1 Article

2 Gel dosimetry with radio-fluorogenic coumarin

Peter A. Sandwall ^{1,*}, Brandt P. Bastow ², Henry B. Spitz ², Howard R. Elson ², Michael Lamba ²,
 William B. Connick ² and Henry Fenichel ²

- 5 ¹ OhioHealth; Mansfield, Ohio
- 6 ² University of Cincinnati; Cincinnati, Ohio
- 7 * Correspondence: pasandwall@gmail.com; Tel.: +1-513-307-8156
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9 Abstract: In radiotherapy, accurate deposition of energy to the targeted volume is vital to ensure 10 effective treatment. Gel dosimeters are attractive detection systems, as tissue substitutes with 11 potential to yield three-dimensional dose distributions. Radio-fluorogenesis is creation fluorescent 12 chemical products in response to energy deposition from high-energy radiation. This report shares 13 studies of a radio-fluorogenic gel dosimetry system, gelatin with coumarin-3-carboxlyic acid 14 (C3CA), for the quantification of imparted energy. Aqueous solutions exposed to ionizing radiation 15 result in the production of hydroxyl free radicals through water radiolysis. Interactions between 16 hydroxyl free radicals and coumarin-3-carboxylic acid produce a fluorescent product. 17 7-hydroxy-coumarin-3-carboxylic acid has a blue (445 nm) emission, following UV to near UV 18 (365-405 nm) excitation. Effects of C3CA concentration and pH buffers were investigated for this 19 system. The response of the system was explored with respect to strength, type, and exposure rate 20 of high-energy radiation. Results show a linear dose response relationship with a dose-rate 21 dependency and no energy or type dependencies. This report supports the utility of gelatin-C3CA 22 for phantom studies of radio-fluorogenic processes.

- 23 Keywords: gel dosimetry; radiation dosimetry; radio-fluorogenic gel, luminescent dosimetry
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25 1. Introduction

26 Advancements in radiation therapy technology have supported study of tissue-equivalent gels 27 containing active chemical sensors for the measurement of absorbed dose of radiation. Gel 28 dosimeters have radiological properties similar to biological tissue and are suitable substitutes with 29 the potential to resolve three-dimensional dose distributions. The development of gel dosimeters 30 was dormant for many years, but has recently been developing at a rapid pace. The first reported use 31 of a gel dosimeter was in 1950 with the colorimetric dye methylene blue [1]. Other early 32 investigators explored chloral hydrate and trichloroethylene in agar [2]. Gelatin with ferricyanide, 33 Fricke-type, gel dosimeters were first studied using colorimetric methods, and later magnetic 34 resonance (MR) imaging [3-5]. Further developments introduced polymer and leuco-dye systems 35 [6-7]. Recently, a radio-fluorogenic polymer system has been introduced [8]. Each of the current gel 36 dosimeters have their own limitations such as rapid diffusion of chemical products with Fricke-type, 37 toxicicty of with polymer systems, intricate fabrication methods with leuco-dyes, and the 38 water-insolubility of radio-fluorogenic polymers [9]. The hunt for the ideal sensor element and gel 39 substrate is ongoing.

Two of the most common gel substrates are agarose and gelatin. Gelatin is derived from bovine or porcine collagen; primary element of skin, bone, and connective tissue. Agarose is a polysaccharide isolated from agar with highest gelling potential; agar is derived from seaweed. Gelatin and agarose are both capable of creating hydrogels with low percentages of gelling agent. However, agarose is opaque and induces light scattering, while gelatin is relatively translucent. The opacity of agar makes it less than ideal for optical analysis.³² Clarity and transparency of gelatin is

46 strongly dependent on raw material history, purity, and preparation. Commercial gelatin consists of 47 tropocollagen rods in the order of 300 nm in length with 1.5 nm diameter [10]. Raw material is 48 processed with acid or base solutions yielding "Type A" (hydrogen chloride) or "Type B" (sodium 49 hydroxide). Type A is denser than type B with a greater intrinsic viscosity [11]. Gelation speed also 50 affects rigidity with structure a function of formation temperature, slow gelatin yields increased 51 organization and orientation of chain elements with greater lateral bonding, this results in the 52 formation of fine well-ordered lattices [12]. Additionally, gelation is not susceptible to ionic effects 53 [10]. Derived from biological tissue with well understood mechanisms of gelation, gelatin is an 54 attractive substrate for exploration of optically active sensors.

55 Radio-fluorogenic sensors are chemical elements that allow for dosimetry, quantification of 56 energy deposition from of ionizing radiation, through measurement of molecular fluorescence. 57 Fluorescent detection methods are particularly promising due to their ability to form selective 58 high-resolution images. Initially reported by Day and Stein in 1949, fluorescence spectroscopy can be 59 used to determine absorbed dose in aqueous solutions of aromatic compounds [13-15]. Ionizing 60 radiation initiates radiolysis of water, yielding hydroxyl free radicals that hydroxylate aromatic 61 compounds via electrophilic substitution. Numerous aromatic compounds are recognized as 62 radio-fluorogenic, with hydroxylation producing fluorescent products. The first fluorescent sensor 63 investigated for radiation dosimetry was aqueous benzoic acid [15]. Other potential sensors are 64 terephthalic, trimesic, and pyromellitic acid [16-18]. Each improved the yield of fluorescent products 65 by restraining positions for substitution. However, each of those compounds possesses excitation 66 wavelengths unsuitable for a gel substrate. Rayleigh scattering is wavelength dependent, 67 proportional to $1/\lambda^4$, resulting in rapid reduction of transmission for shorter wavelengths of light. 68 Organic gels are naturally turbid due to their macromolecular nature, thus it is preferable to use 69 longer excitation wavelengths with greater potential for penetration. Fluorescence of aromatic 70 compounds is due to their conjugated system of alternating single and double-bonds; overlapping 71 pi-orbitals allow for delocalization of electrons. Larger conjugated systems require less energy for 72 excitation [19]. Selection of a multi-cyclic radio-fluorogenic sensor would provide a more attractive 73 fluorescent product; ideally with excitation from visible light. Multi-cyclic coumarin-3-carboxylic 74 acid (C3CA) is a sensor candidate.

Aqueous C3CA has been identified as chemical dosimeter for application to radiotherapy with favorable traits including linear dose response, reproducibility, and long-term stability [20]. The radio-fluorogenic mechanism of C3CA has been studied within aqueous solution [21]. Positive features of C3CA include high solubility in aqueous solutions, simple organic composition, and favorable excitation and emission spectra. C3CA reacts with hydroxyl radicals to yield the fluorescent product, 7-hydroxycoumarin-3-carboxylic acid (7HOC3CA), Figure 1.

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Figure 1. Hydroxyl radicals react with C3CA to yield 7HOC3CA through hydrogen abstraction,
 transfer, and substitution.

The present investigation explored C3CA in gelatin as a potential radio-fluorogenic detector. Concentration effects of C3CA were studied and the influence of pH buffers was investigated with

respect to relative fluorescent yield. Ionizing radiation response was examined subject to dose, rate,energy, and type for megavoltage electron and photon energies.

89 2. Materials and Methods

Solutions were readied with water from EASYpure water purification system (Barnstead International). Reagents were procured from Fisher Scientific (Baltimore, MD): 98% C3CA, C10H6O4 (Acros Organics, Baltimore, MD) and 99% 7HOC3CA, C10H6O5, MW 206.16 (Infodine Chemical Company; Hillsborough, NJ), sodium bicarbonate, sodium hydroxide, phosphate buffered saline, and food grade porcine type A gelatin (bloom strength 260, pH 5, and viscosity 40). Aliquots were separated, irradiated, and analyzed. Samples were stored at low temperature (5° C) to inhibit microbial growth.

97 To allow for dispersion, gelatin was 'wet,' placed in a beaker to soak with half the total volume 98 of water for 20 minutes. C3CA was brought into solution by boiling a small volume in a separate 99 beaker. After sufficient 'wetting', aqueous C3CA solution was added with the remaining portion of 100 water and temperature of gel solution raised to 35° C; care was taken to ensure the temperature 101 remained below 40° C to prevent denaturation. Gel was maintained at 35° C for 90 minutes, or until 102 optically clear and free of visible colloidal structures. The solution was then removed from heat and 103 pipetted into poly-methyl-methacrylate (PMMA) cuvettes. Gels were left to cool overnight at 104 ambient temperature. For initial pH buffer studies, 7% gelatin solutions were made with 7HOC3CA 105 to mimic the radio-fluorogenic product. Sodium bicarbonate/hydroxide and phosphate buffered 106 saline (PBS) solutions were prepared with 0.9 mM C3CA and 0.1 mM 7HOC3CA. For concentration 107 and dose response studies, 1 mM, 5 mM, 10 mM, and 20 mM C3CA solutions were prepared with 108 the sodium bicarbonate/hydroxide buffer.

109 Irradiations were conducted with a C-series high-energy medical linear accelerator (linac) 110 (Varian Medical Systems; Palo Alto, CA), with two megavoltage (MV) photon and five electron 111 energies; 6 and 23 and 6, 9, 12, 15, and 20 MeV. Irradiations were conducted with a polystyrene 112 phantom containing a void for 4 cuvettes; the phantom was designed expressly to provide geometry 113 favorable for establishment of electronic equilibrium. A computed tomography (CT) scan was 114 carried out on the phantom, images were imported into Eclipse treatment planning system (Varian 115 Medical Systems; Palo Alto, CA), and nominal dose calculated.

Instrumental analysis was conducted with a Cary Eclipse fluorescence spectrophotometer (Varian, Inc.; Pal Alto, California). Excitation and emission slit widths were set to 5 nm, emission scans were performed and peak emission values recorded and plotted. The dose response curve was created by plotting intensity of 445nm emission versus nominal dose.

120 **3. Results**

121 3.1. pH Response

The influence of pH buffers on fluorescent response was examined. Several solutions yielded various pH's; deionized water (pH 6.0), phosphate buffered saline (pH 7.4), and sodium hydroxide with sodium bicarbonate (pH 10). Results show positive correlation between pH and quantum yield. A spectral shift of the excitation maxima was also demonstrated. Specifically, peak excitation shifted from 365nm in normal (pH 7) solution to 405 nm in basic (pH = 10) solution, Figure 2.



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128Figure 2. Spectral curves of excitation and emission for solutions of 7% gelatin with 0.9mM C3CA129and 0.1mM 7HOC3CA.

130 3.2. C3CA Concentration

With sodium hydroxide with sodium bicarbonate solutions, varying the concentration of C3CA
 revealed a stronger response with concentrations 5 mM and greater, Figure 3. The peak normalized

133 response, fluorescent intensity divided by dose, produced a decreasing exponential curve, Figure 4.

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Figure 3. Nominal dose plotted against intensity for concentrations of C3CA in 7% gelatin. Error bars

137 represent relative 5% error.



139 **Figure 4.** Nominal dose plotted versus normalized response. Error bars represent relative 5% error.

140 3.3. Dose Response

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141 Dose response was studied with ionizing radiation with respect to type, rate, and energy. 142 Relative response was measured with respect to 445nm emissions and plotted against nominal dose, 143 Figure 5. Repeated measures, using four samples for each data point, demonstrated relative error 144 less than 1%. A linear response was observed in the range investigated (R > 0.99), independent of 145 type (photon or electron) and energy (9 MeV, 6 MV, and 23 MV), Figure 6. A strong negative 146 correlation (R > 0.99) with dose rate was observed; the intensity of normalized fluorescent response 147 decreased with increasing dose rate, Figure 7. Using a definition of three times the standard 148 deviation of the background, the minimum detectable amount (MDA) was extrapolated from 9MeV 149 electron data and estimated to be 1.5Gy, Figure 8.



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Figure 5. Nominal dose plotted against intensity, demonstrating a linear response (R>.99). Linear equations inlaid, error bars represent 5% relative error.



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Figure 6. Normalized response plotted against nominal dose for 23 MV, 6 MV, and 9 MeV beams, error bars represent 5% relative error.



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158 4. Discussion

159 The basic solutions (pH 10) were observed to double emission intensity and shift the peak 160 excitation wavelength from 365 nm to 405 nm. Transition between excited and ground states, the 161 energy gap, is known to be influenced by the micro-environment through molecular motion, 162 collision, rotational and translational diffusion, and formation of complexes. Smaller quantum yields are observed with large energy gaps due to availability of alternative relaxation pathways. The 163 164 observed increase in quantum yield is consistent with previous studies in aqueous solution; 165 however, greater than previously observed (385 nm) [22]. The increased spectral shift may be due to 166 interactions with gelatin, additional study could clarify these effects.

167 The dose response was notably more pronounced for concentrations of C3CA above 5 mM. The 168 normalized response curves of various concentrations of C3CA suggest saturation, diminishing 169 population of radio-fluorogenic reactants in the dose range studied. Future work investigating 170 absolute yields and a larger range of doses would be beneficial.

171 Normalized data demonstrate an independent linear response with respect to dose, energy, and 172 type of ionizing radiation (electron and photon). With respect to type, an independent response is 173 expected since photon dose deposition is predominately by delta rays, secondary electrons. A dose 174 rate dependency was observed, consistent with other findings [20]. Previously suggested to be due

to metallic impurities in C3CA and alleviated by successive distillations. The MDA was estimated tobe 1.5 Gy, this value should be determined rigorously, by study of an expanded dose range.

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178 5. Conclusions

179 Optical imaging of biomarkers is an active area of study with C3CA a recognized radiation 180 activated sensor for fluorescent imaging [18]. Investigators have explored the use of coumarin 181 attached to peptide ligands, designed for DNA binding, with potential for assessment of radiological 182 response [23]. Other work is currently studying the application of fluorescent labels for radiometric 183 assay [24, 25]. Advances in the fabrication of gelatin based phantom materials with 3D printability 184 make further study particularly attractive [26]. Further study radio-fluorescent sensors in a gelatin 185 matrix would help advance these prospective *in vivo* applications.

The potential of C3CA in gelatin for determination of spatial dose distributions has been demonstrated in a separate report [27]. The use of planar laser induced fluorescence (PLIF) has been shown as a method to yield high-resolution three-dimensional images [28]. This method of image collection and analysis has been recognized and is currently being explored with polymer based radio-fluorogenic gel [29]. However, it is the author's belief that the greatest depth of penetration and finest imaging resolution will be obtained by applying methods of two-photon excitation microscopy.

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203 References

205	Kele	nences
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205	1.	Day, M. J., and Stein, G. Chemical effects of ionizing radiation in some gels. <i>Nature</i> , 1950 , 166(4212), 146.
206	2.	Andrews, H. L., Murphy, R. E., and LeBrun, E. J. Gel dosimeter for depth-dose measurements. <i>Review of</i>
207		scientific instruments, 1957 , 28(5), 329-332.
208	3.	Fricke, H., and Hart, E. J. The oxidation of the ferrocyanide, arsenite and selenite ions by the irradiation of
209		their aqueous solutions with x-rays. The Journal of Chemical Physics, 1935, 3(9), 596-596.
210	4.	Stein, G. and Tomkiewicz, M., Radiation chemistry of gelatin gels containing ferricyanide, radiation
211		research, 1970 , 43(1), 25-33.
212	5.	Gore, J. C., and Kang, Y. S. Measurement of radiation dose distributions by nuclear magnetic resonance
213		(NMR) imaging. Physics in Medicine & Biology, 1984, 29(10), 1189.
214	6.	Maryanski, M. J., Ibbott, G. S., Eastman, P., Schulz, R. J., and Gore, J. C. Radiation therapy dosimetry using
215		magnetic resonance imaging of polymer gels. <i>Medical physics</i> , 1996 , 23(5), 699-705.
216	7.	Adamovics, J., and Maryanski, M. J. Characterisation of PRESAGE TM : A new 3-D radiochromic solid
217		polymer dosemeter for ionising radiation. Radiation protection dosimetry, 2006, 120(1-4), 107-112.
218	8.	Warman, J. M., De Haas, M. P., and Luthjens, L. H. High-energy radiation monitoring based on
219		radio-fluorogenic co-polymerization. I: small volume in situ probe. Physics in Medicine & Biology, 2009,
220		54(10), 3185.
221	9.	Doran, S. J. The history and principles of chemical dosimetry for 3-D radiation fields: Gels, polymers and
222		plastics. Applied Radiation and Isotopes, 2009, 67(3), 393-398.
223	10.	Veis, A. The Macromolecular Chemistry of Gelatin; Academic Press, New York, NY, 1964.
224	11.	Djagny, K.B., Wang, Z. and Xu, S. Gelatin: a valuable protein for food and pharmaceutical industries:
225		Review, Critical reviews in food science and nutrition, 2001, 41(6), 481-492.
226	12.	Roussenova, M., Enrione, J., Diaz-Calderon, P., Taylor, A. J., Ubbink, J., and Alam, M. A. A nanostructural
227		investigation of glassy gelatin oligomers: molecular organization and interactions with low molecular
228		weight diluents. New Journal of Physics, 2012, 14(3), 035016.
229	13.	Day, M.J. and Stein, G. Chemical Measurement of Ionizing Radiations, Nature, 1949, 164(1), 671-672.
230	14.	Stein, G. and Weiss, J. Detection of Free Hydroxyl Radicals by Hydroxylation of Aromatic Compounds,
231		<i>Nature</i> , 1950 , 166(4235), 1104-1105.
232	15.	Armstrong, W. A. and Grant, D. W. A highly sensitive chemical dosimeter for ionizing radiation. Nature,
233		1958 , 182(4637), 747-747.
234	16.	Barreto, J. C., Smith, G. S., Strobel, N. H., McQuillin, P. A., and Miller, T. A. Terephthalic acid: a dosimeter
235		for the detection of hydroxyl radicals in vitro. <i>Life sciences</i> , 1994 , 56(4), PL89-PL96.
236	17.	Matthews, R. W. Aqueous chemical dosimetry, The International Journal of Applied Radiation and Isotopes,
237		1982 , 33(11), 1159-1170.
238	18.	Nadrowitz, R., Coray, A., Boehringer, T., Dunst, J., and Rades, D. (2012). A liquid fluorescence dosimeter
239		for proton dosimetry. 2012, Physics in Medicine & Biology, 57(5), 1325.
240	19.	Lakowicz, J. R. Principles of Fluorescence Spectroscopy, 3rd ed.; Springer, New York, NY, 2006
241	20.	Collins, A.K., Makrigiorgos, G.M., and Svensson, G.K. Coumarin chemical dosimeter for radiation
242		therapy. <i>Medical physics</i> , 1994 , 21(11), 1741-1747.
243	21.	Yamashita, S., Baldacchino, G., Maeyama, T., Taguchi, M., Muroya, Y., Lin, M., Kimura, A., Murakami, T.

- and Katsumura, Y. Mechanism of radiation-induced reactions in aqueous solution of
 coumarin-3-carboxylic acid: effects of concentration, gas and additive on fluorescent product yield. *Free radical research*, 2012, 46(7), 861-871.
- 247 22. Dai, X., Rollin, E., Bellerive, A., Hargrove, C., Sinclair, D., Mifflin, C., & Zhang, F. Wavelength shifters for water Cherenkov detectors. *Nuclear Instruments and Methods in Physics Research Section A: Accelerators, Spectrometers, Detectors and Associated Equipment*, 2008, 589(2), 290-295.
- 250 23. Perry, C. C., Tang, V. J., Konigsfeld, K. M., Aguilera, J. A., and Milligan, J. R. Use of a coumarin-labeled
 251 hexa-arginine peptide as a fluorescent hydroxyl radical probe in a nanoparticulate plasmid DNA
 252 condensate. *The Journal of Physical Chemistry B*, 2011, 115(32), 9889-9897.
- 253 24. Gallina ME, Kim TJ, Vasquez J, Tuerkcan S, Abbyad P, and Pratx G. Single-cell analysis of radiotracers' uptake by fluorescence microscopy: direct and droplet approach. *Imaging, Manipulation, and Analysis of Biomolecules, Cells, and Tissues*, 2017, 10068, 100680Y. International Society for Optics and Photonics.

- 25. Gallina ME, Kim TJ, Shelor M, Vasquez J, Mongersun A, Kim M, Tang SK, Abbyad P, and Pratx G. Toward
 a Droplet-Based Single-Cell Radiometric Assay. *Analytical chemistry* 2017, 89(12):6472-81.
- 26. Dahal E, Badal A, Zidan A, Alayoubi A, Hagio T, Glick SJ, Badano A, and Ghammraoui B. Stable
 gelatin-based phantom materials with tunable x-ray attenuation properties and 3D printability for x-ray
 imaging. *Physics in medicine and biology.* 2018 Apr 10.
- 261 27. Sandwall, P. A., Spitz, H. B., Elson, H. R., Lamba, M. A., Connick, W. B., & Fenichel, H. Measuring the photon depth dose distribution produced by a medical linear accelerator in a water-equivalent radio-fluorogenic gel. *Journal of Radioanalytical and Nuclear Chemistry*, 2016, 307(3), 2505-2508.
- 264 28. Sandwall, P., Spitz, H., Elson, H., Lamba, M., Connick, W., & Fenichel, H. Radio-fluorogenic dosimetry with violet diode laser-induced fluorescence. In *Medical Imaging: Physics of Medical Imaging* 2014 (Vol. 9033, p. 90333Y). International Society for Optics and Photonics.
- 267 29. Yao T, Gasparini A, De Haas MP, Luthjens LH, Denkova AG, and Warman JM. A tomographic UV-sheet
 268 scanning technique for producing 3D fluorescence images of x-ray beams in a radio-fluorogenic gel.
 269 *Biomedical Physics & Engineering Express.* 2017, 3(2):027004.