

1 Article

2 Application of a novel S3 nanowire gas sensor device 3 in parallel with GC-MS for the identification of rind 4 percentage of grated Parmigiano Reggiano

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15 **Abstract:** Parmigiano Reggiano cheese is one of the most appreciated and consumed food
16 worldwide, especially in Italy, for its high content of nutrients and for its taste. However, these
17 characteristics make this product subject to counterfeiting in different forms. In this study, a novel
18 method based on an electronic nose has been developed in order to investigate the potentiality of
19 this tool to distinguish rind percentage in grated Parmigiano Reggiano packages that should be
20 lower than 18%. Different samples in terms of percentage, seasoning and rind working process were
21 considered to tackle the problem at 360°. In parallel, GC-MS technique was used to give a name to
22 the compounds that characterize Parmigiano and to relate them with sensors responses. Data
23 analysis consisted of two stages: multivariate analysis (PLS) and classification made in a hierarchical
24 way with PLS-DA and ANNs. Results are promising in terms of correct classification of the samples.
25 The classification rate is higher for ANNs than PLS-DA, reaching also 100% values.

26 **Keywords:** electronic nose, nanowire gas sensors, food quality control, Parmigiano Reggiano,
27 multivariate data analysis, artificial neural network
28

29 1. Introduction

30 Parmigiano Reggiano (PR) cheese is one of the most typical Italian food and one of the oldest
31 traditional cheeses produced in Europe. It is also the most important Protected Designation of Origin
32 (PDO) Italian cheese in terms of commercial importance [1]. Its production is regulated by the
33 Parmigiano Reggiano Cheese Consortium (CFPR). According to European Regulation 510/2006, this
34 designation can be exclusively assigned to the cheese made following a traditional established
35 production technology in a restricted area of Italy (provinces of Parma, Reggio Emilia, Modena,
36 Mantova and Bologna) from milk produced in the same area [2].

37 PR can be found on the market in different forms. It can be portioned or grated and cannot be
38 subjected to any treatment like lyophilization, drying and freezing [3]. All the procedures, that must
39 be followed to obtain the original PR, make this cheese a high-value product. That leads to a final
40 product that has various nutritional properties: its dry weight is mostly composed of proteins and
41 lipids, is lactose- and galactose-free and rich in organic acids, such as lactic acid, acetic acid, propionic
42 and butyric acids [4]; the semi-fat composition due to natural creaming of skimmed unpasteurized
43 milk [5] is produced by cattle that consumes only locally grown forage because supply of silage and
44 fermented feeds is not permitted [6].

45 For these reasons PR has a high cost, if compared to similar hard cheeses. This encourages the
46 appearance on the market of counterfeited products that bear PR brand at a lower price. The rate of
47 fraud is estimated between 20% and 40%, the latter predominantly in the grated form. [7].

48 As established in the procedural guideline, grated PR cheese must follow some technical and
49 technological parameters: moisture no less than 25% and no more than 35%, at least 12 months of
50 ripening, rind not over 18%, typical amino-acid composition of the cheese, absence of additives, not
51 crumbly in aspect and with homogeneous particles with a diameter inferior to 0.5 mm and not
52 exceeding 25% [8].

53 In order to distinguish if a PR cheese package conforms to the rules, the aromatic profile of grated
54 PR can be analyzed thanks to the volatile organic compounds (VOCs). VOCs of various dairy
55 products have received a great deal of attention in the last years. Till now, about 600 volatile
56 compounds have been identified for cheese [9]. However, only a small part of these compounds is
57 responsible for cheese flavor [10]. Cheese aroma is considered the result of the equilibrium between
58 various VOCs that separately do not reflect the overall odor [11]. Hydrocarbons, alcohols, aldehydes,
59 ketones, esters and lactones were the major classes of compounds found in the neutral fraction of
60 cheese [12].

61 In this work, an electronic nose has been used in order to analyze rind percentage in grated PR
62 cheese through emitted VOCs. In the last years, this kind of devices has received numerous attentions
63 for its potentialities; it has been applied in various fields, as environment [13-14], health [15-16] and
64 food with excellent results. Regarding food applications, some examples of electronic nose
65 applications are the detection of microorganisms in tomato sauce [17] and of different molds in coffee
66 [18], the determination of shelf life of milk [19], the finding of additives in fruit juices [20]. These few
67 examples show how e-noses have the potentiality to be used in different ways to assess food quality
68 and identity.

69 Placed side by side with e-nose analysis, Gas Chromatography coupled with Solid Phase Micro
70 Extraction (SPME) was used. SPME has received much attention in the literature to find VOCs that
71 characterize food matrices. Many foods have been studied, including dairy products, such as milk
72 [21], butter [22] and cheese [23-24].

73 The aim of the work is to recognize rind percentages of the sample under analysis with an
74 innovative and rapid methodology in order to identify eventual frauds and therefore have an
75 affordable and reliable instrument to reduce them, making the most of the possible differences
76 between the products such as VOCs presence and amount.

77 2. Materials and Methods

78 2.1. Samples preparation and experimental design

79 Analyzed samples were packaged under vacuum at the headquarters of CFPR. They came from
80 two different ripening stages: 12 and 24 months. For each of these, five different combinations of
81 pulp-rind were prepared (expressed in rind percentage): 0%, 18%, 26%, 45% and 100%. In addition,
82 two kind of rind working processes were considered: washed-rind (WR) and scraped-rind (SR). The
83 only exceptions are represented by 0% samples, for which only the 24 month ripening was taking
84 into account, and 100% samples, for which there is one for WR and one for SR that correspond to 24
85 month and 12 month seasoning respectively. For each sample, 14 several replicas were arranged for
86 a total of 210 (14 replicas x 15 samples).

87 Samples were stored at 4°C until the moment they were prepared for the analysis. The amount
88 of 2 g of grated cheese were positioned in 20 mL glass headspace vials and sealed with a metal cap
89 with a PTFE-silicon membrane, crimped with an aluminum crimp.

91 2.2 GC-MS Analysis

92
93 The Gas Chromatograph (GC) used during the analyses was a Shimadzu GC2010 PLUS (Kyoto,
94 KYT, Japan), equipped with a Shimadzu single quadrupole Mass Spectrometer (MS) MS-QP2010 Ultra

95 (Kyoto, KYT, Japan) and with an autosampler HT280T (HTA s.r.l., Brescia, Italy). The GC-MS analysis
96 was coupled with the Solid-Phase Micro Extraction (SPME) method in order to find the most
97 significant VOCs that allow to recognize the different kinds of cheeses.

98 The fiber used for the adsorption of volatiles was a DVB/CAR/PDMS – 50/30 μm (Supelco Co.
99 Bellefonte, PA, USA). The fiber was exposed to the headspace of the vials after heating the samples
100 in the HT280T oven thermostatically regulated at 50°C for 15 min with the aim to create the headspace
101 equilibrium. The length of the fiber in the headspace was kept constant. Desorption of volatiles took
102 place in the injector of the GC-MS for 6 min at 250°C.

103 The gas chromatograph was operated in the direct mode throughout the run with the mass
104 spectrometer in electron ionization (EI) mode (70 eV). GC separation was performed on a MEGA-
105 WAX capillary column (30m \times 0.25mm \times 0.25 μm , Agilent Technologies, Santa Clara, CA, USA).
106 Ultrapure helium (99.99%) was used as the carrier gas at the constant flow rate of 1.3 mL/min. The
107 following GC oven temperature programming was applied: at the beginning, the column was held
108 at 40 °C for 8 min, and then it was programmed from 40 to 190 °C at 4 °C/min; then, the temperature
109 was maintained at 190°C for 5 min. Next, the temperature was raised from 190°C to 210°C with a rate
110 of 5°C/min; finally, 210°C were maintained for 5 min.

111 The GC-MS interface was kept at 200 °C. The mass spectra were collected over the range of 45 to
112 500 m/z in the Total Ion Current (TIC) mode, with scan intervals being 0.3 s. The identification of
113 the volatile compounds was carried out using the NIST11 and the FFNSC2 libraries of mass spectra.

114 Each sample was analyzed one time.

115 2.3 S3 Analysis

116 The innovative Small Sensors System S3 device used in the present work has been completely
117 designed and constructed at SENSOR Laboratory (University of Brescia, Italy) in collaboration with
118 NASYS S.r.l., spin-off of the University of Brescia. The tool comprises a metal oxide (MOX) gas
119 sensors array, flow sensors, temperature and humidity sensors, fluidodynamic system, electronic
120 control system. In particular, the sensor used in this study are 8 MOX gas sensors. Three of them are
121 nanowires of MOX as presented in [25-26]. Two of them are tin oxides nanowires sensors, both
122 grown by means of Vapor Liquid Solid technique [27] using a gold catalyst on the alumina substrate,
123 but one has also been functionalised with gold clusters; the third sensor has an active layer of copper
124 oxide nanowires. The working temperature is 350°C, 350°C and 400°C respectively. Other three
125 sensors are prepared with Rheotaxial Growth and Thermal Oxidation (RGTO) thin film technology;
126 one is tin oxide functionalised with gold clusters (working at 400°C), while the other two are pure tin
127 oxide (working at 300°C and at 400°C respectively).

128 Finally, the last two are commercial MOX sensors produced by Figaro Engineering Inc. (Osaka,
129 Japan). In particular, they are the TGS2611 and TGS2602, that are sensitive to natural gases and to
130 odorous gases like ammonia, respectively, according to the datasheet of the company. Commercial
131 sensors have been mounted on our e-nose in order to evaluate the performances of nanowire sensors.
132 Details of S3 sensors made at SENSOR Laboratory are summarised in Table 1.

133 The MOX nanowires are gas sensors with a high sensitivity to a broad range of chemicals; they
134 exhibit physical properties that are significantly different from their polycrystalline counterpart. The
135 nanowires have a high degree of crystallinity, atomically sharp terminations and an extraordinary
136 length-to-width ratio, resulting in enhanced sensing capability as well as long-term material stability
137 for prolonged operation. In addition, the three-dimensional network formed by the nanowires
138 increases the adsorption surface and the catalytic activity, improving the response and decreasing
139 the instrument threshold [28].

140 S3 analyses the head space (HS), i.e. the volatile fraction of the samples formed when the
141 equilibrium of the solid-liquid phase and the vapor phase of all volatile compounds is reached. The
142 creation of the HS depends on the test substance (vapor pressure) and the conditioning temperature
143 of the sample. The compounds are extracted at the equilibrium point between the solid phase and
144 the vapor in a dynamic head space. This characteristic allows a non destructive samples analyses. In
145 this case, the sensor base line is obtained with the air of the surrounding environment, no gas cylinder
146 of chromatographic air is required (an essential feature that makes it a portable instrument). The

147 environmental air was filtered using a small metal cylinder (21.5 cm in length, 5 cm of diameter) filled
148 with activated carbons.

149 **Table 1.** Type, composition, morphology and operating temperature of S3 sensors made at SENSOR
150 Laboratory.

Type	Composition	Morphology	Operating temperature (°C)
SnO ₂ Au	SnO ₂ functionalised with Au clusters	RGTO	400°C
SnO ₂	SnO ₂	RGTO	300°C
SnO ₂	SnO ₂	RGTO	400°C
SnO ₂ Au+Au	SnO ₂ grown with Au and functionalised with gold clusters	Nanowire	350°C
SnO ₂ Au	SnO ₂ grown with Au	Nanowire	350°C
CuO	CuO	Nanowire	400°C

151
152 The volatile fraction is then aspirated and transported to the sensor chamber to be analyzed. In
153 order to avoid any influence of the surrounding environment to the sensor response, the chamber
154 has been thermostated and isolated. To prevent the absorption of volatile substances that could be
155 released during subsequent analysis, the chamber and the connection between the elements tires are
156 made using steel. The air is flown into the sensor chamber using a pump through a needle valve. That
157 is used to adjust the total airflow, which is measured by a flowmeter downstream of the pump.

158 The instrument was also provided with the auto-sampler head space system HT2010H,
159 supporting a 42 loading sites carousel and a shaking oven to equilibrate the sample head space. The
160 vials were placed in a randomized mode into the carousel. Each vial was incubated at 50°C for 5
161 minutes into the auto-sampler oven, by shaking it for 1 minute during the incubation. The sample
162 head space was then extracted from the vial in dynamic head space path and released into the carried
163 flow (80 sccm).

164 The analysis timeline can be divided in three different steps, for a duration of 420 seconds (7
165 minutes) per sample, that are preceded by a step of warm-up that allows the achievement of the base
166 line for the entire system:

- 167 • *injection*: the sample HS is flowed in the sensor chamber for 60 seconds (it is the actual
168 analysis time); then, for 30 seconds environmental air flows through the same tube to clean it from
169 any residual VOCs;
- 170 • *restore*: it starts when *injection* period is finished and, in this step, filtered air is flowed into
171 the sensors camber. In this time (330 seconds) the sensors restore the original condition of the base
172 line.

173 Thanks to the processor integrated in the S3 instrument, the frequency at which the equipment
174 works is equal to 1 Hz.

175 2.4 Data Analysis

176 Data analysis was performed using MATLAB® R2015a software (MathWorks, USA). First of all,

177 sensors responses in terms of resistance (Ω) were normalized compared to the first value of the
178 acquisition (R_0). As a feature for all the sensors, the difference between the first value and the
179 minimum value during the analysis time was calculated. Hence, the dataset was composed by $\Delta R/R_0$
180 parameters.

181 In the second step, the normal distribution of the variables was checked using the Jarque-Bera
182 (JB) test with a significance level chosen equal to 0.05. This test is a goodness-of-fit test of whether
183 sample data have the skewness and kurtosis matching a normal distribution. The null hypothesis is
184 a joint hypothesis of both the skewness and the excess kurtosis being zero.

185 Based on the test result, Partial Least Squares (PLS) method was used both to view how the
186 groups of samples were represented thanks to the considered volatile compounds and to sensors
187 responses, respectively, and to build the model that was used to classify the samples themselves. PLS
188 is a statistical method that combines features from Principal Component Analysis (PCA) and multiple
189 regression (MR). PCA is a statistical method that uses an orthogonal transformation in order to pass
190 from the variable space X to a space of uncorrelated variables called principal components (PC). The
191 aim is to reduce the number of variables to obtain a better representation of data. In addition to this,
192 in PLS also the response matrix Y is decomposed and the principal component of X are rotated in the
193 direction of maximum correlation with the principal components of Y . The new calculated variables
194 are called latent variables (LV). Another advantage of using PLS instead of PCA is that it is not
195 required that variables have a normal distribution.

196 Finally, classification was performed comparing two different classifiers: Partial Least Squares
197 Discriminant Analysis (PLS-DA) and Artificial Neural Networks (ANNs). PLS-DA was successfully
198 applied in different fields where products had to be recognized according to their place of origin or
199 the presence of contamination, as in milk [29], honey [30], wine [31] and cheese [32]. ANNs are
200 complex structures that try to mimic what human brain does. They are formed by elemental units
201 called neurons that works like real neurons: once information arrives, they elaborate it and give an
202 output. Each neuron is characterized by an activation function and coefficients of connectivity called
203 weights. Overall structure is mainly composed by an input layer, hidden layers and an output layer
204 [33]. ANNs can be used to resolve regression and classification problems or function approximation.
205 They found a lot of space in food applications in order to analyze data collected with electronic noses
206 [34-37]. In this work, a feed-forward ANN trained with Levenberg-Marquardt algorithm is used.

207 For PLS, dataset was splitted in two parts, training set and test set, using Venetian Blinds (VB)
208 as cross-validation procedure. This method divides the whole dataset in j cross-validation groups; in
209 each one, one sample is put in the test set and the others in the training set on the first step.
210 Subsequently, in every group the sample after the previous one is taken into the test set and the others
211 in the training set, and so on. In this work, the number of cross-validation groups was chosen equal
212 to 10.

213 For the classification with PLS-DA, a toolbox made for MATLAB® and released by Milano
214 Chemometrics was used [38]. Instead, ANNs were created using the function *mntool* of the same
215 software. This tool allows to do a random split of dataset in test and training set by default.

216 3. Results and Discussion

217 3.1 GC-MS Analysis results

218 From the comparison between samples chromatograms, substantial differences were found. The
219 main difference between 12 months and 24 months ripened grated PR lies in the amount of fatty acids
220 that characterize this product. They are acetic acid, butanoic acid, hexanoic acid, octanoic acid, n-
221 decanoic acid and their presence is much greater in 24 months PR. This result is widely confirmed
222 from literature. Indeed, it is well known that these fatty acids are the results of fermentation processes
223 especially in butter and seasoned cheese. Some studies revealed that the amount of acetic acid and
224 butanoic acid doubles up in the period from 12 and 24 months [39-40].

225 Differences were also found comparing samples with different percentage of rind and same rind
226 working process; the same trend is valid both for 12 months and 24 months ripened PR. It turned out

227 that increasing the quantity of rind, the presence of three compounds increases, too. Besides butanoic
 228 and hexanoic acid, 2-nonanone has the same behavior. It is a member of the class of methyl ketones
 229 and it can be found in several foods, among which milk and cheese [41]. It is produced by oxidative
 230 degradation of fatty acids [42]. These results suggest that both the fermentation and the degradation
 231 happen more near the rind than in the central part of the cheese.
 232

233 3.2 S3 Analysis results

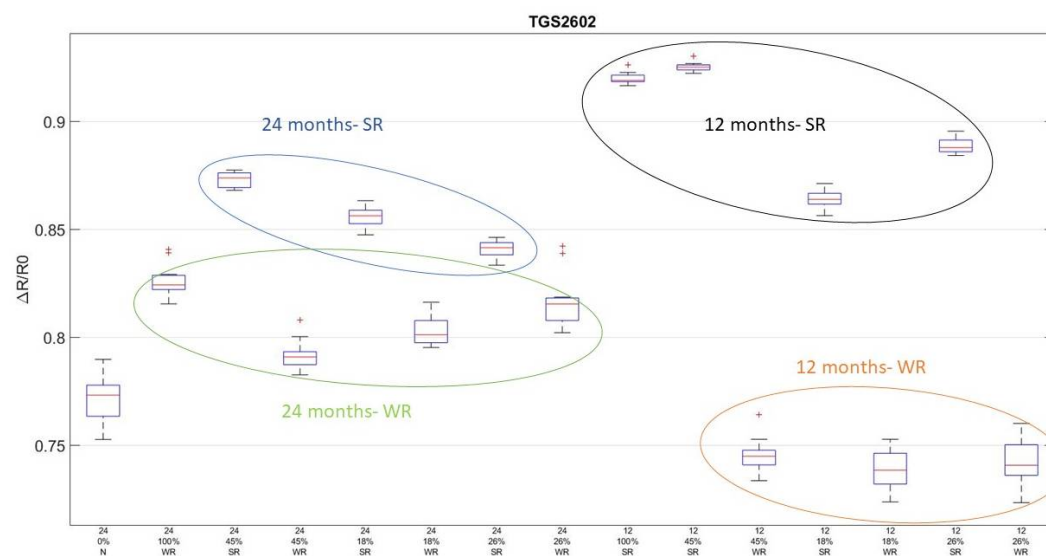
234 Once data were acquired, at first sensors responses were checked. Since the first measures of
 235 each session were very different from the others, they were discarded. Consequently, there is a
 236 different number of replicas for each sample. Most likely, experimental conditions of first acquisitions
 237 were not the same of the following measures in terms of temperature of the autosampler oven where
 238 vials were put in, as explained in section 2.3 S3 Analysis. In Table 2, a detailed description of number
 239 of samples that were considered for the following analysis is shown.

240

241 **Table 2.** Considered samples divided for ripening stage, rind percentage and rind working
 242 processes.

	0%	18%		26%		45%		100%	
12 month	-	11	12	13	11	12	14	-	14
24 month	12	14	14	13	13	11	13	13	-
		WR	SR	WR	SR	WR	SR	WR	SR

243 In Figure 1, boxplots of TGS2602 response that include $\Delta R/R_0$ for each sample are shown. This
 244 sensor represents the general trend that can be observed for all the sensors. Obviously, since different
 245 sensing materials are used, there are differences in terms of the highlighted groups overlapping. On
 246 the left part of the figure, there are 24-months seasoned samples: in the upper part grated cheese with
 247 SR while in the lower PR with WR. On the right part, there are 12-months ripened samples and they
 248 follow the same trend. The first boxplot is relative to samples of 0% rind; its $\Delta R/R_0$ is different respect
 249 to all the other group, but it is more similar to WR grated PR both seasoning stage. This result reflects
 250 the fact that they are characterized by a bigger amount of humidity.

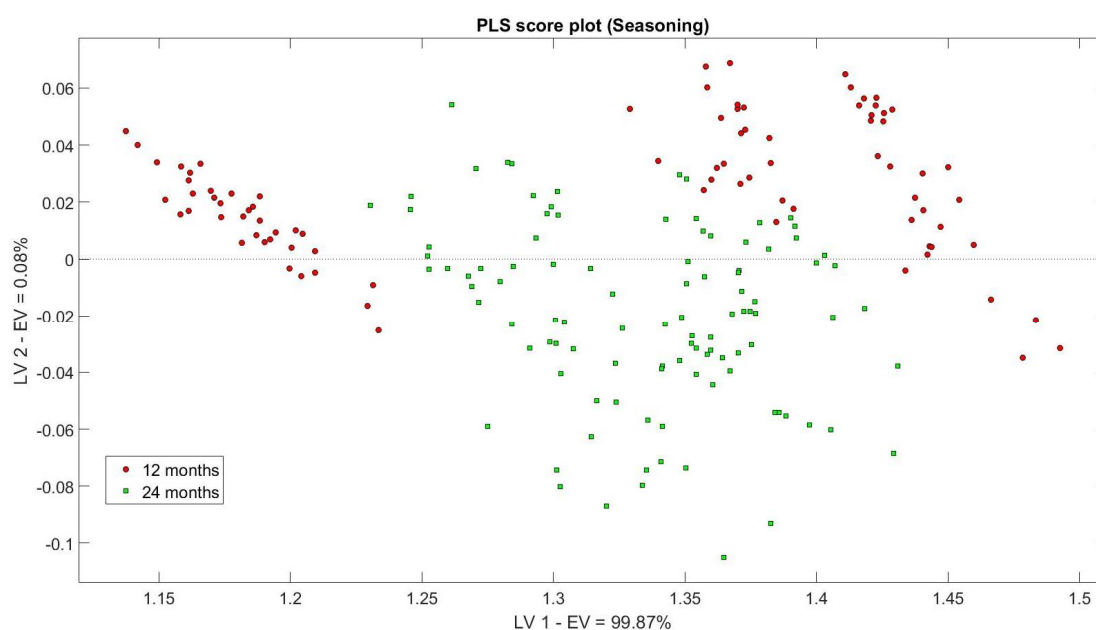


251

252 **Figure 1.** Boxplots of TGS2602 feature $\Delta R/R_0$. Four groups are highlighted: in blue 24 months-SR, in
 253 green 24 months-WR, in black 12 months-SR and in orange 12 months-WR.

254 After checked general sensors performances, JB-test was applied to the dataset. It results that
 255 only 4 of the eight parameters followed a normal distribution ($p < 0.05$); they correspond to the features
 256 extracted by the two tin oxide nanowires and RGTO sensors. That is the main reason for which PLS
 257 has been chosen. In Figure 2, PLS score plot was made considering the first two LV, for a total
 258 explained variance equal to 99.95% (99.87% for LV1 and 0.08% for LV2). In the plot measures are
 259 divided by seasoning degree. It can be observed that 24 months class is in the central part of the
 260 graph, while the other one is divided in the left and right part.

261 For this reason, classification techniques are used in a hiercachical way. In addition, another
 262 motive for this choice is to simplify classification models since this is a 15-class problem. Hence in the
 263 first step, classifiers were used to distinguish the seasoning degree; in a second step, for each ripening
 264 state the different working processes were discriminated; finally, ring percentage has been taken into
 265 account. In Figure 3, a scheme of the steps is shown.

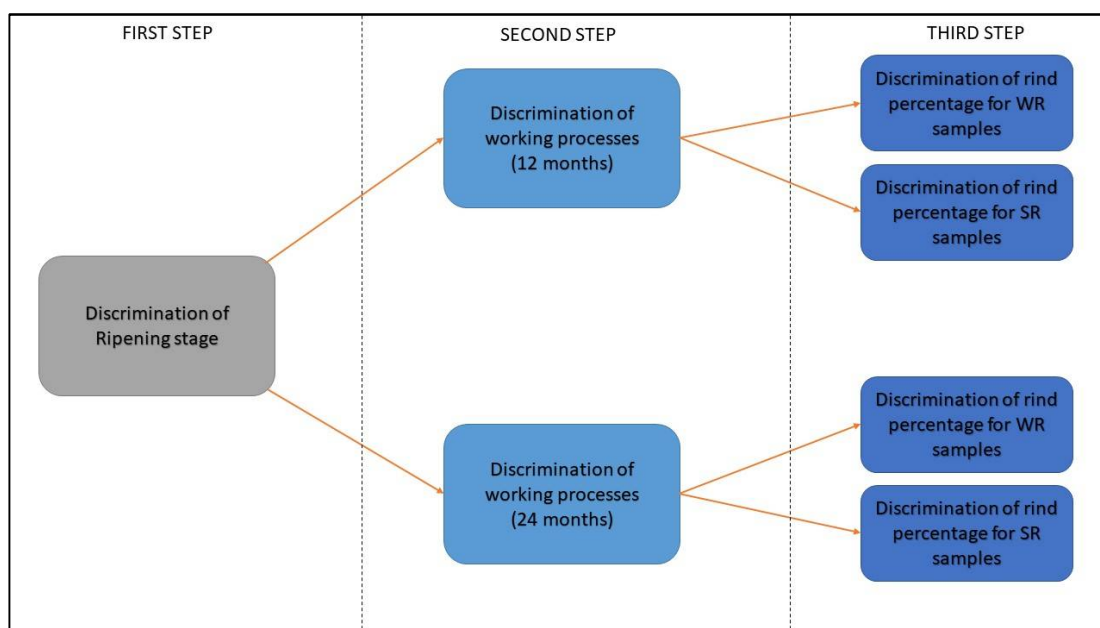


266

267

268

Figure 2. PLS score plot for all the measures divided for seasoning degree: in red circle 12 months, in green square 24 months. Total explained variance equal to 99.95% in first two LV.



269

270

Figure 3. Step by step scheme for classification analysis.

271 Regarding ANNs structures, three different ones were considered, one for each step. In the first
 272 case, a two layers architecture with 3 neurons in the input layer and 1 in the output layer was
 273 considered. For the second stage, the same number of layers was used, but in the first one two
 274 neurons were put. Finally, the third ANN had the same structure of the previous ones, but with 6
 275 neurons in the input layer. For all the neurons, hyperbolic tangent sigmoid transfer function was
 276 chosen.

277 In Table 3, overall classification rate of the two classifiers is put side by side. In general, ANN
 278 classification rates are better than those of PLS-DA. Indeed, ANN is able to recognize correctly all the
 279 samples based on seasoning and rind working processes. Although PLS-DA performances are lower,
 280 it can reach good classification rates. The distinction between rind percentage shows that both
 281 classifier can classify samples with SR better than those with WR. A possible interpretation of this
 282 results could be the different amount of humidity: WR samples have a higher content of humidity
 283 because of water treatment and the adsorption sites this could cause the occupation of by water
 284 molecules instead of the ones that characterize the volatile fingerprint of the samples.

285 **Table 3.** Classification rates of PLS-DA and ANNs divided per steps.

	First step Ripening stage	Second step Working processes	Third step Rind percentage
PLD-DA	94.7%	12 months:100%	WR: 61.1%
			SR: 90.2%
		24 months:79%	WR:90.2%
			SR: 95%
ANN	100%	12 months:100%	WR: 63.8%
			SR: 96.1%
		24 months:100%	WR: 58.8%
			SR: 100%

286

287 4. Conclusions

288 This study was aimed to verify the possible discrimination between grated PR with different
 289 rind percentage with an electronic nose, taking into account other two variables: seasoning degree
 290 and working processes of rind. In parallel, a consolidated technique, *i.e.* SPME GC-MS, has been used
 291 to understand which VOCs characterized analyzed samples. This combined analysis has produced
 292 promising results that pave the way to assess cheeses quality and avoid frauds.

293 First of all, with GC-MS, the VOCs that characterize grated cheeses have been individuated. The
 294 results regarding PR are compliant with those found in literature. Indeed, fatty acids that describe
 295 aroma and taste profile of PR have been found in greater quantity for 24 months seasoned samples
 296 as compared to 12 months ones. In addition, VOCs, whose amount is bigger in rind compared to
 297 pulp, was found and they are acquiescent with chemical reactions that take places in this product.

298 The multivariate statistical analysis made with PLS indicated how to proceed during the
 299 classification stage. A hierarchical approach was used, both for PLS-DA and ANNs. ANNs
 300 classification rates are the highest, suggesting that in future they could be improved to increase their
 301 performances. These first results are encouraging and further research is in progress in order to add
 302 more samples and to have more statistical significance of the achieved results.

303

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 305 samples.

306 The authors declare no conflict of interest.

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