1 Review

2 Epigenetic targeting of aberrant transcriptional

3 modulation in pancreatic cancer

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Abstract: While the mortality rates of cancer are generally declining, pancreatic cancer persists to be an exception with a 5-year-survival rate of less than 7%. Late diagnosis and resistance to conventional therapies contribute to high mortality rates in spite of the remarkable recent advances in cancer management and research. Consequently, there is an urgent need to find new and unconventional therapeutic targets to improve prognosis and survival of pancreatic cancer patients. In this review, we discuss the transcriptional effects of the most widely used epigenetic inhibitors in pancreatic cancer focusing on Bromodomain and Extraterminal domain (BET) and Histone Deacetylase (HDAC) inhibitors, which are currently highly promising therapeutic options. We suggest that these inhibitors can be better utilized at lower doses which exploit their transcriptional modulatory effects on pancreatic cancer transcriptional programs directed by specific factors such as MYC and FOXA1, rather than simply based on their anti-proliferative effects. This approach can potentially help avoid the intolerable adverse events frequently elicited by the use of these treatments at higher doses. In particular, we underscore the crucial role of distal regulatory elements in mediating the specific effects of these epigenetic inhibitors and propose using them in a more selective and prudent manner.

Keywords: BET inhibitors; HDAC inhibitors; pancreatic cancer; aberrant transcription; enhancers; transcription factors; distal regulatory elements; MYC; FOXA1; BRD4

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1. Introduction.

- While the mortality rates of cancer are generally declining, pancreatic cancer persists to be an exception with a 5-year-survival rate of less than 7% [1,2]. Late diagnosis and resistance to conventional therapies contribute to high mortality rates in spite of the remarkable recent advances in cancer management and research [3]. Consequently, there is an urgent need to find new and unconventional therapeutic targets to improve prognosis and survival of pancreatic cancer patients.
- 33 In addition to various genomic mutations, such as KRAS and TP53, that play a role in the 34 pathophysiology of pancreatic cancer, other mutations and signaling pathways play an equally 35 important role by affecting transcription of entire subsets of genes, irrespective of genomic sequence 36 [2-8]. Epigenetic pathways affect transcription either via modulation of histone modifications which 37 can be activating or silencing, DNA methylation-mediated silencing, non-coding RNAs, and 38 alteration of chromatin accessibility [9]. This meshwork provides the cells with various tools that can 39 dramatically affect its transcriptome without the need to induce any irreversible changes at the level 40 of the genome. Moreover, in contrast to the permanent and largely "all or nothing" effects of 41 genomic mutations, modulation of the epigenome allows for more subtle, reversible changes in
- 42 genome regulation.

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Epigenetics represents a promising target in pancreatic cancer for various reasons. Firstly, epigenetic modifications are mainly mediated by enzymes and proteins whose activity can (at least in principle) be targeted by small molecule inhibitors. Secondly, many epigenetic pathways were found to be deregulated in pancreatic cancer, suggesting a crucial role of epigenetic regulation in this malignancy [4-7]. Additionally, pancreatic cancer, among others, was found to be addicted to the activation of aberrant transcriptional programs that not only drive the development and progression of cancer, but are also crucial for the maintenance of detrimental malignant characteristics such as metastasis and chemoresistance [10,11]. Epigenetic pathways were found to be major drivers of the perturbations of such programs in a highly intricate and context-specific manner [12]. Due to its tremendous pliability, epigenetic modulation provides an optimal tool to be hijacked in cancer development and progression creating a specific dependence of cancer cells on these pathways. Accordingly, many epigenetic inhibitors are currently under investigation for the treatment of numerous malignancies, including pancreatic cancer [3,7,13,14]. However, results from these studies so far have been unexpectedly modest and, in some cases, these inhibitors were associated with intolerable toxicities. Many of these studies use epigenetic inhibitors, whether alone or in combination, in a method akin to other more conventional drugs with defined targets like chemotherapy and monoclonal antibodies. On the other hand, epigenetic inhibitors target transcriptional regulation in a complex and unconventional manner and this should be taken into consideration when investigating these drugs.

Numerous reviews have deeply discussed and extensively summarized the recent advances of targeting epigenetics in cancer, in general, as well as in pancreatic cancer in particular [3,9,15-20]. In this review, we focus on the transcriptional mechanisms of the most widely used epigenetic inhibitors in pancreatic cancer focusing on Bromodomain and Extraterminal domain (BET) and Histone Deacetylase (HDAC) inhibitors, which represent promising therapeutic options. We suggest that these inhibitors can be better utilized for their transcriptional modulation, rather than solely on their anti-proliferative effects, which can lead to intolerable adverse events. Moreover, we underscore the crucial role of distal regulatory elements in mediating the specific effects of these epigenetic inhibitors and propose using them in a more selective and prudent manner.

2. BET inhibitors in pancreatic cancer

The BET family of proteins consists of BRD2, BRD3, BRD4, and the testis-specific BRDT [21]. All BET family members contain two conserved bromodomains which enable them to recognize acetylation marks on the chromatin, in addition to an extraterminal domain which enables interactions with other proteins [22,23]. The BET family has attracted much attention due to its significant role in gene transcription regulation in addition to its implication in the development of the particularly aggressive NUT midline carcinoma, which is characterized by the presence of a BRD4-NUT fusion gene [24,25]. The synthesis of the prototype BET inhibitor JQ1, which competitively binds to the acetylation-recognizing hydrophobic pockets in all BET members, has marked an explosion in the number of studies investigating the role of BET family members in gene transcription regulation due to their promising anti-proliferative effects in different cancer types [25,26]. However, many aspects are still unknown about the role of these factors in driving transcriptional activation and the best way to leverage their context-specific effects.

2.1. Role and effects of BET inhibitors in pancreatic cancer

Interestingly, a general screen for limiting epigenetic regulators in pancreatic cancer identified two members of the BET family, BRD2 and BRD3, as major drivers in pancreatic cancer growth and progression [27]. In pancreatic cancer cell lines, BET inhibition exerts anti-proliferative effects by selectively targeting inflammatory and oncogenic pathways [28]. This effect was also observed even in pancreatic cells with chemoresistant and highly migratory phenotypes [29]. Concordantly, BET inhibitors were found to decrease tumor growth in patient-derived xenografts by attenuating inflammatory pathways in cancer cells and their associated fibroblasts [27,30,31]. Recently,

Andricovich et al. [32] showed that pancreatic cells deficient for the lysine demethylase, KDM6A, are more sensitive to BET inhibition due to the activation of BET-dependent super enhancers. Moreover, the *MYC* proto-oncogene, perhaps the best known target for BET inhibition, was successfully used to subgroup 55 pancreatic cancer patient-derived xenografts based on its expression and accurately predicted sensitivity to BET inhibition via apoptosis with high expression of *MYC* correlating with more BET dependence [33].

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Thus, it has become very clear that BET inhibitors have a perceptible anti-proliferative effect in pancreatic cancer and display a very promising potential as highly effective and selective therapeutic agents. These inhibitory effects may largely be due to BET members being crucial for driving aberrant transcriptional programs and can be heightened via specific deficiencies which lead to increased transcriptional dependence on BET family members. This supports the current efforts to identify certain subgroups of patients who may be more responsive to BET inhibition. However, it is important to note that the effects of BET inhibition are varied and highly specific for different tumor types and subgroups.

2.2 BET inhibition and metastasis in pancreatic cancer

108 Metastasis is a major contributing factor to the very poor prognosis of the majority of pancreatic 109 cancer patients [34]. Consequently, therapeutic agents that attenuate and/or prevent metastasis can 110 be of extreme benefit to patients. Recently, it was uncovered that pancreatic mouse organoids from 111 metastatic pancreatic ductal adenomas show a marked reprogramming in their enhancer landscape 112 compared to organoids originated from normal pancreata, early neoplastic (PanIN) lesions or 113 primary tumors [35]. The same pattern of reprogramming was observed in pancreatic cancer 114 patients [36] and also in other cancer types including osteosarcoma, ependymoma, and 115 rhabdomyosarcoma [37-39]. Therefore, it is clear that distal regulatory elements play a significant 116 role in activating metastatic programs in different cancer types. This strongly suggests that targeting 117 enhancers can be a highly efficient approach in managing metastatic pancreatic cancer and 118 potentially preventing metastasis from primary sites.

119 Interestingly, recent studies implied that BET family members play an important role in modulating 120 gene transcription through regulation of the 3D chromatin structure [40-42]. This structure creates 121 specific compartmentalization which enable enhancers to interact with and affect specific target 122 genes [43,44]. Moreover, we have previously shown that BET-dependent genes are not necessarily 123 highly enriched for BRD4, the best well-studied member of the BET family [45]. Instead, while the 124 genes that were highly affected by JQ1 treatment did not have a defined pattern of occupancy for 125 BRD4 at their respective transcription starting site (TSS), they did display a better correlation with 126 tissue-specific BRD4-occupied enhancers. Consistently, Cao et al. [46] reported that expression of 127 genes can be predicted in part by the activity of their enhancers alone. While this phenomena was 128 observed in other systems, it is highly probable that gene dependence follows the same specificity 129 paradigms in various contexts [47]. Indeed, BET inhibitors were observed to exert marked 130 anti-proliferative effects in metastatic melanoma via deactivating the super enhancer of the 131 oncogenic Adhesion Molecule With Ig Like Domain 2 (AMIGO2) [48]. The BET-dependent super 132 enhancer activating Aldehyde Dehydrogenase 2 Family (ALDH), which promotes resistance to 133 chemotherapy and disease recurrence, was identified as a promising target in ovarian cancer [49]. In 134 the highly metastatic Merkel cell carcinoma, BET inhibition is highly effective as it downregulates 135 MYC by targeting its putative super enhancer [50]. Altogether, BET inhibitors are strong candidates 136 for treating metastatic pancreatic cancer and can potentially be used as adjuvant therapies to prevent 137 metastasis and disease recurrence if found safe in further clinical studies. These inhibitors in 138 particular can play this unique role because, as previously discussed, reprogramming of distal 139 regulatory elements is a hallmark of metastatic phenotype and these enhancers are frequently 140 particularly sensitive to BET inhibition.

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141 2.3 Effect of BET inhibition on distal regulatory elements

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142 Recent efforts focusing on uncovering the mechanisms by which BET inhibitors affect the 143 proliferation of cancer cells in different systems largely confirmed the implication of distal 144 regulatory elements in mediating the observed effects. Interestingly, a particular subtype of these 145 elements called "super enhancers" are remarkably associated with BET members and can be partly 146 defined by the intensity of BRD4 occupancy at these regions [21,51]. We have previously tested the 147 validity of the super enhancer concept which suggests that enhancers follow the rule of the vital few 148 (the Pareto principle) where a low percentage of regulatory elements are responsible for most effects 149 on the regulation of gene transcription [52]. While we have failed to discern such a pattern, many 150 studies have indeed defined certain dependencies of crucial enhancers, whether "super" or 151 "typical", on BET members.

Interestingly, BRD4 was observed to be more preferentially enriched at enhancers in diffuse large B-cell lymphoma and BET-dependent super enhancers of master regulatory transcription factors and were correlated with the anti-proliferative effects of JQ1 observed in this system [53]. Metabolic changes promoting proliferation in a specific subgroup of melanoma were reported to occur via upregulation of the BET-dependent super enhancer for PPARG Coactivator 1 Alpha (PGC-1α) [54]. In castration-resistant prostate cancer, BRD4 was localized at the BET inhibitor-sensitive enhancer regions of the driver oncogene, Transmembrane Protease, Serine 2 (TMPRSS2) [55,56]. In general, these studies imply that specific BET-dependent enhancers are activated in cancerous cells, rendering them more sensitive to BET inhibition and providing a specific therapeutic target. Intriguingly, recent methods that can detect nascent RNA such as SLAM-seq have shown that the effects of BET inhibition are dose-dependent and that high doses of BET inhibitors can lead to universal pausing of transcription while low concentrations affected specific subsets of genes in leukemia cells [57]. These hypersensitive genes were not necessarily controlled by super enhancers and correlated only in sub-clusters with other transcription factors and co-activators. This underscores the complexity of BET-orchestrated specific gene regulation, which may be associated with specific clusters of enhancers, but not exclusively explained by a "super enhancer" model.

168 In summary, investigating the efficacy of BET inhibitors in pancreatic cancer is highly justified given 169 the promising anti-proliferative effects seen in different models. Additionally, recent data from 170 pancreatic cancer and other systems identify a general pattern of BET-mediated activation of 171 enhancers, or clusters thereof, that play a major role in driving detrimental aberrant transcriptional 172 programs like metastasis. However, the mechanism of action of BET inhibitors is still not fully 173 understood and is likely highly complex and context-dependent. Many challenges exist in the 174 investigation of the role of these inhibitors including the limitations of current techniques, 175 measuring the anti-proliferative effects for BET inhibitors as a general read-out for efficacy, and the 176 prejudiced focus of research on only one member of the BET family, BRD4. While BRD2 and BRD3 177 share a high homology with BRD4, their functions and specific roles are not well-defined [58,59]. 178 BRD2 has garnered some attention due to recent reports that it frequently co-localizes with CTCF, an 179 important insulator protein that demarcates transcriptional boundaries [42]. Both BRD2 and BRD3 180 affect gene transcription through different mechanisms than BRD4, namely by interaction with 181 specific transcription factors like E2F and GATA1, respectively [60,61]. In leukemia, the knockdown 182 of either BRD2 or BRD3 failed to recapitulate the effects of BET inhibition or BRD4 manipulation, 183 implying that they have a lesser role in this system, in contrast to the reported role in pancreatic 184 cancer [27,57]. Adding to the complexity is the proposed various roles of the different 185 bromodomains within each member, namely BD1 and BD2 [62]. A deeper understanding of each of 186 the BET proteins as well as the importance of each of their bromodomains will enable us to use these 187 and second generation agents safely and effectively and successfully combine them with other 188 agents. Generally, BET inhibitors exert promising synergistic effects in pancreatic cancer with other 189 agents, such as chemotherapeutic drugs like gemcitabine [30] and epigenetic inhibitors like HDAC 190 inhibitors [63].

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191 3. HDAC inhibitors in pancreatic cancer

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192 Histone modifications can be active marks like acetylation of histone 3 at lysine 27 (H3K27ac) and 193 methylation of histone 3 at lysine 4 (H3K4me1), or repressing marks like trimethylation of histone 3 194 at lysine 27 (H3K27me3) or ubiquitination of histone 2A at lysine 119 (H2Aub) [64]. Histone 195 acetylation is a marker associated with transcriptional activation and its manipulation can be 196 beneficial in attenuating detrimental pathways in pancreatic cancer. HDAC inhibitors which inhibit 197 the removal of protein lysine acetylation marks have a broader effect on gene expression [65] in 198 comparison to BET inhibitors. HDAC inhibitors are classified based on their homology to yeast into 199 different classes (Class I-IV) [66]. These classes differ in their domains, expression and effects, and 200 their different roles are still to be clearly defined. HDAC inhibitors differ in their potency and 201 selectivity but mainly show promising anti-proliferative effects in cancer with the main rationale of 202 reactivating silenced tumor suppressor genes and reversing deregulated deacetylation of histones in 203 cancer [67,68].

3.1. Role and effects of HDAC inhibitors in pancreatic cancer

HDAC inhibitors were observed to induce p53-mediated pro-apoptotic effects in pancreatic cancer cells [69]. Additionally, they selectively inhibited proliferation of pancreatic cancer cells by affecting their aerobic metabolism and rendering them more sensitive to glycolytic inhibition [70,71]. In general, specific HDAC inhibitors that targeted certain HDAC classes showed variable effects with pan-HDAC inhibitors causing the most marked anti-proliferative effects [72]. While HDAC inhibition was found to potentiate the effects of gemcitabine in vitro and was reported to overcome its resistance, a clinical study combining HDAC inhibitors with gemcitabine in pancreatic cancer patients was prematurely terminated because the observed benefits did not outweigh the marked adverse events [73-76]. HDAC inhibitors were also reported to suppress metastasis as HDACs were described to mediate the repressor action of the Zinc Finger E-Box Binding Homeobox 1 (ZEB1) on the promoter of the known epithelial marker, calcium-dependent adhesion protein-1 (CDH1) [77-79]. Interestingly, we have also shown that HDAC inhibition attenuates epithelial-to-mesenchymal transition (EMT) and decreases stem-like properties in pancreatic cancer cells [80]. Given that both BET and HDAC inhibition show anti-proliferative and metastasis-suppressive effects in pancreatic cancer, Mazur et al. [63] combined these two agents in vitro and in vivo and observed a synergistic effect mediated by upregulation of the pro-apoptotic p57 protein. Synergy between those two agents appears at first glance to be paradoxical due to the fact that HDAC inhibitors stabilize a histone mark whose recognition is blocked by BET inhibition. However, both agents may work by attacking related transcriptional mechanisms. As previously mentioned, distal regulatory elements play a significant role in the mechanism of action of BET inhibitors. In contrast, the effects of HDAC inhibitors at these regions are less often reported. Recent studies have investigated the role of HDAC inhibitors on enhancer activity, which might further clarify the mechanisms by which these inhibitors act and identify new approaches to use them safely and effectively.

3.2. HDAC inhibition role at distal regulatory elements

229 As HDAC inhibitors stabilize a histone mark associated with active transcription, it is expected that 230 it will lead mainly and directly to an upregulation of dependent genes. Surprisingly, we have 231 detected a significant set of genes that are downregulated upon treatment with selective inhibitors of 232 class I HDACs in the highly metastatic pancreatic cell line L3.6pl [80]. We have observed that while 233 promoters of these downregulated genes gain acetylation as expected, individual associated distal 234 regulatory elements of these genes show a dramatic loss of acetylation and better correlation with 235 gene regulation. This has also been observed in the colorectal cancer cell line, HCT116, where 236 treatment with an HDAC inhibitor also leads to the loss of H3K27ac at certain enhancer regions in a 237 concentration-dependent manner [81]. However, this decrease is not universally observed at all 238 enhancers as an increase of H3K27ac by HDAC inhibition at the enhancer of the pro-apoptotic B cell 239 lymphoma-2 like 11 (BIM) gene was reported in triple negative breast cancer [82]. Consistent with a

particular effect on enhancer activity, HDAC inhibition has been shown to repress enhancer RNA (eRNA) expression to a higher extent than BET inhibition in breast cancer cells [83].

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Therefore, it can be concluded that HDAC inhibitors affect distal regulatory elements in a manner that is distinct from promoters, probably decreasing acetylation at a subset of specific enhancers, thereby affecting gene transcription in a more complex and diverse manner. Such regulation can be partially explained by the status of activation of the enhancer itself prior to treatment [81]. Enhancers can either be in an active state and marked by H3K27ac and H3K4me1, inactive with only H3K27me3, or poised with a lower threshold to be activated via being marked by the H3K4me1 active mark and H3K27me3 repressive mark [84]. Sanchez et al. [81] demonstrated that enhancers which are already active are usually inactivated by HDAC inhibition while poised enhancers show a tendency to be activated. Another very important aspect that may play a central role in defining the response of an enhancer to HDAC inhibition is the repertoire of transcription factors expressed in the cellular system and their importance for cellular phenotype and tumorigenic gene expression. It was reported that HDAC inhibitors not only exert their effects on acetylated histones, but also on acetylated transcription factors [85,86]. Notably, acetylation of the pioneer transcription factor, Forkhead Box A1 (FOXA1) was shown to directly and negatively affect its ability to bind chromatin [87]. Remarkably, FOXA1 was recently shown to be specifically enriched in enhancer regions that are gained in metastatic pancreatic organoids [35]. Accordingly, we postulate that HDAC inhibitors can be used to attenuate the binding of FOXA1 to these enhancers leading to their deactivation. Indeed, enhancer regions that we identified as being lost following HDAC inhibitor treatment were found to be enriched for FOXA1 occupancy in another pancreatic cancer cell line [88] (Figure 1a). Interestingly, genes targeted by this mechanism were unaffected by BET inhibitor treatment [88]. Thus, combined treatment with HDAC and BET inhibitors can potentially simultaneously target different sets of activated enhancers in order to synergistically and more effectively decrease the activation of reprogrammed enhancers activating aberrant transcriptional programs like metastasis. This presents a model in which these two apparently counteracting agents can work together forming a successful therapeutic regimen in metastatic pancreatic cancer (Figure 1b).

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4. Enhancers as an optimal paradigm for therapeutic targeting of pancreatic cancer

270 Based on the evidence discussed above, it is clear that distal regulatory elements play a special role 271 in the scope of the effects of BET and HDAC inhibitors. Thus, it is highly probable that these 272 elements will emerge as a major target of therapy in multiple diseases in the upcoming years. Many 273 positive attributes contribute to the adequacy of enhancers as a target for therapy and manipulation. 274 To activate a certain gene, the transcriptional machinery has to be recruited by transcription factors 275 to the promoters of these genes [89]. Diversity in transcription factor recruitment and abundance are 276 thus very important in regulating gene activation in different contexts and systems. Distal 277 regulatory elements provide a platform with vast variability and substantial magnitude for 278 recruitment of various transcription factors, thereby enabling regulation of gene transcription in a 279 temporal and spatial manner. This means that in certain systems, driver oncogenic pathways can be 280 activated by different transcription factors and enhancers, thus creating a dependence which can be 281 specifically targeted. For example, different enhancers drive the activation of MYC in various 282 systems. In colorectal cancer, the long non-coding RNA, Colon Cancer Associated Transcript 1 283 (CCAT1) is highly active and plays a significant role in MYC activation [90,91]. Consistent with the 284 importance of the Wnt signaling pathway in colorectal cancer, we observed that this enhancer is 285 highly occupied by Wnt-responsive β-catenin-dependent transcription factor TCF7L2 (Figure 2) [91]. 286 This implies that Wnt-signaling mediated activation of MYC in colorectal cancer utilizes a specific 287 mechanisms of activation which can be potentially be targeted by HDAC inhibitors, as they were 288 reported to deplete TCF7L2 [92]. Interestingly, CCAT1 was also reported to be play a tumorigenic 289 role in pancreatic cancer [93]. In other systems like prostate cancer, MYC is activated by a different 290 enhancer called Prostate Cancer Associated Transcript 1 (PCAT1). Consistent, with the androgen

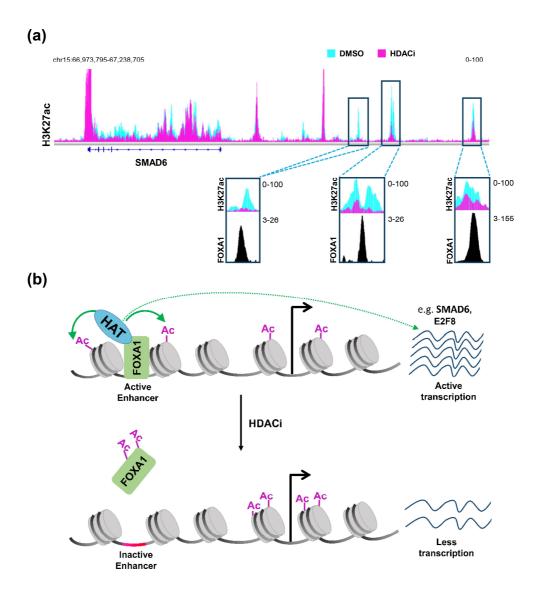


Figure 1. HDAC inhibition and FOXA1 at distal regulatory elements. (a) H3K27ac profiles in L3.6pl cells at *SMAD6*, a gene that is downregulated upon treatment with class I HDAC inhibitor (HDACi) showing distal regulatory regions that dramatically lose H3K27ac and co-localize with FOXA1 in the CFPAC1 pancreatic cancer cell line; (b) A schematic model showing an enhancer region activated by FOXA1 leading to acetylation of histones by a histone acetyltransferase (HAT), which leads to the activation of the gene. Upon treatment with HDACi, increased FOXA1 acetylation attenuates its binding to chromatin leading to downregulation of the gene.

receptor (AR) being a primary driver of prostate cancer, we observed this enhancer to be particularly occupied by AR in LNCaP prostate cancer cells [90,94-96]. Alternatively, the *PVT1* oncogene is active in many other systems like leukemia and plays a similar role as an enhancer of the *MYC* gene [97]. Analogous to the tumor- and context-specific effects shown for TCF7L2 and AR in colorectal and prostate cancers, respectively, we observed an enrichment of the hematopoietic transcription factor GATA-2 on the *MYC* enhancer within the *PVT1* gene [91]. Together, these examples show the complexity by which diverse distal regulatory elements utilize specific transcription factor repertoires to induce common oncogenic pathways. We postulate that distinct, but similar transcription factor networks will also be discovered in pancreatic cancer which can be specifically targeted by inhibitors of BET, HDAC or other epigenetic regulators. Importantly, activation of

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oncogenes such as *MYC* by alternative BET-independent transcriptional pathways can lead to resistance to BET inhibitors in leukemia cells [98]. Thus, identifying which enhancers are specifically active in pancreatic cancer cells and identifying and targeting their dependencies will play an important role in the optimal application of BET inhibitors in the treatment of pancreatic cancer. Furthermore, targeting tumor-specific enhancer regions will be more likely to spare normal cells and may possibly lead to less long term adverse effects [99]. Future studies will test the validity of this rationale upon successful prolongation of patient survival.

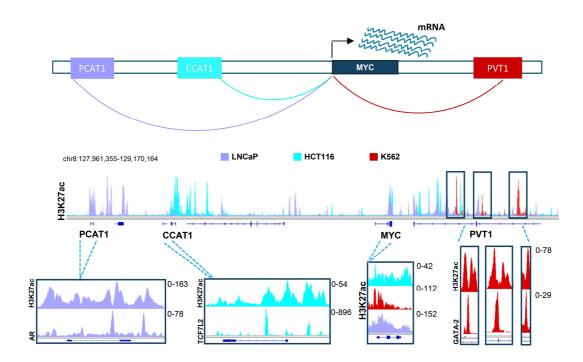


Figure 2. A schematic model showing distal regulatory regulatory mechanisms controlling the expression of the *MYC* gene in different systems, namely in prostate cancer (shown in light blue), colorectal cancer (shown in aqua), and leukemic cell lines (shown in orange). The H3K27ac layered profile in these different cell lines uncover differential activation of unique enhancers in each system. These enhancers are specifically enriched with driver transcription factors (androgen receptor, AR; TCF7L2 and GATA2), which are hallmarks of these tumor types.

5. Targeting transcription factors in pancreatic cancer: a code for specificity

Enhancers are known to be highly-bound by specific transcription factors that mediate transcriptional activation of target gene expression through the recruitment of other activators and transcription initiators [100]. When transcription factors co-localize with a factor of interest, BETs for example, these factors will naturally play a significant role in its mechanism and effects. Interestingly, the BET context-specific effects discerned in the vast body of literature available can probably be explained by the different interacting factors at specific sites where BET members co-localize with other transcription factors. BET-dependent enhancers require BET members and certain transcription factors for their activation, while other BET-independent enhancers might have other factors and activation pathways which make them tolerant to the loss of one of many activators upon BET inhibition. Furthermore, the different expression levels of these transcription factors in different systems, whether absent or highly or lowly expressed, in addition to their pioneering potential can also play a role in enhancer dependency and activity.

Paradoxically, super enhancers are by definition normally highly enriched for transcription factor binding, which can theoretically render them less dependent on one particular factor [101].

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323 However, this is not usually the case given that, as previously discussed, BET inhibition has the 324 ability to turn off certain BET-dependent super enhancers but deletion of individual components of 325 "super enhancers" does not equally impair the activation of the target gene [53,54,90,102,103]. Thus, 326 it appears that BET proteins serve to integrate the activity of transcription factors at BET-dependent 327 enhancers in a manner such that the sum of the activity of the transcription factor binding is greater 328 than that of the components. Subsequently, once a better understanding of transcription factor 329 function at enhancers and promoters and the dependence of each on BET proteins has been 330 achieved, targeting specific transcription factors or their upstream signaling pathways, possibly 331 alone or in combination with BET inhibitors, can provide us with a new layer of specific gene 332 transcriptional manipulation.

333 However, therapeutically targeting transcription factors can be quite challenging. One approach can 334 be targeting signaling pathways that control the activity of the transcription factor of interest. For 335 example, the transcription factor Endothelial PAS Domain Protein 1 (EPAS1 or HIF2A) was 336 demonstrated to play a role in promoting pancreatic cancer in cells and mice [104]. A crosstalk with 337 Wnt-signaling was identified in this system which uncovers a new target that can potentially be 338 inhibited. Furthermore, TGFβ signaling was reported to cooperate with mutant p53 to mediate distal 339 metastasis in pancreatic cancer mouse models [105]. The activation of NF-κB, which also promotes 340 EMT in pancreatic cancer, can also be inhibited by blocking its activation via $I\kappa B\alpha$ phosphorylation 341 [106]. Another approach to target transcription factors is to attenuate their recruitment by affecting 342 their ability to bind chromatin as previously mentioned for FOXA1 and its acetylation. This can also 343 be achieved by designing specific inhibitors that prevent the binding of DNA to a certain 344 transcription factor [107], however this approach has proven to be very difficult for therapeutic 345 application. Another approach can be targeting cooperating transcription factors. For example, 346 NF-κB cooperates with ETS transcription factors to recruit BET members to activate genes, rendering 347 these sensitive to BET inhibition [108,109]. In general, transcription factors can play specific roles in a 348 context-dependent manner based on the combinatorial repertoire of transcription factors expressed, 349 thereby enabling a given transcription factor to activate a different set of genes and programs, 350 dependent upon the expression of other factors. Therefore, identifying important transcription 351 factors playing a role in aberrant transcriptional activation may uncover specific targets that can be 352 manipulated by available inhibitors.

6. Conclusion: Unconventional epigenetic agents should be used in unconventional ways

Epigenetic agents are a special subclass of drugs whose targets and effects are dependent on the epigenetic and transcriptional landscape of each system. In general, a major trend is seen where low concentrations of these agents affect hypersensitive dependent genes and higher concentrations frequently display a more universal effect. So far, gene transcription modulatory agents have been used to initiate cell cycle arrest and/or apoptosis and mainly administered at high levels just under their maximum tolerable dose (MTD). Administering these agents at these doses likely influences their specificity and probably promotes many of the intolerable adverse effects that might lead to premature termination of clinical studies. In fact, higher doses of such inhibitors, which lead to cell cycle arrest, may in fact impede the activity of many chemotherapeutic agents, while lower concentrations which elicit specific transcriptional reprogramming may have minimal effects on their own, but significantly synergize with other therapies. In this review, we summarized data pertaining to the effects of BET and HDAC inhibitors, two of the most promising epigenetic agents, in pancreatic cancer. We draw the conclusion that these agents likely mediate their specific effects through distal regulatory elements. By investigating the potential utility of these agents in lower concentrations, we may be able to uncover their potential as safe adjuvant therapies in combination with other standard of care treatments to manage and prevent recurrence of pancreatic cancer and various malignancies in general.

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372 Appendix A

- Chromatin immune-precipitation profiles which are shown in Figure 1 and 2 as examples were downloaded from the Encyclopedia of DNA Elements (ENCODE) consortium when available (H3K27ac in HCT116: GSM2534277; TCF7L2 in HCT116: GSM782123; H3K27ac in K562: GSM733656;
- 376 GATA-2 in K562: GSM935373) [91]. Other profiles were downloaded from the European Nucleotide
- 377 Archive (H3K27ac in LNCaP: SRR2566837 [94]; AR in LNCaP: SRR4025870 [95]; H3K27ac in L3.6pl:
- 378 SRR5042516,18-21 [80]; FOXA1 in CFPAC1: SRR1736462 [88]). Reads were mapped to the hg19
- genome using BOWTIE/2.2.5 and converted to bam using SAMTOOLS/1.4. DEEPTOOLS/2.4.0 was
- $380\,$ used to produce bigwig files with ignoring the duplicates and extending the reads for 200 base pairs.
- 381 Bigwig files were viewed using IGV 2.4.
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- 385 Author Contributions: F.H.H. and S.A.J have conceived the ideas for this work and wrote the manuscript.
- 386 Conflicts of Interest: The authors declare no conflict of interest.

387 References

- Malvezzi, M.; Carioli, G.; Bertuccio, P.; Boffetta, P.; Levi, F.; La Vecchia, C.; Negri, E. European cancer mortality predictions for the year 2017, with focus on lung cancer. *Ann Oncol* **2017**, *28*, 1117-1123.
- 390 2. Waddell, N.; Pajic, M.; Patch, A.M.; Chang, D.K.; Kassahn, K.S.; Bailey, P.; Johns, A.L.; Miller, D.; 391 Nones, K.; Quek, K., *et al.* Whole genomes redefine the mutational landscape of pancreatic cancer. *Nature* **2015**, *518*, 495-501.
- 393 3. Hessmann, E.; Johnsen, S.A.; Siveke, J.T.; Ellenrieder, V. Epigenetic treatment of pancreatic cancer: is there a therapeutic perspective on the horizon? *Gut* **2017**, *66*, 168-179.
- 395 4. Jakel, C.; Bergmann, F.; Toth, R.; Assenov, Y.; van der Duin, D.; Strobel, O.; Hank, T.; Kloppel, G.; 396 Dorrell, C.; Grompe, M., *et al.* Genome-wide genetic and epigenetic analyses of pancreatic acinar cell carcinomas reveal aberrations in genome stability. *Nat Commun* **2017**, *8*, 017-01118.
- Jia, J.; Parikh, H.; Xiao, W.; Hoskins, J.W.; Pflicke, H.; Liu, X.; Collins, I.; Zhou, W.; Wang, Z.; Powell, J., et al. An integrated transcriptome and epigenome analysis identifies a novel candidate gene for pancreatic cancer. *BMC Med Genomics* **2013**, *6*, 1755-8794.
- 401 6. McCleary-Wheeler, A.L.; Lomberk, G.A.; Weiss, F.U.; Schneider, G.; Fabbri, M.; Poshusta, T.L.; Dusetti, N.J.; Baumgart, S.; Iovanna, J.L.; Ellenrieder, V., et al. Insights into the epigenetic mechanisms controlling pancreatic carcinogenesis. *Cancer Lett* 2013, 328, 212-221.
- 404 7. Bailey, P.; Chang, D.K.; Nones, K.; Johns, A.L.; Patch, A.M.; Gingras, M.C.; Miller, D.K.; Christ, A.N.; 405 Bruxner, T.J.; Quinn, M.C., *et al.* Genomic analyses identify molecular subtypes of pancreatic cancer. *Nature* **2016**, *531*, 47-52.
- 407 8. Vogelstein, B.; Kinzler, K.W. The multistep nature of cancer. *Trends in genetics*: TIG 1993, 9, 138-141.
- 408 9. Iguchi, E.; Safgren, S.L.; Marks, D.L.; Olson, R.L.; Fernandez-Zapico, M.E. Pancreatic Cancer, A 409 Mis-interpreter of the Epigenetic Language. *Yale J Biol Med* **2016**, *89*, 575-590.
- 410 10. Bradner, J.E.; Hnisz, D.; Young, R.A. Transcriptional Addiction in Cancer. Cell 2017, 168, 629-643.
- 411 11. Sharma, S.V.; Settleman, J. Oncogene addiction: setting the stage for molecularly targeted cancer therapy. *Genes Dev* **2007**, *21*, 3214-3231.
- 413 12. Alekseyenko, A.A.; Walsh, E.M.; Wang, X.; Grayson, A.R.; Hsi, P.T.; Kharchenko, P.V.; Kuroda, M.I.; 414 French, C.A. The oncogenic BRD4-NUT chromatin regulator drives aberrant transcription within large topological domains. *Genes Dev* 2015, 29, 1507-1523.
- 416 13. Iwahashi, S.; Utsunomiya, T.; Imura, S.; Morine, Y.; Ikemoto, T.; Arakawa, Y.; Saito, Y.; Ishikawa, D.; 417 Shimada, M. Effects of valproic acid in combination with S-1 on advanced pancreatobiliary tract cancers: clinical study phases I/II. *Anticancer research* 2014, 34, 5187-5191.

- 419 14. Abdelfatah, E.; Kerner, Z.; Nanda, N.; Ahuja, N. Epigenetic therapy in gastrointestinal cancer: the right combination. *Therapeutic advances in gastroenterology* **2016**, *9*, 560-579.
- 421 15. Lomberk, G.A.; Iovanna, J.; Urrutia, R. The promise of epigenomic therapeutics in pancreatic cancer. *Epigenomics* **2016**, *8*, 831-842.
- 423 16. Neureiter, D.; Jager, T.; Ocker, M.; Kiesslich, T. Epigenetics and pancreatic cancer: pathophysiology and novel treatment aspects. *World J Gastroenterol* **2014**, 20, 7830-7848.
- 425 17. Silverman, B.R.; Shi, J. Alterations of Epigenetic Regulators in Pancreatic Cancer and Their Clinical Implications. *Int J Mol Sci* **2016**, *17*.
- 427 18. Quilichini, E.; Haumaitre, C. Implication of epigenetics in pancreas development and disease. *Best* 428 *Pract Res Clin Endocrinol Metab* **2015**, 29, 883-898.
- 429 19. Klieser, E.; Swierczynski, S.; Mayr, C.; Schmidt, J.; Neureiter, D.; Kiesslich, T.; Illig, R. Role of histone deacetylases in pancreas: Implications for pathogenesis and therapy. *World J Gastrointest Oncol* **2015**, 7, 431 473-483.
- Damaskos, C.; Garmpis, N.; Karatzas, T.; Nikolidakis, L.; Kostakis, I.D.; Garmpi, A.; Karamaroudis, S.;
 Boutsikos, G.; Damaskou, Z.; Kostakis, A., *et al.* Histone Deacetylase (HDAC) Inhibitors: Current
 Evidence for Therapeutic Activities in Pancreatic Cancer. *Anticancer Res* **2015**, *35*, 3129-3135.
- Loven, J.; Hoke, H.A.; Lin, C.Y.; Lau, A.; Orlando, D.A.; Vakoc, C.R.; Bradner, J.E.; Lee, T.I.; Young, R.A. Selective inhibition of tumor oncogenes by disruption of super-enhancers. *Cell* **2013**, *153*, 320-334.
- Dhalluin, C.; Carlson, J.E.; Zeng, L.; He, C.; Aggarwal, A.K.; Zhou, M.M. Structure and ligand of a histone acetyltransferase bromodomain. *Nature* **1999**, *399*, 491-496.
- 439 23. Shi, J.; Vakoc, C.R. The mechanisms behind the therapeutic activity of BET bromodomain inhibition. *Mol Cell* **2014**, *54*, 728-736.
- 441 24. French, C.A. Pathogenesis of NUT midline carcinoma. *Annu Rev Pathol* **2012**, 7, 247-265.
- Filippakopoulos, P.; Qi, J.; Picaud, S.; Shen, Y.; Smith, W.B.; Fedorov, O.; Morse, E.M.; Keates, T.; Hickman, T.T.; Felletar, I., *et al.* Selective inhibition of BET bromodomains. *Nature* **2010**, *468*, 1067-1073.
- 444 26. Fu, L.L.; Tian, M.; Li, X.; Li, J.J.; Huang, J.; Ouyang, L.; Zhang, Y.; Liu, B. Inhibition of BET bromodomains as a therapeutic strategy for cancer drug discovery. *Oncotarget* **2015**, *6*, 5501-5516.
- 446 27. Huang, Y.; Nahar, S.; Nakagawa, A.; Fernandez-Barrena, M.G.; Mertz, J.A.; Bryant, B.M.; Adams, C.E.;
 447 Mino-Kenudson, M.; Von Alt, K.N.; Chang, K., et al. Regulation of GLI Underlies a Role for BET
 448 Bromodomains in Pancreatic Cancer Growth and the Tumor Microenvironment. Clinical cancer research
 449 : an official journal of the American Association for Cancer Research 2016, 22, 4259-4270.
- 450 28. Leal, A.S.; Williams, C.R.; Royce, D.B.; Pioli, P.A.; Sporn, M.B.; Liby, K.T. Bromodomain inhibitors, JQ1 and I-BET 762, as potential therapies for pancreatic cancer. *Cancer letters* **2017**, *394*, 76-87.
- 452 29. Sahai, V.; Kumar, K.; Knab, L.M.; Chow, C.R.; Raza, S.S.; Bentrem, D.J.; Ebine, K.; Munshi, H.G. BET bromodomain inhibitors block growth of pancreatic cancer cells in three-dimensional collagen.

 454 *Molecular cancer therapeutics* **2014**, *13*, 1907-1917.
- 455 30. Yamamoto, K.; Tateishi, K.; Kudo, Y.; Hoshikawa, M.; Tanaka, M.; Nakatsuka, T.; Fujiwara, H.; 456 Miyabayashi, K.; Takahashi, R.; Tanaka, Y., *et al.* Stromal remodeling by the BET bromodomain inhibitor JQ1 suppresses the progression of human pancreatic cancer. *Oncotarget* **2016**, *7*, 61469-61484.
- 458 31. Sherman, M.H.; Yu, R.T.; Tseng, T.W.; Sousa, C.M.; Liu, S.; Truitt, M.L.; He, N.; Ding, N.; Liddle, C.; Atkins, A.R., *et al.* Stromal cues regulate the pancreatic cancer epigenome and metabolome. *Proc Natl Acad Sci U S A* **2017**, *114*, 1129-1134.
- 461 32. Andricovich, J.; Perkail, S.; Kai, Y.; Casasanta, N.; Peng, W.; Tzatsos, A. Loss of KDM6A Activates Super-Enhancers to Induce Gender-Specific Squamous-like Pancreatic Cancer and Confers Sensitivity to BET Inhibitors. *Cancer Cell* 2018, 33, 512-526.
- 464 33. Bian, B.; Bigonnet, M.; Gayet, O.; Loncle, C.; Maignan, A.; Gilabert, M.; Moutardier, V.; Garcia, S.; 465 Turrini, O.; Delpero, J.R., *et al.* Gene expression profiling of patient-derived pancreatic cancer xenografts predicts sensitivity to the BET bromodomain inhibitor JQ1: implications for individualized medicine efforts. *EMBO Mol Med* **2017**, *9*, 482-497.

- 468 34. Uccello, M.; Moschetta, M.; Mak, G.; Alam, T.; Henriquez, C.M.; Arkenau, H.T. Towards an optimal treatment algorithm for metastatic pancreatic ductal adenocarcinoma (PDA). *Curr Oncol* **2018**, 25, e90-e94.
- 471 35. Roe, J.S.; Hwang, C.I.; Somerville, T.D.D.; Milazzo, J.P.; Lee, E.J.; Da Silva, B.; Maiorino, L.; Tiriac, H.; 472 Young, C.M.; Miyabayashi, K., et al. Enhancer Reprogramming Promotes Pancreatic Cancer Metastasis. *Cell* 2017, 170, 875-888.
- 474 36. McDonald, O.G.; Li, X.; Saunders, T.; Tryggvadottir, R.; Mentch, S.J.; Warmoes, M.O.; Word, A.E.; 475 Carrer, A.; Salz, T.H.; Natsume, S., *et al.* Epigenomic reprogramming during pancreatic cancer progression links anabolic glucose metabolism to distant metastasis. *Nat Genet* **2017**, *49*, 367-376.
- 477 37. Morrow, J.J.; Bayles, I.; Funnell, A.P.W.; Miller, T.E.; Saiakhova, A.; Lizardo, M.M.; Bartels, C.F.; 478 Kapteijn, M.Y.; Hung, S.; Mendoza, A., et al. Positively selected enhancer elements endow osteosarcoma cells with metastatic competence. *Nat Med* 2018, 24, 176-185.
- 480 38. Mack, S.C.; Pajtler, K.W.; Chavez, L.; Okonechnikov, K.; Bertrand, K.C.; Wang, X.; Erkek, S.; 481 Federation, A.; Song, A.; Lee, C., *et al.* Therapeutic targeting of ependymoma as informed by oncogenic enhancer profiling. *Nature* **2018**, *553*, 101-105.
- 483 39. Gryder, B.E.; Yohe, M.E.; Chou, H.C.; Zhang, X.; Marques, J.; Wachtel, M.; Schaefer, B.; Sen, N.; Song, Y.; Gualtieri, A., et al. PAX3-FOXO1 Establishes Myogenic Super Enhancers and Confers BET Bromodomain Vulnerability. Cancer Discov 2017, 7, 884-899.
- 486 40. Chen, H.S.; De Leo, A.; Wang, Z.; Kerekovic, A.; Hills, R.; Lieberman, P.M. BET-Inhibitors Disrupt Rad21-Dependent Conformational Control of KSHV Latency. *PLoS Pathog* **2017**, *13*.
- 488 41. Cheung, K.L.; Zhang, F.; Jaganathan, A.; Sharma, R.; Zhang, Q.; Konuma, T.; Shen, T.; Lee, J.Y.; Ren, C.; 489 Chen, C.H., *et al.* Distinct Roles of Brd2 and Brd4 in Potentiating the Transcriptional Program for Th17 Cell Differentiation. *Mol Cell* **2017**, *65*, 1068-1080.
- 491 42. Hsu, S.C.; Gilgenast, T.G.; Bartman, C.R.; Edwards, C.R.; Stonestrom, A.J.; Huang, P.; Emerson, D.J.; 492 Evans, P.; Werner, M.T.; Keller, C.A., *et al.* The BET Protein BRD2 Cooperates with CTCF to Enforce 493 Transcriptional and Architectural Boundaries. *Mol Cell* **2017**, *66*, 102-116.
- 494 43. Ko, J.Y.; Oh, S.; Yoo, K.H. Functional Enhancers As Master Regulators of Tissue-Specific Gene Regulation and Cancer Development. *Mol Cells* **2017**, *40*, 169-177.
- 496 44. Kaiser, V.B.; Semple, C.A. When TADs go bad: chromatin structure and nuclear organisation in human disease. *F1000Res* **1000**, 24.
- 498 45. Najafova, Z.; Tirado-Magallanes, R.; Subramaniam, M.; Hossan, T.; Schmidt, G.; Nagarajan, S.; 499 Baumgart, S.J.; Mishra, V.K.; Bedi, U.; Hesse, E., *et al.* BRD4 localization to lineage-specific enhancers is associated with a distinct transcription factor repertoire. *Nucleic Acids Res* **2017**, *45*, 127-141.
- 501 46. Cao, Q.; Anyansi, C.; Hu, X.; Xu, L.; Xiong, L.; Tang, W.; Mok, M.T.S.; Cheng, C.; Fan, X.; Gerstein, M., 612 61. Reconstruction of enhancer-target networks in 935 samples of human primary cells, tissues and 613 621 621. Cell lines. *Nat Genet* 2017, 49, 1428-1436.
- 504 47. Xie, W.; Nagarajan, S.; Baumgart, S.J.; Kosinsky, R.L.; Najafova, Z.; Kari, V.; Hennion, M.; Indenbirken, D.; Bonn, S.; Grundhoff, A., *et al.* RNF40 regulates gene expression in an epigenetic context-dependent manner. *Genome biology* **2017**, *18*, 32.
- 507 48. Fontanals-Cirera, B.; Hasson, D.; Vardabasso, C.; Di Micco, R.; Agrawal, P.; Chowdhury, A.; Gantz, M.; 508 de Pablos-Aragoneses, A.; Morgenstern, A.; Wu, P., et al. Harnessing BET Inhibitor Sensitivity Reveals 509 AMIGO2 as a Melanoma Survival Gene. *Molecular cell* 2017, 68, 731-744.e739.
- 510 49. Yokoyama, Y.; Zhu, H.; Lee, J.H.; Kossenkov, A.V.; Wu, S.Y.; Wickramasinghe, J.M.; Yin, X.; Palozola, 511 K.C.; Gardini, A.; Showe, L.C., *et al.* BET Inhibitors Suppress ALDH Activity by Targeting ALDH1A1 Super-Enhancer in Ovarian Cancer. *Cancer research* **2016**, *76*, 6320-6330.
- 513 50. Sengupta, D.; Kannan, A.; Kern, M.; Moreno, M.A.; Vural, E.; Stack, B., Jr.; Suen, J.Y.; Tackett, A.J.; Gao, L. Disruption of BRD4 at H3K27Ac-enriched enhancer region correlates with decreased c-Myc expression in Merkel cell carcinoma. *Epigenetics* **2015**, *10*, 460-466.
- 516 51. Hnisz, D.; Abraham, B.J.; Lee, T.I.; Lau, A.; Saint-Andre, V.; Sigova, A.A.; Hoke, H.A.; Young, R.A. Super-enhancers in the control of cell identity and disease. *Cell* **2013**, *155*, 934-947.

- 518 52. Hamdan, F.H.; Johnsen, S.A. Super enhancers new analyses and perspectives on the low hanging fruit. *Transcription* **2018**, *9*, 123-130.
- 520 53. Chapuy, B.; McKeown, M.R.; Lin, C.Y.; Monti, S.; Roemer, M.G.; Qi, J.; Rahl, P.B.; Sun, H.H.; Yeda, K.T.; Doench, J.G., *et al.* Discovery and characterization of super-enhancer-associated dependencies in diffuse large B cell lymphoma. *Cancer cell* **2013**, *24*, 777-790.
- 523 54. Gelato, K.A.; Schockel, L.; Klingbeil, O.; Ruckert, T.; Lesche, R.; Toedling, J.; Kalfon, E.; Heroult, M.; 524 Lejeune, P.; Monning, U., et al. Super-enhancers define a proliferative PGC-1alpha-expressing melanoma subgroup sensitive to BET inhibition. Oncogene 2018, 37, 512-521.
- 526 55. Huang, Y.; Jiang, X.; Liang, X.; Jiang, G. Molecular and cellular mechanisms of castration resistant prostate cancer. *Oncology letters* **2018**, *15*, 6063-6076.
- 528 56. Faivre, E.J.; Wilcox, D.; Lin, X.; Hessler, P.; Torrent, M.; He, W.; Uziel, T.; Albert, D.H.; McDaniel, K.; 529 Kati, W., *et al.* Exploitation of Castration-Resistant Prostate Cancer Transcription Factor Dependencies by the Novel BET Inhibitor ABBV-075. *Molecular cancer research: MCR* **2017**, *15*, 35-44.
- 531 57. Muhar, M.; Ebert, A.; Neumann, T.; Umkehrer, C.; Jude, J.; Wieshofer, C.; Rescheneder, P.; Lipp, J.J.; 532 Herzog, V.A.; Reichholf, B., et al. SLAM-seq defines direct gene-regulatory functions of the BRD4-MYC axis. Science (New York, N.Y.) 2018.
- 534 58. Taniguchi, Y. The Bromodomain and Extra-Terminal Domain (BET) Family: Functional Anatomy of BET Paralogous Proteins. *Int J Mol Sci* **2016**, *17*.
- 536 59. Andrieu, G.; Belkina, A.C.; Denis, G.V. Clinical trials for BET inhibitors run ahead of the science. *Drug Discov Today Technol* **2016**, *19*, 45-50.
- 538 60. Lamonica, J.M.; Deng, W.; Kadauke, S.; Campbell, A.E.; Gamsjaeger, R.; Wang, H.; Cheng, Y.; Billin, 539 A.N.; Hardison, R.C.; Mackay, J.P., *et al.* Bromodomain protein Brd3 associates with acetylated GATA1 to promote its chromatin occupancy at erythroid target genes. *Proc Natl Acad Sci U S A* **2011**, *108*, 2.
- 541 61. Denis, G.V.; McComb, M.E.; Faller, D.V.; Sinha, A.; Romesser, P.B.; Costello, C.E. Identification of transcription complexes that contain the double bromodomain protein Brd2 and chromatin remodeling machines. *J Proteome Res* 2006, *5*, 502-511.
- 544 62. Picaud, S.; Wells, C.; Felletar, I.; Brotherton, D.; Martin, S.; Savitsky, P.; Diez-Dacal, B.; Philpott, M.; 545 Bountra, C.; Lingard, H., et al. RVX-208, an inhibitor of BET transcriptional regulators with selectivity for the second bromodomain. Proceedings of the National Academy of Sciences of the United States of America 2013, 110, 19754-19759.
- 548 63. Mazur, P.K.; Herner, A.; Mello, S.S.; Wirth, M.; Hausmann, S.; Sanchez-Rivera, F.J.; Lofgren, S.M.; 549 Kuschma, T.; Hahn, S.A.; Vangala, D., *et al.* Combined inhibition of BET family proteins and histone deacetylases as a potential epigenetics-based therapy for pancreatic ductal adenocarcinoma. *Nat Med* 551 **2015**, *21*, 1163-1171.
- Allis, C.D.; Jenuwein, T. The molecular hallmarks of epigenetic control. *Nat Rev Genet* **2016**, *17*, 487-500.
- 553 65. Suraweera, A.; O'Byrne, K.J.; Richard, D.J. Combination Therapy With Histone Deacetylase Inhibitors (HDACi) for the Treatment of Cancer: Achieving the Full Therapeutic Potential of HDACi. Frontiers in oncology 2018, 8, 92.
- Barneda-Zahonero, B.; Parra, M. Histone deacetylases and cancer. *Molecular oncology* **2012**, *6*, 579-589.
- 557 67. Li, Y.; Seto, E. HDACs and HDAC Inhibitors in Cancer Development and Therapy. *Cold Spring Harbor perspectives in medicine* **2016**, *6*.
- Vancurova, I.; Uddin, M.M.; Zou, Y.; Vancura, A. Combination Therapies Targeting HDAC and IKK in Solid Tumors. *Trends in pharmacological sciences* **2018**, 39, 295-306.
- 561 69. Chen, Y.J.; Wang, W.H.; Wu, W.Y.; Hsu, C.C.; Wei, L.R.; Wang, S.F.; Hsu, Y.W.; Liaw, C.C.; Tsai, W.C. Novel histone deacetylase inhibitor AR-42 exhibits antitumor activity in pancreatic cancer cells by affecting multiple biochemical pathways. *PloS one* **2017**, *12*, e0183368.
- 564 70. He, M.; Qiao, Z.; Wang, Y.; Kuai, Q.; Li, C.; Wang, Y.; Jiang, X.; Wang, X.; Li, W.; He, M., et al. Chidamide Inhibits Aerobic Metabolism to Induce Pancreatic Cancer Cell Growth Arrest by Promoting Mcl-1 Degradation. *PloS one* **2016**, *11*, e0166896.

- 567 71. Dalla Pozza, E.; Manfredi, M.; Brandi, J.; Buzzi, A.; Conte, E.; Pacchiana, R.; Cecconi, D.; Marengo, E.; 568 Donadelli, M. Trichostatin A alters cytoskeleton and energy metabolism of pancreatic adenocarcinoma cells: An in depth proteomic study. *Journal of cellular biochemistry* **2018**, 119, 2696-2707.
- Wang, G.; He, J.; Zhao, J.; Yun, W.; Xie, C.; Taub, J.W.; Azmi, A.; Mohammad, R.M.; Dong, Y.; Kong, W., et al. Class I and class II histone deacetylases are potential therapeutic targets for treating pancreatic cancer. *PloS one* **2012**, *7*, e52095.
- 573 73. Cai, M.H.; Xu, X.G.; Yan, S.L.; Sun, Z.; Ying, Y.; Wang, B.K.; Tu, Y.X. Depletion of HDAC1, 7 and 8 by
 574 Histone Deacetylase Inhibition Confers Elimination of Pancreatic Cancer Stem Cells in Combination
 575 with Gemcitabine. *Scientific reports* **2018**, *8*, 1621.
- 576 74. Chan, E.; Chiorean, E.G.; O'Dwyer, P.J.; Gabrail, N.Y.; Alcindor, T.; Potvin, D.; Chao, R.; Hurwitz, H. Phase I/II study of mocetinostat in combination with gemcitabine for patients with advanced pancreatic cancer and other advanced solid tumors. *Cancer chemotherapy and pharmacology* **2018**, *81*, 355-364.
- 580 75. Lee, H.S.; Park, S.B.; Kim, S.A.; Kwon, S.K.; Cha, H.; Lee, D.Y.; Ro, S.; Cho, J.M.; Song, S.Y. A novel HDAC inhibitor, CG200745, inhibits pancreatic cancer cell growth and overcomes gemcitabine resistance. *Scientific reports* 2017, 7, 41615.
- 583 76. Samulitis, B.K.; Pond, K.W.; Pond, E.; Cress, A.E.; Patel, H.; Wisner, L.; Patel, C.; Dorr, R.T.; 584 Landowski, T.H. Gemcitabine resistant pancreatic cancer cell lines acquire an invasive phenotype with collateral hypersensitivity to histone deacetylase inhibitors. *Cancer biology & therapy* **2015**, *16*, 43-51.
- 586 77. Pan, Y.; Wang, L.; Kang, S.G.; Lu, Y.; Yang, Z.; Huynh, T.; Chen, C.; Zhou, R.; Guo, M.; Zhao, Y. Gd-Metallofullerenol Nanomaterial Suppresses Pancreatic Cancer Metastasis by Inhibiting the Interaction of Histone Deacetylase 1 and Metastasis-Associated Protein 1. *ACS nano* **2015**, *9*, 6826-6836.
- 589 78. Meidhof, S.; Brabletz, S.; Lehmann, W.; Preca, B.T.; Mock, K.; Ruh, M.; Schuler, J.; Berthold, M.; Weber, S.; Burk, U., et al. ZEB1-associated drug resistance in cancer cells is reversed by the class I HDAC inhibitor mocetinostat. *EMBO molecular medicine* **2015**, 7, 831-847.
- 592 79. Aghdassi, A.; Sendler, M.; Guenther, A.; Mayerle, J.; Behn, C.O.; Heidecke, C.D.; Friess, H.; Buchler, 593 M.; Evert, M.; Lerch, M.M., *et al.* Recruitment of histone deacetylases HDAC1 and HDAC2 by the transcriptional repressor ZEB1 downregulates E-cadherin expression in pancreatic cancer. *Gut* 2012, 61, 439-448.
- 596 80. Mishra, V.K.; Wegwitz, F.; Kosinsky, R.L.; Sen, M.; Baumgartner, R.; Wulff, T.; Siveke, J.T.; Schildhaus, 597 H.U.; Najafova, Z.; Kari, V., *et al.* Histone deacetylase class-I inhibition promotes epithelial gene expression in pancreatic cancer cells in a BRD4- and MYC-dependent manner. *Nucleic acids research* 2017, 45, 6334-6349.
- 600 81. Sanchez, G.J.; Richmond, P.A.; Bunker, E.N.; Karman, S.S.; Azofeifa, J.; Garnett, A.T.; Xu, Q.; Wheeler, G.E.; Toomey, C.M.; Zhang, Q., et al. Genome-wide dose-dependent inhibition of histone deacetylases studies reveal their roles in enhancer remodeling and suppression of oncogenic super-enhancers. Nucleic acids research 2018, 46, 1756-1776.
- 604 82. Huang, J.P.; Ling, K. EZH2 and histone deacetylase inhibitors induce apoptosis in triple negative breast cancer cells by differentially increasing H3 Lys(27) acetylation in the BIM gene promoter and enhancers. *Oncology letters* **2017**, *14*, 5735-5742.
- Greer, C.B.; Tanaka, Y.; Kim, Y.J.; Xie, P.; Zhang, M.Q.; Park, I.H.; Kim, T.H. Histone Deacetylases Positively Regulate Transcription through the Elongation Machinery. *Cell reports* **2015**, *13*, 1444-1455.
- 609 84. Creyghton, M.P.; Cheng, A.W.; Welstead, G.G.; Kooistra, T.; Carey, B.W.; Steine, E.J.; Hanna, J.; 610 Lodato, M.A.; Frampton, G.M.; Sharp, P.A., et al. Histone H3K27ac separates active from poised 611 enhancers and predicts developmental state. Proceedings of the National Academy of Sciences of the United 612 States of America 2010, 107, 21931-21936.
- Tao, R.; Zhang, B.; Li, Y.; King, J.L.; Tian, R.; Xia, S.; Schiavon, C.R.; Dong, J.T. HDAC-mediated deacetylation of KLF5 associates with its proteasomal degradation. *Biochemical and biophysical research communications* **2018**.
- Wang, L.; Beier, U.H.; Akimova, T.; Dahiya, S.; Han, R.; Samanta, A.; Levine, M.H.; Hancock, W.W. Histone/protein deacetylase inhibitor therapy for enhancement of Foxp3+ T-regulatory cell function

- posttransplantation. American journal of transplantation: official journal of the American Society of Transplantation and the American Society of Transplant Surgeons 2018.
- Kohler, S.; Cirillo, L.A. Stable chromatin binding prevents FoxA acetylation, preserving FoxA chromatin remodeling. *The Journal of biological chemistry* **2010**, 285, 464-472.
- 622 88. Diaferia, G.R.; Balestrieri, C.; Prosperini, E.; Nicoli, P.; Spaggiari, P.; Zerbi, A.; Natoli, G. Dissection of transcriptional and cis-regulatory control of differentiation in human pancreatic cancer. *Embo J* **2016**, 35, 595-617.
- 625 89. Sainsbury, S.; Bernecky, C.; Cramer, P. Structural basis of transcription initiation by RNA polymerase II. *Nature reviews. Molecular cell biology* **2015**, *16*, 129-143.
- 627 90. McCleland, M.L.; Mesh, K.; Lorenzana, E.; Chopra, V.S.; Segal, E.; Watanabe, C.; Haley, B.; Mayba, O.; Yaylaoglu, M.; Gnad, F., et al. CCAT1 is an enhancer-templated RNA that predicts BET sensitivity in colorectal cancer. *The Journal of clinical investigation* **2016**, 126, 639-652.
- An integrated encyclopedia of DNA elements in the human genome. *Nature* **2012**, 489, 57-74.
- Gotze, S.; Coersmeyer, M.; Muller, O.; Sievers, S. Histone deacetylase inhibitors induce attenuation of Wnt signaling and TCF7L2 depletion in colorectal carcinoma cells. *International journal of oncology* **2014**, 45, 1715-1723.
- 634 93. Yu, Q.; Zhou, X.; Xia, Q.; Shen, J.; Yan, J.; Zhu, J.; Li, X.; Shu, M. Long non-coding RNA CCAT1 that can be activated by c-Myc promotes pancreatic cancer cell proliferation and migration. *American journal of translational research* **2016**, *8*, 5444-5454.
- Taberlay, P.C.; Achinger-Kawecka, J.; Lun, A.T.; Buske, F.A.; Sabir, K.; Gould, C.M.; Zotenko, E.; Bert, S.A.; Giles, K.A.; Bauer, D.C., *et al.* Three-dimensional disorganization of the cancer genome occurs coincident with long-range genetic and epigenetic alterations. *Genome research* **2016**, *26*, 719-731.
- Shukla, S.; Cyrta, J.; Murphy, D.A.; Walczak, E.G.; Ran, L.; Agrawal, P.; Xie, Y.; Chen, Y.; Wang, S.;
 Zhan, Y., et al. Aberrant Activation of a Gastrointestinal Transcriptional Circuit in Prostate Cancer
 Mediates Castration Resistance. Cancer cell 2017, 32, 792-806.e797.
- Walsh, A.L.; Tuzova, A.V.; Bolton, E.M.; Lynch, T.H.; Perry, A.S. Long noncoding RNAs and prostate carcinogenesis: the missing 'linc'? *Trends in molecular medicine* **2014**, *20*, 428-436.
- 545 Seng, C.; Yu, X.; Lai, J.; Yang, L.; Chen, S.; Li, Y. Overexpression of the long non-coding RNA PVT1 is correlated with leukemic cell proliferation in acute promyelocytic leukemia. *J Hematol Oncol* **2015**, *8*, 015-0223.
- 648 98. Rathert, P.; Roth, M.; Neumann, T.; Muerdter, F.; Roe, J.S.; Muhar, M.; Deswal, S.; Cerny-Reiterer, S.; 649 Peter, B.; Jude, J., *et al.* Transcriptional plasticity promotes primary and acquired resistance to BET inhibition. *Nature* **2015**, 525, 543-547.
- Ganesan, A. Epigenetics: the first 25 centuries. *Philosophical transactions of the Royal Society of London.*Series B, Biological sciences **2018**, 373.
- 653 100. Peng, X.L.; So, K.K.; He, L.; Zhao, Y.; Zhou, J.; Li, Y.; Yao, M.; Xu, B.; Zhang, S.; Yao, H., et al. MyoD-654 and FoxO3-mediated hotspot interaction orchestrates super-enhancer activity during myogenic 655 differentiation. *Nucleic acids research* **2017**, 45, 8785-8805.
- Pott, S.; Lieb, J.D. What are super-enhancers? *Nature genetics* **2015**, 47, 8-12.
- Hay, D.; Hughes, J.R.; Babbs, C.; Davies, J.O.J.; Graham, B.J.; Hanssen, L.; Kassouf, M.T.; Marieke Oudelaar, A.M.; Sharpe, J.A.; Suciu, M.C., *et al.* Genetic dissection of the alpha-globin super-enhancer in vivo. *Nat Genet* **2016**, *48*, 895-903.
- Shin, H.Y.; Willi, M.; HyunYoo, K.; Zeng, X.; Wang, C.; Metser, G.; Hennighausen, L. Hierarchy within the mammary STAT5-driven Wap super-enhancer. *Nat Genet* **2016**, *48*, 904-911.
- Zhang, Q.; Lou, Y.; Zhang, J.; Fu, Q.; Wei, T.; Sun, X.; Chen, Q.; Yang, J.; Bai, X.; Liang, T.
 Hypoxia-inducible factor-2alpha promotes tumor progression and has crosstalk with Wnt/beta-catenin signaling in pancreatic cancer. *Molecular cancer* 2017, 16, 119.
- Zhong, Y.; Macgregor-Das, A.; Saunders, T.; Whittle, M.C.; Makohon-Moore, A.; Kohutek, Z.A.; Foling, J.; Herbst, B.T.; Javier, B.M.; Cope, L., et al. Mutant p53 Together with TGFbeta Signaling

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Peer-reviewed version available at Epigenomes 2018, 2, 8; doi:10.3390/epigenomes2020008

667 668		Influence Organ-Specific Hematogenous Colonization Patterns of Pancreatic Cancer. <i>Clinical cancer research: an official journal of the American Association for Cancer Research</i> 2017 , 23, 1607-1620.
669 670 671 672	106.	Nomura, A.; Majumder, K.; Giri, B.; Dauer, P.; Dudeja, V.; Roy, S.; Banerjee, S.; Saluja, A.K. Inhibition of NF-kappa B pathway leads to deregulation of epithelial-mesenchymal transition and neural invasion in pancreatic cancer. <i>Laboratory investigation; a journal of technical methods and pathology</i> 2016 , <i>96</i> , 1268-1278.
673 674 675 676	107.	Nhili, R.; Peixoto, P.; Depauw, S.; Flajollet, S.; Dezitter, X.; Munde, M.M.; Ismail, M.A.; Kumar, A.; Farahat, A.A.; Stephens, C.E., <i>et al.</i> Targeting the DNA-binding activity of the human ERG transcription factor using new heterocyclic dithiophene diamidines. <i>Nucleic acids research</i> 2013 , <i>41</i> , 125-138.
677 678 679	108.	Huang, M.; Zeng, S.; Zou, Y.; Shi, M.; Qiu, Q.; Xiao, Y.; Chen, G.; Yang, X.; Liang, L.; Xu, H. The suppression of bromodomain and extra-terminal domain inhibits vascular inflammation by blocking NF-kappaB and MAPK activation. <i>British journal of pharmacology</i> 2017 , <i>174</i> , 101-115.
680 681 682	109.	Hensel, T.; Giorgi, C.; Schmidt, O.; Calzada-Wack, J.; Neff, F.; Buch, T.; Niggli, F.K.; Schafer, B.W.; Burdach, S.; Richter, G.H. Targeting the EWS-ETS transcriptional program by BET bromodomain inhibition in Ewing sarcoma. <i>Oncotarget</i> 2016 , <i>7</i> , 1451-1463.
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