

1 Article

## 2 A Novel Approach of Ultraviolet Germicidal Irradiation 3 to Reduce Air Pollution in Indoor Environments

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11

12 **Abstract:** This study examined the use of high dosages of ultraviolet germicidal irradiation (UVGI) (253.7  
13 nm) to deal with various concentrations of air pollutants, such as formaldehyde (HCHO), total volatile organic  
14 compounds (TVOC), under various conditions of humidity. We also estimated the emission of ozone as a  
15 secondary pollutant of UVGI as treatment. A number of irradiation methods were applied for various durations  
16 in field studies to examine the efficiency of removing HCHO, TVOC, bacteria, and fungi. The removal  
17 efficiency of air pollutants (HCHO and bacteria) through long-term exposure to UVGI appears to increase with  
18 time. The effects on TVOC and fungi concentration were insignificant in the first week; however, improvements  
19 were observed in the second week. No differences were observed among the various irradiation methods in this  
20 study regarding the removal of HCHO and TVOC; however significant differences were observed in the  
21 removal of bacteria and fungi.

22

23 **Keywords:** ultraviolet radiation; bioaerosol; formaldehyde; total volatile organic compounds; indoor air  
24 quality

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### 26 1. Introduction

27 Airborne microorganisms, such as bacteria, endotoxins, fungi, fungal toxin viruses, and actinobacteria can  
28 cause allergies, irritation, and contagious diseases [1]. Ultraviolet (UV) light can be used for the removal of  
29 these pollutants, with antibacterial efficiency determined by the wavelength of the light applied. The World  
30 Health Organization defined four ranges of ultraviolet light: VUV (100-200nm), UV-C (200-280 nm), UV-B  
31 (280-320 nm) and UV-A (20-400 nm). UV-A and UV-B share a similar antibacterial mechanism, which  
32 involves breaking single strands of DNA and destroying the cell membrane in microorganisms [2,3]. However,  
33 many organisms possess a repair mechanism capable of combatting these effects. The antibacterial mechanism  
34 of UV-C comprises both physical and biochemical processes. The absorption of UV light by the DNA  
35 pyrimidine can result in the formation of pyrimidine dimers capable of altering the double helix structure of  
36 DNA, thereby interfering with DNA duplication and eventually leading to cell death [4,5,6].

37 Ultraviolet germicidal irradiation (UVGI) is generated using low pressure mercury vapor. Within the UV-  
38 C band of the electromagnetic spectrum (100-290 nm), more than 90% of the output irradiation is at a  
39 wavelength of 253.7 nm [7,8]. UVGI has been applied in disinfection and sterilization for treating water,  
40 disinfecting surfaces, and preventing the spread of disease through the air [3,9]. However, most previous studies

41 have focused on the mechanisms involved in the disinfection of microorganisms and appropriate dosages. Few  
42 researchers have addressed the issues of removal efficiency, irradiation duration, or the methods of irradiation  
43 used for particular air pollutants.

44 UVGI is generally applicable in three areas: inside the pipes used for mechanical ventilation, return air  
45 units, and any indoor area [4]. The DNA of contagious air-borne pathogens is damaged by the energy of UVGI  
46 light, which interferes with its duplication, rendering the organisms noncontagious. However, the likelihood  
47 that this damage will lead to cell death varies according to the type of organism and its exposure to UVGI [10].  
48 Unfortunately, UVGI can cause erythema, photokeratitis, and conjunctivitis. The American Conference of  
49 Governmental Industrial Hygienists (ACGIH) has established a maximum exposure limit of UVGI ( $0.2 \mu\text{W cm}^{-2}$ )  
50 for human eyes and skin, which is the equivalent of exposure for 8 hours at a threshold limit of  $6 \text{ mJ cm}^{-2}$   
51 [11]. Upward irradiation is commonly used to prevent or minimize exposure [12,13]. In such cases, organisms  
52 must be directed into upper areas by forced air (mechanical) or natural ventilation (buoyancy-driven) to  
53 facilitate disinfection by UVGI [10].

54 The mechanism by which UV light removes air pollutants is photochemical dissociation, which occurs at  
55 wavelengths ranging between 100 and 1,000 nm. This process involves the absorption of photons by molecules,  
56 resulting in the excitation of their electrons enabling them to jump from low- to high-energy states. Moving  
57 from the ground state to an excited state destabilizes the photons, resulting in the release of light or heat or a  
58 reaction with other molecules when the molecules return to their original ground state. Excited electrons can  
59 break the chemical bonds, thereby altering the physical and chemical properties of the molecule [14,15].

60 The photons associated with UV light of shorter wavelengths are more energetic, making them better able  
61 to remove air pollutants. In the direct photolysis of organic pollutants, Burrows *et al.* [16], Lin [17], and Ao *et al.* [18],  
62 demonstrated the ability of UV-C in breaking down formaldehyde and toluene molecules. However,  
63 the photolysis of 4-nitrophenol could not be achieved using UV light at a wavelength of 365 nm due to the C-  
64 N, C-C, C=C, C-H, and C-O bonds within the structure. The C-N bond (maximum wavelength of 392.7 nm)  
65 can be broken by UV light at a wavelength of 365 nm; however, the maximum wavelength of the other bonds  
66 range from 196.1 nm to 346.1 nm, rendering the UV light too weak for the direct photolysis of 4-nitrophenol  
67 [19].

68 The elimination of air pollutants by UVGI at wavelengths less than 290 nm involves direct photolysis, in  
69 which molecules that absorb light energy enter a chemically active state that breaks their chemical bonds,  
70 resulting in further dissociation reactions or the promotion of reactions with other substances [18]. In Shie *et al.*  
71 [20], it was indicated that UV light of shorter wavelengths is more efficient for the removal of formaldehyde  
72 (HCHO). The means by which photolysis occurs is determined by the chemical bonds in the molecules as well  
73 as the energy provided by UVGI. The efficiency of UVGI for the removal of pollutants is also determined by  
74 the dosage of UV light, the number of UV light lamps in a given area, and the method of irradiation as well as  
75 the relative humidity (RH), temperature, air flow, and mixing of air in the environment [21,22].

76 A review of relevant literature revealed that most research on UV lighting techniques have focused on  
77 sterilization mechanisms and the quantity of disinfectants on the surfaces of microorganisms, rather than on the  
78 efficiency with which air pollutants are removed. This study examined the efficiency of using high dosages of  
79 UVGI (253.7 nm) for the removal the air pollutants (HCHO and TVOC) of various concentrations from  
80 environmental chambers under various levels of relative humidity. We also conducted field tests to investigate  
81 the efficiency of various UVGI irradiation methods (downward irradiation, upward irradiation, and upper space  
82 irradiation) in the removal of indoor air pollutants over various periods of time. Finally, experiment results were  
83 applied to the indoor air quality mass balance (IAQMB) model to predict the concentrations of indoor air

84 pollutants following irradiation for various durations to verify the field test results. This enabled us to establish  
 85 a set of optimal operating conditions for the use of UVGI in the removal of indoor air pollutants.

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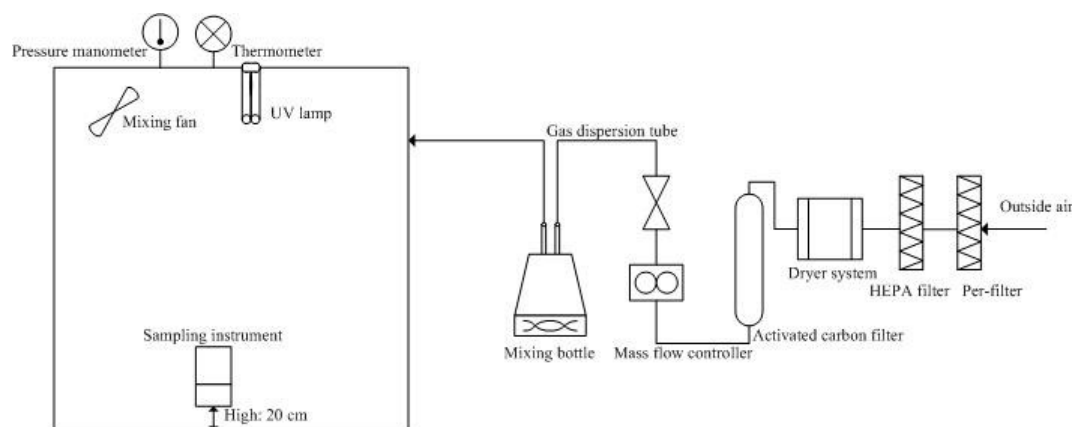
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## 88 2. Materials and Methods

### 89 2.1 UVGI experiments in environment chambers

90 This study employed UVGI lamps (XH-20, 20W-UVC) containing low-pressure mercury-vapor for the  
 91 emission of short-wave UV radiation (253.7 nm). The lamps are 13 cm long with a radius of 1.9 cm. The UVGI  
 92 lamp has been treated to block wavelength at 180 nm. The experiment was designed to explore the effects of  
 93 initial concentration and relative humidity on the removal efficiency of UVGI. The size of the stainless steel  
 94 and glass chamber was  $1.0 \text{ m} \times 1.0 \text{ m} \times 1.5 \text{ m}$  ( $1.5 \text{ m}^3$ ). Prior to the experiments, 75% ethanol was sprayed on  
 95 the inner walls of the chamber, which were then wiped with water ( $>60^\circ\text{C}$ ) to promote vaporization. We tested  
 96 the chamber for leakage and ensured that background concentrations of HCHO and TVOC were lower than  
 97 0.05 ppm by setting instruments for monitoring air quality at 20 cm above ground level. The UVGI luminaire  
 98 was installed 1 m above the air quality monitor instruments. Initially, the chamber was sampled over a 12 h  
 99 period to evaluate air quality without air exchange ( $\text{ACH}=0$ ), as shown in Fig. 1.

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Figure 1. Schematic diagram of the UVGI experiment system

103

104 HCHO was produced by dissolving 4 g of paraformaldehyde in 200 g of the distilled water. The  
 105 paraformaldehyde solution bottle was connected with zero gas to produce formaldehyde gas by aeration (10  
 106 l/min). Formaldehyde was injected into the chamber through a pipe until the concentration reached the level  
 107 required for each experiment. TVOC was produced by spraying volatile organic compounds onto a stainless  
 108 steel plate placed within an acrylic container connected with zero gas. By injecting different ratios of zero gas,  
 109 the required concentration of TVOC was obtained. The relative humidity of the chamber was controlled by  
 110 injecting various ratios of dry zero gas. The conditions used in these experiments were as follows: high ( $1.0 \pm$   
 111  $0.1$  ppm) and low ( $0.5 \pm 0.1$  ppm) initial concentrations of HCHO, high ( $3.0 \pm 0.1$  ppm) and low ( $1.4 \pm 0.1$  ppm)  
 112 initial concentrations of TVOC, and high ( $70 \pm 2\%$ ) and low ( $40 \pm 2\%$ ) relative humidity (RH) of both HCHO  
 113 and TVOC. The experiments were repeated twice for each test condition to confirm the reproducibility of the  
 114 results.

115 During the application of UV light for the removal of HCHO and TVOC, we employed a 2B Model 202  
 116 instrument with UV absorption at 254 nm (2B Technologies, Inc., USA) to measure the concentration of O<sub>3</sub>, to  
 117 confirm whether UVGI generates secondary pollutants [23].

118

## 119 2.2 UVGI experimental in field studies

120 Four sites were selected for the field tests in this study: an underground parking lot, a kitchen waste area,  
 121 an integrated *traditional Chinese* and *western medicine* clinic (Clinic A), and a *medical cosmetics clinic* (Clinic  
 122 B). Samples (two each morning and two each afternoon) were obtained from each site over the period from  
 123 November 15, 2009 to April 20, 2010 to determine the removal efficiency of UVGI. Test subjects included  
 124 TVOC, HCHO, bacteria, and fungi. Background concentrations were measured prior to the initiation of UVGI  
 125 irradiation. The UVGI irradiation plans implemented in the test sites are presented in Table 1.

126

127 Table.1 Details of building information and UVGI application for field studies.

<i>Case</i>	<i>Parking lot</i>	<i>Kitchen waste area</i>	<i>Clinic A</i>	<i>Clinic B</i>
Building age (year)	3	3	>15	<6 months
Volume (m <sup>3</sup> )	322	756	250	80
Number of population	0	0	10 - 15	10 - 15
Air ventilation type	Mechanical ventilation (air blower)		Natural ventilation	Mechanical ventilation (fan coil unit)
UVGI luminaire	Upward irradiation		Downward irradiation	Upper space irradiation
Number of UVGI lamp fixture	8	8	6	6

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129 A Formaldemeter htv-m instrument (PPM Technology, Caernarfon, UK) was used to measure HCHO,  
 130 which had been placed in a calibration tube in the test environment for at least one hour to balance its  
 131 temperature with that of the environment location, with references to the accompanying temperature and  
 132 formaldehyde concentration table to calibrate the instrument. A ppbRAE-3000 VOC monitor (RAE Systems  
 133 Inc., USA) was used to measure TVOC. To ensure the accuracy of experiment data, zero-point calibration was  
 134 performed using an activated carbon tube and span calibration using 10 ppm of isobutylene (C<sub>4</sub>H<sub>8</sub>) gas, with  
 135 calibration error less than 10%. Table 2 lists the specifications of the instruments used for measuring air  
 136 pollutants.

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Table 2. Details of instruments for indoor air quality sampler.

<i>Item</i>	<i>Instrument/Model</i>	<i>Principle</i>	<i>Detection range</i>	<i>Resolution</i>
HCHO	PPM Technology/PPM Formaldehyde htv-m	Electrochemical	0-10 ppm	0.01 ppm
TVOC	RAE/ppbRAE 3000-10.6 eV	Photo-ionization detector	1ppb~10,000 ppm	1 ppb
Ozone	2B/Model 202	UV Absorption at 254 nm	1.5ppb-100 ppb	0.1ppb
Bacteria/f ungi	Thermo/Anderson one- stage sampler	Impacting on agar with incubation (Q : 28.3 Lpm)	Stage 0 (8~24µm) Stage 1 (1~8µm)	-

140

141 Bioaerosol sampling was performed using an Andersen one-stage sampler (Thermo electron corporation,  
142 USA) with 200 holes and air throughput of 28.3 l min<sup>-1</sup>. Sampling methods for bacteria and fungi were based  
143 on standards E301.12C and E401.12 C as set out by the Environmental Protection Administration (EPA) of  
144 Taiwan [24,25]. Collected fungi were placed on malt extract agar (MEA) medium and incubated at 25 °C for 3  
145 days. Collected bacteria were placed on tryptic soy agar (TSA) medium and incubated at 30 °C for 1 day. Two  
146 duplicate samples of bacteria and fungi were also measured. The difference in flow rate (28.3 l/min), as  
147 measured using a hot wire anemometer before and after sampling, was maintained at <10% (± 2 l/min). The  
148 number of colony-forming units per cubic meter of air (CFU/m<sup>3</sup>) was calculated using Eq. (1):

149

$$\text{Bioaerosol conc. (CFU m}^{-3}\text{)} = \frac{[(\text{Avg count for stage 0}) + (\text{Avg count for stage 1})]}{\text{flow rate (28.3 l/min)} \times \text{Sampling time (L)}} \times 1000 \frac{\text{L}}{\text{m}^3} \quad (1)$$

150

### 151 2.3 UVGI removal efficiency of air pollutants

152 Removal efficiency was calculated using Eq. (2); however, the deposition and adsorption of air pollutants  
153 proved difficult to estimate in practical applications. Thus, the calculation of UVGI removal efficiency did not  
154 include the natural rate of decline in the percentage of air pollutants that can be expected to occur in the field,  
155 as shown in Eq. (3):

156

$$\text{UVGI removal efficiency}_{\text{test chamber}}(\%) = \frac{[(C_{UVGI \text{ before}} - C_{UVGI \text{ after}})/C_{UVGI \text{ before}}] - [(C_o - C_i)/C_o]}{1 - [(C_o - C_i)/C_o]} \times 100 \% \quad (2)$$

157

$$\text{UVGI removal efficiency}_{\text{field}}(\%) = \frac{(C_{UVGI \text{ before}} - C_{UVGI \text{ after}})}{C_{UVGI \text{ before}}} \times 100 \% \quad (3)$$

158

159 where  $C_{UVGI \text{ before}}$  is the concentration of air pollutants before prior to the application of UVGI,  $C_{UVGI \text{ after}}$  is  
160 the concentration of air pollutants after using UVGI,  $C_o$  is initial concentration of air pollutants, and  $C_i$  is final  
161 concentration of air pollutants.

## 162 2.4 Measurement of UV irradiance and calculation of dosages

163 We calculated the radiation view factor to estimate UVGI intensity [26], as shown in Eq. (4). The intensity  
 164 of UVGI was measured using an UV-C light meter (Lutron electronic enterprise CO., LTD.; model: UVC-254)  
 165 with a 254 nm sensor in order to quantify the difference between calculated and measured data associated with  
 166 UVGI intensity at various distances from the lamp, as shown in Fig. 2.

167

$$F = \frac{l/r}{\pi(x/r)} \left[ \frac{1}{l/r} \left( \tan^{-1} \frac{l/r}{(x/r)^2 - 1} \right) - \tan^{-1} \left( \frac{\sqrt{(x/r) - 1}}{\sqrt{(x/r) + 1}} \right) + \frac{(1 + (x/r)^2) + (l/r)^2 - 2(x/r)}{\sqrt{((1 + (x/r)^2) + (l/r)^2)((1 - (x/r)^2) + (l/r)^2)}} \right] \quad (4)$$

$$\tan^{-1} \left( \frac{\sqrt{((1 + (x/r)^2) + (l/r)^2)((x/r) - 1)}}{\sqrt{((1 - (x/r)^2) + (l/r)^2)((x/r) + 1)}} \right)$$

168

169 where  $x$  is distance from the lamp (cm),  $l$  is length of the lamp segment (cm), and  $r$  is the radius of the  
 170 lamp (cm).

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172 Irradiation intensity at any given point is determined according to surface irradiation intensity ( $I_{sur}$ ), as  
 173 shown in Eq. (5):

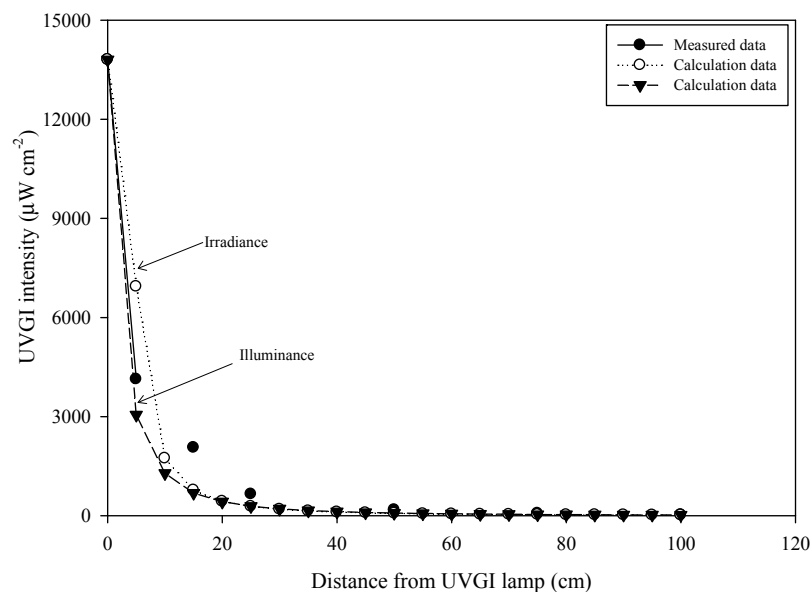
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$$I_{sur} (\mu W cm^{-2}) = \frac{E_{UV} F}{2\pi r l} \quad (5)$$

175

176 where  $I_{sur}$  is the UV intensity at  $x,y,z$  point,  $E_{UV}$  is power output of lamp ( $W cm^{-2}$ ),  $F$  is the radiation view  
 177 factor,  $r$  is the radius of the lamp, and  $l$  is the length of the lamp segment (cm).

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179

180 Figure 2. Calculated data and measured data of UVGI intensity at various distances from the UVGI lamp.

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183 The efficiency of UVGI in removing air pollutants depends heavily on whether the energy it generates is  
 184 sufficient to break the chemical bonds in question. We therefore used the Planck equation (Eq. (6)) to derive  
 185 the photon energy of UVGI at various wavelengths, which were then converted into electron volts (eV) (1J =  
 186  $6.25 \times 10^{18}$  eV).

187

$$E = hv = \frac{hc}{\lambda} \quad (6)$$

188

189 where  $E$  is the energy of a photon (J),  $h$  is the Planck constant ( $6.626 \times 10^{-34}$  J s<sup>-1</sup>),  $\nu$  is the frequency of light  
 190 (s<sup>-1</sup>),  $c$  is the speed ( $3 \times 10^8$  m s<sup>-1</sup>), and  $\lambda$  is the wavelength (nm).

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### 193 3. Results and discussion

#### 194 3.1 Efficiency of air pollutant removal under various levels of relative humidity and various initial 195 concentrations of pollutants

196 Table 3 summarizes the test results showing that in cases of high relative humidity (RH), the concentration  
 197 of HCHO was reduced from 1.0 to 0.54 ppm over a period of 12 hours, which represents a removal rate of  $15.97$   
 198  $\pm 0.03\%$ . In cases of low RH, the concentration of HCHO was reduced from 1.0 to 0.44 ppm in the same period  
 199 of time, representing a removal rate of  $32.60 \pm 0.09\%$ . In cases of high RH, TVOC concentration was reduced  
 200 from 3.0 to 2.51 ppm (removal rate of  $7.12 \pm 0.17\%$ ), and in cases of low RH, this was reduced from 3.0 to 2.36  
 201 ppm (removal rate of  $13.56 \pm 0.08\%$ ). Water molecules are able to block the partial energy of UVGI with a  
 202 consequent effect on the removal of organic substances. As a result, UVGI was shown to be more efficient in  
 203 the removal of HCHO and TVOC in cases of low RH than in cases of high RH.

204

205 Table 3. Removal efficiency of duplicate analysis for chemical air pollutants.

Item	Test condition	High conc./ Low RH		High conc./ High RH		Low conc./ High RH	
		1.0 ppm/40%RH	1.0 ppm/40%RH	1.0 ppm/70% RH	1.0 ppm/70% RH	0.5 ppm/70%RH	0.5 ppm/70%RH
HCHO	Removal efficiency (%)	#1	#2	#1	#2	#1	#2
		32.54	32.66	15.95	15.99	18.39	17.88
	Avg. $\pm$ S.D. (%)	32.6 $\pm$ 0.09		15.97 $\pm$ 0.03		18.14 $\pm$ 0.36	
	Removal deviation rate (%)	-		11.76		-	
TVOC	Test condition	3.0 ppm/40%RH		3.0 ppm/70%RH		1.4 ppm/70%RH	
	Removal efficiency (%)	13.61	13.5	7.24	7.00	6.10	5.75
	Avg. $\pm$ S.D. (%)	13.56 $\pm$ 0.08		7.12 $\pm$ 0.17		5.93 $\pm$ 0.25	
	Removal deviation rate (%)	-		4.55		-	
		-		0.84			

206 S.D., standard deviation.

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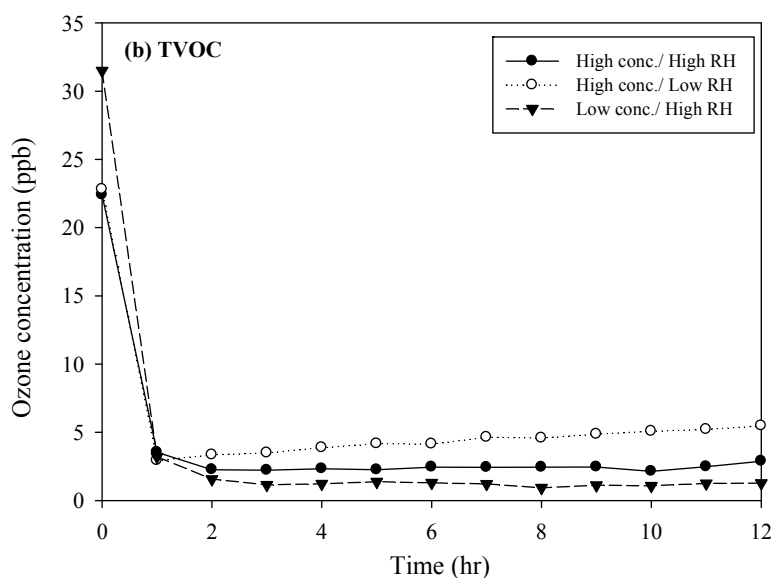
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209 The initial concentration of pollutants has a direct impact on removal efficiency. UVGI was shown to  
 210 achieve removal rates of  $15.97 \pm 0.03\%$  and  $18.14 \pm 0.36\%$  in cases with high and low concentrations of HCHO,  
 211 respectively. However, the removal rate of TVOC at high concentration was  $7.12 \pm 0.17\%$  and  $5.93 \pm 0.25\%$   
 212 for low concentrations. Thus, UVGI was shown to be more efficient in the removal of HCHO at low  
 213 concentrations and TVOC at high concentrations. Factors other than the distance to the UVGI source affect the  
 214 effectiveness of UVGI in the removal of air pollutants, including the photon energy of the UVGI as well as the  
 215 nature of the chemical bonds in the VOCs. Repeated experiments presented an error rate between 0.03% and  
 216 0.36% in the HCHO and TVOC removal rates, indicating the good reproducibility of the experiment results.

217 During the experiments, we simultaneously tested  $O_3$  concentrations to confirm whether the use of UVGI  
 218 resulted in the generation of secondary pollutants (see Figs. 3 (a) and (b)). The experiment conditions included  
 219 high formaldehyde concentration with high humidity (A), high formaldehyde concentration with low humidity  
 220 (B), and low formaldehyde concentration with high humidity (C). While gauging the removal rates, we tested  
 221 the background concentrations of  $O_3$  in the test chambers, the results of which were: (A) 20.2 ppb, (B) 20.6  
 222 ppb, and (C) 15.6 ppb, respectively. After 12 hours of continuous testing, the  $O_3$  concentrations dropped to (A)  
 223 1.3 ppb, (B) 1.3 ppb, and (C) 1.2 ppb. We also tested high concentration of TVOC with high humidity (D), high  
 224 TVOC concentration with low humidity (E), and low TVOC concentration with high humidity (F). We again  
 225 tested the background concentrations of  $O_3$  during removal, the results of which were (D) 22.4 ppb, (E) 22.8  
 226 ppb, and (F) 31.5 ppb. After 12 hours of continuous testing, the  $O_3$  concentrations dropped to (D) 2.9 ppb, (E)  
 227 5.5 ppb, and (F) 1.3 ppb.

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229

230 Figure 3. Ozone concentration during the UVGI operation (a) HCHO experiments, (b) TVOC experiments.

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232 Our results indicate that with UVGI, the  $O_3$  response mechanisms involves the absorption of UV light (200  
 233 - 280 nm) by ozone and resulting in its breaking down into oxygen molecules and atoms, thereby reducing the  
 234 concentration of  $O_3$  in the test chambers after just one or two hours of UV irradiation.

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### 238 3.2 Efficiency of air pollutant removal through long-term exposure to UVGI

239 The efficiency of chemical air pollutants removal by long-term exposure to UVGI is presented in Figs.  
240 4(a) and (b). The background concentration of HCHO was lowest in the area used for kitchen waste, presenting  
241 an average concentration of 0.04 ppm (0.01–0.06 ppm). After one week of UVGI, the average concentration of  
242 HCHO measured 0.03 ppm (0.03–0.04 ppm), indicating a removal rate of 17.1%. After two weeks of irradiation,  
243 the average concentration of HCHO declined to 0.02 ppm, representing a removal rate of 45.9%. The indoor  
244 background concentration of HCHO was relatively low; therefore the subsequent removal rates were  
245 insignificant. The average background concentrations of HCHO were 0.20 to 0.33 ppm in the underground  
246 parking lot, Clinic A, and Clinic B. The high concentrations of HCHO in the underground parking lot were due  
247 to the incomplete combustion of organic substances in the exhaust emissions of motor vehicles. Poor ventilation  
248 at the site exacerbated the accumulation of pollutants [27,28].

249 In the clinics, the fumes produced by materials used in building renovation as well as the volatile medical  
250 sterilizers used in the clinics produced considerable quantities of VOCs. After one week of UVGI, the average  
251 concentrations of HCHO in the underground parking lot, Clinic A, and Clinic B measured 0.16 to 0.27 ppm,  
252 which represent removal rates of 16.7 to 29.8%. After two weeks, the average concentrations of HCHO declined  
253 to 0.05–0.20 ppm, increasing the removal rates to 40.1–76.2%.

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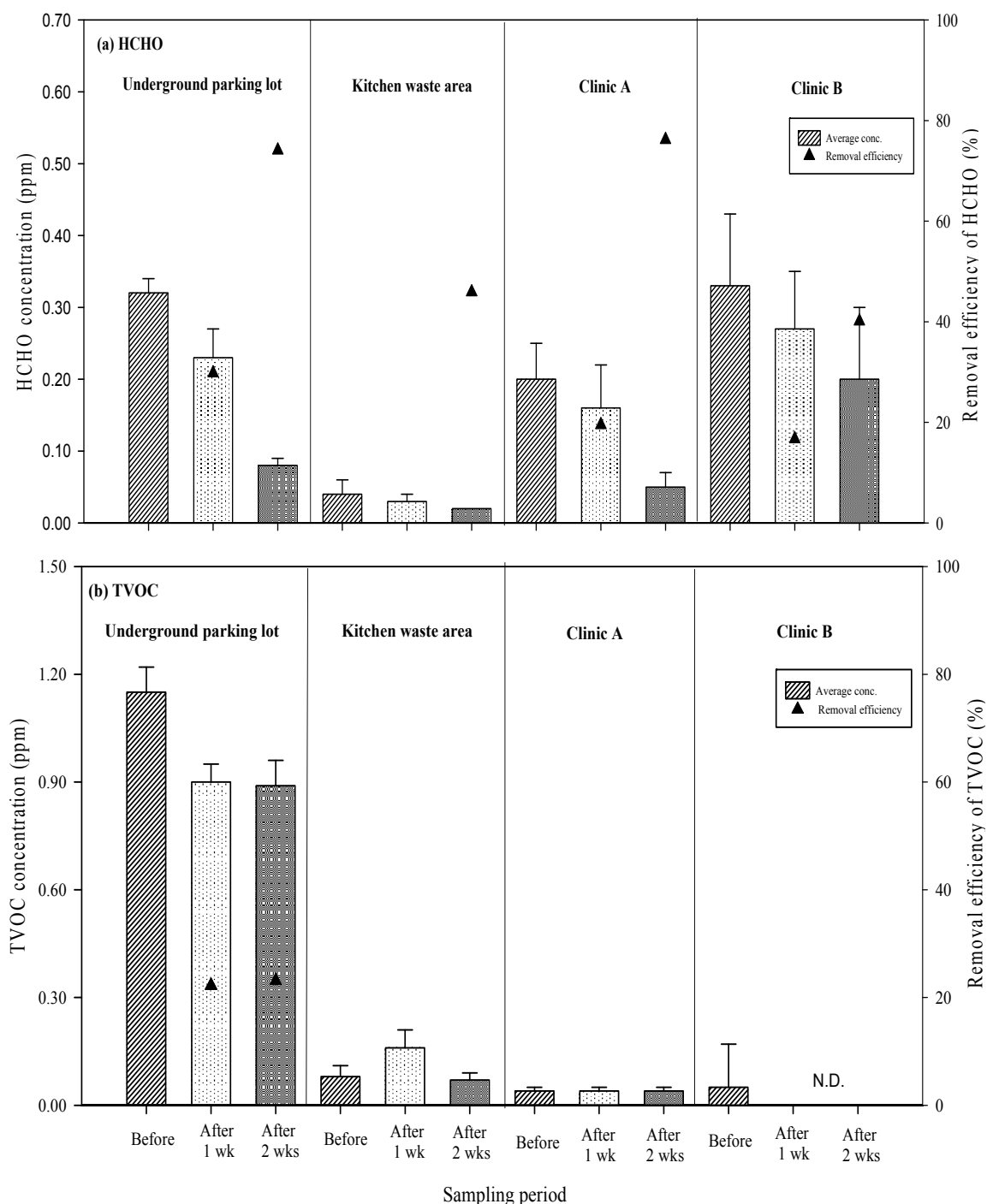


Figure 4. Efficiency of chemical air pollutants (a) HCHO, (b) TVOC removal by long-term UVGI exposure.

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281 With the exception of the underground parking lot, all of the background concentrations of TVOC were  
 282 low. After one week of UVGI treatment in the parking lot, the average concentration of TVOC measured 0.90  
 283 ppm (0.84–0.95 ppm), representing a removal rate of 22.2%. After two weeks of irradiation, the average  
 284 concentration of TVOC decreased to 0.89 ppm (0.81–0.96 ppm), for a removal rate of 23.1%. In Clinic A and  
 285 Clinic B, the average concentration of TVOC after UVGI irradiation for two weeks (<0.001–0.04 ppm) was  
 286 close to the background concentration (0.04–0.05 ppm), indicating that the amount of pollutants removed was

287 insignificant. After one week of irradiation, the average concentration of TVOC was 0.16 ppm (0.13–0.21 ppm),  
288 which was higher than the background concentration (0.08 ppm, 0.05–0.11 ppm). After the second week, the  
289 results were still close to the background concentration (0.07 ppm, 0.06–0.09 ppm). We postulate that these  
290 poor removal rates were due to the fact that this study did not focus on a single VOC.

291 The composition of TVOC varies from site to site. The photon energy (4.89 eV) produced by the UVGI  
292 irradiation in this study is sufficient to break single-bond molecules such as in C-C or C-H but not O-O, as the  
293 energy of their bonds ranges from 65.0 to 119.1 kcal/mol (equivalent to 2.8 to 5.2 eV). The photon energy of  
294 UVGI is also insufficient to break the chemical bonds of multiple-bond molecules such as C=O, C=C, and C≡C  
295 (6.3 to 8.7 eV). According to Kuo *et al.* [29] and Kim *et al.* [30], the primary constituents of VOCs from motor  
296 vehicle exhaust are toluene, benzene, xylene, and ethylbenzene. The molecular structures of these chemical  
297 substances comprise mainly C-H bonds, which require a minimum wavelength of 289.7 nm (equivalent to 4.3  
298 eV) to promote breakage. The photon energy of the UVGI in this study (4.89 eV) was higher than that required  
299 to break C-H bonds; therefore, the removal efficiency of TVOC was higher in the underground parking lot.

300 The efficiency of long-term exposure to UVGI in the removal of HCHO and TVOC was determined by  
301 comparing HCHO readings with background concentrations. After one week of UVGI irradiation, removal rates  
302 ranged from 17.1–29.8%, whereas after two weeks of UVGI irradiation produced removal rates ranging from  
303 40.1–76.2%. After the second week of UVGI, the removal rates of HCHO measured 23.4–56.7% higher than  
304 those of the first week. Formaldehyde removal by UVGI irradiation is associated with the amount of UVGI  
305 energy received by the bonds ( $\text{HCHO} + h\nu \rightarrow \text{H} + \text{HCO}\bullet$ ). The molecular formula of HCHO indicates that the  
306 C-H and C=O bonds require 98.7 kcal/mol and 176.0 kcal/mol of energy to break. This corresponds to maximum  
307 wavelengths of 289.7 nm and 162.4 nm [31], which can be respectively converted into  $6.862 \times 10^{-19}$  J and  
308  $1.223 \times 10^{-18}$  J of photon energy using the Planck equation, and are equivalent to 4.3 eV and 7.6 eV of energy.  
309 The wavelength of the UVGI in this study was 253.7 nm, which is equivalent to 4.89 eV, which is higher than  
310 the energy present in the C-H bonds in HCHO (98.7 kcal/mol; 4.3 eV). Thus, direct photolysis is able to break  
311 the C-H bonds but not the C=O bonds. As a result, the UVGI in this study was able to remove some, but not all,  
312 of the HCHO.

313 After one week of UVGI irradiation, the concentration levels of TVOC in the kitchen waste area, Clinic  
314 B, and Clinic A were either greater than or equal to the background concentrations. Only the underground  
315 parking lot displayed a positive removal rate of 22.2%. After two weeks of UVGI, the underground parking lot,  
316 kitchen waste area, Clinic A, and Clinic B displayed TVOC removal rates of 11.0 to 100%, demonstrating the  
317 effectiveness of UVGI at all four sites.

318 The respective average background concentrations of microbiological air pollutants in the underground  
319 parking lot, kitchen waste area, Clinic A, and Clinic B are presented in Figs. 5(a) and (b). After one week of  
320 UVGI, the average concentrations of bacteria measured between 277 CFU m<sup>-3</sup> and 440 CFU m<sup>-3</sup>, which indicate  
321 removal rates of 8.8 to 64.0%. After two weeks, the average bacteria concentrations measured between 145 and  
322 639 CFU m<sup>-3</sup>, which indicates removal rates of -32.7 to 84.0%. With the exception of Clinic B, the removal  
323 rates of bacteria all ranged between 47.7 % and 84.0 %.

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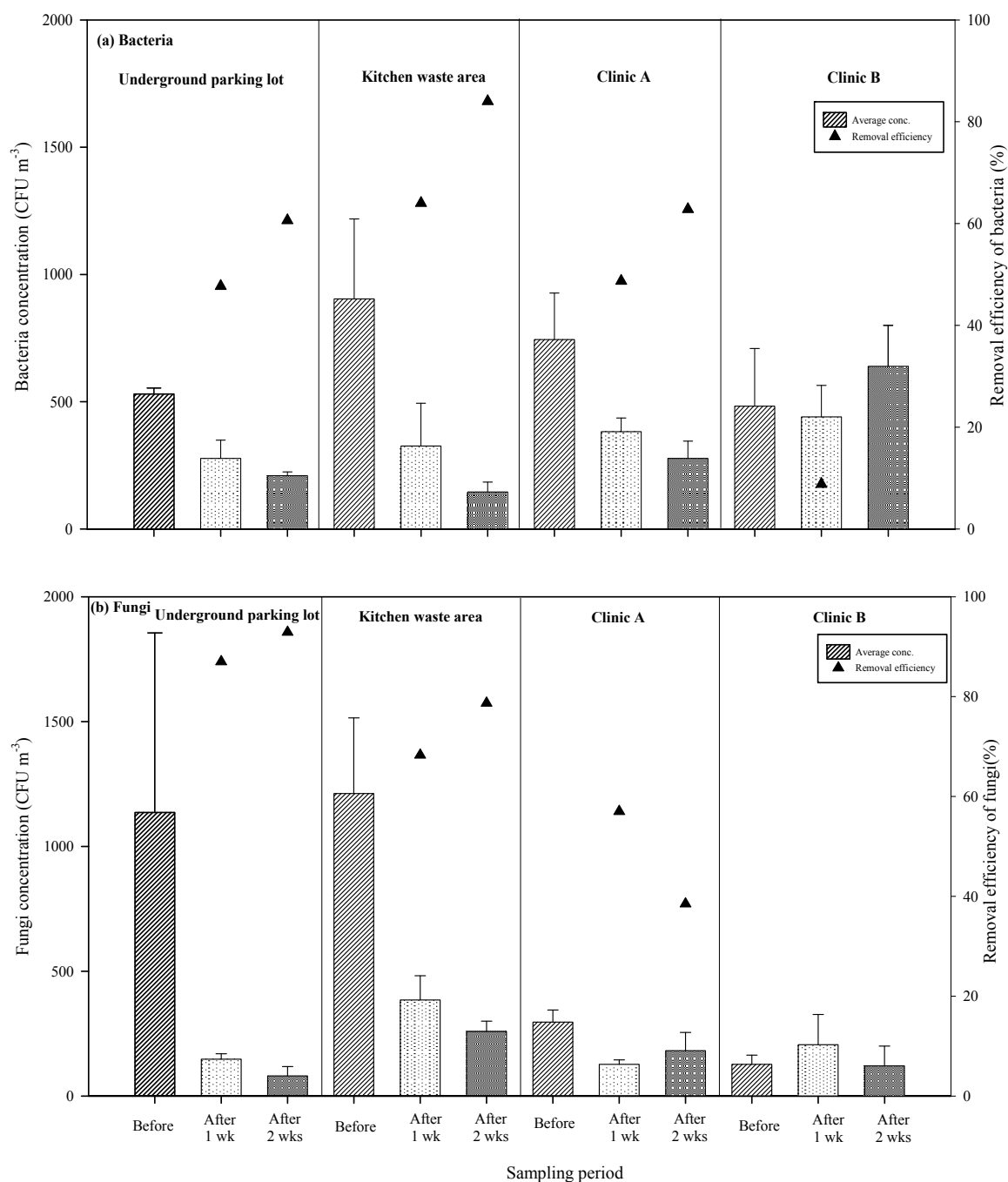


Figure 5. Efficiency of microbiological air pollutants (a) Bacteria, (b) Fungi removal by long-term UVGI exposure.

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After one week of UVGI irradiation, the average concentrations of fungi dropped significantly to 127–385 CFU m<sup>-3</sup>, which indicates removal rates of 57.0 to 87.0% in the underground parking lot, the kitchen waste area, and Clinic A; however, no significant effects were observed in Clinic B. Two weeks of UVGI irradiation lowered the average concentrations of fungi at the four sites to 81–259 CFU m<sup>-3</sup>, representing removal rates of 4.8 to 92.9%. These results indicate that with the exception of Clinic B, the fungi removal rates at all the sites ranged between 38.5% and 92.9%. The lack of effective sterilizing at Clinic B may be due to the particular strains of microorganisms at that site and/or the FCU ventilation system. The efficiency with which UV light can remove microorganisms depends on the UV dosage, irradiation time, the type of microorganism and it

339 sensitivity to UV light, and how long the microorganisms remain within the UV irradiated area.

340 With regard to the efficiency of long-term UVGI at removing bacteria and fungi, one week of UVGI  
341 reduced the background concentrations of bacteria by 8.8 to 64%, whereas two weeks of UVGI decreased the  
342 background concentrations by 60.6 to 84.0% in the kitchen waste area, underground parking lot and Clinic A.  
343 With the exception of Clinic B, the removal rates after the second week of UVGI were 12.9 to 20.0% higher  
344 than those of the first week. The poor sterilization at Clinic B may be because the strains and forms of  
345 microorganisms present at that site are less sensitive to UVGI [6]. One week of UVGI decreased the background  
346 concentrations of fungi by 57.0 to 68.3% and two weeks resulted in removal rates ranging between 4.8% and  
347 92.9%. The rate of fungi removal in the underground parking lot and the kitchen waste area increased by 5.9 to  
348 10.4% after the second week; however, in the clinics, we failed to observe any increase with time. Furthermore,  
349 the underground parking lot and the kitchen waste area were more humid than the clinics. Hydration and re-  
350 hydration can alter protein structures, thereby influencing the enzymes and nucleic acids involved in DNA  
351 repair. The hydration of biopolymer cell walls also moderates the influence of relative humidity on the  
352 sterilization effects of UVGI [32].

353 The effects of UVGI on the removal of bacteria and fungi differed slightly from those reported by  
354 Memarzadeh et al. in 2010 [33]. The cell walls of fungal spores are rigid structures, markedly different from  
355 the cell walls of prokaryotic bacteria. The DNA in the proteins of thick inner layers of chitin or cellulose can  
356 render fungi more resistant to UV light, such that higher UV doses are required for sterilization [34]. In clinic  
357 B, UVGI was shown to be inefficient in the removal of bacteria and fungi, perhaps due to the use of mechanical  
358 ventilation (a fan coil unit). The type of indoor ventilation and location of intake and exhaust ports can have a  
359 significant influence on the vertical mixing of air [21]. UVGI irradiation in Clinic A exhibited good removal  
360 efficiency with regard to bacteria but very poor removal efficiency when dealing with fungi. Open windows  
361 and doors can influence the movement of aerosols and the primary source of the fungi was the outside  
362 environment; therefore, the increase in indoor concentrations can be attributed to swift airflow preventing  
363 microorganisms from being sufficiently exposed to UVGI [35,36].

364

### 365 *3.3 Efficiency of air pollutant removal using various UVGI irradiation methods*

366 HCHO and TVOC were removed using the UVGI irradiation methods shown in Figs. 6 (a) and (b).  
367 Downward irradiation was the most effective approach to HCHO removal, followed by upward irradiation.  
368 Upper space irradiation proved the least effective. Downward irradiation for two weeks reduced background  
369 concentrations of HCHO by 76.2% (from 0.20 to 0.05 ppm), while upward irradiation reduced HCHO by 71.7%  
370 (from 0.18 to 0.05 ppm). Upper space irradiation for two weeks reduced background concentration of HCHO  
371 by 40.1 % (from 0.33 to 0.20 ppm). Starting with a TVOC background concentration of 0.05 ppm (<0.001–0.17  
372 ppm). Upper space UVGI irradiation for two weeks resulted in the total elimination of TVOC, representing a  
373 removal rate of 100%. Upward irradiation for two weeks reduced TVOC background concentrations by 22.26%  
374 (from 0.62 to 0.48 ppm). Downward irradiation for two weeks resulted in TVOC background concentrations  
375 falling negligibly from between 0.04 and 0.05 ppm to 0.04 ppm (0.03–0.05 ppm). UV photons can break C-C  
376 bonds and degrade organic substances; however, the composition of VOCs in indoor air tends to be complex.  
377 Good removal efficiency can only be achieved if the indoor TVOC have bonds that UV photons are capable of  
378 breaking. Furthermore, the efficiency with which air pollutants are removed by UV light also depends on the  
379 UV irradiation time, UV intensity, and mixing of air [37].

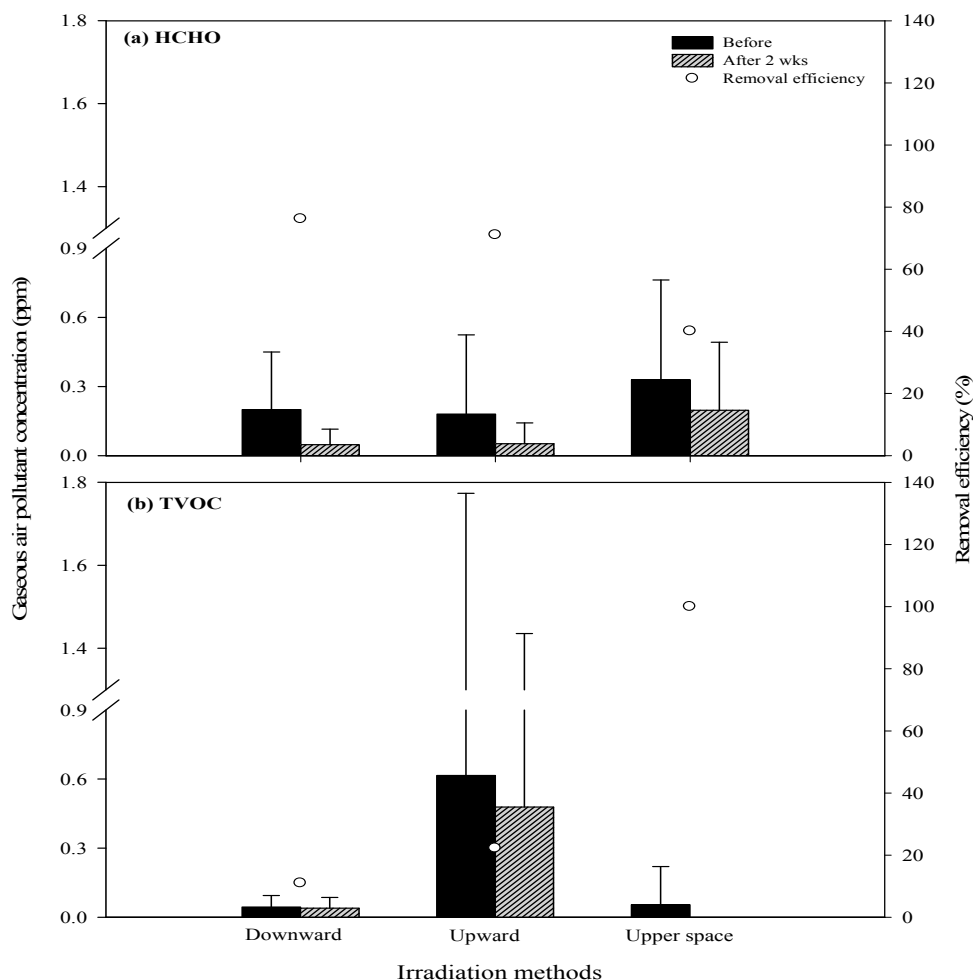


Figure 6. Efficiency of chemical air pollutants (a) HCHO, (b) TVOC removal by varying UVGI irradiation methods.

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381 Upward irradiation proved the most effective at removing microbiological air pollutants (Figs. 7 (a) and  
 382 (b)), followed by downward irradiation and upper space irradiation. Upward irradiation for two weeks reduced  
 383 the background concentrations of bacteria from 716 CFU m<sup>-3</sup> (476–1218 CFU m<sup>-3</sup>) to 177 CFU m<sup>-3</sup> (111–224  
 384 CFU m<sup>-3</sup>) and that of fungi from 1174 CFU m<sup>-3</sup> (444–1855 CFU m<sup>-3</sup>) to 169 CFU m<sup>-3</sup> (61–300 CFU m<sup>-3</sup>),  
 385 representing removal rates of 75.3% and 85.6%, respectively. Downward irradiation for two weeks reduced the  
 386 background concentration of bacteria from 744 CFU m<sup>-3</sup> (564–927 CFU m<sup>-3</sup>) to 277 CFU m<sup>-3</sup> (218–345 CFU  
 387 m<sup>-3</sup>) and that of fungi from 296 CFU m<sup>-3</sup> (255–345 CFU m<sup>-3</sup>) to 182 CFU m<sup>-3</sup> (127–255 CFU m<sup>-3</sup>), representing  
 388 removal rates of 62.8% and 38.5%, respectively. Downward irradiation for two weeks reduced the background  
 389 concentration of bacteria from 482 CFU m<sup>-3</sup> (236–709 CFU m<sup>-3</sup>) to 639 CFU m<sup>-3</sup> (436–800 CFU m<sup>-3</sup>) and that  
 390 of fungi from 127 CFU m<sup>-3</sup> (55–164 CFU m<sup>-3</sup>) to 121 CFU m<sup>-3</sup> (36–200 CFU m<sup>-3</sup>), which were higher than or  
 391 equal to the background concentrations. Sterilization involves a number of factors, including the ventilation  
 392 rate, the intensity of UV irradiation, the physiology and species of the bacteria, airflow distribution, relative  
 393 humidity, and photoreactivity [7]. Our results are consistent with the findings of Brickner *et al.* in 2003 [22],  
 394 which showed that bacteria are easier to eliminate than fungi. However, the UVGI dosage required for  
 395 sterilization varies considerably according to the microorganisms, and single-stranded nucleic acids tend to be  
 396 more sensitive to the effects of UV light than are double-stranded nucleic acids [6].

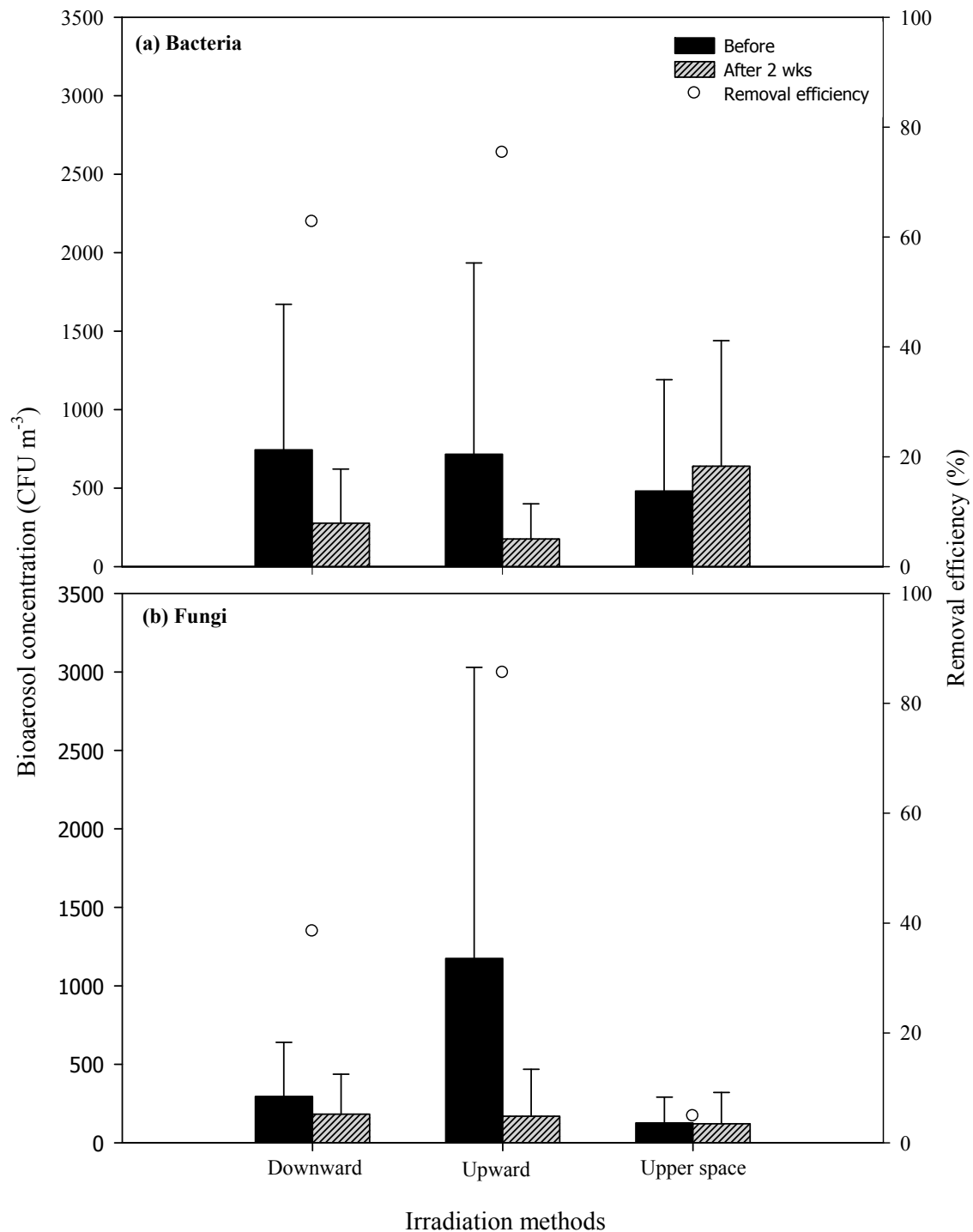


Figure 7. Efficiency of microbiological air pollutants (a) Bacteria, (b) Fungi removal by varying UVGI irradiation methods.

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With regard to the removal of HCHO or TVOC, there are no observable differences in the three UVGI methods (Table 4). In the removal of bacteria, upward irradiation (p-value: 0.031) and downward irradiation (p-value: 0.027) proved significantly more efficient than upper space irradiation. In the removal of fungi, upward irradiation is more efficient than downward irradiation (p-value: 0.007). These findings are consistent with those of Miller and MacHer [4]. A closer distance to the ceiling enables narrowband UVGI to kill the biological PMs carried to the upper space by upward airflow [38]. Downward irradiation and upward irradiation



404 are more direct than upper space irradiation with regard to the removal of air pollutants. The inefficiency of  
 405 upper space irradiation may be due to the fact that the UVGI source is placed within the FCU system in the  
 406 ceiling, which makes air mixing, particularly vertical air mixing, a critical factor. Poor convection in the indoor  
 407 airflow can prevent air pollutants from being transported to the UV irradiation area to be eliminated [39].

408

409 Table 4. Student's t test (two sample unequal variance) for various irradiation methods.

<i>Air pollutant</i>	<i>HCHO</i>	<i>TVOC</i>	<i>Bacteria</i>	<i>Fungi</i>
<i>UVGI luminaires</i>				
Upward with downward	0.606	0.212	0.500	0.007**
Upward with upper space	0.103	0.404	0.031*	0.035
Downward with upper space	0.109	0.268	0.027*	0.244

410 \* p-value < 0.05

411 \*\*p-value < 0.01

412

#### 413 4. Conclusions

414 This study tested the efficiency of UVGI in the removal of HCHO and TVOC at various concentrations  
 415 and under conditions with different levels of relative humidity. Our results indicate that removal efficiency is  
 416 higher when dealing with low concentrations of HCHO than when dealing with higher concentrations. When  
 417 dealing with TVOC, removal efficiency is higher when concentrations are higher. Removal efficiency of both  
 418 HCHO and TVOC is better in conditions of low humidity. Relative humidity produced greater fluctuations in  
 419 the removal rates than did the initial concentrations of pollutants. Under conditions of high relative humidity,  
 420 water molecules can provide a barrier to UVGI irradiation, thereby weakening its ability to break down organic  
 421 compounds. Moreover, ozone was not emitted as an air pollutant during UVGI operations in environment  
 422 chambers. This proves that the removal of HCHO and TVOC in our experiments was not achieved by the O<sub>3</sub>  
 423 produced by the UVGI.

424 The application of UVGI for one week resulted in HCHO removal rates ranging from 17.1 to 29.8%,  
 425 while treatment for two weeks resulted in removal rates between 40.1% and 76.2%. This represents an increase  
 426 of between 23.4% and 56.7%. No effects were apparent in the UVGI treatment of TVOC after the first week;  
 427 however, the effect produced noticeable results after the second week. One week of UVGI treatment produced  
 428 bacteria removal rates between 8.8% and 64%, whereas two weeks resulted in removal rates ranging from 60.6  
 429 to 84.0% (with the exception of Clinic B). The removal rates after the second week of UVGI were 12.9 to 20.0%  
 430 higher than those of the first week. After one week of UVGI, the fungi removal rates ranged between 57.0%  
 431 and 68.3%, and after two weeks, the removal rates ranged between 4.8% and 92.9%. Therefore, the removal  
 432 rates of fungi in the underground parking lot and the kitchen waste area only increased by 5.9 to 10.4% after  
 433 the second week, but those in the medical establishments did not increase.

434 No significant differences were observed in the removal rates of HCHO or TVOC, such that the  
 435 background concentrations of air pollutants were lower or close to the concentrations obtained after UV  
 436 irradiation. Upward and downward irradiation methods were shown to be considerably more efficient in the  
 437 removal of bacteria than was upper space irradiation. Upward irradiation was more efficient in the removal of  
 438 fungi than was downward irradiation. A closer distance to the ceiling made it possible for the narrowband UVGI  
 439 to kill the biological PMs carried into the upper space by upward airflow.

440

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