

Review

Oral Mucositis: Melatonin Gel an Effective New Treatment

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Abstract: The current treatment for cervico-facial cancer involves radio and/or chemotherapy. Unfortunately, cancer therapies can lead to local and systemic complications such as mucositis, which is the most common dose-dependent complication in the oral cavity and gastrointestinal tract. Mucositis can cause a considerably reduced quality of life in cancer patients already suffering from physical and psychological exhaustion. However, melatonin, whose role in the treatment of mucositis has recently been investigated, offers an effective alternative therapy in the prevention and/or management of radio and/or chemotherapy-induced mucositis. This review focuses on the pathobiology and management of mucositis in order to improve the quality of cancer patients' lives.

Keywords: mucositis; radiotherapy; chemotherapy; pathophysiology; management; melatonin

Overview of mucositis pathobiology

Mucositis, one of the most severe toxic side effects of cancer therapy, can affect the entire gastrointestinal tract, with the oral cavity being the most common affected site. It presents in virtually all head and neck cancer patients receiving chemo and/or radiotherapy, in 60–85% of those receiving myeloablative therapy for stem-cell transplantation and in 20–40% of patients receiving conventional chemotherapy [1-3]. The use of concomitant chemotherapy and/or targeted agents increases the risk of mucositis.

Oral lesions lead to considerably decreased quality of life in these patients due to solid and liquid food dysphagia, dysarthria and odynophagia, resulting in depression in some patients, who often require percutaneous endoscopic gastrostomy tube insertion [4]. In addition, mucositis lesions may represent a gateway to opportunistic infections, can complicate cancer treatment and extend hospitalization [5-6]. On the other hand, given its dose-limiting toxicity for both chemo and radiotherapy, mucositis can have a direct impact on survival rates [7].

Pathophysiology of mucositis

Recent developments in mucositis research have highlighted multiple factors which contribute to mucosal injury [8]. A five-phase chronological process has been proposed: initiation, primary damage response (upregulation and message generation), signal amplification, ulceration and the healing phase [9]. Mucositis commences when gastrointestinal (GI) mucosa are exposed to cytotoxic agents, resulting in cellular DNA damage and cell death, mainly through the generation of oxidative

stress and reactive oxygen species (ROS) formation. ROS directly induce tissue injury and trigger a cascade of inflammatory pathways [10]. Ortiz et al. have also observed a mitochondrial oxidation response to radiation with subsequent mitochondrial dysfunction [11].

The progression of mucositis is characterized by significant inflammatory mediator up-regulation due to the activation of the NF- κ B pathway (upregulation and message generation phase). This is followed by the signaling and amplification phase, during which, once activated by chemotherapy and ROS, NF- κ B promotes the expression of multiple pro-inflammatory molecules, including inducible nitric oxide synthase (iNOS), cyclooxygenase-2 (COX-2), TNF α , and pro-IL-1 β , and ensures feedback amplification of the NF- κ B-dependent signaling pathway [10]. Furthermore, mitochondrial ROS production promotes inflammation by activating a multi-protein cytoplasmic complex, such as the NLRP3 (NACHT, LRR and PYD domains-containing protein 3) inflammasome. NLRP3 inflammasome assembly leads to caspase-1 activation and subsequent cleavage of pro-cytokines such as pro-IL-1 β , pro-IL-18 and pro-IL-33 into their mature form [11-12] resulting in ulceration (ulceration phase). Therefore, the NF κ B pathway, mitochondrial dysfunction and subsequent NLRP3 inflammasome activation are the three main players involved in the development of oral mucositis, which amplify the whole inflammatory process *via* positive feedback loops, thus prolonging tissue injury and ending in the healing phase.

During initiation phase of mucositis, patients begin to develop erythema followed by focal areas of oral mucosal desquamation [13], which mainly occur at the submucosa and basal epithelium level. Therefore, although no evident damage to mucosal integrity is observed clinically, the tissue biology is altered [10]. The progression of mucositis is then prolonged and severe, mucosal integrity is breached, ulceration begins to form and the patient starts to have a burning sensation. Atrophic changes occur in the GI mucosa, culminating in tissue injury and stem cell death. GI epithelial integrity and function are destroyed and impaired, respectively. A fibrinous exudate, or pseudomembrane, containing bacteria covers the ulcer. Bacterial colonization at the mucosa ulcers further induces inflammation by stimulating infiltration and activation of macrophages, which occurs approximately two weeks after therapy [10]. Cell wall residues originating from colonizing bacteria penetrate the submucosa, where they activate macrophages in the infiltrate [14]. This can lead to prolonged hospitalization, the need for parenteral nutrition, severe pain, risk of infection and sepsis and increased risk of morbidity and mortality.

The final stage of mucositis pathobiology is the healing process. Epithelial cells controlled by signals secreted by the extracellular matrix, which are then downregulated to avoid hyperplasia, migrate, grow and differentiate to form a wound. With the healing process under way, symptoms begin to abate [9], and healing is completed within 4 weeks after the final dose of radiation. Unfortunately, even after full replenishment of the epithelium, the structure of the reconstituted submucosa differs from its pre-radiotherapy state [15].

On the other hand, radiation-induced damage to healthy intestine tissue is a common side-effect caused by out-of-field or scattered radiation [16]. We recently demonstrated that tongue irradiation also induces intestinal damage. Typical macroscopic traces of mucositis were also detected in the small intestines of irradiated animals, including a significant decrease in villus height and morphological alterations associated with substantial intestinal architecture changes [17]. Given the involvement of mitochondrial oxidative stress, bio-energetic impairment and

subsequent NLRP3 inflammasome activation in the development of radiotherapy-induced gut toxicity, oral irradiation of rats also resulted in increased small intestinal damage.

Mucositis management

As there is no effective therapy for mucositis or its associated pain, a large number of studies have been conducted in this field. Strategies for managing oral mucositis include preventative measures and therapeutic approaches (**Figure 1**) [13].

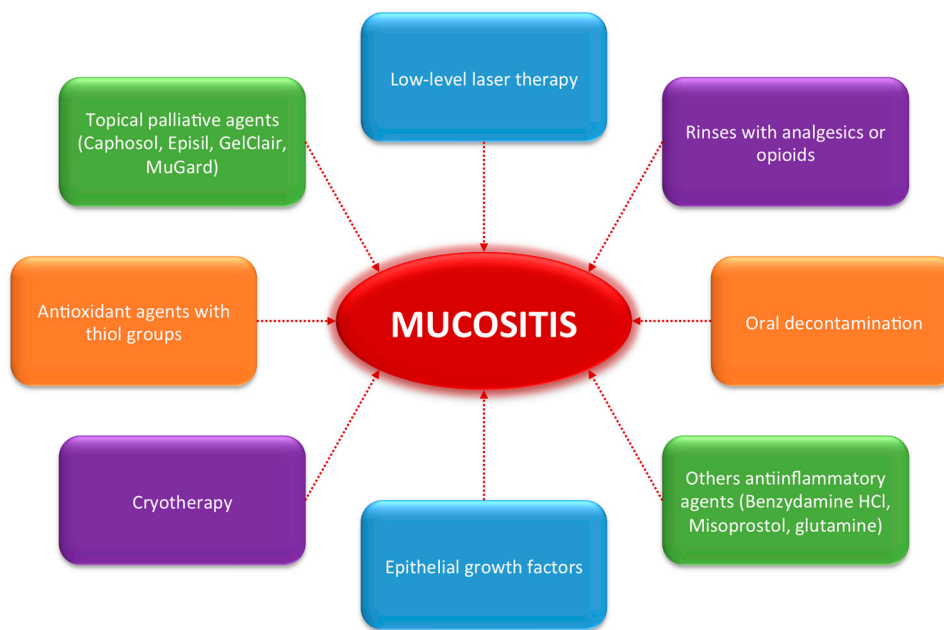


Figure 1: Actual strategies for managing oral mucositis

Current supportive measures to reduce the risk and severity of oral mucositis include improved oral hygiene, which eliminates the presence of any irritants to the oral mucosa [18]. Systemic analgesics are also recommended, as tissue injury activates nociceptive receptors which increase pain alongside the underlying tissue damage [7].

Several topical palliative agents, including Caphosol, Episil, GelClair and MuGard, have been approved for the treatment of mucositis, are aimed at alleviating pain and improving the patient's quality of life. However, data on the efficacy of these agents in controlling mucositis-related symptoms are scarce [13,19-22].

A combination of treatments, such as local rinses with a 2% viscous lidocaine solution, magic mouthwash preparations, a topical morphine solution and other systemic analgesics, are used to control pain [23]. Frequent rinsing with sodium chloride solution helps to keep the mucosa moist, reduces caking of secretions and soothes inflamed/ulcerated mucosa. An oral rinse containing doxepin appears to be effective for easing acute oral mucositis pain caused by radiation therapy (with or without chemotherapy) [24]. However, there is no significant evidence to suggest that

these mouthwashes are effective [25]. Another type of topical agent is the transdermal patch, composed of fentanyl, which is a potent, fast and short acting, synthetic opioid analgesic, although transdermal fentanyl causes mild dizziness, gastrointestinal reactions and itching [26].

Oral decontamination, involving treatments such as Nystatin and chlorhexidine [27], may result in significant positive outcomes in the prevention of ulcerations linked to oral mucositis. Recently, Silva et al. [28] tested a soft pastille formulation consisting of 0.25% lidocaine and 78,000 IU Nystatin, which was found to facilitate accurate drug administration by physicians and to enable patients to control drug retention time in the mouth in order to manage the pain treatment process. On the other hand, chlorhexidine is an effective broad-spectrum antiplaque antiseptic agent [29]. However, these drugs are not very effective in reducing the severity or incidence of mucositis [30-31].

Cryotherapy, during which patients suck on ice chips for 30 minutes prior to and during chemotherapy infusion [32-35], has been shown to effectively attenuate the onset and severity of mucositis in patients undergoing chemotherapy with 5-fluorouracil and melphalan. However, it is only effective for short bolus chemotherapeutic infusions [36], may not be tolerated by some subjects and thus cannot play a significant role in radiation-induced oral mucositis treatments [23].

Several antioxidant agents to prevent mucositis or to reduce its severity have been tested. One of the first drugs used to treat mucositis was amifostine [13], a thiol compound which is dephosphorylated to an active metabolite and acts as a potent ROS scavenger. However, given its limited and inconsistent results, amifostine is not recommended for the prevention of oral mucositis in patients receiving either chemotherapy or radiotherapy alone [37]. N-acetyl cysteine (NAC) is another antioxidant containing thiol groups, which stimulates glutathione synthesis and scavenges free radicals. In addition to its antioxidant properties, NAC prevents NF- κ B activation which increases the inflammatory response. In a double-blind, randomized, placebo-controlled trial, NAC significantly reduced severe oral mucositis incidence [53].

Given that a reduction in the proliferative capacity of oral epithelial cells is thought to play a role in mucositis pathogenesis, various growth factors capable of increasing epithelial cell proliferation have been studied with regard to oral mucositis management. Although palifermin, an epithelial-specific growth factor, is the only agent approved for the prevention of oral mucositis in bone-marrow transplant patients [38], it is also associated with adverse side effects, requires intravenous administration and is expensive [39]. Other growth factors, such as velifermin, filgrastim and argramostim, have been analyzed for use in the treatment of mucositis [40]. Smad7, which has recently received considerable attention [41], was initially identified as a TGF- β superfamily signaling antagonist, which blocks TGF- β -induced growth inhibition and apoptosis in keratinocytes [42] and reduces inflammation by antagonizing NF- κ B activation. All of these characteristics may make Smad7 beneficial in the treatment of oral mucositis [41].

Several anti-inflammatory agents have produced good results in studies of oral mucositis in animals; there is still conflicting evidence, however, on the efficacy of these agents in reducing the severity of mucositis in humans [13]. Benzydamine HCl is a non-steroidal anti-inflammatory drug that inhibits pro-inflammatory cytokines including TNF- α and IL-1 β . It has been administered in an intravenous formulation recommended for the prevention of oral mucositis in patients with head and neck cancer receiving moderate-dose radiation therapy without concomitant chemotherapy

[43]. A long list of anti-inflammatory drugs has produced inconsistent results with regard to the prevention of chemotherapy-induced oral mucositis. This is the case for Misoprostol, a synthetic prostaglandin E1 analog, which has anti-inflammatory mucosa protection properties. However, the overall results of using misoprostol mouthwash in the prevention of radiation-induced oral mucositis in head and neck cancer patients were negative [44-45]. While glutamine, a nonessential amino acid, may mitigate mucosal injury by reducing pro-inflammatory cytokine production and cytokine-related apoptosis [46-47], it produced inconsistent results in the prevention of chemotherapy-induced oral mucositis [48-50].

Multiple studies have indicated that the application of low-level laser therapy (LLLT) reduces the incidence and, by hastening oral re-epithelialization, favorably influences oral mucositis outcomes in patients undergoing standard, hematopoietic stem cell transplantation and myeloablative chemotherapy [51-56]. Although the mechanism involved in these benefits is not understood, it has been suggested that LLLT may reduce ROS and/or pro-inflammatory cytokine levels which contribute to mucositis pathogenesis [23]. However, many of the pathways stimulated by LLLT are associated with undesirable tumor behaviors and/or treatment responses [57].

Melatonin: a new treatment for mucositis

Melatonin (N-acetyl-5-methoxytryptamine), a hormone synthesized from tryptophan, is produced by the pineal gland; it has been detected in multiple extrapineal organ tissues at much higher concentrations than in the pineal gland [58]. It is a potent free radical scavenger with anti-oxidant properties, which increases the expression and activity of endogenous antioxidant enzymes such as superoxide dismutase (SOD), catalase, glutathione peroxidase (GPx), glutathione reductase (GRd) and γ -glutamyl-cystein synthase. This special class of antioxidant generates a series of metabolites that are also free radical scavengers when scavenging free radicals [59-63]. In other words, as compared to other antioxidants, melatonin is more effective in preventing damage caused by oxidative stress. Capable of crossing cell membranes and of easily reaching all cell compartments, it is taken up by mitochondria and can maintain mitochondrial homeostasis in different experimental models [58,64-67]. Melatonin increases membrane fluidity, electron transfer chain (ETC) complex activity, ATP production and mitochondrial membrane potential, while reducing oxidative stress and closing mitochondrial permeability transition pores (MPTPs) [68]. Its important anti-inflammatory effects include expression inhibition of iNOS/i-mtNOS, COX-2 and pro-inflammatory cytokines such as IL-1 β or TNF- α . Many of these properties are attributed to the inhibition of NF- κ B-dependent innate immune pathway activation [69-70], and we recently showed that melatonin blunts NLRP3 inflammasome activation under different experimental conditions [11,71-72].

A) Melatonin in the oral cavity

We recently demonstrated that melatonin is synthesized in oral mucosa, where it is involved in autocrine/paracrine signaling by binding to MT1, MT2, and ROR γ receptors, thus suggesting that it plays a role in normal oral mucosal physiology [11]. We found that aralkylamine N-acetyltransferase (AANAT) and acetylserotonin O-methyltransferase (ASMT), the two main enzymes in melatonin synthesis, are expressed in oral mucosa and also in salivary glands. However, melatonin, which is also found in saliva, is believed to passively enter the mucous through the circulatory system in

salivary glands (parotid, submaxillary and sublingual glands) [58,73]. Thus, as the quantity of melatonin entering the oral cavity is proportional to salivary flow, xerostomia, which actually aggravates periodontal status, could be associated with oral pathologies [74]. Together, these data, which are corroborated by the literature, demonstrate that melatonin has important implications for the prevention of oral cavity diseases by limiting tissue damage through free radicals and by stimulating immune responses. Almughrabi et al. demonstrated the relationship between low levels of melatonin in saliva and the increased severity of oral pathologies such as gingivitis, chronic periodontitis and aggressive periodontitis [75]. The beneficial effects of topical melatonin applications in patients with periodontal diseases, as evidenced by improvements in gingival index and pocket depth, have been established by Gomez-Moreno et al. [76]. Melatonin has also proven to be an effective treatment for oral infections such as herpes [74]. Kara et al. showed that it decreases proinflammatory cytokines in gingivitis and periodontitis [77]. Cutando et al. [78] showed that melatonin treatment in patients with diabetes significantly reduced the gingival index, pocket depth and RANKL levels and also increased osteoprotegrin concentrations. Due to its anti-inflammatory and anti-oxidative effects, the severity of gingival and periodontal inflammation was reduced. Other studies have demonstrated that melatonin could be a beneficial therapy after surgical procedures in the oral cavity by preventing inflammatory and infectious complications induced by oxidative stress [79]. Cutando et al. [79] showed that topically applied melatonin in the evacuated sockets following tooth removal reduced oxidative stress and inflammation and accelerated the healing process. Thus, by being directly deposited in the oral cavity, it has the capacity to treat oral disorders and pathologies by reducing inflammatory responses in the gingiva and periodontium [74].

B) Melatonin as a radio-protective agent

For many years, the radio-protective effects of melatonin have been observed in different experimental models as well as organs and tissues. Tan et al. [80] were the first to report its ability to protect against electromagnetic radiation emitted by ultraviolet light, with subsequent studies reporting its effectiveness in protecting against ionizing radiation. Vijayalaxmi et al., who carried out a series of experiments to study the radio-protective effects of melatonin *in vitro* and *in vivo*, showed that melatonin guarded against gamma radiation-induced cell damage in blood lymphocytes [81-84]. Other authors have reported that it prevents hemolysis in irradiated human red blood cells [85]. Pretreatment with melatonin guards erythrocytes, granulocytes, macrophages, megakaryotes and T cells against radiation-induced cellular injury and also inhibits splenocyte apoptosis in whole-body irradiated mice [86]. It also prevents radiation-induced damage in retinal cells [87], thymocytes [88] and bone-marrow cells in mice [89]. Several *in vivo* studies reveal that it increases the survival rate of animals exposed to radiation. Pre-treatment with 250 mg/kg melatonin raised the survival rate of lethally whole-body irradiated (9.5 Gy) mice [90] to approximately 43% and to 85% at a dose of 8.15 Gy [91]. Iwata et al. demonstrated that pre-treatment with intraperitoneally administered melatonin at a dosage of 150 mg/kg showed a survival rate of around 100% for mice following a radiation dose of 7.5 Gy [92], with an amelioration in radiation-induced injury in radio-sensitive organs such as bone marrow, spleen and gastrointestinal [93]. In all these experiments, melatonin was observed to protect against radiation-induced genotoxicity in both the somatic and germ cells of mice. Its radio-protective impact was also demonstrated in the testis and

ovary of irradiated rodents [94-97]. Melatonin also plays an important protective role against radiation-induced damage to the thyroid gland [98] and the small intestine [99], which is one of the most radio-sensitive organs, resulting in inflammation-induced radiation enteritis. Its administration prior to irradiation guards against intestinal damage caused by X-rays [100]. It prevents mucosal intestinal damage caused by radiotherapy by countering structural changes in the small intestine (as evidenced by the villous pattern of the intestinal mucosa) [101] and also inhibits gut bacterial translocation to the spleen, liver and kidney [86].

The mechanisms involved in melatonin as a radioprotective agent have been attributed to its antioxidant properties, which reduce radiation-induced DNA damage and lipid peroxidation, and to its protection of the immune system by reducing apoptosis through the inhibition of p53 and Bax and by enhancing anti-apoptotic protein Bcl-2 [86,93,102]; it is also involved in repairing lesions in cellular DNA [103].

In patients, administration of melatonin resulted in amelioration of hypotension, myelotoxicity, and lymphocytopenia associated with radiotherapy [104]. Therefore, its use as a prophylactic agent could reduce morbidity and limit radiation-induced injury in cancer patients under radiotherapy [86,102].

B) Use of melatonin in the prevention of radiation-induced mucositis

Together, given the data summarized in this review, highlighting its anti-inflammatory and antioxidant function in the oral cavity and its potential effectiveness in protecting against ionizing radiation, melatonin could, in our view, play a beneficial role in the prevention of mucositis. We demonstrated that the application of a melatonin gel to oral mucosa can totally prevent oral mucositis in irradiated rats [11]. The purpose of this study was to investigate the pathophysiology of oral mucositis and how melatonin can prevent its development. We observed that melatonin gel is capable of preventing mucosal disruption and the emergence of ulcers. Furthermore, it can prevent the loss of proliferative progenitor stem cells caused by radiation and enhance their capacity to repopulate the tissue. In addition, ionizing radiation exposure causes oxidative damage to DNA, which arrests the cell cycle, inhibits growth and increases cell death [86]. Thus, protection of cellular DNA is of the utmost importance in reducing radiation-induced cellular perturbation and also in the proliferation/differentiation of normal cells. Melatonin treatment significantly reduces radiation-induced DNA degradation. Therefore, given the high dependence of both epidermis and mucosal epithelia on resident self-renewing stem cells, the therapeutic interventions using melatonin described above, which can reduce the deleterious effects of radiation on normal epithelial stem cells, could have a considerable impact on the quality of life of cancer patients.

On the other hand, it is well known that irradiation inhibits the activity of mitochondrial ETC complexes which enhances electron leakage and subsequent superoxide anion ($O_2^{\bullet -}$) generation, leading to persistent oxidative stress, responsible, at least partially, for radiation-induced cell death in normal human fibroblast cells [105]. Given the induction of mitochondrial dysfunction by ionizing radiation, we assessed whether mitochondrial damage is involved in radiation-induced mucositis. We also determined whether the known improvement by melatonin of mitochondrial function could protect oral mucosa against deleterious radiation effects. In our experiments, we found that the application of melatonin gel decreased the radiation-induced

oxidation responses of mitochondria. We also observed that the protective effect of melatonin was mediated by the increased expression and activity of mitochondrial antioxidant enzymes such as GRd. This reduced the mitochondrial GSSG/GSH ratio and restored mitochondrial GSH homeostasis [64], thus enabling mitochondria to recover from post-radiation oxidative stress. This is in line with the finding that melatonin gel increases ETC protein expression and activity and also expands citrate synthase activity, reflecting the increase in mitochondrial mass, resulting in a full recovery of their bio-energetic capacity and prevention of cell death. These findings suggest that the reduction in mitochondrial damage by melatonin by preserving mitochondrial structure and function may underlie its efficacy in preventing mucositis. The prevention of mitochondrial damage in irradiated oral mucosa may therefore constitute one of the mechanisms by which melatonin protects against cell death. Interestingly, we found that it inhibits mitochondrial-dependent apoptosis by decreasing the Bax/Bcl-2 ratio. It also significantly inhibits p53 and Bax protein expression, while anti-apoptotic Bcl-2 protein expression increases considerably. Consequently, at least part of its radio-protective capacity could depend on the inhibitory action of p53-related signaling proteins, which prevents the opening of MPTPs and thus blocks the release of cytochrome c to the cytosol. In this regard, radiation induces MPTPs to open up and mitochondrial DNA (mtDNA) to translocate to the cytosol. In addition to ROS released from damaged mitochondria to the cytosol, mtDNA promotes the assembly of NLRP3 inflammasome, a multiprotein complex which activates caspase-1 [106]. Once activated, caspase-1 converts NF- κ B-dependent pro-inflammatory cytokines, including pro-IL-1 β , into a mature form. Thus, the activation of both NLRP3 inflammasome and NF- κ B innate immunity pathways leads to overproduction of IL-1 β , TNF- α and other pro-inflammatory mediators [106]. This double-stranded, innate immune response by the NF- κ B and NLRP3 inflammasome pathways could explain the difficulty of finding an effective anti-mucositis therapy. Furthermore, the presence of impaired mitochondria underlying these inflammatory responses makes the search for a specific treatment more difficult. Our results highlight the importance of scavenging ROS and the protection of mitochondria from irradiation in order to suppress inflammasome activation and consequently the development of oral mucositis. This would, in turn, help to explain the therapeutic benefits of melatonin gel in combating oral mucositis (**Figure 2**).

On the other hand, as discussed above, epithelial crypts in the intestinal region contain the highest proliferating cells [107], which are extremely sensitive to ionizing radiation [108]. Recent studies have reported that, even when the intestine is outside the irradiation field, radiation-induced damage to healthy intestine tissue is a common side-effect of out-of-field or scattered radiation. This could explain why typical signs of mucositis were detected in the small intestine following tongue irradiation. Histological analysis revealed crypt loss, lower villi numbers and shorter lengths in the gastrointestinal region, leading to the conclusion that radiation enteropathy prevention requires protection of the small intestine. Melatonin gel treatment in the mouth, which was observed to reduce intestinal morpho-pathological changes [17], is therefore also associated with improved preservation of both small intestinal and oral mucosa.

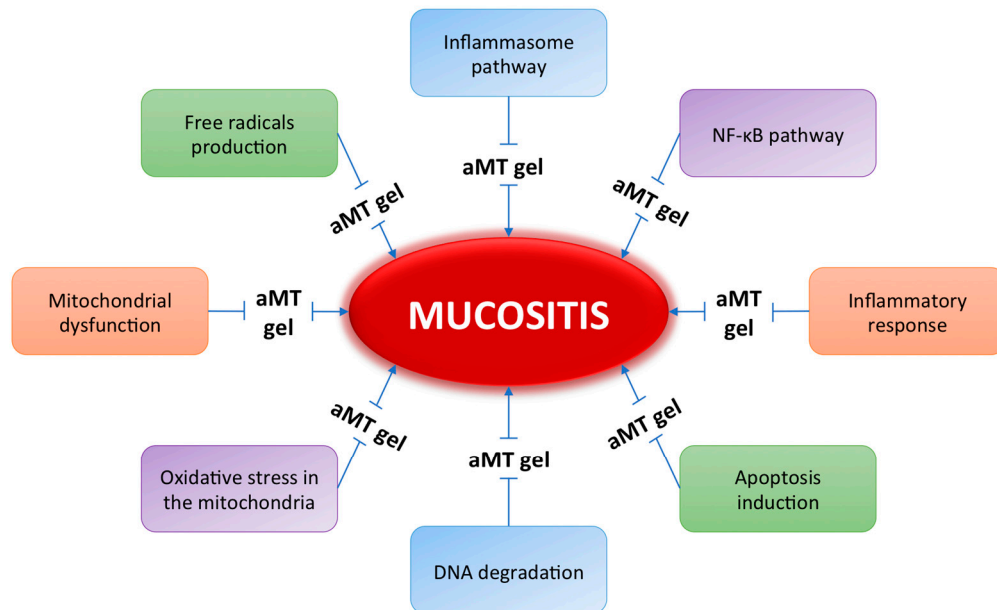


Figure 2: Molecular pathways of melatonin gel to prevent oral mucositis

In addition, a number of studies have shown that melatonin contains remarkable oncostatic properties. Its antiproliferative properties have been demonstrated in an extensive variety of tumors, including breast, endometrial, prostate, colon and ovarian cancers as well as choriocarcinoma, melanoma, neuroblastoma and osteosarcoma among others [109-110]. Its oncostatic mechanisms are associated with several hallmarks of cancer. Its anticancer action directly inhibits the proliferation and growth of tumor cells [111]. Melatonin protects normal cells from apoptosis, while, at the same time, promoting apoptotic cell death in several types of cancer cell [112]. Its immuno-modulatory anticancer action also augments antitumor immune response [113]. Additionally, melatonin has an important metabolic effect, which decreases glucose uptake by cancer cells [114] and inhibits tumor growth through the suppression of the uptake of linoleic acid and its metabolism by the tumor to the mitogenic molecule 13-HODE [115]. Its anti-angiogenic and anti-metastatic properties have also been reported in numerous studies [116-118]. Given its capacity to increase the efficacy of anticancer drugs, these data show that melatonin can be used not only to treat mucositis due to the absence of adverse side effects, but also can be used in cancer co-treatment programs.

Conclusions

In summary, oral mucositis is a clinically important, deleterious consequence of chemo and radiotherapy, for which no effective treatment has been found to date. Mucositis lesions can be painful, affect nutrition and quality of life and have a significant economic impact. The pathogenesis of oral mucositis is multifactorial and complex, and not all mucositis can be prevented. Once

mucositis has developed, therapy should focus on supportive care, which aims to maintain hydration, provide appropriate caloric intake through enteral or parenteral nutritional support, relieve pain and to prevent infection. This review discusses current clinical practices in the management of oral mucositis and emphasizes that no standard therapeutic approach has been developed for patients suffering from oral mucositis. Thus, basic, translational and clinical research into how to prevent and treat oral and gastrointestinal mucositis continues. Our findings, showing that melatonin reduces irradiation toxicity and prevents treatment-induced mucositis, indicate that, with its oncostatic and cytoprotective properties, it constitutes an innovative, adjuvant strategy in the treatment of cancer. We demonstrated that treatment with melatonin gel protects rats from post-radiation oral mucositis, prevents duodenal inflammation and necrosis and restores mucosal endogenous melatonin levels in irradiated animals. A clinical trial of this gel, which is under patent, is currently underway to test for the prevention of oral mucositis in head and neck cancer patients.

Acknowledgments: Part of this study was partially funded by the following grants: SAF2009-14037 from the Spanish Ministry of Economy and Competitiveness (MINECO), CB/10/00238 from the Carlos III Health Institute, GREIB.PT_2010_04 from the CEIBiotic Program of the University of Granada, Spain, and CTS-101 from the Innovacion, Science and Business Council, Junta de Andalucía, Spain. Finally, we wish to thank Michael O'Shea for proofreading the paper.

Conflicts of Interest: The authors declare no conflict of interest.

Abbreviations

| | |
|----------------|--------------------------------------------------|
| ApoE | apolipoprotein E |
| COX-2 | Cyclooxygenase-2 |
| CTCAE | Common Terminology Criteria for Adverse Events |
| EGF | epidermal growth factor |
| G-CSF | granulocyte colony-stimulating factor |
| GIT | gastrointestinal tract |
| GM-CSF | granulocyte-macrophage colony stimulating factor |
| IL-1 | Interleukin-1 |
| MAPK | mitogen-activated protein kinase |
| MMP | matrix metalloproteinase |
| mTOR | mammalian target of rapamycin |
| NAC | N-acetyl cysteine |
| NCI | National Cancer Institute |
| NF- κ B | nuclear factor kappa-B |
| NLRP3 | NACHT, LRR and PYD domains-containing protein 3 |
| NSAID | non-steroidal anti-inflammatory drug |
| PARs | protease-activated receptors |
| PGE | Prostaglandin E |
| PLAG | 1-palmitoyl-2-linoleoyl-3-acetyl-rac-glycerol |
| rhEGF | recombinant human EGF |
| ROS | reactive oxygen species |
| SAP | serum amyloid-P |
| Smad7 | mothers against decapentaplegic homolog 7 |
| TCM | traditional Chinese medicine |
| TGF- β | transforming growth factor beta |
| TNF- α | tumor necrosis factor- α |

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