Image Features Based on Characteristic Curves and Local Binary Patterns for Automated HER2 Scoring

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Abstract: This paper presents novel feature descriptors and classification algorithms for automated scoring of HER2 in Whole Slide Images (WSI) of breast cancer histology slides. Since a large amount of processing is involved in analyzing WSI images, the primary design goal has been to keep the computational complexity to the minimum possible level and to use simple, yet robust feature descriptors that can provide accurate classification of the slides. We propose two types of feature descriptors that encode important information about staining patterns and the percentage of staining present in ImmunoHistoChemistry (IHC) stained slides. The first descriptor is called a characteristic curve which is a smooth non-increasing curve that represents the variation of percentage of staining with saturation levels. The second new descriptor introduced in this paper is an LBP feature curve which is also a non-increasing smooth curve that represents the local texture of the staining patterns. Both descriptors show excellent interclass variance and intraclass correlation, and are suitable for the design of automatic HER2 classification algorithms. This paper gives the detailed theoretical aspects of the feature descriptors and also provides experimental results and comparative analysis.

Keywords: medical image classification; local binary patterns; characteristic curves; whole slide image processing; automated HER2 scoring

1. Introduction

The most commonly used method for breast cancer grading is the ImmunoHistoChemistry (IHC) test which is a staining process performed on biopsy samples of breast cancer tissues [1]. The IHC stained slides are normally observed under a microscope by pathologists to determine the level of over-expression of Human Epidermal Growth factor Receptor 2 (HER2) protein in cancer cells. The tissue sample is then assigned a HER2 score of 0 to 3+ representing the grade of cancer present in the sample [2]. Manual grading and annotations of breast cancer slides are time consuming, and there are huge maintenance costs associated with collecting, archiving, and transporting tissue specimens. It is also well documented that manual grading can have significant variability in pathologist assessments due to the subjective process of determining the intensity and uniformity of staining in the presence of variable staining patterns and heterogeneity of tumor grade [3]. Automated methods can also suffer from errors due to inaccuracies in the training algorithm and its inability to segment faint and complex tissue structures [4].

In the rapidly growing field of digital pathology, several Whole Slide Image (WSI) processing algorithms are currently being developed as diagnostic tools to help pathologists in the assessment of disease patterns [5]. WSIs have a pyramidal structure to enable optimized viewing across multiple magnification levels, and they provide a high resolution overview of the entire slide [5,6]. Typically at 40x magnification, the images have a resolution of approximately 0.25 microns per pixel. At this resolution, a slide region of size 15mm x 15mm could correspond to 60,000 x 60,000 pixels. WSIs were originally used as a computer aided digital microscopy tool, where pathologists could view different parts of a sample at different magnifications to improve the accuracy of their scores [3].
Powerful computational algorithms are being developed to automatically extract features related to cytological and protein structures in the image for accurately quantifying biomarkers like HER2 [7]. In [8], the authors used adaptive thresholding and watershed algorithm for cell segmentation.

Recently, an online contest was organized by the University of Warwick in conjunction with the UK/Ireland Pathology Society annual meeting 2016, with the aim of advancing research in the field of automated HER2 scoring algorithms [9]. This contest was the primary motivation for our research work presented in this paper. Our algorithm (registered with team name UC-CSSE-CGIP) performed exceedingly well in the contest, obtaining the second best points score of 390 out of 420 and the overall seventh position in the combined leader board [10]. The teams that were on the top of the leader board, including our team, were invited to submit a very brief (one paragraph) summary of the algorithms used for inclusion in a journal paper prepared by the contest organizers [11].

WSIs contain voluminous amounts of data. One of the primary design goals has been to keep the computational complexity to the minimum possible level and to develop an efficient method that can process relevant tiles of an input WSI image quickly and classify the image into one of the four classes corresponding to the four HER2 scores. The second design goal was to have a feature set whose correlation to the percentage of membrane staining in the given sample could be easily visualized and interpreted by pathologists. The third design goal was to reduce the amount of information redundancy in the feature set by extracting a minimal set of characteristic features that would adequately represent the staining pattern and the percentage of staining. This paper presents two types of feature descriptors that have shown excellent intraclass correlation and interclass variance in our experimental analysis involving a large collection of WSI images. The first descriptor is called characteristic curves and they represent the variation of the percentage of staining in an image tile with saturation levels of the staining colour. The second descriptor is based on local binary patterns [12], and they encode information about the local texture variation in the image with saturation levels. The paper provides a detailed description of the WSI processing stages, development and selection of features, and the experimental analysis performed. We hope that the methods presented in this paper will contribute significantly to the development of faster and accurate automatic HER2 scoring techniques in the area of breast cancer histopathology.

The paper is organized as follows: The next section gives a description of the dataset used, an outline of HER2 assessment scheme and an overview of the stages of the processing pipeline. Section 3 provides an introduction to a novel set of features called characteristic curves and discusses their computational aspects and properties. Section 4 gives an overview of local binary patterns, their computation, and introduces another set of feature descriptors called LBP feature curves. Section 5 gives a brief description of a classification algorithm using the proposed feature descriptors for classifying histopathological images based on their HER2 scores. and Section 5 presents experimental results and comparative analysis. Section 6 presents experimental results and analysis. Section 7 concludes the paper with a summary of important aspects of the proposed features and outlines future research directions.

2. Materials and Methods

2.1 HER2 Assessment

The amplification of HER2 genes and correspondingly the over-expression of HER2 protein receptors play an important role in the development of breast cancer. The assessment of HER2 protein over-expression is done using the ImmunoHistoChemistry (IHC) test based on the percentage of membrane staining observed in tumor cells as well as the intensity of staining [2]. The mapping between the level of membrane staining and the reported HER2 score is shown in Table 1.
Table 1. Correlation between the intensity and percentage of membrane staining and the assigned HER2 scores [2].

<table>
<thead>
<tr>
<th>HER2 Score</th>
<th>Assessment</th>
<th>Staining Pattern</th>
</tr>
</thead>
<tbody>
<tr>
<td>0</td>
<td>Negative</td>
<td>No staining is observed, or membrane staining is observed in less than 10% of tumor cells</td>
</tr>
<tr>
<td>1+</td>
<td>Negative</td>
<td>A faint/barely perceptible membrane staining is detected in greater than 10% of tumor cells. The cells exhibit incomplete membrane staining.</td>
</tr>
<tr>
<td>2+</td>
<td>Weakly Positive</td>
<td>A weak to moderate membrane staining is observed in greater than 10% of tumor cells.</td>
</tr>
<tr>
<td>3+</td>
<td>Positive</td>
<td>A strong complete membrane staining is observed in greater than 10% of tumor cells.</td>
</tr>
</tbody>
</table>

A few sample tiles from WSI images of IHC stained slides are given in Figure 1 along with the HER2 scores to show the variations of the scores with the level of membrane staining seen in the images.

Figure 1. WSI tiles showing different levels of staining and corresponding HER2 scores.

2.2 Dataset

The dataset used in this research work was provided by the University of Warwick as part of the online HER2 scoring contest [9]. Permission was granted by the contest organizers to participating teams for the use of the dataset for research and academic purposes. The dataset consisted of a total of 172 whole slide images in Nano-zoomer Digital Pathology (NDPI) format. These WSIs were extracted from 86 cases of patients with invasive breast carcinomas [11]. For each case, WSIs of both Hematoxylin and Eosin (H&E) stained and IHC stained slides were provided. There were two HER2 scoring contests, and the number of WSIs provided for training and testing the classification algorithm is given in Table 2.

Table 2. Number of WSIs provided for training and testing the classification algorithm.

<table>
<thead>
<tr>
<th>Ground Truth HER2 Score</th>
<th>Number of WSIs</th>
<th>Contest-1 No. of WSIs</th>
<th>Contest-2 No. of WSIs</th>
</tr>
</thead>
<tbody>
<tr>
<td>0</td>
<td>13</td>
<td>28</td>
<td>6</td>
</tr>
<tr>
<td>1+</td>
<td>13</td>
<td></td>
<td></td>
</tr>
<tr>
<td>2+</td>
<td>13</td>
<td></td>
<td></td>
</tr>
<tr>
<td>3+</td>
<td>13</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Total:</td>
<td>52</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

2.3 Processing Stages

Various stages of the processing pipeline are shown in Figure 2. We used the OpenSlide API [13] to read WSIs of IHC stained slides, and a region of interest (ROI) containing a significant portion...
of the imaged tissue is extracted from the middle segment of the image. Rectangular tiles of size 1800 x 1200 pixels at 20x magnification that contain at most 20% background pixels are then created and used as inputs for the method that computes LBP features and characteristic curves. At least six tiles at randomly selected locations within the ROI are generated for each WSI. The remaining part of the pipeline thresholds the input tiles and computes the LBP features and also the percentage of staining in the tissue sample to obtain the characteristic curves. These steps are detailed in the following sections.

Figure 2. Processing stages in the extraction of characteristic curves and LBP features

3. Characteristic Curves

Curve based automated analysis of immunohistochemical images have been tried in the past with limited success [14]. In this section, we introduce a novel feature vector called a characteristic curve. An important parameter in HER2 assessment is the percentage of membrane staining perceived in an image segment. Assuming that we can compute the percentage of membranes stained in a particular colour range (this computation will be discussed in detail below), we can analyse the variations in this percentage value with respect to changes in the colour saturation threshold. Specifically, if \([h, s, v]\) represent the stain colour components in HSV space, and if \(p(\text{slow})\) denotes the percentage of staining with colour in the range given by the following inequalities:

\[
\begin{align*}
    h_1 & \leq h < h_2 \\
    s & > \text{slow} \\
    v_1 & \leq v < v_2,
\end{align*}
\]

then, the variation of \(p(\text{slow})\) plotted against \(\text{slow}\) gives the characteristic curve (or the percentage-saturation curve) of the image. In Equation 1, \([h_1, h_2]\) denote fixed hue thresholds specifying allowable variations in the hue value, and similarly \([v_1, v_2]\) denote value thresholds. Since we specify only the lower bound for saturation, progressively increasing \(\text{slow}\), typically from 0.1 to 0.5, produces a non-increasing characteristic curve (Figure 3).

The base components of the stain colour \([h, s, v]\) are computed using the training set where the given percentage of staining is above 80%. While computing the percentage of staining for the test (or cross-validation) sets, it is important to eliminate not only the background region but also other segments that are not part of the membrane region such as connective tissues, lobules and nuclei.
These regions can be segmented using colour (nuclei are stained in a distinctly different colour) or using a distance measure evaluated in colour space over a neighborhood mask around each pixel (for identifying regions of nearly constant colour value).

Figure 3. Intermediate stages in the generation of a characteristic curve.

Figure 3 shows thresholded images with stained regions in red colour as the value of \( s_{low} \) is increased from 0.1 to 0.4. The resulting characteristic curve is also shown. The characteristics curves have the property that they are always monotonically decreasing smooth curves. They allow accurate polynomial approximations using cubic curves. The shape of the curve can be directly matched with the staining patterns given in the HER2 assessment guidelines (Table 1) for a straightforward interpretation of the derived score (Figure 4). For example, the characteristic curve always lies below the 10% threshold when the score is 0, and only a small initial segment of the curve lies above the 10% mark when the score is 1. If the score is 3+, the curve lies completely above the 30% mark showing a strong and complete membrane staining. As seen in Figure 4, the curve passes through a much wider range of values of percentage staining when the score is 2+.

Figure 4. Variations in the shapes of the characteristic curves with different levels of staining.
The properties of the characteristic curve outlined above, particularly the fact that the curve is non-increasing, can be used for developing a naive rule-based classification algorithm as follows.

- If \( z_0 = p(0.1) < 10\% \), then the whole curve lies below 10\%, and the score is 0
- Else if \( z_{n-1} = p(0.5) > 30\% \), then the whole curve lies above 30\%, and the score is 3+
- Else if \( 10\% \leq z_0 = p(0.1) < 40\% \) and \( p(0.2) < 15\% \), the score is 1+
- Else if \( p(0.4) < 15\% \), then the score is 2+
- Else, the score is 3+

The rules were formed by analyzing the shapes of characteristic curves for several image tiles with ground truth values of HER2 scores assigned by pathologists. Note that for the above simple classification algorithm, we sample the curve at only four key points \( p(0.1), p(0.2), p(0.4), \) and \( p(0.5) \). We outlined the rule based algorithm here primarily to show the feature representation capability of the characteristic curves.

4. Local Binary Patterns

4.1 LBP Computation

Local binary patterns (LBP) are powerful feature descriptors used for texture analysis and classification [12]. The binary pattern is derived by comparing the intensity at each pixel with its eight neighbors and encoding the information in an 8-bit integer value. This encoding can be viewed as a transformation of the input image into an LBP image as shown in Figure 9. The histogram of the LBP image is generally used for texture classification. In the area of medical image analysis, LBP methods have been successfully used in characterizing disease patterns [15-17] and automated diagnosis [18]. Local binary patterns have also been used for analyzing histopathological images and detecting mitotic cells [19,20]. Several variants of LBP features such as hierarchical LPB have also been proposed for specific applications like retinal vein occlusion recognition [21].

Figure 5. The intermediate steps in the computation of the LBP histogram of an image.

As an example, an input image and its LBP image are shown in Figure 6.

Figure 6. (a) A sample input image for LBP computation; (b) The corresponding LBP image.
As discussed in Section 3, we first obtain a thresholded image using a hue range \([h_1, h_2]\) and saturation values with \(s > s_{\text{low}}\). The pixels passing the threshold test are converted to gray level by mapping \(h_1\) to 0 and \(h_2\) to 255. This gray level image is used as the input for LBP computation. The LBP histogram of such images contain predominant features that represent the texture characteristics of the staining patterns.

### 4.2 LBP Feature Curves

An LBP histogram is shown in Figure 7. The important LBP features are highlighted based on their magnitudes. The LBP histogram contains 256 values \(L_i, \ i = 0...255\). We propose the following set of five LBP features for classification:

\[
V = \{L_2, L_{32}, L_{128}, L_{223}, L_{253}\} \quad (2)
\]

Each feature in the above set can generate a feature curve as detailed below. Consider one of the LBP feature components, say \(L_{32}\). When the input image's saturation threshold \(s_{\text{low}}\) is varied from 0.1 to 0.4 as discussed in Section 3, we get the corresponding variation in \(L_{32}\). This variation in the values of \(L_{32}\) shows a non-increasing trend very similar to that of the characteristic curve.

The values of the LBP feature curves are normalized to remove any variations due to changes in image size as follows:

\[
L_i' = L_i \cdot 100/(w \cdot h)
\]
where, \( w, h \) denote the width and the height of the input image. The variations of three LBP feature components \( L'_32, L'_128, \) and \( L'_253 \) with the saturation threshold for images with HER2 scores 0, 1+, 2+ and 3+ are shown in Figure 8.

The LBP feature curves bear similarity with characteristic curves in that they do not contain high frequency variations and are non-increasing. Further, as can be seen in Figure 8, LBP feature curves also show excellent discriminating power between the four HER2 classes, making them suitable for use as feature vectors in classification algorithms.

5. HER2 Classification and Scoring

In this section, we outline a ‘one-vs-all’ multi-class classification algorithm using logistic regression. Logistic regression was chosen to minimize the computational complexity. Higher order methods such as neural networks could also be designed with the use of the feature vectors proposed in this paper. For a given training example with index \( j \), the points sampled along its characteristic curve or LBP feature curve \( x^{(i)} = p(s_i), i = 1..n, \ j = 1..m \) are used as features. The class labels are denoted by \( y_j \in [0, 3], \ j = 1..m \). We denote the feature matrix by \( X \in \mathbb{R}^{m \times (n+1)} \), the output vector of labels by \( Y \in \mathbb{R}^{m \times 1} \), and the classifier parameter vector for each class by \( \theta_k \in \mathbb{R}^{(n+1) \times 1}, \ k = 1..4 \). Here, class-1 corresponds to the set of training examples with HER2 score 1+, class-2 with HER2 score 2+, class-3 with HER2 score 3+ and class-4 with HER2 score 0. We then have the following equations for the hypothesis functions \( H \), the cost function and the gradient functions:

\[
H = g(X\theta_k)
\]

where, \( H \in \mathbb{R}^{m \times 1} \), and \( g() \) denotes the sigmoid function. The cost function \( J(\theta) \) is then given by

\[
J(\theta_k) = -\frac{1}{m} \left( Y^T \log(H) + (1-Y)^T \log(1-H) \right)
\]

and the gradient function vector \( J'(\theta_k) \) is defined as

\[
J'_k(\theta) = \frac{1}{m} (X^T (H - Y)), \ k = 1..4.
\]

For prediction, the points \( x_i \) on the characteristic curve or the LBP feature curve of a given sample are combined with the trained values of class parameters \( \theta_k \) for each class \( k = 1..4 \), and the class that gives the maximum value for \( g(x/ \theta_k) \) is chosen. In the next section, we provide the result of classification experiments using the above methods.

6. Experimental Results and Analysis

We used features computed from 52 WSIs with 3 tiles at 20x from each image (comprising of 156 images) and their ground truth values as the training data. Another set of 3 tiles from each of the 52 cases formed the cross-validation set. Out of the total of 156 image tiles in the cross-validation set, 39 belonged to each of the four classes corresponding to four HER2 scores. For generating feature vectors for classification using logistic regression, it was found that a step size of 0.02 for the saturation threshold would provide an adequate number of 20 points (features) within the saturation range \( slow \in [0.1, 0.5] \). The feature matrix \( X \) in Equation 3 therefore had the dimension 156x20.

The gradient descent algorithm used 100 iterations to converge to the solution with a learning rate of 0.001 (Figure 9).
Figure 9. Convergence of the cost functions of the four-class logistic regression algorithm.

Figure 9 gives a sample plot of cost function values for each of the four classes, where the characteristic curves consisting of 20 features for each sample were used as feature vectors. The confusion matrix in Table 3 summarizes the results for each class and gives the overall accuracy achieved.

<table>
<thead>
<tr>
<th>Predicted</th>
<th>0</th>
<th>1+</th>
<th>2+</th>
<th>3+</th>
<th>Precision</th>
<th>Recall</th>
</tr>
</thead>
<tbody>
<tr>
<td>Actual</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>0</td>
<td>37</td>
<td>2</td>
<td>0</td>
<td>0.86</td>
<td>0.95</td>
</tr>
<tr>
<td></td>
<td>1+</td>
<td>6</td>
<td>29</td>
<td>4</td>
<td>0.83</td>
<td>0.74</td>
</tr>
<tr>
<td></td>
<td>2+</td>
<td>0</td>
<td>4</td>
<td>34</td>
<td>0.87</td>
<td>0.87</td>
</tr>
<tr>
<td></td>
<td>3+</td>
<td>0</td>
<td>0</td>
<td>1</td>
<td>38</td>
<td>0.97</td>
</tr>
</tbody>
</table>

Table 3. Confusion matrix for the multi-class logistic regression algorithm

The smoothness and monotonically decreasing properties of the characteristic curve can be effectively made use of in reducing the dimensionality of the features in the logistic regression algorithm. As in the case of the rule based classification method, we can sample the curve at only four key points, p(0.1), p(0.2), p(0.4), and p(0.5), and also use the slope information at those points, p'(0.1), p'(0.2), p'(0.4), and p'(0.5) to get a feature vector of size 8 instead of 20. The cost functions converge to almost similar values with only a slight increase in the magnitudes. The confusion matrix obtained by running the algorithm with the reduced set of features of the characteristic curve is shown in Table 4.

<table>
<thead>
<tr>
<th>Predicted</th>
<th>0</th>
<th>1+</th>
<th>2+</th>
<th>3+</th>
<th>Precision</th>
<th>Recall</th>
</tr>
</thead>
<tbody>
<tr>
<td>Actual</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>0</td>
<td>37</td>
<td>2</td>
<td>0</td>
<td>0.80</td>
<td>0.95</td>
</tr>
<tr>
<td></td>
<td>1+</td>
<td>8</td>
<td>24</td>
<td>7</td>
<td>0.75</td>
<td>0.61</td>
</tr>
<tr>
<td></td>
<td>2+</td>
<td>1</td>
<td>6</td>
<td>31</td>
<td>0.79</td>
<td>0.79</td>
</tr>
<tr>
<td></td>
<td>3+</td>
<td>0</td>
<td>0</td>
<td>1</td>
<td>38</td>
<td>0.97</td>
</tr>
</tbody>
</table>

Table 4. Confusion matrix for the multi-class logistic regression algorithm with the reduced feature set.

As seen in Table 4, reducing the dimensionality of the feature set from 20 to 8 only affected the recall rates of classes 1 and 2.

Experimental analysis using LBP feature curves also gave reasonably good levels of accuracy. Only one LBP feature curve selected from the set in Equation 2 was used in our analysis, and the feature vector consisted of 20 sample points along the curve. We give below the results for the feature curve $L_{32}$ (Table 5).
Table 5. Confusion matrix for the multi-class logistic regression algorithm with the LBP feature vector $L_3$.

<table>
<thead>
<tr>
<th>Actual</th>
<th>0</th>
<th>1+</th>
<th>2+</th>
<th>3+</th>
<th>Precision</th>
<th>Recall</th>
</tr>
</thead>
<tbody>
<tr>
<td>0</td>
<td>38</td>
<td>1</td>
<td>0</td>
<td>0</td>
<td>0.84</td>
<td>0.97</td>
</tr>
<tr>
<td>1+</td>
<td>6</td>
<td>27</td>
<td>6</td>
<td>0</td>
<td>0.82</td>
<td>0.69</td>
</tr>
<tr>
<td>2+</td>
<td>1</td>
<td>5</td>
<td>32</td>
<td>1</td>
<td>0.84</td>
<td>0.82</td>
</tr>
<tr>
<td>3+</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>39</td>
<td>0.98</td>
<td>1.00</td>
</tr>
</tbody>
</table>

The texture characteristics represented by LBP features were useful in resolving some of the ambiguous cases for scores 0 and 3+ where the texture features are highly distinguishable, providing higher recall rates for those two scores. The LBP features also gave higher false positives for score 2+. Overall, logistic regression with 20 feature points computed from the characteristic curves gave the highest accuracy of 88.5%.

Analyzing the staining patterns in tiles that were wrongly classified revealed a common problem in the automatic extraction of tiles from WSIs. Some of the samples with scores 1+ and 2+ had large tissue regions without any staining. The example shown in Figure 10 contains a tissue sample at 10x magnification with an assigned score of 2+.

Figure 10. An example showing two tile positions with varying image characteristics within the same WSI.

In Figure 10, the tile on the top did not contain any stained membrane regions and was assigned a ground truth value of 2+ at the training stage, and a predicted value of 0 at the cross-validation stage. This tile could have been a valid part of any WSI with a score 0, and therefore there is no way by which such tiles can be identified and discarded by the automatic tile extraction method. Manually identifying such tiles from the training and cross-validation sets significantly improved the scores of the classification algorithms. The tile on the bottom half of Figure 10 was assigned the correct score of 2+.

7. Conclusions and future work

This paper has introduced two novel feature descriptors viz., characteristic curves and LBP feature curves that could be effectively used in classification algorithms for automated scoring of HER2 in breast cancer histology slides. The computational aspects of both types of descriptors and their shape feature representation capabilities in embedding information about the staining patterns and the percentage of staining present in images with different HER2 scores have been discussed in detail. Both descriptors have similar geometrical attributes in that they are both smooth non-increasing curves. Experimental analysis have shown that both descriptors have excellent in-
terclass variance and intraclass correlation properties that make them useful for applications in
classification algorithms. Results of experimental analysis done using a comprehensive WSI dataset
provided by the University of Warwick[9] has also been presented. The results show that the fea-
tures used with a multi-class classification algorithm such as logistic regression can provide very
good levels of accuracy. The paper also outlined computational stages in the overall processing
pipeline for automatic HER2 scoring using WSI files as inputs.

Experimental results given in the paper also show the need for further improving the discrim-
inating power of the features. Future work is directed towards performing detailed feature analysis
to select a combination of features from characteristic curves and LBP feature curves. Further
analysis is required for accurate identification of membrane morphology and region segmentation,
particularly for samples with an assigned HER2 score 1+. It is also necessary to assess the repro-
ducibility of results, specifically inter-scanner variability [23] of the rule-based classification algo-

rithm as the rules were formed using data produced by a single scanner. Future work is also directed


References

1. Hicks, D.G.; Schiffhauer, L. Standardized assessment of the HER2 status in breast cancer by

immunohistochemistry.  Lab. Med. 2015, 42(8), 459-467,  DOI: 10.1309/LMGZZS8CT50DBGTW


2015, 68, 93-99, DOI: 10.1136/jclinpath-2014-202571


interpretation of HER2 immunohistochemical expression with unaided and computer aided digital


and pathologist-generated tumour segmentations for immunohistochemical scoring of breast tissue

microarrays.  Br J Cancer 2015, 113(7), 1075-1080, DOI: 10.1038/bjc.2015.309


70(1), 59-73, DOI: 10.1016/j.ymeth.2014.06.015

6. Farahani, N.; Parwani, A.V.; Pantanowitz, L. Whole slide imaging in pathology: advantages, limitations


patients using large FISH images. Proceedings of 25th Signal Processing and Communications

Applications Conference, Antalya, Turkey, 15-18 May 2017, 1-4, DOI: 10.1109/SIU.2017.7960428

9. Department of Computer Science, University of Warwick: Her2 Scoring Contest. Available online:

http://www2.warwick.ac.uk/fac/sci/dcs/research/combi/research/bic/her2contest/ (accessed on 15 Nov

2016).

10. Department of Computer Science, University of Warwick: Her2 Contest Results. Available online:

http://www2.warwick.ac.uk/fac/sci/dcs/research/combi/research/bic/her2contest/outcome (accessed on 15

Nov 2016).

11. Quaiser, T., et.al. HER2 Challenge Contest: A detailed assessment of HER2 scoring algorithms and man

vs machine in whole slide images of breast cancer tissues. Histopathology 2017. DOI: 10.1111/his.13333

12. Pietkainen, M.; Zhao, G.; Hadid, A.; Ahonen, T. Computer Vision Using Local Binary Patterns,


14. Livanos, G.; Zervakis, M.; Giakos, G.C. Automated analysis of immunohistochemical images based on

curve evolution approaches. Proceedings of IEEE conference of Imaging Systems and Techniques, Beijing,

China, 22-23 Oct 2013, 112-115, DOI: 10.1109/IST.2013.6729673

15. Sørensen, L.; Shaker, S.B.; Bruijne, M. de. Quantitative analysis of pulmonary emphysema using local

binary patterns, IEEE Transactions on Medical Imaging 2010, 29(2), 559-569, DOI: 10.1109/TMI.2009.2038575.


