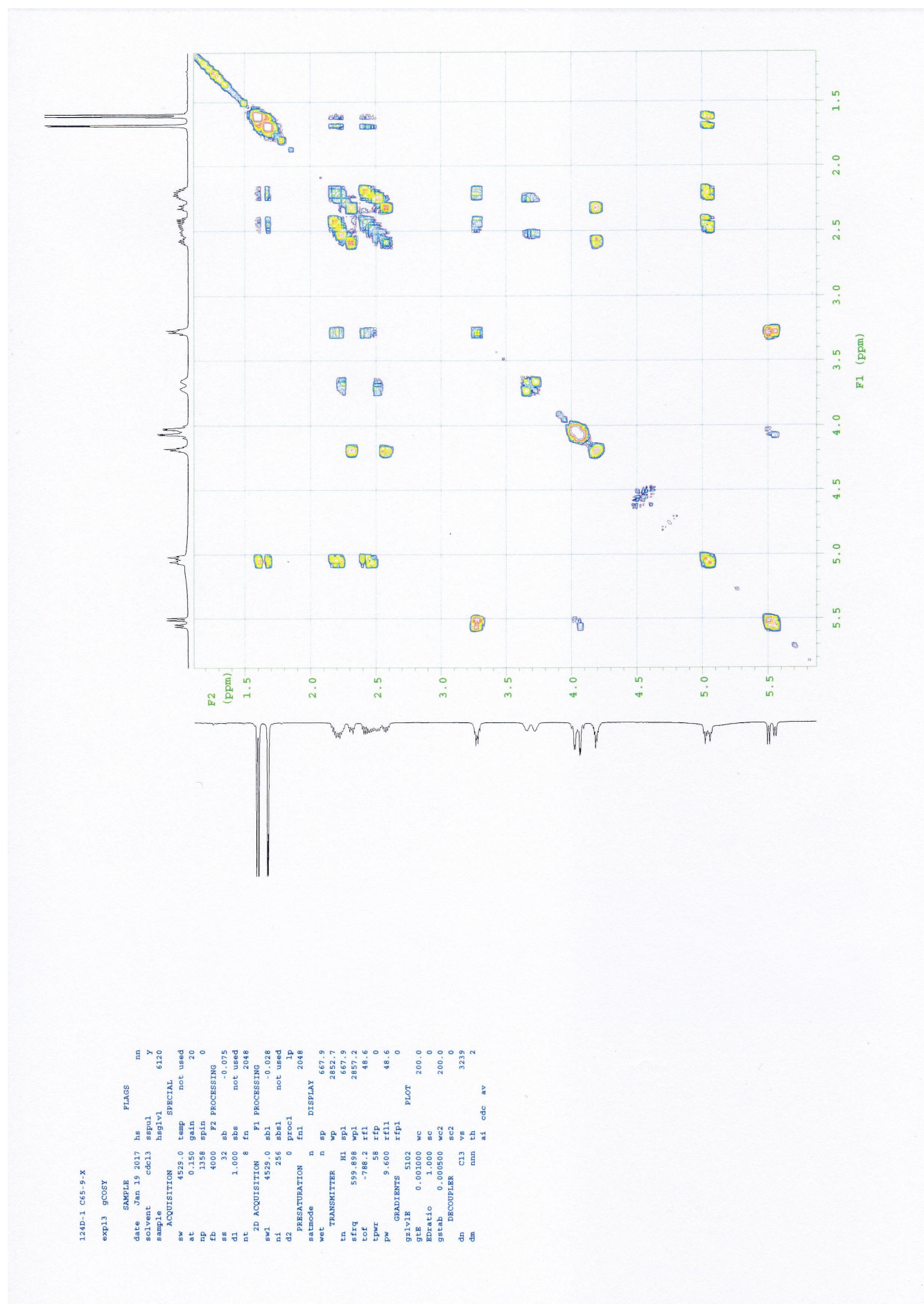


Figure S16 NOESY of 3

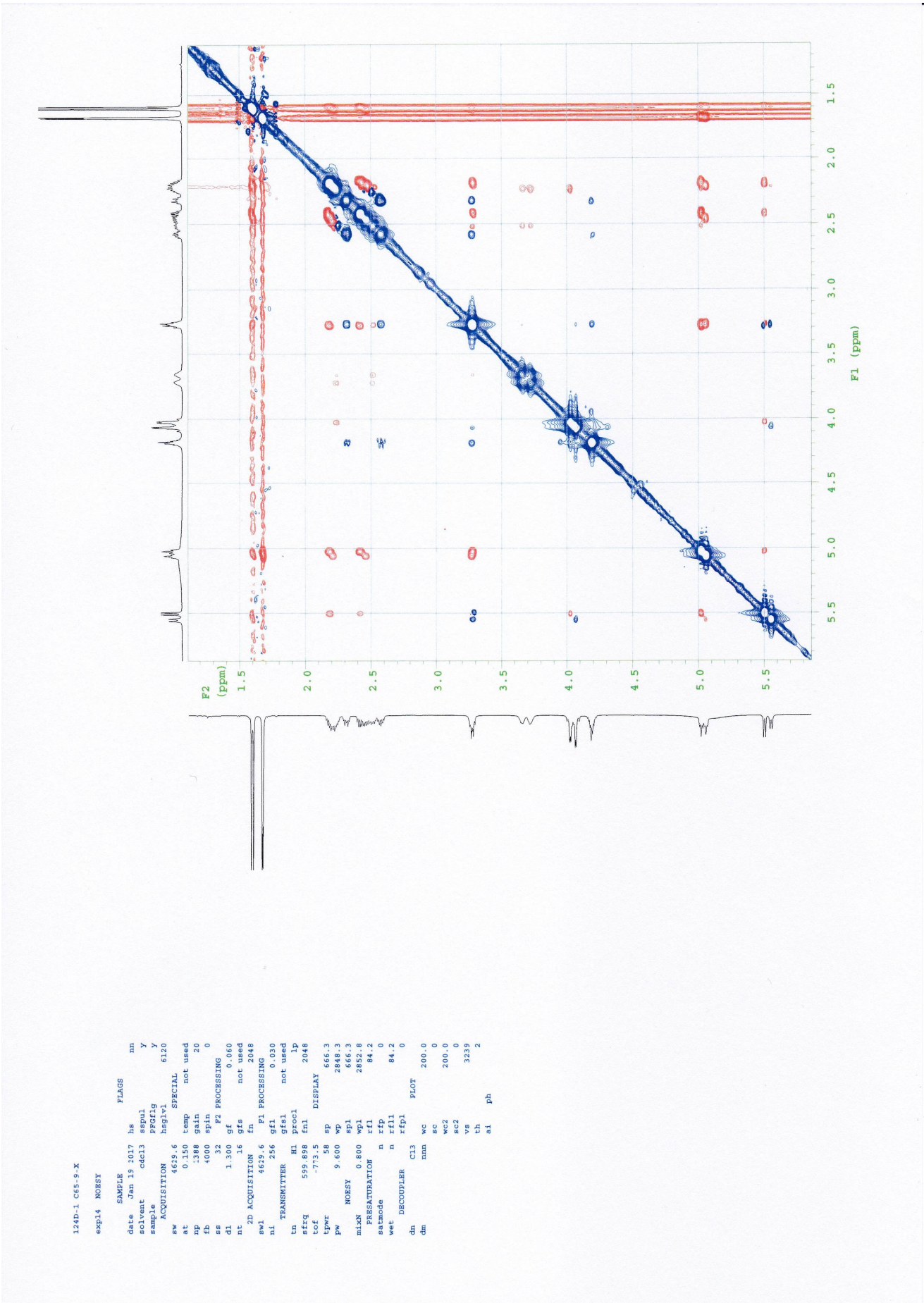


Figure S17 HMQC of 3

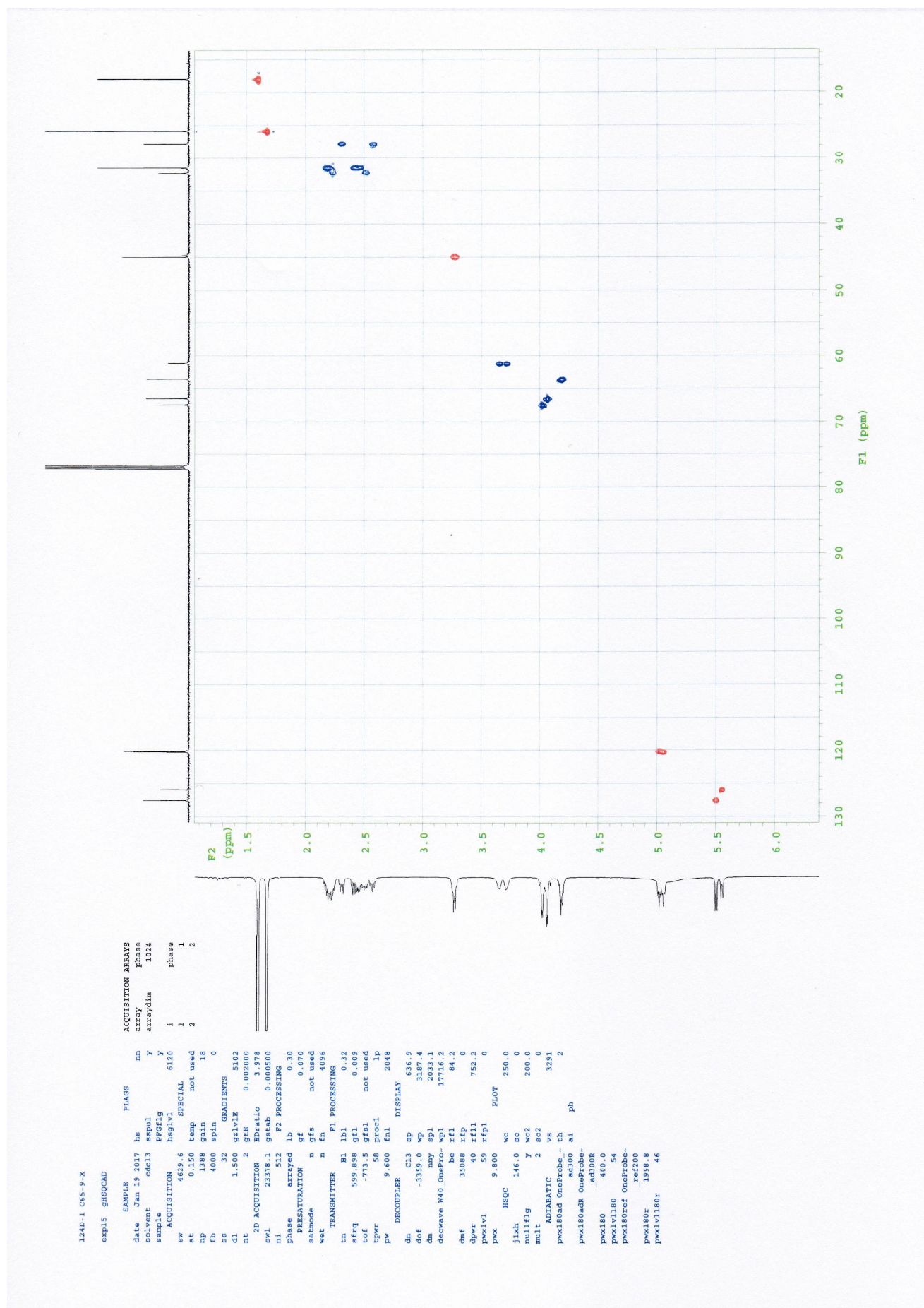


Figure S18 HMBC of 3

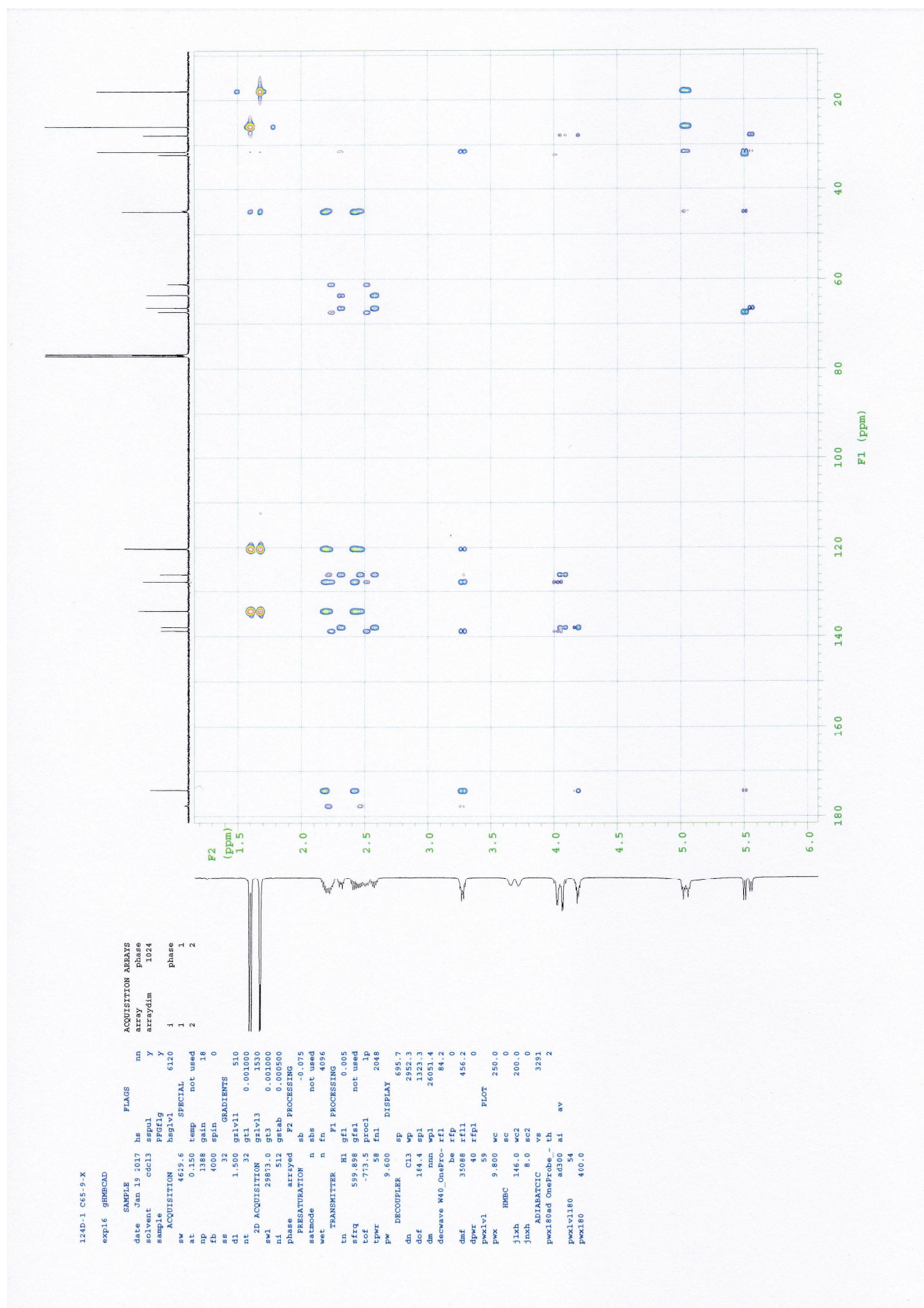


Figure S19 ^1H NMR spectra of 4 in CDCl_3

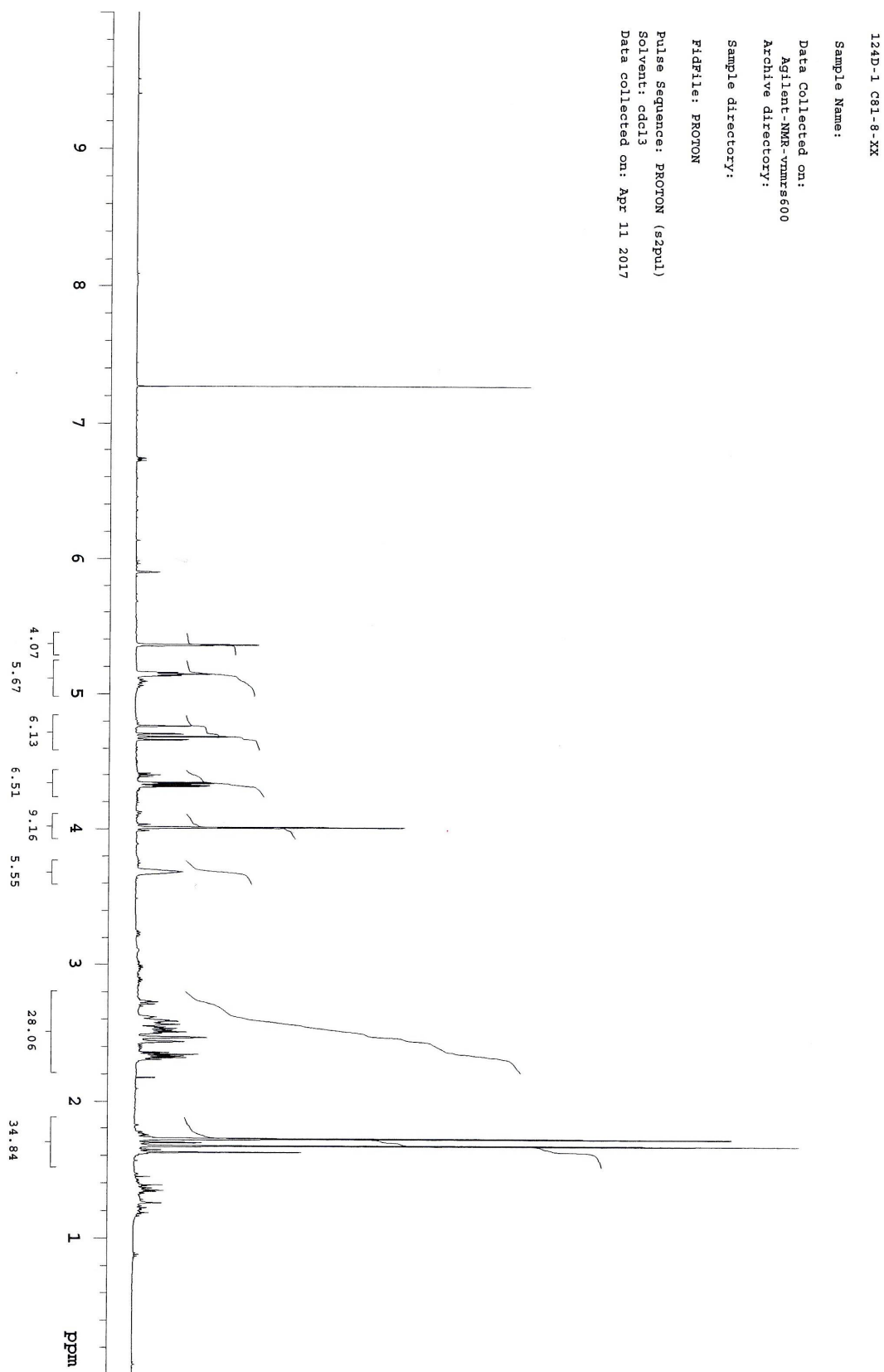


Figure S20 ^{13}C NMR spectra of 4 in CDCl_3

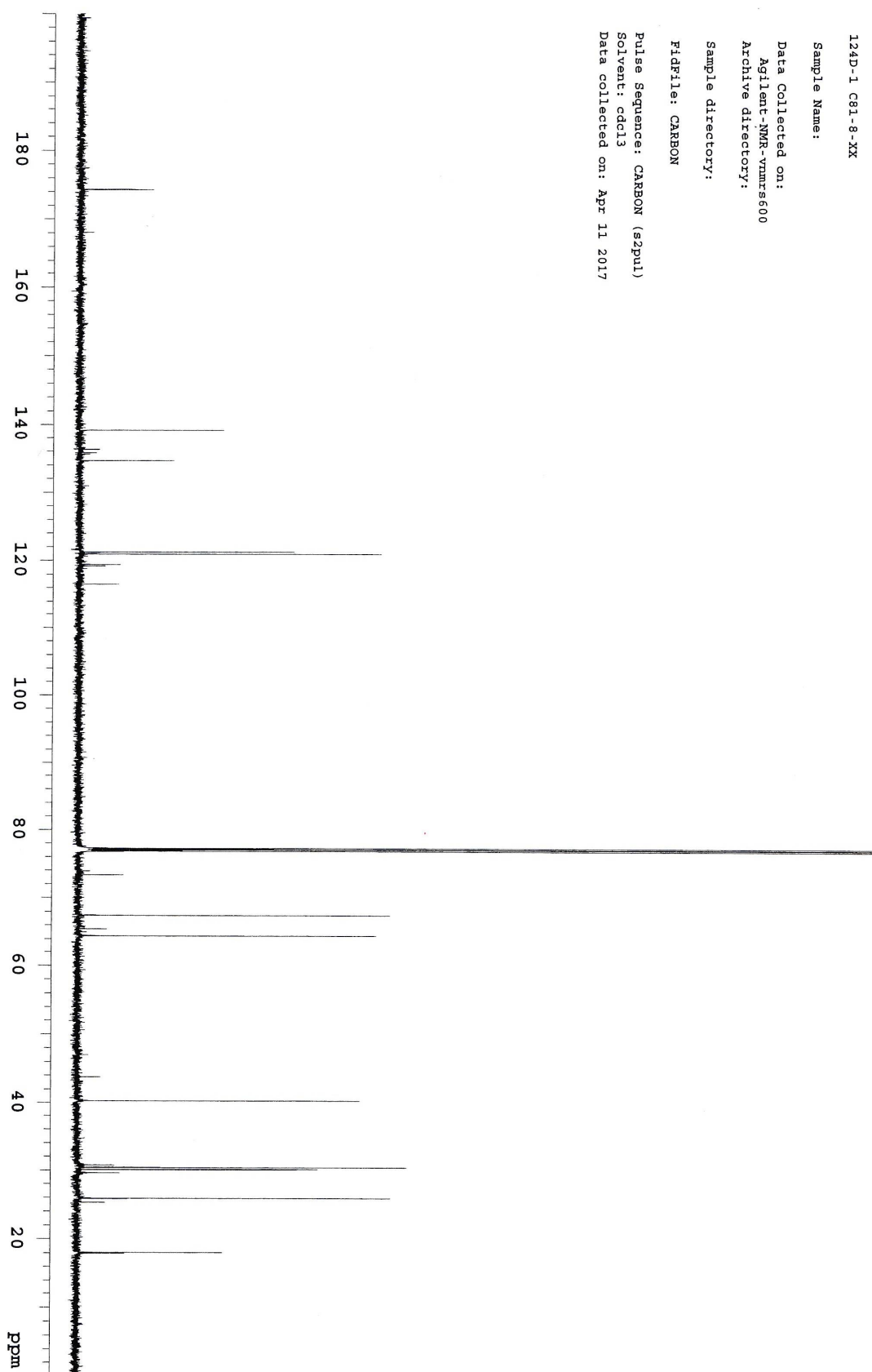


Figure S21 ^1H - ^1H COSY of 4

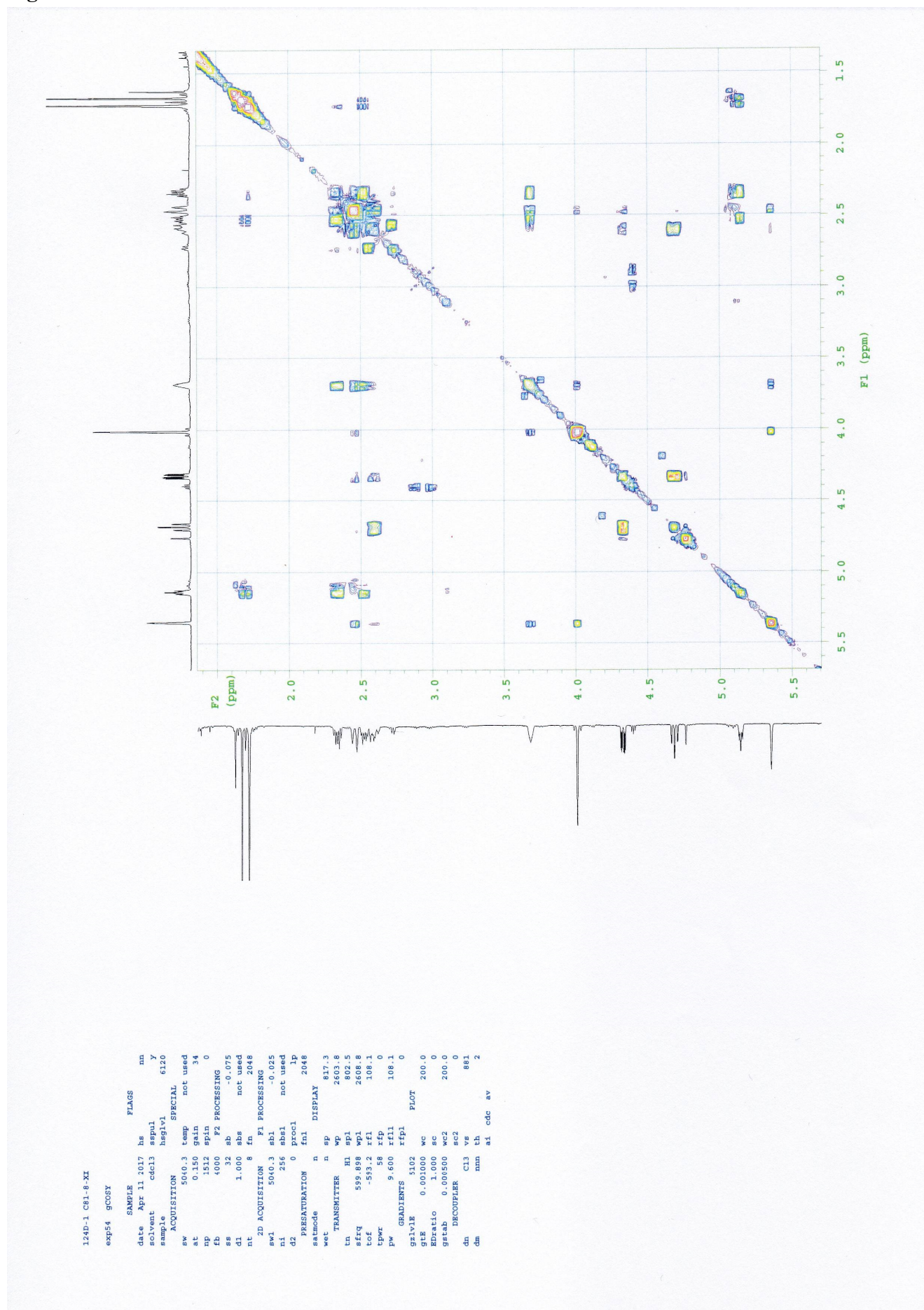


Figure S22 NOESY of 4⁺

124D-1 C81-8-XX

exp57 NOESY

SAMPLE		FLAGS	
date	Apr 11 2017	hs	nn
solvent	cccl3	aspl	y
sample	peclg	peclg	6120
sw	4867.7	SPECIAL	
at	0.150	temp	not used
np	1442	gain	34
fb	4000	spin	0
ss	32	P2 PROCESSING	
d1	1.300	Gf	0.060
nt	16	Gfs	not used
2D ACQUISITION	fn	2048	
sw1	4867.7	F1 PROCESSING	
ni	256	Gf1	0.030
tn	H1	proc1	lp
sfrq	599.898	fn1	2048
tof	-738.1	DISPLAY	
tpwr	58	sp	816.4
pw	9.600	wp	2601.0
mixN	NOESY	spl	802.3
PRESATURATION	rfl	2605.7	
satmode	n	rfl	136.7
wet	n	rfl1	136.7
DECOUPLER	cl13	rfl1	0
dm	nmn	wc	200.0
	sc	0	
	wc2	200.0	
	sc2	0	
	vs	881	
	th	cdc	2
	ai	ph	

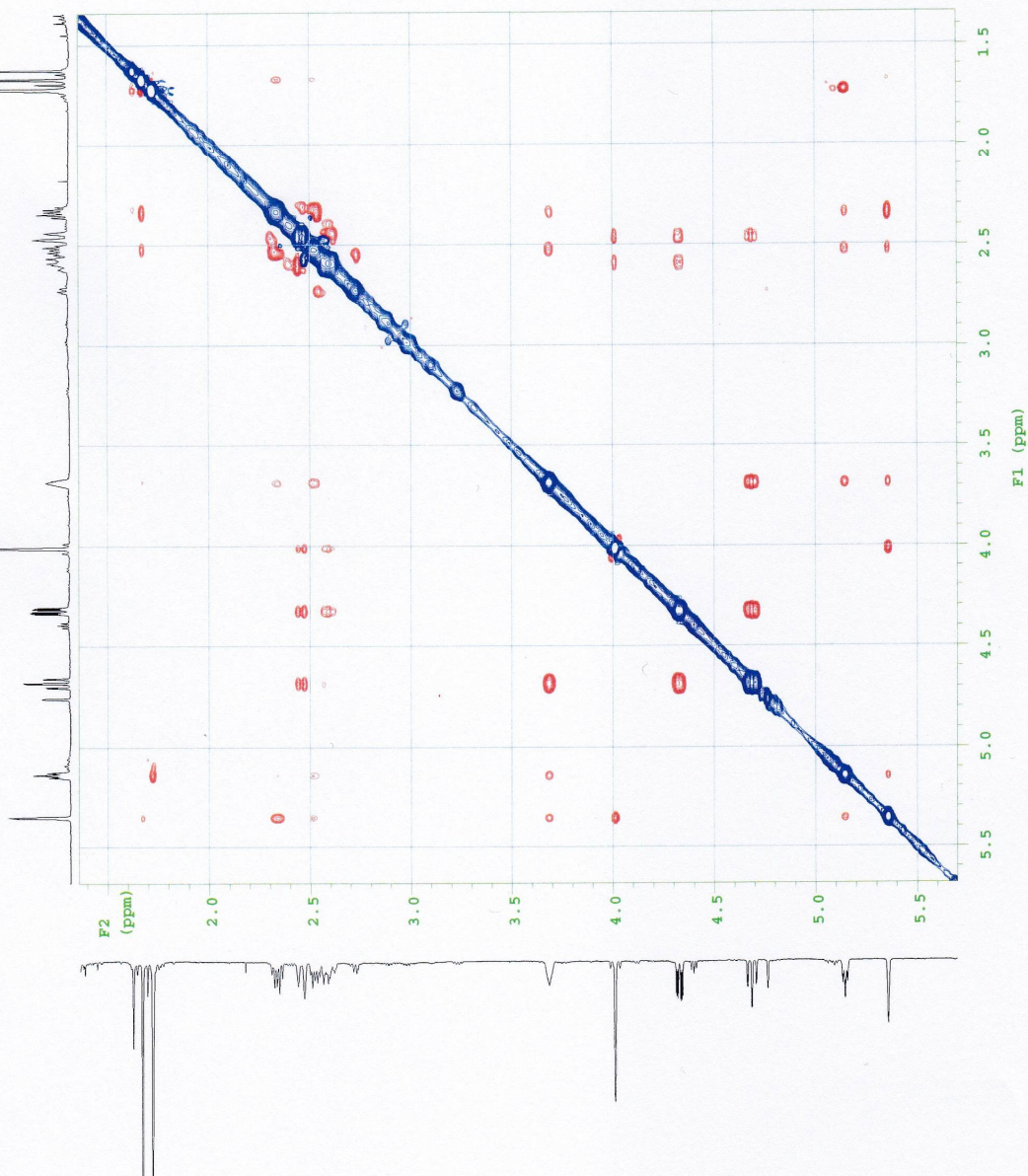


Figure S23 HMQC of 4

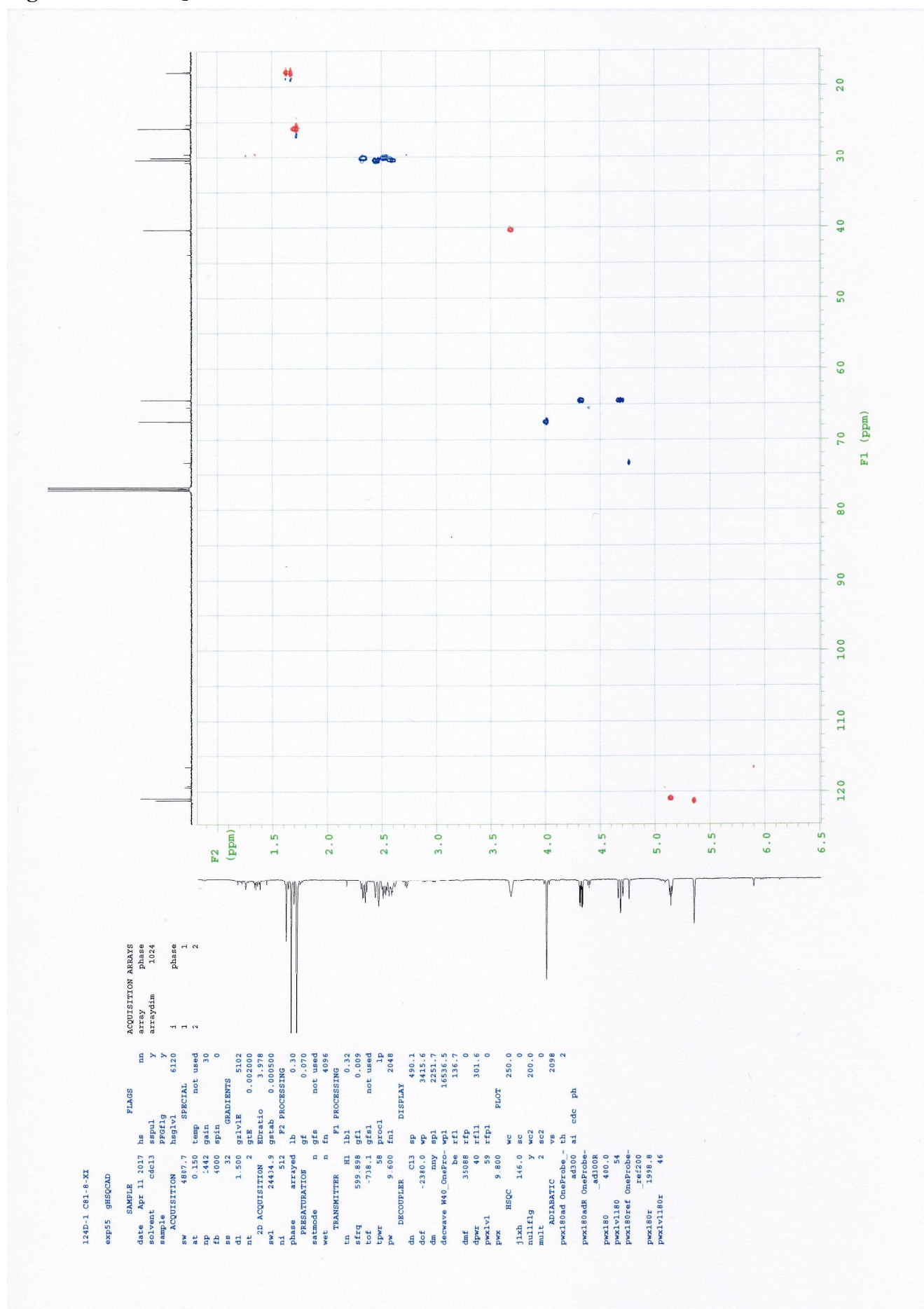


Figure S24 HMBC of 4

124D-1 C81-8 XX

exp60 gHBCAD

SAMPLE		FLAGS		ACQUISITION ARRAYS	
date	Apr 11 2017	hs	nm	array	phase
solvent	cdcl3	sepul	y	arraydim	1024
sample					
PROBHD					
ACQUISITION	hsq1v1	6120	i		
SW	4607.7	SPECIAL			
at	0.150	temp	not used		
ap	1442	gain	30		
ab	4000	spin	0		
ds	24				
dt	1.592	g1v11	510		
nt	0.001000				
2D ACQUISITION	g1v13	0.001000			
sd1	30464.6	g1v13	1530		
ni	512	getab	0.000500		
Phase	arrayed	P2 PROCESSING			
PRESATURATION	sb	-0.075			
satmode	n	sbs	not used		
wet	n	fn	4096		
TRANSMITTER		P1 PROCESSING			
tn	H1	gfl	0.005		
sfrq	599.898	gfa1	not used		
tof	-738.1	procl	1p		
tpwr	58	fn1	2048		
pw	9.600	DISPLAY			
DECOUPLER	sp	1253.1			
dn	C13	wp	2115.1		
dof	639.9	sp1	2053.8		
dm	nm	wpl	25139.2		
decwave	W40	OnePro-	rfl	136.7	
be	rfl	0			
dmf	35088	rfl1	296.5		
dpwr	40	rfl1	0		
pw1v1	59	PLOT			
pw2	9.800	wc	250.0		
sc					
j1xh	146.0	wc2	200.0		
j2xh	8.0	sc2	0		
ADIBATIC	vs	2098			
pw180ad	OneProbe-	th	2		
ad300	ai	cdc	av		
pw1v1180	54				
pw2180	400.0				

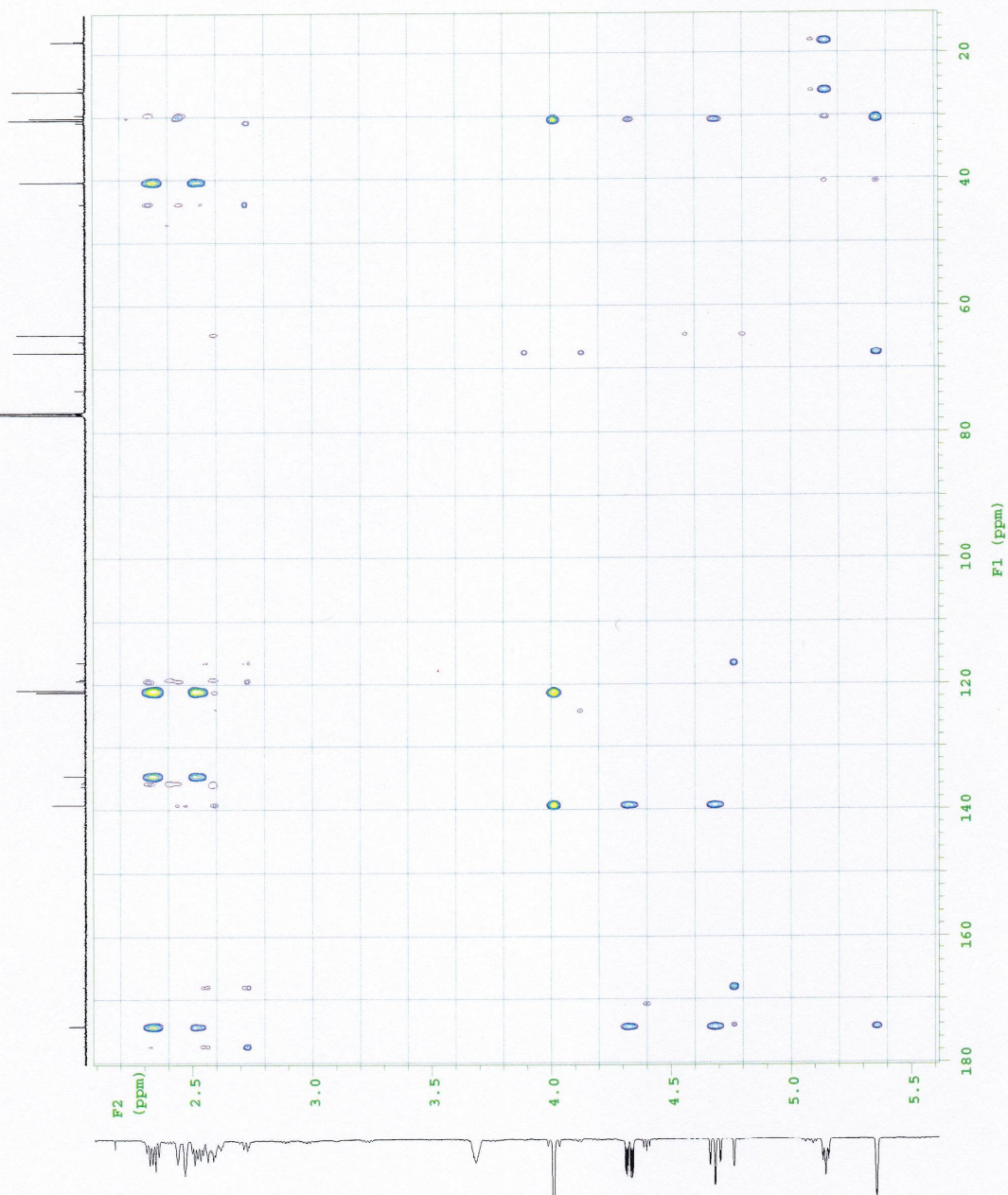
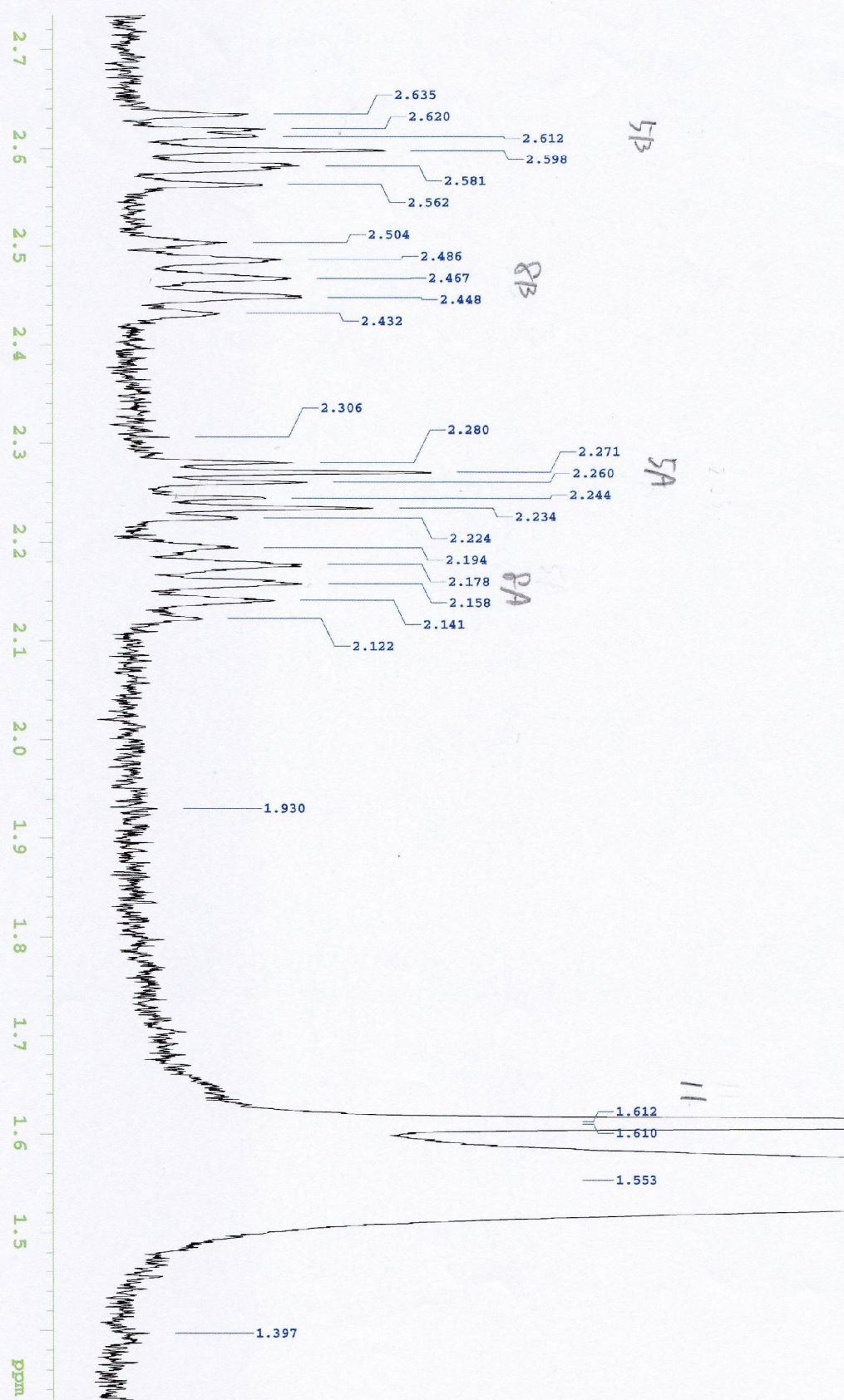
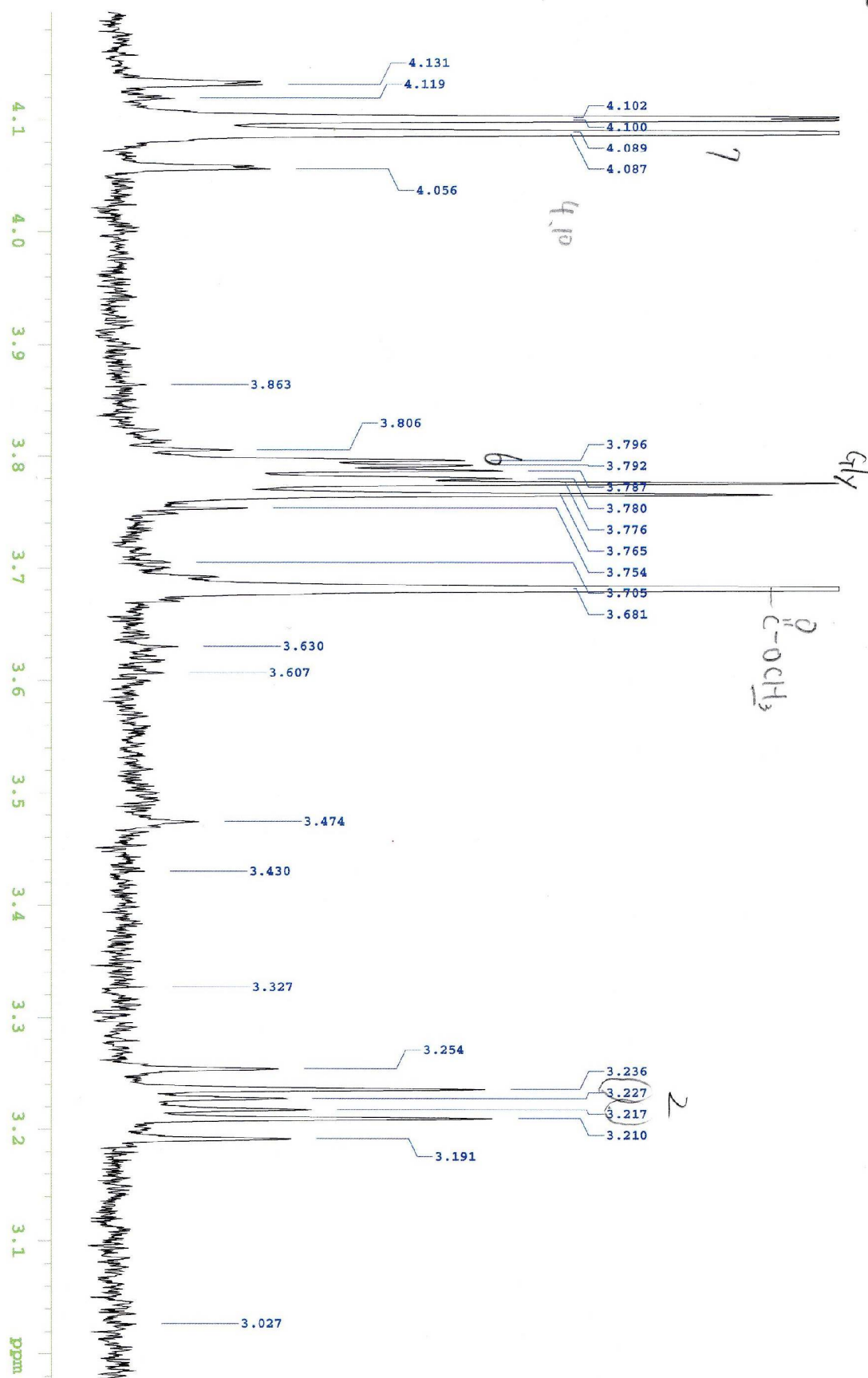


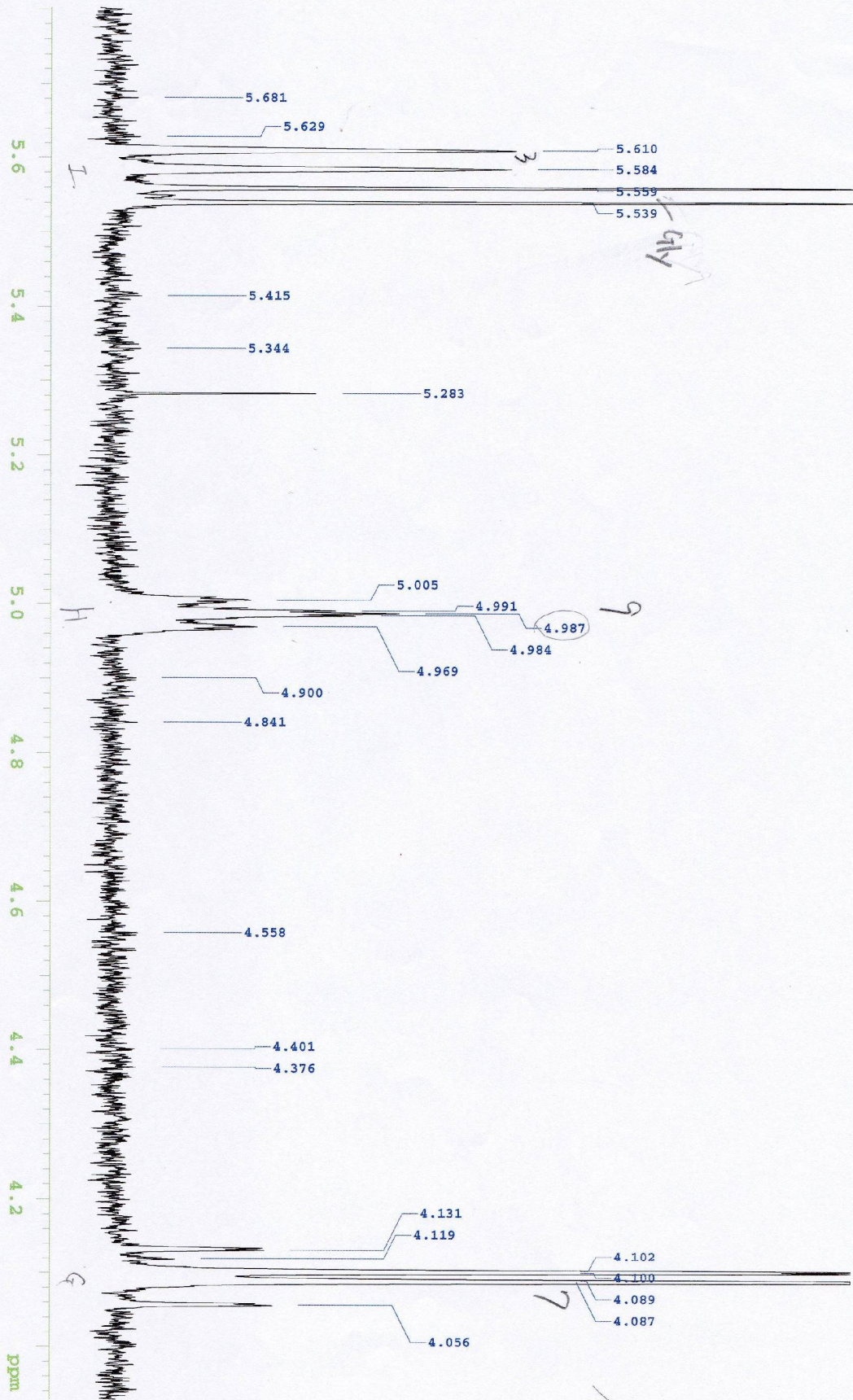
Figure S25 ^1H NMR spectra of 1a in CDCl_3



S-2



2-3



S-4

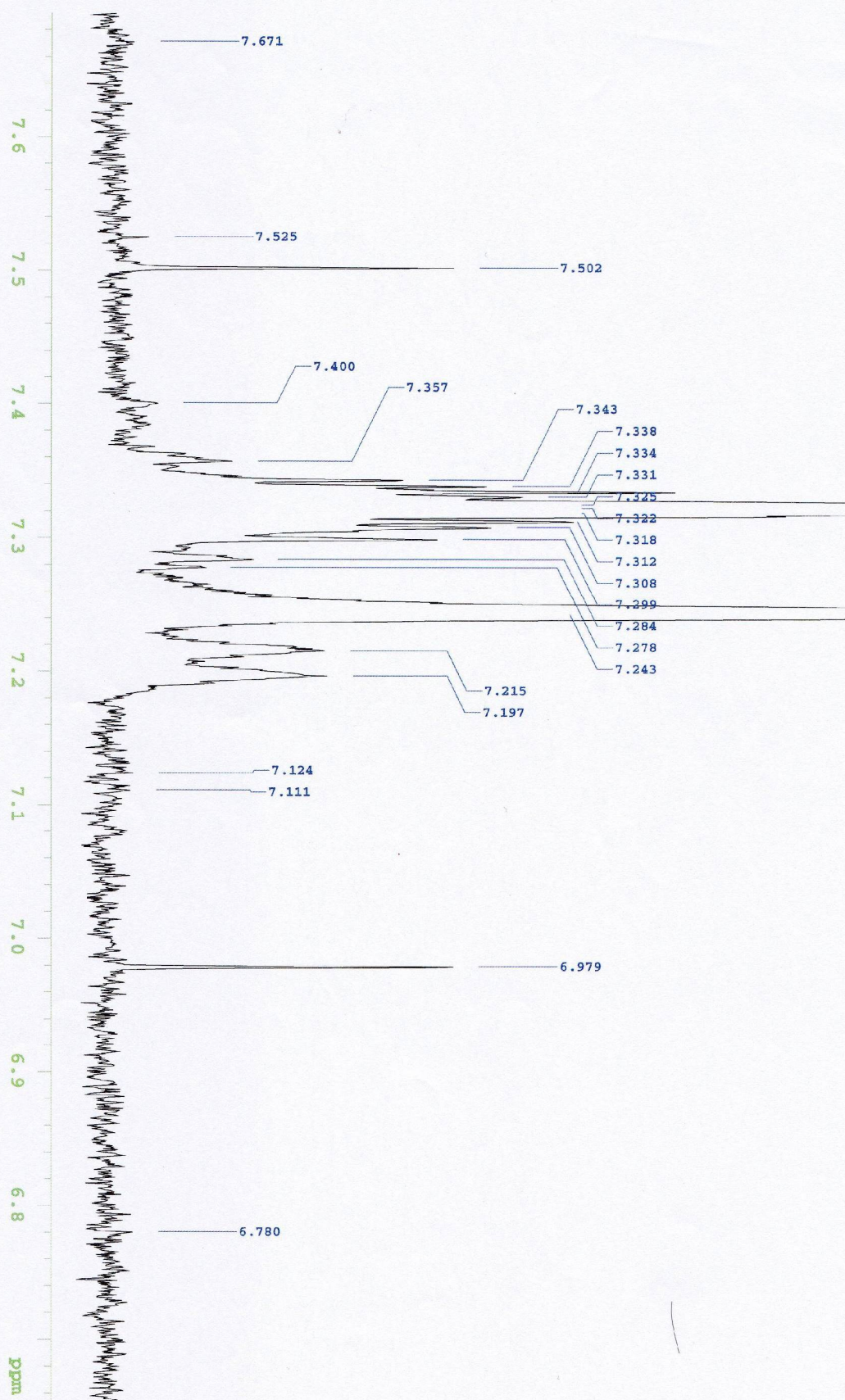
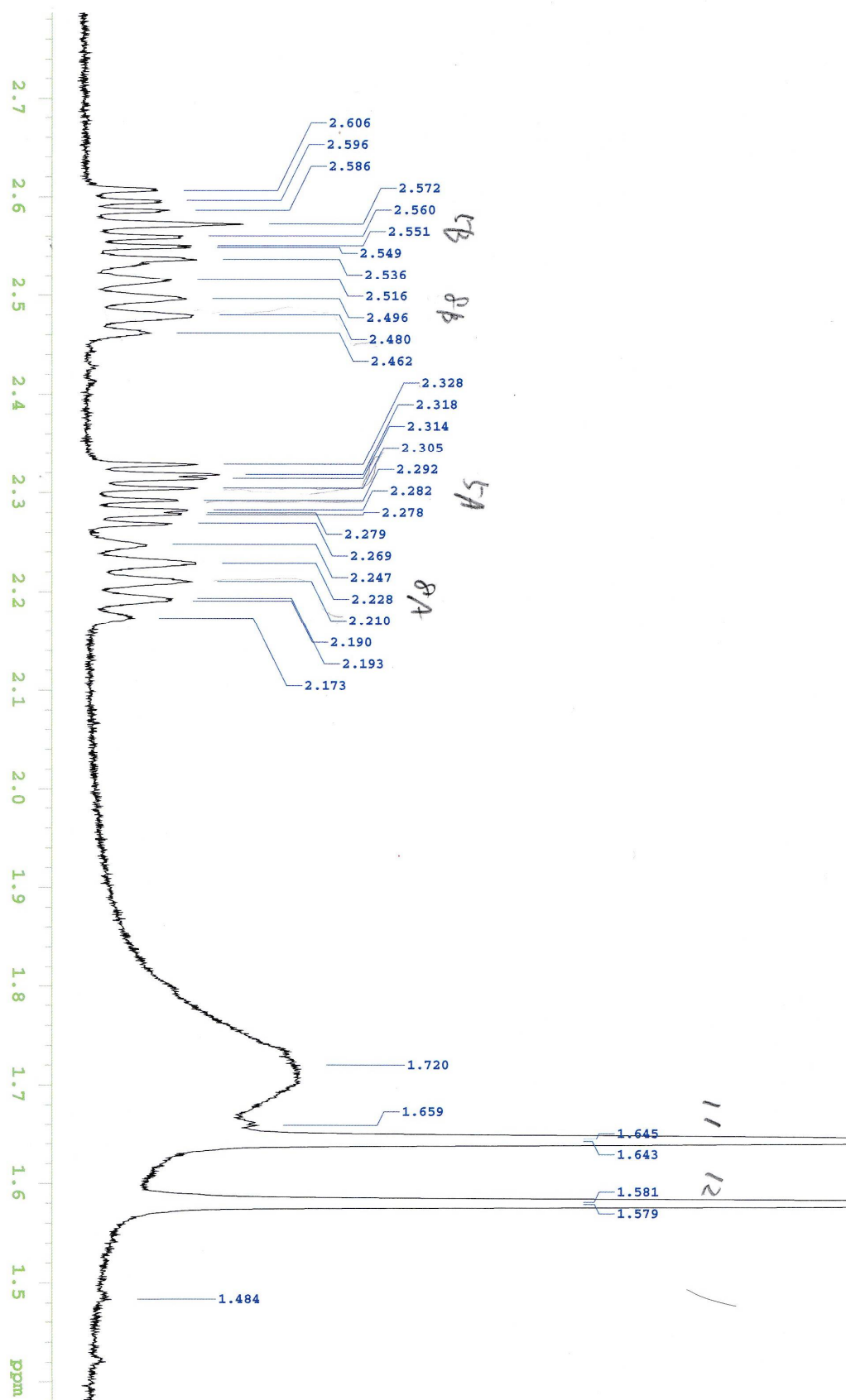
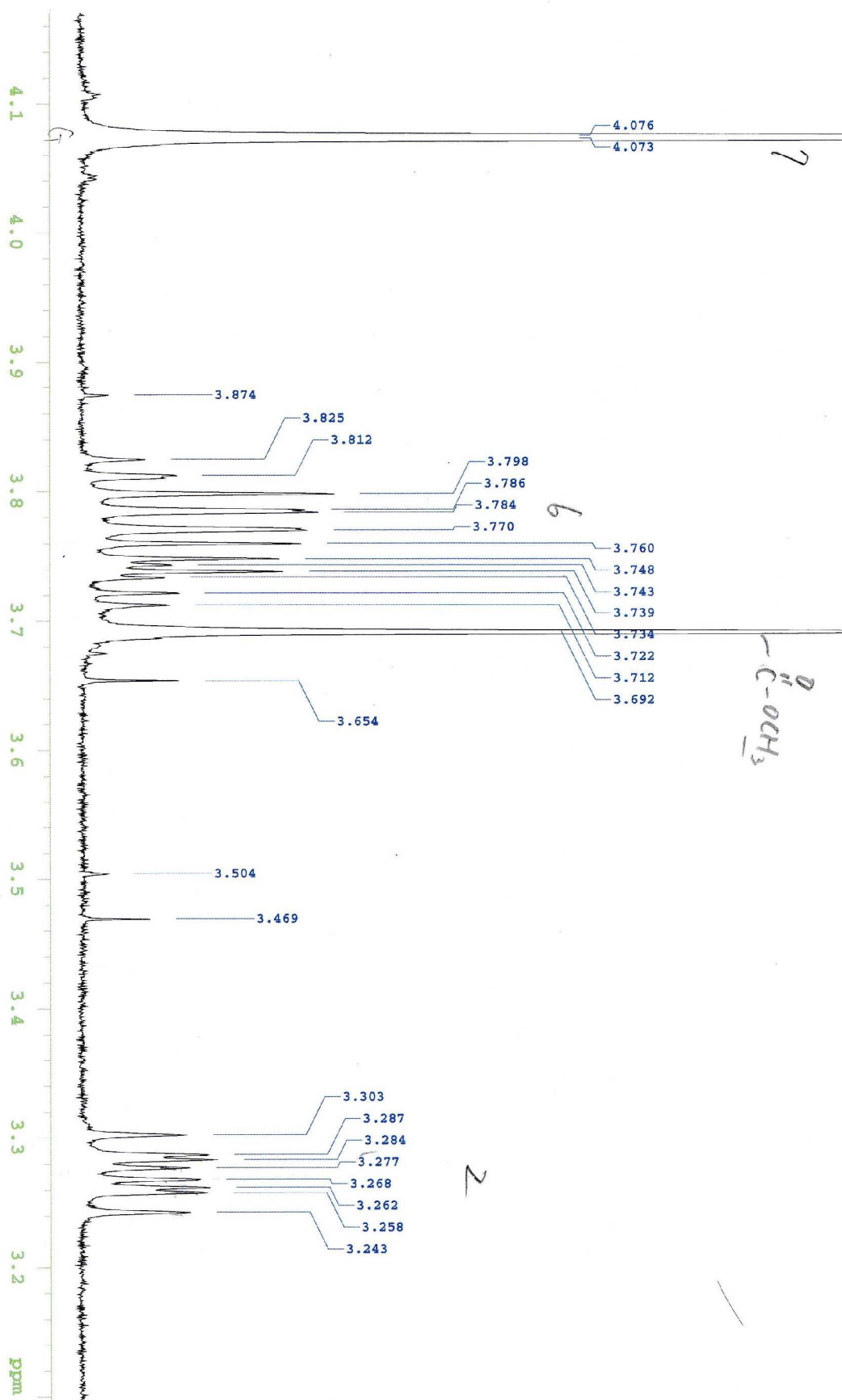


Figure S26 ^1H NMR spectra of 2a in CDCl_3



R-1

R-2



R-3

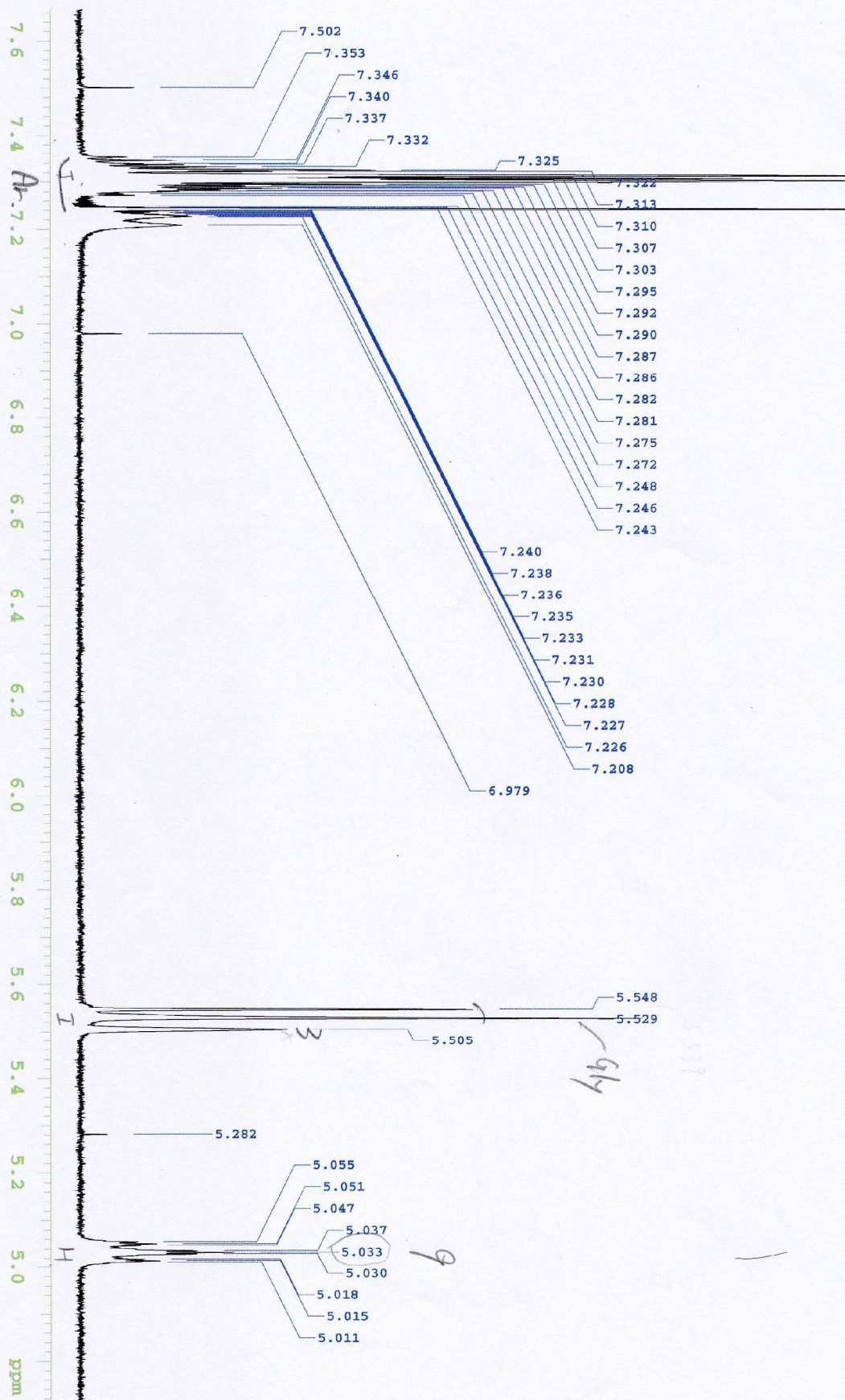


Figure S27 ^1H NMR spectra of 2b in CDCl_3

