

Review

# Linoleic Acid Derivative DCP-LA Prevents Tau Phosphorylation by Targeting GSK-3 $\beta$

Tomoyuki Nishizaki<sup>1,\*</sup>

<sup>1</sup> Innovative Bioinformation Research Organization, 2-3-14 Katsuragi, Kita-ku, Kobe 6751-1223, Japan; [tnishizaki@bioresorganization.com](mailto:tnishizaki@bioresorganization.com) (T.N.)

\* Correspondence: [tnishizaki@bioresorganization.com](mailto:tnishizaki@bioresorganization.com) (T.N.)

**Abstract:** Abnormal Tau phosphorylation and aggregation into neuronal paired helical filaments and neurofibrillary tangles cause tauopathies, a class of neurodegenerative diseases, that include Alzheimer's disease, frontotemporal dementia and parkinsonism linked to chromosome 17, progressive supranuclear palsy, Pick's disease, and corticobasal degeneration. Glycogen synthase kinase-3 $\beta$  (GSK-3 $\beta$ ) is the most critical kinase to phosphorylate Tau. We have developed the linoleic acid derivative 8-[2-(2-pentyl-cyclopropylmethyl)-cyclopropyl]-octanoic acid (DCP-LA), with cyclopropane rings instead of *cis*-double bonds, as an anti-dementia drug. DCP-LA serves as a selective activator of PKC $\epsilon$  and a potent inhibitor of protein tyrosine phosphatase 1B (PTP1B). DCP-LA prevents Tau phosphorylation due to PKC $\epsilon$ -mediated direct inactivation of GSK-3 $\beta$ , to PKC $\epsilon$ /Akt-mediated inactivation of GSK-3 $\beta$ , and to receptor tyrosine kinase-mediated inactivation of GSK-3 $\beta$  in association with PTP1B inhibition. DCP-LA targeting GSK-3 $\beta$ , thus, could become a valid drug for treatment of tauopathies.

**Keywords:** DCP-LA; PKC $\epsilon$ ; protein tyrosine phosphatase 1B; Akt; GSK-3 $\beta$ ; tauopathy

## 1. Introduction

Alzheimer's disease (AD), a neurodegenerative disease, is characterized by extensive deposition of amyloid  $\beta$  ( $A\beta$ ) called amyloid plaque and formation of neurofibrillary tangles (NFTs).  $A\beta$  has been long thought to be the most causative factor in AD, and therefore, a variety of AD drugs targeting  $A\beta$  have been developed. No beneficial result, however, has been obtained even though  $A\beta$  was cleaned up from the brain. A current challenge, therefore, has focused upon Tau protein.

Tau, a microtubule-associated protein, is abundantly expressed in neurons of the central nervous system. Tau is upregulated during neuronal development, to promote generation of cell processes and establish cell polarity [1]. Microtubules are the tracks for motor proteins bearing intracellular cargo transport [2,3]. Tau polymerizes and stabilizes microtubules by interacting with tubulin, and modulates microtubule dynamics including axonal transport [4-7].

When hyperphosphorylated, Tau detaches from the microtubules and forms insoluble fibrils, referred to as paired helical filaments (PHFs), followed by NFTs comprising aggregation of PHFs [8,9]. Aggregation of hyperphosphorylated Tau is responsible for tauopathies, a class of neurodegenerative diseases, that include not only AD but frontotemporal dementia and parkinsonism linked to chromosome 17, progressive supranuclear palsy, Pick's disease, and corticobasal degeneration [10].

Tau is phosphorylated by serine/threonine protein kinases such as GSK-3 $\beta$ , cyclin-dependent kinase 5 (Cdk5)/p25, extracellular signal-regulated kinase 2 (ERK2), p70 ribosomal protein S6 kinase (p70-S6K), microtubule affinity-regulating kinase (MARK), SAD kinase (SADK), protein kinase A (PKA), calcium/calmodulin-dependent protein kinase II (CaMKII) or tyrosine kinases such as Fyn and c-Abl [11-15]. Of the protein kinases GSK-3 $\beta$  plays a central role in Tau phosphorylation relevant to tauopathies. The present review shows that the linoleic acid derivative DCP-LA is capable of preventing Tau phosphorylation by targeting GSK-3 $\beta$ .

## 2. Tau phosphorylation in the AD brain

Tau from the AD brain contains eleven Ser/Thr-Pro sites, that are phosphorylated by proline-directed kinases, and nine Ser/Thr-X sites, that are phosphorylated by non-proline-directed kinases. GSK-3 $\beta$ , Cdk5/p25, ERK2, and p70-S6K belong to proline-directed kinases, which phosphorylate Tau at Thr181, Ser202/T205, Thr212/S214,

Thr231/Ser235, and Ser396/Ser404 on Ser-Pro or Thr-Pro motifs in the regions flanking the repeat domains [11-13]. MARK, SADK, PKA, and CaMKII belong to non-proline-directed kinases, which phosphorylate Tau at Ser262, Ser 320, Ser324, and Ser356 on KXGS motifs in the repeat domains (R1-R4) [12,14,15]. The non-receptor tyrosine kinases (non-RTKs) Fyn and c-Abl phosphorylate Tau at Tyr-18 and Tyr-394 [12].

GSK-3 $\beta$  is enriched in the brain and preferentially expressed in the hippocampus. GSK-3 $\beta$  acts as the main executioner of Tau phosphorylation in PHFs [16,17]. Intriguingly, GSK-3 accelerates the rate of Tau phosphorylation several-fold, if Tau is pre-phosphorylated by priming kinases such as non-proline-directed kinases [18-20]. Of Tau phosphorylation sites, Ser396 phosphorylation is a key step in the PHF formation [21]. Once a priming kinase phosphorylates Tau at Ser404, GSK-3 $\beta$  phosphorylates Tau at Ser400, followed by sequential phosphorylation of Ser396 [21]. GSK-3 $\beta$ , alternatively, phosphorylates Tau at Ser202 directly, but Thr231 phosphorylation requires for Ser235 pre-phosphorylation [21].

### 3. Interaction between A $\beta$ and GSK-3 $\beta$

GSK-3 $\beta$  is originally in the active form. GSK-3 $\beta$  is inactivated by being phosphorylated at Ser9 and activated by being phosphorylated at Tyr216 [22].

A $\beta$  activates the non-RTK Fyn, that in turn, activates GSK-3 $\beta$  by phosphorylating at Tyr216, leading to Tau phosphorylation and accumulation in neurons [23,24]. A $\beta$ , alternatively, inhibits/inactivates phosphoinositide 3-kinase (PI3K), thereby suppressing GSK-3 $\beta$ -Ser9 phosphorylation, to activate GSK-3 $\beta$  [25]. Chronic exposure of A $\beta$  downregulates Akt phosphorylation, to activate GSK-3 $\beta$  and increase Tau phosphorylation [26]. Soluble A $\beta$  oligomers inhibit insulin signaling relevant to Akt activation, to activate GSK-3 $\beta$  and increase Tau phosphorylation [27]. Intracellular A $\beta$ <sub>1-42</sub> promotes Tau phosphorylation and induces neuronal loss [28]. GSK-3 $\beta$  exacerbates A $\beta$ -induced neurotoxicity and cell death [29].

Amyloid precursor protein (APP) intracellular domain (AICD), that is produced from  $\gamma$ -secretase-mediated APP cleavage, activates GSK-3 $\beta$  [30] or enters the nucleus and activates gene transcription, increasing the GSK-3 $\beta$  mRNA and protein [31]. C-terminal fragments of APP stimulate GSK-3 $\beta$  activation, to increase Tau phosphorylation and induce apoptosis [32].

Aging, inflammation, and stress also activate GSK-3 $\beta$ , causing an initial Tau phosphorylation, responsible for mild cognitive impairment (MCI), a preliminary group of AD. A $\beta$  further activates GSK-3 $\beta$  and accelerates Tau phosphorylation, leading to progression into

AD from MCI [33,34].

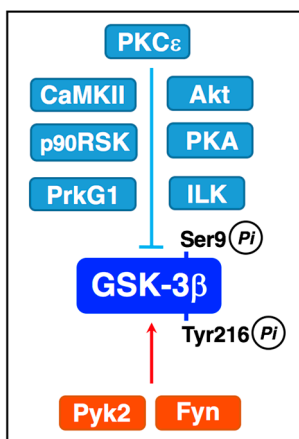
#### 4. Regulation of GSK-3 $\beta$ activity

The serine/threonine protein kinases such as PKC $\epsilon$  [35], Akt [35], PKA [36], integrin-linked kinase (ILK) [37], CaMKII [38], p90 ribosomal protein S6 kinase (p90RSK) [39], and cGMP-dependent protein kinase 1 (PrkG1) [40] inactivate GSK-3 $\beta$  by directly phosphorylating at Ser9 (Figure 1). Conversely, inhibition of those kinases allows relative activation of GSK-3 $\beta$ .

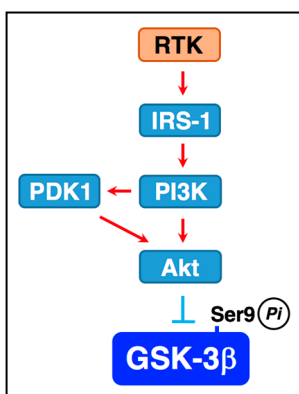
Pyk2 binds to and activates SH2 and SH3 domain-containing proteins like Src kinases. Pyk2 and Fyn activate GSK-3 $\beta$  by phosphorylating at Tyr216 directly [23,41] (Figure 1).

Akt1 is activated by being phosphorylated at Thr308 and Ser473 through the major pathway along a RTK/insulin receptor substrate 1 (IRS-1)/PI3K/3-phosphoinositide-dependent protein kinase 1 (PDK1)/Akt axis, to phosphorylate GSK-3 $\beta$  at Ser9 and inactivate GSK-3 $\beta$  [35] (Figure 2). Accordingly, activation of RTK induces inactivation of GSK-3 $\beta$ ; conversely, inhibition of RTK induces activation of GSK-3 $\beta$ . For example, insulin or insulin-like growth-factor 1 (IGF1) binds to and activates the RTK insulin receptor involving GSK-3 $\beta$  inactivation, which occurs still in the brain.

Lines of evidence have pointed to other pathways for regulation of GSK-3 $\beta$  activity. AMP-activated protein kinase (AMPK) phosphorylates and inactivates GSK-3 $\beta$  [42], while AMPK by itself phosphorylates Tau at Ser262 [43]. A $\beta$ <sub>1-42</sub> upregulates expression of adenylate kinase-1 (AK1), that inhibits AMPK, causing GSK-3 $\beta$  activation [43]. AMPK is activated due to inhibition of protein phosphatase 2A (PP2A) through a sphingosine-1-phosphate receptor 1 (S1PR1) linked to G $_i$  protein [44], thereby inactivating GSK-3 $\beta$ .



**Figure 1.** Protein kinases bearing phosphorylation of GSK-3 $\beta$ . PKC $\epsilon$ , Akt, PKA, ILK, CaMKII, p90RSK, and PrkG1 phosphorylate GSK-3 $\beta$  at Ser9 and inactivate GSK-3 $\beta$ . Pyk2 and Fyn phosphorylate GSK-3 $\beta$  at Tyr216 and activate GSK-3 $\beta$ .



**Figure 2.** RTK-mediated GSK-3 $\beta$  inactivation. Akt is activated through a pathway along a RTK/IRS-1/PI3K/PDK1/Akt axis and inactivates GSK-3 $\beta$  by phosphorylating at Ser9.

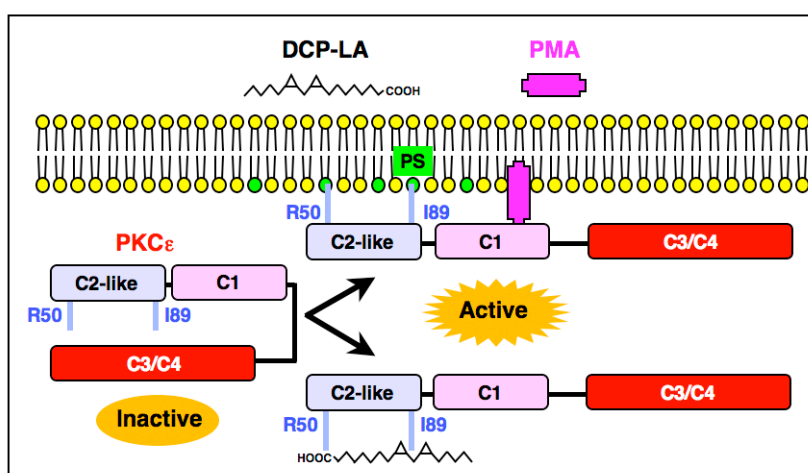
## 5. Linoleic acid derivative DCP- LA

PKC is classified into the conventional PKC isozymes  $\alpha$ ,  $\beta$ I,  $\beta$ II, and  $\gamma$ , the novel PKC isozymes  $\delta$ ,  $\epsilon$ ,  $\eta$ , and  $\theta$ , the atypical PKC isozymes  $\iota/\lambda$  and  $\zeta$ , and the PKC-like isozymes  $\mu$  and  $\nu$ . All the PKCs have the phosphatidylserine (PS) binding site and are activated by diacylglycerol (DG) [45]. Of the PKC isozymes *cis*-unsaturated free fatty acids (uFFAs) such as arachidonic, linoleic, linolenic, oleic, and docosahexaenoic acid directly interact with the novel PKC isozymes [45]. uFFAs are very unstable, and therefore, we have originally synthesized the linoleic acid derivative 8-[2-(2-pentyl-cyclopropylmethyl)-cyclopropyl]-octanoic acid (DCP-LA),

with cyclopropane rings instead of *cis*-double bonds, that exhibits stable bioactivities [46].

The primary site of action of DCP-LA is PKC $\epsilon$ . DCP-LA activates PKC $\epsilon$  selectively and directly still in the absence of DG and calcium [47]. DCP-LA binds to the PS binding/associating sites Arg50 and Ile89 in the C2-like domain of PKC $\epsilon$  at the carboxyl-terminal end and the cyclopropane rings, respectively, which are distinct from the phorbol 12-myristate 13-acetate (PMA) binding site in the C1 domain [48] (Figure 3).

DCP-LA, on the other hand, acts as a potent inhibitor of protein tyrosine phosphatase 1B (PTP1B) through its direct interaction [49].



**Figure 3.** A schematic diagram for DCP-LA- and PMA-induced PKC $\epsilon$  activation. Inactive form of PKC $\epsilon$  binds to PS in the inner leaflet of the lipid bilayer at Arg50 and Ile89, to make PKC $\epsilon$  an open frame and activate PKC $\epsilon$  partially, allowing PMA to bind to the C1 domain and activate PKC $\epsilon$  fully. In contrast, DCP-LA activates PKC $\epsilon$  in the cytosol by directly interacting with Arg50 and Ile89 in the C2-like domain, regardless of PS.

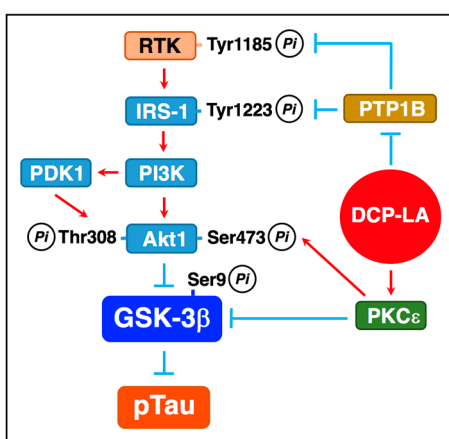
## 6. DCP-LA inactivates GSK-3 $\beta$ by cooperation of PKC $\epsilon$ activation and PTP1B inhibition

PKC $\epsilon$ , activated by DCP-LA, inactivates GSK-3 $\beta$  by directly phosphorylating at Ser9 (Figure 4) [35]. Activated PKC $\epsilon$ , alternatively, activates Akt by directly phosphorylating at the serine residue, followed by inactivation of GSK-3 $\beta$  [35] (Figure 4).

When activated, RTK phosphorylates its own receptor at Tyr1185 and activates IRS-1 by phosphorylating at Tyr1222. Activated IRS-1 recruits and activates PI3K, which produces phosphatidylinositol 3,4,5-triphosphate (PIP $_3$ ) by phosphorylating phosphatidylinositol 4,5-bisphosphate (PIP $_2$ ). PIP $_3$  binds to and activates PDK1. PI3K and/or PDK1 activate Akt by

phosphorylating at the serine and threonine residues. RTK and IRS-1 are inactivated through PTP1B-mediated tyrosine dephosphorylation. DCP-LA-induced PTP1B inhibition, therefore, represses inactivation of RTK and IRS-1, allowing relative Akt activation through a RTK/IRS-1/PI3K/PDK1/Akt pathway, to phosphorylate and inactivate GSK-3 $\beta$  [35] (Figure 4).

PKC $\epsilon$  activation or PTP1B inhibition is capable of inactivating GSK-3 $\beta$  each independently. In experiments using PC-12 cells, PKC $\epsilon$  overexpression and PTP1B deficiency activate Akt and inactivate GSK-3 $\beta$  synergistically [35]. This indicates that DCP-LA enables more efficient inactivation of GSK-3 $\beta$  by cooperation of PKC $\epsilon$  activation and PTP1B inhibition [35] (Figure 4).



**Figure 4.** DCP-LA-induced suppression of Tau phosphorylation. PKC $\epsilon$ , activated by DCP-LA, inactivates GSK-3 $\beta$  by phosphorylating Ser9 directly or through a PKC $\epsilon$ /Akt pathway, to restrain Tau phosphorylation (pTau). DCP-LA-induced PTP1B inhibition, alternatively, activates Akt through a RTK/IRS-1/PI3K/PDK1/Akt pathway by repressing tyrosine dephosphorylation of RTK and IRS-1, followed by GSK-3 $\beta$ -Ser9 phosphorylation and inactivation of GSK-3 $\beta$ , to restrain pTau.

## 7. DCP-LA prevents Tau phosphorylation by targeting GSK-3 $\beta$

A $\beta_{1-42}$  activates GSK-3 $\beta$  by reducing GSK-3 $\beta$ -Ser9 phosphorylation, leading to an enhancement in the Tau phosphorylation at Ser202/Thr205 and Ser396, and the A $\beta_{1-42}$ -induced Tau phosphorylation is neutralized by DCP-LA [35]. This indicates that DCP-LA prevents Tau phosphorylation by inactivating GSK-3 $\beta$  with PKC $\epsilon$  activation and PTP1B inhibition.

5xFAD mouse, as an animal model of AD, is APP/presenilin 1 (PS1) double transgenic mice that coexpress five familial forms of AD mutations such as the Swedish/London/Florida

mutations and the M146L/L286V mutations [50]. The  $A\beta_{1-42}$  levels in the 5xFAD mouse brain rise in an age-dependent manner [50]. The GSK-3 $\beta$  activity is enhanced in parallel with  $A\beta_{1-42}$  rise and Tau-Ser396 phosphorylation, responsible for PHF formation, is accelerated in the hippocampus of 5xFAD mice [51]. DCP-LA suppresses the GSK-3 $\beta$  activation and Tau-Ser396 phosphorylation in the hippocampus of 5xFAD mice [35]. DCP-LA, thus, has the potential to restrain Tau-Ser396 hyperphosphorylation efficiently by activating PKC $\epsilon$  and inhibiting PTP1B simultaneously.

#### **8. DCP-LA ameliorates cognitive disorders by facilitating hippocampal synaptic transmission**

Tau hyperphosphorylation causes tauopathies. Cognitive decline in association with tauopathies including AD, however, could not be improved only by suppression of Tau phosphorylation.

DCP-LA promotes vesicular transport of  $\alpha 7$  ACh receptor towards the cell surface in a PKC $\epsilon$ -dependent manner [52,53]. DCP-LA-induced increase in  $\alpha 7$  ACh receptor on the plasma membrane at presynaptic terminals stimulates presynaptic glutamate release [52,54]. DCP-LA, alternatively, promotes exocytosis of  $\alpha$ -amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid (AMPA) receptor, a major postsynaptic excitatory receptor, through CaMKII activation due to PP-1 inhibition [55]. Those effects of DCP-LA on  $\alpha 7$  ACh receptor and AMPA receptor induce a long-lasting facilitation of hippocampal synaptic transmission, to enhance cognitive functions [46,56].

Indeed, DCP-LA ameliorates spatial learning and memory decline in 5xFAD mice [35]. Moreover, DCP-LA improves  $A\beta_{1-40}$ - and mutant  $A\beta$ -induced spatial learning deficits in rats [57,58], scopolamine-induced spatial learning and memory impairment in rats [57], spatial learning and memory deterioration in senescence accelerated mice [59,60]. Overall, DCP-LA could improve cognitive disorders in a variety of dementias.

Interestingly, DCP-LA protects neurons from oxidative stress-induced apoptosis by inhibiting caspase-3/-9 activation [61]. This indicates that DCP-LA could also prevent progression of neuronal loss in the AD brain.

#### **9. Conclusions**



DCP-LA restrains Tau phosphorylation efficiently due to PKC $\epsilon$ -mediated direct inactivation of GSK-3 $\beta$ , to PKC $\epsilon$ /Akt-mediated inactivation of GSK-3 $\beta$ , and to RTK/IRS-1/PI3K/PDK1/Akt-mediated inactivation of GSK-3 $\beta$  in association with PTP1B inhibition. In addition, DCP-LA induces a long-lasting facilitation of hippocampal synaptic transmission, to enhance cognitive functions. Taken together, DCP-LA could not only prevent Tau phosphorylation but improve cognitive impairments associated with tauopathies. Consequently, DCP-LA could be developed as a promising drug for prevention and therapy of tauopathies.

**Conflict of Interest:** The author declares no conflict of interest.

## References

1. Drubin, D.G.; Kirschner, M.W. Tau protein function in living cells. *J. Cell. Biol.* **1986**, *103*, 2739-2746.
2. Hirokawa, N.; Takemura, R. Molecular motors and mechanisms of directional transport in neurons. *Nat. Rev. Neurosci.* **2005**, *6*, 201-214.
3. Mandelkow, E.; von Bergen, M.; Biernat, J.; Mandelkow, E.M. Structural principles of tau and the paired helical filaments of Alzheimer's disease. *Brain Pathol.* **2007**, *17*, 83-90.
4. Garcia, M.L.; Cleveland, D.W. Going new places using an old MAP: tau, microtubules and human neurodegenerative disease. *Curr. Opin. Cell Biol.* **2001**, *13*, 41-48.
5. Binder, L.I.; Guillozet-Bongaarts, A.L.; Garcia-Sierra, F.; Berry, R.W. Tau, tangles, and Alzheimer's disease. *Biochim. Biophys. Acta* **2005**, *1739*, 216-223.
6. Cuchillo-Ibanez, I.; Seereeram, A.; Byers, H.L.; Leung, K.Y.; Ward, M.A.; Anderton, B.H.; Hanger, D.P. Phosphorylation of tau regulates its axonal transport by controlling its binding to kinesin. *FASEB J.* **2008**, *22*, 3186-3195.
7. Trinczek, B.; Ebner, A.; Mandelkow, E.M.; Mandelkow, E. Tau regulates the attachment/detachment but not the speed of motors in microtubule-dependent transport of single vesicles and organelles. *J. Cell Sci.* **1999**, *112*, 2355-2367.
8. Grundke-Iqbal, I.; Iqbal, K.; Tung, Y.C.; Quinlan, M.; Wisniewski, H.M.; Binder, L.I. Abnormal phosphorylation of the microtubule-associated protein  $\tau$  (tau) in Alzheimer cytoskeletal pathology. *Proc. Natl. Acad. Sci. U.S.A.* **1986**, *83*, 4913-4917.
9. Brion, J.P.; Couck, A.M.; Passareiro, E.; Flament-Durand, J. Neurofibrillary tangles of Alzheimer's disease: an immunohistochemical study. *J. Submicrosc. Cytol.* **1985**, *17*, 89-96.
10. Kneynsberg, A.; Combs, B.; Christensen, K.; GerardoMorfini, G.; Nicholas M. Kanaan, N.M. Axonal degeneration in tauopathies: disease relevance and underlying mechanisms. *Front. Neurosci.* **2017**, *11*, 572.
11. Patrick, G.N.; Zukerberg, L.; Nikolic, M.; de la Monte, S.; Dikkes, P.; Tsai, L.H. Conversion of p35 to p25 deregulates Cdk5 activity and promotes neurodegeneration. *Nature* **1999**, *402*, 615-622.
12. Jeganathan, S.; Hascher, A.; Chinnathambi, S.; Biernat, J.; Mandelkow, E.M.; Mandelkow, E. Proline-directed pseudo-phosphorylation at AT8 and PHF1 epitopes induces a compaction of the paperclip folding of Tau and generates a pathological (MC-1) conformation. *J. Biol. Chem.* **2008**, *283*, 32066-32076.
13. Pei, J.J.; Björkdahl, C.; Zhang, H.; Zhou, X.; Winblad, B. p70 S6 kinase and tau in

- Alzheimer's disease. *J. Alzheimers Dis.* **2008**, *14*, 385-392.
14. Ren, Q.G.; Wang, Y.J.; Gong, W.G.; Wu, D.; Tang, X.; Li, X.L.; Wu, F.F.; Bai, F.; Xu, L.; Zhang, Z.J. Escitalopram ameliorates Tau hyperphosphorylation and spatial memory deficits induced by protein kinase A activation in Sprague Dawley rats. *J. Alzheimers Dis.* **2015**, *47*, 61-71.
  15. Yamamoto, H.; Yamauchi, E.; Taniguchi, H.; Ono, T.; Miyamoto, E. Phosphorylation of microtubule-associated protein tau by Ca<sup>2+</sup>/calmodulin-dependent protein kinase II in its tubulin binding sites. *Arch Biochem. Biophys.* **2002**, *408*, 255-262.
  16. Hernandez, F.; Lucasa, J.J.; Avila, J. GSK3 and tau: two convergence points in Alzheimer's disease. *J. Alzheimers Dis.* **2013**, *33 Suppl 1*, S141-144.
  17. Imahori K. The biochemical study on the etiology of Alzheimer's disease. *Proc. Jpn. Acad. Ser. B. Phys. Biol. Sci.* **2010**, *86*, 54-61.
  18. Singh, T.J.; Haque, N.; Grundke-Iqbal, I.; Iqbal, K. Rapid Alzheimer-like phosphorylation of tau by the synergistic actions of non-proline-dependent protein kinases and GSK-3. *FEBS Lett.* **1995**, *358*, 267-272.
  19. Cohen, P.; Frame, S. The renaissance of GSK3. *Nat. Rev. Mol. Cell Biol.* **2001**, *2*, 769-776.
  20. Beurel, E.; Grieco, S.F.; Jope, R.S. Glycogen synthase kinase-3 (GSK3): regulation, actions, and diseases. *Pharmacol. Ther.* **2015**, *148*, 114-131.
  21. Li, T.; Paudel, H.K. Glycogen synthase kinase 3 $\beta$  phosphorylates Alzheimer's disease-specific Ser396 of microtubule-associated protein tau by a sequential mechanism. *Biochemistry* **2006**, *45*, 3125-3133.
  22. Forde, J.E.; Dale, T.C. Glycogen synthase kinase 3: a key regulator of cellular fate. *Cell. Mol. Life Sci.* **2007**, *64*, 1930-1944.
  23. Li, C.; Götz, J. Somatodendritic accumulation of Tau in Alzheimer's disease is promoted by Fyn-mediated local protein translation. *EMBO J.* **2017**, pii:e201797724.
  24. Hu, M.; Waring, J.F.; Gopalakrishnan, M.; Li, J. Role of GSK-3 $\beta$  activation and  $\alpha$ 7 nAChRs in A $\beta$ <sub>1-42</sub>-induced tau phosphorylation in PC12 cells. *J. Neurochem.* **2008**, *106*, 1371-1377.
  25. Takashima, A. GSK-3 $\beta$  and memory formation. *Front. Mol. Neurosci.* **2012**, *5*, 47.
  26. Abbott, J.; Howlett, D.R.; Francis, P.T.; Williams, R.J. A $\beta$ <sub>1-42</sub> modulation of Akt phosphorylation via  $\alpha$ 7 nAChR and NMDA receptors. *Neurobiol. Aging* **2008**, *29*, 992-1001.
  27. Tokutake, T.; Kasuga, K.; Yajima, R.; Sekine, Y.; Tezuka, T.; Nishizawa, M.; Ikeuchi T. Hyperphosphorylation of Tau induced by naturally secreted amyloid- $\beta$  at nanomolar

- concentrations is modulated by insulin-dependent Akt-GSK3 $\beta$  signaling pathway. *J. Biol. Chem.* **2012**, *287*, 35222-35233.
28. Rebeck, G.W.; Hoe, H.S.; Moussa, C.E.  $\beta$ -Amyloid<sub>1-42</sub> gene transfer model exhibits intraneuronal amyloid, gliosis, tau phosphorylation, and neuronal loss. *J. Biol. Chem.* **2010**, *285*, 7440-7446.
  29. Takashima, A.; Noguchi, K.; Sato, K.; Hoshino, T.; Imahori, K. Tau protein kinase I is essential for amyloid  $\beta$  protein-induced neurotoxicity. *Proc. Natl. Acad. Sci. U.S.A.* **1993**, *90*, 7789-7793.
  30. Ryan, K.A.; Pimplikar, S.W. Activation of GSK-3 and phosphorylation of CRMP2 in transgenic mice expressing APP intracellular domain. *J. Cell Biol.* **2005**, *171*, 327-335.
  31. Balaraman, Y.; Limaye, A.R.; Levey, A.I.; Srinivasan, S. Glycogen synthase kinase 3 $\beta$  and Alzheimer's disease: pathophysiological and therapeutic significance. *Cell. Mol. Life Sci.* **2006**, *63*, 1226-1235.
  32. Kim, H.S.; Kim, E.M.; Lee, J.P.; Park, C.H.; Kim, S.; Seo, J.H.; Chang, K.A.; Yu, E.; Jeong, S.J.; Chong, Y.H.; Suh, Y.H. C-terminal fragments of amyloid precursor protein exert neurotoxicity by inducing glycogen synthase kinase-3 $\beta$  expression. *FASEB J.* **2003**, *17*, 1951-1953.
  33. Ballatore, C.; Lee, V.M.; Trojanowski, J.Q. Tau-mediated neurodegeneration in Alzheimer's disease and related disorders. *Nat. Rev. Neurosci.* **2007**, *8*, 663-672.
  34. Hurtado, D.E.; Molina-Porcel, L.; Iba, M.; Aboagye, A.K.; Paul, S.M.; Trojanowski, J.Q.; Lee, V.M. A $\beta$  accelerates the spatiotemporal progression of tau pathology and augments tau amyloidosis in an Alzheimer mouse model. *Am. J. Pathol.* **2010**, *177*, 1977-1988.
  35. Kanno, T.; Tsuchiya, A.; Tanaka, A.; Nishizaki, T. Combination of PKC $\epsilon$  activation and PTP1B inhibition effectively suppresses A $\beta$ -induced GSK-3 $\beta$  activation and Tau phosphorylation. *Mol. Neurobiol.* **2016**, *53*, 4787-4797.
  36. Shelly, M.; Lim, B.K.; Cancedda, L.; Heilshorn, S.C.; Gao, H.; Poo, M.M. Local and long-range reciprocal regulation of cAMP and cGMP in axon/dendrite formation. *Science* **2010**, *327*, 547-552.
  37. Naska, S.; Park, K.J.; Hannigan, G.E.; Dedhar, S.; Miller, F.D.; Kaplan, D.R. An essential role for the integrin-linked kinase-glycogen synthase kinase-3 $\beta$  pathway during dendrite initiation and growth. *J. Neurosci.* **2006**, *26*, 13344-13356.
  38. Song, B.; Lai, B.; Zheng, Z.; Zhang, Y. Luo, J.; Wang, C.; Chen, Y.; Woodgett, J.R.; Li, M. Inhibitory phosphorylation of GSK-3 by CaMKII couples depolarization to neuronal

- survival. *J. Biol. Chem.* **2010**, *285*, 41122-41134.
39. Valerio, A.; Ghisi, V.; Dossena, M.; Tonello, C.; Giordano, A.; Frontini, A.; Ferrario, M.; Pizzi, M.; Spano, P.; Carruba, M.O.; Nisoli, E. Leptin increases axonal growth cone size in developing mouse cortical neurons by convergent signals inactivating glycogen synthase kinase-3 $\beta$ . *J. Biol. Chem.* **2006**, *281*, 12950-12958.
  40. Zhao, Z.; Wang, Z.; Gu, Y.; Feil, R.; Hofmann, F.; Ma, L. Regulate axon branching by the cyclic GMP pathway via inhibition of glycogen synthase kinase 3 in dorsal root ganglion sensory neurons. *J. Neurosci.* **2009**, *29*, 1350-1360.
  41. Sayas, C.L.; Ariaens, A.; Ponsioen, B.; Moolenaar, W.H. GSK-3 is activated by the tyrosine kinase Pyk2 during LPA1-mediated neurite retraction. *Mol. Biol. Cell* **2006**, *17*, 1834-1844.
  42. Song, J.S.; Kim, E.K. Choi, Y.W.; Oh, W.K.; Kim, Y.M. Hepatocyte-protective effect of nectandrin B, a nutmeg lignan, against oxidative stress: Role of Nrf2 activation through ERK phosphorylation and AMPK-dependent inhibition of GSK-3 $\beta$ . *Toxicol. Appl. Pharmacol.* **2016**, *307*, 138-149.
  43. Galasso, A.; Cameron, C.S.; Frenguelli, B.G.; Moffat, K.G. An AMPK-dependent regulatory pathway in tau-mediated toxicity. *Biol. Open* **2017**, pii: bio.022863.
  43. Park, H.; Kam, T.I.; Kim, Y.; Choi, H.; Gwon, Y.; Kim, C.; Koh, J.Y.; Jung, Y.K. Neuropathogenic role of adenylate kinase-1 in A $\beta$ -mediated tau phosphorylation via AMPK and GSK3 $\beta$ . *Hum. Mol. Genet.* **2012**, *21*, 2725-2737.
  44. St-Cyr Giguère, F.; Attiori Essis, S.; Chagniel, L.; Germain, M.; Cyr, M.; Massicotte, G. The sphingosine-1-phosphate receptor 1 agonist SEW2871 reduces Tau-Ser262 phosphorylation in rat hippocampal slices. *Brain Res.* **2017**, *1658*, 51-59.
  45. Steinberg, S.F. Structural basis of protein kinase C isoform function. *Physiol. Rev.* **2008**, *88*, 1341-1378.
  46. Tanaka, A.; Nishizaki, T. The newly synthesized linoleic acid derivative FR236924 induces a long-lasting facilitation of hippocampal neurotransmission by targeting nicotinic acetylcholine receptors. *Bioorg. Med. Chem. Lett.* **2003**, *13*, 1037-1040.
  47. Kanno, T.; Yamamoto, H.; Yaguchi, T.; Hi, R.; Mukasa, T.; Fujikawa, H.; Nagata, T.; Yamamoto, S.; Tanaka, A.; Nishizaki, T. The linoleic acid derivative DCP-LA selectively activates PKC- $\epsilon$ , possibly binding to the phosphatidylserine binding site. *J. Lipid Res.* **2006**, *47*, 1146-1156.
  48. Kanno, T.; Tsuchiya, A.; Shimizu, T.; Mabuchi, M.; Tanaka, A.; Nishizaki, T. DCP-LA activates cytosolic PKC $\epsilon$  by interacting with the phosphatidylserine binding/associating

- sites Arg50 and Ile89 in the C2-like domain. *Cell. Physiol. Biochem.* **2015**, *37*, 193-200.
49. Tsuchiya, A.; Kanno, T.; Nagaya, H.; Shimizu, T.; Tanaka, A.; Nishizaki, T. PTP1B inhibition causes Rac1 activation by enhancing receptor tyrosine kinase signaling. *Cell. Physiol. Biochem.* **2014**, *33*, 1097-1106.
50. Oakley, H.; Cole, S.L.; Logan, S.; Maus, E.; Shao, P.; Craft, J.; Guillozet-Bongaarts, A.; Ohno, M.; Disterhoft, J.; Van Eldik, L.; Berry, R.; Vassar, R. Intraneuronal  $\beta$ -amyloid aggregates, neurodegeneration, and neuron loss in transgenic mice with five familial Alzheimer's disease mutations: potential factors in amyloid plaque formation. *J. Neurosci.* **2006**, *26*, 10129-40.
51. Kanno, T.; Tsuchiya, A.; Nishizaki, T. Hyperphosphorylation of Tau at Ser396 occurs in the much earlier stage than appearance of learning and memory disorders in 5XFAD mice. *Behav. Brain Res.* **2014**, *274*, 302-306.
52. Kanno, T.; Tanaka, A.; Nishizaki, T. Linoleic acid derivative DCP-LA stimulates vesicular transport of  $\alpha 7$  ACh receptors towards surface membrane. *Cell. Physiol. Biochem.* **2012**, *30*, 75-82.
53. Kanno, T.; Tsuchiya, A.; Tanaka, A.; Nishizaki, T. The linoleic acid derivative DCP-LA increases membrane surface localization of the  $\alpha 7$  ACh receptor in a protein 4.1N-dependent manner. *Biochem. J.* **2013**, *450*, 303-309.
54. Shimizu, T.; Kanno, T.; Tanaka, A.; Nishizaki, T.  $\alpha, \beta$ -DCP-LA selectively activates PKC- $\epsilon$  and stimulates neurotransmitter release with the highest potency among 4 diastereomers. *Cell. Physiol. Biochem.* **2011**, *27*, 149-158.
55. Kanno, T.; Yaguchi, T.; Nagata, T.; Tanaka, A.; Nishizaki, T. DCP-LA stimulates AMPA receptor exocytosis through CaMKII activation due to PP-1 inhibition. *J. Cell. Physiol.* **2009**, *221*, 183-188.
56. Yamamoto, S.; Kanno, T.; Nagata, T.; Yaguchi, T.; Tanaka, A.; Nishizaki, T. The linoleic acid derivative FR236924 facilitates hippocampal synaptic transmission by enhancing activity of presynaptic  $\alpha 7$  acetylcholine receptors on the glutamatergic terminals. *Neuroscience* **2005**, *130*, 207-213.
57. Nagata, T.; Yamamoto, S.; Yaguchi, T.; Iso, H.; Tanaka, A.; Nishizaki, T. The newly synthesized linoleic acid derivative DCP-LA ameliorates memory deficits in animal models treated with amyloid- $\beta$  peptide and scopolamine. *Psychogeriatrics* **2005**, *5*, 122-126.
58. Nagata, T.; Tomiyama, T.; Mori, H.; Yaguchi, T.; Nishizaki, T. DCP-LA neutralizes mutant amyloid  $\beta$  peptide-induced impairment of long-term potentiation and spatial learning.

- Behav. Brain Res.* **2010**, *206*, 151-154.
59. Yaguchi, T.; Nagata, T.; Mukasa, T.; Fujikawa, H.; Yamamoto, H.; Yamamoto, S.; Iso, H.; Tanaka, A.; Nishizaki, T. Linoleic acid derivative DCP-LA improves learning impairment in SAMP8. *Neuroreport* **2006**, *17*, 105-108.
60. Kanno, T.; Yaguchi, T.; Shimizu, T.; Tanaka, A.; Nishizaki, T. 8-[2-(2-Pentyl-cyclopropylmethyl)-cyclopropyl]-octanoic acid and its diastereomers improve age-related cognitive deterioration. *Lipids* **2012**, *47*, 687-695.
61. Yaguchi, T.; Fujikawa, H.; Nishizaki, T. Linoleic acid derivative DCP-LA protects neurons from oxidative stress-induced apoptosis by inhibiting caspase-3/-9 activation. *Neurochem. Res.* **2010**, *35*, 712-717.