

Review

# Amino Acids as Building Blocks for Carbonic Anhydrase Inhibitors

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**Abstract:** Carbonic Anhydrases (CAs) are a superfamily of metalloenzymes widespread in all life kingdoms, classified into seven genetically different families ( $\alpha$ - $\theta$ ). These enzymes catalyse the reversible hydration of carbonic anhydride ( $\text{CO}_2$ ), generating bicarbonate ( $\text{HCO}_3^-$ ) and protons ( $\text{H}^+$ ). Fifteen isoforms of human CA (hCA I-XV) have been isolated, their presence being fundamental for the regulation of many physiological processes. In addition, overexpression of some isoforms has been associated with the outbreak or the progression of several diseases. For this reason, for a long time CA inhibitors (CAIs) are used in the control of glaucoma and as diuretics. Furthermore, the search for new potential CAIs for other pharmacological applications is a very active field. Amino acids constitute the smallest fundamental monomers of protein and, due to their useful bivalent chemical properties, are widely used in organic chemistry. Both proteinogenic and non-proteinogenic amino acids have been extensively used to synthesize CAIs. This article provides an overview of the different strategies that have been used to design new CAIs containing amino acids, and how these bivalent molecules influence the properties of the inhibitors.

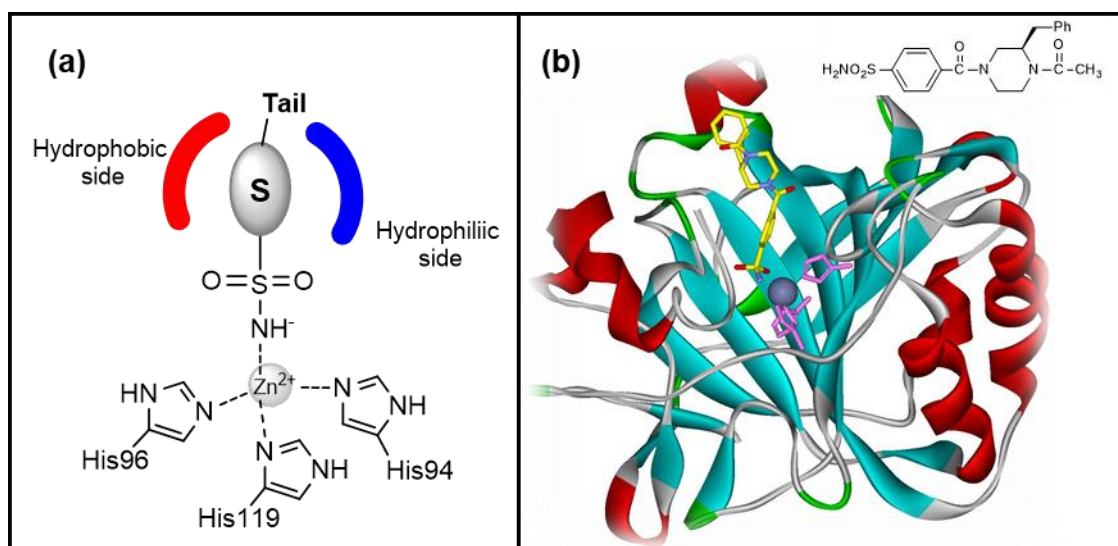
**Keywords:** carbonic anhydrase, enzyme inhibition, metalloenzymes, amino acid, glaucoma, tumors

## 1. Introduction

The interconversion between  $\text{CO}_2$  and  $\text{HCO}_3^-$  is a fundamental process for the successful flow of biochemical processes in all living cells [1]. The metabolic conversion is ensured by the hydration of carbonic anhydride, that leads to the corresponding soluble ion, bicarbonate, and the release of a proton [2,3]. Due to the high number of physiological process in which these chemical species are essential, the optimal balance between them is vital for all life forms [2,4–7]. This equilibrium is regulated by a superfamily of metalloenzyme, the Carbonic Anhydrases (CAs, EC 4.2.1.1) [1]. By accelerating this normally slow reaction, CAs fulfil the metabolic needs connected to  $\text{CO}_2$ /bicarbonate and protons [8]. In fact, CAs are among the most effective catalysts known in nature [3] as their turnover number ( $k_{\text{cat}}$ ) can reach the value of  $10^6 \text{ s}^{-1}$  [9]. Seven genetically different families of these ubiquitous enzymes were identified ( $\alpha$ - $\eta$ ) [2]; all of them possess a bivalent metal ion fundamental for the catalysis, as the apoenzyme is devoid of activity [3]. Mammalian CAs belongs to the  $\alpha$ -family and they are characterized by the presence of a bivalent zinc ion within the active site [1]. This  $\text{Zn}^{2+}$  coordinates a water molecule, or a hydroxide ion in the activated form of the enzyme, making the hydration of  $\text{CO}_2$  a fast process [1,2,10].

Human carbonic anhydrases (hCAs) are widespread in the organism, varying for tissue distribution and subcellular localization. Fifteen  $\alpha$ -isozymes (hCA I-XV) were isolated and characterized so far, but only twelve of them are catalytically active [4,5,11]. hCAs are involved in pH and  $\text{CO}_2$  homeostasis but also in the regulation of many crucial physiological processes, such as gluconeogenesis, lipogenesis or electrolyte secretion in a variety of tissues and organs [2,4–6]. hCA I and II are the widest expressed isoforms and, together with hCA IV, were identified in the anterior chamber of the eye, being responsible in this organ for the bicarbonate production in this organ, the main constituent of aqueous humour [5,12]. hCA IX and XII can be defined as tumor-associated proteins, due to their massive expression in many hypoxic cancers [4,5,13,14]. These isozymes are

transmembrane proteins, with an extracellular catalytic domain; they are fundamental for the survival of tumor cells under stressful conditions, due to their ability to generate a differential pH microenvironment, resulting in increased tumor growth [5,13,15,16]. While CA activators do not have for the moment any approved pharmacological application [17], CA inhibitors (CAIs) are commonly used in therapy, mainly as anti-glaucoma agents, anti-epileptics and diuretics, while other therapeutic applications are still under investigation (anti-cancers, anti-obesity agents, and others) [5,12,18–21].



**Figure 1.** (a) Schematization of the interaction between a generic sulfonamide and the Zn<sup>2+</sup> in the CA active site. (b) X-ray structure of (S)-4-(4-acetyl-3-benzylpiperazine-1-carbonyl)-benzenesulfonamide (with carbons in yellow, discussed in section 7) bounded in the active site of hCA I (PDB entry 6EVR) [79]. The sulfonamide group interacts with the Zn<sup>2+</sup> (dark grey sphere) which is coordinated by three His residues (in purple).

## 2. Carbonic Anhydrases Inhibitors (CAIs)

In the CAs active site the bivalent metal ion normally possesses a tetrahedral geometry, since it interacts with three amino acids moieties and coordinates the water molecule/hydroxide ion. In  $\alpha$ -CAs, the Zn<sup>2+</sup> ligands are three histidine residues (Figure 1) and, together with other key amino acids, they form a very particular and strictly conserved catalytic site within each family of carbonic anhydrases [1–3]. It is therefore possible to recognize two different regions in the cavity of the active site, a first half composed exclusively of hydrophobic amino acids and a second one where only hydrophilic residues are present [22], leading to a “bipolar” site architecture [2].

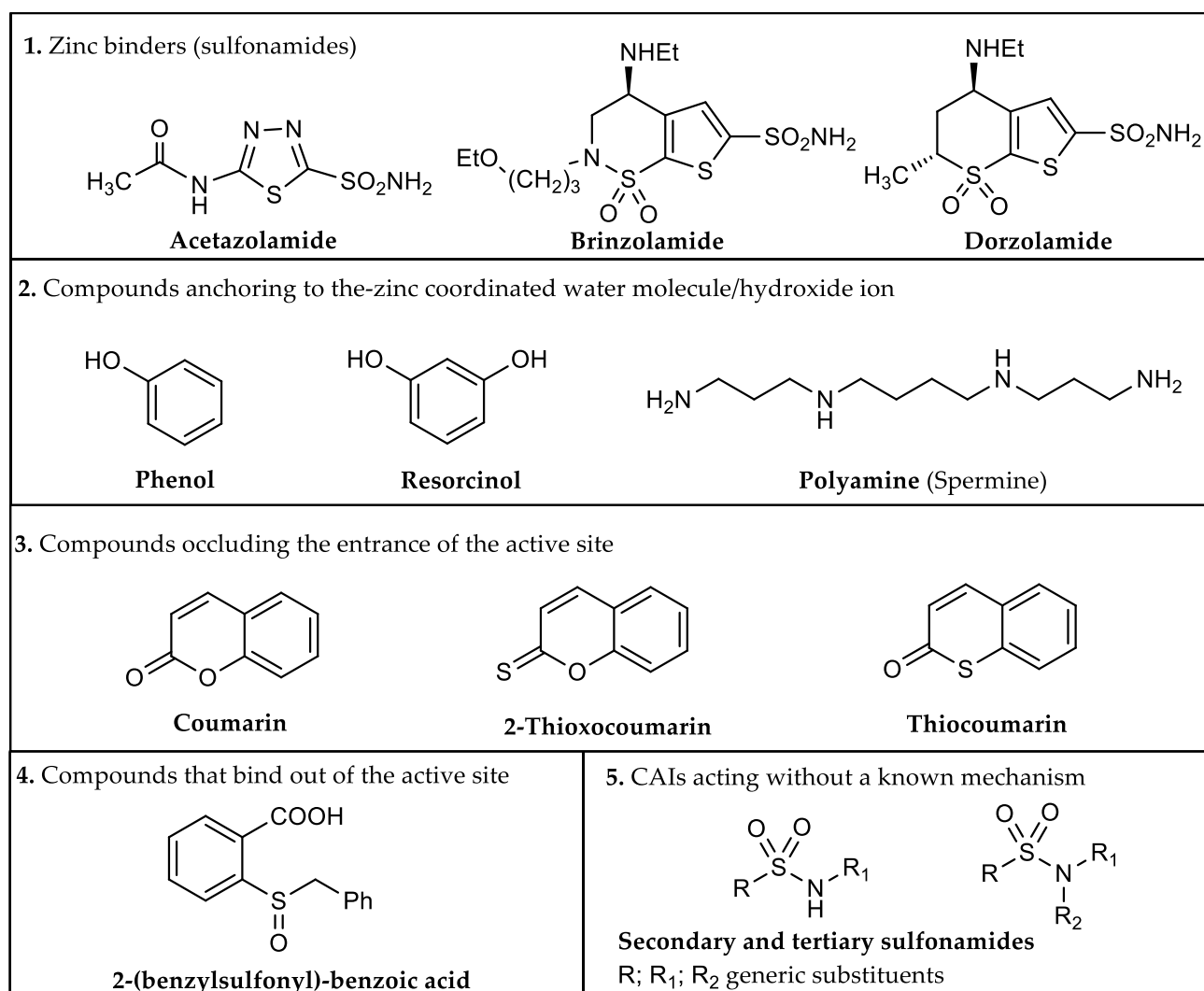
Depending on the mechanism of action, five different classes of CAIs are known (Figure 2) [21,23]:

1. Zinc binders, i.e. compounds that chelates the bivalent metal ion of the active site, interrupting the coordination of the water molecule/hydroxide ion and consequently the enzymatic activity [21,23,24]. The mechanism is schematized in figure 1A: the scaffold of these molecules (reported as “S”) may interact with one or both the halves of the active site, stabilizing the interaction with the ion in a tetrahedral geometry. This is the most important class of inhibitors, to which belongs sulfonamides and their isosteres (sulfamates or sulfamides), dithiocarbamates, hydroxamate, etc [3,23]. Sulfonamides are the most widely studied CAIs with at least 20 compounds in clinical use for decades [23]. Some examples (Acetazolamide, Brinzolamide and Dorzolamide) are shown in figure 2.

2. Compounds that anchor to the-zinc coordinated water molecule/hydroxide ion, such as phenols and polyamines [3,23,25].

3. Compounds occluding the entrance of the active site (coumarins and their isosteres) [3,23,26].

4. Compounds that bind out of the active site, such as 2-(benzylsulfonyl)benzoic acid [23,27].  
 5. CAIs acting without a known mechanism; such as secondary/tertiary sulfonamides, imatinib, etc [3,23].



**Figure 2:** The five classes of Carbonic Anhydrases Inhibitors

As we can notice in **figure 2**, the five types of inhibitors possess various structures; chemical differences in their pharmacophoric moieties result in different structure activity relationship. Despite this premise, a common problem of CAIs, and in particular of zinc binders, is the lack of selectivity for a specific isoform or the absence of a good water solubility. Many efforts have been done in the last decades to explore chemical modifications of the CAIs structures that could have a positive effect on these properties. In particular, the aim of this review is to provide an overview of the investigated applications of amino acid moieties in the resolution of these problems. This paper is focused on CAIs that carry amino acids or their derivatives in their structures, and how these molecules influence the properties of these inhibitors. Both proteinogenic and non-proteinogenic amino acids have been considered in this analysis and, depending on their synthetic use, five different strategies have been identified.

Amino acids are the primary building blocks of protein and, of the over 300 naturally occurring, 22 constitute the monomer units of these biological molecules [28]. Amino acids are widely used in organic chemistry, they constitute the smallest fundamental pieces in solid phase peptide synthesis but, due to their characteristics, they are also commonly used in solution synthesis [29]. Their properties range from acidic to basic due to the contemporary presence of a carboxyl and an amino

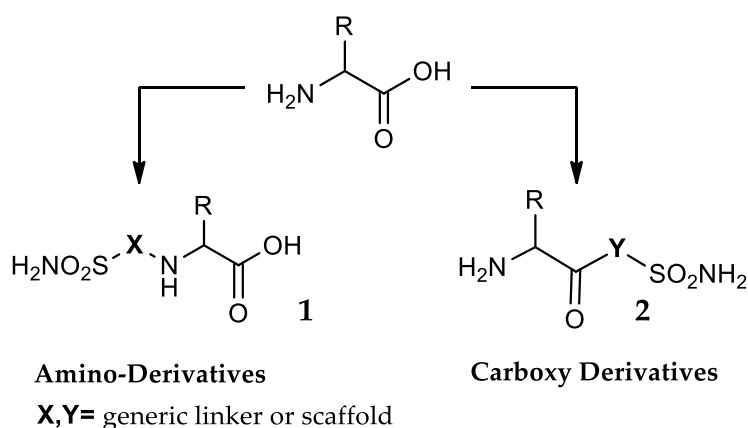
group that, in addition, allow the possibility to easily generate different kind of connections to link the amino acid to other molecules. Beside this, a number of amino acids have reactive groups in their side chains that can be chemically modified [28]. Furthermore, the use of natural  $\alpha$ -amino acids allows to obtain a broad range of derivatives, with the possibility, starting from enantiopure amino acids, to further define the stereochemistry of the side chain. Thanks to their ionizing properties that could be exploited to form salts, the carboxy and the amino moieties ensure also a good water solubility to these derivatives.

### 3-Amino Acyl as a water-solubilizing tail

Glaucoma is a group of optical neuropathies associated with progressive loss of visual field, leading to visual impairment and blindness. There is a general agreement that increased intraocular pressure (IOP) is the most important risk factor for the outbreak and progression of this disease [12,30]. It was demonstrated [31] that in glaucomatous patients the inhibition of CAs leads to a reduction of IOP and an amelioration of the symptoms. Due to the wide distribution of the different CA isozymes in the body, the administration of systemic inhibitors usually elicits undesired side effects [12,30,32]. For this reason, the treatment of glaucoma is mainly performed with topically administered CAIs [12,33]; two drugs, Brinzolamide and Dorzolamide (**Figure 2**), are available in the clinic. These two inhibitors often produce local side effects, such as ocular burning, stinging sensation, superficial punctate keratitis, blurred vision and reddening of the eye. These problems are mainly owed to the strong acidic pH of their solution, as they are both administered as hydrochloride salts [12,34].

In order to reduce these inconveniences, many efforts have been made for the development of new topically administered CAIs. Among others, a deeply investigated method is the attachment of water-solubilising tails to known effective CAIs. This method is particularly important for sulfonamides, very potent anti-glaucoma agents often endowed of a low water-solubility.

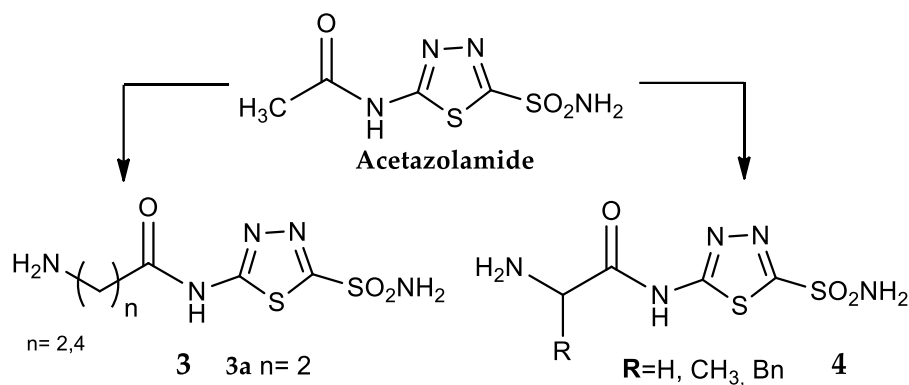
The use of amino acids for this purpose was investigated by many research groups, with a broad variety of approaches. The bivalent chemical nature of these synthons allows obtaining two possible series of products, the amino-substituted compounds **1** and the carboxy derivatives **2** (**Figure 3**). In **1**, the free carboxyl group can be treated with strong bases to give salts or can be further functionalized. Also compounds **2** can be endowed with good hydrophilic properties thanks to the free basic amino group: such compounds are ideal for ophthalmologic applications, since the water solutions of their salts with strong acids have a weakly acidic pH, preferred over those of alkaline pH [35].



**Figure 3.**

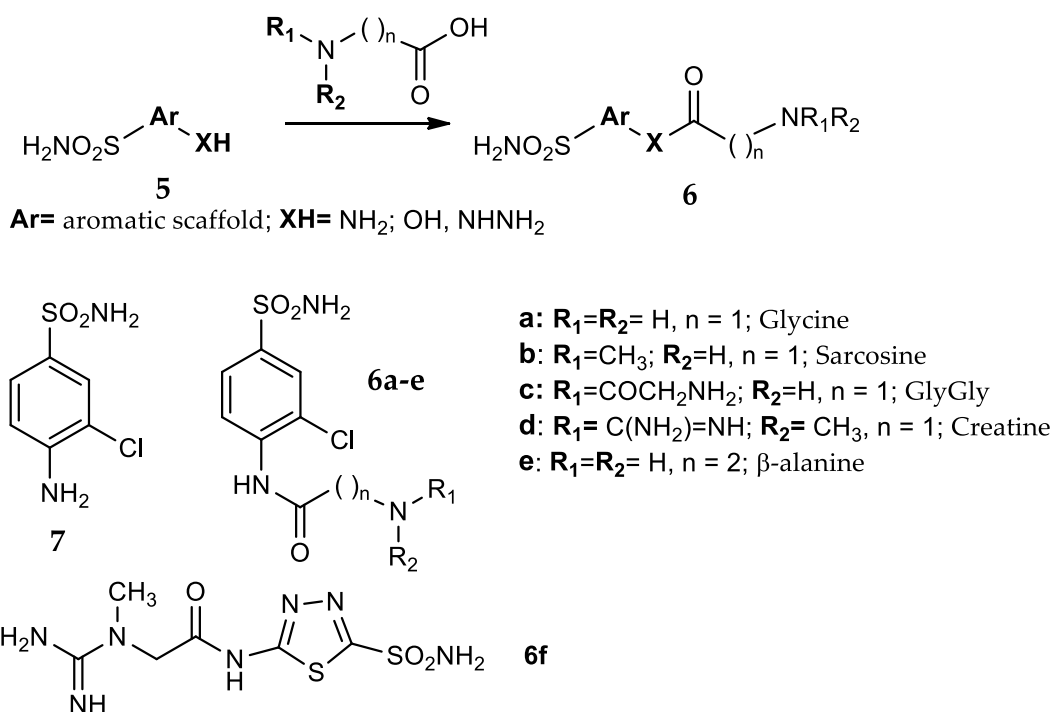
Antonaroli et al. [36], Blackburn's [37] group, and later Barboiu et al [38] reported some structural manipulation of Acetazolamide, where the N-acetyl residue was substituted with different aminoacids, giving compounds with general formula **3** and **4** (**figure 4**). The structural modification of this first-generation CA inhibitor led to interesting products: the salts of compounds **3** and **4**

showed a very good water solubility, associated with effective inhibitory activities against the enzyme. As example, the  $\beta$ -alanyl derivatives **3a** was three times more potent on hCA II and over a hundred times on hCA I than the reference drug Dorzolamide. Only against the isozymes hCA IV, this compound showed an activity three times lower than Dorzolamide.



**Figure 4.**

A similar approach was also investigated by Scozzafava et al in two separate works [35,39]. Twenty six different sulfonamides containing amino, imino, hydrazino or hydroxyl groups (general formula **5**, **figure 5**) reacted with the carboxy moiety of five different glycine derivatives (glycine, sarcosine, creatine, gly-gly and  $\beta$ -alanine). The extremely versatile nature of the carboxyl group allowed to generate a large library of new compounds (general formula **6**), differing not only in the structure of the two reagents, but also for the characteristics of the new formed carboxy-derived bonds (i.e. amidic, esteric and hydrazidic bonds).



**Figure 5.**

All the 130 obtained compounds **6** possess very good water solubility as salts of strong acids. They showed a wide range of inhibitory activities against three CA isozymes, hCA I, II and bCA IV. The addition of the aminoacyl/dipeptidyl moiety generally led to an increase of CA inhibitory



properties with respect to the corresponding parent sulfonamide. The five amino acid derivatives **6a-f** can be taken as example: they were from 3 to 18 times more potent than the precursor **7**. In addition, the most active compounds of the series were selected for in vivo studies. The topical application directly into the eye showed IOP-lowering effects both in normotensive and glaucomatous rabbits, a frequently used animal model of glaucoma [40,41]. As an example, the acetazolamide derivative **6f** was a more efficient and long-lasting IOP lowering agent compared to the reference drug dorzolamide.

It is also possible to enhance the basic nature of the free  $\text{NH}_2$  group by conversion into a guanidine moiety, as reported by Ceruso et al [42]. The carboxyl group of two amino acids,  $\text{N}\alpha$ -acetylysine and  $\gamma$ -aminobutyric acid, was connected to a benzyl or phenethylamine carrying a sulfonamide group, obtaining a first series of basic compounds (**8a** and **9a**, figure 6); subsequently the amino groups were treated with  $\text{N,N'$ -di-Boc- $\text{N''}$ -trifluoromethane-sulfonylguanidine, obtaining the guanidine derivatives **8b** and **9b**. This transformation enhanced the solubility of the hydrochloride salts but not their potency, since the guanidine derivatives showed  $\text{K}_i$  values almost in the same range of the amino-precursor on hCA I, hCA II and *Porphyromonas gingivalis*  $\gamma$ -CA. The same products were also tested against the two tumor-associated isoforms hCA IX and hCA XII [43]. Due to their transmembrane localization and their extracellular catalytic domain, the use of hydrophilic CAIs, endowed with poor cellular permeability, could in principle selectively target these isoforms. The effect of the terminal guanidine moiety on activity was relevant only on hCA XII for some GABA-derivatives: as example **9c** showed a  $\text{K}_i$  value almost nine times lower than the corresponding amino precursor.

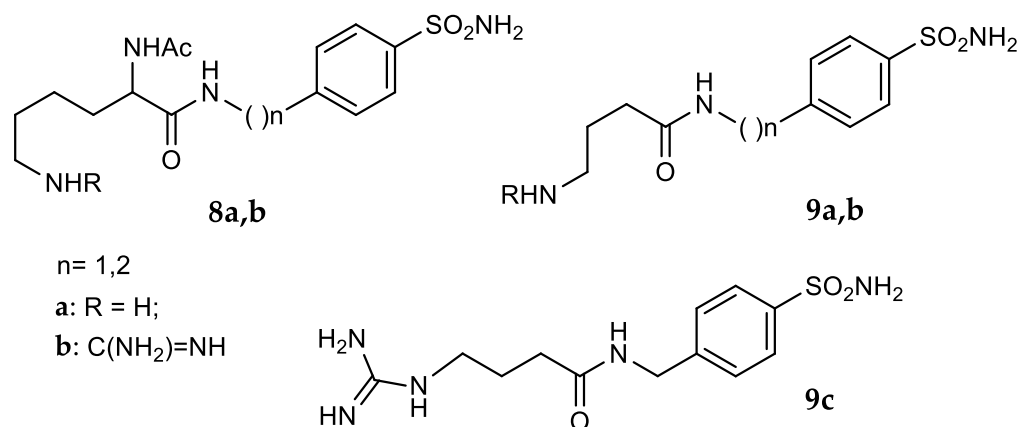


Figure 6.

The addition of the amino acid by means of the amino group, instead of the carboxy one, was investigated by Casini et. al in two separate, but closely related papers [34][44]. Two series of sulfonamides were synthesized by the reaction of 4-isothiocyanatobenzenesulfonamide **10a** or 4-isothiocyanatomethyl-benzenesulfonamide **10b** with 33 different amino acids and oligopeptides, forming a thioureido linkage between these two portions (general formula **11**, figure 7). Many newly obtained compounds possess very good water-solubility properties; the presence of at least one free carboxyl group allow the easily formation of sodium salts. Their solutions showed pH values in the range of 6.5-7.0 and, due to these optimal values, no or modest eye irritation effects were observed. Strong inhibitory properties were detected for many derivatives against hCA I, hCA II and bCA IV and, thanks to their strong tendency to concentrate in ocular fluids and tissues, almost 20 compounds showed effective IOP-lowering properties after topical administration.

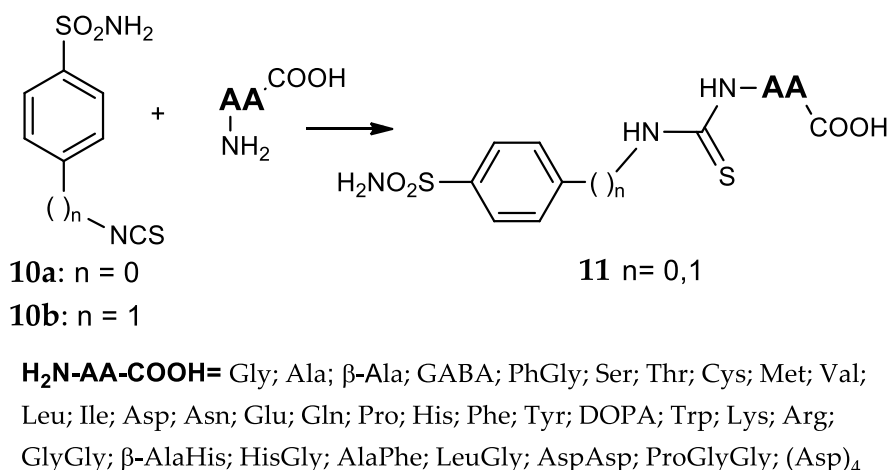
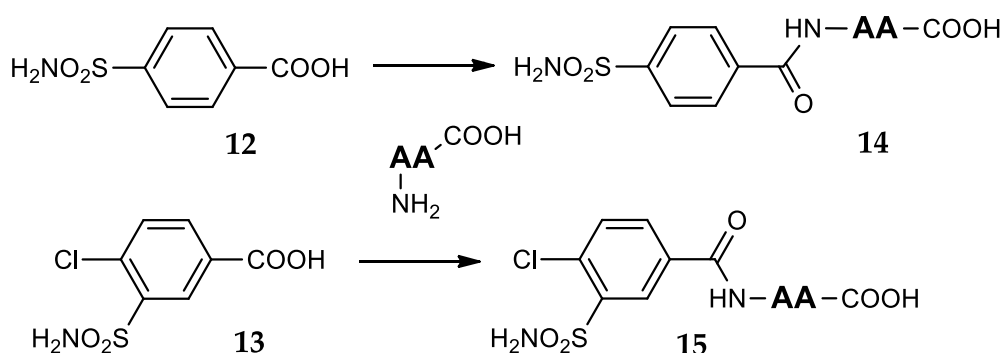


Figure 7.

Using 4-carboxybenzenesulfonamide **12** or 4-chloro-3-sulfamoylbenzoic acid **13** (figure 8) Mincione et al. [45] synthesized a new series of inhibitors, where different amino acids or dipeptides were directly linked to the sulfonamide through an amidic bond (general formula **14** and **15**, figure 8). As previously said for the thioureido derivatives **11**, many newly obtained compounds were able to inhibit three CA isozymes (hCA I, hCA II and bCA IV) showing  $K_i$  values in the nanomolar range. The corresponding carboxylate salts, with their good water-solubility, were investigated in IOP-lowering in-vivo experiments. The topical administration of these solutions, which possessed pH values in the neutrality range, showed very important and long lasting IOP-lowering effects in rabbits, stronger and longer-lasting on the glaucomatous animals than on the normotensive ones.



**H<sub>2</sub>N-AA-COOH**: Gly; β-Ala; GABA; Ala; Val; Leu; Ile; α-PhGly; Ser; β-PhSer; Thr; Cys; Met; Asp; Asn; Glu; Gln; His; Phe; Tyr; DOPA; GlyGly; β-AlaHis; HisGly; HisPhe; AlaPhe; LeuGly

Figure 8.

#### 4. The tail approach to Carbonic Anhydrases Inhibitors using Amino acids

As reported in section 2, CAIs fall into five categories, according to their mechanism of action [21]. Probably, the most important CAIs problem is the lack of selectivity for a specific isoform, due to the well conserved catalytic site among the isozymes. A possible method to gain selectivity is the so called "Tail Approach" [21,23,24,32]. This strategy is based on the structural modification of a portion on the inhibitor (**Tail** in figure 1A), usually in a position not fundamental for the Zn-chelating activity, that could potentially interact with the regions surrounding the active site, where the amino acids variability is higher. On these basis, the chemical differences in the amino acids side chains and the possibility to use also "non-α" amino acids make these molecules optimal candidates for the "tail" role (Figure 9). As is simple to imagine, there is an extremely high variety of possible synthetic

pathways that can be used to obtain different compounds by using the carboxy (as in 16) or the amino moiety (as in 17).

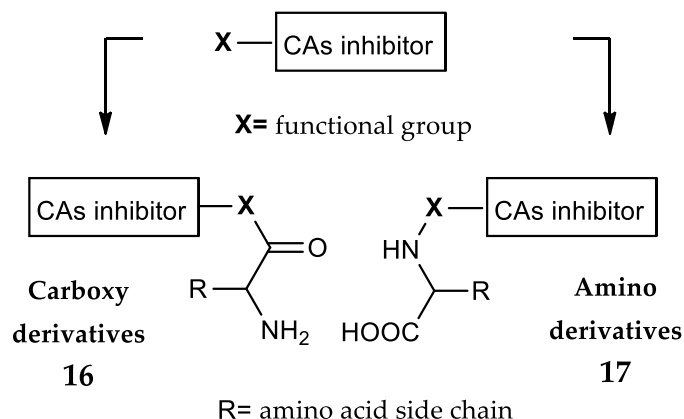


Figure 9.

An interesting work, focused on the investigation of enzyme-inhibitor interactions, was performed by the groups of Whitesides and Christianson [46–48]: in order to explore if secondary interactions away from the active site could influence inhibitory activity, they prepared three series of derivatives (general formula 18–20, figure 10).

At first these researchers synthesized a series of oligoglycine- and oligo(ethylene glycol)-linked benzenesulfonamides (18), whose dissociation constant on bCA II ( $K_d$ , measured by means of a competitive fluorescence-based assay) was found in the high micromolar range and independent of the polymer length [47]. Later, they condensed several tripeptides to 4-sulfamoylbenzoic acid, obtaining compounds with general formula 19, endowed with  $K_d$  values in the nanomolar range on hCA II [48]. The complex with 19a was analyzed by means of X-ray crystallography: the phenylglycine moiety was placed near Phe131 and Pro202, establishing hydrophobic interactions and validating the initial design.

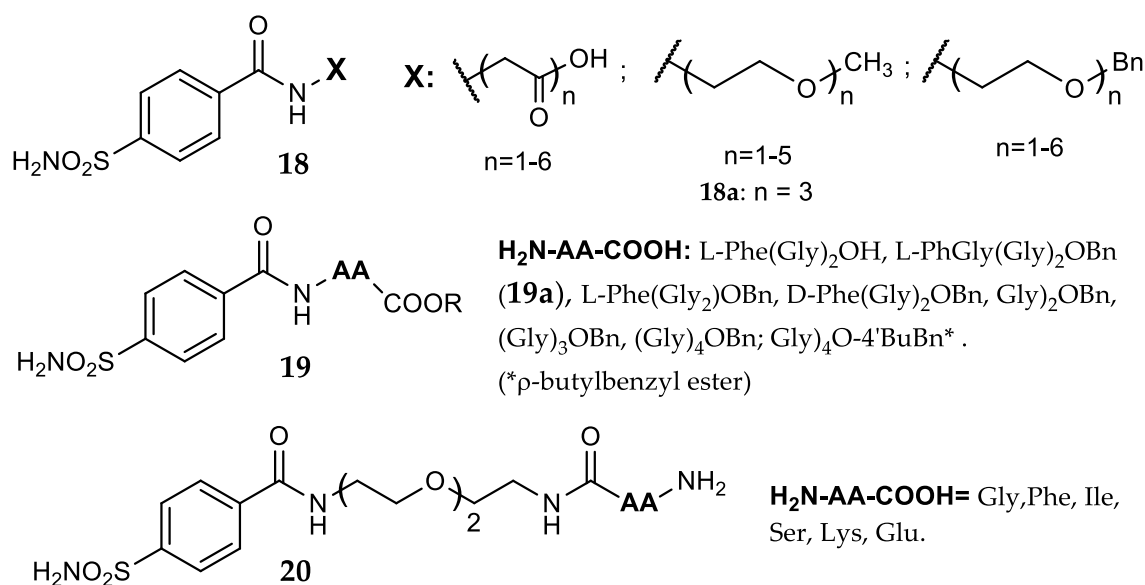
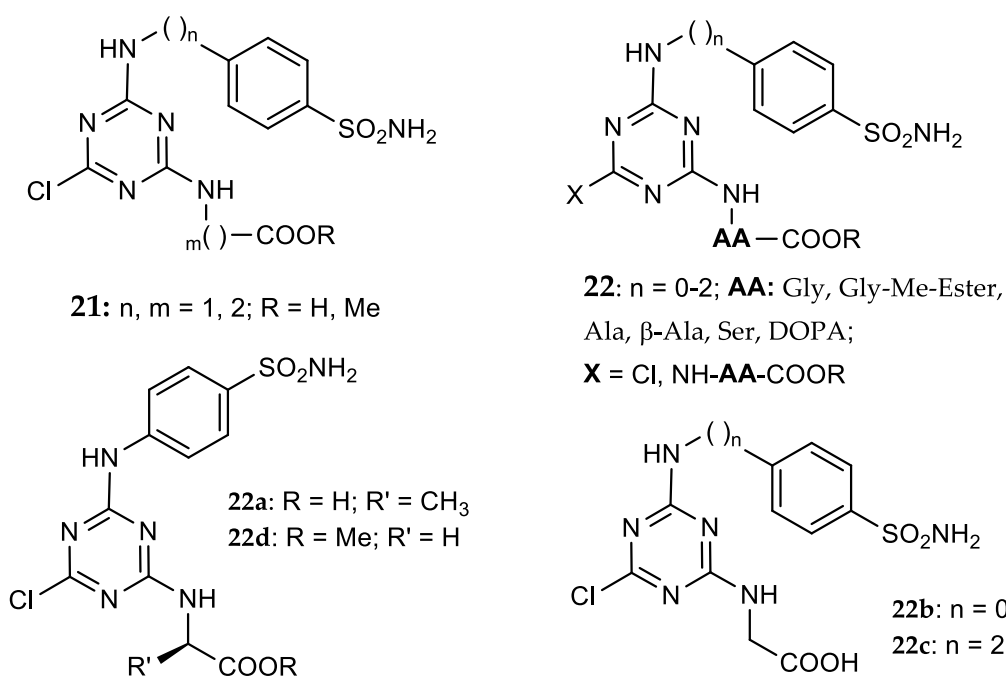


Figure 10.



The compound carrying three ethylene glycol moieties (**18a**) was therefore chosen by Borjack et al as primary scaffold for their final investigation [46]. Different amino acids were added to the terminal portion of the poly-ethylene glycol chain; compounds **20** were obtained, carrying a free amino group, and endowed with increased activity ( $K_d$  values from 2 to 10 times lower than the parent compound **18a**). This flexible and quite long linker was able to direct the amino acid toward enzyme surfaces far from the zinc ion, in a region where it can stabilize the binding through side interactions. Since in this series of compounds the best terminal pendants were lipophilic amino acids, these researchers suggested that the linker chain delivered the terminal group close to a hydrophobic region composed by Pro201, Pro202 and Leu198, even if the X-ray structures did not reveal ordered folds of the ethylene glycol linker.

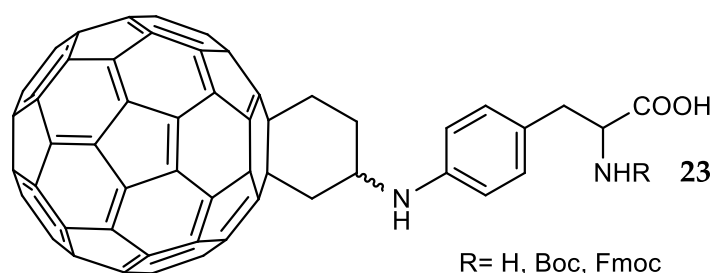
Garaj et al [49] used the tail approach on a series of novel sulfonamides incorporating the 1,3,5-triazine moiety, obtaining a small set of compounds with affinity from nanomolar to micromolar on hCA I, hCA II and hCA IX. In this series, glycine ( $m=1$ ) and  $\beta$ -alanine ( $m=2$ ) derivatives **21** (Figure 10) showed good potency on hCA IX, with  $K_i$  values 20-30 times lower than on hCA I and II, but independently of  $m$ . The library of these derivatives was later expanded by Carta et al. [50], who investigated, among several substituents, a wider number of amino acids in position 2 or 2,4 of the triazinyl ring (general formula **22**, figure 11). Many compounds showed nanomolar  $K_i$  values against the transmembrane tumor-associated CA IX and XII, in addition to CA XIV; the latter isoform is not associated with cancers but is widespread in many tissues such as the kidneys, liver and brain among others [3]. As for **21**, higher  $K_i$  values were found against the off-target isoforms hCA I and II. In some cases, some selectivity was obtained: as an example, compound **22a** showed a  $K_i$  0.96 nM on hCA IX, being on this isoform 6, 10, >550 and >1300 times more potent than on hCA XII, XIV, II and I, respectively. Two derivatives (**22b** and **22c**) displayed significant activity also on hCA VII. The binding of the analogue **22d** was analyzed by means of X-ray crystallography on hCA II: the Cl atom was engaged in contacts with the side-chain atoms of Ile91 and Gln92 while the triazinyl ring made  $\pi$ -stacking interaction with the phenyl ring of Phe131. The glycine moiety was directed toward the rim of the active site, apparently not engaged in positive interactions.



**Figure 11.**

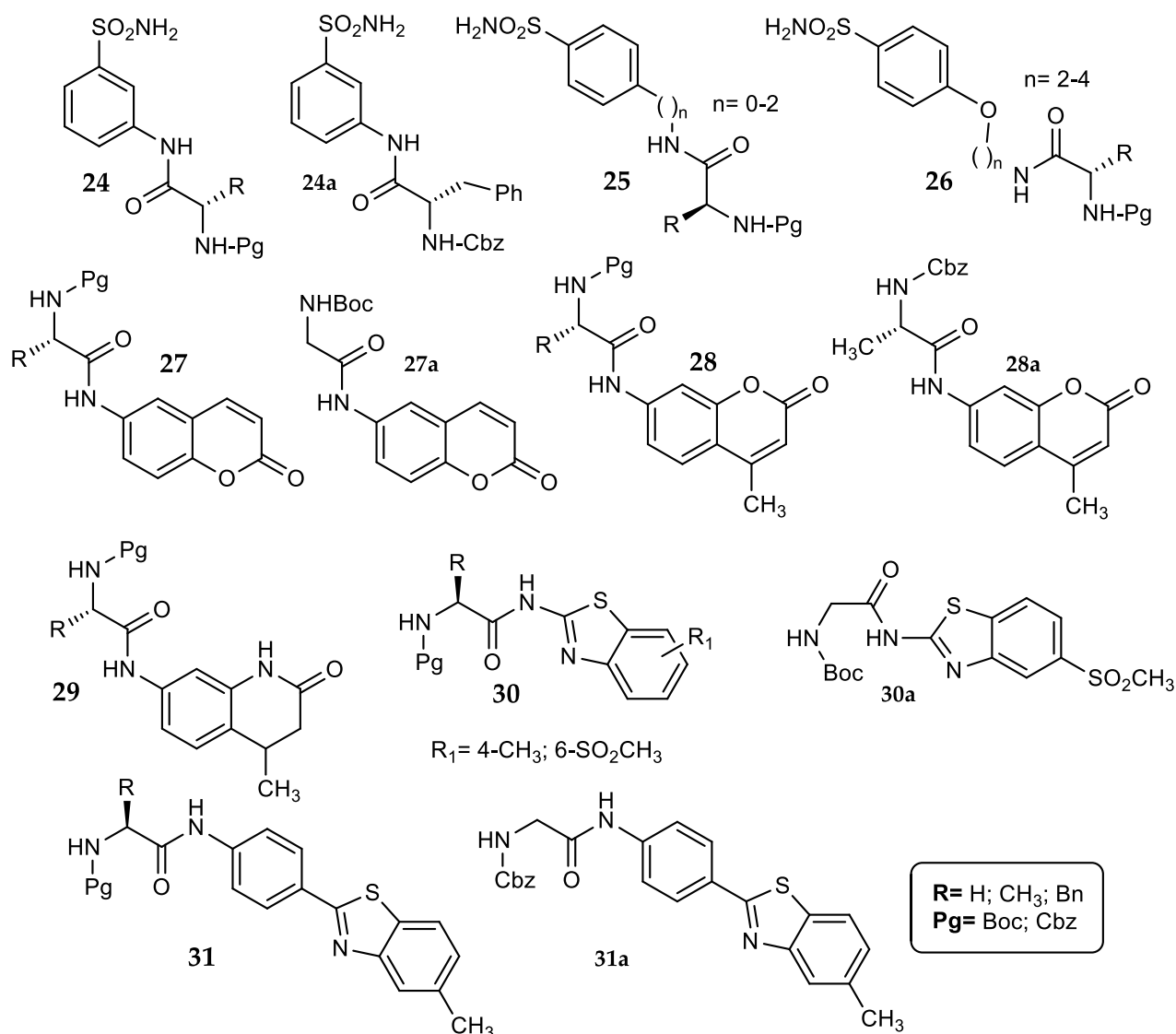
Amino acids were the ideal candidates for the side decoration of fullerene ( $C_{60}$ ), an atypical potential CA inhibitor [51]. Fullerene derivatives possess some interesting properties: as an example, Fullevir [52], the sodium salt of fullerene-polyhydro-polyaminocaproic acid, displays significant

antiviral, antibacterial and anticancer activity [53–56]. In addition, fullerene possesses a diameter of about 1 nm, a size similar to the width of the active site entrance of most CA isozymes [57]. In order to improve the very low water solubility and to insert groups that could provide selectivity by interacting with residues of the entrance of the catalytic site, some phenylalanine derivatives were investigated as fullerene pendants (general formula **23**, **figure 12**). Inhibitory activity was measured on a panel of hCA isozymes (I, II, III, IV, VA, VB, VI, VII, IX, XII, XIII, XIV, XV), finding  $K_i$  values in the micromolar range and no selectivity. Computational studies suggested that the inhibition mechanism is the occlusion of the active site through the fullerene cage; it is therefore possible that this bulky and extremely rigid scaffold prevents pendants from interacting with crucial amino acid residues, located in a deeper area of the enzyme. This hypothesis seems confirmed by the very low difference in activity measured for the synthesized fullerenes, including those not carrying amino acids pendants.



**Figure 12.**

A wide investigation of the “amino acid-tail approach” was performed by Küçükbay et al. They synthesised carboxy-derivatives linking a N-protected amino acid to three different types of CA inhibitors, obtaining amino-sulfonamides (**24–26**) [58], coumarins (**27, 28**), tetrahydroquinolinones **29** [59] and benzothiazoles **30, 31** [60] (**Figure 13**).

**Figure 13.**

Compounds **24-31** were tested against four CA isozymes (hCA I, II, IV and XII). The results were extremely different between the three series. Sulfonamides **24-26** were the most active and sensitive to structural variation, with  $K_i$  values mainly in the nanomolar range. By analysing the data, it is indeed possible to note appreciable differences in activity when the N-protected amino acid was varied, even if it is not simple to infer clear structure activity relationship. Few compounds were endowed with some selectivity toward CA XII: on this isoform, for instance, compound **24a** ( $K_i$  9.5 nM) is 18, 47 and 201 times more potent than on CA IV, II and I, respectively.

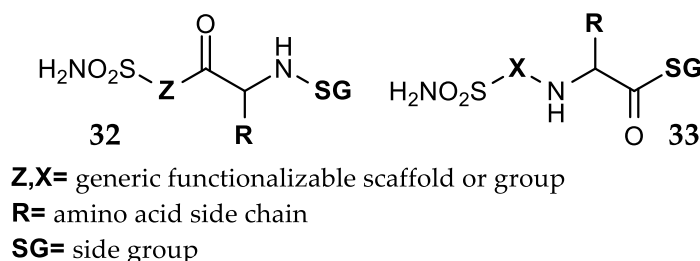
Dihydroquinolinones **29** were inactive on the four isoforms, while coumarins **27** and **28** showed micromolar  $K_i$  values only against hCA IV and XII [59]. As a matter of fact, this scaffold is already known as preferential inhibitor of the tumor-associated hCA IX and XII [23,61,62]; the addition of an aminoacid tail abolished activity against the off-target isozymes hCA I and II. Moreover, two derivatives (**27a** and **28a**, **figure 13**) were completely inactive also against hCA IV, thus showing selectivity for hCA XII.

Benzoathiazoles **30** and **31** showed micromolar  $K_i$  values almost exclusively on the ubiquitous isoform hCA I, with only three compounds being active also on hCA II. Surprisingly, one N-Boc-glycine derivative (**30a**, **figure 13**) showed activity also on hCA XII, with a  $K_i$  value nine time lower than that registered against hCA I. Another compound, **31a**, showed activity only on CA II, albeit with low potency ( $K_i$  10  $\mu$ M).

It is interesting to note that compounds **24-31** inhibit CA by different mechanisms, owing to the diverse scaffolds carrying the amino acid tail. The analysis of this research suggests that the “amino acid tail approach” could be interesting for the derivatization of different kind of CA inhibitors. The pretty simple and quick synthetic methods available to link an amino acid moiety to a generic group allow to obtain a large library of structurally-related compounds, where it is easy to study the effects of the substitution on activity. In addition, the hydrophilic nature of these molecules could be useful to enhance their water-solubility.

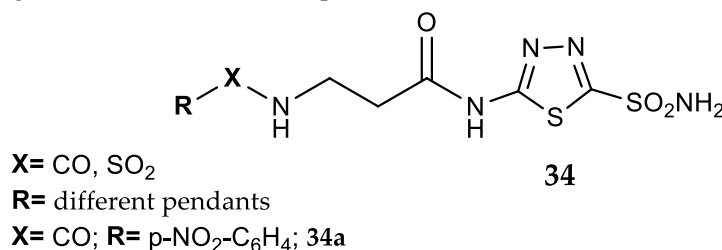
## 5. Amino acids as linkers

The bifunctional chemical nature of the amino acids can be used to link together two different substituents, generating molecules with new characteristics. A generic sulfonamide could be indeed connected to another chemical entity through these bivalent linkers (generic formula **32** and **33**, **figure 14**). The possibility to protect the amino or the carboxyl moieties, in addition to their different reactivity, allows the easily and selective functionalization of these chemical groups. This strategy could be considered an upgrade of the “amino acid tail approach”, on these compounds, beyond the amino acid chain, the side group (SG in **figure 14**) represents an additional point of variation.



**Figure 14.**

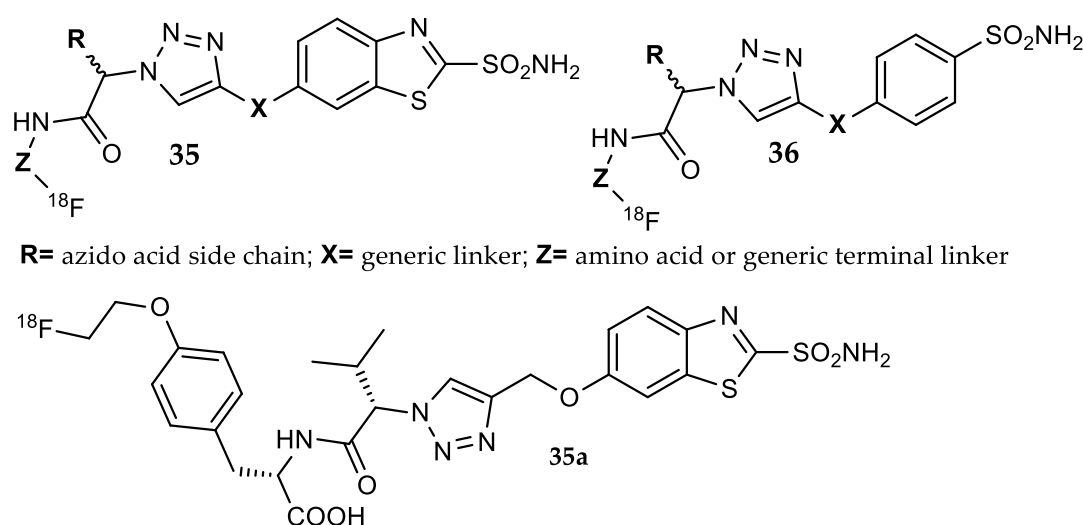
A first example is the previously cited work of Barboiu et al. [38], where compound **3a** (**figure 4**) was further functionalized on the free NH<sub>2</sub> with various pendants, affording a wider series of 5-substituted thiadiazole-2-sulfonamides (general formula **34**, **figure 15**). In order to obtain derivatives with different physico-chemical properties, compound **3a** (**Figure 4**) was reacted with 30 different electrophilic reagents (i.e. alkyl/aryl sulfonyl chlorides or fluorides, sulfonic acid cyclic anhydrides or acyl chlorides) and the corresponding products were then tested against isozymes hCA I, II and bCA IV. This series is characterized by a potent inhibitory activity on hCA II, most of the measured K<sub>i</sub> values being in the low nanomolar range on this isoform, but in the micromolar range against hCA I and bCA IV. We can cite derivative **34a** as example: the Barboiu group determined a K<sub>i</sub> of 0.75 nM on hCA II while the values measured against hCA I and bCA were respectively 125 and 100 nM. There is indeed a strong selectivity for the human isoform II confirming the evidence that this scaffold could be a promising candidate for the development of selective hCA II inhibitors.



**Figure 15.**

Kolb et al. patented a series of radioactively labelled sulfonamides [63], with high affinity for hCA IX, that could be potentially used as PET tracers for imaging the hCA IX expression on the surface of cancer cells. The group identified a benzothiazole or p-substituted phenyl ring as ideal

aromatic scaffolds to carry the sulfonamidic moiety and used click-chemistry to functionalize the inhibitor portion with different groups; a  $^{18}\text{F}$  atom was inserted on this tail as radionuclide. They designed four series of compounds and two of them (general formula **35** and **36**, **figure 16**) were prepared using an amino acid derivative (azido acid) as building block for the construction of the triazole central ring. In addition, the use of a second amino acid as radionuclide-carrying portion was investigated. With such a strategy, these researchers were able to obtain a large library of compounds, with different chemical characteristics and many possible points of structural variation. Furthermore, the use of L or D amino and azido acids allowed the stereochemical control of the side chain and, consequently, its orientation toward the regions of the enzyme. One of the most promising compound, VM-4037A (**35a**), was tested in tumor xenograft models expressing hCA-IX [64]. In addition, its biodistribution and radiation dose were determined in healthy human volunteers [65]; the high uptake of **35a** in liver and kidney indicated that the compound was not suitable for the detection of hCA IX in these tissues.



**R**= azido acid side chain; **X**= generic linker; **Z**= amino acid or generic terminal linker

**Figure 16.**

A quite different approach was used by the Matulis group [66,67] for the derivatization of *p*-amino benzenesulfonamide. By reacting the *p*- amino group with acrylic acid, Rutkauskas et al obtained a series of compounds where a  $\beta$ -alanyl moiety is directly linked to the aromatic scaffold (general formula **37** and **38**, **figure 17**). In this instance, the amino acid linker was generated in situ and then functionalized on the free  $\beta$ -carboxy moiety. In addition, the versatile nature of this group allowed its transformation into heterocycles, such as triazoles, oxadiazoles or thiadiazoles [66]. Only in few instances the activity of this first series of compounds ( $K_d$  values determined by a fluorescent thermal shift assay on CA I, II, VI, VII, XII and XIII) was below  $1\ \mu\text{M}$ . The functionalization of the alanyl nitrogen and substitution on the benzenesulfonamide ring were also investigated, adding other possible points of variation [67]. As one can imagine, this large number of different products showed a wide range of activities, with  $K_d$  values on CA I, II, XII and XIII from nanomolar to micromolar. The selectivity of some compounds was further tested on a larger panel of hCAs, with some interesting findings. For instance, **39** (**figure 17**) showed good potency on CA VB and CA IX ( $K_d = 5$  and  $43\ \text{nM}$ , respectively) while being almost inactive on the other isoforms. The crystallographic studies performed on **37a** and hCA II allowed to understand how the flexible  $\beta$ -alanyl linker was folded in the enzyme cavity.

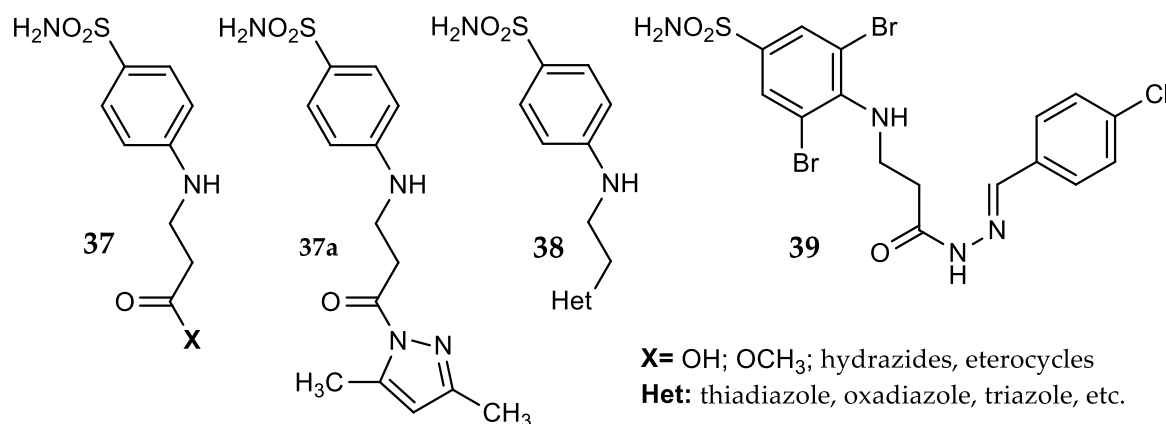


Figure 17.

Ceruso et al. [68] extended the chain using a  $\gamma$ -aminobutyric acid moiety as linker between two different benzenesulfonamides and various aromatic pendants. Compared to **37**, the amino acid orientation was inverted, leaving a free amino group that was reacted with aryl isocyanates. The obtained molecules (general formula **40**, figure 18) were tested against 13 hCA isoforms, showing good inhibition potency against hCA I, II, VII and XII. This series lack isoform selectivity, but a deeper investigation on the effects of the chain length and of the chemical nature of the terminal aromatic pendants could potentially lead to more interesting derivatives.

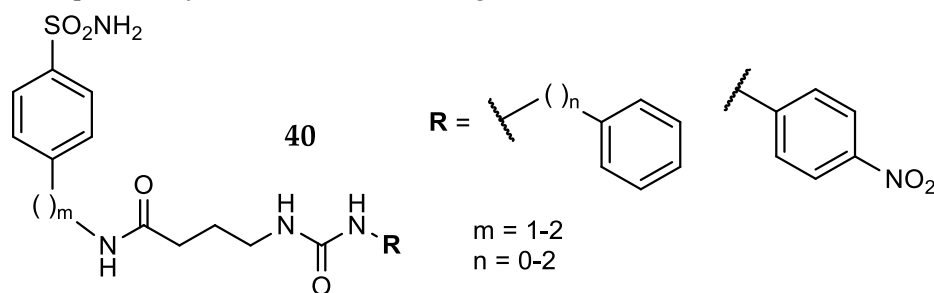


Figure 18.

Moeker et al. [69] studied an innovative approach using a sulfamate as zinc binding group (ZBG). These researchers designed a series of glycosylated compounds with poor membrane permeability that could potentially lead to hCA IX selective inhibitors, owing the extracellular localization of this enzyme. The general structure of these derivatives (**41**, figure 19) is characterized by a free or acetylated glucose moiety, linked to a panel of primary and secondary amino alcohols through a sulfonamide bridge. Two hydroxylated amino acid (L-Serine methyl ester and L-4-(R)-hydroxyproline methyl ester) were investigated as hydroxylated linkers and transformed into the corresponding sulfamate (**41a** and **41b**). In such a way, the ZBG is directly placed on the chiral amino acid side chain. All the synthesized molecules showed nanomolar  $K_i$  values against the tumor-associated isoforms hCA IX and XII [5,13], and lower potency against the off-target isozymes hCA I and II. Particularly interesting is the proline-derivative **41a**, with  $K_i$  values of 2 and 1 nanomolar respectively on hCA IX and XII and more than 350 times lower potency on hCA I and II. This innovative method is interesting not only from a synthetic point of view, but also because it allows to keep fixed the ZBG and functionalize both the amino and carboxy portions of the amino acid with different residues.

These examples demonstrate that a chiral amino acid can serve either as building block either as linker between side pendants.



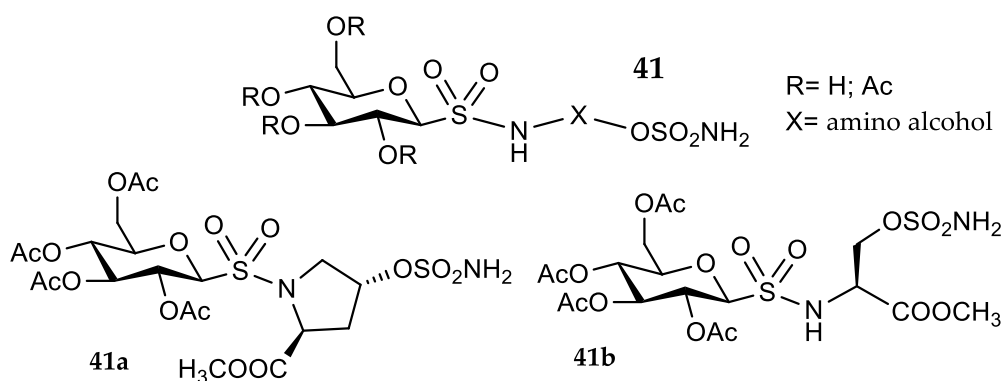


Figure 19.

## 6. Dual CAs and MMPs inhibitors

Matrix metalloproteinases (MMPs) are zinc-containing endopeptidases, capable of degrading the extracellular matrix proteins; up-regulation of specific isoforms is associated to various pathologies, including some metastatic cancers [70–72]. These enzymes are potently inhibited by hydroxamates, but also other chemical groups are known to be effective Zn-chelating agents [73]. Since hCA IX and XII are overexpressed in tumors [5,13], the simultaneous inhibition of both enzymes should give a synergic effect.

On these basis, dual inhibitors were designed [74,75], i.e. compounds able to interact with the binding regions of both metalloenzymes. Some amino acids were investigated as structural backbone of the new dual inhibitors; the derivatization of the carboxy and the amino moieties gave sulfonylated amino acid hydroxamates and N-alkyloxy amino acid hydroxamates (general formulas **42–44**, figure 20). Compounds contemporary carrying a hydroxamic and a protic-sulfonyl groups are therefore provided with two different chelating moieties that could potentially interact with both metalloenzymes. Compounds **42** were strong dual inhibitors with  $K_i$  values in the low-medium nanomolar range, their inhibitory properties against the two enzymes depending on some structural features [74]. For instance, compounds **42** carrying a NH moiety ( $X = H$ ) were more active on CA (isoform I, II and IV) than on MMPs (isoforms 1, 2, 8 and 9). When X was a benzyl group, a potent activity was found on both enzymes, while a substituent on the benzyl moiety almost abolished activity on CAs. Unfortunately, these substances were not tested on the tumor-associated CA isoforms (IX and XII).

Compounds **43** and **44** were in general less potent on both hCAs and MMPs, probably due to the completely absence of protic sulfonyl moiety that could behave as ZBG [75].

These studies confirm that amino acids, by showing good synthetic properties, are interesting synthones also for the design of hybrid inhibitors, where an important need is the possibility to easily modify many positions with different kind of substituents.

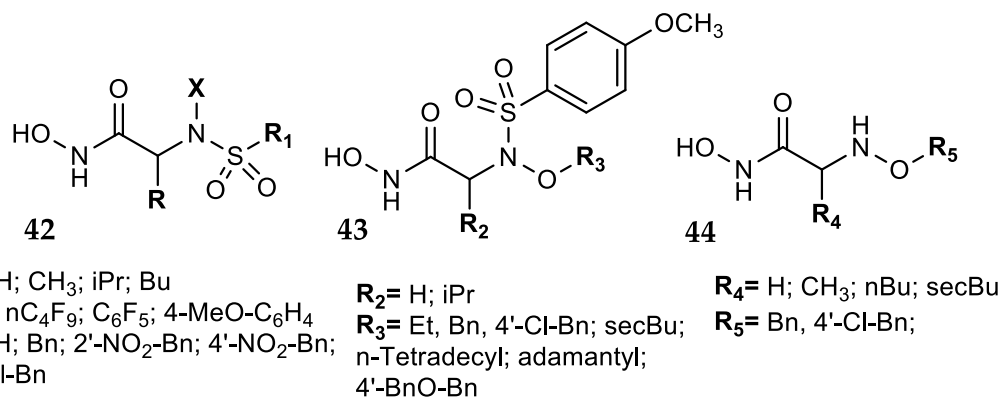
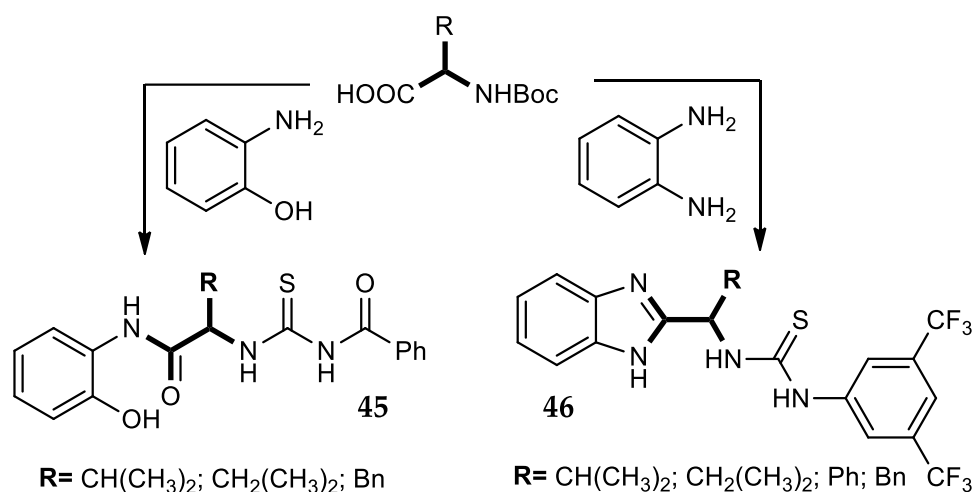


Figure 20.

## 7-Other approaches

Especially from a synthetic point of view, another interesting approach to the design of novel CAIs is the one reported in a paper by Korkmaz N. et al [76]. These researchers, aiming to investigate innovative inhibitors, synthesized a series of thiourea derivatives lacking a classic ZBG. As we can notice from general structures **45** and **46** in **figure 21**, there are not pharmacophoric moieties that could be immediately associated with an inhibitory activity on the enzyme. Nevertheless, micromolar  $K_i$  values were measured for these series of compounds against hCA I and II. By analysing the structures, we can observe that lipophilic amino acids were used as linker between two aromatic portions. For **46**, the amino acid carboxy group reacted with o-phenyldiamine giving a benzimidazole ring. Compounds **45** and **46** products were not extremely potent, but these synthetic pathways could be interesting for the side derivatization of sulfonamidic molecules.



**Figure 21.**

Another scaffold lacking a classic chelating moiety is Probenecid, a drug used treat gout and hyperuricemia. Probenecid, probably thanks to the free carboxyl that coordinates the zinc ion, possesses a weak inhibitory activity against CAs. Mollica et al. [77] investigated the effects of adding different L-amino acid, obtaining probenecid-based amide derivatives (general formula **47**, **figure 22**). This modification shifted the carboxy group away from the aromatic ring and introduced a chiral linker. With the aim to study the impact of the chemical modification of this terminal COOH, these researchers synthesized also the corresponding primary amides. All the compounds showed an interesting profile, their inhibitory activity against hCA IX and XII being higher than against the off-target isozymes hCA I and II. The transformation of the COOH group into CONH<sub>2</sub> was also interesting, resulting in a remarkable lower affinity against hCA II. As example, both the methionine derivatives **47a** and **47b** were very active against the two tumor-associated isoforms, but while the carboxy derivatives possesses a low residual activity against hCA I and II, the corresponding amide is completely inactive toward these isoforms. This paper provides a useful overview of the application of the “amino acid tail approach” to an atypical zinc-chelating molecule, confirming the synthetic and pharmacological benefits that could be obtained from this method.

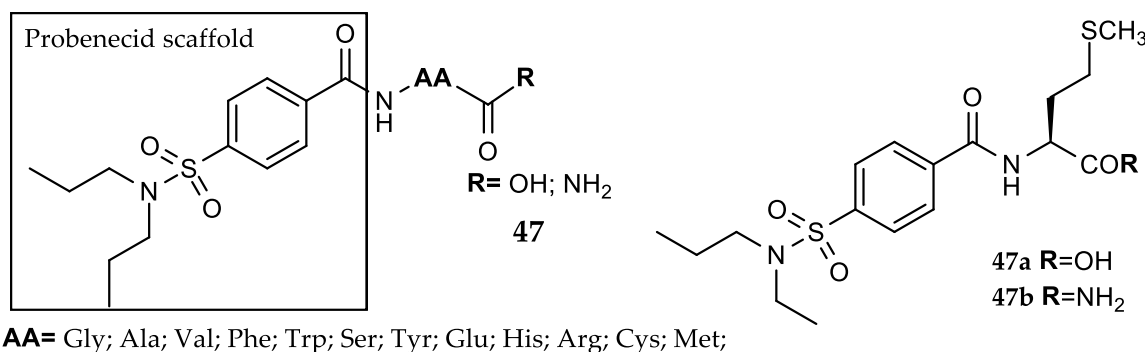


Figure 22.

In this section we report also the paper published by Fidan I. et al [78], in which four aromatic sulfonamides were directly functionalized on the amidic NH<sub>2</sub> group with glycine and phenylalanine (general formula 48, figure 23). The derivatization of the zinc binding group with substituents able to interact with the amino acids of the catalytic site was aimed at obtaining an enhancement in the activity of these secondary sulfonamides. However, it must be reminded that the architecture of the active site is very well conserved among the different CA isoforms, so this modification may not confer selectivity to such derivatives. In general, the new compounds did not show a better inhibition profile against hCA I and II as compared to their primary sulfonamide precursors. A substitution directly on the ZBG, probably due to steric hindrance, seems not to be a convenient way to enhance the potency of this kind of sulfonamides. Despite these negative results, this approach should be further investigated, maybe with longer linkers between the sulfonamidic nitrogen and the pendants.

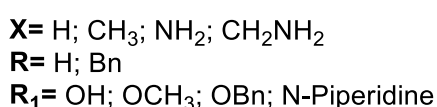
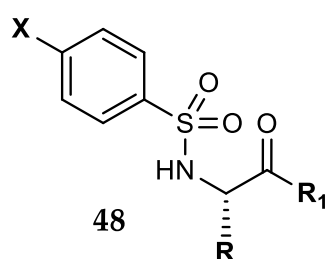


Figure 23.

Chiral pool synthesis was used by Chiaramonte et al in order to prepare enantiopure CA inhibitors [79]. Starting from L or D-phenylalanine, both enantiomers of two series of 2-benzylpiperazines (49 and 50, figure 24), carrying a sulfamoyl-benzoic moiety as ZBG, were obtained. The amino acid was directly used to build the piperazine scaffold, allowing the stereochemical control of the side benzyl group. With this method, it was possible to take advantage of the versatile synthetic properties of the synthon: the reduction of the intermediate diketopiperazine 51 converted the amino acid into an alkyl derivative without affecting the absolute configuration. The piperazines were further decorated on one nitrogen atom with the ZBG and with different alkyl/acetyl/sulfonyl groups on the other one, with the aim to look for selectivity toward the CA isoforms. All the synthesized products were able to inhibit four pharmacological-relevant CA isozymes (hCA I, II, IV and IX), showing K<sub>i</sub> values ranging from low nanomolar to micromolar; hCA IX was the least sensitive isoform. The hydrochloric salts of two basic piperazines (49a and 50a, figure 24), showing very good water solubility, were then selected for in-vivo tests in a rabbit model of glaucoma. Both derivatives were able to reduce IOP with a potency and efficacy similar to the reference drug

Dorzolamide, confirming that this series is promising for the development of new anti-glaucoma agents. The binding mode was further investigated with X-ray analysis and the structure of the complex of compound **49b**, bounded in the hCA I active site, was solved. In the image, reported in **figure 1B** (section 1), is possible to appreciate the interaction between the sulfonamidic group and the  $\text{Zn}^{2+}$ , while the benzyl group is deeply inserted in the lipophilic cavity of the enzyme, where it establishes Van der Waals interactions with some hydrophobic amino acid residues. This synthetic method allowed building a chiral piperazine scaffold, incorporating the amino acid-skeleton. Starting from different amino acids, it should be possible to modify the nature of this substituent and at the same time to control its stereochemistry.

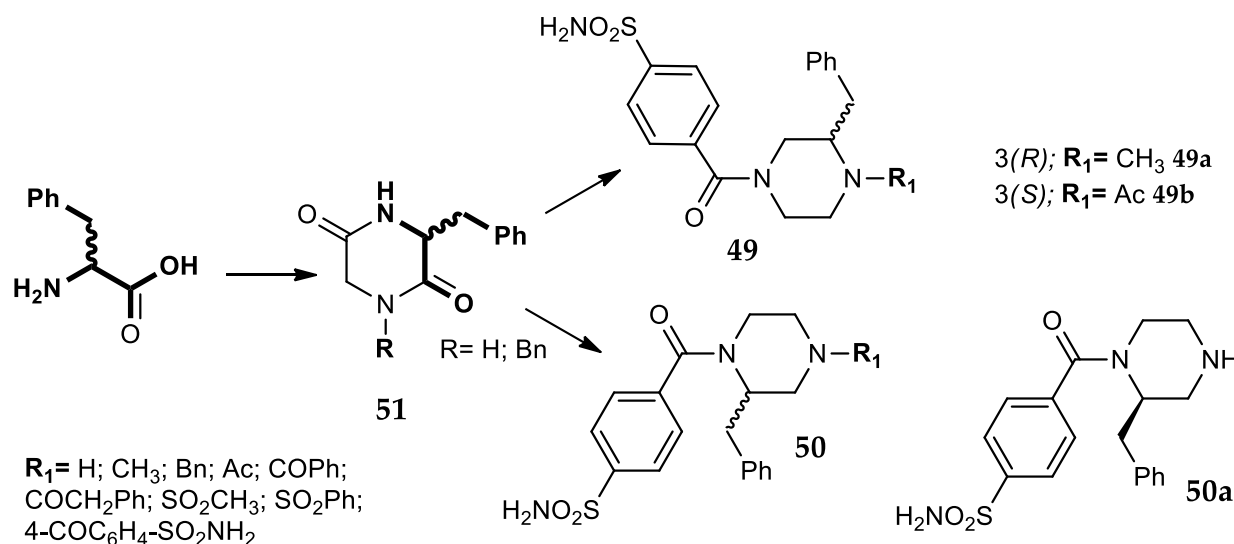


Figure 24.

## 8. Conclusions

Amino acids are useful synthons. Thanks to their characteristics, there are several potential applications in chemical synthesis. The possibility to use both proteinogenic and non-proteinogenic amino acids allow to prepare many related derivatives, where the different side chains modulate the properties. The research of new CAIs or the structural modification of existing inhibitors is a dynamic field with many perspectives but also a lot of still unsolved challenges, the most important being the lack of selectivity of sulfonamides. The use of amino acids or their derivatives, could sometimes provide a smart solution both to synthetic or to pharmacological problems.

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**Conflicts of interests:** The authors declare no conflict of interest, except C.T. Supuran who declares conflict of interest, being co-author of many patents describing CAIs.

**Abbreviation list:** CA: carbonic anhydrase; hCA: human carbonic anhydrase; bCA: bovine carbonic anhydrase; CAIs: carbonic anhydrases inhibitors; MMPs: matrix metalloproteinases; IOP: intraocular pressure; Ala: alanine; Asn: asparagine; Asp: aspartic acid; Arg: arginine;  $\beta$ -Ala: beta-alanine;  $\beta$ -PhSer: beta-phenylserine; Boc: tert-butyloxycarbonyl; Cys: cysteine; DOPA: (3',4'-dihydroxy)phenylalanine; Fmoc: fluorenylmethyloxycarbonyl; GABA:  $\gamma$ -aminobutyric acid; Gly: glycine; Gln: glutamine; Glu: glutamic acid; Ile: isoleucine;  $K_{\text{cat}}$ : turnover number;  $K_a$ : dissociation constant;  $K_i$ : inhibition constant; Leu: leucine; Lys: lysine; Met: methionine; PET: positron emission tomography; Phe: phenylalanine; PhGly: phenylglycine; Pro: proline; Ser: serine; Thr: threonine; Trp: tryptophan; Tyr: tyrosine; Val: valine; ZBG: zinc binding group

## References.

1. Kupriyanova, E.; Pronina, N.; Los, D. Carbonic anhydrase — a universal enzyme of the carbon-based life. *Photosynthetica* **2017**, *55*, 3–19, doi:10.1007/s11099-017-0685-4.
2. Supuran, C. T. Structure and function of carbonic anhydrases. *Biochem. J.* **2016**, *473*, 2023–2032, doi:10.1042/BCJ20160115.
3. Supuran, C. T.; De Simone, G. Carbonic Anhydrases: An Overview. *Carbon. Anhydrases as Biocatal. From Theory to Med. Ind. Appl.* **2015**, 3–13, doi:10.1016/B978-0-444-63258-6.00001-9.
4. D'Ambrosio, K.; De Simone, G.; Supuran, C. T. Human Carbonic Anhydrases: Catalytic Properties, Structural Features, and Tissue Distribution. *Carbon. Anhydrases as Biocatal. From Theory to Med. Ind. Appl.* **2015**, 17–30, doi:10.1016/B978-0-444-63258-6.00002-0.
5. Supuran, C. T. Carbonic anhydrases: Novel therapeutic applications for inhibitors and activators. *Nat. Rev. Drug Discov.* **2008**, *7*, 168–181, doi:10.1038/nrd2467.
6. Supuran, C. Carbonic Anhydrases and Metabolism. *Metabolites* **2018**, *8*, 25, doi:10.3390/metabo8020025.
7. Smith, K. S.; Jakubzick, C.; Whittam, T. S.; Ferry, J. G. Carbonic anhydrase is an ancient enzyme widespread in prokaryotes. *Proc. Natl. Acad. Sci.* **1999**, *96*, 15184–15189, doi:10.1073/pnas.96.26.15184.
8. Khalifah, R. G. The Carbon Dioxide Hydration Activity of Carbonic Anhydrase. *J. Biol. Chem.* **1971**, *246*, 2561–2573.
9. Lindskog, S. Structure and mechanism of carbonic anhydrase. *Pharmacol. Ther.* **1997**, *74*, 1–20, doi:10.1016/S0163-7258(96)00198-2.
10. Silverman, D. N.; Lindskog, S. The Catalytic Mechanism of Carbonic Anhydrase: Implications of a Rate-Limiting Protolysis of Water. *Acc. Chem. Res.* **1988**, *21*, 30–36, doi:10.1021/ar00145a005.
11. Alterio, V.; Di Fiore, A.; D'Ambrosio, K.; Supuran, C. T.; De Simone, G. Multiple binding modes of inhibitors to carbonic anhydrases: How to design specific drugs targeting 15 different isoforms? *Chem. Rev.* **2012**, *112*, 4421–4468.
12. Masini, E.; Carta, F.; Scozzafava, A.; Supuran, C. T. Antiglaucoma carbonic anhydrase inhibitors: a patent review. *Expert Opin. Ther. Pat.* **2013**, *23*, 705–716, doi:10.1517/13543776.2013.794788.
13. Neri, D.; Supuran, C. T. Interfering with pH regulation in tumours as a therapeutic strategy. *Nat. Rev. Drug Discov.* **2011**, *10*, 767–777, doi:10.1038/nrd3554.
14. Supuran, C. T. Carbonic anhydrase inhibition and the management of hypoxic tumors. *Metabolites* **2017**, *7*, doi:10.3390/metabo7030048.
15. Singh, S.; Lomelino, C. L.; Mboge, M. Y.; Frost, S. C. Cancer Drug Development of Carbonic Anhydrase Inhibitors beyond the Active Site. **2018**, doi:10.3390/molecules23051045.
16. Mboge, M.; Mahon, B.; McKenna, R.; Frost, S. Carbonic Anhydrases: Role in pH Control and Cancer. *Metabolites* **2018**, *8*, 19, doi:10.3390/metabo8010019.
17. Supuran, C. T. Carbonic anhydrase activators. *Future Med. Chem.* **2018**, *10*, 561–573, doi:10.4155/fmc-2017-0223.
18. Carta, F.; Supuran, C. T. Diuretics with carbonic anhydrase inhibitory action: a patent and literature review (2005 – 2013). *Expert Opin. Ther. Pat.* **2013**, *23*, 681–691, doi:10.1517/13543776.2013.780598.
19. Monti, S. M.; Supuran, C. T.; De Simone, G. Anticancer carbonic anhydrase inhibitors: a patent review (2008 – 2013). *Expert Opin. Ther. Pat.* **2013**, *23*, 737–749, doi:10.1517/13543776.2013.798648.
20. Scozzafava, A.; Supuran, C. T.; Carta, F. Antiobesity carbonic anhydrase inhibitors: a literature and patent review. *Expert Opin. Ther. Pat.* **2013**, *23*, 725–35, doi:10.1517/13543776.2013.790957.
21. Supuran, C. T. Advances in structure-based drug discovery of carbonic anhydrase inhibitors. *Expert Opin. Drug Discov.* **2017**, *12*, 61–88, doi:10.1080/17460441.2017.1253677.
22. De Simone, G.; Alterio, V.; Supuran, C. T. Exploiting the hydrophobic and hydrophilic binding sites for designing carbonic anhydrase inhibitors. *Expert Opin. Drug Discov.* **2013**, *8*, 793–810, doi:10.1517/17460441.2013.795145.
23. Supuran, C. T. How many carbonic anhydrase inhibition mechanisms exist? *J. Enzyme Inhib. Med. Chem.* **2016**, *31*, 345–360, doi:10.3109/14756366.2015.1122001.
24. Supuran, C. T. Structure-based drug discovery of carbonic anhydrase inhibitors. *J. Enzyme Inhib. Med. Chem.* **2012**, *27*, 759–772.
25. Karioti, A.; Carta, F.; Supuran, C. T. Phenols and polyphenols as Carbonic anhydrase inhibitors. *Molecules* **2016**, *21*, 12–17, doi:10.3390/molecules21121649.



26. Maresca, A.; Temperini, C.; Pochet, L.; Masereel, B.; Scozzafava, A.; Supuran, C. T. Deciphering the mechanism of carbonic anhydrase inhibition with coumarins and thiocoumarins. *J. Med. Chem.* **2010**, *53*, 335–344, doi:10.1021/jm901287j.
27. D'Ambrosio, K.; Carradori, S.; Monti, S. M.; Buonanno, M.; Secci, D.; Vullo, D.; Supuran, C. T.; De Simone, G. Out of the active site binding pocket for carbonic anhydrase inhibitors. *Chem. Commun.* **2015**, *51*, doi:10.1039/c4cc07320g.
28. Bischoff, R.; Schlüter, H. Amino acids: Chemistry, functionality and selected non-enzymatic post-translational modifications. *J. Proteomics* **2012**, *75*, 2275–2296.
29. Jaradat, D. M. M. Thirteen decades of peptide synthesis: key developments in solid phase peptide synthesis and amide bond formation utilized in peptide ligation. *Amino Acids* **2018**, *50*, 39–68.
30. Schmidl, D.; Schmetterer, L.; Garhöfer, G.; Popa-Cherecheanu, A. Pharmacotherapy of Glaucoma. *J. Ocul. Pharmacol. Ther.* **2015**, *31*, 63–77, doi:10.1089/jop.2014.0067.
31. Sugrue, M. F. Pharmacological and ocular hypotensive properties of topical carbonic anhydrase inhibitors. *Prog. Retin. Eye Res.* **2000**, *19*, 87–112.
32. Supuran, C. T.; Casini, A.; Scozzafava, A. Development of sulfonamide carbonic anhydrase inhibitors. In *Carbonic Anhydrase*; 2004; pp. 67–147.
33. Maren, T. H. Carbonic anhydrase: chemistry, physiology, and inhibition. *Physiol. Rev.* **1967**, *47*, 595–781, doi:10.1152/physrev.1967.47.4.595.
34. Casini, A.; Scozzafava, A.; Mincione, F.; Menabuoni, L.; Ilies, M. A.; Supuran, C. T. Carbonic anhydrase inhibitors: Water-soluble 4-sulfamoylphenylthioureas as topical intraocular pressure-lowering agents with long-lasting effects. *J. Med. Chem.* **2000**, *43*, 4884–4892, doi:10.1021/jm001051+.
35. Scozzafava, A.; Briganti, F.; Mincione, G.; Menabuoni, L.; Mincione, F.; Supuran, C. T. Carbonic anhydrase inhibitors: Synthesis of water-soluble, aminoacyl/dipeptidyl sulfonamides possessing long-lasting intraocular pressure-lowering properties via the topical route. *J. Med. Chem.* **1999**, *42*, 3690–3700, doi:10.1021/jm9901879.
36. Antonaroli, S.; Bianco, A.; Brufani, M.; Cellai, L.; Baido, G. Lo; Potier, E.; Bonomi, L.; Perfetti, S.; Fiaschi, A. I.; Segre, G. Acetazolamide-like Carbonic Anhydrase Inhibitors with Topical Ocular Hypotensive Activity. *J. Med. Chem.* **1992**, *35*, 2697–2703, doi:10.1021/jm00092a021.
37. Jayaweera, G. D. S. A.; MacNeil, S. A.; Trager, S. F.; Blackburn, G. M. Synthesis of 2-substituted-1,3,4-thiadiazole-5-sulphonamides as novel water-soluble inhibitors of carbonic anhydrase. *Bioorganic Med. Chem. Lett.* **1991**, *1*, 407–410, doi:10.1016/S0960-894X(00)80266-8.
38. Barboiu, M.; Supuran, C. T.; Menabuoni, L.; Scozzafava, A.; Mincione, F.; Briganti, F.; Mincione, G. Carbonic anhydrase inhibitors. Synthesis of topically effective intraocular pressure lowering agents derived from 5-(omega-aminoalkylcarboxamido)-1,3,4-thiadiazole-2-sulfonamide. *J. Enzyme Inhib.* **1999**, *15*, 23–46.
39. Supuran, C. T.; Briganti, F.; Menabuoni, L.; Mincione, G.; Mincione, F.; Scozzafava, A. Carbonic anhydrase inhibitors - Part 78. Synthesis of water-soluble sulfonamides incorporating  $\beta$ -alanyl moieties, possessing long lasting- intraocular pressure lowering properties via the topical route. *Eur. J. Med. Chem.* **2000**, *35*, 309–321, doi:10.1016/S0223-5234(00)00130-6.
40. Maren, T. H.; Brechue, W. F.; Bar-Ilan, A. Relations among IOP reduction, ocular disposition and pharmacology of the carbonic anhydrase inhibitor ethoxzolamide. *Exp. Eye Res.* **1992**, *55*, 73–79, doi:10.1016/0014-4835(92)90094-9.
41. Brechue, W. F.; Maren, T. H. pH and drug ionization affects ocular pressure lowering of topical carbonic anhydrase inhibitors. *Investig. Ophthalmol. Vis. Sci.* **1993**, *34*, 2581–2587.
42. Ceruso, M.; Del Prete, S.; Alothman, Z.; Osman, S. M.; Scozzafava, A.; Capasso, C.; Supuran, C. T. Synthesis of sulfonamides with effective inhibitory action against *Porphyromonas gingivalis*  $\gamma$ -carbonic anhydrase. *Bioorganic Med. Chem. Lett.* **2014**, *24*, 4006–4010, doi:10.1016/j.bmcl.2014.06.024.
43. Ceruso, M.; Bragagni, M.; Alothman, Z.; Osman, S. M.; Supuran, C. T. New series of sulfonamides containing amino acid moiety act as effective and selective inhibitors of tumor-associated carbonic anhydrase XII. *J. Enzyme Inhib. Med. Chem.* **2015**, *30*, 430–434, doi:10.3109/14756366.2014.942659.
44. Casini, A.; Scozzafava, A.; Mincione, F.; Menabuoni, L.; Supuran, C. T. Carbonic anhydrase inhibitors: Synthesis of water soluble sulfonamides incorporating a 4-sulfamoylphenylmethylthiourea scaffold, with potent intraocular pressure lowering properties. *J. Enzyme Inhib. Med. Chem.* **2002**, *17*, 333–343, doi:10.1080/1475636021000010950.
45. Mincione, F.; Starnotti, M.; Menabuoni, L.; Scozzafava, A.; Casini, A.; Supuran, C. T. Carbonic anhydrase inhibitors: 4-Sulfamoyl-benzenecarboxamides and 4-chloro-3-sulfamoyl-benzenecarboxamides with strong



- topical antiglaucoma properties. *Bioorganic Med. Chem. Lett.* **2001**, *11*, 1787–1791, doi:10.1016/S0960-894X(01)00303-1.
46. Boriack, P. A.; Christianson, D. W.; Kingery-Wood, J.; Whitesides, G. M. Secondary Interactions Significantly Removed from the Sulfonamide Binding Pocket of Carbonic Anhydrase II Influence Inhibitor Binding Constants. *J. Med. Chem.* **1995**, *38*, 2286–2291, doi:10.1021/jm00013a004.
47. Jain, A.; Huang, S. G.; Whitesides, G. M. Lack of Effect of the Length of Oligoglycine- and Oligo(ethylene glycol)-Derived para-Substituents on the Affinity of Benzenesulfonamides for Carbonic Anhydrase II in Solution. *J. Am. Chem. Soc.* **1994**, *116*, 5057–5062, doi:10.1021/ja00091a005.
48. Jain, A.; Whitesides, G. M.; Alexander, R. S.; Christianson, D. W. Identification of Two Hydrophobic Patches in the Active-Site Cavity of Human Carbonic Anhydrase II by Solution-Phase and Solid-State Studies and Their Use in the Development of Tight-Binding Inhibitors. *J. Med. Chem.* **1994**, *37*, 2100–2105, doi:10.1021/jm00039a023.
49. Garaj, V.; Puccetti, L.; Fasolis, G.; Winum, J. Y.; Montero, J. L.; Scozzafava, A.; Vullo, D.; Innocenti, A.; Supuran, C. T. Carbonic anhydrase inhibitors: Novel sulfonamides incorporating 1,3,5-triazine moieties as inhibitors of the cytosolic and tumour-associated carbonic anhydrase isozymes I, II and IX. *Bioorganic Med. Chem. Lett.* **2005**, *15*, 3102–3108, doi:10.1016/j.bmcl.2005.04.056.
50. Carta, F.; Garaj, V.; Maresca, A.; Wagner, J.; Avvaru, B. S.; Robbins, A. H.; Scozzafava, A.; McKenna, R.; Supuran, C. T. Sulfonamides incorporating 1,3,5-triazine moieties selectively and potently inhibit carbonic anhydrase transmembrane isoforms IX, XII and XIV over cytosolic isoforms i and II: Solution and X-ray crystallographic studies. *Bioorganic Med. Chem.* **2011**, *19*, 3105–3119, doi:10.1016/j.bmc.2011.04.005.
51. Innocenti, A.; Durdagi, S.; Doostdar, N.; Amanda Strom, T.; Barron, A. R.; Supuran, C. T. Nanoscale enzyme inhibitors: Fullerenes inhibit carbonic anhydrase by occluding the active site entrance. *Bioorganic Med. Chem.* **2010**, *18*, 2822–2828, doi:10.1016/j.bmc.2010.03.026.
52. Nosik, D. N.; Lyalina, I. K.; Kalnina, L. B.; Lobach, O. A.; Chataeva, M. S.; Rasnetsov, L. D. The antiretroviral agent Fullevir. *Vopr. Virusol.* **2009**, *54*, 41–43.
53. Sijbesma, R.; Srdanov, G.; Wudl, F.; Castoro, J. A.; Wilkins, C.; Friedman, S. H.; DeCamp, D. L.; Kenyon, G. L. Synthesis of a Fullerene Derivative for the Inhibition of HIV Enzymes. *J. Am. Chem. Soc.* **1993**, *115*, 6510–6512, doi:10.1021/ja00068a006.
54. Kang, B.; Yu, D.; Dai, Y.; Chang, S.; Chen, D.; Ding, Y. Cancer-cell targeting and photoacoustic therapy using carbon nanotubes as “bomb” agents. *Small* **2009**, *5*, 1292–1301, doi:10.1002/smll.200801820.
55. Horie, M.; Fukuhara, A.; Saito, Y.; Yoshida, Y.; Sato, H.; Ohi, H.; Obata, M.; Mikata, Y.; Yano, S.; Niki, E. Antioxidant action of sugar-pendant C<sub>60</sub>-fullerenes. *Bioorganic Med. Chem. Lett.* **2009**, *19*, 5902–5904, doi:10.1016/j.bmcl.2009.08.067.
56. Chaudhuri, P.; Paraskar, A.; Soni, S.; Mashelkar, R. A.; Sengupta, S. Fullereneol-cytotoxic conjugates for cancer chemotherapy. *ACS Nano* **2009**, *3*, 2505–2514, doi:10.1021/nn900318y.
57. Alterio, V.; Di Flore, A.; D'Ambrosio, K.; Supuran, C. T.; De Simone, G. X-ray crystallography of carbonic anhydrase inhibitors and its importance in drug design. *Drug Des. Zinc-Enzyme Inhib.* **2009**, 73–138.
58. Küçükbay, F. Z.; Küçükbay, H.; Tanc, M.; Supuran, C. T. Synthesis and carbonic anhydrase I, II, IV and XII inhibitory properties of N-protected amino acid-sulfonamide conjugates. *J. Enzyme Inhib. Med. Chem.* **2016**, *31*, 1476–1483, doi:10.3109/14756366.2016.1147438.
59. Küçükbay, F. Z.; Küçükbay, H.; Tanc, M.; Supuran, C. T. Synthesis and carbonic anhydrase inhibitory properties of amino acid – coumarin/quinolinone conjugates incorporating glycine, alanine and phenylalanine moieties. *J. Enzyme Inhib. Med. Chem.* **2016**, *31*, 1198–1202, doi:10.3109/14756366.2015.1113173.
60. Küçükbay, F. Z.; Buğday, N.; Küçükbay, H.; Tanc, M.; Supuran, C. T. Synthesis, characterization and carbonic anhydrase inhibitory activity of novel benzothiazole derivatives. *J. Enzyme Inhib. Med. Chem.* **2016**, *31*, 1221–1225, doi:10.3109/14756366.2015.1114931.
61. Lomelino, C. L.; Supuran, C. T.; McKenna, R. Non-classical inhibition of carbonic anhydrase. *Int. J. Mol. Sci.* **2016**, *17*, doi:10.3390/ijms17071150.
62. Žalubovskis, R. In a search for selective inhibitors of carbonic anhydrases: coumarin and its bioisosteres -- synthesis and derivatization. *Chem. Heterocycl. Compd.* **2015**, *51*, 607–612, doi:10.1007/s10593-015-1748-8.
63. Kolb, H. C.; Walsh, J. C.; Kasi, D.; Mocharla, V.; Wang, B.; Gangadharmath, U. B.; Duclos, B. A.; Chen, K.; Zhang, W.; Chen, G.; Padgett, H. C.; Karimi, F.; Scott, P. J. H.; Gao, Z.; Liang, Q.; Collier, T. L.; Zhao, T.; Xia, C. Development of triazole derivatives as molecular imaging probes for carbonic anhydrase-IX using click chemistry. *PCT Int. Appl.* **2008**, 188pp.

64. Peeters, S. G. J. A.; Dubois, L.; Lieuwes, N. G.; Laan, D.; Mooijer, M.; Schuit, R. C.; Vullo, D.; Supuran, C. T.; Eriksson, J.; Windhorst, A. D.; Lambin, P. [18F]VM4-037 MicroPET Imaging and Biodistribution of Two In Vivo CAIX-Expressing Tumor Models. *Mol. Imaging Biol.* **2015**, *17*, 615–619, doi:10.1007/s11307-015-0831-y.
65. Doss, M.; Kolb, H. C.; Walsh, J. C.; Mocharla, V. P.; Zhu, Z.; Haka, M.; Alpaugh, R. K.; Chen, D. Y. T.; Yu, J. Q. Biodistribution and radiation dosimetry of the carbonic anhydrase IX imaging agent [(18)F]VM4-037 determined from PET/CT scans in healthy volunteers. *Mol. Imaging Biol.* **2014**, *16*, 739–746, doi:10.1007/s11307-014-0730-7.
66. Rutkauskas, K.; Zubrienė, A.; Tumosienė, I.; Kantminienė, K.; Kažemėkaitė, M.; Smirnov, A.; Kazokaitė, J.; Morkūnaitė, V.; Čapkauskaitė, E.; Manakova, E.; Gražulis, S.; Beresnevičius, Z.; Matulis, D. 4-Amino-substituted Benzenesulfonamides as Inhibitors of Human Carbonic Anhydrases. *Molecules* **2014**, *19*, 17356–17380, doi:10.3390/molecules191117356.
67. Vaškevičienė, I.; Paketurytė, V.; Zubrienė, A.; Kantminienė, K.; Mickevičius, V.; Matulis, D. N-Sulfamoylphenyl- and N-sulfamoylphenyl-N-thiazolyl-β-alanines and their derivatives as inhibitors of human carbonic anhydrases. *Bioorg. Chem.* **2017**, *75*, 16–29, doi:10.1016/j.bioorg.2017.08.017.
68. Ceruso, M.; Antel, S.; Vullo, D.; Scozzafava, A.; Supuran, C. T. Inhibition studies of new ureido-substituted sulfonamides incorporating a GABA moiety against human carbonic anhydrase isoforms I–XIV. *Bioorganic Med. Chem.* **2014**, *22*, 6768–6775, doi:10.1016/j.bmc.2014.10.041.
69. Moeker, J.; Mahon, B. P.; Bornaghi, L. F.; Vullo, D.; Supuran, C. T.; McKenna, R.; Poulsen, S. A. Structural insights into carbonic anhydrase IX isoform specificity of carbohydrate-based sulfamates. *J. Med. Chem.* **2014**, *57*, 8635–8645, doi:10.1021/jm5012935.
70. Shay, G.; Lynch, C. C.; Fingleton, B. Moving targets: Emerging roles for MMPs in cancer progression and metastasis. *Matrix Biol.* **2015**, *44–46*, 200–6, doi:10.1016/j.matbio.2015.01.019.
71. Zhong, Y.; Lu, Y. T.; Sun, Y.; Shi, Z. H.; Li, N. G.; Tang, Y. P.; Duan, J. A. Recent opportunities in matrix metalloproteinase inhibitor drug design for cancer. *Expert Opin. Drug Discov.* **2018**, *13*, 75–87.
72. Bonnans, C.; Chou, J.; Werb, Z. Remodelling the extracellular matrix in development and disease. *Nat. Rev. Mol. Cell Biol.* **2014**, *15*, 786–801.
73. Hidalgo, M.; Eckhardt, S. G. Development of Matrix Metalloproteinase Inhibitors in Cancer Therapy. *JNCI J. Natl. Cancer Inst.* **2001**, *93*, 178–193.
74. Scozzafava, A.; Supuran, C. T. Carbonic anhydrase and matrix metalloproteinase inhibitors: Sulfonylated amino acid hydroxamates with MMP inhibitory properties act as efficient inhibitors of CA isozymes I, II, and IV, and N-hydroxysulfonamides inhibit both these zinc enzymes. *J. Med. Chem.* **2000**, *43*, 3677–3687, doi:10.1021/jm000027t.
75. Nuti, E.; Orlandini, E.; Nencetti, S.; Rossello, A.; Innocenti, A.; Scozzafava, A.; Supuran, C. T. Carbonic anhydrase and matrix metalloproteinase inhibitors. Inhibition of human tumor-associated isozymes IX and cytosolic isozyme I and II with sulfonylated hydroxamates. *Bioorganic Med. Chem.* **2007**, *15*, 2298–2311, doi:10.1016/j.bmc.2007.01.023.
76. Korkmaz, N.; Obaidi, O. A.; Senturk, M.; Astley, D.; Ekin, D.; Supuran, C. T. Synthesis and biological activity of novel thiourea derivatives as carbonic anhydrase inhibitors. *J. Enzyme Inhib. Med. Chem.* **2015**, *30*, 75–80, doi:10.3109/14756366.2013.879656.
77. Mollica, A.; Costante, R.; Akdemir, A.; Carradori, S.; Stefanucci, A.; Macedonio, G.; Ceruso, M.; Supuran, C. T. Exploring new Probenecid-based carbonic anhydrase inhibitors: Synthesis, biological evaluation and docking studies. *Bioorganic Med. Chem.* **2015**, *23*, 5311–5318, doi:10.1016/j.bmc.2015.07.066.
78. Fidan, I.; Salmas, R. E.; Arslan, M.; Şentürk, M.; Durdagi, S.; Ekin, D.; Şentürk, E.; Coşgun, S.; Supuran, C. T. Carbonic anhydrase inhibitors: Design, synthesis, kinetic, docking and molecular dynamics analysis of novel glycine and phenylalanine sulfonamide derivatives. *Bioorganic Med. Chem.* **2015**, *23*, 7353–7358, doi:10.1016/j.bmc.2015.10.009.
79. Chiaramonte, N.; Bua, S.; Ferraroni, M.; Nocentini, A.; Bonardi, A.; Bartolucci, G.; Durante, M.; Lucarini, L.; Chiapponi, D.; Dei, S.; Manetti, D.; Teodori, E.; Gratteri, P.; Masini, E.; Supuran, C. T.; Romanelli, M. N. 2-Benzylpiperazine: A new scaffold for potent human carbonic anhydrase inhibitors. Synthesis, enzyme inhibition, enantioselectivity, computational and crystallographic studies and in vivo activity for a new class of intraocular pressure lowering agents. *Eur. J. Med. Chem.* **2018**, *151*, 363–375, doi:10.1016/j.ejmech.2018.04.002.