

1 Review

2 Applications of metals for bone regeneration

3 Kristina Glenske ^{1,†}, Phil Donkiewicz ^{2,†}, Alexander Köwitsch ², Nada Milosevic-Oljaca ¹,
4 Sven Rofall ², Jörg Franke ³, Ole Jung ⁴, Reinhard Schnettler ⁵, Sabine Wenisch ^{1,†,*} and
5 Mike Barbeck ^{2,4,†,*}

6 1 Clinic of Small Animals, c/o Institute of Veterinary Anatomy, Histology and Embryology, Justus
7 Liebig University of Giessen, Giessen, Germany

8 2 botiss biomaterials, Berlin, Germany

9 3 Clinic for Trauma Surgery and Orthopedics, Elbe Kliniken Stade-Buxtehude, Stade, Germany

10 4 Department of Oral and Maxillofacial Surgery, University Hospital Hamburg-
11 Eppendorf, Hamburg, Germany

12 5 University Medical Center, Justus Liebig University of Giessen, Giessen, Germany

13

14 + Authors contributed equally

15 * Correspondence:

16 (1) Dr. Mike Barbeck: mike.barbeck@icloud.com; Tel.: +49-176-81022467

17 (2) Prof. Dr. Sabine Wenisch: Sabine.Wenisch@vetmed.uni-giessen.de; Tel.: +49-641-9938111

18

19 **Abstract:** The regeneration of bone tissue is a main purpose of most therapies in dental medicine.
20 For bone regeneration, calcium phosphate (CaP)-based substitute materials based on natural (allo-
21 and xenografts) and synthetic origins (alloplastic materials) are applied for guiding the regeneration
22 processes. The optimal bone substitute has to act as a substrate for bone ingrowth into a defect,
23 while it should be resorbed even in the time frame needed for complete regeneration up to the
24 condition of *restitution ad integrum*. In this context, the modes of action of CaP-based substitute
25 materials have been frequently investigated and it has been shown that such materials strongly
26 influence regenerative processes such as osteoblast growth or differentiation and also on osteoclastic
27 resorption due to different physicochemical properties of the materials. However, the material
28 characteristics needed for the required ratio between the formation of new bone tissue and material
29 degradation has not been found until now. The addition of different substances such as collagen or
30 growth factors and also of different cell types have already been tested but did not allow for
31 sufficient or prompt application. Moreover, metals or metal ions are differently used as basis or as
32 supplement for different materials in the field of bone regeneration. Moreover, it has already been
33 shown that different metal ions are integral components of bone tissue playing functional roles in
34 the physiological cellular environment as well as in the course of bone healing. The present review
35 focuses on frequently used metals as integral parts of materials designated for bone regeneration
36 with the aim to give an overview of currently existing knowledge about the effects of metals in the
37 field of bone regeneration.

38 **Keywords:** metals, dental regeneration, bioactivity, tissue regeneration, bone

39

40 1. Introduction

41 The regeneration of bone is of special interest, most notably, in dental medicine. For the
42 regeneration of bone tissue of the jaw and also within the sinus cavity autografts are still the so-called
43 “gold standard” due to their osteoinductive, osteogenic and osteoconductive regenerative capacities
44 [1]. These properties are based on the different components of the transplanted bone tissue: Beside
45 the calcified bone matrix, the different bone cell types, i.e., osteoblasts, osteocytes and osteoclasts,
46 and the connective tissue including the vasculature and, thus, endothelial cells, as well as different

47 other cell types such as macrophages (so-called “osteomacs”) and fibroblasts amongst others are
48 components of autografts [2]. Additionally, bone-associated proteins such as members of the bone
49 morphogenetic protein (BMP) family or osteopontin, osteonectin and osteocalcin beside matrix- and
50 cell-related metal ions are integral parts of autografts. Altogether, an autograft represents a
51 physiologically active transplant as all of these components allow to support the bone regeneration
52 process after implantation into a defect side [3, 4]. However, the application of autografts requires for
53 harvest of healthy bone tissue from another part of the body, i.e., from extraoral locations such as the
54 hip crest or intraoral localizations such as the mandibular ramus. Thus, one of the disadvantages of
55 the application of bony autografts is the second defect side that is created for harvesting of the bone
56 tissue. Beside different complications that could have been accompanied with this second surgical
57 intervention, the amount of bone tissue from other locations is often limited and, thus, is not sufficient
58 to fill a bone defect [5].

59 Beside autografts a variety of so-called bone substitute materials has been developed within the
60 last decades to overcome the issues with bone autografts. In this context, two main material classes
61 are differentiated: bone substitutes based on “natural” precursors and synthetic materials [6]. The
62 natural-based bone substitute materials are mainly processed outgoing from human or animal bone
63 (allo- and xenografts). For the manufacturing of allogenic bone substitutes bone tissue from living
64 donors, i.e., from femoral heads, or of dead donors is used, while xenografts are mainly processed
65 from bovine bone (or recently porcine bone). Furthermore, different natural-based materials based
66 on a variety of biopolymers such as silk fibroin amongst many others have been analyzed for
67 application as bone substitutes within the last decades [7, 8].

68 Moreover, different synthetic bone substitute materials have been developed and most of these
69 materials that are clinically applied are based on calcium phosphates such as hydroxyapatite (HAp)
70 or beta tricalcium-phosphate (β -TCP) [9]. Even mixtures of this compounds have been shown to
71 provide good healing results based on the combined degradation behavior. Moreover, a variety of
72 other synthetic materials also combined with techniques such as three-dimensional printing
73 procedures have been tested and have shown to be suitable for bone regeneration [10-12].

74 However, the regenerative properties of all the afore-mentioned biomaterials are restricted
75 particular in comparison to autografts as most of the bone substitute materials provide only a basis
76 for osteoconductive bone growth [13]. Altogether, up to date no bone substitute material has been
77 developed that features comparable regenerative capacities compared to autografts.
78 Different strategies have been originated to overcome even this issue. A first group of concepts
79 includes synthetic bone substitute materials with controllable material characteristics such as the
80 porosity or the (nano-) topography of synthetic bone substitutes [14]. It has been suggested that even
81 these special material properties, which have often stated to mimic the characteristics of the bony
82 extracellular and calcified matrix and, thus, being “biomimetic”, allow for induction of bone growth
83 [15]. Interestingly, many publications including *in vitro* studies and *in vivo* analyses within ectopic
84 tissues such as the subcutaneous connective tissue describe osteoinductive properties of especially
85 developed synthetic bone substitute materials [16]. However, the suspected osteoinductive
86 properties of such materials have never been revealed in clinical studies indicating that such concept
87 is still not tenable.

88 A second concept group includes the addition of different biologically active agents such as collagen
89 or hyaluronic acid or osteoinductive molecules such as members of the bone morphogenetic protein
90 (BMP) family [17-20]. In this context, it has been shown that the combination of synthetic bone
91 substitutes with extracellular matrix proteins such as collagen leads to diverse regenerative results.
92 On the one hand, the polymer addition can allow to increase the bony integration behavior, while
93 other results report about significantly lower bone growth rates for such a material composition
94 compared to the bone substitute material alone [21-24]. In case of an addition of molecules such as
95 BMPs different other issues have been realized, although a variety of studies has shown their
96 exceptional regenerative properties [25-27]. This results from the facts that the underlying
97 regenerative mechanisms of BMPs are not yet understood and possible side effects are not well-
98 known, especially since such molecules are usually administered in non-physiological doses

99 (thousands to millions times the amount normally found in the body) [28]. Additionally, such
100 molecules are still very expensive although being also available as recombinant proteins compared
101 to other bone substitute materials [27]. Additionally, the effect of the immobilized growth factor also
102 depends on the amount released within a certain timeframe. Hence, the material properties such as
103 porosity play a significant role [29].

104 A further group of tissue engineering concepts includes the addition of different cell types to bone
105 substitute materials. Most often osteoblasts and their precursor cells are used for such material-cell-
106 combinations even based on the fact that this cell type is mainly involved in bone regeneration by
107 deposition of the organic extracellular matrix and its following mineralization [30]. In this context,
108 also mesenchymal stem cells are of special interest as this cell type represents the earliest cellular step
109 in osteoblastic differentiation [31]. Furthermore, also the additions of different other cell types that
110 directly or indirectly support the bone growth process have been examined [32]. For example, the
111 influence of different endothelial cell types such as human dermal microvascular endothelial cells
112 (HDMEC) in mono- or co-culture with bone substitute materials have been analyzed as a fast and
113 sufficient vascularization is an important factor for bone tissue regeneration [33, 34]. Additionally,
114 blood cells or “inflammatory” cells such as cell types of the monocyte/macrophage line have been
115 used to increase the regenerative properties of bone substitutes [35]. This concept is based on the
116 assumption that such cell types express different molecules that are involved in (bone) tissue healing
117 and might induce or at least increase the process of bone regeneration [36-38]. In this context, a broad
118 spectrum of scaffolds combined different blood cells - for example platelet-rich plasma (PRP) or
119 platelet-rich fibrin (PRF) - obtained by simple centrifugation from freshly drawn venous blood have
120 also suggested to increase or even induce bone regeneration [39-41]. The assumption of such concepts
121 is that both the obtained cells and moreover growth factors present within the blood should stimulate
122 (bone) tissue regeneration [42]. However, all of these tissue engineering concepts also did not find
123 their way into the clinic as they are either not applicable in acute surgical situations due to the long
124 time spans needed for cell isolation and co-cultivation with a bone substitute or their clinical efficacy
125 has still not been proven by scientific analyses such as in case of PRP or PRF concepts.

126 A further concept is the application or the combination of different metals or metal ions with bone
127 substitute materials in the field of bone regeneration. Different metal ions are essential components
128 of different tissues such as calcium phosphates in case of the extracellular calcified bone matrix or
129 integral component of cells or proteins that are regulating essential cellular processes such as
130 proliferation or differentiation [43-45]. Altogether, the different metal ions have functional roles in
131 the physiological cellular environment and also in the course of bone healing. Thus, the application
132 of metal ions in combination with the above-mentioned bone substitutes or solely is of special interest
133 for bone regeneration [46-48]. To give an overview of the regenerative potential of the different metal
134 ions the present review summarizes the knowledge about their involvement in cellular processes and
135 the bone healing process. Additionally, further focus is on studies that already analyzed the
136 regenerative potential of bone substitutes including metals.

137 **2. Bone tissue healing and approaches for material-related support**

138 The process of bone tissue healing is based on different factors. Primarily, the bone related cells, i.e.,
139 osteoblasts and also osteoclasts and also their precursors, are involved in this process [49]. In this
140 context, most bone substitute materials allow for the osteoconductive ingrowth of osteoblasts and
141 mesenchymal progenitors acting as a scaffold structure [50, 51]. Hereafter, osteoblasts produce the
142 extracellular organic bone matrix, which mainly consists of collagen type 1 and hydroxyapatite is
143 crystallized on the collagen fibrils. Moreover, osteoblasts trigger and promote the crystallization by
144 secretion and expression of various other proteins or receptors such as RANKL and GDF5 [52]. Thus,
145 osteoblasts and their precursors are always a first starting point for different concepts that should
146 improve bone healing [53]. Interestingly, also different ions such as Mg²⁺ ions have influence on
147 osteoblastic growth, proliferation or differentiation (for further details see paragraph 3) [54, 55].

148 Moreover, influences on bone-resorbing cells and their precursors, i.e., multinucleated osteoclasts
149 and hematopoietic stem cells as well as the different intermediate stages, are of great interest in the
150 field of bone tissue regeneration [56]. This is based on the fact that a molecular cross-talk between
151 osteoblast and osteoclasts has been revealed and additionally it has been shown that both cell types
152 are organized in so-called [57, 58]. On the one hand, osteoblasts play an important role in
153 osteoclastogenesis and bone resorption based on the expression of different molecules such as the
154 receptor activator of NF- κ B ligand (RANKL), the macrophage-colony stimulating factor (M-CSF),
155 interleukin (IL)-1 β , IL-6 and IL-11 amongst others [52, 59, 60]. Furthermore, osteoblasts also express
156 different inhibiting molecules such as osteoprotegerin (OPG), the granulocyte-macrophage-colony-
157 stimulating factor (GM-CSF), IL-3, IL-12 and IL-18, which lead to the conclusion that a balanced
158 control of bone remodeling is prevailed. On the other hand, different coupling factors are nowadays
159 known that are expressed by osteoclasts such as the tartrate-resistant acid phosphatase (TRAP),
160 sphingosine 1-phosphate (S1P), bone morphogenetic protein 6 (BMP-6), hepatocyte growth factor
161 (HGF) and collagen triple helix repeat containing 1 (CTHRC1) amongst different others inducing
162 osteoblastic growth or bone formation [61]. Thus, this cell type constitutes a further approach for
163 enhancement of bone regeneration. In this context, it has already been shown that ions such as Sr²⁺
164 ions can influence bone formation via depression of osteoclast-mediated bone resorption (for further
165 details see paragraph 3).

166 Additionally, other cell types such as endothelial cells are involved in the process of bone tissue
167 healing as a sufficient vasculature and the related transport of both nutrients and metabolic end
168 products is a basic factor for bone formation [33]. Thus, this cell type and functional blood vessels are
169 also a key factor in the regeneration process. In this context, both the process of bone healing and
170 angiogenesis are directly coupled via different local factors [62]. Primarily, the so-called hypoxia-
171 inducible factor 1-alpha (HIF-1 α) pathway is induced by local hypoxia affected by a bone injury as a
172 key mechanism for coupling bone growth to angiogenesis [63]. The induction of this pathway results
173 in an increased expression of the vascular endothelial growth factor (VEGF), one of the most
174 important and strongest angiogenic cytokines, also expressed by osteoblasts and also by cell types
175 such as macrophages [64]. The expression of VEGF leads to blood vessel ingrowth within the defect
176 area and also has direct influence on osteoblast growth and proliferation as wells as matrix deposition
177 [64].

178 Moreover, the connection between the immune system and the bone tissue metabolism and
179 regeneration has been recognized in more detail in the last years. In this context, it has been revealed
180 that a special subtype of the macrophage line within bone tissue, so-called osteomacs, are a further
181 key element for bone formation [65]. Interestingly, these osteal macrophages are also integrated into
182 resting bone tissue and are enriched at sites of bone formation combined with the inflammatory
183 process following bone injury [37]. Following their activation, osteomacs have shown to promote
184 osteoblastogenesis and matrix deposition via the nuclear factor (NF)- κ B signaling pathway which is
185 important for their pro-osteogenic function [66]. Furthermore, it has been revealed that a direct cell-
186 cell-contact between osteomacs and osteoblasts takes place ensuring the osteoblastic maintenance
187 and homeostasis via sequestosome 1/p62-dependent low-level activity of NF- κ B [65, 67].

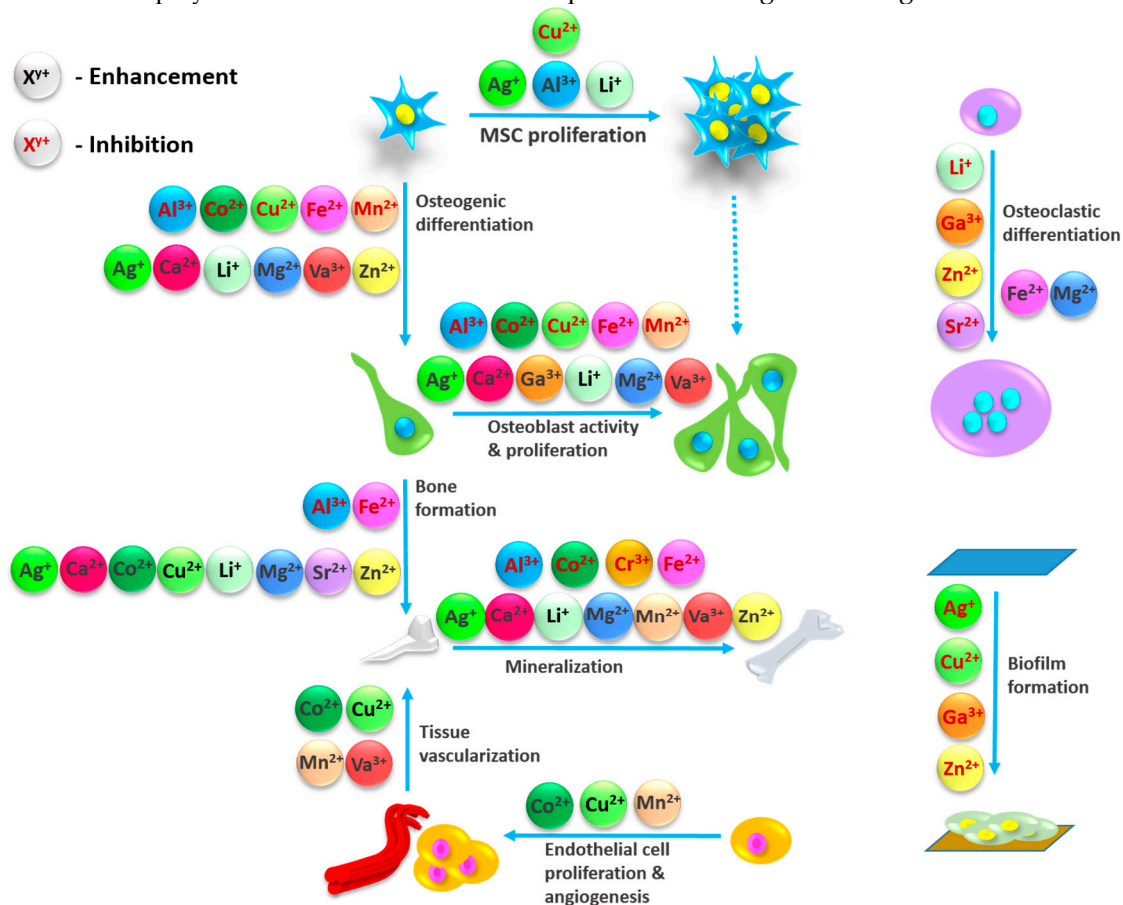
188 Finally, influence on different inflammatory cells even reacting to an implanted bone substitute
189 allows for influence on the process of bone healing [68, 69]. Thus, research in the field of biomaterial-
190 induced inflammation is more and more in the focus of bone regeneration research. In this context, it
191 has been revealed in the last decades that nearly all bone substitute materials induce an inflammatory
192 cascade, the so-called "foreign body response to biomaterials", after its application [70]. In this
193 cascade the initial accumulation of proteins, which is highly specific for every biomaterial dependent
194 on its respective physicochemical characteristics, causes the further binding of a first generation of
195 different cells and following inductions of specific signaling pathways [71, 72]. This first generation
196 of cells within an implantation bed furthermore guides the further cellular processes via expression
197 if different molecules or cytokines [73]. Interestingly, it has been revealed that in this inflammatory

198 cascade macrophages and their fused end stages, the so-called multinucleated giant cells (MNGCs),
 199 are cellular key components [74, 75]. Those cell types have shown to express both pro- and anti-
 200 inflammatory molecules such as VEGF that guide the integration behavior and factors such as the
 201 implant bed vascularization of bone substitute materials [64, 76]. Additionally, other cell types such
 202 as granulocytes or thrombocytes have supposed and revealed to have eminent influence on this tissue
 203 reaction cascade, which finally leads to different outcomes of the bone regeneration process. In this
 204 context, material factors such as the chemical composition or physical material properties such as the
 205 porosity or the surface structure but also the involvement of different ions such as Cu^{2+} ions have
 206 shown to allow for influence on the inflammatory tissue reaction to a bone substitute material (for
 207 further details see paragraph 3) [71, 77, 78].

208

209 3. Metal ions, their physiological functionalities and role in bone healing

210 Metals are widely accepted as implant material since a few decades. Even when a solid metal is
 211 applied to physiological environment it is always in equilibrium with its ions. These metal ions are
 212 responsible for a variety of biochemical functions which are important for the different steps of bone
 213 regeneration as they influence the equilibrium between osteoblasts, osteoclasts and osteocytes. Thus,
 214 metals and their corresponding ions which have an influence on the process of bone healing should
 215 be mentioned here (Figure 1). We will also line out the impact on different states of tissue formation
 216 and the interplay between related metal ions in processes leading to bone regeneration.



217

218

219

220

Figure 1: Influence of metal ions on the variety of processes involved in bone regeneration.

3.1. Aluminum (Al^{3+})

221

222

Aluminum does not belong to the group of trace elements, is not involved in any physiological functions and is consequently not essential for the human organism [79]. Intake of larger elevated

223 quantities of aluminum is associated with toxic effects leading to serious adverse reactions including
224 anemia, encephalopathy and osteoporosis as Al^{3+} ions compete with essential ions like Fe^{2+} [80-82].
225 Investigation of the specific reaction of human neural cells to aluminum exposure for example
226 showed that concentrations as little as 100 nM of aluminum sulfate significantly elevated atypical,
227 pro-inflammatory and pro-apoptotic gene expression [83].

228 Reports about the functions of aluminum in bone formation are ambiguous. Positive impacts of
229 aluminum supplementation on the osteogenesis in beagles as well as in osteopenic rats was
230 previously demonstrated and initiated a further interest for the investigation of aluminum in tissue
231 engineering [84, 85]. In contrast to these findings, expression of osteoblast activity markers was
232 substantially lower while expression of apoptotic markers was increased when treated with
233 aluminum, demonstrating impaired cellular activity and survival and a clear link between aluminum
234 intoxication and compromised bone formation [86]. Quarles and colleagues put their findings on the
235 positive impact of aluminum on de novo bone formation into context with the contradicting literature
236 and suggested that aside from discrepancies in the applied model/organism, aluminum
237 concentrations and time of exposure, a paradoxical impact of aluminum on mesenchymal progenitors
238 and mature osteoblasts could be the main reason for these dissimilar observations [85]. This
239 hypothesis was further supported by another group, which demonstrated that aluminum ions
240 provoked a chemotrophic stimulus in preosteoblasts while having an inhibitory effect on osteoblasts
241 [87].

242 Negative impacts of aluminum on osteoblast function, however, are prevailing in the contemporary
243 literature. An in vivo study in rats assessed the effects of aluminum exposure on the uptake of bone
244 mineral elements, trace elements and bone mineral density. The level of analyzed trace elements were
245 significantly lower with aluminum exposure and deposition of calcium, phosphorus and magnesium
246 was decreased in comparison to the control population. Bone mineral density in the femur
247 metaphysis of the aluminum-treated group was also significantly lower compared to the control,
248 resulting in pronounced bone loss [88]. Altogether, aluminum does not seem to contribute to bone
249 and tissue healing but to have a rather opposing impact in this process so that, aside from favorable
250 mechanical properties, application of aluminum in implantable medical devices offers no
251 scientifically evident benefits. Furthermore, other bioceramics such as zirconia oxide are
252 progressively emerging as bioinert alternative to aluminum oxide [89].

253

254 3.2 Calcium (Ca^{2+})

255 Calcium is an important functional component of biodegradable calcium phosphate-based
256 biomaterials designated for bone regeneration in orthopedics, trauma surgery, and in dentistry (for
257 reviews: [90-92]).

258 Calcium is the most common mineral of the body and is primary stored in the skeleton [93]. Calcium
259 homeostasis is tightly regulated by parathyroid hormone (PTH) and calcitonin which regulate
260 calcium serum levels by stimulating (PTH) or inhibiting (calcitonin) bone resorption – mediated by
261 osteoclasts. During bone remodeling bone resorbing osteoclasts can create local concentrations of
262 extracellular calcium ions up to 40 mM [94]. These microenvironmental increases are known to
263 inhibit resorption activity of osteoclasts, and to stimulate proliferation and differentiation of
264 mesenchymal stromal cells [93, 95-99] and osteoblasts [100, 101].

265 During the 1980s extracellular calcium was shown to activate an extracellular G-protein-coupled
266 receptor, termed calcium sensor receptor (CaSR) [102]. The CaSR is expressed in cells of the
267 hematopoietic lineage, such as in monocytes [103], and osteoclasts [104] as well as in cells of the
268 mesenchymal lineage [93, 99, 101, 105, 106]. Regarding the high responsiveness of the cells of the bone
269 to extracellular calcium, elevated levels enhance proliferation chemotaxis and osteogenic
270 differentiation of bone marrow-derived mesenchymal stromal cells in a dose-dependent manner by

271 activating the CaSR [93, 99]. Downstream, the intracellular pathway induces phosphorylation of
272 extracellular signal-regulated protein kinases 1 and 2 (ERK 1/2) [99] which are part of the MAPK
273 signaling pathway playing an important role in regulating cell proliferation in various mammalian
274 cells [107]. The activation of the CaSR in response to extracellular calcium levels also stimulates
275 phospholipase C (PLC) and induces sustained increase of cytosolic calcium in rat calvarial osteoblasts
276 [106]. The activation of PLC results in generation of inositol 1,4,5-trisphosphate (IP₃) and triggers IP₃-
277 receptor-mediated calcium release from endoplasmic reticulum. As a result, store operated calcium
278 entry (SOCE) mediates extracellular calcium entry into the cells for endoplasmic reticulum -calcium
279 store filling [108]. In addition to the effects mediated by the CaSR and the SOCE route voltage gated
280 calcium channels may also serve as structural units accounting for calcium entry into osteoblasts
281 [106], and osteogenic differentiation of osteoprogenitors [96, 109].

282 Given the superior significance to modulate cellular functions, variations of extracellular calcium in
283 the milimolar range result in proliferation, survival and chemotaxis as well as in differentiation of
284 osteoblasts [101, 106] and bone marrow derived mesenchymal stromal cells (MSCs) [93, 96, 99].
285 Optimal conditions to stimulate proliferation of rat calvarial osteoblasts include extracellular calcium
286 concentration in the range of 3 mM and 10mM, respectively [101]. Proliferation of bone marrow
287 derived MSCs harvested from different species (i .e. human, porcine, rat) is effectively supported by
288 concentrations of 4mM [99], 7.8 mM [96], and 10 mM [93]. Additionally, osteogenic differentiation
289 capacity of human bone-derived MSCs is stimulated in response to extracellular calcium
290 concentrations in the range of 10 mM and 20 mM [98].

291 According to the pivotal role of calcium in cellular functions and to composition of natural bone,
292 various calcium phosphate based materials have been developed for bone replacement therapies [90-
293 92]. Incorporation of the calcium phosphate phases modulates bioactivity of the biomaterials, and as
294 pointed out in previous studies, high bioactivity is equivalent to calcium phosphate binding capacity
295 and causes depletion of calcium in close vicinity to the biomaterial [97, 104, 110]. Calcium phosphate
296 deposition along the surface of bone substitute materials represents an advantageous property to
297 support osseointegration. However, the calcium deficient microenvironment in close vicinity to the
298 materials remains obscure – especially considering the aforementioned calcium-dependent effects on
299 osteoblasts and progenitor cells. It has been shown that osteoprogenitors – as in the case of bone-
300 derived MSCs – can overcome calcium deficiency when they are cultured in combination with highly
301 bioactive xerogels [97]. The mechanism by which the cells maintain their functional integrity even in
302 response to calcium levels next to zero is still not clear. Given the fact that the materials with high
303 bioactivity are composites, it might be concluded that the beneficial effects on cell survival,
304 proliferation and differentiation are mediated in large part by ionic dissolution products such as silica
305 [97, 111] or phosphate ions [112, 113]. According to this, it has been postulated that best results of
306 osteogenic differentiation of osteoblast progenitors along with bone formation may be expected when
307 calcium phosphate based materials dissociate easily to calcium and phosphate ions [113].

308

309 3.3 Chromium

310 The physiological function of chromium in human is currently under debate. Though, some cellular
311 functions of chromium have been reported, in 2014 the European food safety authority officially
312 removed it from their list of essential micronutrients [114, 115]. The impact of chromium exposition
313 on osteoblasts was investigated in several studies, whereby only toxic effects, causing reduced DNA,
314 RNA and protein synthesis, were reported [116, 117]. Furthermore, chromium suppressed
315 collagenase activity in osteoblasts, which reduced collagen formation and deposition and also
316 negatively affects new bone formation [117].

317 In the field of reconstructive medicine, cobalt-chromium (CoCr) is one of the main alloys used for
318 total hip arthroplasty. However, Co²⁺ ion release from CoCr surfaces has been reported to severely

319 impact mesenchymal stem cells by altering osteogenic gene expression, affecting osteogenic lineage
320 differentiation and compromising the mineralization process [118]. The impairment of bone
321 formation by chromium and cobalt was further analyzed by the effect of these ions on the expression
322 of various TGF- β isoforms and mineralization in MG-63 and SaOs2 osteosarcoma cells as well as in
323 primary human osteoblasts. While Co²⁺ decreased the expression of different TGF- β isoforms in all
324 investigated cell types, Cr³⁺ had no impact in this manner. Cr³⁺ on the other hand strongly inhibited
325 the mineralization process of these cells in vitro, whereas Co²⁺, within the range of the tested
326 concentrations, showed no inhibitory effects on mineralization [119].

327

328 3.4 Cobalt (Co²⁺)

329 As cobalt is a compound of cobalamin, it is an essential trace element, which stimulates the
330 production of red blood cells and promotes angiogenesis by activating hypoxia-inducible
331 transcription factors (HIF) [120-122]. Previous studies demonstrated a rather unfavorable effect of
332 Co²⁺ ions released from CoCr surfaces on osteogenic lineage differentiation of hMSCs, TGF- β isoform
333 expression in osteoblasts and the mineralization process, whereby recent data indicates that the
334 impaired mineralization reported by Schröck and colleagues rather results from Cr³⁺ ion release than
335 from Co²⁺ ions [118, 119].

336 The angiogenic capacities of cobalt ions sparked the idea of incorporating this metal into different
337 materials used for bone healing in order to stimulate vascularization of implanted grafting materials,
338 enhance remodeling processes and thus, support the overall regeneration process. The impact of Co²⁺
339 ions incorporated into calcium phosphate (CaP) coatings for poly-lactic acid (PLA) particles on new
340 blood vessel formation was studied in an intramuscular implantation model in goats. The
341 inflammatory reaction following a 12-week implantation course demonstrated no pathologic
342 differences in PLA particles coated with solely CaPs or coated with Co²⁺ containing CaPs. Formation
343 of blood vessels was significantly increased when Co²⁺ containing CaP coated PLA particles were
344 implanted and vessel size was notably increased, suggesting a positive impact of Co²⁺ on
345 vascularization in vivo [123].

346 The impact of Co²⁺ containing CaPs on osteoporotic alveolar bone regeneration was further
347 investigated in rats. Biocompatibility assessment of the material was approved for epithelial Caco-2
348 and osteoblastic MC3T3-E1 cells, whereby no toxic effects in Caco-2 cells, however, considerable
349 decrease in cell viability and impairment of cytoskeletal organization was observed in MC3T3-E1
350 cells. Despite the negative impact of Co²⁺ ions on osteogenic cells, hydroxyapatite (HAp)
351 nanoparticles doped with Co²⁺ demonstrated dose-dependent acceleration of osteogenesis,
352 osteoporotic bone regeneration and graft material substitution in comparison to HA-nanoparticles
353 without Co²⁺. The authors listed several hypothesis for their observations including increased
354 transport of Ca²⁺ ions into the extracellular fluids facilitated by the moderate toxicity of Co²⁺ ions as
355 well as increased cytokine production and release, which could potentially boost aminopeptidase
356 activity together with migration and proliferation of endothelial cells [124].

357 The combination of Co²⁺ HAp nanoparticles with blood or plasma rich in growth factors (PRGF) was
358 shown to induce the generation of large quantities of osteoblasts, increased mineralization and
359 accelerated bone regeneration [124]. Taking into consideration that recent studies demonstrated
360 impaired growth factor expression and osteogenic lineage determination in hMSCs exposed to Co²⁺,
361 these observations seem reasonable, as blood and PRGF may compensate this lower expression and,
362 thus, enable proper osteogenic lineage differentiation [119]. Furthermore, the study implicates that
363 bone mineral containing scaffolds as presented in this study are suitable for cobalt incorporation, as
364 cobalt does not impair but rather seems to support the mineralization process [119, 124].

365 Similar findings were made by another group who developed a hydrogel with incorporated Co²⁺
366 ions. Hydrogels solely doped with Co²⁺ did not increase the amount of regenerated bone volume,

367 bone surface and bone surface density in a rat model in vivo, whereas addition of BMP2 to the
368 hydrogel did. The observed gain was even more pronounced with the simultaneous loading of Co^{2+}
369 and BMP2 onto the hydrogel, which again favors the hypothesis of a synergistic effect of Co^{2+} in
370 conjunction with growth factors in graft vascularization and bone regeneration [125]. Increased
371 collagen deposition, new bone formation and bone hardness was also reported for cobalt-containing
372 bioglasses compared to bioglasses without cobalt in critical size defects in the rabbit's femur in vivo
373 [126]. Additionally, the authors showed that inclusion of both strontium and cobalt into the bioactive
374 glasses even further ameliorated the bone regeneration process.

375 3.5 Copper

376 While Cu^{2+} is the most stable oxidation state in aqueous solution it can also be present as Cu^+ in
377 human body exhibiting diverse properties and functions [127]. Together with iron and zinc, copper
378 is one of the most important metals for humans and especially needed to generate Cu-proteins which
379 have enzyme functions. Cu-proteins have three main functions in living organisms such as
380 participation in electron-transfer reaction, transport of oxygen and transport or storage of the metal
381 itself.

382 Therefore, copper is involved in multiple physiological functions, the regulation of bone metabolism
383 and turnover among them. Cu imbalances also affect the nervous system and can lead to vascular
384 abnormalities in the human body. The impact of copper deficiency on skeletal growth and
385 development was previously assessed in several studies [128, 129]. Copper caught attention in the
386 field of bone regeneration because of its antibacterial properties and its ability to stimulate collagen
387 fiber deposition and angiogenesis, which represents the first step towards the formation of vital and
388 vascularized tissue [130-132]. The effect of copper-doped silicate bioceramics on vascularization was
389 subjected to several studies and a positive impact on the expression of angiogenic growth factors in
390 human umbilical vein endothelial cells (HUVECs) and human dermal fibroblasts (HDFs) in response
391 to Cu^{2+} released from copper silicate bioceramics was recently reported by Kong and colleagues [133].
392 Thus, the release of Cu^{2+} ions from porous matrices like bioactive glass should facilitate the ingrowth
393 of bone into the scaffold matrix [134].

394 Current data supports enhanced osteogenic differentiation of mesenchymal stem cells mediated by
395 copper supplementation. Early studies on the effect of copper on MSCs derived from
396 postmenopausal women demonstrated reduced proliferation, a 2-fold enhancement of differentiation
397 into osteoblasts and increased calcium deposition, while alkaline phosphatase activity was
398 considerably diminished in these cells but shifted to an earlier timepoint [135]. Similar findings on the
399 suppression of alkaline phosphatase activity mediated by copper exposition was observed in rat
400 MSCs by Li and colleagues, whereby they reported a clear reduction in osteogenic differentiation of
401 rat MSCs concomitant with the reduction of several osteogenic genes, alkaline phosphatase activity
402 and bone nodule formation. In addition, cytoskeletal abnormalities during osteogenesis was found
403 in these cells. The process of ectopic bone formation in a rat model was also significantly impaired
404 by presence of copper and while vascularization in the regenerated soft tissue was promoted,
405 collagen formation was strongly inhibited [136].

406 These findings are supported by a study conducted with pre-osteoblastic MC3T3-E1 cells cultured
407 on copper containing bioglasses. While no effects on proliferation and alkaline phosphatase activity
408 of these cells was noted with scaffolds doped with 0.4 to 0.8 wt.% CuO, 2.0% showed a significant
409 reduction on both. In an in vivo approach in rat calvarial defects showed that this higher
410 concentration of Cu^{2+} ions also substantially reduced new bone formation from $46 \pm 8\%$ to $0.8 \pm 0.7\%$,
411 while lower concentrations showed no such impairment. On the other hand, the authors found a
412 stimulatory effect on blood vessel formation in dependence of the copper content of the scaffolds
413 with the biggest impact seen for the highest concentration of 2.0% CuO [137]. Benefits of copper
414 supplementation in the regeneration of critical-sized calvarial defects in rats were further reported

415 by the comparison of chitosan scaffolds and chitosan scaffolds doped with copper. Analysis of micro-
416 CT scans after 4 weeks of healing indicated twice the amount of bone volume in the defects treated
417 with copper containing chitosan scaffolds as compared to scaffolds without copper [138].

418

419 3.6 Gallium (Ga^{3+})

420 Gallium is a metal which serves no known essential functions in human. While currently being
421 investigated in cancer treatment because of its anti-proliferative properties resulting from the
422 interference with iron-dependent cellular functions, studies also demonstrated that short term
423 gallium treatment reduces bone turnover in vivo and increases calcium content of bone in patients
424 suffering from cancer-related hypercalcemia [139]. Furthermore, gallium has the potential to disrupt
425 microbial iron utilization by interacting with iron-binding bacterial molecules called siderophores. In
426 this manner, gallium downregulates the bacterial iron uptake and impairs their growth [140].
427 Gallium-EDTA coated titanium chips exhibited significant antimicrobial activity against *Escherichia*
428 *Coli* for more than 28 days after coating, underscoring a promising application of gallium-based
429 coatings for effective prevention of biofilm formation, which could be used in dental and orthopedic
430 reconstructive surgery [141]. Additionally, gallium coated titanium implants showed superior
431 antibacterial properties in vivo and consequently more effective prevention of biofilm formation than
432 silver coatings [142].

433 Several studies analyzed the effect of gallium administration on osteoclasts and osteoblasts. While
434 osteoclastic lineage differentiation and resorption activity was lowered by gallium, no impact on
435 viability and proliferation of osteoblasts was noted [143]. In an in vivo approach using a rabbit
436 femoral defect model, gallium-loaded calcium phosphate cements showed no superiority over
437 calcium phosphate cements without gallium in terms of bone healing, whereby the authors implied
438 that no effect was observed due to the little resorption of the material and consequently low release
439 of Ga^{3+} -ions [144]. In a subsequently conducted study the gallium release from Ga-CaP was optimized
440 and re-evaluated for its beneficial properties in bone healing. Upregulation of osteoblastic marker
441 expression was observed in primary human osteoblasts cultured on the Ga-CaP, whereby late
442 osteoclastic markers were downregulated in primary human monocytes which were previously
443 induced towards the osteoclast lineage.

444 The in vivo properties of Ga-loaded CaPs in new bone formation were assessed in a murine bone
445 defect healing model; aside from an enhanced total defect-fill, Ga-CaPs also promoted the synthesis
446 of mature organized collagen [145]. With respect to the current literature, gallium holds a set of
447 promising qualities for future applications in tissue engineering.

448

449 3.7 Iron (Fe^{2+})

450 Iron is one of the most important ions in the human organism as it is essential for a variety of cellular
451 processes [146-148]. Different cellular effects such as the synthesis of deoxyribonucleic acid (DNA)
452 and ribonucleic acid (RNS), proteins, electron transport processes, cellular proliferation and
453 differentiation are related to iron ions [149, 150]. These effects are based on the involvement of iron
454 ions mainly as components of enzyme molecules, such as oxidases, catalases, peroxidases, aconitases,
455 ribonucleotide reductases and nitric oxide synthases amongst others [150-152]. As coordinating ion in
456 the center of hemoglobin and myoglobin, iron is an essential trace element, required for oxygen
457 transport and regulation of several metabolic enzymes [153, 154]. Further, iron is the loosely bound
458 ion component of the procollagen proline hydroxylase and the procollagen lysine hydroxylase [155].
459 Both enzymes effect the hydroxylation of proline and lysine residues in precursors of collagen. Large
460 amounts of iron released from iron-containing implants, however, may cause excessive iron levels in
461 the blood. Here, the free iron can react with peroxides and trigger the formation of free radicals which

462 are highly reactive and damage lipids, proteins, DNA as well as cellular structures [156, 157].
463 Additionally, hemochromatosis has been demonstrated to result in osteoporosis mediated by
464 increased ferroxidase activity of ferritin and in vitro experiments demonstrated inhibition of
465 osteogenic lineage differentiation in human osteoblasts concomitant with decreased calcification
466 caused by iron overload [158-160]. In vivo experiments in zebrafish larvae demonstrated that the
467 mechanism by which iron-overload causes impaired osteoblast function and mineralization is based
468 on the increased generation of reactive oxygen species. Application of deferoxamine, an iron chelator
469 capable of removing whole-body iron, ameliorated the iron-induced negative effects on osteoblastic
470 marker expression and mineralization [161]. Same was observed for hepcidin, a regulator of iron-
471 uptake, which is also capable of removing whole body iron. Likewise, hepcidin downregulation
472 elevates iron level and causes iron-overload mediated interference with osteogenesis [162].

473 Iron exposure of human bone marrow mesenchymal stem cells (BMSCs) decreased their
474 differentiation towards the osteogenic lineage as well as extracellular matrix mineralization with a
475 total block of lineage commitment at a concentration of 50 μ M. In vivo experiments in mice were able
476 to reproduce these findings. The inhibitory effect of iron, however, was specific for osteogenic lineage
477 differentiation, whereas no impact on chondrogenesis and adipogenesis was noted [163].
478 Furthermore, the promotion of osteoclast formation mediated by iron was previously reported,
479 which additionally underscores the unfavorable features of iron for the purpose of biomedical tissue
480 engineering [164]. In contrast to these previous results, Wang and colleagues reported positive impact
481 of iron oxide nanoparticles (IONPs) on the osteogenic differentiation of human BMSCs in vitro
482 mediated by MAPK signaling. The authors speculated that the negative impact of iron on
483 osteogenesis observed in previous studies resulted from increased ROS formation and ferritin
484 activity, whereby this process is proposed to be prevented by nanoparticle formulations [165].
485 Moreover, Zhao and colleagues analyzed both effects of excessive and low body iron conditions on
486 osteoblast activity [166]. The results showed that an increased iron concentration inhibited
487 osteoblastic activity in a concentration-dependent manner, while a mild iron deficiency lead to an
488 increase of the cellular activity. In contrast, a severe low iron level completely inhibited osteoblastic
489 differentiation. An enhanced osteoclast formation is one result of an increased iron concentration
490 while osteogenic stimuli are blocked at the same conditions [167]. Thus, further studies will have to
491 clearly determine the potential benefits of iron in tissue engineering.

492

493 3.8 Lithium (Li^+)

494 Lithium is a non-essential trace element and consequently fulfills no known functions in the human
495 organism. However, due to its beneficial impact in the treatment of psychological disorders, lithium
496 has been widely introduced into medical applications [168]. Among the various mechanisms of action
497 that have been proposed for lithium in this manner the stimulation of neural progenitor cell
498 proliferation by the Wnt/ β -catenin pathway, which leads to increase of the brains grey matter, is
499 widely accepted [169, 170]. Interestingly, the proliferation of other cell types such as MSCs is also
500 regulated by the Wnt/ β -catenin pathway, suggesting that lithium might also modulate the
501 proliferation of these cells [171]. In fact, a recent study reported increased proliferation of hMSCs
502 stimulated by lithium-mediated Wnt/ β -catenin signaling in vitro [172]. Additionally, previous
503 studies reported this pathway to be a main regulator of osteoblastogenesis, which made lithium
504 application in the field of tissue engineering even more appealing [173]. Though few studies reported
505 beneficial impact of lithium supplementation on bone mineral density and a reduction of the risk of
506 fracturing, the molecular mechanisms by which lithium facilitates these effects are not completely
507 elucidated yet [174, 175]. In a transcriptome-based approach in order to identify the impact of lithium
508 on osteoblastogenesis, Satija and colleagues reported diminishing proliferation of hMSCs treated
509 with lithium, however, decreased expression of adipogenic and osteoclastogenic factors
510 accompanied by the induction of osteoblastogenic markers associated to collagen-1 deposition and
511 mineralization, whereby similar results were also reported by other groups [176-178]. Systemic

512 lithium application exhibited beneficial effects on bone healing following distraction osteotomy in
513 the tibia of rats. Bone mineral density, quantity of newly formed mature bone tissue and bone mass
514 regeneration were increased in rats who received a lithium solution through gastric gavage in
515 comparison to those receiving a saline solution, pointing to accelerated callus ossification and bone
516 healing mediated by lithium [179].

517 To further utilize the beneficial effects of lithium on bone regeneration, various biodegradable lithium
518 containing scaffolds have been developed and tested for their potential in bone regeneration,
519 whereby preliminary experiments on lithium release, toxicity and osteoblastic cell activity on such
520 scaffolds were promising [180, 181]. In vitro experiments comparing pure HAp with lithium-doped
521 HAp scaffolds demonstrated increased osteoblast activity, resulting in accelerated material
522 degradation, whereby the degradation products exhibited no toxic impacts on osteoblasts, however,
523 enhanced their proliferation. Additionally, compressive strength testing revealed favorable
524 mechanical properties of lithium-doped HAp scaffolds [182]. Further evidence on the beneficial
525 impact of lithium incorporation into calcium phosphate cement scaffolds on bone healing was
526 recently demonstrated. Lithium release from this material stimulated the proliferation and
527 differentiation of osteoblasts in vitro by Wnt/ β -catenin activation. Application of lithium-doped
528 calcium phosphate cements significantly increased osteogenesis and defect repair in vivo and
529 showed superior osteoconduction and osteointegration compared to pure calcium phosphate
530 cements [183]. Overall, the literature emphasizes that lithium regulates growth and development of
531 osteogenic progenies while suppressing osteoclast development, whereby identification of the exact
532 mechanisms of lithium orchestrating either differentiation or proliferation of osteoblasts represents a
533 pivotal goal for future clinical applications. Nonetheless, lithium seems to directly regulate and
534 benefit osteogenic lineage cells, whereas other metallic ions, such as copper and cobalt, rather seem
535 to impact bone regeneration by their impact on endothelial cells and accelerated vascularization.

536

537 3.9 Magnesium (Mg^{2+})

538 Magnesium is an alkaline earth metal and belongs to group 2 metals of the periodic table. The
539 mammalian body consists of approximately 0.4 g magnesium/kg body weight [184]. More than 90 %
540 is bound and stored in bone, muscle and non-muscular soft tissue [184, 185], while only a small
541 amount (1% - 5%) [185] resides in extracellular fluids [186] in form of ionized / free magnesium (55-
542 70%) or is bound to proteins and anions [184].

543 Magnesium is an important intracellular cation [185-187] as it is cofactor for more than 300 enzymatic
544 reactions, essential for synthesis of proteins and nucleic acids [185, 188], and for transport of both,
545 potassium and calcium ions [185]. Magnesium is also crucial for transphosphorylation of ATP, and
546 changes of intracellular magnesium levels might influence several pathways [189].

547 As magnesium maintains bone strength [185] and bone formation capacity [184] adequate dietary
548 magnesium plays a major role in musculoskeletal health, and is relevant to prevent osteoporosis
549 [190]. In contrast, magnesium deficiency exerts negative effects on rat bone metabolism, systemic
550 bone mass [191], and contributes to osteoporosis in humans [189]. It has been proposed that the effects
551 of magnesium deficiency might be the result of increased levels of $TNF\alpha$, IL-1 [192], and NF- κ B ligand
552 (RANKL), along with decreased serum levels of osteoprotegerin (OPG) [193].

553 According to the superior role of magnesium in cellular functions, magnesium-based materials are
554 regarded as promising candidates for bone replacement therapies due to stimulation capacity of bone
555 cell differentiation in vitro [194-197] and bone formation in vivo [198-201]. Currently available
556 materials include different magnesium containing compounds such as oxides, phosphates and
557 silicates that are used as bone cements, bone scaffolds or implant coatings. Overviews of the different
558 magnesium-based materials – such as bioceramics, e.g. magnesium phosphates ($MgO-P_2O_5$),

559 calcium magnesium phosphates (CaO-MgO-P₂O₅), and magnesium glasses (SiO₂-MgO) [202] are
560 given in recent systematic reviews [203-215].

561 Numerous in vitro studies attend to the effects of magnesium ions on cells of the bone, in terms of
562 enhancing proliferation and migration as well as ALP activity of human osteosarcoma MG-63 cells
563 [216], increasing viability and differentiation capacity of a human osteoblast cell line (hFOB1.19,
564 ATCC) [217], cell proliferation of bone marrow derived stromal cells (BMSC), and expression of α 2
565 und α 3 integrins [218]. However, additional data provide evidence that the effects of magnesium ions
566 develop dose-dependently [217]. Concentrations of about 1- 3 mM Mg²⁺ stimulate gap junctional
567 intercellular communication (GJIC) of osteoblasts [217], while viability, proliferation and
568 differentiation of human BMSCs are ensured by concentrations in the range of 2.5 – 10 mM [216, 218-
569 220].

570 In contrast, decreased mineralization capacity and matrix deposition of BMSCs have been observed
571 in response to magnesium concentrations higher than 1.3 mM Mg²⁺ [221-223]. According to the role
572 of magnesium as a physiological calcium antagonist [222], it has been suggested that magnesium
573 substitution for calcium in hydroxyapatite structure [224] and/or modulations of intracellular calcium
574 oscillations with consecutive suppression of spontaneous ATP release and inactivation purinergic
575 receptors are responsible for the decreased mineralization capacity of the cells [221]. Additionally,
576 magnesium has a competitive role against Matrix gla protein (MGP) suggested as a potent inhibitor
577 of HAp crystal growth during mineralization [225]. These results are consistent with emerging
578 studies demonstrating significant suppression of mitochondrial accumulation of calcium ions in
579 MSCs [222], and inhibition of excess calcium-induced mineralization in response to high extracellular
580 magnesium [226]. Similarly, decreased intracellular calcium concentration and decreased calcium
581 influx have been observed when MSCs have been cultured in presence of high magnesium
582 concentration [223]. Competition between calcium and magnesium ions for same ion transporters,
583 such as transient receptor potential cation channel, subfamily M, member 7 (TRPM7) [223] and/or
584 inhibition of expression of calcium-sensing receptor (CaSR) [226] might be responsible for the
585 decreased mineralization capacity. In terms of how high concentrations of Mg²⁺ ions modulate bone
586 cell metabolism and bone cell function, the Wnt/ β -catenin anti-calcifying pathway and the
587 magnesium transporter SLC41A1 have been shown to be involved in magnesium-mediated signaling
588 of BMSCs [223].

589 The high grade of biodegradability which avoids second surgery for implant removal and prevents
590 formation of foreign body giant cells in close vicinity of permanent implants has been designated as
591 a major advantage of the magnesium-based materials [227]. As architecture and pore structural
592 conditions of magnesium-enriched scaffolds greatly influence bone formation and remodeling
593 activities [228] hydrogen gas released during degradation of magnesium-enriched scaffolds enlarges
594 pre-existing pores, and expands the space for invading cells and blood vessels [201]. Given these
595 beneficial effects, magnesium-based materials have emerged as a new class of biodegradable
596 biomaterials for bone tissue engineering – referred to as next-generation biomaterials [227].

597 However, considering the rapid degradation rates, magnesium-based implants are still not
598 commonly used in clinical practice [212, 227, 229]. The “high magnesium microenvironment” created
599 by rapid corrosion of magnesium alloys might disturb calcium-dependent processes and physiology
600 of the cells localized in close vicinity to the implants [222]. Therefore, the balance between calcium
601 and magnesium ions is not only crucial for bone physiology [222] but also for successful
602 osseointegration of magnesium-based materials.

603 Additionally, due to rapid corrosion rates magnesium-based implants hold the risks of structural
604 failure and toxic responses immediately after implantation [227]. In the course of degradation
605 magnesium hydroxide and hydrogen gas are produced both of which cause detrimental effects on
606 cells and tissue localized close to the implant [188, 230]. Controllable in vivo corrosion rates, in terms

607 of establishing sufficient corrosion protection methods on different levels might represent promising
608 tools to overcome these disadvantages [188, 212, 227, 229, 230].

609

610 3.10 Manganese (Mn^{2+})

611 Manganese is an essential element and crucial for the proper function of multitudinous enzymes in
612 living organisms [231]. Divalent cations such as Mn^{2+} are furthermore known to influence cell
613 migration by modulating focal adhesion organization via integrins and actin stress fiber formation
614 [232, 233]. These properties make manganese an interesting candidate for improving ingrowth and
615 integration of bone grafts and other implantable materials alike. The impact of manganese on MG-63
616 osteoblastic cells was evaluated in order to confirm this theoretical benefit of manganese
617 supplementation in the process of new bone formation. Manganese supplementation reduced cell
618 proliferation, migration, ERK/MAPK-signaling and collagen I as well as alkaline phosphatase
619 expression in a dose-dependent manner. Interestingly, mRNA level of bone sialo protein (BSP) were
620 increased by manganese exposition, whereas BSP protein level were not elevated [234].

621 Interestingly, doping alumina tubes with manganese significantly enhanced tissue maturation and
622 osteogenesis in vivo in rats, whereby the authors noted that the surface structure of the alumina tubes
623 was altered by manganese incorporation which made it impossible to distinguish whether the
624 observations resulted from the phase composition or the surface topography modification [235].
625 However, manganese is also reported to hold insulin-mimetic properties and other substances within
626 this class such as VAC increased fracture site vascularization by local application, which lead to the
627 hypothesis that manganese might also accelerate fracture healing [236, 237]. In fact a group reported
628 significant increase in mechanical properties of bone, mineralized tissue formation and VEGF-
629 expression in a rat femoral fracture model when manganese chloride ($MnCl_2$) was supplemented.
630 Additionally, blood vessel density was dramatically increased by $MnCl_2$ treatment, suggesting
631 increased vascularization, fracture healing and osteogenesis and implicating a potential function for
632 manganese in tissue engineering [238].

633

634 3.11 Silver (Ag^+)

635 Due to its antimicrobial properties, silver has a long-time history in application for medical purposes,
636 whereas the investigation of potential functions of silver in bone regeneration is a quite recent
637 occurrence [239]. Analysis of the tissue response to silver acetate coated Dacron vascular grafts
638 implanted into the dorsal skinfold chamber in mice revealed higher functional capillary density
639 without affecting inflammatory host tissue response, collagen formation, apoptosis and cell
640 proliferation as compared to uncoated grafts [240]. Furthermore, functionalization of silver
641 nanoparticles in tissue regeneration has already been introduced into commercially available wound
642 dressings, as the exhibit outstanding anti-microbial and anti-inflammatory properties [241-243].
643 Additional arguments for utilization of silver nanoparticles instead of other silver formulations like
644 silver nitrate in tissue engineering were recently reported by Quin and colleagues [244]. They showed
645 that the lowest toxic concentration of silver nanoparticles on urine derived stem cells was
646 substantially higher than that assessed for silver nitrate. More interestingly, however, was the
647 reported promotion of osteogenic lineage induction and actin polymerization of these cells, which
648 was only observed for AgNPs, however, not for $AgNO_3$ [244]. In fact, stimulatory impact of AgNPs
649 on the mineralization of MC3T3-E1 osteoblastic cells maintained by miRNA mediated increased
650 expression of genes associated with bone formation was previously reported [245].

651 In order to identify putative impacts of AgNPs in the process of osteogenic lineage induction, the
652 entire transcriptome of MC3T3-E1 cells in response to AgNP exposure was analyzed. Here the
653 authors found that, aside from the upregulation of different bone morphogenic proteins important
654 for osteogenesis, the enhancement of osteoclastic marker expression was the most pronounced

655 transcription-based alteration [246]. Based on the stimulatory properties of AgNPs on keratinocyte
656 proliferation and migration and fibroblast differentiation, which contributes to the promotion of
657 wound contraction, the impact of AgNPs on proliferation and differentiation of MSCs was analyzed
658 [247, 248]. AgNPs successfully promoted MSC proliferation and osteogenic differentiation in vitro.
659 In vivo experiments using a femoral fracture model in mice supports the preliminary observations,
660 as AgNPs encapsulated in collagen were able to accelerate callus formation and fracture gap closure.
661 Though the exact impact of AgNPs in this process remains elusive, the authors suggested possible
662 chemotactic impact of AgNPs on MSCs and fibroblasts as well as induction of MSC proliferation and
663 osteogenic differentiation to be responsible for the observed effects [249]. Despite the here reported
664 beneficial impacts of AgNPs hard-and soft-tissue related cells, further studies will have to elucidate
665 the clinical practicability relevance of AgNPs application in promotion of osteogenesis.

666

667 3.12 Strontium (Sr^{2+})

668 Strontium (Sr) is an alkaline earth metal and belongs to the group 2 elements of the periodic table.
669 Although it is considered as a non-essential element there is growing interest concerning the effects
670 of Sr on cells of the bone. This interest is based upon the fact that strontium ranelate is used in Europe
671 as a therapeutic drug for treatment of osteoporosis since 2004. Osteoporosis is a serious systemic
672 skeletal disorder, and is becoming a major health problem due to rapid population aging. As
673 osteoporosis leads to dramatic changes of the skeleton in terms of markedly decreased bone mass
674 and reduced bone quality as well as altered architecture on the macroscopic and microscopic level
675 the disease is associated with high incidence of osteoporotic fractures.

676 The use of Sr for the treatment of osteoporosis is based upon its dual mode of action: Sr influences
677 both, osteoblasts and osteoclast, and gives rise to increased bone formation capacity of osteoblasts,
678 and decreased bone resorption activity of osteoclasts [250-254]. Due to its similarity with calcium, the
679 effects of Sr are mediated in large part by the calcium sensing receptor (CaSR) which is a membrane-
680 bound receptor expressed in osteoblasts and osteoclasts [255-258]. In response to Sr, intracellular
681 signaling pathways are activated resulting in enhanced proliferation and differentiation of
682 mesenchymal stem cells and osteoblasts along with increased mineralization and deposition of
683 extracellular matrix [250, 255, 259] – at least by activating the Wnt/Catenin signal pathway [250, 260].
684 Additionally, in response of activating this pathway, OPG (osteoprotegerin) levels of osteoblasts and
685 their precursors increase whereas RANKL (receptor activator of nuclear factor κ B ligand) expression
686 of the cells decreases [261]. The expression patterns in favor of OPG suppress differentiation of
687 osteoclasts and limit the extent of bone resorption. Similar effects are observable in the course of
688 direct interaction of Sr with the extracellular domain of the CaSR: downstream cascades stimulate
689 diacylglycerol (DAG)-protein kinase C (PKC) β II which in turn induces osteoclast apoptosis [257]. In
690 a recent in vitro study Sr could be detected by means of mass spectrometry within the cytoplasm of
691 osteoclasts which were cultivated in combination with a Sr-enriched calcium phosphate cement. Cell
692 differentiation of the osteoclasts was obviously delayed [262]. However, the mechanism by which the
693 ions enter the cells, and to what extent intracellular Sr deposition influences cell signaling must still
694 be clarified.

695 Beside the beneficial effects on bone metabolism, systemic administration of strontium ranelate
696 increases the risk of cardiovascular diseases [263]. Therefore, its use is restricted to patients who show
697 no signs of heart and circulatory diseases.

698 For the benefit of osteoporotic patients and in the light of the effects of Sr on bone remodeling,
699 combination of Sr with bone substitutes might represent a successful approach to overcome the
700 adverse effects of systemic administration of strontium ranelate. Accordingly, Sr is used for apatite
701 coatings of orthopedic and dental implants [264-266], and is incorporated into different bone cements
702 [262, 267-273]. Because of their subsequent substitution by natural bone in the course of physiological

703 remodeling, it has been proposed that calcium phosphate-based cements ensure the local release of
704 Sr [274], and therefore might represent ideal bone substitutes for the osteoporotic bone. According to
705 this suggestion, stable incorporation of Sr into the crystal lattice of the bone mineral is based upon
706 remodeling activities of osteoblasts and osteoclasts (for a review see [275]), and Sr uptake is especially
707 high in newly formed bone tissue [276]. So placed at the disposal of the bone cells, Sr might locally
708 regulate their activities as well as the bone healing process in the course of further remodeling.

709

710 3.13 Vanadium (V^{+})

711 Vanadium is a trace element present in basically all living organisms and is predominantly stored
712 within the bone tissue [277]. Because of its growth factor mimicking properties, it was previously
713 suggested that vanadium might positively influence osteogenesis [278, 279]. An early study
714 analyzing the impact of vanadium derivatives on osteoblast-like UMR106 cells reported enhanced
715 proliferation, alkaline phosphatase activity and even differentiation [280]. As insulin
716 supplementation ameliorates negative effects of diabetes on bone regeneration and local insulin
717 treatment enhances fracture healing in healthy rats, the insulin-mimetic properties of vanadium are
718 currently being investigated as safe and cost-efficient alternative to insulin supplementation [281,
719 282].

720 Intramedullary delivery of an organic vanadium salt (vanadyl acetylacetonate) in a rat femoral
721 fracture model significantly promoted cell proliferation, vascular endothelial growth, callus cartilage
722 formation and mineralization and considerably increased torque to failure compared to treatment
723 with saline control solutions [236]. A vanadium-loaded collagen scaffold was recently described by
724 Cortizo and colleagues; although vanadium loading increased membrane permeability, no changes
725 in the collagen structure were observed. Furthermore, attachment, growth and osteoblastic as well as
726 chondrocytic differentiation of rBMPCs was improved by loading vanadyl acetylacetonate onto
727 collagen membranes [283]. Vanadium coating of titanium implants was also shown to enhance
728 fibroblast attachment and proliferation, which suggests potential benefits in soft tissue healing by
729 vanadium treatment [284]. Taken together, published data demonstrates vanadium to be an
730 interesting metal with great potential in regulating both angiogenesis and osteogenesis, however,
731 further studies are required to support these preliminary findings.

732

733 3.14 Zinc (Zn^{2+})

734 Zinc is an essential trace element which is pivotal for proper immune system functioning, cell division
735 and for skeletal development and therefore has been implemented into biomaterials for orthopedic
736 and dental applications [285-287]. Further, zinc and zinc alloys are promising biomaterials as load-
737 bearing scaffolds as they hold similar mechanical properties like mammalian bone. Especially Zn^{2+} ions
738 have a multitude of physiological functions. Zinc led to increased ECM mineralization in hMSC
739 culture by promoting the expression of ALP and osteopontin [288]. Also for SMCs a concentration-
740 dependent behavior was found in presence of Zn^{2+} *in vitro*. In the range 80-120 μM a change in
741 biological response was observed by inhibition of viability and proliferation [289]. When Zn was used
742 in different titan coatings the measured expression of Zn-transporters (ZnT1 and ZIP1) suggests that
743 cells prefer Zn^{2+} present at the biomaterial interface rather than plain diffusion of Zn^{2+} ions in the
744 surrounding medium [290]. Additional studies on the actions of zinc supplementation in
745 osteogenesis reported enhanced collagen deposition and mineralization of osteoblast like MC3T3-E1
746 cells, antagonizing effects on osteoclastogenesis with simultaneous promotion of osteoblastogenic
747 differentiation and increased osteoblast activity mediated by zinc supplementation in a
748 concentration-dependent manner [291-293]. Zinc phosphate-loaded barrier membranes showed
749 excellent anti-microbial properties, capable of inhibiting bacterial colonization upon membrane
750 exposure and avoiding potential infections [294]. To further analyze beneficial properties of zinc in
751 GBR procedures, cross-linked gelatin membranes loaded with zinc hydroxyapatite powder were

752 compared to cross-linked collagen membranes in a rat calvarial defect model. After a period of 6
753 weeks, bone defect fill was $80 \pm 2\%$, $60 \pm 5\%$ and $40 \pm 2\%$ for the zinc-loaded gelatin membrane, the
754 collagen membrane and the unfilled control group, respectively, demonstrating the tremendous
755 potential for the application of zinc in bone regeneration approaches [295]. Antibacterial effects,
756 excellent biocompatibility and stimulatory impact on the activity of osteoblast-like MG63 cells was
757 also recently reported for nanocomposites of carboxylated graphene oxide sheets decorated with zinc
758 oxide nanoparticles, emphasizing the potential application of zinc in nanoparticle formulations for
759 tissue engineering [296].

760 Zinc ions released from zinc-doped tricalcium phosphates were able to enhance TRAP and ALP
761 activity of hBMSCs and to regulate multinuclear giant cell formation and activity of RAW264.7
762 macrophages [297]. De novo bone formation in a canine ectopic implantation model was only
763 induced by the addition of zinc to TCPs, however, not by TCPs alone, whereby the rate of new bone
764 formation was coherent with zinc concentration [297]. Zinc is also an attractive candidate for the
765 development of coatings in order to promote the integration of implants. Regarding this matter, a
766 study analyzed rBMSC activity in response to zinc-loaded titanium oxide coatings and the impact of
767 zinc-supplementation on osseointegration in a rat implantation model. In comparison to TiO₂
768 coatings without zinc, osteogenic gene expression was upregulated in rBMSCs cultivated on zinc-
769 doped TiO₂ coatings and early-stage new bone formation as well as bone contact ratio were increased
770 *in vivo* [290]. Yu and colleagues further reported increased osteogenic differentiation and
771 mineralized matrix deposition in rat bone marrow-derived pericytes (BM-PCs) and significant
772 promotion of new bone formation around titanium implants in osteopenic rabbits with the
773 application of zinc-modified calcium silicate coatings. Molecular analysis revealed that zinc exerts
774 these actions by regulating the TGF- β /Smad signaling pathway, which is pivotal for
775 osteoblastogenesis [298]. Reports about zinc in biomedical applications for tissue engineering,
776 especially with regards to the positive impacts on osteoblastogenesis, osteoblast activity and tissue
777 mineralization, are promising for improving implant osseointegration, accelerating bone
778 regeneration and inhibiting biofilm formation.

779

780 3.15 Others

781 There are other metals and their corresponding ions which have been demonstrated to have an effect
782 onto bone regeneration process [195]. Webster et al. have shown a higher adsorption of calcium,
783 vitronectin and collagen on yttrium-doped HAp [299]. Further, zirconium and also molybdenum are
784 used in different metal alloys which are used for orthopedic and dental applications [300]. The latter
785 metals are primarily used to achieve specific material properties. There are additional metals which
786 play a role as implant material especially titanium which builds up a very stable oxide layer, and
787 thus, can be considered almost inert in physiological conditions [301]. Nevertheless, for titanium and
788 its alloys it was shown that released titanium enhanced the release of bone resorbing cytokines from
789 LPS-stimulated monocyte cultures [302]. Long-term *in vivo* studies in baboons revealed an increased
790 titanium ion concentration in urine as well as enhanced levels in tissues [303]. Nevertheless, no toxic
791 effects were observed up to 8 years of implantation.

792 4. Conclusions

793 The existing bone substitute materials only provide osteoconductive healing capacities and most
794 of the newly developed tissue engineering strategies are still not applicable in the daily clinical
795 routine. The presented overview of the physiological mode of action of different metal ions and their
796 influence on the process of bone tissue regeneration has shown that their addition to existing bone
797 substitute materials may to alter different issues like inflammation and foreign body response or the
798 onset of bone regeneration as well as material durability. Another important problem is the
799 availability and the cost of suitable bone grafting material for the increasing need of an aging
800 population.

801 It is obvious that different parameters play an important role for the use or the combination of metals
802 with existing biomaterials. Further, it has been demonstrated that the concentration of the released
803 metal ions plays a crucial role for the bone formation process. Thereby, it would be beneficial to have
804 the ions present in close vicinity of the implanted biomaterial as bone regeneration should preferably
805 occur directly at the implant site. On the other hand metals can be incorporated to scaffolds which
806 support a continuous release to support early induction of osteoblast differentiation as they can
807 control transcriptional regulators like Runx2 and therefore osteogenesis.
808 There is still ongoing work investigating specific effects as well as possible synergistic effects of metal
809 ions with other synthetic materials on the differentiation into osteogenic lineage. Therefore, it is
810 necessary to plan and run additional experiments and studies in almost every scientific field to
811 develop the suitable biomaterial for the patients need.

812

813 **Author Contributions:** K. G., P. D., A. K., N. M., S. R., J. F., O. J., R. S., S. W. and M. B. wrote the paper. R.S., S.W.
814 and M.B. contributed materials.
815

816 **Conflicts of Interest:** The authors declare no conflict of interest.

817 References

- 818 1. Sakkas, A.; Wilde, F.; Heufelder, M.; Winter, K.; Schramm, A., Autogenous bone grafts in oral
819 implantology—is it still a “gold standard”? A consecutive review of 279 patients with 456
820 clinical procedures. *International journal of implant dentistry* **2017**, *3*, (1), 23. doi:
821 10.1186/s40729-017-0084-4
- 822 2. Sandberg, O. H.; Aspenberg, P., Inter-trabecular bone formation: a specific mechanism for
823 healing of cancellous bone: A narrative review. *Acta orthopaedica* **2016**, *87*, (5), 459-465. DOI:
824 10.1080/17453674.2016.1205172
- 825 3. Garbuz, D. S.; Masri, B. A.; Czitrom, A. A., Biology of allografting. *Orthopedic Clinics* **1998**, *29*,
826 (2), 199-204. DOI: [https://doi.org/10.1016/S0030-5898\(05\)70318-7](https://doi.org/10.1016/S0030-5898(05)70318-7)
- 827 4. Eagan, M. J.; McAllister, D. R., Biology of allograft incorporation. *Clinics in sports medicine*
828 **2009**, *28*, (2), 203-214. DOI: <https://doi.org/10.1016/j.csm.2008.10.009>
- 829 5. Dimitriou, R.; Mataliotakis, G. I.; Angoules, A. G.; Kanakaris, N. K.; Giannoudis, P. V.,
830 Complications following autologous bone graft harvesting from the iliac crest and using the
831 RIA: a systematic review. *Injury* **2011**, *42*, S3-S15. doi: 10.1016/j.injury.2011.06.015
- 832 6. Mano, J. F.; Sousa, R. A.; Boesel, L. F.; Neves, N. M.; Reis, R. L., Bioinert, biodegradable and
833 injectable polymeric matrix composites for hard tissue replacement: state of the art and recent
834 developments. *Composites Science and Technology* **2004**, *64*, (6), 789-817.
835 <https://doi.org/10.1016/j.compscitech.2003.09.001>
- 836 7. Kuboyama, N.; Kiba, H.; Arai, K.; Uchida, R.; Tanimoto, Y.; Bhawal, U. K.; Abiko, Y.;
837 Miyamoto, S.; Knight, D.; Asakura, T., Silk fibroin-based scaffolds for bone regeneration.
838 *Journal of Biomedical Materials Research Part B: Applied Biomaterials* **2013**, *101*, (2), 295-302. DOI:
839 10.1002/jbm.b.32839
- 840 8. Jung, S. W.; Byun, J.-H.; Oh, S. H.; Kim, T. H.; Park, J.-S.; Rho, G.-J.; Lee, J. H., Multivalent
841 ion-based in situ gelling polysaccharide hydrogel as an injectable bone graft. *Carbohydrate*
842 *polymers* **2018**, *180*, 216-225. DOI: <https://doi.org/10.1016/j.carbpol.2017.10.029>
- 843 9. Dard, M.; Larjava, H., Hydroxyapatite/beta-tricalcium phosphate biphasic ceramics as
844 regenerative material for the repair of complex bone defects. *Journal of Biomedical Materials*
845 *Research Part B: Applied Biomaterials* **2017**. DOI: 10.1002/jbm.b.34049
- 846 10. Basha, R. Y.; Doble, M., Design of biocomposite materials for bone tissue regeneration.
847 *Materials Science and Engineering: C* **2015**, *57*, 452-463.
848 <https://doi.org/10.1016/j.msec.2015.07.016>
- 849 11. Gentile, P.; Chiono, V.; Carmagnola, I.; Hatton, P. V., An overview of poly (lactic-co-glycolic)
850 acid (PLGA)-based biomaterials for bone tissue engineering. *International journal of molecular*
851 *sciences* **2014**, *15*, (3), 3640-3659. doi:10.3390/ijms15033640
- 852 12. Wang, J.; Wu, D.; Zhang, Z.; Li, J.; Shen, Y.; Wang, Z.; Li, Y.; Zhang, Z.-Y.; Sun, J.,
853 Biomimetically Ornamented Rapid Prototyping Fabrication of an Apatite–Collagen–
854 Polycaprolactone Composite Construct with Nano–Micro–Macro Hierarchical Structure for
855 Large Bone Defect Treatment. *ACS applied materials & interfaces* **2015**, *7*, (47), 26244-26256.
856 DOI: 10.1021/acsami.5b08534
- 857 13. Miron, R.; Zhang, Y., Osteoinduction: a review of old concepts with new standards. *Journal*
858 *of dental research* **2012**, *91*, (8), 736-744. DOI: [doi/pdf/10.1177/0022034511435260](https://doi.org/10.1177/0022034511435260)

- 859 14. Gong, T.; Xie, J.; Liao, J.; Zhang, T.; Lin, S.; Lin, Y., Nanomaterials and bone regeneration.
860 *Bone research* **2015**, *3*, 15029. doi:10.1038/boneres.2015.29
- 861 15. Wu, S.; Liu, X.; Yeung, K. W.; Liu, C.; Yang, X., Biomimetic porous scaffolds for bone tissue
862 engineering. *Materials Science and Engineering: R: Reports* **2014**, *80*, 1-36. DOI:
863 <https://doi.org/10.1016/j.mser.2014.04.001>
- 864 16. García-Gareta, E.; Coathup, M. J.; Blunn, G. W., Osteoinduction of bone grafting materials
865 for bone repair and regeneration. *Bone* **2015**, *81*, 112-121. DOI:
866 <https://doi.org/10.1016/j.bone.2015.07.007>
- 867 17. Bae, S. E.; Choi, J.; Joung, Y. K.; Park, K.; Han, D. K., Controlled release of bone
868 morphogenetic protein (BMP)-2 from nanocomplex incorporated on hydroxyapatite-formed
869 titanium surface. *Journal of controlled release* **2012**, *160*, (3), 676-684. DOI:
870 <https://doi.org/10.1016/j.jconrel.2012.04.021>
- 871 18. Wang, J.; Guo, J.; Liu, J.; Wei, L.; Wu, G., BMP-functionalised coatings to promote
872 osteogenesis for orthopaedic implants. *International journal of molecular sciences* **2014**, *15*, (6),
873 10150-10168. doi:10.3390/ijms150610150
- 874 19. Fernandez-Yague, M. A.; Abbah, S. A.; McNamara, L.; Zeugolis, D. I.; Pandit, A.; Biggs, M.
875 J., Biomimetic approaches in bone tissue engineering: Integrating biological and
876 physicommechanical strategies. *Advanced drug delivery reviews* **2015**, *84*, 1-29. DOI:
877 <https://doi.org/10.1016/j.addr.2014.09.005>
- 878 20. Faruq, O.; Kim, B.; Padalhin, A. R.; Lee, G. H.; Lee, B.-T., A hybrid composite system of
879 biphasic calcium phosphate granules loaded with hyaluronic acid–gelatin hydrogel for bone
880 regeneration. *Journal of biomaterials applications* **2017**, *32*, (4), 433-445. DOI:
881 <https://doi.org/10.1177/0885328217730680>
- 882 21. Tanaka, K.; Goto, T.; Miyazaki, T.; Morita, Y.; Kobayashi, S.; Takahashi, T., Apatite-coated
883 hyaluronan for bone regeneration. *Journal of dental research* **2011**, *90*, (7), 906-911. DOI:
884 <https://doi.org/10.1177/0022034511404070>
- 885 22. Kuttappan, S.; Mathew, D.; Nair, M. B., Biomimetic composite scaffolds containing
886 bioceramics and collagen/gelatin for bone tissue engineering-A mini review. *International*
887 *journal of biological macromolecules* **2016**, *93*, 1390-1401.
888 DOI: <https://doi.org/10.1016/j.ijbiomac.2016.06.043>
- 889 23. Lyons, F. G.; Gleeson, J. P.; Partap, S.; Coghlan, K.; O'Brien, F. J., Novel microhydroxyapatite
890 particles in a collagen scaffold: a bioactive bone void filler? *Clinical Orthopaedics and Related*
891 *Research*® **2014**, *472*, (4), 1318-1328.
892 DOI: <https://doi.org/10.1007/s11999-013-3438-0>
- 893 24. Förster, Y.; Bernhardt, R.; Hintze, V.; Möller, S.; Schnabelrauch, M.; Scharnweber, D.;
894 Rammelt, S., Collagen/glycosaminoglycan coatings enhance new bone formation in a critical
895 size bone defect—A pilot study in rats. *Materials Science and Engineering: C* **2017**, *71*, 84-92.
896 DOI: <https://doi.org/10.1016/j.msec.2016.09.071>
- 897 25. Vo, T. N.; Kasper, F. K.; Mikos, A. G., Strategies for controlled delivery of growth factors and
898 cells for bone regeneration. *Advanced drug delivery reviews* **2012**, *64*, (12), 1292-1309. DOI:
899 <https://doi.org/10.1016/j.addr.2012.01.016>
- 900 26. Boerckel, J. D.; Kolambkar, Y. M.; Dupont, K. M.; Uhrig, B. A.; Phelps, E. A.; Stevens, H. Y.;
901 García, A. J.; Guldberg, R. E., Effects of protein dose and delivery system on BMP-mediated

- 902 bone regeneration. *Biomaterials* **2011**, *32*, (22), 5241-5251. DOI:
903 <https://doi.org/10.1016/j.biomaterials.2011.03.063>
- 904 27. Haidar, Z. S.; Hamdy, R. C.; Tabrizian, M., Delivery of recombinant bone morphogenetic
905 proteins for bone regeneration and repair. Part B: Delivery systems for BMPs in orthopaedic
906 and craniofacial tissue engineering. *Biotechnology letters* **2009**, *31*, (12), 1825-1835. DOI
907 <https://doi.org/10.1007/s10529-009-0100-8>
- 908 28. Wozney, J. M., The bone morphogenetic protein family and osteogenesis. *Molecular*
909 *reproduction and development* **1992**, *32*, (2), 160-167.
910 DOI: 10.1002/mrd.1080320212
- 911 29. Hänseler, P.; Ehrbar, M.; Kruse, A.; Fischer, E.; Schibli, R.; Ghayor, C.; Weber, F. E., Delivery
912 of BMP-2 by two clinically available apatite materials: In vitro and in vivo comparison.
913 *Journal of Biomedical Materials Research Part A* **2015**, *103*, (2), 628-638. DOI: 10.1002/jbm.a.35211
- 914 30. Florencio-Silva, R.; Sasso, G. R. d. S.; Sasso-Cerri, E.; Simões, M. J.; Cerri, P. S., Biology of bone
915 tissue: structure, function, and factors that influence bone cells. *BioMed research international*
916 **2015**, 2015.
917 DOI: <http://dx.doi.org/10.1155/2015/421746>
- 918 31. Diomedede, F.; Gugliandolo, A.; Scionti, D.; Merciaro, I.; Cavalcanti, M. F.; Mazzon, E.;
919 Trubiani, O., Biotherapeutic Effect of Gingival Stem Cells Conditioned Medium in Bone
920 Tissue Restoration. *International journal of molecular sciences* **2018**, *19*, (2), 329.
921 doi:10.3390/ijms19020329
- 922 32. Bianco, P.; Robey, P. G., Stem cells in tissue engineering. *Nature* **2001**, *414*, (6859), 118.
923 doi:10.1038/35102181
- 924 33. Kanczler, J.; Oreffo, R., Osteogenesis and angiogenesis: the potential for engineering bone.
925 *Eur Cell Mater* **2008**, *15*, (2), 100-114.
926 DOI: 10.22203/eCM.v015a08
- 927 34. Santos, M. I.; Unger, R. E.; Sousa, R. A.; Reis, R. L.; Kirkpatrick, C. J., Crosstalk between
928 osteoblasts and endothelial cells co-cultured on a polycaprolactone–starch scaffold and the
929 in vitro development of vascularization. *Biomaterials* **2009**, *30*, (26), 4407-4415. DOI:
930 <https://doi.org/10.1016/j.biomaterials.2009.05.004>
- 931 35. Du, C.; Cui, F.; Zhu, X.; De Groot, K., Three-dimensional nano-HAp/collagen matrix loading
932 with osteogenic cells in organ culture. *Journal of Biomedical Materials Research Part A* **1999**, *44*,
933 (4), 407-415.
934 DOI: 10.1002/(SICI)1097-4636(19990315)44:4<407::AID-JBM6>3.0.CO;2-T
- 935 36. Mistry, A. S.; Mikos, A. G., Tissue engineering strategies for bone regeneration. In
936 *Regenerative medicine II*, Springer: 2005; pp 1-22.
937 DOI: <https://doi.org/10.1007/b99997>
- 938 37. Alexander, K. A.; Chang, M. K.; Maylin, E. R.; Kohler, T.; Müller, R.; Wu, A. C.; Van Rooijen,
939 N.; Sweet, M. J.; Hume, D. A.; Raggatt, L. J., Osteal macrophages promote in vivo
940 intramembranous bone healing in a mouse tibial injury model. *Journal of bone and mineral*
941 *research* **2011**, *26*, (7), 1517-1532.
942 DOI: 10.1002/jbmr.354

- 943 38. Kim, Y.-H.; Furuya, H.; Tabata, Y., Enhancement of bone regeneration by dual release of a
944 macrophage recruitment agent and platelet-rich plasma from gelatin hydrogels. *Biomaterials*
945 **2014**, *35*, (1), 214-224.
946 DOI: <https://doi.org/10.1016/j.biomaterials.2013.09.103>
- 947 39. Yamada, Y.; Ueda, M.; Naiki, T.; Takahashi, M.; Hata, K.-I.; Nagasaka, T., Autogenous
948 injectable bone for regeneration with mesenchymal stem cells and platelet-rich plasma:
949 tissue-engineered bone regeneration. *Tissue engineering* **2004**, *10*, (5-6), 955-964. DOI:
950 <https://doi.org/10.1089/1076327041348284>
- 951 40. Weibrich, G.; Hansen, T.; Kleis, W.; Buch, R.; Hitzler, W., Effect of platelet concentration in
952 platelet-rich plasma on peri-implant bone regeneration. *Bone* **2004**, *34*, (4), 665-671. DOI:
953 <https://doi.org/10.1016/j.bone.2003.12.010>
- 954 41. Simonpieri, A.; Del Corso, M.; Vervelle, A.; Jimbo, R.; Inchingolo, F.; Sammartino, G.; M
955 Dohan Ehrenfest, D., Current knowledge and perspectives for the use of platelet-rich plasma
956 (PRP) and platelet-rich fibrin (PRF) in oral and maxillofacial surgery part 2: Bone graft,
957 implant and reconstructive surgery. *Current pharmaceutical biotechnology* **2012**, *13*, (7), 1231-
958 1256.
959 DOI: <https://doi.org/10.2174/138920112800624472>
- 960 42. Civinini, R.; Macera, A.; Redl, B.; Innocenti, M., Blood-derived growth factors. *Clinical Cases*
961 *in Mineral and Bone Metabolism* **2010**, *7*, (3), 194.
- 962 43. Barrère, F.; van Blitterswijk, C. A.; de Groot, K., Bone regeneration: molecular and cellular
963 interactions with calcium phosphate ceramics. *International journal of nanomedicine* **2006**, *1*, (3),
964 317.
- 965 44. Inzana, J. A.; Olvera, D.; Fuller, S. M.; Kelly, J. P.; Graeve, O. A.; Schwarz, E. M.; Kates, S. L.;
966 Awad, H. A., 3D printing of composite calcium phosphate and collagen scaffolds for bone
967 regeneration. *Biomaterials* **2014**, *35*, (13), 4026-4034. DOI:
968 <https://doi.org/10.1016/j.biomaterials.2014.01.064>
- 969 45. Bernhardt, A.; Schamel, M.; Gbureck, U.; Gelinsky, M., Osteoclastic differentiation and
970 resorption is modulated by bioactive metal ions Co^{2+} , Cu^{2+} and Cr^{3+} incorporated into
971 calcium phosphate bone cements. *PLoS one* **2017**, *12*, (8), e0182109. DOI:
972 <https://doi.org/10.1371/journal.pone.0182109>
- 973 46. Philippart, A.; Gómez-Cerezo, N.; Arcos, D.; Salinas, A. J.; Boccardi, E.; Vallet-Regi, M.;
974 Boccaccini, A. R., Novel ion-doped mesoporous glasses for bone tissue engineering: Study of
975 their structural characteristics influenced by the presence of phosphorous oxide. *Journal of*
976 *Non-Crystalline Solids* **2017**, *455*, 90-97. DOI: <https://doi.org/10.1016/j.jnoncrysol.2016.10.031>
- 977 47. Cattalini, J.; Hoppe, A.; Pishbin, F.; Roether, J.; Boccaccini, A.; Lucangioli, S.; Mouriño, V.,
978 Novel nanocomposite biomaterials with controlled copper/calcium release capability for
979 bone tissue engineering multifunctional scaffolds. *Journal of The Royal Society Interface* **2015**,
980 *12*, (110), 20150509.
981 DOI: <http://dx.doi.org/10.1098/rsif.2015.0509>
- 982 48. Xia, W.; Grandfield, K.; Schwenke, A.; Engqvist, H., Synthesis and release of trace elements
983 from hollow and porous hydroxyapatite spheres. *Nanotechnology* **2011**, *22*, (30), 305610.
984 DOI: <https://doi.org/10.1088/0957-4484/22/30/305610>

- 985 49. Maes, C.; Kobayashi, T.; Selig, M. K.; Torrekens, S.; Roth, S. I.; Mackem, S.; Carmeliet, G.;
986 Kronenberg, H. M., Osteoblast precursors, but not mature osteoblasts, move into developing
987 and fractured bones along with invading blood vessels. *Developmental cell* **2010**, *19*, (2), 329-
988 344.
989 DOI: <https://doi.org/10.1016/j.devcel.2010.07.010>
- 990 50. Mastrogiacomo, M.; Scaglione, S.; Martinetti, R.; Dolcini, L.; Beltrame, F.; Cancedda, R.;
991 Quarto, R., Role of scaffold internal structure on in vivo bone formation in macroporous
992 calcium phosphate bioceramics. *Biomaterials* **2006**, *27*, (17), 3230-3237. DOI:
993 <https://doi.org/10.1016/j.biomaterials.2006.01.031>
- 994 51. Woodard, J. R.; Hilldore, A. J.; Lan, S. K.; Park, C.; Morgan, A. W.; Eurell, J. A. C.; Clark, S.
995 G.; Wheeler, M. B.; Jamison, R. D.; Johnson, A. J. W., The mechanical properties and
996 osteoconductivity of hydroxyapatite bone scaffolds with multi-scale porosity. *Biomaterials*
997 **2007**, *28*, (1), 45-54.
998 DOI: <https://doi.org/10.1016/j.biomaterials.2006.08.021>
- 999 52. Blair, H. C.; Larrouture, Q. C.; Li, Y.; Lin, H.; Beer-Stoltz, D.; Liu, L.; Tuan, R. S.; Robinson, L.
1000 J.; Schlesinger, P. H.; Nelson, D. J., Osteoblast differentiation and bone matrix formation in
1001 vivo and in vitro. *Tissue Engineering Part B: Reviews* **2017**, *23*, (3), 268-280. DOI:
1002 <https://doi.org/10.1089/ten.teb.2016.0454>
- 1003 53. Boyan, B.; Bonewald, L.; Paschalis, E.; Lohmann, C.; Rosser, J.; Cochran, D.; Dean, D.;
1004 Schwartz, Z.; Boskey, A., Osteoblast-mediated mineral deposition in culture is dependent on
1005 surface microtopography. *Calcified tissue international* **2002**, *71*, (6), 519-529. DOI:
1006 <https://doi.org/10.1007/s00223-001-1114-y>
- 1007 54. Galli, S.; Stocchero, M.; Andersson, M.; Karlsson, J.; He, W.; Lilin, T.; Wennerberg, A.; Jimbo,
1008 R., The effect of magnesium on early osseointegration in osteoporotic bone: a histological and
1009 gene expression investigation. *Osteoporosis International* **2017**, *28*, (7), 2195-2205. DOI:
1010 <https://doi.org/10.1007/s00198-017-4004-5>
- 1011 55. Park, J. W.; Kim, Y. J.; Jang, J. H.; Song, H., Osteoblast response to magnesium ion-
1012 incorporated nanoporous titanium oxide surfaces. *Clinical oral implants research* **2010**, *21*, (11),
1013 1278-1287.
1014 DOI: [10.1111/j.1600-0501.2010.01944.x](https://doi.org/10.1111/j.1600-0501.2010.01944.x)
- 1015 56. Arai, F.; Miyamoto, T.; Ohneda, O.; Inada, T.; Sudo, T.; Brasel, K.; Miyata, T.; Anderson, D.
1016 M.; Suda, T., Commitment and differentiation of osteoclast precursor cells by the sequential
1017 expression of c-Fms and receptor activator of nuclear factor κ B (RANK) receptors. *Journal of*
1018 *Experimental Medicine* **1999**, *190*, (12), 1741-1754. DOI: [10.1084/jem.190.12.1741](https://doi.org/10.1084/jem.190.12.1741)
- 1019 57. Martin, T. J.; Sims, N. A., Osteoclast-derived activity in the coupling of bone formation to
1020 resorption. *Trends in molecular medicine* **2005**, *11*, (2), 76-81.
1021 DOI: <https://doi.org/10.1016/j.molmed.2004.12.004>
- 1022 58. Tanaka, Y.; Nakayamada, S.; Okada, Y., Osteoblasts and osteoclasts in bone remodeling and
1023 inflammation. *Current Drug Targets-Inflammation & Allergy* **2005**, *4*, (3), 325-328. DOI:
1024 <https://doi.org/10.2174/1568010054022015>
- 1025 59. Wong, B. R.; Josien, R.; Lee, S. Y.; Sauter, B.; Li, H.-L.; Steinman, R. M.; Choi, Y., TRANCE
1026 (tumor necrosis factor [TNF]-related activation-induced cytokine), a new TNF family

- 1027 member predominantly expressed in T cells, is a dendritic cell-specific survival factor.
1028 *Journal of Experimental Medicine* **1997**, 186, (12), 2075-2080.
- 1029 60. Hirano, T., Interleukin 6 and its receptor: ten years later. *International reviews of immunology*
1030 **1998**, 16, (3-4), 249-284. DOI: 10.3109/08830189809042997
- 1031 61. Quinn, J. M.; Gillespie, M. T., Modulation of osteoclast formation. *Biochemical and biophysical*
1032 *research communications* **2005**, 328, (3), 739-745.
1033 DOI: <https://doi.org/10.1016/j.bbrc.2004.11.076>
- 1034 62. Saran, U.; Piperni, S. G.; Chatterjee, S., Role of angiogenesis in bone repair. *Archives of*
1035 *biochemistry and biophysics* **2014**, 561, 109-117.
1036 DOI: <https://doi.org/10.1016/j.abb.2014.07.006>
- 1037 63. Wang, Y.; Wan, C.; Deng, L.; Liu, X.; Cao, X.; Gilbert, S. R.; Boussein, M. L.; Faugere, M.-C.;
1038 Guldberg, R. E.; Gerstenfeld, L. C., The hypoxia-inducible factor α pathway couples
1039 angiogenesis to osteogenesis during skeletal development. *The Journal of clinical investigation*
1040 **2007**, 117, (6), 1616-1626. doi:10.1172/JCI31581
- 1041 64. Schindeler, A.; McDonald, M. M.; Bokko, P.; Little, D. G. In *Bone remodeling during fracture*
1042 *repair: the cellular picture*, Seminars in cell & developmental biology, 2008; Elsevier: 2008; pp
1043 459-466.
1044 DOI: <https://doi.org/10.1016/j.semcd.2008.07.004>
- 1045 65. Batoon, L.; Millard, S. M.; Raggatt, L. J.; Pettit, A. R., Osteomacs and Bone Regeneration.
1046 *Current osteoporosis reports* **2017**, 15, (4), 385-395.
1047 DOI: <https://doi.org/10.1007/s11914-017-0384-x>
- 1048 66. Mise-Omata, S.; Alles, N.; Fukazawa, T.; Aoki, K.; Ohya, K.; Jimi, E.; Obata, Y.; Doi, T., NF-
1049 κ B RELA-deficient bone marrow macrophages fail to support bone formation and to
1050 maintain the hematopoietic niche after lethal irradiation and stem cell transplantation.
1051 *International immunology* **2014**, 26, (11), 607-618.
1052 DOI: <https://doi.org/10.1093/intimm/dxu062>
- 1053 67. Chang, K. H.; Sengupta, A.; Nayak, R. C.; Duran, A.; Lee, S. J.; Pratt, R. G.; Wellendorf, A. M.;
1054 Hill, S. E.; Watkins, M.; Gonzalez-Nieto, D., p62 is required for stem cell/progenitor retention
1055 through inhibition of IKK/NF- κ B/Ccl4 signaling at the bone marrow macrophage-osteoblast
1056 niche. *Cell reports* **2014**, 9, (6), 2084-2097. DOI: <https://doi.org/10.1016/j.celrep.2014.11.031>
- 1057 68. Coleman, D.; King, R.; Andrade, J., The foreign body reaction: a chronic inflammatory
1058 response. *Journal of Biomedical Materials Research Part A* **1974**, 8, (5), 199-211. DOI:
1059 10.1002/jbm.820080503
- 1060 69. Loi, F.; Córdova, L. A.; Pajarinen, J.; Lin, T.-h.; Yao, Z.; Goodman, S. B., Inflammation, fracture
1061 and bone repair. *Bone* **2016**, 86, 119-130.
1062 DOI: <https://doi.org/10.1016/j.bone.2016.02.020>
- 1063 70. Ghanaati, S.; Barbeck, M.; Detsch, R.; Deisinger, U.; Hilbig, U.; Rausch, V.; Sader, R.; Unger,
1064 R. E.; Ziegler, G.; Kirkpatrick, C. J., The chemical composition of synthetic bone substitutes
1065 influences tissue reactions in vivo: histological and histomorphometrical analysis of the
1066 cellular inflammatory response to hydroxyapatite, beta-tricalcium phosphate and biphasic
1067 calcium phosphate ceramics. *Biomedical materials* **2012**, 7, (1), 015005.
1068 DOI: <https://doi.org/10.1088/1748-6041/7/1/015005>

- 1069 71. Trindade, R.; Albrektsson, T.; Tengvall, P.; Wennerberg, A., Foreign body reaction to
1070 biomaterials: on mechanisms for buildup and breakdown of osseointegration. *Clinical implant*
1071 *dentistry and related research* **2016**, 18, (1), 192-203.
1072 DOI: 10.1111/cid.12274
- 1073 72. Martin, P.; Leibovich, S. J., Inflammatory cells during wound repair: the good, the bad and
1074 the ugly. *Trends in cell biology* **2005**, 15, (11), 599-607.
1075 DOI: <https://doi.org/10.1016/j.tcb.2005.09.002>
- 1076 73. Mantovani, A.; Sica, A.; Sozzani, S.; Allavena, P.; Vecchi, A.; Locati, M., The chemokine
1077 system in diverse forms of macrophage activation and polarization. *Trends in immunology*
1078 **2004**, 25, (12), 677-686.
1079 DOI: <https://doi.org/10.1016/j.it.2004.09.015>
- 1080 74. Barbeck, M.; Booms, P.; Unger, R.; Hoffmann, V.; Sader, R.; Kirkpatrick, C. J.; Ghanaati, S.,
1081 Multinucleated giant cells in the implant bed of bone substitutes are foreign body giant
1082 cells—New insights into the material-mediated healing process. *Journal of Biomedical Materials*
1083 *Research Part A* **2017**, 105, (4), 1105-1111.
1084 DOI: 10.1002/jbm.a.36006
- 1085 75. Anderson, J. M., Multinucleated giant cells. *Current opinion in hematology* **2000**, 7, (1), 40-47.
1086 DOI: 10.1097/00062752-200001000-00008
- 1087 76. Mountziaris, P. M.; Mikos, A. G., Modulation of the inflammatory response for enhanced
1088 bone tissue regeneration. *Tissue Engineering Part B: Reviews* **2008**, 14, (2), 179-186. DOI:
1089 <https://doi.org/10.1089/ten.teb.2008.0038>
- 1090 77. Sussman, E. M.; Halpin, M. C.; Muster, J.; Moon, R. T.; Ratner, B. D., Porous implants
1091 modulate healing and induce shifts in local macrophage polarization in the foreign body
1092 reaction. *Annals of biomedical engineering* **2014**, 42, (7), 1508-1516.
1093 DOI: <https://doi.org/10.1007/s10439-013-0933-0>
- 1094 78. Ahmadzadeh, E.; Talebnia, F.; Tabatabaei, M.; Ahmadzadeh, H.; Mostaghaci, B.,
1095 Osteoconductive composite graft based on bacterial synthesized hydroxyapatite
1096 nanoparticles doped with different ions: From synthesis to in vivo studies. *Nanomedicine:*
1097 *Nanotechnology, Biology and Medicine* **2016**, 12, (5), 1387-1395.
1098 DOI: <https://doi.org/10.1016/j.nano.2016.01.020>
- 1099 79. Exley, C., Human exposure to aluminium. *Environmental Science: Processes & Impacts* **2013**, 15,
1100 (10), 1807-1816. DOI: 10.1039/C3EM00374D
- 1101 80. Priyadarshi, A.; Shapiro, J. I. In *Hematology: Issues in the dialysis patient: Erythropoietin resistance*
1102 *in the treatment of the anemia of chronic renal failure*, Seminars in dialysis, 2006; Wiley Online
1103 Library: 2006; pp 273-278.
1104 DOI: 10.1111/j.1525-139X.2006.00172.x
- 1105 81. Kawahara, M., Effects of aluminum on the nervous system and its possible link with
1106 neurodegenerative diseases. *Journal of Alzheimer's Disease* **2005**, 8, (2), 171-182.
1107 DOI: 10.3233/JAD-2005-8210
- 1108 82. Aaseth, J.; Boivin, G.; Andersen, O., Osteoporosis and trace elements—an overview. *Journal of*
1109 *Trace Elements in Medicine and Biology* **2012**, 26, (2-3), 149-152.
1110 DOI: <https://doi.org/10.1016/j.jtemb.2012.03.017>

- 1111 83. Lukiw, W. J.; Percy, M. E.; Kruck, T. P., Nanomolar aluminum induces pro-inflammatory and
1112 pro-apoptotic gene expression in human brain cells in primary culture. *Journal of inorganic*
1113 *biochemistry* **2005**, *99*, (9), 1895-1898.
1114 DOI: <https://doi.org/10.1016/j.jinorgbio.2005.04.021>
- 1115 84. Gomez-Alonso, C.; Menendez-Rodriguez, P.; Virgos-Soriano, M.; Fernandez-Martin, J.;
1116 Fernandez-Coto, M.; Cannata-Andia, J., Aluminum-induced osteogenesis in osteopenic rats
1117 with normal renal function. *Calcified tissue international* **1999**, *64*, (6), 534-541. DOI:
1118 <https://doi.org/10.1007/s002239900645>
- 1119 85. Quarles, L.; Gitelman, H.; Drezner, M., Induction of de novo bone formation in the beagle. A
1120 novel effect of aluminum. *The Journal of clinical investigation* **1988**, *81*, (4), 1056-1066.
1121 doi:10.1172/JCI113417
- 1122 86. Li, X.; Han, Y.; Guan, Y.; Zhang, L.; Bai, C.; Li, Y., Aluminum induces osteoblast apoptosis
1123 through the oxidative stress-mediated JNK signaling pathway. *Biological trace element research*
1124 **2012**, *150*, (1-3), 502-508.
1125 DOI: <https://doi.org/10.1007/s12011-012-9523-5>
- 1126 87. Kidder, L. S.; Klein, G. L.; Gundberg, C. M.; Seitz, P. K.; Rubin, N. H.; Simmons, D. J., Effects
1127 of aluminum on rat bone cell populations. *Calcified tissue international* **1993**, *53*, (5), 357-361.
1128 DOI <https://doi.org/10.1007/BF01351843>
- 1129 88. Li, X.; Hu, C.; Zhu, Y.; Sun, H.; Li, Y.; Zhang, Z., Effects of aluminum exposure on bone
1130 mineral density, mineral, and trace elements in rats. *Biological trace element research* **2011**, *143*,
1131 (1), 378-385.
1132 DOI: <https://doi.org/10.1007/s12011-010-8861-4>
- 1133 89. Piconi, C.; Condo, S. G.; Kosmač, T., Alumina-and zirconia-based ceramics for load-
1134 bearing applications. In *Advanced ceramics for dentistry*, Elsevier: 2014; pp 219-253. DOI:
1135 <https://doi.org/10.1016/B978-0-12-394619-5.00011-0>
- 1136 90. Zhang, J.; Liu, W.; Schnitzler, V.; Tancret, F.; Bouler, J.-M., Calcium phosphate cements for
1137 bone substitution: chemistry, handling and mechanical properties. *Acta biomaterialia* **2014**, *10*,
1138 (3), 1035-1049.
1139 DOI: <https://doi.org/10.1016/j.actbio.2013.11.001>
- 1140 91. Prati, C.; Gandolfi, M. G., Calcium silicate bioactive cements: biological perspectives and
1141 clinical applications. *Dental materials* **2015**, *31*, (4), 351-370.
1142 DOI: <https://doi.org/10.1016/j.dental.2015.01.004>
- 1143 92. Eliaz, N.; Metoki, N., Calcium phosphate bioceramics: a review of their history, structure,
1144 properties, coating technologies and biomedical applications. *Materials* **2017**, *10*, (4), 334.
1145 doi:10.3390/ma10040334
- 1146 93. González-Vázquez, A.; Planell, J. A.; Engel, E., Extracellular calcium and CaSR drive
1147 osteoinduction in mesenchymal stromal cells. *Acta biomaterialia* **2014**, *10*, (6), 2824-2833. DOI:
1148 <https://doi.org/10.1016/j.actbio.2014.02.004>
- 1149 94. Silver, I.; Murrills, R.; Etherington, D., Microelectrode studies on the acid microenvironment
1150 beneath adherent macrophages and osteoclasts. *Experimental cell research* **1988**, *175*, (2), 266-
1151 276.
1152 DOI: [https://doi.org/10.1016/0014-4827\(88\)90191-7](https://doi.org/10.1016/0014-4827(88)90191-7)

- 1153 95. Maeno, S.; Niki, Y.; Matsumoto, H.; Morioka, H.; Yatabe, T.; Funayama, A.; Toyama, Y.;
1154 Taguchi, T.; Tanaka, J., The effect of calcium ion concentration on osteoblast viability,
1155 proliferation and differentiation in monolayer and 3D culture. *Biomaterials* **2005**, *26*, (23),
1156 4847-4855.
1157 DOI: <https://doi.org/10.1016/j.biomaterials.2005.01.006>
- 1158 96. Barradas, A. M.; Fernandes, H. A.; Groen, N.; Chai, Y. C.; Schrooten, J.; van de Peppel, J.; van
1159 Leeuwen, J. P.; van Blitterswijk, C. A.; de Boer, J., A calcium-induced signaling cascade
1160 leading to osteogenic differentiation of human bone marrow-derived mesenchymal stromal
1161 cells. *Biomaterials* **2012**, *33*, (11), 3205-3215. DOI:
1162 <https://doi.org/10.1016/j.biomaterials.2012.01.020>
- 1163 97. Wagner, A.-S.; Glenske, K.; Henß, A.; Kruppke, B.; Rößler, S.; Hanke, T.; Moritz, A.; Rohnke,
1164 M.; Kressin, M.; Arnhold, S., Cell behavior of human mesenchymal stromal cells in response
1165 to silica/collagen based xerogels and calcium deficient culture conditions. *Biomedical Materials*
1166 **2017**, *12*, (4), 045003.
1167 DOI: <https://doi.org/10.1088/1748-605X/aa6e29>
- 1168 98. Wagner, A.-S.; Glenske, K.; Wolf, V.; Fietz, D.; Mazurek, S.; Hanke, T.; Moritz, A.; Arnhold,
1169 S.; Wenisch, S., Osteogenic differentiation capacity of human mesenchymal stromal cells in
1170 response to extracellular calcium with special regard to connexin 43. *Annals of Anatomy-
1171 Anatomischer Anzeiger* **2017**, *209*, 18-24.
1172 DOI: <https://doi.org/10.1016/j.aanat.2016.09.005>
- 1173 99. Ye, J.; Ai, W.; Zhang, F.; Zhu, X.; Shu, G.; Wang, L.; Gao, P.; Xi, Q.; Zhang, Y.; Jiang, Q.,
1174 Enhanced proliferation of porcine bone marrow mesenchymal stem cells induced by
1175 extracellular calcium is associated with the activation of the calcium-sensing receptor and
1176 ERK signaling pathway. *Stem cells international* **2016**, 2016.
1177 DOI: <http://dx.doi.org/10.1155/2016/6570671>
- 1178 100. Huang, Z.; Cheng, S.-L.; Slatopolsky, E., Sustained activation of the extracellular signal-
1179 regulated kinase pathway is required for extracellular calcium stimulation of human
1180 osteoblast proliferation. *Journal of Biological Chemistry* **2001**, *276*, (24), 21351-21358. DOI
1181 10.1074/jbc.M010921200
- 1182 101. Dvorak, M. M.; Siddiqua, A.; Ward, D. T.; Carter, D. H.; Dallas, S. L.; Nemeth, E. F.; Riccardi,
1183 D., Physiological changes in extracellular calcium concentration directly control osteoblast
1184 function in the absence of calciotropic hormones. *Proceedings of the National Academy of
1185 Sciences of the United States of America* **2004**, *101*, (14), 5140-5145. DOI:
1186 <https://doi.org/10.1073/pnas.0306141101>
- 1187 102. Brown, E. M.; MacLeod, R. J., Extracellular calcium sensing and extracellular calcium
1188 signaling. *Physiological reviews* **2001**, *81*, (1), 239-297.
1189 DOI: <https://doi.org/10.1152/physrev.2001.81.1.239>
- 1190 103. Olszak, I. T.; Poznansky, M. C.; Evans, R. H.; Olson, D.; Kos, C.; Pollak, M. R.; Brown, E. M.;
1191 Scadden, D. T., Extracellular calcium elicits a chemokinetic response from monocytes in vitro
1192 and in vivo. *The Journal of clinical investigation* **2000**, *105*, (9), 1299-1305. doi:10.1172/JCI9799
- 1193 104. Glenske, K.; Wagner, A.-S.; Hanke, T.; Cavalcanti-Adam, E. A.; Heinemann, S.; Heinemann,
1194 C.; Kruppke, B.; Arnhold, S.; Moritz, A.; Schwab, E. H., Bioactivity of xerogels as modulators
1195 of osteoclastogenesis mediated by connexin 43. *Biomaterials* **2014**, *35*, (5), 1487-1495.

- 1196 DOI: <https://doi.org/10.1016/j.biomaterials.2013.11.002>
- 1197 105. Yamauchi, M.; Yamaguchi, T.; Kaji, H.; Sugimoto, T.; Chihara, K., Involvement of calcium-
- 1198 sensing receptor in osteoblastic differentiation of mouse MC3T3-E1 cells. *American Journal of*
- 1199 *Physiology-Endocrinology and Metabolism* **2005**, 288, (3), E608-E616. DOI:
- 1200 <https://doi.org/10.1152/ajpendo.00229.2004>
- 1201 106. Hu, F.; Pan, L.; Zhang, K.; Xing, F.; Wang, X.; Lee, I.; Zhang, X.; Xu, J., Elevation of
- 1202 extracellular Ca²⁺ induces store-operated calcium entry via calcium-sensing receptors: a
- 1203 pathway contributes to the proliferation of osteoblasts. *PLoS One* **2014**, 9, (9), e107217. DOI:
- 1204 <https://doi.org/10.1371/journal.pone.0107217>
- 1205 107. Zhang, W.; Liu, H. T., MAPK signal pathways in the regulation of cell proliferation in
- 1206 mammalian cells. *Cell research* **2002**, 12, (1), 9. doi:10.1038/sj.cr.7290105
- 1207 108. Parekh, A. B.; Putney Jr, J. W., Store-operated calcium channels. *Physiological reviews* **2005**, 85,
- 1208 (2), 757-810. DOI: <https://doi.org/10.1152/physrev.00057.2003>
- 1209 109. Wen, L.; Wang, Y.; Wang, H.; Kong, L.; Zhang, L.; Chen, X.; Ding, Y., L-type calcium channels
- 1210 play a crucial role in the proliferation and osteogenic differentiation of bone marrow
- 1211 mesenchymal stem cells. *Biochemical and biophysical research communications* **2012**, 424, (3), 439-
- 1212 445.
- 1213 DOI: <https://doi.org/10.1016/j.bbrc.2012.06.128>
- 1214 110. Heinemann, S.; Heinemann, C.; Wenisch, S.; Alt, V.; Worch, H.; Hanke, T., Calcium
- 1215 phosphate phases integrated in silica/collagen nanocomposite xerogels enhance the
- 1216 bioactivity and ultimately manipulate the osteoblast/osteoclast ratio in a human co-culture
- 1217 model. *Acta biomaterialia* **2013**, 9, (1), 4878-4888.
- 1218 DOI: <https://doi.org/10.1016/j.actbio.2012.10.010>
- 1219 111. Hoppe, A.; Guldal, N. S.; Boccaccini, A. R., A review of the biological response to ionic
- 1220 dissolution products from bioactive glasses and glass-ceramics. *Biomaterials* **2011**, 32, (11),
- 1221 2757-2774. DOI: <https://doi.org/10.1016/j.biomaterials.2011.01.004>
- 1222 112. Beck, G. R., Inorganic phosphate as a signaling molecule in osteoblast differentiation. *Journal*
- 1223 *of cellular biochemistry* **2003**, 90, (2), 234-243.
- 1224 DOI: 10.1002/jcb.10622
- 1225 113. Shih, Y.-R. V.; Hwang, Y.; Phadke, A.; Kang, H.; Hwang, N. S.; Caro, E. J.; Nguyen, S.; Siu,
- 1226 M.; Theodorakis, E. A.; Gianneschi, N. C., Calcium phosphate-bearing matrices induce
- 1227 osteogenic differentiation of stem cells through adenosine signaling. *Proceedings of the*
- 1228 *National Academy of Sciences* **2014**, 111, (3), 990-995. DOI:
- 1229 <https://doi.org/10.1073/pnas.1321717111>
- 1230 114. Hoffman, N. J.; Penque, B. A.; Habegger, K. M.; Sealls, W.; Tackett, L.; Elmendorf, J. S.,
- 1231 Chromium enhances insulin responsiveness via AMPK. *The Journal of nutritional biochemistry*
- 1232 **2014**, 25, (5), 565-572.
- 1233 DOI: <https://doi.org/10.1016/j.jnutbio.2014.01.007>
- 1234 115. EFSA, E. F. S. A., Scientific Opinion on Dietary Reference Values for Chromium. *EFSA Journal*
- 1235 **2014**, 12, (10), 3845. doi: 10.2903/j.efsa.2014.3845
- 1236 116. Raghunathan, V. K.; Tettey, J. N.; Ellis, E. M.; Grant, M. H., Comparative chronic in vitro
- 1237 toxicity of hexavalent chromium to osteoblasts and monocytes. *Journal of Biomedical Materials*
- 1238 *Research Part A* **2009**, 88, (2), 543-550.

- 1239 DOI: 10.1002/jbm.a.31893
- 1240 117. Ning, J.; Henderson, C.; Grant, M., The cytotoxicity of chromium in osteoblasts: effects on
1241 macromolecular synthesis. *Journal of Materials Science: Materials in Medicine* **2002**, 13, (1), 47-
1242 52. DOI: <https://doi.org/10.1023/A:1013630401959>
- 1243 118. Schröck, K.; Lutz, J.; Mändl, S.; Hacker, M. C.; Kamprad, M.; Schulz-Siegmund, M., Co (II)-
1244 mediated effects of plain and plasma immersion ion implanted cobalt-chromium alloys on
1245 the osteogenic differentiation of human mesenchymal stem cells. *Journal of Orthopaedic*
1246 *Research* **2015**, 33, (3), 325-333.
1247 DOI: 10.1002/jor.22765
- 1248 119. Drynda, S.; Drynda, A.; Kekow, J.; Lohmann, C., AB0074 Influence of cobalt and chromium
1249 ions on TGF-BETA expression and mineralization of bone forming cells IN-VITRO. In BMJ
1250 Publishing Group Ltd: 2017.
1251 DOI: <http://dx.doi.org/10.1136/annrheumdis-2017-eular.6218>
- 1252 120. Fan, W.; Crawford, R.; Xiao, Y., Enhancing in vivo vascularized bone formation by cobalt
1253 chloride-treated bone marrow stromal cells in a tissue engineered periosteum model.
1254 *Biomaterials* **2010**, 31, (13), 3580-3589.
1255 DOI: <https://doi.org/10.1016/j.biomaterials.2010.01.083>
- 1256 121. Yuan, Y.; Hilliard, G.; Ferguson, T.; Millhorn, D. E., Cobalt inhibits the interaction between
1257 hypoxia-inducible factor- α and von Hippel-Lindau protein by direct binding to hypoxia-
1258 inducible factor- α . *Journal of Biological Chemistry* **2003**, 278, (18), 15911-15916. DOI:
1259 <https://doi.org/10.1096/fj.10-162107>
- 1260 122. Tanaka, T.; Kojima, I.; Ohse, T.; Ingelfinger, J. R.; Adler, S.; Fujita, T.; Nangaku, M., Cobalt
1261 promotes angiogenesis via hypoxia-inducible factor and protects tubulointerstitium in the
1262 remnant kidney model. *Laboratory investigation* **2005**, 85, (10), 1292.
1263 doi:10.1038/labinvest.3700328
- 1264 123. Birgani, Z. T.; Fennema, E.; Gijbels, M. J.; de Boer, J.; van Blitterswijk, C. A.; Habibovic, P.,
1265 Stimulatory effect of cobalt ions incorporated into calcium phosphate coatings on
1266 neovascularization in an in vivo intramuscular model in goats. *Acta biomaterialia* **2016**, 36,
1267 267-276. DOI: <https://doi.org/10.1016/j.actbio.2016.03.031>
- 1268 124. Ignjatović, N.; Ajduković, Z.; Savić, V.; Najman, S.; Mihailović, D.; Vasiljević, P.; Stojanović,
1269 Z.; Uskoković, V.; Uskoković, D., Nanoparticles of cobalt-substituted hydroxyapatite in
1270 regeneration of mandibular osteoporotic bones. *Journal of Materials Science: Materials in*
1271 *Medicine* **2013**, 24, (2), 343-354.
1272 DOI: <https://doi.org/10.1007/s10856-012-4793-1>
- 1273 125. Perez, R. A.; Kim, J.-H.; Buitrago, J. O.; Wall, I. B.; Kim, H.-W., Novel therapeutic core-shell
1274 hydrogel scaffolds with sequential delivery of cobalt and bone morphogenetic protein-2 for
1275 synergistic bone regeneration. *Acta biomaterialia* **2015**, 23, 295-308. DOI:
1276 <https://doi.org/10.1016/j.actbio.2015.06.002>
- 1277 126. Kargozar, S.; Lotfibakhshai, N.; Ai, J.; Mozafari, M.; Milan, P. B.; Hamzehlou, S.; Barati,
1278 M.; Bairo, F.; Hill, R. G.; Joghataei, M. T., Strontium-and cobalt-substituted bioactive glasses
1279 seeded with human umbilical cord perivascular cells to promote bone regeneration via
1280 enhanced osteogenic and angiogenic activities. *Acta biomaterialia* **2017**, 58, 502-514. DOI:
1281 <https://doi.org/10.1016/j.actbio.2017.06.021>

- 1282 127. Festa, R. A.; Thiele, D. J., Copper: an essential metal in biology. *Current Biology* **2011**, *21*, (21),
1283 R877-R883. DOI: <https://doi.org/10.1016/j.cub.2011.09.040>
- 1284 128. Strain, J., A reassessment of diet and osteoporosis—possible role for copper. *Medical*
1285 *hypotheses* **1988**, *27*, (4), 333-338.
1286 DOI: [https://doi.org/10.1016/0306-9877\(88\)90016-3](https://doi.org/10.1016/0306-9877(88)90016-3)
- 1287 129. Strause, L. G.; Hegenauer, J.; Saltman, P.; Cone, R.; Resnick, D., Effects of long-term dietary
1288 manganese and copper deficiency on rat skeleton. *The Journal of nutrition* **1986**, *116*, (1), 135-
1289 141. DOI: <https://doi.org/10.1093/jn/116.1.135>
- 1290 130. Ziche, M.; Jones, J.; Gullino, P. M., Role of prostaglandin E1 and copper in angiogenesis.
1291 *Journal of the National Cancer Institute* **1982**, *69*, (2), 475-482.
1292 DOI: <https://doi.org/10.1093/jnci/69.2.475>
- 1293 131. Dan, Z.; Ni, H.; Xu, B.; Xiong, J.; Xiong, P., Microstructure and antibacterial properties of AISI
1294 420 stainless steel implanted by copper ions. *Thin solid films* **2005**, *492*, (1-2), 93-100. DOI:
1295 <https://doi.org/10.1016/j.tsf.2005.06.100>
- 1296 132. Gérard, C.; Bordeleau, L.-J.; Barralet, J.; Doillon, C. J., The stimulation of angiogenesis and
1297 collagen deposition by copper. *Biomaterials* **2010**, *31*, (5), 824-831. DOI:
1298 <https://doi.org/10.1016/j.biomaterials.2009.10.009>
- 1299 133. Kong, N.; Lin, K.; Li, H.; Chang, J., Synergy effects of copper and silicon ions on stimulation
1300 of vascularization by copper-doped calcium silicate. *Journal of Materials Chemistry B* **2014**, *2*,
1301 (8), 1100-1110. DOI: 10.1039/C3TB21529F
- 1302 134. Wu, C.; Zhou, Y.; Xu, M.; Han, P.; Chen, L.; Chang, J.; Xiao, Y., Copper-containing
1303 mesoporous bioactive glass scaffolds with multifunctional properties of angiogenesis
1304 capacity, osteostimulation and antibacterial activity. *Biomaterials* **2013**, *34*, (2), 422-433. DOI:
1305 <https://doi.org/10.1016/j.biomaterials.2012.09.066>
- 1306 135. Rodríguez, J. P.; Rios, S.; Gonzalez, M., Modulation of the proliferation and differentiation of
1307 human mesenchymal stem cells by copper. *Journal of cellular biochemistry* **2002**, *85*, (1), 92-100.
1308 DOI: 10.1002/jcb.10111
- 1309 136. Li, S.; Wang, M.; Chen, X.; Li, S. F.; Li-Ling, J.; Xie, H. Q., Inhibition of osteogenic
1310 differentiation of mesenchymal stem cells by copper supplementation. *Cell proliferation* **2014**,
1311 *47*, (1), 81-90. DOI: 10.1111/cpr.12083
- 1312 137. Lin, Y.; Xiao, W.; Bal, B. S.; Rahaman, M. N., Effect of copper-doped silicate 13–93 bioactive
1313 glass scaffolds on the response of MC3T3-E1 cells in vitro and on bone regeneration and
1314 angiogenesis in rat calvarial defects in vivo. *Materials Science and Engineering: C* **2016**, *67*, 440-
1315 452.
1316 DOI:<https://doi.org/10.1016/j.msec.2016.05.073>
- 1317 138. D'Mello, S.; Elangovan, S.; Hong, L.; Ross, R. D.; Sumner, D. R.; Salem, A. K., Incorporation
1318 of copper into chitosan scaffolds promotes bone regeneration in rat calvarial defects. *Journal*
1319 *of Biomedical Materials Research Part B: Applied Biomaterials* **2015**, *103*, (5), 1044-1049. DOI:
1320 10.1002/jbm.b.33290
- 1321 139. Warrell Jr, R. P.; Alcock, N. W.; Bockman, R. S., Gallium nitrate inhibits accelerated bone
1322 turnover in patients with bone metastases. *Journal of Clinical Oncology* **1987**, *5*, (2), 292-298.
1323 DOI: 10.1200/JCO.1987.5.2.292

- 1324 140. Ross-Gillespie, A.; Weigert, M.; Brown, S. P.; Kümmerli, R., Gallium-mediated siderophore
1325 quenching as an evolutionarily robust antibacterial treatment. *Evolution, medicine, and public*
1326 *health* **2014**, 2014, (1), 18-29.
1327 DOI: <https://doi.org/10.1093/emph/eou003>
- 1328 141. Zhu, Y.; Qiu, Y.; Chen, R.; Liao, L., The Inhibition of Escherichia Coli Biofilm Formation by
1329 Gallium Nitrate-Modified Titanium. *Journal of nanoscience and nanotechnology* **2015**, 15, (8),
1330 5605-5609.
1331 DOI: <https://doi.org/10.1166/jnn.2015.10305>
- 1332 142. Cochis, A.; Azzimonti, B.; Della Valle, C.; Chiesa, R.; Arciola, C. R.; Rimondini, L., Biofilm
1333 formation on titanium implants counteracted by grafting gallium and silver ions. *Journal of*
1334 *biomedical materials research Part A* **2015**, 103, (3), 1176-1187. DOI: 10.1002/jbm.a.35270
- 1335 143. Verron, E.; Masson, M.; Khoshniat, S.; Duplomb, L.; Wittrant, Y.; Baud'huin, M.; Badran, Z.;
1336 Bujoli, B.; Janvier, P.; Scimeca, J. C., Gallium modulates osteoclastic bone resorption in vitro
1337 without affecting osteoblasts. *British journal of pharmacology* **2010**, 159, (8), 1681-1692.
1338 doi:10.1111/j.1476-5381.2010.00665.x
- 1339 144. Mellier, C.; Fayon, F.; Boukhechba, F.; Verron, E.; LeFerrec, M.; Montavon, G.; Lesoeur, J.;
1340 Schnitzler, V.; Massiot, D.; Janvier, P., Design and properties of novel gallium-doped
1341 injectable apatitic cements. *Acta biomaterialia* **2015**, 24, 322-332.
1342 DOI: <https://doi.org/10.1016/j.actbio.2015.05.027>
- 1343 145. Strazic Geljic, I.; Melis, N.; Boukhechba, F.; Schaub, S.; Mellier, C.; Janvier, P.; Laugier, J. P.;
1344 Bouler, J. M.; Verron, E.; Scimeca, J. C., Gallium enhances reconstructive properties of a
1345 calcium phosphate bone biomaterial. *Journal of tissue engineering and regenerative medicine*
1346 **2017**. DOI: 10.1002/term.2396
- 1347 146. Beard, J. L., Iron biology in immune function, muscle metabolism and neuronal functioning.
1348 *The Journal of nutrition* **2001**, 131, (2), 568S-580S.
1349 DOI: <https://doi.org/10.1093/jn/131.2.568S>
- 1350 147. Abbaspour, N.; Hurrell, R.; Kelishadi, R., Review on iron and its importance for human
1351 health. *Journal of research in medical sciences: the official journal of Isfahan University of Medical*
1352 *Sciences* **2014**, 19, (2), 164.
- 1353 148. Lieu, P. T.; Heiskala, M.; Peterson, P. A.; Yang, Y., The roles of iron in health and disease.
1354 *Molecular aspects of medicine* **2001**, 22, (1-2), 1-87.
1355 DOI: [https://doi.org/10.1016/S0098-2997\(00\)00006-6](https://doi.org/10.1016/S0098-2997(00)00006-6)
- 1356 149. Andrews, N. C., Disorders of iron metabolism. *New England Journal of Medicine* **1999**, 341, (26),
1357 1986-1995. DOI: 10.1056/NEJM199912233412607
- 1358 150. Conrad, M. E.; Umbreit, J. N.; Moore, E. G., Iron absorption and transport. *The American*
1359 *journal of the medical sciences* **1999**, 318, (4), 213-229.
1360 DOI: [https://doi.org/10.1016/S0002-9629\(15\)40626-3](https://doi.org/10.1016/S0002-9629(15)40626-3)
- 1361 151. Ponka, P., Cell biology of heme. *The American journal of the medical sciences* **1999**, 318, (4), 241-
1362 256. DOI: [https://doi.org/10.1016/S0002-9629\(15\)40628-7](https://doi.org/10.1016/S0002-9629(15)40628-7)
- 1363 152. Lill, R., Function and biogenesis of iron-sulphur proteins. *Nature* **2009**, 460, (7257), 831.
1364 doi:10.1038/nature08301
- 1365 153. Ponka, P.; Beaumont, C.; Richardson, D. R. In *Function and regulation of transferrin and ferritin*,
1366 *Seminars in hematology*, 1998; 1998; pp 35-54.

- 1367 154. Abboud, S.; Haile, D. J., A novel mammalian iron-regulated protein involved in intracellular
1368 iron metabolism. *Journal of Biological Chemistry* **2000**, 275, (26), 19906-19912. doi:
1369 10.1074/jbc.M000713200
- 1370 155. Aravind, L.; Koonin, E. V., The DNA-repair protein AlkB, EGL-9, and leprecan define new
1371 families of 2-oxoglutarate-and iron-dependent dioxygenases. *Genome biology* **2001**, 2, (3),
1372 research0007. 1.
- 1373 156. Eaton, J. W.; Qian, M., Molecular bases of cellular iron toxicity¹². *Free Radical Biology and*
1374 *Medicine* **2002**, 32, (9), 833-840.
1375 DOI: [https://doi.org/10.1016/S0891-5849\(02\)00772-4](https://doi.org/10.1016/S0891-5849(02)00772-4)
- 1376 157. Diamond, T.; Stiel, D.; Posen, S., Osteoporosis in hemochromatosis: iron excess, gonadal
1377 deficiency, or other factors? *Annals of internal medicine* **1989**, 110, (6), 430-436. DOI:
1378 10.7326/0003-4819-110-6-430
- 1379 158. Zarjou, A.; Jeney, V.; Arosio, P.; Poli, M.; Zavaczki, E.; Balla, G.; Balla, J., Ferritin ferroxidase
1380 activity: a potent inhibitor of osteogenesis. *Journal of Bone and Mineral Research* **2010**, 25, (1),
1381 164-172. doi: 10.1359/jbmr.091002.
- 1382 159. Weinberg, E. D., Iron loading: a risk factor for osteoporosis. *Biometals* **2006**, 19, (6), 633-635.
1383 DOI <https://doi.org/10.1007/s10534-006-9000-8>
- 1384 160. Weinberg, E., Role of iron in osteoporosis. *Pediatric endocrinology reviews: PER* **2008**, 6, 81-85.
- 1385 161. Chen, B.; Yan, Y.-L.; Liu, C.; Bo, L.; Li, G.-F.; Wang, H.; Xu, Y.-J., Therapeutic effect of
1386 deferoxamine on iron overload-induced inhibition of osteogenesis in a zebrafish model.
1387 *Calcified tissue international* **2014**, 94, (3), 353-360.
1388 DOI <https://doi.org/10.1007/s00223-013-9817-4>
- 1389 162. Jiang, Y.; Yan, Y.; Wang, X.; Zhu, G.; Xu, Y.-J., Hepcidin inhibition on the effect of
1390 osteogenesis in zebrafish. *Biochemical and biophysical research communications* **2016**, 476, (1), 1-
1391 6. DOI: <https://doi.org/10.1016/j.bbrc.2016.05.118>
- 1392 163. Balogh, E.; Tolnai, E.; Nagy Jr, B.; Nagy, B.; Balla, G.; Balla, J.; Jeney, V., Iron overload inhibits
1393 osteogenic commitment and differentiation of mesenchymal stem cells via the induction of
1394 ferritin. *Biochimica et Biophysica Acta (BBA)-Molecular Basis of Disease* **2016**, 1862, (9), 1640-1649.
1395 DOI: <https://doi.org/10.1016/j.bbadis.2016.06.003>
- 1396 164. Jia, P.; Xu, Y. J.; Zhang, Z. L.; Li, K.; Li, B.; Zhang, W.; Yang, H., Ferric ion could facilitate
1397 osteoclast differentiation and bone resorption through the production of reactive oxygen
1398 species. *Journal of Orthopaedic Research* **2012**, 30, (11), 1843-1852. DOI: 10.1002/jor.22133
- 1399 165. Wang, Q.; Chen, B.; Cao, M.; Sun, J.; Wu, H.; Zhao, P.; Xing, J.; Yang, Y.; Zhang, X.; Ji, M.,
1400 Response of MAPK pathway to iron oxide nanoparticles in vitro treatment promotes
1401 osteogenic differentiation of hBMSCs. *Biomaterials* **2016**, 86, 11-20. DOI:
1402 <https://doi.org/10.1016/j.biomaterials.2016.02.004>
- 1403 166. Zhao, G.-y.; Zhao, L.-p.; He, Y.-f.; Li, G.-F.; Gao, C.; Li, K.; Xu, Y.-j., A comparison of the
1404 biological activities of human osteoblast hFOB1. 19 between iron excess and iron deficiency.
1405 *Biological trace element research* **2012**, 150, (1-3), 487-495. DOI <https://doi.org/10.1007/s12011-012-9511-9>
- 1407 167. Jeney, V., Clinical impact and cellular mechanisms of iron overload-associated bone loss.
1408 *Frontiers in pharmacology* **2017**, 8, 77.
1409 DOI: <https://doi.org/10.3389/fphar.2017.00077>

- 1410 168. Geddes, J. R.; Burgess, S.; Hawton, K.; Jamison, K.; Goodwin, G. M., Long-term lithium
1411 therapy for bipolar disorder: systematic review and meta-analysis of randomized controlled
1412 trials. *American Journal of Psychiatry* **2004**, 161, (2), 217-222. DOI:
1413 <https://doi.org/10.1176/appi.ajp.161.2.217>
- 1414 169. Su, H.; Chu, T.-H.; Wu, W., Lithium enhances proliferation and neuronal differentiation of
1415 neural progenitor cells in vitro and after transplantation into the adult rat spinal cord.
1416 *Experimental neurology* **2007**, 206, (2), 296-307.
1417 DOI: <https://doi.org/10.1016/j.expneurol.2007.05.018>
- 1418 170. Mora, A.; Sabio, G.; Risco, A. M. a.; Cuenda, A.; Alonso, J. C.; Soler, G.; Centeno, F., Lithium
1419 blocks the PKB and GSK3 dephosphorylation induced by ceramide through protein
1420 phosphatase-2A. *Cellular signalling* **2002**, 14, (6), 557-562.
1421 DOI: [https://doi.org/10.1016/S0898-6568\(01\)00282-0](https://doi.org/10.1016/S0898-6568(01)00282-0)
- 1422 171. De Boer, J.; Wang, H. J.; Van Blitterswijk, C., Effects of Wnt signaling on proliferation and
1423 differentiation of human mesenchymal stem cells. *Tissue engineering* **2004**, 10, (3-4), 393-401.
1424 DOI: <https://doi.org/10.1089/107632704323061753>
- 1425 172. Zhu, Z.; Yin, J.; Guan, J.; Hu, B.; Niu, X.; Jin, D.; Wang, Y.; Zhang, C., Lithium stimulates
1426 human bone marrow derived mesenchymal stem cell proliferation through GSK-3 β -
1427 dependent β -catenin/Wnt pathway activation. *The FEBS journal* **2014**, 281, (23), 5371-5389.
1428 DOI: 10.1111/febs.13081
- 1429 173. Hartmann, C., A Wnt canon orchestrating osteoblastogenesis. *Trends in cell biology* **2006**, 16,
1430 (3), 151-158. DOI: <https://doi.org/10.1016/j.tcb.2006.01.001>
- 1431 174. Vestergaard, P.; Rejnmark, L.; Mosekilde, L., Reduced relative risk of fractures among users
1432 of lithium. *Calcified tissue international* **2005**, 77, (1), 1-8.
- 1433 175. Zamani, A.; Omrani, G. R.; Nasab, M. M., Lithium's effect on bone mineral density. *Bone* **2009**,
1434 44, (2), 331-334. DOI <https://doi.org/10.1007/s00223-004-0258-y>
- 1435 176. Satija, N. K.; Sharma, D.; Afrin, F.; Tripathi, R. P.; Gangenahalli, G., High throughput
1436 transcriptome profiling of lithium stimulated human mesenchymal stem cells reveals
1437 priming towards osteoblastic lineage. *PLoS One* **2013**, 8, (1), e55769.
1438 DOI: <https://doi.org/10.1371/journal.pone.0055769>
- 1439 177. Moon, J. S.; Ko, H. M.; Park, J. i.; Kim, J. H.; Kim, S. H.; Kim, M. S., Inhibition of human
1440 mesenchymal stem cell proliferation via Wnt signaling activation. *Journal of cellular*
1441 *biochemistry* **2018**. DOI: 10.1002/jcb.26326
- 1442 178. Tang, L.; Chen, Y.; Pei, F.; Zhang, H., Lithium chloride modulates adipogenesis and
1443 osteogenesis of human bone marrow-derived mesenchymal stem cells. *Cellular Physiology and*
1444 *Biochemistry* **2015**, 37, (1), 143-152.
1445 DOI: <https://doi.org/10.1159/000430340>
- 1446 179. Wang, X.; Zhu, S.; Jiang, X.; Li, Y.; Song, D.; Hu, J., Systemic administration of lithium
1447 improves distracted bone regeneration in rats. *Calcified tissue international* **2015**, 96, (6), 534-
1448 540. DOI: <https://doi.org/10.1007/s00223-015-0004-7>
- 1449 180. da Silva, J. G.; Babb, R.; Salzechner, C.; Sharpe, P. T.; Brauer, D. S.; Gentleman, E.,
1450 Optimisation of lithium-substituted bioactive glasses to tailor cell response for hard tissue
1451 repair. *Journal of materials science* **2017**, 52, (15), 8832-8844.
1452 DOI: <https://doi.org/10.1007/s10853-017-0838-7>

- 1453 181. Cai, Y.; Guo, L.; Shen, H.; An, X.; Jiang, H.; Ji, F.; Niu, Y., Degradability, bioactivity, and
1454 osteogenesis of biocomposite scaffolds of lithium-containing mesoporous bioglass and
1455 mPEG-PLGA-b-PLL copolymer. *International journal of nanomedicine* **2015**, *10*, 4125.
1456 doi: 10.2147/IJN.S82945
- 1457 182. Wang, Y.; Yang, X.; Gu, Z.; Qin, H.; Li, L.; Liu, J.; Yu, X., In vitro study on the degradation of
1458 lithium-doped hydroxyapatite for bone tissue engineering scaffold. *Materials Science and*
1459 *Engineering: C* **2016**, *66*, 185-192.
1460 DOI: <https://doi.org/10.1016/j.msec.2016.04.065>
- 1461 183. Li, L.; Peng, X.; Qin, Y.; Wang, R.; Tang, J.; Cui, X.; Wang, T.; Liu, W.; Pan, H.; Li, B.,
1462 Acceleration of bone regeneration by activating Wnt/ β -catenin signalling pathway via
1463 lithium released from lithium chloride/calcium phosphate cement in osteoporosis. *Scientific*
1464 *reports* **2017**, *7*, 45204. doi:10.1038/srep45204
- 1465 184. Jahnen-Dechent, W.; Ketteler, M., Magnesium basics. *Clinical kidney journal* **2012**, *5*, (Suppl_1),
1466 i3-i14. DOI: <https://doi.org/10.1093/ndtplus/sfr163>
- 1467 185. Okuma, T., Magnesium and bone strength. In Elsevier: 2001.
- 1468 186. Arnaud, M. J., Update on the assessment of magnesium status. *British Journal of Nutrition*
1469 **2008**, *99*, (S3), S24-S36.
1470 DOI: <https://doi.org/10.1017/S000711450800682X>
- 1471 187. Vormann, J., Magnesium: nutrition and metabolism. *Molecular aspects of medicine* **2003**, *24*, (1-
1472 3), 27-37. DOI: [https://doi.org/10.1016/S0098-2997\(02\)00089-4](https://doi.org/10.1016/S0098-2997(02)00089-4)
- 1473 188. Weng, L.; Webster, T. J., Nanostructured magnesium has fewer detrimental effects on
1474 osteoblast function. *International journal of nanomedicine* **2013**, *8*, 1773.
1475 doi: 10.2147/IJN.S39031
- 1476 189. Castiglioni, S.; Leidi, M.; Carpanese, E.; Maier, J. A., Extracellular magnesium and in vitro
1477 cell differentiation: different behaviour of different cells. *Magnesium research* **2013**, *26*, (1), 24-
1478 31. DOI: 10.1684/mrh.2013.0330
- 1479 190. Welch, A. A.; Skinner, J.; Hickson, M., Dietary Magnesium May Be Protective for Aging of
1480 Bone and Skeletal Muscle in Middle and Younger Older Age Men and Women: Cross-
1481 Sectional Findings from the UK Biobank Cohort. *Nutrients* **2017**, *9*, (11), 1189.
1482 doi:10.3390/nu9111189
- 1483 191. Belluci, M. M.; Giro, G.; del Barrio, R. A. L.; Pereira, R. M. R.; Marcantonio, E.; Orrico, S. R.
1484 P., Effects of magnesium intake deficiency on bone metabolism and bone tissue around
1485 osseointegrated implants. *Clinical oral implants research* **2011**, *22*, (7), 716-721. DOI:
1486 10.1111/j.1600-0501.2010.02046.x
- 1487 192. Rude, R.; Gruber, H.; Norton, H.; Wei, L.; Frausto, A.; Kilburn, J., Reduction of dietary
1488 magnesium by only 50% in the rat disrupts bone and mineral metabolism. *Osteoporosis*
1489 *international* **2006**, *17*, (7), 1022-1032.
1490 DOI: <https://doi.org/10.1007/s00198-006-0104-3>
- 1491 193. Bae, Y. J.; Kim, M.-H., Calcium and magnesium supplementation improves serum
1492 OPG/RANKL in calcium-deficient ovariectomized rats. *Calcified tissue international* **2010**, *87*,
1493 (4), 365-372.
1494 DOI: <https://doi.org/10.1007/s00223-010-9410-z>

- 1495 194. Chen, X.; Liao, X.; Huang, Z.; You, P.; Chen, C.; Kang, Y.; Yin, G., Synthesis and
1496 characterization of novel multiphase bioactive glass-ceramics in the CaO-MgO-SiO₂ system.
1497 *Journal of Biomedical Materials Research Part B: Applied Biomaterials* **2010**, 93, (1), 194-202. DOI:
1498 10.1002/jbm.b.31574
- 1499 195. Mouriño, V.; Cattalini, J. P.; Boccaccini, A. R., Metallic ions as therapeutic agents in tissue
1500 engineering scaffolds: an overview of their biological applications and strategies for new
1501 developments. *Journal of the Royal Society Interface* **2011**,
1502 DOI: 10.1098/rsif.2011.0611
- 1503 196. Hussain, A.; Bessho, K.; Takahashi, K.; Tabata, Y., Magnesium calcium phosphate as a novel
1504 component enhances mechanical/physical properties of gelatin scaffold and osteogenic
1505 differentiation of bone marrow mesenchymal stem cells. *Tissue Engineering Part A* **2011**, 18,
1506 (7-8), 768-774.
1507 DOI: <https://doi.org/10.1089/ten.tea.2011.0310>
- 1508 197. Yoshizawa, S.; Chaya, A.; Verdelsis, K.; Bilodeau, E. A.; Sfeir, C., An in vivo model to assess
1509 magnesium alloys and their biological effect on human bone marrow stromal cells. *Acta*
1510 *biomaterialia* **2015**, 28, 234-239.
1511 DOI: <https://doi.org/10.1016/j.actbio.2015.08.037>
- 1512 198. Witte, F.; Kaese, V.; Haferkamp, H.; Switzer, E.; Meyer-Lindenberg, A.; Wirth, C.;
1513 Windhagen, H., In vivo corrosion of four magnesium alloys and the associated bone
1514 response. *Biomaterials* **2005**, 26, (17), 3557-3563.
1515 DOI: <https://doi.org/10.1016/j.biomaterials.2004.09.049>
- 1516 199. Witte, F.; Ulrich, H.; Palm, C.; Willbold, E., Biodegradable magnesium scaffolds: Part II: Peri-
1517 implant bone remodeling. *Journal of biomedical materials research Part A* **2007**, 81, (3), 757-765.
1518 DOI: 10.1002/jbm.a.31293
- 1519 200. Galli, S.; Naito, Y.; Karlsson, J.; He, W.; Andersson, M.; Wennerberg, A.; Jimbo, R.,
1520 Osteoconductive potential of mesoporous titania implant surfaces loaded with magnesium:
1521 an experimental study in the rabbit. *Clinical implant dentistry and related research* **2015**, 17, (6),
1522 1048-1059. doi: 10.1111/cid.12211
- 1523 201. Brown, A.; Zaky, S.; Ray Jr, H.; Sfeir, C., Porous magnesium/PLGA composite scaffolds for
1524 enhanced bone regeneration following tooth extraction. *Acta biomaterialia* **2015**, 11, 543-553.
1525 doi: 10.1016/j.actbio.2014.09.008
- 1526 202. Nabiyouni, M.; Brückner, T.; Zhou, H.; Gbureck, U.; Bhaduri, S. B., Magnesium-based
1527 Bioceramics in Orthopedic Applications. *Acta biomaterialia* **2017**. doi: 10.1016/j.actbio.2017
- 1528 203. Raman, R.; Harandi, S. E., Resistance of Magnesium Alloys to Corrosion Fatigue for
1529 Biodegradable Implant Applications: Current Status and Challenges. *Materials* **2017**, 10, (11),
1530 1316. doi: 10.3390/ma10111316
- 1531 204. Yang, J.; Koons, G. L.; Cheng, G.; Zhao, L.; Mikos, A. G.; Cui, F.-Z., A review on the
1532 exploitation of biodegradable magnesium-based composites for medical applications.
1533 *Biomedical Materials* **2017**. doi: 10.1088/1748-605X/aa8fa0
- 1534 205. Pogorielov, M.; Husak, E.; Solodivnik, A.; Zhdanov, S., Magnesium-based biodegradable
1535 alloys: Degradation, application, and alloying elements. *Interventional Medicine and Applied*
1536 *Science* **2017**, 9, (1), 27-38. doi: 10.1556/1646.9.2017.1.04

- 1537 206. Yazdimamaghani, M.; Razavi, M.; Vashae, D.; Moharamzadeh, K.; Boccaccini, A. R.; Tayebi,
1538 L., Porous magnesium-based scaffolds for tissue engineering. *Materials Science and*
1539 *Engineering: C* **2017**, 71, 1253-1266. doi: 10.1016/j.msec.2016.11.027
- 1540 207. Ibrahim, H.; Esfahani, S. N.; Poorganji, B.; Dean, D.; Elahinia, M., Resorbable bone fixation
1541 alloys, forming, and post-fabrication treatments. *Materials Science and Engineering: C* **2017**, 70,
1542 870-888. doi: 10.1016/j.msec.2016.09.069
- 1543 208. Zhao, D.; Witte, F.; Lu, F.; Wang, J.; Li, J.; Qin, L., Current status on clinical applications of
1544 magnesium-based orthopaedic implants: A review from clinical translational perspective.
1545 *Biomaterials* **2017**, 112, 287-302. doi: 10.1016/j.biomaterials.2016.10.017
- 1546 209. Li, X.; Liu, X.; Wu, S.; Yeung, K.; Zheng, Y.; Chu, P. K., Design of magnesium alloys with
1547 controllable degradation for biomedical implants: From bulk to surface. *Acta biomaterialia*
1548 **2016**, 45, 2-30. doi: 10.1016/j.actbio.2016.09.005
- 1549 210. Willbold, E.; Weizbauer, A.; Loos, A.; Seitz, J. M.; Angrisani, N.; Windhagen, H.; Reifenrath,
1550 J., Magnesium alloys: A stony pathway from intensive research to clinical reality. Different
1551 test methods and approval-related considerations. *Journal of Biomedical Materials Research Part*
1552 *A* **2017**, 105, (1), 329-347. doi: 10.1002/jbm.a.35893
- 1553 211. Agarwal, S.; Curtin, J.; Duffy, B.; Jaiswal, S., Biodegradable magnesium alloys for orthopaedic
1554 applications: A review on corrosion, biocompatibility and surface modifications. *Materials*
1555 *Science and Engineering: C* **2016**, 68, 948-963. doi: 10.1016/j.msec.2016.06.020
- 1556 212. Kuśnierczyk, K.; Basista, M., Recent advances in research on magnesium alloys and
1557 magnesium–calcium phosphate composites as biodegradable implant materials. *Journal of*
1558 *biomaterials applications* **2017**, 31, (6), 878-900. DOI:10.1177/0885328216657271
- 1559 213. Saleh, M. M.; Touny, A.; Al-Omair, M. A.; Saleh, M., Biodegradable/biocompatible coated
1560 metal implants for orthopedic applications. *Bio-medical materials and engineering* **2016**, 27, (1),
1561 87-99. doi: 10.3233/BME-161568
- 1562 214. Heise, S.; Virtanen, S.; Boccaccini, A. R., Tackling Mg alloy corrosion by natural polymer
1563 coatings—A review. *Journal of Biomedical Materials Research Part A* **2016**, 104, (10), 2628-2641.
1564 doi: 10.1002/jbm.a.35776
- 1565 215. Tian, P.; Liu, X., Surface modification of biodegradable magnesium and its alloys for
1566 biomedical applications. *Regenerative biomaterials* **2015**, 2, (2), 135-151. doi: 10.1093/rb/rbu013
- 1567 216. Lu, J.; Wei, J.; Yan, Y.; Li, H.; Jia, J.; Wei, S.; Guo, H.; Xiao, T.; Liu, C., Preparation and
1568 preliminary cytocompatibility of magnesium doped apatite cement with degradability for
1569 bone regeneration. *Journal of Materials Science: Materials in Medicine* **2011**, 22, (3), 607-615. doi:
1570 10.1007/s10856-011-4228-4
- 1571 217. He, L.; Zhang, X.; Liu, B.; Tian, Y.; Ma, W., Effect of magnesium ion on human osteoblast
1572 activity. *Brazilian Journal of Medical and Biological Research* **2016**, 49, (7). doi: 10.1590/1414-
1573 431X20165257
- 1574 218. Leem, Y. H.; Lee, K. S.; Kim, J. H.; Seok, H. K.; Chang, J. S.; Lee, D. H., Magnesium ions
1575 facilitate integrin alpha 2-and alpha 3-mediated proliferation and enhance alkaline
1576 phosphatase expression and activity in hBMSCs. *Journal of tissue engineering and regenerative*
1577 *medicine* **2016**, 10, (10). doi: 10.1002/term.1861

- 1578 219. Yoshizawa, S.; Brown, A.; Barchowsky, A.; Sfeir, C., Magnesium ion stimulation of bone
1579 marrow stromal cells enhances osteogenic activity, simulating the effect of magnesium alloy
1580 degradation. *Acta biomaterialia* **2014**, *10*, (6), 2834-2842. doi: 10.1016/j.actbio.2014.02.002
- 1581 220. Leidi, M.; Dellera, F.; Mariotti, M.; Maier, J. A., High magnesium inhibits human osteoblast
1582 differentiation in vitro. *Magnesium research* **2011**, *24*, (1), 1-6. doi: 10.1684/mrh.2011.0271
- 1583 221. Zhang, L.; Yang, C.; Li, J.; Zhu, Y.; Zhang, X., High extracellular magnesium inhibits
1584 mineralized matrix deposition and modulates intracellular calcium signaling in human bone
1585 marrow-derived mesenchymal stem cells. *Biochemical and biophysical research communications*
1586 **2014**, *450*, (4), 1390-1395. doi: 10.1016/j.bbrc.2014.07.004
- 1587 222. Li, Y.; Wang, J.; Yue, J.; Wang, Y.; Yang, C.; Cui, Q., High magnesium prevents matrix vesicle-
1588 mediated mineralization in human bone marrow-derived mesenchymal stem cells via
1589 mitochondrial pathway and autophagy. *Cell biology international* **2017**. doi: 10.1002/cbin.10888
- 1590 223. Tsao, Y.-T.; Shih, Y.-Y.; Liu, Y.-A.; Liu, Y.-S.; Lee, O. K., Knockdown of SLC41A1 magnesium
1591 transporter promotes mineralization and attenuates magnesium inhibition during
1592 osteogenesis of mesenchymal stromal cells. *Stem cell research & therapy* **2017**, *8*, (1), 39. doi:
1593 10.1186/s13287-017-0497-2
- 1594 224. Roy, M.; Nishimoto, S., Matrix Gla protein binding to hydroxyapatite is dependent on the
1595 ionic environment: calcium enhances binding affinity but phosphate and magnesium
1596 decrease affinity. *Bone* **2002**, *31*, (2), 296-302. DOI: [https://doi.org/10.1016/S8756-3282\(02\)00821-9](https://doi.org/10.1016/S8756-3282(02)00821-9)
- 1597
1598 225. O'Young, J.; Liao, Y.; Xiao, Y.; Jalkanen, J.; Lajoie, G.; Karttunen, M.; Goldberg, H. A.; Hunter,
1599 G. K., Matrix Gla protein inhibits ectopic calcification by a direct interaction with
1600 hydroxyapatite crystals. *Journal of the American Chemical Society* **2011**, *133*, (45), 18406-18412.
1601 doi: 10.1021/ja207628k
- 1602 226. Nakatani, S.; Mano, H.; Ryanghyok, I.; Shimizu, J.; Wada, M., Excess magnesium inhibits
1603 excess calcium-induced matrix-mineralization and production of matrix gla protein (MGP)
1604 by ATDC5 cells. *Biochemical and biophysical research communications* **2006**, *348*, (3), 1157-1162.
1605 DOI:10.1016/j.bbrc.2006.07.180
- 1606 227. Ding, W., Opportunities and challenges for the biodegradable magnesium alloys as next-
1607 generation biomaterials. *Regenerative biomaterials* **2016**, *3*, (2), 79-86. doi: 10.1093/rb/rbw003
- 1608 228. Kim, J.-A.; Lim, J.; Naren, R.; Yun, H.-s.; Park, E. K., Effect of the biodegradation rate
1609 controlled by pore structures in magnesium phosphate ceramic scaffolds on bone tissue
1610 regeneration in vivo. *Acta biomaterialia* **2016**, *44*, 155-167. doi: 10.1016/j.actbio.2016.08.039
- 1611 229. Bornapour, M.; Celikin, M.; Cerruti, M.; Pekguleryuz, M., Magnesium implant alloy with low
1612 levels of strontium and calcium: The third element effect and phase selection improve bio-
1613 corrosion resistance and mechanical performance. *Materials Science and Engineering: C* **2014**,
1614 *35*, 267-282. doi: 10.1016/j.msec.2013.11.011
- 1615 230. Weng, L.; Webster, T. J., Nanostructured magnesium increases bone cell density.
1616 *Nanotechnology* **2012**, *23*, (48), 485105. doi: 10.1088/0957-4484/23/48/485105
- 1617 231. Schramm, V. L., *Manganese in metabolism and enzyme function*. Elsevier: 2012.
- 1618 232. Dormond, O.; Ponsonnet, L.; Hasmim, M.; Foletti, A.; Rüegg, C., Manganese-induced
1619 integrin affinity maturation promotes recruitment of $\alpha V\beta 3$ integrin to focal adhesions in

- endothelial cells: evidence for a role of phosphatidylinositol 3-kinase and Src. *Thrombosis and haemostasis* **2004**, *91*, (01), 151-161. DOI:10.1160/TH03-11-0728
233. Mould, A. P.; Akiyama, S. K.; Humphries, M. J., Regulation of Integrin $\alpha 5\beta 1$ -Fibronectin Interactions by Divalent Cations EVIDENCE FOR DISTINCT CLASSES OF BINDING SITES FOR Mn^{2+} , Mg^{2+} , AND Ca^{2+} . *Journal of Biological Chemistry* **1995**, *270*, (44), 26270-26277. doi: 10.1074/jbc.270.44.26270
234. Lüthen, F.; Bulnheim, U.; Müller, P. D.; Rychly, J.; Jesswein, H.; Nebe, J. B., Influence of manganese ions on cellular behavior of human osteoblasts in vitro. *Biomolecular engineering* **2007**, *24*, (5), 531-536. DOI:10.1016/j.bioeng.2007.08.003
235. Pabbruwe, M. B.; Standard, O. C.; Sorrell, C. C.; Howlett, C. R., Bone formation within alumina tubes: effect of calcium, manganese, and chromium dopants. *Biomaterials* **2004**, *25*, (20), 4901-4910. DOI:10.1016/j.biomaterials.2004.01.005
236. Paglia, D. N.; Wey, A.; Park, A. G.; Breitbart, E. A.; Mehta, S. K.; Bogden, J. D.; Kemp, F. W.; Benevenia, J.; O'connor, J. P.; Lin, S. S., The effects of local vanadium treatment on angiogenesis and chondrogenesis during fracture healing. *Journal of Orthopaedic Research* **2012**, *30*, (12), 1971-1978. doi: 10.1002/jor.22159
237. Subasinghe, S.; Greenbaum, A. L.; McLean, P., The insulin-mimetic action of Mn^{2+} : Involvement of cyclic nucleotides and insulin in the regulation of hepatic hexokinase and glucokinase. *Biochemical medicine* **1985**, *34*, (1), 83-92. [https://doi.org/10.1016/0006-2944\(85\)90064-X](https://doi.org/10.1016/0006-2944(85)90064-X)
238. Hreha, J.; Wey, A.; Cunningham, C.; Krell, E. S.; Brietbart, E. A.; Paglia, D. N.; Montemurro, N. J.; Nguyen, D. A.; Lee, Y. J.; Komlos, D., Local manganese chloride treatment accelerates fracture healing in a rat model. *Journal of Orthopaedic Research* **2015**, *33*, (1), 122-130. doi: 10.1002/jor.22733
239. Russell, A.; Hugo, W., 7 antimicrobial activity and action of silver. In *Progress in medicinal chemistry*, Elsevier: 1994; Vol. 31, pp 351-370. [https://doi.org/10.1016/S0079-6468\(08\)70024-9](https://doi.org/10.1016/S0079-6468(08)70024-9)
240. Jeanmonod, P.; Laschke, M. W.; Gola, N.; von Heesen, M.; Glanemann, M.; Dold, S.; Menger, M. D.; Moussavian, M. R., Silver acetate coating promotes early vascularization of Dacron vascular grafts without inducing host tissue inflammation. *Journal of vascular surgery* **2013**, *58*, (6), 1637-1643. doi:10.1016/j.jvs.2013.02.012
241. Rigo, C.; Ferroni, L.; Tocco, I.; Roman, M.; Munivrana, I.; Gardin, C.; Cairns, W. R.; Vindigni, V.; Azzena, B.; Barbante, C., Active silver nanoparticles for wound healing. *International journal of molecular sciences* **2013**, *14*, (3), 4817-4840. doi: 10.3390/ijms14034817
242. Hebeish, A.; El-Rafie, M.; El-Sheikh, M.; Seleem, A. A.; El-Naggar, M. E., Antimicrobial wound dressing and anti-inflammatory efficacy of silver nanoparticles. *International journal of biological macromolecules* **2014**, *65*, 509-515. doi: 10.1016/j.ijbiomac.2014.01.071
243. Tian, J.; Wong, K. K.; Ho, C. M.; Lok, C. N.; Yu, W. Y.; Che, C. M.; Chiu, J. F.; Tam, P. K., Topical delivery of silver nanoparticles promotes wound healing. *ChemMedChem* **2007**, *2*, (1), 129-136. DOI:10.1002/cmdc.200600171
244. Qin, H.; Zhu, C.; An, Z.; Jiang, Y.; Zhao, Y.; Wang, J.; Liu, X.; Hui, B.; Zhang, X.; Wang, Y., Silver nanoparticles promote osteogenic differentiation of human urine-derived stem cells at noncytotoxic concentrations. *International journal of nanomedicine* **2014**, *9*, 2469. doi: 10.2147/IJN.S59753

- 1663 245. Mahmood, M.; Li, Z.; Casciano, D.; Khodakovskaya, M. V.; Chen, T.; Karmakar, A.; Dervishi,
1664 E.; Xu, Y.; Mustafa, T.; Watanabe, F., Nanostructural materials increase mineralization in
1665 bone cells and affect gene expression through miRNA regulation. *Journal of cellular and*
1666 *molecular medicine* **2011**, 15, (11), 2297-2306. doi: 10.1111/j.1582-4934.2010.01234.x
- 1667 246. Qing, T.; Mahmood, M.; Zheng, Y.; Biris, A. S.; Shi, L.; Casciano, D. A., A genomic
1668 characterization of the influence of silver nanoparticles on bone differentiation in MC3T3-E1
1669 cells. *Journal of Applied Toxicology* **2018**, 38, (2), 172-179. doi: 10.1002/jat.3528
- 1670 247. Kwan, K. H.; Liu, X.; To, M. K.; Yeung, K. W.; Ho, C.-m.; Wong, K. K., Modulation of collagen
1671 alignment by silver nanoparticles results in better mechanical properties in wound healing.
1672 *Nanomedicine: Nanotechnology, Biology and Medicine* **2011**, 7, (4), 497-504. doi:
1673 10.1016/j.nano.2011.01.003
- 1674 248. Liu, X.; Lee, P. y.; Ho, C. m.; Lui, V. C.; Chen, Y.; Che, C. m.; Tam, P. K.; Wong, K. K., Silver
1675 nanoparticles mediate differential responses in keratinocytes and fibroblasts during skin
1676 wound healing. *ChemMedChem* **2010**, 5, (3), 468-475. doi: 10.1002/cmdc.200900502
- 1677 249. Zhang, R.; Lee, P.; Lui, V. C.; Chen, Y.; Liu, X.; Lok, C. N.; To, M.; Yeung, K. W.; Wong, K. K.,
1678 Silver nanoparticles promote osteogenesis of mesenchymal stem cells and improve bone
1679 fracture healing in osteogenesis mechanism mouse model. *Nanomedicine: Nanotechnology,*
1680 *Biology and Medicine* **2015**, 11, (8), 1949-1959. doi: 10.1016/j.nano.2015.07.016
- 1681 250. Yang, F.; Yang, D.; Tu, J.; Zheng, Q.; Cai, L.; Wang, L., Strontium enhances osteogenic
1682 differentiation of mesenchymal stem cells and in vivo bone formation by activating
1683 Wnt/catenin signaling. *Stem cells* **2011**, 29, (6), 981-991. doi: 10.1002/stem.646
- 1684 251. Reginster, J.-Y.; Seeman, E.; De Vernejoul, M.; Adami, S.; Compston, J.; Phenekos, C.;
1685 Devogelaer, J.-P.; Curiel, M. D.; Sawicki, A.; Goemaere, S., Strontium ranelate reduces the
1686 risk of nonvertebral fractures in postmenopausal women with osteoporosis: Treatment of
1687 Peripheral Osteoporosis (TROPOS) study. *The journal of clinical endocrinology & metabolism*
1688 **2005**, 90, (5), 2816-2822. DOI:10.1210/jc.2004-1774
- 1689 252. Saidak, Z.; Marie, P. J., Strontium signaling: molecular mechanisms and therapeutic
1690 implications in osteoporosis. *Pharmacology & therapeutics* **2012**, 136, (2), 216-226. doi:
1691 10.1016/j.pharmthera.2012.07.009
- 1692 253. Marie, P. J., Strontium ranelate: a dual mode of action rebalancing bone turnover in favour
1693 of bone formation. *Current opinion in rheumatology* **2006**, 18, S11-S15.
1694 DOI:10.1097/01.bor.0000229522.89546.7b
- 1695 254. Ammann, P., Strontium ranelate: a novel mode of action leading to renewed bone quality.
1696 *Osteoporosis international* **2005**, 16, (1), S11-S15. DOI:10.1007/s00198-004-1809-9
- 1697 255. Bonnelye, E.; Chabadel, A.; Saltel, F.; Jurdic, P., Dual effect of strontium ranelate: stimulation
1698 of osteoblast differentiation and inhibition of osteoclast formation and resorption in vitro.
1699 *Bone* **2008**, 42, (1), 129-138. DOI:10.1016/j.bone.2007.08.043
- 1700 256. Chattopadhyay, N.; Quinn, S. J.; Kifor, O.; Ye, C.; Brown, E. M., The calcium-sensing receptor
1701 (CaR) is involved in strontium ranelate-induced osteoblast proliferation. *Biochemical*
1702 *pharmacology* **2007**, 74, (3), 438-447. DOI:10.1016/j.bcp.2007.04.020
- 1703 257. Hurtel-Lemaire, A. S.; Mentaverri, R.; Caudrillier, A.; Cournarie, F.; Wattel, A.; Kamel, S.;
1704 Terwilliger, E. F.; Brown, E. M.; Brazier, M., The calcium-sensing receptor is involved in

- 1705 strontium ranelate-induced osteoclast apoptosis new insights into the associated signaling
1706 pathways. *Journal of Biological Chemistry* **2009**, *284*, (1), 575-584. doi: 10.1074/jbc.M801668200
- 1707 258. Takaoka, S.; Yamaguchi, T.; Yano, S.; Yamauchi, M.; Sugimoto, T., The Calcium-sensing
1708 Receptor (CaR) is involved in strontium ranelate-induced osteoblast differentiation and
1709 mineralization. *Hormone and metabolic research* **2010**, *42*, (09), 627-631. DOI: 10.1055/s-0030-
1710 1255091
- 1711 259. Barbara, A.; Delannoy, P.; Denis, B.; Marie, P., Normal matrix mineralization induced by
1712 strontium ranelate in MC3T3-E1 osteogenic cells. *Metabolism-Clinical and Experimental* **2004**,
1713 *53*, (4), 532-537. <https://doi.org/10.1016/j.metabol.2003.10.022>
- 1714 260. Rybchyn, M. S.; Slater, M.; Conigrave, A. D.; Mason, R. S., An Akt-dependent increase in
1715 canonical Wnt signaling and a decrease in sclerostin protein levels are involved in strontium
1716 ranelate-induced osteogenic effects in human osteoblasts. *Journal of Biological Chemistry* **2011**,
1717 *286*, (27), 23771-23779. doi: 10.1074/jbc.M111.251116
- 1718 261. Brennan, T. C.; Rybchyn, M. S.; Green, W.; Atwa, S.; Conigrave, A. D.; Mason, R. S.,
1719 Osteoblasts play key roles in the mechanisms of action of strontium ranelate. *British journal
1720 of pharmacology* **2009**, *157*, (7), 1291-1300. doi: 10.1111/j.1476-5381.2009.00305.x
- 1721 262. Schumacher, M.; Wagner, A.; Kokesch-Himmelreich, J.; Bernhardt, A.; Rohnke, M.; Wenisch,
1722 S.; Gelinsky, M., Strontium substitution in apatitic CaP cements effectively attenuates
1723 osteoclastic resorption but does not inhibit osteoclastogenesis. *Acta biomaterialia* **2016**, *37*, 184-
1724 194. doi: 10.1016/j.actbio.2016.04.016
- 1725 263. Atteritano, M.; Catalano, A.; Santoro, D.; Lasco, A.; Benvenga, S., Effects of strontium ranelate
1726 on markers of cardiovascular risk in postmenopausal osteoporotic women. *Endocrine* **2016**,
1727 *53*, (1), 305-312. doi: 10.1007/s12020-015-0721-8
- 1728 264. Li, Y.; Li, Q.; Zhu, S.; Luo, E.; Li, J.; Feng, G.; Liao, Y.; Hu, J., The effect of strontium-
1729 substituted hydroxyapatite coating on implant fixation in ovariectomized rats. *Biomaterials*
1730 **2010**, *31*, (34), 9006-9014. doi: 10.1016/j.biomaterials.2010.07.112
- 1731 265. Forsgren, J.; Engqvist, H., A novel method for local administration of strontium from implant
1732 surfaces. *Journal of Materials Science: Materials in Medicine* **2010**, *21*, (5), 1605-1609. doi:
1733 10.1007/s10856-010-4022-8
- 1734 266. Xin, Y.; Jiang, J.; Huo, K.; Hu, T.; Chu, P. K., Bioactive SrTiO₃ nanotube arrays: strontium
1735 delivery platform on Ti-based osteoporotic bone implants. *ACS nano* **2009**, *3*, (10), 3228-3234.
1736 doi: 10.1021/nn9007675
- 1737 267. Li, Y.; Leong, J.; Lu, W.; Luk, K.; Cheung, K.; Chiu, K.; Chow, S., A novel injectable bioactive
1738 bone cement for spinal surgery: a developmental and preclinical study. *Journal of Biomedical
1739 Materials Research Part A* **2000**, *52*, (1), 164-170. DOI: 10.1002/1097-4636(200010)52:1<164::AID-
1740 JBM21>3.0.CO;2-R
- 1741 268. Alkhraisat, M. H.; Moseke, C.; Blanco, L.; Barralet, J. E.; Lopez-Carbacos, E.; Gbureck, U.,
1742 Strontium modified biocements with zero order release kinetics. *Biomaterials* **2008**, *29*, (35),
1743 4691-4697. doi: 10.1016/j.biomaterials.2008.08.026
- 1744 269. Dagang, G.; Kewei, X.; Yong, H., The influence of Sr doses on the in vitro biocompatibility
1745 and in vivo degradability of single-phase Sr-incorporated HAP cement. *Journal of Biomedical
1746 Materials Research Part A* **2008**, *86*, (4), 947-958. DOI:10.1002/jbm.a.31687

- 1747 270. Tadier, S.; Bareille, R.; Siadous, R.; Marsan, O.; Charvillat, C.; Cazalbou, S.; Amédée, J.; Rey,
1748 C.; Combes, C., Strontium-loaded mineral bone cements as sustained release systems:
1749 Compositions, release properties, and effects on human osteoprogenitor cells. *Journal of*
1750 *Biomedical Materials Research Part B: Applied Biomaterials* **2012**, 100, (2), 378-390. doi:
1751 10.1002/jbm.b.31959
- 1752 271. Romieu, G.; Garric, X.; Munier, S.; Vert, M.; Boudeville, P., Calcium–strontium mixed
1753 phosphate as novel injectable and radio-opaque hydraulic cement. *Acta biomaterialia* **2010**, 6,
1754 (8), 3208-3215. doi: 10.1016/j.actbio.2010.02.008
- 1755 272. Pina, S.; Torres, P.; Goetz-Neunhoeffler, F.; Neubauer, J.; Ferreira, J., Newly developed Sr-
1756 substituted α -TCP bone cements. *Acta biomaterialia* **2010**, 6, (3), 928-935. doi:
1757 10.1016/j.actbio.2009.09.001
- 1758 273. Zhang, W.; Shen, Y.; Pan, H.; Lin, K.; Liu, X.; Darvell, B. W.; Lu, W. W.; Chang, J.; Deng, L.;
1759 Wang, D., Effects of strontium in modified biomaterials. *Acta Biomaterialia* **2011**, 7, (2), 800-
1760 808. <https://doi.org/10.1016/j.actbio.2010.08.031>
- 1761 274. Ginebra, M.-P.; Canal, C.; Espanol, M.; Pastorino, D.; Montufar, E. B., Calcium phosphate
1762 cements as drug delivery materials. *Advanced drug delivery reviews* **2012**, 64, (12), 1090-1110.
1763 doi: 10.1016/j.addr.2012.01.008
- 1764 275. Querido, W.; Rossi, A. L.; Farina, M., The effects of strontium on bone mineral: A review on
1765 current knowledge and microanalytical approaches. *Micron* **2016**, 80, 122-134.
1766 <https://doi.org/10.1016/j.micron.2015.10.006>
- 1767 276. Dahl, S.; Allain, P.; Marie, P.; Mauras, Y.; Boivin, G.; Ammann, P.; Tsouderos, Y.; Delmas, P.;
1768 Christiansen, C., Incorporation and distribution of strontium in bone. *Bone* **2001**, 28, (4), 446-
1769 453. [https://doi.org/10.1016/S8756-3282\(01\)00419-7](https://doi.org/10.1016/S8756-3282(01)00419-7)
- 1770 277. Poucheret, P.; Verma, S.; Grynepas, M. D.; McNeill, J. H., Vanadium and diabetes. *Molecular*
1771 *and Cellular Biochemistry* **1998**, 188, (1-2), 73-80. <https://doi.org/10.1023/A:1006820522587>
- 1772 278. Nobes, C. D.; Hawkins, P.; Stephens, L.; Hall, A., Activation of the small GTP-binding
1773 proteins rho and rac by growth factor receptors. *Journal of cell science* **1995**, 108, (1), 225-233.
- 1774 279. Barrio, D.; Etcheverry, S., Vanadium and bone development: putative signaling pathways.
1775 *Canadian journal of physiology and pharmacology* **2006**, 84, (7), 677-686. DOI:10.1139/y06-022
- 1776 280. Cortizo, A. M.; Etcheverry, S. B., Vanadium derivatives act as growth factor–mimetic
1777 compounds upon differentiation and proliferation of osteoblast-like UMR106 cells. *Molecular*
1778 *and cellular biochemistry* **1995**, 145, (2), 97-102. <https://doi.org/10.1007/BF00935481>
- 1779 281. Gandhi, A.; Beam, H. A.; O'Connor, J. P.; Parsons, J. R.; Lin, S. S., The effects of local insulin
1780 delivery on diabetic fracture healing. *Bone* **2005**, 37, (4), 482-490.
1781 DOI:10.1016/j.bone.2005.04.039
- 1782 282. Dedania, J.; Borzio, R.; Paglia, D.; Breitbart, E. A.; Mitchell, A.; Vaidya, S.; Wey, A.; Mehta, S.;
1783 Benevenia, J.; O'connor, J. P., Role of local insulin augmentation upon allograft incorporation
1784 in a rat femoral defect model. *Journal of Orthopaedic Research* **2011**, 29, (1), 92-99. doi:
1785 10.1002/jor.21205
- 1786 283. Cortizo, A. M.; Ruderman, G.; Mazzini, F. N.; Molinuevo, M. S.; Mogilner, I. G., Novel
1787 vanadium-loaded ordered collagen scaffold promotes osteochondral differentiation of bone
1788 marrow progenitor cells. *International journal of biomaterials* **2016**, 2016. doi:
1789 10.1155/2016/1486350

- 1790 284. Jarrell, J. D.; Dolly, B.; Morgan, J. R., Controlled release of vanadium from titanium oxide
1791 coatings for improved integration of soft tissue implants. *Journal of Biomedical Materials*
1792 *Research Part A* **2009**, 90, (1), 272-281. doi: 10.1002/jbm.a.32093
- 1793 285. Rink, L., Zinc and the immune system. *Proceedings of the Nutrition Society* **2000**, 59, (4), 541-
1794 552. <https://doi.org/10.1017/S0029665100000781>
- 1795 286. MacDonald, R. S., The role of zinc in growth and cell proliferation. *The Journal of nutrition*
1796 **2000**, 130, (5), 1500S-1508S. <https://doi.org/10.1093/jn/130.5.1500S>
- 1797 287. Yamaguchi, M., Role of zinc in bone formation and bone resorption. *The Journal of Trace*
1798 *Elements in Experimental Medicine* **1998**, 11, (2-3), 119-135. DOI: 10.1002/(SICI)1520-
1799 670X(1998)11:2/3<119::AID-JTRA5>3.0.CO;2-3
- 1800 288. Zhu, D.; Su, Y.; Young, M. L.; Ma, J.; Zheng, Y.; Tang, L., Biological responses and
1801 mechanisms of human bone marrow mesenchymal stem cells to Zn and Mg biomaterials.
1802 *ACS applied materials & interfaces* **2017**, 9, (33), 27453-27461. doi: 10.1021/acsami.7b06654
- 1803 289. Ma, J.; Zhao, N.; Zhu, D., Bioabsorbable zinc ion induced biphasic cellular responses in
1804 vascular smooth muscle cells. *Scientific reports* **2016**, 6, 26661. doi: 10.1038/srep26661
- 1805 290. Qiao, Y.; Zhang, W.; Tian, P.; Meng, F.; Zhu, H.; Jiang, X.; Liu, X.; Chu, P. K., Stimulation of
1806 bone growth following zinc incorporation into biomaterials. *Biomaterials* **2014**, 35, (25), 6882-
1807 6897. doi: 10.1016/j.biomaterials.2014.04.101
- 1808 291. Nagata, M.; Lönnerdal, B., Role of zinc in cellular zinc trafficking and mineralization in a
1809 murine osteoblast-like cell line. *The Journal of nutritional biochemistry* **2011**, 22, (2), 172-178. doi:
1810 10.1016/j.jnutbio.2010.01.003
- 1811 292. Yamaguchi, M.; Weitzmann, M. N., Zinc stimulates osteoblastogenesis and suppresses
1812 osteoclastogenesis by antagonizing NF- κ B activation. *Molecular and cellular biochemistry* **2011**,
1813 355, (1-2), 179. doi: 10.1007/s11010-011-0852-z
- 1814 293. Seo, H.-J.; Cho, Y.-E.; Kim, T.; Shin, H.-I.; Kwun, I.-S., Zinc may increase bone formation
1815 through stimulating cell proliferation, alkaline phosphatase activity and collagen synthesis
1816 in osteoblastic MC3T3-E1 cells. *Nutrition research and practice* **2010**, 4, (5), 356-361. doi:
1817 10.4162/nrp.2010.4.5.356
- 1818 294. Chou, A. H.; LeGeros, R. Z.; Chen, Z.; Li, Y., Antibacterial effect of zinc phosphate
1819 mineralized guided bone regeneration membranes. *Implant dentistry* **2007**, 16, (1), 89-100.
1820 DOI:10.1097/ID.0b013e318031224a
- 1821 295. Chou, J.; Komuro, M.; Hao, J.; Kuroda, S.; Hattori, Y.; Ben-Nissan, B.; Milthorpe, B.; Otsuka,
1822 M., Bioresorbable zinc hydroxyapatite guided bone regeneration membrane for bone
1823 regeneration. *Clinical oral implants research* **2016**, 27, (3), 354-360. doi: 10.1111/clr.12520
- 1824 296. Chen, J.; Zhang, X.; Cai, H.; Chen, Z.; Wang, T.; Jia, L.; Wang, J.; Wan, Q.; Pei, X., Osteogenic
1825 activity and antibacterial effect of zinc oxide/carboxylated graphene oxide nanocomposites:
1826 Preparation and in vitro evaluation. *Colloids and Surfaces B: Biointerfaces* **2016**, 147, 397-407.
1827 <https://doi.org/10.1016/j.colsurfb.2016.08.023>
- 1828 297. Luo, X.; Barbieri, D.; Davison, N.; Yan, Y.; de Bruijn, J. D.; Yuan, H., Zinc in calcium
1829 phosphate mediates bone induction: in vitro and in vivo model. *Acta biomaterialia* **2014**, 10,
1830 (1), 477-485. <https://doi.org/10.1016/j.actbio.2013.10.011>
- 1831 298. Yu, J.; Xu, L.; Li, K.; Xie, N.; Xi, Y.; Wang, Y.; Zheng, X.; Chen, X.; Wang, M.; Ye, X., Zinc-
1832 modified calcium silicate coatings promote osteogenic differentiation through TGF- β /Smad

- 1833 pathway and osseointegration in osteopenic rabbits. *Scientific Reports* **2017**, *7*, (1), 3440.
1834 doi:10.1038/s41598-017-03661-5
- 1835 299. Webster, T. J.; Ergun, C.; Doremus, R. H.; Bizios, R., Hydroxylapatite with substituted
1836 magnesium, zinc, cadmium, and yttrium. II. Mechanisms of osteoblast adhesion. *Journal of*
1837 *Biomedical Materials Research Part A* **2002**, *59*, (2), 312-317. DOI: 10.1002/jbm.1247
- 1838 300. Manivasagam, G.; Dhinasekaran, D.; Rajamanickam, A., Biomedical Implants: Corrosion and
1839 its Prevention-A Review. *Recent patents on corrosion science* **2010**.
- 1840 301. Fleck, C.; Eifler, D., Corrosion, fatigue and corrosion fatigue behaviour of metal implant
1841 materials, especially titanium alloys. *International journal of fatigue* **2010**, *32*, (6), 929-935. doi:
1842 10.1016/j.msec.2015.10.006
- 1843 302. Wang, J. Y.; Wicklund, B. H.; Gustilo, R. B.; Tsukayama, D. T., Titanium, chromium and
1844 cobalt ions modulate the release of bone-associated cytokines by human
1845 monocytes/macrophages in vitro. *Biomaterials* **1996**, *17*, (23), 2233-2240.
1846 [https://doi.org/10.1016/0142-9612\(96\)00072-5](https://doi.org/10.1016/0142-9612(96)00072-5)
- 1847 303. Woodman, J.; Jacobs, J.; Galante, J.; Urban, R., Metal ion release from titanium-based
1848 prosthetic segmental replacements of long bones in baboons: A long-term study. *Journal of*
1849 *Orthopaedic Research* **1983**, *1*, (4), 421-430. DOI: 10.1002/jor.1100010411
- 1850