Synthesis and Anti-Tumor Activity of Oleanolic Acid

Derivatives

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Abstract: With the computer drug-aided design and the key amino acid residue fragment of the target protein analyzed on the basis of simulated docking of Survivin and known active small molecules, the active groups capable of binding to the critical sites were determined. After the natural product Oleanolic acid was used as lead compound, then the active groups were introduced on the ring of A, next the carboxyl group at the C-28 position was modified by esterification or amidation, twenty new Oleanolic acid derivatives had been designed and synthesized. SKOV3 and BGC-823 cells were used to screen the antitumor activity in vitro through the standard MTT method. Among the selection, compounds II_3 , III_5 and IV_4 exhibited more potent cytotoxicity than positive drugs.

Keywords: oleanolic acid derivatives; synthesis; anti-tumor activity; molecular docking

1. Introduction

Oleanolic acid (OA) is a pentacyclic triterpenoid compound, which is widely present in plants in the form of dissociation or in combination with sugars ^[1]. An active ingredients, mainly separated from Prunella vulgaris, Honeysuckle, Olive, Forsythia, Aloe Vera and Ligustrum lucidum and other plants ^[2], oleanolic acid has a variety of important pharmacological functions, such as anti-virus ^[3-5], anti-diabetic and hypoglycemic, anti-HBV, and anti-tumor. Our research group has carried out the anti-tumor research in pentacyclic triterpenoid analogues since 2000, which mainly focuses on the extensive structural modification of inactylactic acid, ursolic acid, asiatic acid and in vitro antitumor activity evaluation ^[6-9]. In addition, some progress has been made in guiding the structural transformation of oleanolic acid, ursolic acid and asiatic acid on the basis of the combination of computer-aided design analogue analyte and protein target ^[10-11].

Survivin, a member of apoptotic proteins family, is considered as an important target for anticancer therapy ^[12] due to its function in cell division and suppression. Oleanolic acid (OA) and 5-FU, combined together, impact significantly the human pancreatic cell (Panc-28) apoptosis through inhibiting Survivin ^[13]. By improving the proapoptotic protein Bax and changing the balance of Bcl-2/Bax, OA significantly reduce the expression of anti-tumor apoptotic protein Survivin ^[14]. YM155 has a naphthoquinolidazole structure that inhibits the transformation of Survivin gene through inhibiting the promoter activity ^[15-16]. And it has been in the clinical stage I, II study. SC144, with structure of quinolide hydrazine, can inhibit the IL-6/gp130/Stat3

*Corresponding author. Tel.: 15940588671. E-mail addresses: myq6581@163.com signal axis, bind gp130, induce gp130 phosphorylation and de-glycosylation, prevent Stat3 phosphorylation and nuclear migration, and ultimately inhibit the downstream expression of Survivin^[17]. It is the first oral active inhibitor of gp130. After combining the three-dimensional crystal structure (PDB: 3UIH) of Survivin protein in PDB database, and analyzing the interaction between known Survivin small molecule inhibitor and target enzyme by molecular simulation docking method, and studding the key amino acid residue fragment, we determine the active site that binds to the critical site, which is shown on Figure 1. And the active group fragments in the partial inhibitor structure were introduced into the tricyclic triterpenoid matrix and the main transformation concentrated in the A-ring parallel nitrogen-containing heterocyclic ring, have shown in Figure 2. Twenty new oleanolic acid analogs were designed and synthesized and tested for in vitro activity.



Figure1. Analysis of the Interaction of YM155 and SC144 with 3UIH, the key amino

acids:ASP71,GLU63,GLU65



Figure2. The analytical active groups introduce oleanolic acid and design new derivatives

In this paper, we put together the structural modification of pentacyclic triterpene compounds suggested by Meng's study team, the antitumor compounds which have entered clinical research (Figure 1), and the computer simulation to analyze the structure of the key active groups (Figure 2). We introduce these groups in A ring of oleanolic acid. Through computer molecular docking simulation analysis, the combination mode of $II_{3,s}$ III₅ and IV₄ with Survivin targeting protein displays a strong combining ability. The antitumor activities have been tested *in vitro* by MTT. The results showed that compounds III₅ and IV₄ have more outstanding antitumor activities on SKOV3 and BGC-823 cells than Adriamycin.

2. Results and discussion

2.1. Chemical Synthesis

A total of 20 compounds were designed and synthesized from four categories with OA as original material. The target compound route was shown in Scheme 1. First, the Jones reagent was used to oxidize the oleanolic acid at the 3-position hydroxyl group to the ketone to obtain OA-1. On this basis, the 28-carboxyl group was esterified or amidized to obtain the compound OA-2 and OA-6. Then they react with (T-BuOK) respectively, result in compounds OA-3 and OA-7 under basic

conditions at 50 °C , which are further synthesized to result in olean-2-ene-[2,3-b] pyrazin-12-ene-28-carboxylic acid ester compounds I₁ to I₄, 5'-methyl-olean-2-ene-[2,3-b] pyrazine-12-ene-28-oic acid ester compounds II₁ to II₅, and 5'-methyl-olean-2-ene-[2,3-b] pyrazine-12-ene-28-carboxylic acid amide compounds IV₁ to IV₅. In addition, OA-2 reacted with hydrazine hydrate to gain OA-5, which, after cyclization, forms olean-2-ene-[2,3-b] imidazole-12-ene-28-oic acid ester III₁ to III₅.



Comp.	R_1	Comp.	R ₁	Comp.	R_1	Comp.	R ₂
I_1	$-C_2H_5$	II ₁	$-C_2H_5$	III_1	$-C_2H_5$	IV_1	
I_2	—C ₃ H ₇	II_2	—C ₃ H ₇	III_2	—C ₃ H ₇	IV ₂	
I ₃	$-C_4H_9$	II ₃	$-C_4H_9$	III ₃	—C₄H ₉	IV ₃	
I_4	—C ₅ H ₁₁	II_4	$-C_{5}H_{11}$	III_4		IV_4	
I ₅	$-C_{6}H_{13}$	II_5	—C ₆ H ₁₃	III ₅	—C ₆ H ₁₃	IV ₅	-N_O

$$\label{eq:response} \begin{split} \text{Reagents and conditions:(a)Jones,acetone,r.t.,1h;(b)RBr,K_2CO_3,DMF;(c)t-BuOH,t-BuOK,reflux,5h,50^{\circ}C; \\ (d)EtOH,C_2H_8N_2,reflux,6h;(e)KOH,MnO_2,EtOH,r.t.;(f)EtOH,C_3H_{11}N_2,reflux,6h;(g)N_2H_4`H_2O,EtOH,reflux,1h; \\ (h)HCOOH,reflux,3h;(i)DCM,(COCl)_2,DMF,r.t.4h;(j)amins,DCM,r.t.24h; \end{split}$$

Scheme1 Synthetic routes of target compounds.

2.2. Biological evaluation

The inhibitory activity of target compounds I_{1-5} , II_{1-5} , III_{1-5} , IV_{1-5} on SKOV3 and BGC-823 cells was tested *in vitro* by the MTT method, meanwhile gefitinib and 5-FU acted as the positive control. As shown in Table 1, all the tested compounds indicated

some inhibitory effect on SKOV3 and BGC-823 cells. Among the compounds, II_{35} III₅ and IV₄ showed excellent inhibitory effects on SKOV3 cells (IC₅₀ =8.3µM, IC₅₀ =7.8µM, IC₅₀ =8.0µM) and BGC-823 cells (IC₅₀=6.3µM,IC₅₀ =4.3µM,IC₅₀ =7.9µM), which manifested a 7.1,10.4 and 5.6-fold to enhance the anticancer activity compared to that of Adriamycin, respectively.

Through the analysis of tumor cells morphological variation, we found that the distribution of the diamond BGC-823 cells showed fragmental, cavitation, and apopt - osis gradually after 72 hours. Whereas, the blank control group cell lines had been in malignant proliferation state. By contrast, we can conclude that compounds III₅ and IV_4 have significant inhibitory activity on SKOV3 and BGC-823 cells. The results have shown in Figure 3 and Figure 4.



Figure 3. Morphological changes of SKOV3 cells induced by different compounds, 72 h a.Normal SKOV3 cells b. OA c. III₅ d. IV₄ e.5-FU f. Adriamycin



Figure 4. Morphological changes of BGC-823 cells induced by different compounds, 72 h a.Normal BGC-823 cells b. OA c. III₅ d. IV₄ e.5-FU f. Adriamycin

Compd -	Inhibition	Rate (%) ^a	IC50 (µM) ^b		
Compu.	SKOV3	BGC-823	SKOV3	BGC-823	
OA	12.7	10.3	>50	>50	
I ₁	29.5	30.6	>50	>50	
I_2	35.7	33.2	35.3	36.7	
I ₃	39.2	30.3	30.9	40.1	
I4	32.1	26.5	>50	>50	
I ₅	41.5	43.1	27.5	25.3	
II_1	36.2	37.3	32.6	>50	
II_2	27.9	26.7	>50	>50	
II ₃	57.3	59.6	8.3	6.2	
II4	39.1	38.4	30.2	33.6	
II_5	35.6	37.2	40.3	>50	
III_1	42.6	44.3	26.7	21.3	
III ₂	43.5	41.6	25.6	28.3	
III ₃	56.3	49.2	10.2	17.6	
III4	38.9	36.2	32.6	35.7	
III5	60.9	63.4	7.8	4.3	
IV 1	57.8	55.1	8.0	10.1	
IV 2	37.8	35.6	42.6	>50	
IV ₃	33.4	31.5	>50	>50	
IV_4	62.1	58.3	5.3	7.9	
IV_5	40.1	38.5	29.7	33.6	
5-FU	42.3	45.6	26.3	21.1	
Adriamycin	46.8	37.9	30.6	44.6	

Table 1. Antitumor activity of the target compounds on SKOV3 and BGC-823 cell lines.

a. Inhibitory percentage of cells treated with each compound at a concentration of 10 μ M for 96 h; *b*. The agent concentration that inhibited SKOV3 and BGC-823 cells growth by 50%.

2.3. Molecular docking

Oleanolic acid derivatives I_{1-5} , II_{1-5} , IV_{1-5} have the combined affinity with Survivin protein (PDB code: 3UIH). Molegro Virtual Docker (MVD) is able to predict how protein interacts with macromolecular ligands and its interaction energy. The connection between 3UIH and molecules was assessed by MVD. Scores are expressed as binding free energy (E_{score} kcal/mol). Using MVD 6.0, the binding scores showed that compound II_3 : -76.432, compound III_5 : -86.021, compound IV_4 :-78.851, in comparison with protein's small molecule ligands (-75.542). The lower the energy score is, the stronger the binding affinity becomes. Through intuiting analysis of the combination of compounds and targets by the Discovery Studio 4.0, we found that some compounds interacted with Survivin protein and closely connected with the surrounding amino acids. Based on Figure 5, Figure 6 and Figure 7, compounds II_3 , III_5 and IV_4 were firmly fixed in the hydrophobic pocket and they interacted with key amino acids through hydrophobic bond and H-bond. Key energy could not be measured in the process of the molecular docking. MolDock score of all compounds are shown in Table 2. In this step, we cooperated with Shenyang Pharmaceutical University.



Figure 5. Binding of compound **II**₃ to the active site of Survivin, it exhibited 1 H-bond with ASP71, the hydrogen bonds formed colored in green.



Figure 6. Binding of compound **III**₅ to the active site of Survivin, it exhibited 2 H-bonds with LYS62 and ASP71, the hydrogen bonds formed colored in green.



Figure 7. Binding of compound IV_4 to the active site of Survivin, it exhibited 1 H-bonds with GLU65, the hydrogen bonds formed colored in green.

Compd.	MolDock Score	Compd.	MolDock Score
I ₁	-67.387	III ₁	-74.600
I_2	-69.850	III ₂	-71.915
I ₃	-72.289	III3	-76.533
I ₄	-65.424	III4	-69.951
I5	-73.277	III5	-90.022
II_1	-68.272	IV_1	-74.103
II_2	-64.290	IV_2	-78.620
II ₃	-80.432	IV 3	-60.720
II_4	-69.015	IV_4	-80.851
II_5	-68.397	IV_5	-71.225
Molecule	-75.542		

 Table 2.
 Comparison of energy scores for different compounds with Survivin protein.

3. Conclusions

In summary, four new series of OA derivatives were designed and synthesized. Their antitumor activities on SKOV3 and BGC-823 cell lines were evaluated. All the tested compounds showed some anticancer activity against SKOV3 and BGC-823 cell lines. Molecular docking studies demonstrated that twenty OA derivatives were obtained through structural optimization of the lead compound(OA) and they docked into Survivin protein-tyrosine kinase. Molegro Virtual Docker (MVD) was able to tell if there was good binding affinity of the synthesized all compounds with Survivin protein. Specifically, compounds H_3 , HI_5 and IV_4 exhibited outstanding inhibitory activities on SKOV3 cells ($IC_{50} = 8.3 \mu$ M, $IC_{50} = 7.8 \mu$ M, $IC_{50} = 8.0 \mu$ M) and BGC-823 cells ($IC_{50}=6.3 \mu$ M, $IC_{50}=4.3 \mu$ M, $IC_{50}=7.9 \mu$ M). They showed a 7.1,10.4 and 5.6-fold enhanced anticancer activity, compared with the positive control Adriamycin.As the ester chain of OA increases, the anticancer activity increases. The structure-activity relationships of newly synthesized compounds have shown in Figure 8. Our data indicated that proper structural modification at A ring and C-28 position of OA is necessary to enhance the anticancer activity of Oleanolic acid.



Figure 8. Summarized structure-activity relationships of novel compounds with regard to cancer inhibition.

4. Experimental

4.1. General experimental procedures

The melting points were determined on a Büchi B-540 melting point apparatus produced by Broker Corporation (Flawil, Switzerland) and are uncorrected. ¹HNMR spectra were recorded on Bruker a ARX-300 MHz spectrometers from Bruker Corporation (Ettlingen, Germany) and the solvent is CDCl₃, using trimethylsilane as an internal standard. ESI-MS were measured on a Thermo-Finnigan LCQ equipment from Thermo Finnigan (San Francisco, CA, USA). Thin-layer chromatography (TLC) ware carried out with GF 254, column chromatograph with silica gel (200-300 mesh) obtained from Qing-dao Marine Chemical Factory (Qingdao, China). The reagents were all of analytical grade or chemically pure.

4.2. Preparation of the compounds

4.2.1. 3-Oxo-olean-12-ene-28-oic acid (OA-1)

OA (0.500g) was dissolved in 50 mL of acetone, and allowed to react with the newly prepared Jones' reagent (0.64mL) under ice bath. The end of the reaction was detected by TLC. 15 mL of isopropanol was added to quench its oxidizing property and stirred at room temperature for 30 min. A small amount of saturated sodium chloride solution and moderate ethyl acetate were added to the reaction mixture to extract for three times. The organic layer was dried with anhydrous magnesium sulfate for 4 h. The crude product was purified by silica gel column chromatography (eluent: petroleum ether/ethyl acetate =12/1(V/V)). Then, the solvent were removed to give a powder OA-1, with a yield of 98.0%. m.p. 200.4~202.1°C.

4.2.2. 3-Oxo-olean-12-ene-28-oic acid ethyl ester (OA-2)

To a solution of OA-1(0.500g) in N,N-dimethylformamide (DMF), were added anhydrous K₂CO₃ (30mg, 0.22 mmol) and bromoethane (0.24mL,5.02mmol) at room temperature for 5h. The end of the reaction was detected by TLC. (developing solvent: petroleum ether/ethyl acetate=5/1(V/V)). The crude product was purified by silica gel column chromatography (eluent: petroleum ether/ethyl acetate =15/1(V/V)) to give 0.324 g of white flaky solid OA-2, 57.9%. m.p.189.4~191.1 °C.

4.2.3. 2,3-Dioxo-olean-12-ene-28-oic acid ethyl ester (**OA-3**)

The intermediate OA-2 (0.300g) was dissolved in 10mL tert-butanol(t-BuOH) solution and reacted at 50 °C. After completely dissolving, 0.15 g of potassium tert-butoxide was added and 1.5 mL of tetrahy-drofuran was added as catalyst. The end of the reaction was detected by TLC.(developing solvent: petroleum ether/ethyl acetate=5/1(V/V)),6h reaction end. The organic layer was dried with anhydrous magnesium sulfate for 4 h.The crude product was purified by silica gel column chromatography (eluent: petroleum ether/ethyl acetate =15/1(V/V)) to give 0.245 g of white flaky solid OA-3 in yield 80.24%. m.p.187.7~187.2 °C.

4.2.4. 5', 6'-dihydro-olean-2-ene-[2,3-b]pyrazin-12-ene-28-oic acid ethyl ester (OA-4)

OA-3 (0.300 g) was dissolved in 10 mL of absolute ethanol and 0.5 g of anhydrous magnesium sulfate was added. The supersaturated ethylenediamine-ethanol solution (0.04 mL ethylenediamine) was slowly added to the system, and the mixture was refluxed at 79 °C for 8h. The end of the reaction was detected by TLC.(developing solvent: petroleum ether/ethyl acetate=5/1(V/V)), The crude product was purified by silica gel column chromatography (eluent: petroleum ether/ethyl acetate = 15/1(V/V)) to give 0.186 g of a pale yellow oil, OA-4, in a yield of 72.30 %. m.p.201.1~203.7 °C. 4.2.5 2-hydrazone-3-Oxo-olean-12-ene-28-oic acid ethyl ester (OA-5).

OA-2 (0.300 g, 0.62 mmol) was dissolved in 30 mL of methanol. 1 mL of hydrazine hydrate was added and heated to reflux. The end of the reaction was detected by TLC.(developing solvent: petroleum ether/ethyl acetate=3/1(V/V)), 2h reaction ends. The crude product was purified by silica gel column chromatography (eluent: petroleum ether/ethyl acetate =15/1(V/V)) to give 0.226 g of a yellow oil, OA-5, in a yield of 72.30% . m.p.201.1~203.7 °C.

4.2.6 2-Oxo-olean-12-ene-28-acyl-aniline (**OA-6**)

Take the intermediate OA-1 (0.500g), add 25mL of dry dichloromethane(DCM)and oxalyl chloride (1.20mmol) for 4h, the reaction solvent and unreacted oxalyl chloride were removed by steaming, and the residue was added with 10 mL of cyclohexane, followed by distillation of cyclohexane under reduced pressure and repeated operation three times. Acid chloride was added 15mL DCM, and triethylamine adjusted to pH 9 \sim 10,adding aniline (2.2mmol), room temperature reaction 6h, TLC detection reaction endpoint. The crude product was purified by silica gel column chromatography (eluent: petroleum ether/ethyl acetate = 20/1(V/V)) to give the desired product. 0.396 g of pale yellow crystals of OA-6, yield%. m.p.122.4~124.7 °C.

4.2.7 2,3-Dioxo-olean-12-ene-28-acyl-aniline(**OA-7**)

The intermediate OA-6(0.300g) was dissolved in 10mL tert-butanol(t-BuOH) solution and reacted at 50 °C. After completely dissolving, 0.15 g of potassium tert-butoxide was added and 1.5 mL of tetrahy-drofuran was added as catalyst. The end of the reaction was detected by TLC.(developing solvent: petroleum ether/ethyl acetate=5/1(V/V)),6h reaction end.The organic layer was dried with anhydrous magnesium sulfate for 4 h.The crude product was purified by silica gel column chromatography (eluent: petroleum ether / ethyl acetate =15/1(V/V)) to give 0.245 g of white flaky solid OA-3 in yield 81.67%. m.p.192.1~194.3 °C.

4.2.8 Olean-2-ene-[2,3-b] pyrazin-12-ene-28-oic acid ethyl ester (I_1)

The intermediate OA-4 (0.500 g) was dissolved in 20 mL of absolute ethanol, were added KOH (0.070 g, 1.00 mmol), MnO₂ (0.260, 3.00 mol) and reflux. The end of the reaction was detected by TLC.(developing solvent: petroleum ether/ester = 5/1(V/V)),6h reaction ended. The crude product was purified by silica gel column chromatography (eluent: petroleum ether/ethyl acetate =20/1(V/V)) to give 0.370 g of a white powder as a solid in 73.5% yield. m.p.134.3~138.1 °C.

¹H-NMR(CDCl₃,300MHz) δ :8.41~8.27 (m,2H,NCHCHN) ,5.38~5.29(m,1H,H-12),2.50(t,1H),4.12~4.05(m,2H,COOCH₂CH₃),1.93(t,*J*=7.0Hz,3H,COOCH₂CH₃),1.2 3(s,3H),1.18(s, *J*=7.6Hz, 3H, H-

18),1.14(s,3H),1.12(s,3H),1.02(s,3H),0.96(s,3H),0.95(s,3H),ESI-

MS(*m*/*z*):518.3[M+H]⁺. Elemental anal.(%) calcd. For C₃₄H₅₀N₂O₂: C 78.72,H 9.71,N 5.40,O 6.17;found: C 78.70,H 9.75,N 5.42,O 6.13.

4.2.9 Olean-2-ene-[2,3-b] pyrazin-12-ene-28-oic acid npropyl ester (I2)

According to the same method for compound I₁, compound I₂ was prepared from OA-1 (1.1mmol) and brominated npropane (4.4 mmol), The crude product was purified by silica gel column chromatography (eluent: petroleum ether/ethyl acetate=20/1 (V/V)) to give 0.330 g of a white powder as a solid in 72.4% yield. m.p.146.9~150.1 °C.

¹H-NMR(CDCl₃,300MHz) δ :8.40~8.24 (m,2H,NCHCHN) ,5.43~5.38(m,1H,H-12),1.84(t,1H *J*=7.5Hz,H-

18)4.21(t,2H,*J*=7.5Hz,COOC<u>H</u>₂CH₂CH₃),1.87(t,3H,*J*=11.7Hz,COOCH₂C<u>H</u>₃), 1.32(s,3H),1.25(s,3H),1.13(s,3H),1.12(s,3H),1.02(s,3H),0.94(s,3H),0.83(s,3H), ESI-MS(*m*/*z*):532.4[M+H]⁺. Elemental anal.(%) calcd. For C₃₅H₅₂N₂O₂: C 78.90,H 9.84,N 5.26,O 6.01;found: C 78.88,H 9.81,N 5.28,O 6.04.

4.2.10 Olean-2-ene-[2,3-b] pyrazin-12-ene-28-oic acid nbutyl ester (I3)

According to the same method for compound I₁, compound I₃ was prepared from OA-1 (1.1mmol) and brominated nbutylane (4.4 mmol), The crude product was purified by silica gel column chromatography (eluent: petroleum ether/ethyl acetate=20/1 (V/V)) to give 0.362 g of a white powder as a solid in 73.5% yield. m.p.156.1~158.2 °C. ¹H-NMR(CDCl₃,300MHz) δ :8.24~8.36 (m,2H,NCHCHN) ,5.43~5.38(m,1H,H-12),1.75(t, *J*=7.3Hz,1H,H-

18),4.04(t,J=7.5Hz,2H,COOCH2CH2CH2CH3),1.53(t,3H,J=10.3Hz,

COOCH₂CH₂CH₂CH₂CH₃),1.83~1.78(m,4H,COOCH₂(CH₂)₂CH₃).1.44(s,3H),1.29(s,3H), 1.13(s,3H),1.12(s,3H),0.92(s,3H),0.83(s,3H),0.75(s,3H), ESI-MS(*m/z*):544.7[M+H]⁺. Elemental anal.(%) calcd. For C₃₆H₅₄N₂O₂: C 79.07,H 9.95,N 5.12,O 5.85;found: C 79.03,H 9.99,N 5.10,O 5.87.

4.2.11 Olean-2-ene-[2,3-b] pyrazin-12-ene-28-oic acid npentyl ester(I4)

According to the same method for compound I₁, compound I₃ was prepared from OA-1 (1.1mmol) and brominated npentane (4.4 mmol), The crude product was purified by silica gel column chromatography (eluent: petroleum ether/ethyl acetate=20/1 (V/V)) to give 0.343g of a white powder as a solid in 68.3% yield. m.p.163.2~166.1 °C. ¹H-NMR(CDCl₃,300MHz) δ :8.39~8.25 (m,2H,NCHCHN), 5.45~5.10(m,1H,H-12),2.50(t, *J*=8.0Hz,1H,H-18),4.21(t, *J*=7.5Hz,2H,COOCH₂(CH₂)₃CH₃),1.87(t,3H, *J*=12.3Hz,COOCH₂(CH₂)₃CH₃),

1.67~1.42(m,6H,COOCH₂(C<u>H</u>₂)₃CH₃).1.41(s,3H),1.32(s,3H),1.13(s,3H),1.03(s,3H),0. 98(s,3H),0.86(s,3H),0.82(s,3H), ESI-MS(*m*/*z*):560.4[M+H]⁺. Elemental anal.(%) calcd. For C₃₇H₅₆N₂O₂: C 79.24,H 10.06,N 4.99,O 5.71;found: C 79.26,H 10.09,N 4.97,O 5.68.

4.2.12 Olean-2-ene-[2,3-b] pyrazin-12-ene-28-oic acid nhextyl ester (I₅)

According to the same method for compound I₁, compound I₃ was prepared from OA-1 (1.1mmol) and brominated nhexane (4.4 mmol), The crude product was purified by silica gel column chromatography (eluent: petroleum ether/ethyl acetate=20/1 (V/V)) to give 0.384g of a white powder as a solid in 76.3% yield. m.p.173.2~176.9°C. ¹H-NMR(CDCl₃,300MHz) δ :8.38~8.26 (m,2H,NCHCHN), 5.48~ 5.39(m,1H,H-12),1.74(t, *J*=7.6Hz,1H,H-18),4.13(t, *J*=7.0Hz,2H,COOCH₂(CH₂)₄CH₃),1.53(t, *J*=12.5Hz,3H, COOCH₂(CH₂)₄CH₃),

2.05~1.81(m,8H,COOCH₂(C<u>H</u>₂)₄CH₃).1.47(s,3H),1.29(s,3H),1.25(s,3H),1.05(s,3H),0. 99(s,3H),0.85(s,3H),0.79(s,3H), ESI-MS(*m/z*):574.3[M+H]⁺. Elemental anal.(%) calcd. For C₃₈H₅₈N₂O₂: C 79.39,H 10.17,N 4.87,O 5.57;found: C 79.36,H 10.13,N 4.91,O 5.60.

3.2.13 5'-methyl-olean-2-ene-[2,3-b] pyrazine-12-ene-28-oic acid ethyl ester (II_1)

The intermediate OA-3(0.500g) was dissolved in 20 mL of absolute ethanol and 0.5 g of anhydrous magnesium sulfate was added and the supersaturated 1,2propanediamine-ethanol solution (0.04 mL) was slowly added to the system, the mixture was stirred at 79 °C for 8 h. The solvent was removed by evaporation under reduced pressure. The crude product was purified by silica gel column chromatography (eluent: petroleum ether/ethyl acetate =15/1(V/V)) to give 0.389 g of a white powder as a yield of 73.5%. m.p.156.3~158.1°C.

¹H-NMR(CDCl₃,300MHz) δ :1.62~1.50 (m,1H,NC<u>H</u>CH₃CH₂N) ,1.08 (d, *J*=7.3Hz 3H,NCHC<u>H</u>₃CH₂N) , 1.41~1.23 (m,2H,NCHCH₃C<u>H</u>₂N) , 5.26~5.19(m,1H,H-12),2.65(t, *J*=7.0Hz,1H,H-18),4.21~4.17(m, *J*=8.0Hz,2H,COOC<u>H</u>₂CH₃),1.21(t, *J*=10.4Hz,3H,COOCH₂C<u>H</u>₃),

1.23(s,3H),1.18(s,3H),1.14(s,3H),1.01(s,3H),0.99(s,3H),0.89(s,3H),0.87(s,3H), ESI-MS(*m*/*z*):534.4[M+H]⁺. Elemental anal.(%) calcd. For C₃₅H₅₄N₂O₂: C 78.60,H 10.18,N 5.24,O 5.98;found: C 78.62,H 10.15,N 5.27,O 5.96.

3.2.14 5'-methyl-olean-2-ene- [2,3-b] pyrazine-12-ene-28-oic acid npropyl ester(II₂) According to the same method for compound II₁, compound II₂ was prepared from OA-1 (1.1mmol) and brominated npropane (4.4 mmol), The crude product was purified by silica gel column chromatography (eluent: petroleum ether/ethyl acetate=20/1 (V/V)) to give 0.283g of a white powder as a solid in 40.31% yield. m.p.168.8~171.3°C.

¹H-NMR(CDCl₃,300MHz) δ :1.63~1.55 (m,1H,NC<u>H</u>CH₃CH₂N) ,1.12 (d, *J*=7.8Hz, 3H,NCHC<u>H</u>₃CH₂N) , 1.42~1.21 (m,2H,NCHCH₃C<u>H</u>₂N) , 5.26~5.19(m,1H,H-12),2.67(t,1H,H-18),4.06(t, *J*=7.5Hz, 2H,COOC<u>H</u>₂CH₂CH₃),1.01(t, *J*=11.6Hz, 3H,COOCH₂CH₂C<u>H</u>₂C<u>H</u>₃),1.18(s,3H),1.16(s,3H),1.01(s,3H),1.01(s,3H),0.99(s,3H),0.94(s,3H),0.89(s,3H), ESI-MS(*m*/*z*):548.4[M+H]⁺. Elemental anal.(%) calcd. For C₃₆H₅₆N₂O₂: C 78.78,H 10.28,N 5.10,O 5.83;found: C 78.75,H 10.35,N 5.14,O 5.86. *3.2.15 5'-methyl-olean-2-ene-[2,3-b] pyrazine-12-ene-28-oic acid nbutyl ester (II₃)*

According to the same method for compound II₁, compound II₃ was prepared from OA-1 (1.1mmol) and brominated nbutylane (4.4 mmol), The crude product was purified by silica gel column chromatography (eluent: petroleum ether/ethyl acetate=20/1 (V/V)) to give 0.303g of a white powder as a solid in 60.24% yield. m.p.164.7~167.5°C. ¹H-NMR(CDCl₃,300MHz) δ :1.62~1.55 (m,1H,NCHCH₃CH₂N) ,1.08 (d, *J*=7.0Hz, 3H,NCHCH₃CH₂N) , 1.42~1.21 (m,2H,NCHCH₃CH₂N) 5.24~5.18(m,1H,H-12),2.65(t, *J*=7.5Hz,1H,H-18),4.10(t, *J*=6.5Hz,2H,COOCH₂(CH₂)₂CH₃), 1.83~1.78(m,4H,COOCH₂(CH₂)₂CH₃), 1.01(t, *J*=11.0Hz 3H,COOCH₂CH₂CH₂CH₂CH₂CH₂), 1.21(s,3H),1.17(s,3H),1.04(s,3H),1.03(s,3H),0.93(s,3H),0. 89(s,3H),0.88(s,3H), ESI-MS(*m*/*z*):562.5[M+H]⁺. Elemental anal.(%) calcd. For C₃₇H₅₈N₂O₂: C 78.95,H 10.39,N 4.98,O 5.68;found: C 78.98,H 10.35,N 5.00,O 5.67. *3.2.16 5'-methyl-olean-2-ene-[2,3-b] pyrazine-12-ene-28-oic acid npentyl ester (II*4)

According to the same method for compound II₁, compound II₄ was prepared from OA-1 (1.1mmol) and brominated npentane (4.4 mmol), The crude product was purified by silica gel column chromatography (eluent: petroleum ether/ethyl acetate=20/1 (V/V)) to give 0.322g of a white powder as a solid in 64.51% yield. m.p.179.7~183.5°C. ¹H-NMR(CDCl₃,300MHz) δ :1.61~1.47 (m,1H,NCHCH₃CH₂N) ,1.08 (d, *J*=7.0Hz, 3H,NCHCH₃CH₂N) , 1.42~1.21 (m,2H,NCHCH₃CH₂N) 5.22~5.19(m,1H,H-12),2.61(t, *J*=8.0Hz,1H,H-18), 4.06(t, *J*=6.5Hz,2H,COOCH₂(CH₂)₃CH₃), 1.68~1.39(m,6H, COOCH₂(CH₂)₃CH₃), 1.01(t,

J=12.5Hz,3H,COOCH₂(CH₂)₃C<u>H</u>₃),1.23(s,3H),1.19(s,3H),1.04(s,3H),1.04(s,3H),0.97 (s,3H),0.97(s,3H),0.87(s,3H), ESI-MS(*m*/*z*):576.5[M+H]⁺. Elemental anal.(%) calcd. For C₃₈H₆₀N₂O₂: C 79.11,H 10.48,N 4.86,O 5.55;found: C 79.14,H 10.45,N 4.83,O 5.58.

3.2.17 5'-methyl-olean-2-ene- [2,3-b] pyrazine-12-ene-28-oic acid nhextyl ester (II₅) According to the same method for compound II₁, compound II₄ was prepared from OA-1 (1.1mmol) and brominated nhexane (4.4 mmol), The crude product was purified by silica gel column chromatography (eluent:petroleum ether/ethyl acetate=20/1(V/V)) to give 0.304g of a white powder as a solid in 63.29% yield. m.p.196.2~199.7°C.¹H-NMR(CDCl₃,300MHz) δ:7.52 (s,1H,NCHN) , 5.24~5.19(m,1H,H-12),2.63(t, J=8.0Hz,1H,H-18),4.22~4.02(m,2H,COOCH₂CH₃),1.21(t,J=10.5Hz ,3H,COOCH₂CH₃),1.18(s,3H),0.

18),4.22~4.02(m,2H,COOC<u>H</u>₂CH₃),1.21(t,J=10.5Hz,3H,COOCH₂C<u>H</u>₃),1.18(s,3H),0.
99(s,3H),0.99(s,3H),1.01(s,3H),0.99(s,3H),0.89(s,3H),0.89(s,3H), ESI-MS(*m/z*):506.4[M+H]⁺. Elemental anal.(%) calcd. For C₃₉H₆₂N₂O₂: C 79.27,H
10.58,N 4.74,O 5.41;found: C 79.30,H 10.55,N 4.75,O 5.40.
4.2.18 olean-2-ene- [2,3-b] imidazole-12-ene-28-oic acid ethyl ester (III₁)

A solution of intermediate OA-5 (1.10 mmol) was dissolved in 20 mL of absolute ethanol, and formic acid (1.10 mmol) was added and heated to reflux.3h latter, the product was poured into cold water and filtered. After drying, the crude product was purified by silica gel column chromatography (eluent: petroleum ether/ethyl acetate=20/1(V/V)) to give 0.204 g of white crystals III₁ in 43.29% yield mp 196.2~199.7 °C. ¹H-NMR(CDCl₃,300MHz) δ :7.52 (s,1H,NCHN) , 5.24~5.19(m,1H,H-12),2.63(t, *J*=6.5Hz 1H,H-18),4.22~4.06(m,2H,COOCH₂CH₃),1.21(t,*J*=11.3Hz ,3H,COOCH₂CH₃),1.18(s,3H),0.

99(s,3H),0.99(s,3H),1.01(s,3H),0.99(s,3H),0.89(s,3H),0.89(s,3H), ESI-MS(*m*/*z*):506.4[M+H]⁺. Elemental anal.(%) calcd. For C₃₃H₅₀N₂O₂: C 78.21,H 9.95,N 5.53,O 6.31;found: C 78.24,H 9.97,N 5.49,O 6.29.

4.2.19 olean-2-ene-[2,3-b] imidazole-12-ene-28-oic acid npropyl ester(III2)

According to the same method for compound III₁, compound III₂ was prepared from OA-1 (1.1mmol) and brominated npropane (4.4 mmol), The crude product was purified by silica gel column chromatography (eluent: petroleum ether/ethyl acetate=20/1 (V/V)) to give 0.192g of a white powder as a solid in 40.15% yield. m.p.188.8~192.3°C. ¹H-NMR(CDCl₃,300MHz) δ :7.56 (s,1H,NCHN) , 5.24~5.15(m,1H,H-12),2.61(t, *J*=7.2Hz,1H,H-18),4.06(t, *J*=8.0Hz ,2H,COOC<u>H</u>₂CH₂CH₃), 1.01(t, *J*=10.5Hz,3H,COOCH₂CH₂C<u>H</u>₃), 1.25(s,3H),1.18(s,3H),1.01(s,3H), 1.01(s,3H),0.99(s,3H), 0.87(s,3H),0.87(s,3H),ESI-MS(*m/z*):520.4[M+H]⁺. Elemental anal.(%) calcd. For C₃₄H₅₂N₂O₂: C 78.41,H 10.06,N 5.38,O 6.14;found: C 78.38,H 10.08,N 5.41,O 6.11. *4.2.20 olean-2-ene-* [2,3-*b*] *imidazole-12-ene-28-oic acid nbutyl ester* (**III**₃)

According to the same method for compound III₁, compound III₃ was prepared from OA-1 (1.1mmol) and brominated nbutylane (4.4 mmol), The crude product was purified by silica gel column chromatography (eluent: petroleum ether/ethyl

acetate=20/1 (V/V)) to give 0.231g of a white powder as a solid in 45.35% yield.

m.p.176.3~180.1°C. ¹H-NMR(CDCl₃,300MHz) δ:7.61 (s,1H,NCHN),

5.24~5.19(m,1H,H-12),2.66(t, J=7.5Hz,1H,H-18),4.08(t,2H,

J=7.0Hz ,COOCH₂(CH₂)₂CH₃), 1.54~1.40(m,4H, COOCH₂(CH₂)₂CH₃), 0.90(t, J=12.5Hz,3H,COOCH₂(CH₂)₂CH₃), 1.31(s,3H),1.27(s,3H),1.21(s,3H), 1.18(s,3H),1.01(s,3H), 0.99(s,3H),ESI-MS(*m*/*z*):534.4[M+H]⁺. Elemental anal.(%) calcd. For C₃₅H₅₄N₂O₂: C 78.60,H 10.18,N 5.24,O 5.98;found: C 78.63,H 10.20,N 5.20,O 5.97.

4.2.21 olean-2-ene- [2,3-b] imidazole-12-ene-28-oic acid npentyl ester(III4)

According to the same method for compound III₁, compound III₃ was prepared from OA-1 (1.1mmol) and brominated nbutylane (4.4 mmol), The crude product was purified by silica gel column chromatography (eluent: petroleum ether/ethyl acetate=20/1 (V/V)) to give 0.153g of a white powder as a solid in 37.23% yield. m.p.182.3~185.8°C. ¹H-NMR(CDCl₃,300MHz) δ :7.60 (s,1H,NCHN) , 5.24~5.19(m,1H,H-12),2.62(t, *J*=7.5Hz,1H,H-18),4.04(t,2H, *J*=8.5Hz, COOC<u>H</u>₂(CH₂)₃CH₃), 1.60~1.39(m,6H, COOCH₂(C<u>H₂</u>)₃CH₃), 0.90(t, *J*=11.5Hz 3H,COOCH₂(CH₂)₃C<u>H</u>₃), 1.27(s,3H),1.27(s,3H),1.19(s,3H), 1.18(s,3H),1.01(s,3H), 0.99(s,3H),0.99(s,3H),ESI-MS(*m*/*z*):548.4[M+H]⁺. Elemental anal.(%) calcd. For C₃₆H₅₆N₂O₂: C 78.78,H 10.28,N 5.10,O 5.83;found: C 78.75,H 10.31,N 5.12,O 5.81. *4.2.22 olean-2-ene-[2,3-b] imidazole-12-ene-28-oic acid nhextyl ester(III₅)*

According to the same method for compound II₁, compound II₄ was prepared from OA-1 (1.1mmol) and brominated nhexane (4.4 mmol), The crude product was purified by silica gel column chromatography (eluent:petroleum ether/ethyl acetate=20/1 (V/V)) to give 0.273g of a white powder as a solid in 51.26% yield. m.p.189.2~193.4°C. ¹H-NMR(CDCl₃,300MHz) δ :7.64 (s,1H,NCHN) , 5.23~5.20(m,1H,H-12),2.66(t, *J*=6.5Hz,1H,H-18),4.06(t, *J*=7.5Hz

2H,COOC<u>H</u>₂(CH₂)₄CH₃), 1.60~1.37(m,8H, COOCH₂(C<u>H</u>₂)₄CH₃), 0.83(t, J=10.7Hz ,3H,COOCH₂(CH₂)₄C<u>H</u>₃), 1.32(s,3H),1.25(s,3H), 1.13(s,3H), 1.12(s,3H),1.02(s,3H),0.94(s,3H),0.83(s,3H),ESI-MS(*m*/*z*):562.4[M+H]⁺. Elemental anal.(%) calcd. For C₃₇H₅₈N₂O₂: C 78.95,H 10.39,N 4.98,O 5.68;found: C 78.97,H 10.37,N 5.01,O 5.65.

4.2.23 5'-methyl-olean-2-ene- [2,3-b] pyrazine-12-ene-28-acylanilide (IV_1)

The intermediate OA-7(0.500g) was dissolved in 20 mL of absolute ethanol and 0.5 g of anhydrous magnesium sulfate was added and the supersaturated 1,2propanediamine-ethanol solution (0.04 mL) was slowly added to the system, the mixture was stirred at 79 °C for 8 h. The solvent was removed by evaporation under reduced pressure. The crude product was purified by silica gel column chromatography (eluent: petroleum ether/ethyl acetate =15/1(V/V)) to give 0.235g of a white powder as a yield of 75.32%. m.p.193.1.3~196.4°C. ¹H-NMR (CDCl₃, 300MHz) δ : 1.62~1.49 (m,1H,NCHCH₃CH₂N) ,1.07 (d, *J*=6.5Hz,3H,NCHCH₃CH₂N) , 1.43~1.25 (m,2H,NCHCH₃CH₂N) 5.20 (s, 1H, H-12), 2.22 (t, *J*=7.2Hz,1H, H-18), 7.26~7.09 (m, 1H, CONHC₆H₅), 7.21~7.63 (m, 5H, CONHC₆H₅), 1.25 (s, 3H), 1.18 (s, 3H), 1.02 (s, 3H), 0.95 (s, 3H), 0.96 (s, 3H), 0.87 (s, 3H), 0.73 (s, 3H), ESI-MS(*m*/z):581.4[M+H]⁺. Elemental anal.(%) calcd. For C₃₉H₅₅N₃O: C 80.50,H 9.53,N 7.22,O 2.75;found: C 80.52,H 9.51,N 7.21,O 2.76. *4.2.24 5'-methyl-olean-2-ene- [2,3-b] pyrazine-12-ene-28-acylchloroaniline(IV*₂)

According to the same method for compound IV₁, compound IV₄ was prepared from OA-1 (1.1mmol) and m-chloroaniline (2.2 mmol), The crude product was purified by silica gel column chromatography (eluent:petroleum ether/ethyl acetate=13/1 (V/V)) to give 0.235g of a white powder as a solid in 75.32% yield. m.p.193.1~196.4°C.

¹H-NMR (CDCl₃, 300MHz) δ : 1.62~1.52 (m,1H,NC<u>H</u>CH₃CH₂N) ,1.07 (d, *J*=6.2Hz,3H,NCHC<u>H</u>₃CH₂N) , 1.43~1.25 (m,2H,NCHCH₃C<u>H</u>₂N) 5.20 (s, 1H, H-12), 2.22 (t, *J*=7.5Hz,1H, H-18), 7.26 (m, 1H, CONHC₆H₅), 7.63~7.21 (m, 5H, CONHC₆H₅), 1.25 (s, 3H), 1.18 (s, 3H), 1.02 (s, 3H), 0.95 (s, 3H), 0.96 (s, 3H), 0.87 (s, 3H), 0.73 (s, 3H), ESI-MS(*m*/*z*):581.4[M+H]⁺. Elemental anal.(%) calcd. For C₃₉H₅₄ClN₃O: C 76.00,H 8.83, Cl 5.75,N 6.82,O 2.60;found: C 76.04,H 8.80, Cl 5.77,N 6.80,O 2.59.

4.2.25 5'-methyl-olean-2-ene- [2,3-b] pyrazine-12-ene-28-acylfluaniline(IV₃)

According to the same method for compound IV₁, compound IV₄ was prepared from OA-1 (1.1mmol) and m-fluoroaniline(2.2 mmol), The crude product was purified by silica gel column chromatography (eluent:petroleum ether/ethyl acetate=13/1 (V/V)) to give 0.235g of a white powder as a solid in 58.32% yield. m.p.213.5~215.6°C.

¹H-NMR (CDCl₃, 300MHz) δ : 1.63~1.52 (m,1H,NC<u>H</u>CH₃CH₂N) ,1.10 (d, *J*=6.5Hz,3H,NCHC<u>H</u>₃CH₂N) ,1.39~1.25 (m,2H,NCHCH₃C<u>H</u>₂N) 5.27(s, 1H, H-12), 2.26(t, *J*=7.3Hz,1H, H-18), 7.25 (s, 1H, CONHC₆H₄F), 7.98~7.23 (m, 4H, CONHC₆H₄F), 1.25(s, 3H), 1.17 (s, 3H), 1.02 (s, 3H), 0.97 (s, 3H), 0.96(s, 3H), 0.81 (s, 3H), 0.75(s, 3H), ESI-MS(*m*/*z*):599.4[M+H]⁺. Elemental anal.(%) calcd. For C₃₉H₅₄FN₃O: C 78.09,H 9.07, F 3.17,N 7.00,O 2.67;found: C 78.06,H 9.10, F 3.18,N

7.02,0 2.64.

4.2.26 5'-methyl-olean-2-ene- [2,3-b] pyrazine-12-ene-28-acofluoroaniline(IV4)

According to the same method for compound IV_1 , compound IV_4 was prepared from OA-1 (1.1mmol) and m-fluoroaniline(2.2 mmol), The crude product was purified by silica gel column chromatography (eluent:petroleum ether/ethyl acetate=15/1 (V/V)) to give 0.523g of a white powder as a solid in 62.54% yield. m.p.208.9~211.6°C.

¹H-NMR (CDCl₃, 300MHz) δ: 1.67~1.44 (m,1H,NC<u>H</u>CH₃CH₂N) ,1.08 (d, J=6.5Hz,3H,NCHCH₃CH₂N), 1.43~1.30 (m,2H,NCHCH₃CH₂N) 5.22 (s, 1H, H-12), 2.25(t, J=7.5Hz,1H, H-18), 7.25 (s, 1H, CONHC₆H₄OCH₃), 7.22~7.07 (m, 4H, CONHC₆H₄ OCH₃), 4.05(s, 3H, CONHC₆H₄OCH₃), 1.23(s, 3H), 1.18 (s, 3H), 1.09(s, 3H), 0.95 (s, 3H), 0.94(s, 3H), 0.83(s, 3H), 0.78(s, 3H), ESI-MS(*m*/*z*):611.4[M+H]⁺. Elemental anal.(%) calcd. For C₄₀H₅₇N₃O₂: C 78.51,H 9.39,N 6.87,O 5.23;found: C 78.48,H 9.42,N 6.88,O 5.22.

4.2.27 5'-methyl-olean-2-ene- [2,3-b] pyrazine-12-ene-28-morphomorpholine (IV₅)

According to the same method for compound W_1 , compound W_4 was prepared from OA-1 (1.1mmol) and methoxy aniline (2.2 mmol), The crude product was purified by silica gel column chromatography (eluent:petroleum ether/ethyl acetate=15/1 (V/V)) to give 0.402g of a white powder as a solid in 55.34% yield. m.p.221.6~225.3°C.

¹H-NMR (CDCl₃, 300MHz) δ: 1.67~1.51 (m,1H,NCHCH₃CH₂N) ,1.08 (d, J=6.5Hz,3H,NCHCH₃CH₂N), 1.43~1.30 (m,2H,NCHCH₃CH₂N) 5.22 (s, 1H, H-12), 2.40(t, J=7.0Hz,1H, H-18), 3.38~3.30 (m, 4H, N(CH₂)₂(CH₂)₂O), 3.62-3.52 (m, 4H, N(CH₂)₂(CH₂)₂O), 1.44(s, 3H), 1.26(s, 3H), 1.21(s, 3H), 0.95 (s, 3H), 0.94(s, 3H), 0.85(s, 3H), 0.79(s, 3H), ESI-MS(*m/z*):575.4[M+H]⁺. Elemental anal.(%) calcd. For C37H57N3O2: C 77.17,H 9.98,N 7.30,O 5.56; found: C 77.20,H 9.95,N 7.27,O 5.59.

4.3. Cell Proliferative Assay

The antiproliferative activities of the title compounds were evaluated in vitro using the MTT method against SKOV3 and BGC-823 cell lines, with 5-FU and Adriamycin as the positive control. The negative control contains cells, culture medium, MTT and DMSO. The two tumor cell lines were cultured in RPMI 1640 medium supplemented with 10% fetal bovine serum (FBS). Tumor cells were detached by trypsinisation, seeded at $1.0 \sim 2.0 \times 10^3$ cells each well in 96-well culture plates and incubated in 5% CO₂ at 37 °C. After 24h, the test compounds was added at different indicated concentration of 10⁻⁵, 10⁻⁶, 10⁻⁷, and 10⁻⁸ mol/L for another 48 h incubation. Then MTT solution(100 µL per well) was added and incubated at 37 °C for 4 h. The MTT-formazan formed by metabolically viable cells was dissolved in 150 µL DMSO each well, and monitored by a microplate reader at dual-wavelength of 490 nm, IC₅₀ was defined as the drug concentrations that inhibited the cell number to 50% after 48 h. Each experiment was repeated at least three times and the results averaged.

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