

1 **The Genetic Structure of *Turnip mosaic virus* Population**
2 **Reveals the Rapid Expansion of a New Emergent Lineage**
3 **in China**

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16 **Abstract:** *Turnip mosaic virus* (TuMV) is one of the most widespread and economically
17 important virus infecting both crop and ornamental species of the family *Brassicaceae*.
18 TuMV isolates can be classified to five phylogenetic lineages, basal-B, basal-BR, Asian-BR,
19 world-B and Orchis. To understand the genetic structure of TuMV from radish in China, the
20 3'-terminal genome of 90 TuMV isolates were determined and analyzed with other Chinese
21 isolates available. The results showed that the Chinese TuMV isolates from radish formed
22 three groups: Asian-BR, basal-BR and world-B. More than half of these isolates (52.54%)
23 were clustered to basal-BR group, and could be further divided into three sub-groups. The
24 TuMV basal-BR isolates in the sub-groups I and II were genetically homologous with
25 Japanese ones, while those in sub-group III formed a distinct lineage. Sub-populations of
26 TuMV basal-BR II and III were new emergent and in a state of expansion. The Chinese
27 TuMV radish populations were under negative selection. Gene flow between TuMV
28 populations from Tai'an, Weifang and Changchun was frequent.

29

30 **Keywords:** *Turnip mosaic virus*; *Potyvirus*; Genetic structure; Population; China

31

32 **1. Introduction**

33 Due to the error-prone nature of their RNA-dependent RNA polymerases, populations
34 of plant RNA viruses are genetically heterogeneous and the genetic structure may change
35 with time and environment[1, 2]. Studies of the genetic structure of viruses will provide
36 information about the mechanisms and factors driving their evolution and help us to
37 understand the molecular evolutionary history of viruses in relation to their dispersion and
38 emergence of new epidemics[3].

39 *Turnip mosaic virus* (TuMV) is a species of the largest plant virus genus *Potyvirus*
40 (family *Potyviridae*). TuMV has flexuous filamental particles of 700-750 nm long and can be
41 transmitted by 40-50 species of aphids in a non-persistent manner [4, 5]. The TuMV genome
42 consists of one single-stranded positive sense RNA molecule of approximately 9830
43 nucleotides (nt) and contains a large open reading frame (ORF)[6]. The ORF encodes a
44 polyprotein that will be hydrolyzed to produce ten mature products[7, 8]. A frame-shift
45 protein, P3N-PIPO, was reported to be involved in the pathogenesis and movement of
46 TuMV[9, 10].

47 TuMV can infect plants of 300 species in 43 families, and is probably the most
48 widespread and economically important virus infecting both crop and ornamental species of
49 family *Brassicaceae*[11, 12]. In an extensive survey conducted in 28 countries, TuMV
50 ranked second for crop yield losses[4]. TuMV is a highly variable and has many biological
51 and serological strains[13-16]. According to its host range, TuMV isolates can be classified
52 to two pathotypes, B (mainly infects plants of the genus *Brassica*) and BR (infects plants of
53 both *Brassica* and *Raphanus*). The brassica-infecting TuMV isolates were categorized into
54 four phylogenetic lineages, basal-B, basal-BR, Asian-BR and world-B, which correlated well
55 with their differences in pathogenicity and geographical origin[17]. Most recently, a
56 monophyletic sister lineage called ‘Orchis group’ was detected from wild orchids-infecting
57 TuMV isolates, which are more likely the ancestor of TuMV[18]. As in other potyviruses[19],
58 recombination is a frequent event in the evolution of TuMV. Intra- and inter-lineage
59 recombinants are common in natural populations of TuMV and can be detected throughout
60 the genome[6, 20-22]. The Chinese and Japanese TuMV isolates are part of the same
61 population but are a discrete lineage[22, 23]. The gene flow between sub-populations of
62 TuMV from Vietnam, Japan and China are frequent[20]. The basal-BR isolates have
63 occurred over the whole Japanese islands and have evolved into four sub-lineages[23-25].

64 Previous studies showed that the TuMV isolates of China can be clustered to world-B
65 and Asian-BR groups[10, 17, 24, 26, 27]. However, we have detected the existence of
66 basal-BR isolates in China and reported the complete genomic sequences of two basal-BR
67 isolates that represented two novel recombination patterns[6, 28]. Here, we studied the
68 genetic structure of TuMV population in China and found that the basal-BR group of TuMV
69 was expanding in China.

70 **2. Materials and Methods**

71 *2.1. Virus samples, RNA extraction and sequencing*

72 Samples were collected from radish from Heilongjiang, Jilin and Shandong provinces
73 from 2005 to 2010. All the samples were biologically cloned by three cycles of single lesion
74 isolation in *Chenopodium amaranticolor* and propagated in *B. rapa*. Inoculated plants were
75 maintained in a glasshouse at 25 °C.

76 Total RNAs were extracted from 100 mg TuMV-infected *B. rapa* leaves with the
77 Invitrogen Trizol Kit following instructions of the manufacturer. The 3'-terminal 1.1 kb of
78 TuMV genome were amplified with RT-PCR using primers CP-F (5'-ATC TTC GAA GAT
79 TAC GAA GA-3') and CP-R (5'-CCT TGC TTC CTA TCA AAT G-3') [29]. The fragments
80 were cloned into pMD18-T vector (TaKaRa Biotechnology Dalian Co, Ltd) and sequenced
81 by a ABI PRISM™ 377 DNA Sequencer. For each isolate, at least four clones from two
82 separate PCR were sequenced. In case of any inconsistency, at least two more clones will be
83 sequenced to obtain the consensus sequence.

84 2.2. Recombination analysis

85 The sequences of 90 TuMV isolates and other 28 obtained from the databank were
86 subjected to recombination analyses using the software package RDP3, which assembled
87 programs RDP[30], GENECONV[31], BOOTSCAN[32], MAXCHI[33], CHIMEARA[34]
88 and SISCAN. The sequences were analyzed using the default settings for different detection
89 programs and a Bonferroni-corrected *P*-value cut off of 0.05. The potential recombinants
90 identified by the programs in RDP3 were re-checked using PHYLPRO[35]. The RDP,
91 BOOTSCAN and SISCAN programs were based on phylogenetic methods, whereas
92 GENECONV, MAXCHI and CHIMAERA programs were substitution methods, and the
93 PHYLPRO program was a distance comparison method. Only those sequences with
94 recombination supported by at least three programs or two kinds of methods and with
95 *P*-value $<1.0 \times 10^{-6}$ were regarded as 'clear' recombinants; otherwise, they were called as
96 'tentative' recombinants[23, 25].

97 2.3. Phylogenetic analysis of the TuMV population

98 Sequence alignments were performed using the CLUSTAL W program (Thompson *et*
99 *al.*, 1994). Phylogenetic tree of TuMV isolates excluding the recombinant ones was
100 constructed using methods including Maximum Likelihood (ML) method that are packaged
101 in the MEGA6.0[36]. The CP gene of one *Narcissus yellow stripe virus* (NYSV) isolate was
102 used as outgroup[37]. Bootstrap analysis was repeated 1000 times to evaluate the
103 significance of the internal branches.

104 2.4. Sequences diversity and population demography analysis

105 DnaSP version 5.10 was used to calculate the values of nucleotide diversity, Tajima's D,
106 Fu and Li's D and F tests, haplotype diversity and nucleotide diversity[38-40]. Tajima's D,
107 Fu and Li's D and F tests hypothesize that all mutations are selectively neutral. Tajima's D
108 test depends on the differences between the numbers of segregating sites and the average
109 number of nucleotide differences. Fu and Li's D test is related the differences between the
110 number of singletons (mutations appearing only once among the sequences) and the total
111 numbers of mutations. Fu and Li's F test is based on the differences between the numbers of
112 singletons and the average number of nucleotide differences among all pairs of sequences.
113 Haplotype diversity refers to the frequency and number of haplotypes in the population.
114 Nucleotide diversity estimates the average pairwise differences among sequences. The
115 nucleotide diversities were calculated within and between groups. DnaSP version 5.10[40]
116 was also used to estimate the frequency distribution of the number of pairwise differences
117 among all sequences. Mismatch distribution of all populations were estimated on all pairs of
118 haplotypes present in a population[40]. Mismatch distribution analysis was based on 1 000
119 simulated samples and used to evaluate whether a population had undergone sudden
120 expansion or maintained constant size. In a recently expanded and still intact population, the
121 majority of lineage coalescence events were expected to produce a smooth unimodal Poisson
122 distribution around the time of expansion; otherwise, multimodal and ragged distribution
123 was expected.

124 2.5. Selection pressure, genetic differentiation and gene flow

125 The selection pressure was estimated by d_N/d_S ratio, where d_N represented the average
126 number of non-synonymous substitutions per non-synonymous site and d_S represented the
127 average number of synonymous substitutions per synonymous site. The values of d_N and d_S
128 were estimated separately by using the PBL method[41, 42] implemented in MEGA 6.0.
129 When d_N/d_S ratio =1, it means that neutral selection had occurred; when $d_N/d_S < 1$ or > 1 , it
130 means that negative (purifying) or positive (diversifying) selection, respectively, had
131 occurred. Genetic distances were calculated by Pamilo-Bianchi-Li (PBL) methods[41, 42].

132 Genetic differentiation between populations was examined by three permutation-based
 133 statistical tests, K_s^* , Z and S_{nn} [43, 44]. $P < 0.05$ was considered as the criterion for rejecting
 134 the null hypothesis that there is no genetic differentiation between two subpopulations. The
 135 level of gene flow between populations was measured by estimating F_{st} (the
 136 inter-population component of genetic variation or the standardized variance in allele
 137 frequencies across populations) and Nm using DnaSP 5.10[40]. F_{st} ranges from 0 to 1 for
 138 undifferentiated to fully differentiated populations, respectively. Normally, an absolute value
 139 of $F_{st} > 0.33$ or $Nm < 1$ suggests infrequent gene flow, while absolute value of $F_{st} < 0.33$ or
 140 $Nm > 1$ suggests frequent gene flow.

141 3. Results

142 3.1. Identities between TuMV isolates from radish in China

143 We collected and biologically cloned 101 TuMV isolates from radish from 2004 to 2010
 144 from Beijing, Hebei, Heilongjiang, Henan, Jilin and Shandong provinces, 94 of which are
 145 first reported here. A fragment of 1082 bp covering partial NIB gene (28 bp), complete CP
 146 gene (867 bp) and 3'-UTR (187 bp) was amplified from these isolates. The geographical
 147 origin of each isolate is listed in Table 1.

148 **Table 1.** Recombination sites and possible parent-like isolates

Isolate	Major parent	Minor parent	B-E	Software*	P-value	Z-value [#]
CHBJ1	WF1-04	WF7-06	15-1051	GBS3	2.07×10^{-18}	9.7
CHBJ2	WF1-04	WF7-06	15-1051	GBS3	2.07×10^{-18}	9.65
CHK16	WF-05	R4	17-387	BMCS3	1.028×10^{-14}	9.43
CHK51	WF-05	R4	17-387	BMCS3	1.028×10^{-14}	9.38
R	WF-05	R4	17-697	MCS3	1.028×10^{-14}	8.92
R5	WF-05	R4	17-451	MCS3	1.028×10^{-14}	10.6
WF2-06	WF1-04	WF7-06	15-1051	GBS3	2.07×10^{-18}	10.2
WF3-06	WF1-04	WF7-06	10-1051	GBS3	2.07×10^{-18}	10.2
WF3-07	WF1-04	WF7-06	10-1047	GBS3	2.07×10^{-18}	10.2
WF8-08	WF1-04	WF7-06	10-1051	GBS3	2.07×10^{-18}	10.1
WF10-07	TA15-08	WF3-08	91-786	MCS3	1.18×10^{-8}	7.1
WFLB3	WF7-06	WF1-04	377-657	GBS3	2.07×10^{-18}	10.6

149 The recombination crossover sites within CP-UTR of turnip mosaic virus were detected by the
150 recombination detecting programs.

151 *The programs supporting recombination event. R(RDP), G (Geneconv), B (Bootscan), M (Maxchi),
152 C (Chimaera), S (Siscan) and 3 (3seq). The analysis was carried out with default settings for the different
153 detection methods and a Bonferroni-corrected cutoff of 0.05. The program that has the greatest *P*-value
154 was marked in bold. B-E represents the beginning and ending point of recombination.

155 The cloned sequences excluding primers shared identities of 89.6% - 100% at nt level
156 with other 28 Chinese TuMV radish sequences available in Genbank. These 129 CP gene
157 sequences showed identities of 88.2% - 100% at nt level and 91.3% - 100% at aa level. The
158 identities of 44 TuMV isolates from Weifang were 89.6% -100% at nt level and 95.1% -
159 100% at aa level. Those of 37 isolates from Tai'an were 88.9% - 100% at nt level and 94.1 -
160 100% at aa level. The 12 Changchun isolates shared identities of 90.3% - 100% at nt level
161 and 95.8% - 100% at aa level.

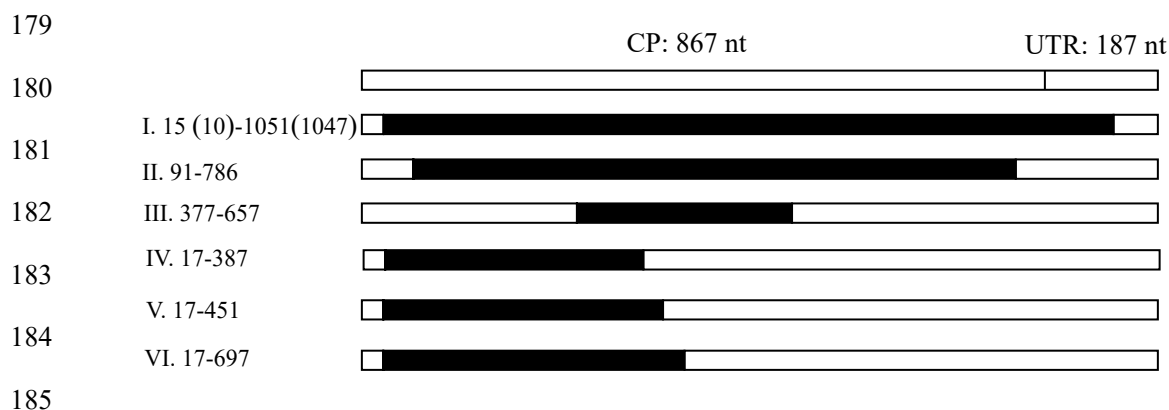
162 3.2. *Recombination analyses*

163 Possible recombination events in the CP-UTR region of 129 radish isolates from China
164 were detected with the program package RDP. Twelve of the sequences (11.4%) analyzed
165 had 'clear' recombination. Among the recombinant isolates, WF0710 was the within-group
166 recombinant of basal-BR isolates , with TA0815 as its major parent and WF0803 as the
167 minor parent; others were between-group recombinants of Asian-BR and world-B isolates,
168 most with WF-05 or WF1-04 of world-B as the major parent and WF7-06 or R4 of Asian-BR
169 as minor parent; WFLB had WF7-06 as its major parent and WF1-04 as minor parent (Table
170 1).

171 The recombination pattern can be classified into six types (Fig. 1). More than 50%
172 recombinants belong to recombination pattern 1, with the recombination site located within
173 UTR. WF0710 belonged to pattern II, WFLB3 to pattern III, CHK16 and CHK51 to pattern
174 IV, R5 to pattern V and R to pattern VI (Fig. 1).

175 3.3. *Phylogenetic analyses*

176 Using ML method, a phylogenetic tree was constructed with the 118 CP-UTR
177 sequences of TuMV (excluding the 11 between-group recombinants) from radish in China.
178 These TuMV isolates were clustered to three lineages corresponding to world-B, Asian-BR

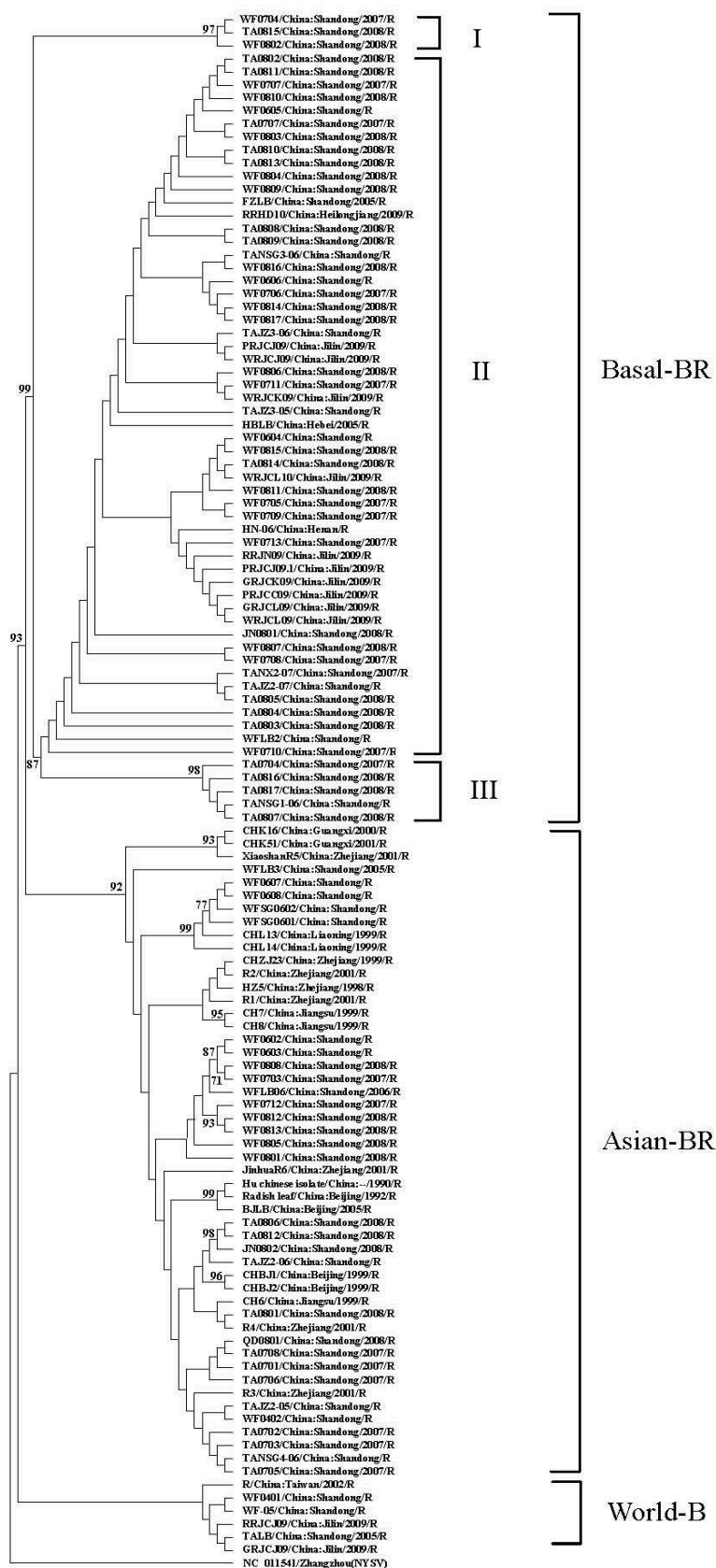


186 **Figure 1.** Recombination patterns in the CP-UTR region of TuMV from radish in China. Twelve
 187 recombinants were divided into 6 recombination patterns. I: CHBJ1, CHBJ2, WF2-06, WF3-06, WF8-08,
 188 WF3-07; II: WF10-07; III: WFLB3; IV: CHK16, CHK51; V: R5; VI: R.

189
 190 and basal-BR (Fig. 2). The world-B lineage contained only six isolates (R, WF0401, WF-05,
 191 TALB, GRJCJ09 and RRJCJ09). The Asian-BR lineage consisted of 50 isolates. The
 192 Basal-BR lineage included 62 isolates which can be further divided into 3 sub-lineages.
 193 Sub-lineage Basal-BR I had three isolates (WF0704, WF0802 and TA0815), all of which
 194 were from Shandong province. Basal-BR II consisted of 54 isolates. Among which 41 were
 195 from Shandong, ten from Jilin, each from Henan, Heilongjiang and Hebei. Basal-BR III
 196 contained five isolates, all of which were found in Tai'an, Shandong province. The genetic
 197 distance values within groups ranged from 0.014 to 0.026, which were 4 to 5 times lower
 198 than those between groups (0.067 to 0.094) (Table 2). The genetic distance values between
 199 sub-groups of basal-BR were 0.033 to 0.041, which were higher than those within
 200 sub-groups but lower than those between groups.

201 The phylogenetic tree constructed with the CP gene could also be divided into three
 202 groups corresponding to world-B, Asian-BR and basal-BR. The genetic distance values
 203 between groups ranged from 0.076 to 0.091, which were higher than those within groups
 204 (0.015 to 0.049). The genetic distance values between sub-groups of basal-BR were 0.032 to
 205 0.049, which were remarkably higher than those within sub-groups(0.004 to 0.015) but
 206 lower than those between groups. Therefore, the classification of these TuMV isolates into
 207 three groups and basal-BR into three sub-groups was reasonable.

208 To further study the genetic structure of TuMV basal-BR sub-populations from China
 209 and Japan, we constructed phylogenetic trees with basal-BR isolates available from both
 210 countries using ML method (Fig. 3). These TuMV isolates were clustered into four lineages,



211

212 **Figure 2.** Maximum Likelihood tree of TuMV isolates from radish in China calculated from the CP

213 -UTR sequencess.

Table 2. Estimates of genetic differentiation among sites (F_{ST}) within each region

Gene	Parameter	WF between WF	TA between TA	WF between TA	WF between CC	TA between CC	CC between CC
CP	Ks(<i>P</i> -value)	3.16314 (1.0000)	3.12325 (1.0000)	3.14501 (0.1200)	3.01985 (0.0080**)	2.90505 (0.0120*)	2.41801 (1.0000)
	Z(<i>P</i> -value)	1935.22093 (1.0000)	1367.97222 (1.0000)	1618.33343 (0.2990)	740.62579 (0.0080**)	534.81963 (0.0060**)	143.22727 (1.0000)
	Snn(<i>P</i> -value)	0.08081 (1.0000)	0.12416 (1.0000)	0.66143 ** (0.0030)	0.88492 (0.0000***)	0.85305 (0.0000***)	0.10000 (1.0000)
	F_{ST}	-0.02326	-0.02778	-0.00436	0.15467	0.15485	-0.09091
	Nm	-11.00	-9.25	-57.56	1.37	1.36	-3.00
	Ks*(<i>P</i> -value)	1.19299 (1.0000)	0.96193 (1.0000)	1.08658 (0.5460)	0.94143 (0.0570) ns	1.13222 (0.0420*)	0.88305 (0.9280) ns
UTR	Z(<i>P</i> -value)	1844.07143 (1.0000)	1362.22222 (1.0000)	1581.52519 (0.4120)	552.71858 (0.0680) ns	723.74704 (0.0470*)	140.95455 (1.0000) ns
	Snn(<i>P</i> -value)	0.34906 (1.0000)	0.39139 (1.0000)	0.52678 (0.1430)	0.68796 (0.0230*)	0.69754 (0.0780) ns	0.31111 (1.0000) ns
	F_{ST}	-0.02381	-0.02778	-0.01268	0.08893	0.11459	-0.09091
	Nm	-10.75	-9.25	-19.96	2.56	1.83	-3.00

215 ns, not significant; *, 0.01<P<0.05; **, 0.001<P<0.01; ***, P<0.001. Determined using 1000 permutations.

216 WF, isolates from Weifang of Shandong province; TA, isolates from Tai'an of Shandong province; CC, isolates from Changchun of Jilin
 217 province.

218

219

224 figure were listed by isolate name/ location of origin/ year of collection /original host (R is for *Raphanus*).
225
226 corresponding to the ones reported by Tomitaka *et al.*[25]. Interestingly, the TuMV basal-BR
227 isolates of sub-groups I and II from both China and Japan formed common clusters, which
228 indicated sub-populations of basal-BR I and II from these two countries were genetically
229 identical. However, those of sub-group III from China and Japan formed separate clusters,
230 indicating that China and Japan had different sub-populations of basal-BR III. Sub-group IV
231 consisted of isolates from Japan only. No Chinese TuMV isolate fell into this sub-group.

232 3.4. Selective pressures acting on TuMV CP genes

233 To estimate the selection pressure acting on TuMV CP genes, we calculated the d_N/d_S
234 ratios for TuMV sub-populations of different collection regions using Pamilo-Bianchi-Li
235 (PBL) method assembled in MEGA version 6.0[36]. The d_N values for TuMV isolates from
236 Weifang, Tai'an and Changchun were 0.005 ± 0.001 , 0.006 ± 0.001 and 0.002 ± 0.001 ,
237 respectively, which were less than the d_S values (0.070 ± 0.007 , 0.098 ± 0.012 and
238 0.014 ± 0.006). Therefore, the values of the d_N / d_S ratio for TuMV *cp* genes were <1 ,
239 indicating that purifying (negative) selection was acting on TuMV *cp* genes. The nucleotide
240 distances were 0.023 ± 0.002 , 0.029 ± 0.003 and 0.005 ± 0.001 , respectively, and showed no
241 significant difference.

242 3.5. Genetic differentiation and gene flow

243 Genetic differentiation and gene flow between and within populations was examined by
244 five permutation-based statistical tests, K_s^* , Z and S_{nn} or F_{st} and N_m . The results showed
245 no genetic differentiation between or within TuMV sub-populations from Tai'an and
246 Weifang in the CP genes or UTR (Table 2).

247 The absolute values of F_{ST} between or within TuMV populations of Tai'an, Weifang
248 and Changchun were all below 0.33, indicating that the gene flow between or within TuMV
249 populations of Tai'an and Weifang, and that with TuMV population of Changchun is most
250 frequent; however, the gene flow between Changchun and Tai'an, and Changchun and
251 Weifang is less frequent. The absolute values of $N_m > 1$ also support the conclusion on gene
252 flow.

253 3.6. Population dynamics

254 The Tajima's D , Fu & Li's D^* , Fu & Li's F^* values for TuMV basal-BR II
255 sub-population from Weifang and Tai'an of Shandong province were negative and the data is
256 significant, which indicated that these sub-populations were in state of increasing (Table 3).

257 Sub-populations of Asian-BR and basal-BR III from Tai'an, Asian-BR from Weifang, and
 258 basal-BR from Changchun were also in a state of increasing, but the data was not significant
 259 (Table 3). Haplotype diversity, ranging from 0.890 to 1.000, had little difference between
 260 groups or sub-groups. The basal-BR II isolates from Tai'an had the lowest nucleotide
 261 diversity of 0.00322, while the Asian-BR isolates from Weifang had the highest one of
 262 0.01159.

263 **Table 3.** Neutrality tests, haplotype and nucleotide diversity of *Turnip mosaic virus*
 264 sub-populations

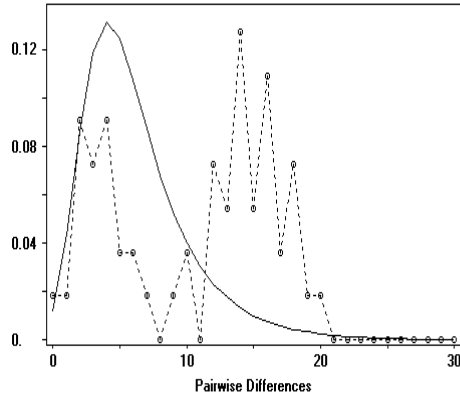
Group		Haplotype diversity	Nucleotide diversity	Tajima's D	Fu and Li's D	Fu and Li's F
Tai'an	basal-BR II	0.890 (0.073)	0.00322 (0.00062)	-2.19554**	-2.72246*	-2.72246**
	basal-BR III	1.000 (0.126)	0.00417 (0.00073)	-1.18441	-1.18441	-1.24511
	Asian-BR	0.939 (0.058)	0.00661 (0.00130)	-0.95449	-0.79711	-0.95432
Weifang	basal-BR II	0.960 (0.031)	0.00490 (0.00073)	-2.39975**	-3.45534 **	-3.66541**
	Asian-BR	0.987 (0.023)	0.01159 (0.00161)	-1.22279	-1.79120	-1.88684
Changchun	basal-BR	0.982 (0.046)	0.00478 (0.00095)	-1.63909	-2.06292	-2.21575

265 The sub-populations less than four isolates were not included. *: $P < 0.05$, **: $P < 0.02$.

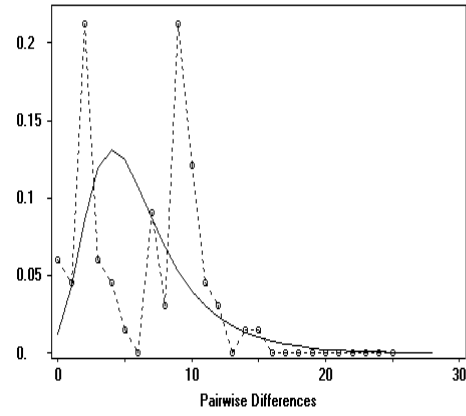
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267 The mismatch distribution of TuMV CP gene and 3'-UTR for the basal-BR II isolates
 268 collected from Weifang, Tai'an, Changchun and basal-BR III were unimodal and smooth,
 269 and fit well with the expected model of sudden expansion, indicating that these
 270 sub-populations were new emergent (Fig. 4). The Asian-BR isolates from Weifang and
 271 Tai'an of Shandong province and Zhejiang were multiple-peaked, ragged, indicating that
 272 these sub-populations were long-existing ones (Fig. 4).

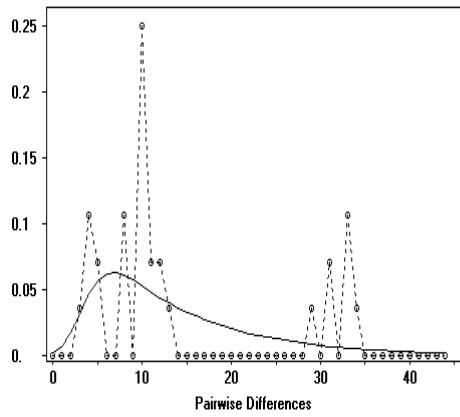
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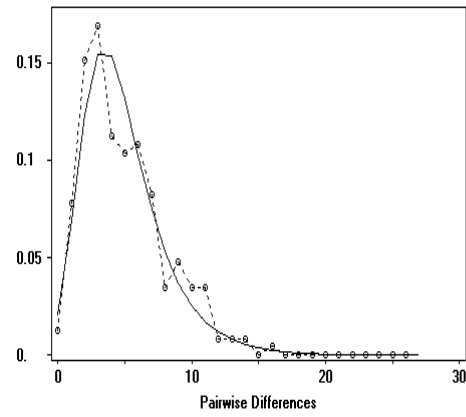
A. Weifang Asian-BR group



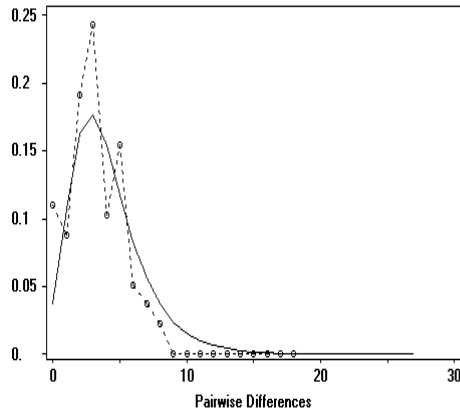
B. Taian Asian-BR group



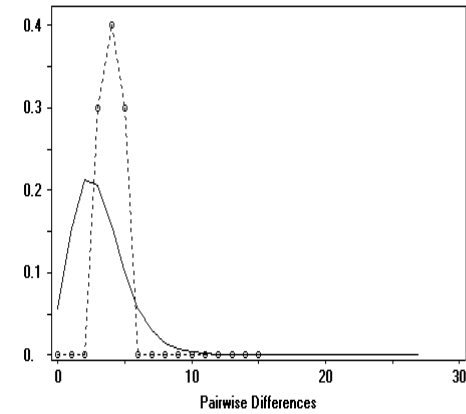
C. Zhejiang Asian-BR group



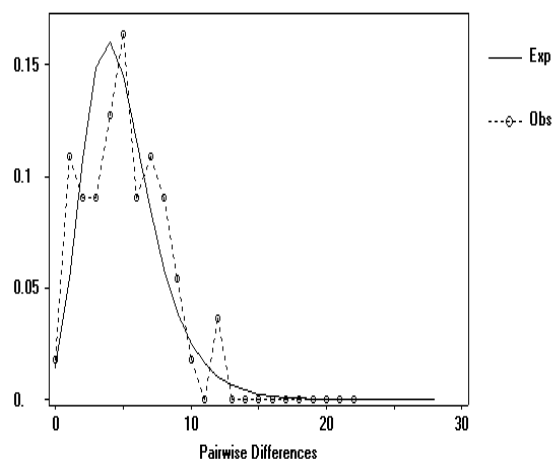
D. Weifang basal-BR II group



E. Tai'an basal-BR II group



F. basal-BR III



G. Changchun basal-BR group

274 **Fig. 4.** The frequency distribution of the number of pairwise nucleotide differences obtained from CP
 275 gene nucleotide sequences. (A) basal-BR II group of Weifang; (B) Asian-BR group of Weifang; (C)
 276 basal-BR II group of Tai'an, (D) basal-BR III group of Tai'an; (E) basal-BR II group of Tai'an, (F)
 277 basal-BR III group; (G) Changchun isolates of basal-BR group. Broken line represents the observed data
 278 and unbroken line represents the expected data. The sub-populations less than four isolates were not
 279 included.

280 4. Discussion

281 In this paper, we studied the molecular structure of TuMV population from China by
 282 analyzing the CP gene sequence of 129 TuMV isolates from radish and comparing them with
 283 41 isolates of basal-BR group from Japan. Our results show that (1) about one-tenth of the
 284 TuMV isolates characterized are recombinants; (2) sub-population of basal-BR expands
 285 rapidly and accounts to more than one half of the isolates detected; (3) isolates of basal-BR
 286 in China evolve to three sub-groups, with sub-groups I and II genetically homologous with
 287 Japanese ones, while sub-group III a distinct lineage; (4) Sub-populations of TuMV
 288 basal-BR II and III are new emergent and in a state of expansion; (5) the TuMV population
 289 of China is under negative selection; (6) frequent gene flow is detected between TuMV
 290 sub-populations from Weifang, Taian and Changchun.

291 Recombination is important in virus evolution and has been detected in many potyvirus
 292 species[19, 20, 24, 45-48]. The percentages of recombinant isolates may account to ten to
 293 sixty-five of isolates studied[24, 47]. Intra- and inter-lineage recombination is very common
 294 in TuMV[6, 18, 22]. The hotspots of recombination sites of TuMV genome are located in the
 295 P1 and CI/6K2/VPg region[21]. Ohshima and colleagues have detected 37 recombination
 296 patterns[6, 21]. Novel recombination patterns of TuMV are increasing[6, 18]. About 10% of

297 the TuMV isolates characterized in this study experienced ‘clear’ recombination event. The
298 percentage is a little lower than previous studies[18, 22]. The underlying reason is that we
299 just analyzed the CP-UTR region, where the crossover sites of TuMV are scarce[18, 22, 23].
300 If longer sequences or the whole genome is included, there will be more recombination
301 events detected.

302 The d_N/d_S ratios are often used to estimate the selection pressure under which viral
303 gene(s) suffered[10, 49]. Positive selection ($d_N/d_S > 1$) may endow the virus more fitness to
304 adapt a new host or environment. However, rapid divergence driven by positive selection has
305 been rarely demonstrated[50]. Like the case of most virus genes, our results show that
306 negative (purifying) selection dominates the evolution of TuMV CP genes. If selection
307 pressure on single residue is estimated, amino acids under positive selection may be sorted
308 out[49].

309 Basal-BR is a new emergent in east Asia and has been detected in Japan and China [6,
310 17, 23, 28]. So far, there has been no basal-BR isolates reported in Vietnam[20]. After its
311 first detection in 2005[28], the population of basal-BR isolates increased rapidly in China
312 and showed characteristics of founder effect. As reported in this research, Basal-BR isolates
313 were detected from samples from Hebei, Henan, Jilin and Shandong provinces, and
314 accounted to more than half of the isolates from Shandong and Jilin provinces. The Chinese
315 basal-BR isolates have evolved to three sub-groups. Among the 48 basal-BR isolates from
316 Weifang and Tai’an of Shandong province, 40 belonged to sub-group II, which represents
317 the prevalent cluster in those areas. Basal-BR III was detected after 2006 and only found in
318 Tai’an of Shandong province. What’s more interesting, sub-groups of basal-BR I and II are
319 genetically homologous to those of Japanese isolates, while sub-groups of basal-BR III
320 from China and Japan are genetically distinct and form separate clusters, indicating that
321 China and Japan had different sub-populations. Another difference is that the prevalent
322 subgroup of Basal-BR is II in China but III in Japan.

323 The gene flow between TuMV isolates of basal-BR II and III from Weifang, Tai’an and
324 Changchun is frequent. But TuMV is transmitted by aphid in a non-persistent manner and
325 there is no evidence of seed transmission reported[4, 5]. It remains unknown how TuMV
326 isolates, especially the new emergent, spread to other places[18, 23, 25]. But TuMV isolates
327 of basal-BR II are prevalent and expanding rapidly in Weifang and Tai’an of Shandong and
328 Changchun of Jilin. Therefore, a program should be launched to evaluate the resistance of
329 commercial available cultivars of cruciferous crops to TuMV isolates, particularly basal-BR
330 II.

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343 References

- 344 1. Roossinck, M. J., Mechanisms of plant virus evolution. *Annual Review of Phytopathology* **1997**, 35,
 345 (35), 191.
- 346 2. Roossinck, M. J., *Plant Virus Evolution*. Springer Berlin Heidelberg: 2008; p 157-164.
- 347 3. Garcíaarenal, F.; Fraile, A.; Malpica, J. M., Variability and genetic structure of plant virus populations.
 348 *Annual Review of Phytopathology* **2001**, 39, (1), 157.
- 349 4. Tomlinson, J. A., Epidemiology and control of virus diseases of vegetables. *Annals of Applied Biology*
 350 **1987**, 110, (3), 661-681.
- 351 5. Shukla, D. D.; Ward, C. W.; Brunt, A. A., The Potyviridae. **1994**.
- 352 6. Wang, H. Y.; Liu, J. L.; Gao, R.; Chen, J.; Shao, Y. H.; Li, X. D., Complete genomic sequence
 353 analyses of Turnip mosaic virus basal-BR isolates from China. *Virus Genes* **2009**, 38, (3), 421-8.
- 354 7. Riechmann, J. L.; Laín, S.; García, J. A., Highlights and prospects of potyvirus molecular biology.
 355 *Journal of General Virology* **1992**, 73 (Pt 1), (4), 1-16.
- 356 8. Urcuqui-Inchima, S.; Haenni, A. L.; Bernardi, F., Potyvirus proteins: a wealth of functions. *Virus*
 357 *Research* **2001**, 74, (1-2), 157.
- 358 9. Chung, B. Y.; Miller, W. A.; Atkins, J. F.; Firth, A. E., An overlapping essential gene in the
 359 Potyviridae. *Proceedings of the National Academy of Sciences of the United States of America* **2008**,
 360 105, (15), 5897-5902.
- 361 10. Wei, T.; Zhang, C.; Hong, J.; Xiong, R.; Kasschau, K. D.; Zhou, X.; Carrington, J. C.; Wang, A.,
 362 Formation of complexes at plasmodesmata for potyvirus intercellular movement is mediated by the
 363 viral protein P3N-PIPO. *Plos Pathogens* **2010**, 6, (6), e1000962.
- 364 11. Edwardson, J. R.; Christie, R. G., The Potyvirus Group. *University of Florida Monograph* **1991**, 3,
 365 699-712.
- 366 12. Shattuck, V. I., The Biology, Epidemiology, and Control of Turnip Mosaic Virus. *Horticultural*
 367 *Reviews* **2010**, 14, 199-238.
- 368 13. Green, S. K.; Deng, T. C., Turnip mosaic virus strains in cruciferous hosts in Taiwan. *Plant Disease*
 369 **1985**, 69, (69), 28-31.
- 370 14. Liu, X. P.; Lu, W. C.; Li, J. L.; Liu, Y. K., A study on TuMV strain differentiation of cruciferous
 371 vegetables from ten provinces in China- Selection of new identification host and strains portion.
 372 *Science Bulletin* **1990**, 35, (20), 1734-1739.
- 373 15. Jenner, C. E.; Keane, G. J.; Jones, J. E.; Walsh, J. A., Serotypic variation in turnip mosaic virus. *Plant*
 374 *Pathology* **1999**, 48, (1), 101-108.
- 375 16. Jenner, C. E.; Walsh, J. A., Pathotypic variation in turnip mosaic virus with special reference to
 376 European isolates. *Plant Pathology* **1996**, 45, (5), 848-856.
- 377 17. Ohshima, K. Y. Y.; Hirota, R.; Hamamoto, T.; Tomimura, K.; Tan, Z. S. T.; Azuhata, F.; Walsh, J. A.;
 378 Fletcher, J.; Chen, J. S.; Gera, A., Molecular evolution of Turnip mosaic virus: evidence of host

- 379 adaptation, genetic recombination and geographical spread. *Journal of General Virology* **2002**, 83, (6),
380 1511-1521.
- 381 18. Nguyen, H. D.; Tomitaka, Y.; Ho, S. Y. W.; Duchêne, S.; Vetten, H. J.; Lesemann, D.; Walsh, J. A.;
382 Gibbs, A. J.; Ohshima, K., Turnip Mosaic Potyvirus Probably First Spread to Eurasian Brassica Crops
383 from Wild Orchids about 1000 Years Ago. *Plos One* **2013**, 8, (2), e55336.
- 384 19. Chare, E. R.; Holmes, E. C., A phylogenetic survey of recombination frequency in plant RNA viruses.
385 *Archives of Virology* **2006**, 151, (5), 933-946.
- 386 20. Nguyen, H. D.; Tran, H. T.; Ohshima, K., Genetic variation of the Turnip mosaic virus population of
387 Vietnam: a case study of founder, regional and local influences. *Virus Research* **2013**, 171, (1),
388 138-149.
- 389 21. Ohshima, K.; Tomitaka, Y.; Wood, J. T.; Minematsu, Y.; Kajiyama, H.; Tomimura, K.; Gibbs, A. J.,
390 Patterns of recombination in turnip mosaic virus genomic sequences indicate hotspots of
391 recombination. *Journal of General Virology* **2007**, 88, (Pt 1), 298-315.
- 392 22. Tan, Z.; Wada, Y.; Chen, J.; Ohshima, K., Inter- and intralineage recombinants are common in natural
393 populations of Turnip mosaic virus. *Journal of General Virology* **2004**, 85, (9), 2683-96.
- 394 23. Tomitaka, Y.; Ohshima, K., A phylogeographical study of the Turnip mosaic virus population in East
395 Asia reveals an 'emergent' lineage in Japan. *Molecular Ecology* **2007**, 15, (14), 4437-4457.
- 396 24. Tomimura, K.; Gibbs, A. J.; Jenner, C. E.; Walsh, J. A.; Ohshima, K., The phylogeny of Turnip
397 mosaic virus; comparisons of 38 genomic sequences reveal a Eurasian origin and a recent 'emergence'
398 in east Asia. *Molecular Ecology* **2003**, 12, (8), 2099-2111.
- 399 25. Tomitaka, Y.; Yamashita, T.; Ohshima, K., The genetic structure of populations of Turnip mosaic
400 virus in Kyushu and central Honshu, Japan. *Journal of General Plant Pathology* **2007**, 73, (3),
401 197-208.
- 402 26. Shi, M. L., Cloning and sequence analysis of HC-Pro genes of Turnip mosaic virus Eurasian isolates.
403 *Acta Phytopathologica Sinica* **2007**, 383-389.
- 404 27. Song, Y., Cloning and sequence analysis of coat protein genes of turnip mosaic virus isolates obtained
405 from shandong. *Scientia Agricultura Sinica* **2005**, 58, (3), 504-510.
- 406 28. Tian, Y. P.; Zhu, X. P.; Liu, J. L.; Yu, X. Q.; Du, J.; Kreuzer, J.; Li, X. D., Molecular Characterization
407 of the 3'-Terminal Region of Turnip mosaic virus Isolates from Eastern China. *Journal of*
408 *Phytopathology* **2007**, 155, (155), 333-341.
- 409 29. Liu, J. L.; Yu, X. Q.; Tian, Y. P.; Du, J.; Zhu, X. P.; Li, X. D.; Yan, D. Y., Molecular characterization
410 and coat protein gene expression of a turnip mosaic virus isolate from radish in Weifang. *Acta*
411 *Horticulturae Sinica* **2006**, 33, 84-88.
- 412 30. Martin, D. P.; Lemey, P.; Lott, M.; Moulton, V.; Posada, D.; Lefevre, P., RDP3: a flexible and fast
413 computer program for analyzing recombination. *Bioinformatics* **2010**, 26, (19), 2462-2463.
- 414 31. Sawyer, S. A., GENECONV: a computer package for the statistical detection of gene conversion.
415 *Washington University in St. Louis* **1999**, <http://www.math.wustl.edu/~sawyer>.
- 416 32. Salminen, M. O.; Carr, J. K.; Burke, D. S.; Mccutchan, F. E., Identification of breakpoints in
417 intergenotypic recombinants of HIV type 1 by bootscanning. *Aids Research & Human Retroviruses*
418 **1995**, 11, (11), 1423-5.
- 419 33. Smith, J. M., Analyzing the mosaic structure of genes. *Journal of Molecular Evolution* **1992**, 34, (2),
420 126-9.
- 421 34. Posada, D.; Crandall, K. A., Evaluation of methods for detecting recombination from DNA sequences:
422 computer simulations. *Proceedings of the National Academy of Sciences of the United States of*
423 *America* **2001**, 98, (24), 13757-62.
- 424 35. Weiller, G. F., Phylogenetic profiles: a graphical method for detecting genetic recombinations in
425 homologous sequences. *Molecular Biology & Evolution* **1998**, 15, (3), 326-35.
- 426 36. Tamura, K.; Stecher, G.; Peterson, D.; Filipski, A.; Kumar, S., MEGA6: Molecular Evolutionary
427 Genetics Analysis version 6.0. *Molecular Biology & Evolution* **2013**, 44, (4), 2725.
- 428 37. Chen, J.; Chen, J. P.; Langeveld, S. A.; Derks, A. F. L. M.; Adams, M. J., Molecular characterization
429 of carla- and potyvirus from Narcissus in China. *Journal of Phytopathology* **2003**, 151, (1), 26-29.
- 430 38. Fu, Y. X.; Li, W. H., Statistical tests of neutrality of mutations. *Genetics* **1993**, 133, (3), 693.
- 431 39. Tajima, F., Statistical method for testing the neutral mutation hypothesis by DNA polymorphism.
432 *Genetics* **1989**, 123, (3), 585-95.
- 433 40. Librado, P.; Rozas, J., DnaSP v5: a software for comprehensive analysis of DNA polymorphism data.
434 *Bioinformatics* **2009**, 25, (11), 1451-1452.
- 435 41. Pamilo, P.; Bianchi, N. O., Evolution of the Zfx and Zfy genes: rates and interdependence between the
436 genes. *Molecular Biology & Evolution* **1993**, 10, (2), 271.
- 437 42. Li, W. H., Unbiased estimation of the rates of synonymous and nonsynonymous substitution. *Journal*
438 *of Molecular Evolution* **1993**, 36, (1), 96-99.

- 439 43. Hudson, R. R., A new statistic for detecting genetic differentiation. *Genetics* **2000**, 155, (4), 2011.
- 440 44. Hudson, R. R.; Boos, D. D.; Kaplan, N. L., A statistical test for detecting geographic subdivision.
- 441 *Molecular Biology & Evolution* **1992**, 9, (1), 138.
- 442 45. Moreno, I. M.; Malpica, J. M.; Díaz-Pendón, J. A.; Moriones, E.; Fraile, A.; García-Arenal, F.,
- 443 Variability and genetic structure of the population of watermelon mosaic virus infecting melon in
- 444 Spain. *Virology* **2004**, 318, (1), 451-60.
- 445 46. Alla G Gagarinova, M. B., Martina V Strömvik, Aiming Wang, Recombination analysis of Soybean
- 446 mosaic virus sequences reveals evidence of RNA recombination between distinct pathotypes. *Virology*
- 447 *Journal* **2008**, 5, (1), 1-8.
- 448 47. Tugume, A. K.; Cuéllar, W. J.; Mukasa, S. B.; Valkonen, J. P. T., Molecular genetic analysis of virus
- 449 isolates from wild and cultivated plants demonstrates that East Africa is a hotspot for the evolution
- 450 and diversification of Sweet potato feathery mottle virus. *Molecular Ecology* **2010**, 19, (15),
- 451 3139-3156.
- 452 48. Zhang, C. L.; Gao, R.; Wang, J.; Zhang, G. M.; Li, X. D.; Liu, H. T., Molecular variability of Tobacco
- 453 vein banding mosaic virus populations. *Virus Research* **2011**, 158, (1-2), 188.
- 454 49. Tian, Y. P.; Liu, J. L.; Zhang, C. L.; Liu, Y. Y.; Wang, B.; Li, X. D.; Guo, Z. K.; Valkonen, J. P.,
- 455 Genetic diversity of Potato virus Y infecting tobacco crops in China. *Phytopathology* **2011**, 101, (3),
- 456 377.
- 457 50. Nielsen, R., Changes in ds/dn in the HIV-1 env gene. *Molecular Biology & Evolution* **1999**, 16, (5),
- 458 711-714.
- 459
- 460