

Article

Not peer-reviewed version

Streptococcus pneumoniae and Influenza (H1N1) virus Genome Study Against Human Genome Sequences – Blastn and DAVID Analysis

[Mark Christopher Arokiaraj](#)^{*}, Jarad Wilson, Jingqiao Lu, [RuoPan Huang](#)

Posted Date: 2 August 2023

doi: 10.20944/preprints202308.0171.v1

Keywords: *Streptococcus pneumoniae*; Influenza (H1N1) virus; computational biology; Blastn; DAVID; NDEx



Preprints.org is a free multidiscipline platform providing preprint service that is dedicated to making early versions of research outputs permanently available and citable. Preprints posted at Preprints.org appear in Web of Science, Crossref, Google Scholar, Scilit, Europe PMC.

Copyright: This is an open access article distributed under the Creative Commons Attribution License which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Article

Streptococcus Pneumoniae and Influenza (H1N1) Virus Genome Study against Human Genome Sequences – Blastn and DAVID Analysis

Mark Christopher Arokiaraj ^{1*}, Jarad Wilson ², Jingqiao Lu ² and RuoPan Huang ²

¹ Cardiology consultant and researcher, Pondicherry, India 605001

² RayBiotech, Peachtree corners, Georgia, USA

* Correspondence: christomark@gmail.com

Abstract: **Aim:** Streptococcus pneumoniae and influenza H1N1 virus are common organisms associated with human infections. These infections could play a significant role in immune regulation. The study was performed to analyse the genome sequences of these organisms with human genome and study its functional significance. **Methods:** The study was performed to analyse the overlapping of genome sequences in Streptococcus pneumoniae and Influenza (H1N1) virus against human genome sequences by BLASTn sequence analysis. The alignments are studied against the corresponding genes for their functional significance with DAVID and NDEx software. **Results:** Several hits or overlapping nucleotide segments were identified. Between streptococcus and homo sapiens 287 overlaps were identified, and among influenza and homo sapiens 124 hits were identified. A wide range of functional significance of these genes were identified, and the results are presented in this study. The results show insights into functional pathways and biological activities associated with the respective vaccinations or infections by these microorganisms. **Conclusion:** The common organisms like Streptococcus pneumoniae and Influenza H1N1 virus actively interact with the immune system and result in a wide range of immune regulations.

Keywords: Streptococcus pneumoniae; influenza (H1N1) virus; computational biology; Blastn; DAVID; NDEx

Introduction

Streptococcus pneumoniae and influenza (H1N1) are common vaccinations given to the elderly general population. Infections by these organisms are associated with mortality and morbidity worldwide, and the regional incidence varies [1]. Vaccinations against these organisms are associated with cardiovascular benefits and also a general reduction in mortality [2–4]. Improvement in trained immunity has been postulated for non-specific protection of these vaccinations against Covid19 [5]. The exact mechanisms of this protection. The study was performed to analyze the genome sequence overlap between these microorganisms and the human genome and to evaluate the possible modifications and influences of these vaccinations on human gene expression.

Methods

Blastn analysis [6,7] was performed by comparing the Streptococcus pneumoniae strain Hu17 and Haemophilus influenza (H1N1) 2009 California genome sequences against the human genome (Homo sapiens Hg38). The genome sequence of Influenza H1N1 2009 California (2009 California) was downloaded from NCBI (<https://www.ncbi.nlm.nih.gov/data-hub/taxonomy/641809/>). The Influenza H1N1 2009 California genome sequences were compared against Homo sapiens hg38 downloaded from NCBI (https://www.ncbi.nlm.nih.gov/assembly/GCF_000001405.26/) with blastn tools (2.13.0+) on a Linux server, respectively. The genome sequence of Streptococcus Pneumonia strain Hu17 (2009 California) was downloaded from NCBI (<https://www.ncbi.nlm.nih.gov/data-hub/taxonomy/641809/>). The Streptococcus pneumoniae strain Hu17 genome sequences were compared against Homo sapiens hg38 downloaded from NCBI (https://www.ncbi.nlm.nih.gov/assembly/GCF_000001405.26/) with blastn tools (2.13.0+) on a Linux

server, respectively. The hits/alignments on the genome of Homo sapiens hg38 reported by the blast analysis were retrieved with customized Python scripts. The regions of hits were annotated with gene information, respectively.

The corresponding genes were studied for functional annotations to study background processes, which could influence physiological changes. The blast analysis results were further processed for functional analysis of the concerned genes through the database for annotation, visualization, and integrated discovery (DAVID) [8] to understand the biological implications of the outcomes.

Results

Based on the Blastn analysis, 124 hits or overlapping sequences were observed in the analysis of H1N1 vs. Homosapiens. 287 hits were observed between the overlapping segments of Streptococcus pneumoniae and Homo sapiens. The results are published in the data repository (supplement data 1 and 2). DAVID analysis showed various gene enrichment related to functions with statistical significance. The results are published in the supplement data repository (supplement data 3). The enrichment values and the level of significance of the genes are also shown in the results. A wide range of enrichment and functional annotations are observed, which give insights into the background processes after the vaccinations or infections with these organisms. The functional annotation chart with biogrid interactions (Figure 1), genes and pathway map (Figure 2), protein domain (Figure 3) and tissue characterisation (Figure 4) derived from DAVID analysis between influenza H1N1 and Homo sapiens indicate the various functional associations with associated genes. Similarly the results of DAVID analysis with Streptococcus pneumoniae and Homo sapiens show the associated protein domain (Figure 5), pathways (Figure 6), genes associated with tissue expression (Figure 7) and biogrid interactions (Supplement Figure S1).

Numerically streptococcus pneumoniae has more functional interactions than influenza H1N1 virus though the extent of biological outcomes is difficult to quantify. NDEx analysis of the overlapping genes showed the pathways associated with the genes and gene ontology analysis (Figures 8 and 9) showed the relevant associated genes (Supplement Figures S2 and S3). Heat shock related pathways and signalling was predominantly involved with S. pneumoniae-homo sapiens overlapping genes (Figure 9). Genes related to histone modifications, myc pathways, DNA damage check point signalling, FGF/IGF, notch signalling were associated with influenza – Homo sapiens overlap related genes (Supplement Figure S2). PRKN, SNCA, STUB1, GPR37, SEPT, TUBA, TUBB genes related pathways concerning proteasomal degradation were associated with S. pneumonia-Homo sapiens overlapping genes (Supplement Figure S3). Some of the other seen by pathway figures similarity were KRAS, EGFR and by INDRO GO similarity ELAVL1 and MAPKAPK2 were some of the pathways identified (Supplement Figure S4) when analysed between influenza H1N1 Vs Homosapiens. Genes like HSF1, GCase and HSPA9 were some of the pathways identified when analysed between Streptococcus pneumoniae Vs Homosapiens (Supplement Figure S5).

Sublist	Category	Term	RT	Genes	Count	%	P-Value	Benjamini
<input type="checkbox"/>	BIOGRID_INTERACTION	vir like m6A methyltransferase associated(VIRMA)	RT	<div></div>	33	26.8	9.7E-6	3.7E-2
<input type="checkbox"/>	BIOGRID_INTERACTION	formyl peptide receptor 1(FPR1)	RT	<div></div>	5	4.1	5.9E-3	1.0E0
<input type="checkbox"/>	BIOGRID_INTERACTION	cystatin 5(CST4)	RT	<div></div>	4	3.3	9.9E-3	1.0E0
<input type="checkbox"/>	BIOGRID_INTERACTION	amylase alpha 1C(AMY1C)	RT	<div></div>	4	3.3	9.9E-3	1.0E0
<input type="checkbox"/>	BIOGRID_INTERACTION	sodium voltage-gated channel beta subunit 2(SCN2B)	RT	<div></div>	4	3.3	1.0E-2	1.0E0
<input type="checkbox"/>	BIOGRID_INTERACTION	protein phosphatase 3 regulatory subunit B, beta(PPP3R2)	RT	<div></div>	3	2.4	1.1E-2	1.0E0
<input type="checkbox"/>	BIOGRID_INTERACTION	KH RNA binding domain containing, signal transduction associated 1(KHDRBS1)	RT	<div></div>	6	4.9	1.1E-2	1.0E0
<input type="checkbox"/>	BIOGRID_INTERACTION	Fas ligand(FASLG)	RT	<div></div>	4	3.3	1.1E-2	1.0E0
<input type="checkbox"/>	BIOGRID_INTERACTION	capicua transcriptional repressor(CIC)	RT	<div></div>	9	7.3	1.3E-2	1.0E0
<input type="checkbox"/>	BIOGRID_INTERACTION	trafficking protein particle complex subunit 2B(TRAPPC2B)	RT	<div></div>	3	2.4	1.3E-2	1.0E0
<input type="checkbox"/>	BIOGRID_INTERACTION	G protein-coupled receptor 17(GPR17)	RT	<div></div>	6	4.9	1.4E-2	1.0E0
<input type="checkbox"/>	BIOGRID_INTERACTION	solute carrier family 18 member A2(SLC18A2)	RT	<div></div>	4	3.3	1.5E-2	1.0E0
<input type="checkbox"/>	BIOGRID_INTERACTION	heterogeneous nuclear ribonucleoprotein L(HNRNPL)	RT	<div></div>	17	13.8	1.9E-2	1.0E0
<input type="checkbox"/>	BIOGRID_INTERACTION	hook microtubule tethering protein 1(HOOK1)	RT	<div></div>	5	4.1	2.3E-2	1.0E0
<input type="checkbox"/>	BIOGRID_INTERACTION	Rac family small GTPase 1(RAC1)	RT	<div></div>	6	4.9	2.5E-2	1.0E0
<input type="checkbox"/>	BIOGRID_INTERACTION	SLIT-ROBO Rho GTPase activating protein 2(SRGAP2)	RT	<div></div>	4	3.3	2.5E-2	1.0E0
<input type="checkbox"/>	BIOGRID_INTERACTION	dynein cytoplasmic 1 light intermediate chain 1(Dync1li1)	RT	<div></div>	3	2.4	2.6E-2	1.0E0
<input type="checkbox"/>	BIOGRID_INTERACTION	par-3 family cell polarity regulator(PARD3)	RT	<div></div>	5	4.1	3.1E-2	1.0E0
<input type="checkbox"/>	BIOGRID_INTERACTION	trafficking protein particle complex subunit 8(TRAPPC8)	RT	<div></div>	3	2.4	3.2E-2	1.0E0
<input type="checkbox"/>	BIOGRID_INTERACTION	proteasome activator subunit 4(PSMF4)	RT	<div></div>	3	2.4	3.3E-2	1.0E0
<input type="checkbox"/>	BIOGRID_INTERACTION	ubiquitin specific peptidase 7(USP7)	RT	<div></div>	7	5.7	3.5E-2	1.0E0
<input type="checkbox"/>	BIOGRID_INTERACTION	protein inhibitor of activated STAT 1(PIAS1)	RT	<div></div>	4	3.3	3.5E-2	1.0E0
<input type="checkbox"/>	BIOGRID_INTERACTION	DDB1 and CUL4 associated factor 15(DCAF15)	RT	<div></div>	5	4.1	3.9E-2	1.0E0
<input type="checkbox"/>	BIOGRID_INTERACTION	lymphocyte transmembrane adaptor 1(LAX1)	RT	<div></div>	2	1.6	4.0E-2	1.0E0
<input type="checkbox"/>	BIOGRID_INTERACTION	ankyrin repeat domain 28(Ankrd28)	RT	<div></div>	2	1.6	4.0E-2	1.0E0
<input type="checkbox"/>	BIOGRID_INTERACTION	gap junction protein alpha 1(GJA1)	RT	<div></div>	7	5.7	4.0E-2	1.0E0
<input type="checkbox"/>	BIOGRID_INTERACTION	butyrophilin like 9(BTNL9)	RT	<div></div>	4	3.3	4.2E-2	1.0E0
<input type="checkbox"/>	BIOGRID_INTERACTION	RAB4A, member RAS oncogene family(RAB4A)	RT	<div></div>	6	4.9	4.3E-2	1.0E0
<input type="checkbox"/>	BIOGRID_INTERACTION	protein kinase cAMP-activated catalytic subunit alpha(PRKACA)	RT	<div></div>	5	4.1	4.3E-2	1.0E0
<input type="checkbox"/>	BIOGRID_INTERACTION	RAB2B, member RAS oncogene family(RAB2B)	RT	<div></div>	3	2.4	4.3E-2	1.0E0
<input type="checkbox"/>	BIOGRID_INTERACTION	RAB11A, member RAS oncogene family(RAB11A)	RT	<div></div>	8	6.5	4.8E-2	1.0E0
<input type="checkbox"/>	BIOGRID_INTERACTION	late endosomal/lysosomal adaptor, MAPK and MTOR activator 1(LAMTOR1)	RT	<div></div>	8	6.5	5.3E-2	1.0E0
<input type="checkbox"/>	BIOGRID_INTERACTION	peroxisome proliferator activated receptor alpha(PPARA)	RT	<div></div>	3	2.4	5.4E-2	1.0E0
<input type="checkbox"/>	BIOGRID_INTERACTION	WD repeat domain, phosphoinositide interacting 2(Wipi2)	RT	<div></div>	2	1.6	5.4E-2	1.0E0
<input type="checkbox"/>	BIOGRID_INTERACTION	chromosome 17 open reading frame 50(C17orf50)	RT	<div></div>	2	1.6	5.4E-2	1.0E0
<input type="checkbox"/>	BIOGRID_INTERACTION	pleckstrin homology and FYVE domain containing 2(PLEKHF2)	RT	<div></div>	4	3.3	5.8E-2	1.0E0
<input type="checkbox"/>	BIOGRID_INTERACTION	trafficking protein particle complex subunit 3L(TRAPPC3L)	RT	<div></div>	2	1.6	5.9E-2	1.0E0
<input type="checkbox"/>	BIOGRID_INTERACTION	estrogen receptor binding site associated antigen 9(EBAG9)	RT	<div></div>	5	4.1	6.0E-2	1.0E0
<input type="checkbox"/>	BIOGRID_INTERACTION	DDB1 and CUL4 associated factor 6(DCAF6)	RT	<div></div>	3	2.4	6.1E-2	1.0E0
<input type="checkbox"/>	BIOGRID_INTERACTION	protein phosphatase 1 catalytic subunit alpha(PPP1CA)	RT	<div></div>	3	2.4	6.2E-2	1.0E0
<input type="checkbox"/>	BIOGRID_INTERACTION	TNF receptor associated factor 1(TRAF1)	RT	<div></div>	5	4.1	6.3E-2	1.0E0
<input type="checkbox"/>	BIOGRID_INTERACTION	lysosomal associated membrane protein 2(LAMP2)	RT	<div></div>	7	5.7	6.5E-2	1.0E0
<input type="checkbox"/>	BIOGRID_INTERACTION	G protein-coupled receptor 45(GPR45)	RT	<div></div>	5	4.1	6.6E-2	1.0E0
<input type="checkbox"/>	BIOGRID_INTERACTION	kinesin family member 14(KIF14)	RT	<div></div>	15	12.2	6.6E-2	1.0E0
<input type="checkbox"/>	BIOGRID_INTERACTION	RAB9A, member RAS oncogene family(RAB9A)	RT	<div></div>	7	5.7	6.7E-2	1.0E0
<input type="checkbox"/>	BIOGRID_INTERACTION	LSM7 homolog, U6 small nuclear RNA and mRNA degradation associated(LSM7)	RT	<div></div>	3	2.4	6.9E-2	1.0E0
<input type="checkbox"/>	BIOGRID_INTERACTION	Cbl proto-oncogene B(CBLB)	RT	<div></div>	3	2.4	7.2E-2	1.0E0
<input type="checkbox"/>	BIOGRID_INTERACTION	chymotrypsin like elastase 2A(CELA2A)	RT	<div></div>	2	1.6	7.3E-2	1.0E0
<input type="checkbox"/>	BIOGRID_INTERACTION	thrombospondin 1(THBS1)	RT	<div></div>	3	2.4	7.8E-2	1.0E0
<input type="checkbox"/>	BIOGRID_INTERACTION	secreted phosphoprotein 1(SPP1)	RT	<div></div>	3	2.4	8.0E-2	1.0E0
<input type="checkbox"/>	BIOGRID_INTERACTION	lysine demethylase 6B(KDM6B)	RT	<div></div>	3	2.4	8.1E-2	1.0E0
<input type="checkbox"/>	BIOGRID_INTERACTION	ST14 transmembrane serine protease matriptase(ST14)	RT	<div></div>	4	3.3	8.2E-2	1.0E0
<input type="checkbox"/>	BIOGRID_INTERACTION	tripartite motif containing 66(TRIM66)	RT	<div></div>	4	3.3	8.2E-2	1.0E0
<input type="checkbox"/>	BIOGRID_INTERACTION	CD2 molecule(CD2)	RT	<div></div>	2	1.6	8.3E-2	1.0E0
<input type="checkbox"/>	BIOGRID_INTERACTION	WW domain containing oxidoreductase(WWOX)	RT	<div></div>	7	5.7	8.5E-2	1.0E0
<input type="checkbox"/>	BIOGRID_INTERACTION	ezrin(EZR)	RT	<div></div>	6	4.9	8.5E-2	1.0E0
<input type="checkbox"/>	BIOGRID_INTERACTION	GABA type A receptor-associated protein(GABARAP)	RT	<div></div>	3	2.4	8.7E-2	1.0E0
<input type="checkbox"/>	BIOGRID_INTERACTION	chemerin chemokine-like receptor 1(CMKLR1)	RT	<div></div>	3	2.4	8.9E-2	1.0E0
<input type="checkbox"/>	BIOGRID_INTERACTION	syntaxin 7(STX7)	RT	<div></div>	7	5.7	9.0E-2	1.0E0
<input type="checkbox"/>	BIOGRID_INTERACTION	calcium voltage-gated channel subunit alpha1 A(CACNA1A)	RT	<div></div>	3	2.4	9.0E-2	1.0E0
<input type="checkbox"/>	BIOGRID_INTERACTION	FHF complex subunit HOOK interacting protein 1B(FHIP1B)	RT	<div></div>	2	1.6	9.2E-2	1.0E0
<input type="checkbox"/>	BIOGRID_INTERACTION	cell division cycle associated 5(Cdca5)	RT	<div></div>	2	1.6	9.2E-2	1.0E0
<input type="checkbox"/>	BIOGRID_INTERACTION	cadherin 3(CDH3)	RT	<div></div>	2	1.6	9.2E-2	1.0E0
<input type="checkbox"/>	BIOGRID_INTERACTION	tripartite motif containing 21(TRIM21)	RT	<div></div>	4	3.3	9.4E-2	1.0E0
<input type="checkbox"/>	BIOGRID_INTERACTION	nucleoporin 214(NUP214)	RT	<div></div>	3	2.4	9.6E-2	1.0E0
<input type="checkbox"/>	BIOGRID_INTERACTION	SEC62 homolog, preprotein translocation factor(SEC62)	RT	<div></div>	6	4.9	9.7E-2	1.0E0
<input type="checkbox"/>	BIOGRID_INTERACTION	trafficking protein particle complex 8(Trappc8)	RT	<div></div>	2	1.6	9.7E-2	1.0E0
<input type="checkbox"/>	BIOGRID_INTERACTION	ERBB receptor feedback inhibitor 1(ERF1)	RT	<div></div>	3	2.4	9.8E-2	1.0E0

Figure 1. Influenza H1N1 virus Vs. Homosapiens - Biogrid interactions.

Sublist	Category	Term	RT	Genes	Count	%	P-Value	Benjamini
<input type="checkbox"/>	REACTOME_PATHWAY	FCER1 mediated Ca+2 mobilization	RT	<div></div>	3	2.4	1.1E-2	1.0E0
<input type="checkbox"/>	REACTOME_PATHWAY	RHOF GTPase cycle	RT	<div></div>	3	2.4	1.8E-2	1.0E0
<input type="checkbox"/>	REACTOME_PATHWAY	Fc epsilon receptor (FCER1) signaling	RT	<div></div>	4	3.3	2.8E-2	1.0E0
<input type="checkbox"/>	REACTOME_PATHWAY	RHOD GTPase cycle	RT	<div></div>	3	2.4	2.9E-2	1.0E0
<input type="checkbox"/>	REACTOME_PATHWAY	Signal Transduction	RT	<div></div>	20	16.3	3.7E-2	1.0E0
<input type="checkbox"/>	REACTOME_PATHWAY	RHOG GTPase cycle	RT	<div></div>	3	2.4	5.1E-2	1.0E0
<input type="checkbox"/>	REACTOME_PATHWAY	RAC1 GTPase cycle	RT	<div></div>	4	3.3	6.3E-2	1.0E0
<input type="checkbox"/>	REACTOME_PATHWAY	RHO GTPase cycle	RT	<div></div>	6	4.9	6.9E-2	1.0E0
<input type="checkbox"/>	REACTOME_PATHWAY	RAC2 GTPase cycle	RT	<div></div>	3	2.4	7.0E-2	1.0E0
<input type="checkbox"/>	REACTOME_PATHWAY	MAPK6/MAPK4 signaling	RT	<div></div>	3	2.4	7.1E-2	1.0E0
<input type="checkbox"/>	REACTOME_PATHWAY	Post-translational protein modification	RT	<div></div>	12	9.8	9.0E-2	1.0E0

Figure 2. Influenza H1N1 virus Vs. Homosapiens – pathways.

Sublist	Category	Term	RT	Genes	Count	%	P-Value	Benjamini
<input type="checkbox"/>	INTERPRO	Src homology-3 domain	RT	<div></div>	6	4.9	5.9E-3	8.7E-1
<input type="checkbox"/>	INTERPRO	Pleckstrin homology-like domain	RT	<div></div>	8	6.5	7.7E-3	8.7E-1
<input type="checkbox"/>	INTERPRO	Armadillo-type fold	RT	<div></div>	7	5.7	9.5E-3	8.7E-1
<input type="checkbox"/>	INTERPRO	ATPase, AAA-type, core	RT	<div></div>	3	2.4	2.9E-2	1.0E0
<input type="checkbox"/>	INTERPRO	RNA recognition motif domain	RT	<div></div>	5	4.1	3.1E-2	1.0E0
<input type="checkbox"/>	INTERPRO	AAA+ ATPase domain	RT	<div></div>	4	3.3	3.4E-2	1.0E0
<input type="checkbox"/>	INTERPRO	Nucleotide-binding, alpha-beta plait	RT	<div></div>	5	4.1	4.9E-2	1.0E0
<input type="checkbox"/>	INTERPRO	Diacylglycerol kinase, accessory domain	RT	<div></div>	2	1.6	4.9E-2	1.0E0
<input type="checkbox"/>	INTERPRO	Pleckstrin homology domain	RT	<div></div>	5	4.1	5.0E-2	1.0E0
<input type="checkbox"/>	INTERPRO	MAD homology 1, Dwarffin-type	RT	<div></div>	2	1.6	5.9E-2	1.0E0
<input type="checkbox"/>	INTERPRO	Diacylglycerol kinase, catalytic domain	RT	<div></div>	2	1.6	7.3E-2	1.0E0
<input type="checkbox"/>	INTERPRO	ATP-NAD kinase-like domain	RT	<div></div>	2	1.6	8.2E-2	1.0E0
<input type="checkbox"/>	INTERPRO	Inorganic polyphosphate/ATP-NAD kinase, domain 1	RT	<div></div>	2	1.6	8.2E-2	1.0E0
<input type="checkbox"/>	INTERPRO	P-loop containing nucleoside triphosphate hydrolase	RT	<div></div>	9	7.3	9.0E-2	1.0E0

Figure 3. Influenza H1N1 virus Vs. Homosapiens - Protein domains.

Sublist	Category	Term	RT	Genes	Count	%	P-Value	Benjamini
<input type="checkbox"/>	UP_TISSUE	Cervix carcinoma	RT	<div></div>	40	32.5	4.4E-3	3.9E-1
<input type="checkbox"/>	UP_TISSUE	Brain	RT	<div></div>	56	45.5	8.4E-3	3.9E-1
<input type="checkbox"/>	UP_TISSUE	Cerebellum	RT	<div></div>	12	9.8	2.1E-2	6.1E-1
<input type="checkbox"/>	UP_TISSUE	Leukemic T-cell	RT	<div></div>	21	17.1	2.6E-2	6.1E-1
<input type="checkbox"/>	UP_TISSUE	Amygdala	RT	<div></div>	10	8.1	3.3E-2	6.2E-1
<input type="checkbox"/>	UP_TISSUE	Platelet	RT	<div></div>	7	5.7	6.0E-2	9.3E-1
<input type="checkbox"/>	UP_TISSUE	Spinal cord	RT	<div></div>	3	2.4	7.8E-2	1.0E0

Figure 4. Influenza H1N1 virus Vs. Homosapiens – Tissue expressions.

Sublist	Category	Term	RT	Genes	Count	%	P-Value	Benjamini
<input type="checkbox"/>	INTERPRO	Peptidase_M1, alanine aminopeptidase/leukotriene A4 hydrolase	RT		5	1.7	5.7E-6	1.8E-3
<input type="checkbox"/>	INTERPRO	Peptidase_M1, membrane alanine aminopeptidase, N-terminal	RT		5	1.7	8.1E-6	1.8E-3
<input type="checkbox"/>	INTERPRO	Heat shock protein 70, conserved site	RT		5	1.7	1.5E-5	1.8E-3
<input type="checkbox"/>	INTERPRO	Heat shock protein 70 family	RT		5	1.7	1.5E-5	1.8E-3
<input type="checkbox"/>	INTERPRO	AAA+ ATPase domain	RT		10	3.5	2.0E-5	1.8E-3
<input type="checkbox"/>	INTERPRO	Cation-transporting P-type ATPase, C-terminal	RT		5	1.7	3.3E-5	1.8E-3
<input type="checkbox"/>	INTERPRO	Cation-transporting P-type ATPase, N-terminal	RT		5	1.7	4.2E-5	1.8E-3
<input type="checkbox"/>	INTERPRO	P-type ATPase, phosphorylation site	RT		6	2.1	4.2E-5	1.8E-3
<input type="checkbox"/>	INTERPRO	P-type ATPase, A domain	RT		6	2.1	4.2E-5	1.8E-3
<input type="checkbox"/>	INTERPRO	Cation-transporting P-type ATPase	RT		6	2.1	4.2E-5	1.8E-3
<input type="checkbox"/>	INTERPRO	P-type ATPase, cytoplasmic domain N	RT		6	2.1	4.2E-5	1.8E-3
<input type="checkbox"/>	INTERPRO	P-loop containing nucleoside triphosphate hydrolase	RT		25	8.7	4.3E-5	1.8E-3
<input type="checkbox"/>	INTERPRO	P-type ATPase, transmembrane domain	RT		6	2.1	4.8E-5	1.9E-3
<input type="checkbox"/>	INTERPRO	Domain of unknown function DUF3358	RT		4	1.4	6.3E-5	2.2E-3
<input type="checkbox"/>	INTERPRO	Carbamoyl-phosphate synthetase large subunit-like, ATP-binding domain	RT		4	1.4	6.3E-5	2.2E-3
<input type="checkbox"/>	INTERPRO	ABC transporter, conserved site	RT		6	2.1	1.2E-4	3.9E-3
<input type="checkbox"/>	INTERPRO	ABC transporter-like	RT		6	2.1	2.0E-4	6.1E-3
<input type="checkbox"/>	INTERPRO	ABC transporter, transmembrane domain, type 1	RT		5	1.7	2.4E-4	6.7E-3
<input type="checkbox"/>	INTERPRO	Pre-ATP-grasp domain	RT		4	1.4	3.1E-4	8.4E-3
<input type="checkbox"/>	INTERPRO	ATP-grasp fold	RT		4	1.4	3.9E-4	1.0E-2
<input type="checkbox"/>	INTERPRO	ATP-grasp fold, subdomain 1	RT		4	1.4	7.2E-4	1.7E-2
<input type="checkbox"/>	INTERPRO	ATPase, alpha/beta subunit, N-terminal	RT		3	1.0	1.1E-3	2.3E-2
<input type="checkbox"/>	INTERPRO	ATPase, alpha/beta subunit, nucleotide-binding domain, active site	RT		3	1.0	1.1E-3	2.3E-2
<input type="checkbox"/>	INTERPRO	ATPase, F1/V1/A1 complex, alpha/beta subunit, nucleotide-binding domain	RT		3	1.0	1.1E-3	2.3E-2
<input type="checkbox"/>	INTERPRO	HAD-like domain	RT		6	2.1	1.5E-3	3.1E-2
<input type="checkbox"/>	INTERPRO	Cyclophilin-like peptidyl-prolyl cis-trans isomerase domain	RT		4	1.4	2.0E-3	4.0E-2
<input type="checkbox"/>	INTERPRO	Galactose-binding domain-like	RT		6	2.1	2.2E-3	4.2E-2
<input type="checkbox"/>	INTERPRO	Chromo domain-like	RT		4	1.4	5.1E-3	9.3E-2
<input type="checkbox"/>	INTERPRO	SNF2-related	RT		4	1.4	5.5E-3	9.4E-2
<input type="checkbox"/>	INTERPRO	Chromo domain/shadow	RT		4	1.4	5.5E-3	9.4E-2
<input type="checkbox"/>	INTERPRO	Helicase, C-terminal	RT		6	2.1	6.6E-3	1.1E-1
<input type="checkbox"/>	INTERPRO	Helicase, superfamily 1/2, ATP-binding domain	RT		6	2.1	7.6E-3	1.2E-1
<input type="checkbox"/>	INTERPRO	Organic anion transporter polypeptide OATP	RT		3	1.0	1.5E-2	2.3E-1
<input type="checkbox"/>	INTERPRO	Kazal domain	RT		4	1.4	1.8E-2	2.3E-1
<input type="checkbox"/>	INTERPRO	Cyclophilin-type peptidyl-prolyl cis-trans isomerase, conserved site	RT		3	1.0	2.1E-2	2.3E-1
<input type="checkbox"/>	INTERPRO	S-adenosylmethionine synthetase, central domain	RT		2	0.7	2.1E-2	2.3E-1
<input type="checkbox"/>	INTERPRO	S-adenosylmethionine synthetase superfamily	RT		2	0.7	2.1E-2	2.3E-1
<input type="checkbox"/>	INTERPRO	S-adenosylmethionine synthetase, C-terminal	RT		2	0.7	2.1E-2	2.3E-1
<input type="checkbox"/>	INTERPRO	S-adenosylmethionine synthetase, N-terminal	RT		2	0.7	2.1E-2	2.3E-1
<input type="checkbox"/>	INTERPRO	S-adenosylmethionine synthetase, conserved site	RT		2	0.7	2.1E-2	2.3E-1
<input type="checkbox"/>	INTERPRO	Phosphoglycerate kinase, conserved site	RT		2	0.7	2.1E-2	2.3E-1
<input type="checkbox"/>	INTERPRO	Isy1-like splicing	RT		2	0.7	2.1E-2	2.3E-1
<input type="checkbox"/>	INTERPRO	ATPase, F1 complex beta subunit/V1 complex, C-terminal	RT		2	0.7	2.1E-2	2.3E-1
<input type="checkbox"/>	INTERPRO	Phosphoglycerate kinase	RT		2	0.7	2.1E-2	2.3E-1
<input type="checkbox"/>	INTERPRO	S-adenosylmethionine synthetase	RT		2	0.7	2.1E-2	2.3E-1
<input type="checkbox"/>	INTERPRO	ATPase, F1/A1 complex, alpha subunit, N-terminal	RT		2	0.7	2.1E-2	2.3E-1
<input type="checkbox"/>	INTERPRO	Phosphoglycerate kinase, N-terminal	RT		2	0.7	2.1E-2	2.3E-1
<input type="checkbox"/>	INTERPRO	Laminin G domain	RT		4	1.4	2.6E-2	2.8E-1
<input type="checkbox"/>	INTERPRO	Carbamoyl-phosphate synthase large subunit, CPSase domain	RT		2	0.7	3.1E-2	2.9E-1
<input type="checkbox"/>	INTERPRO	Carbamoyl-phosphate synthetase, large subunit oligomerisation domain	RT		2	0.7	3.1E-2	2.9E-1
<input type="checkbox"/>	INTERPRO	Calcium-transporting P-type ATPase, subfamily IIA, SERCA-type	RT		2	0.7	3.1E-2	2.9E-1
<input type="checkbox"/>	INTERPRO	Phosphoribosyl pyrophosphate synthetase, conserved site	RT		2	0.7	3.1E-2	2.9E-1
<input type="checkbox"/>	INTERPRO	Carbamoyl-phosphate synthase, small subunit N-terminal domain	RT		2	0.7	3.1E-2	2.9E-1
<input type="checkbox"/>	INTERPRO	Carbamoyl-phosphate synthase, small subunit	RT		2	0.7	3.1E-2	2.9E-1
<input type="checkbox"/>	INTERPRO	Carbamoyl-phosphate synthase, large subunit	RT		2	0.7	3.1E-2	2.9E-1
<input type="checkbox"/>	INTERPRO	Chromo domain	RT		3	1.0	3.3E-2	3.0E-1
<input type="checkbox"/>	INTERPRO	Cadherin	RT		5	1.7	3.7E-2	3.3E-1
<input type="checkbox"/>	INTERPRO	Armadillo-type fold	RT		9	3.1	3.9E-2	3.4E-1
<input type="checkbox"/>	INTERPRO	Cadherin-like	RT		5	1.7	4.0E-2	3.4E-1
<input type="checkbox"/>	INTERPRO	Biotin-binding site	RT		2	0.7	4.2E-2	3.4E-1
<input type="checkbox"/>	INTERPRO	Methylglyoxal synthase-like domain	RT		2	0.7	4.2E-2	3.4E-1
<input type="checkbox"/>	INTERPRO	Beta-galactosidase 1-like	RT		2	0.7	4.2E-2	3.4E-1
<input type="checkbox"/>	INTERPRO	Glycoside hydrolase, family 35	RT		2	0.7	4.2E-2	3.4E-1
<input type="checkbox"/>	INTERPRO	Ribose-phosphate diphosphokinase	RT		2	0.7	5.2E-2	3.9E-1
<input type="checkbox"/>	INTERPRO	Biotin carboxylation domain	RT		2	0.7	5.2E-2	3.9E-1
<input type="checkbox"/>	INTERPRO	Pyridine nucleotide-disulphide oxidoreductase, dimerisation	RT		2	0.7	5.2E-2	3.9E-1
<input type="checkbox"/>	INTERPRO	Pyridine nucleotide-disulphide oxidoreductase, class I, active site	RT		2	0.7	5.2E-2	3.9E-1
<input type="checkbox"/>	INTERPRO	Biotin carboxylase, C-terminal	RT		2	0.7	5.2E-2	3.9E-1
<input type="checkbox"/>	INTERPRO	Carbamoyl-phosphate synthase, large subunit, N-terminal	RT		2	0.7	5.2E-2	3.9E-1
<input type="checkbox"/>	INTERPRO	Rudiment single hybrid motif	RT		2	0.7	6.2E-2	4.5E-1
<input type="checkbox"/>	INTERPRO	Glutamine amidotransferase	RT		2	0.7	6.2E-2	4.5E-1
<input type="checkbox"/>	INTERPRO	Sodium/potassium-transporting P-type ATPase, subfamily IIC	RT		2	0.7	7.2E-2	5.1E-1
<input type="checkbox"/>	INTERPRO	Pleckstrin homology domain	RT		7	2.4	7.5E-2	5.2E-1
<input type="checkbox"/>	INTERPRO	FAD/NAD-linked reductase, dimerisation	RT		2	0.7	8.2E-2	5.7E-1
<input type="checkbox"/>	INTERPRO	Kelch-like protein, gigaxonin	RT		3	1.0	8.6E-2	5.8E-1
<input type="checkbox"/>	INTERPRO	Protein kinase, catalytic domain	RT		10	3.5	8.7E-2	5.9E-1
<input type="checkbox"/>	INTERPRO	BEN domain	RT		2	0.7	9.2E-2	6.1E-1

Figure 5. Streptococcus pneumoniae Vs. Homo sapiens - protein domain.

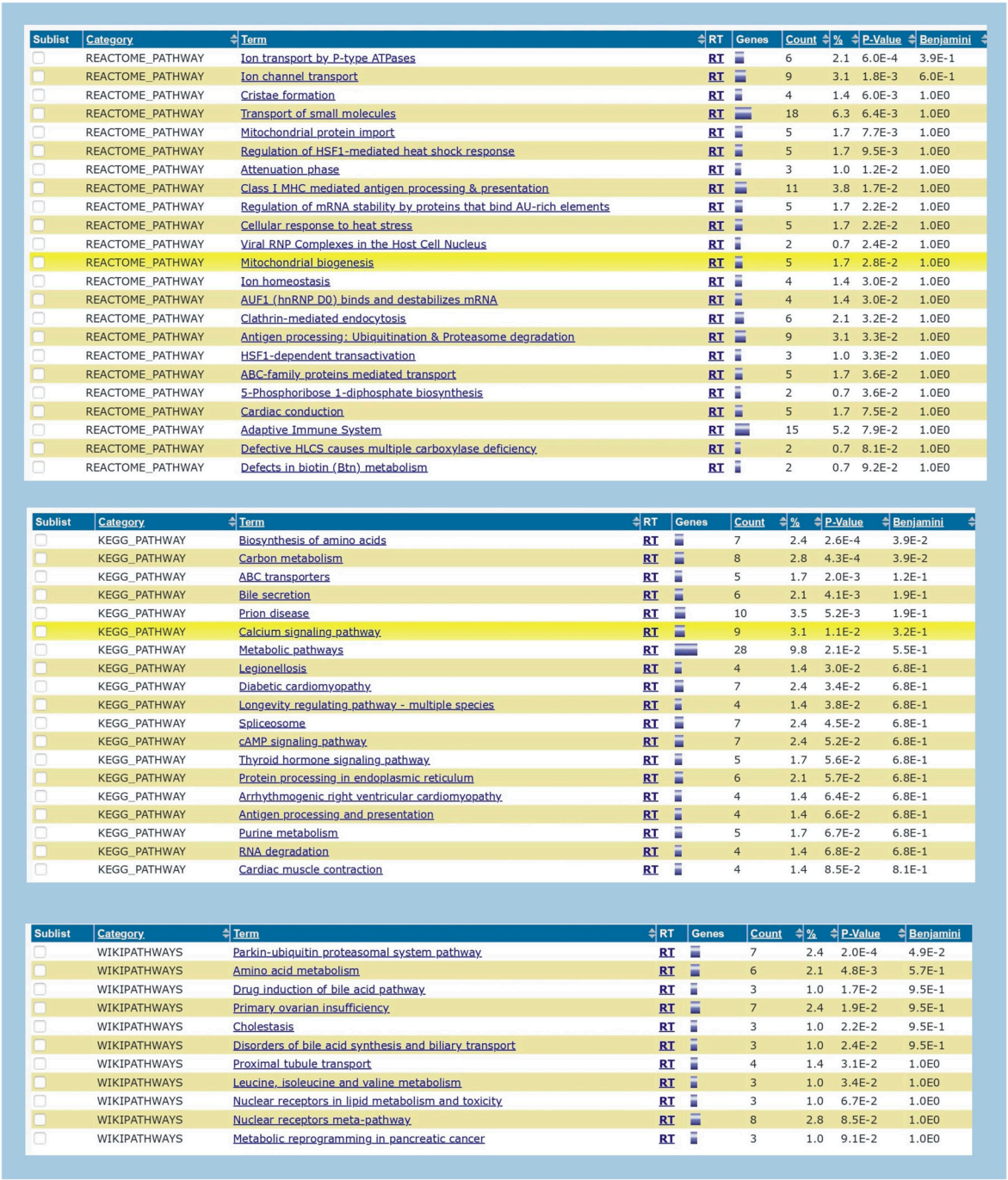


Figure 6. Streptococcus pneumoniae Vs. Homo sapiens – genes and pathways.




























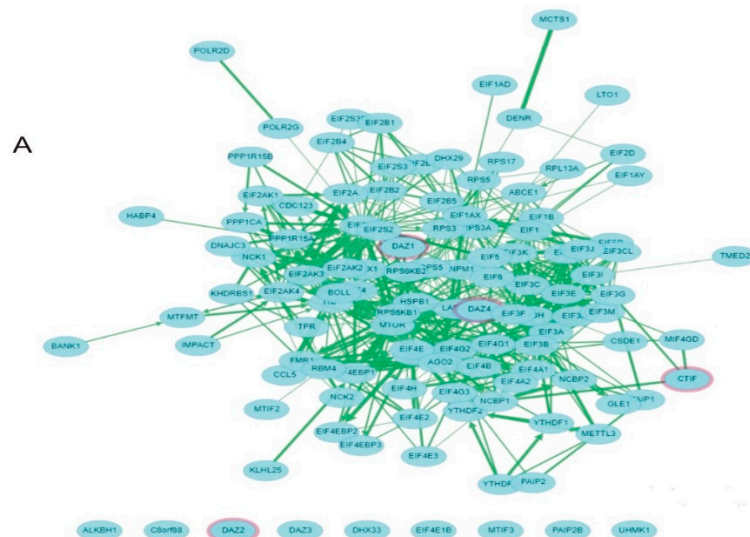
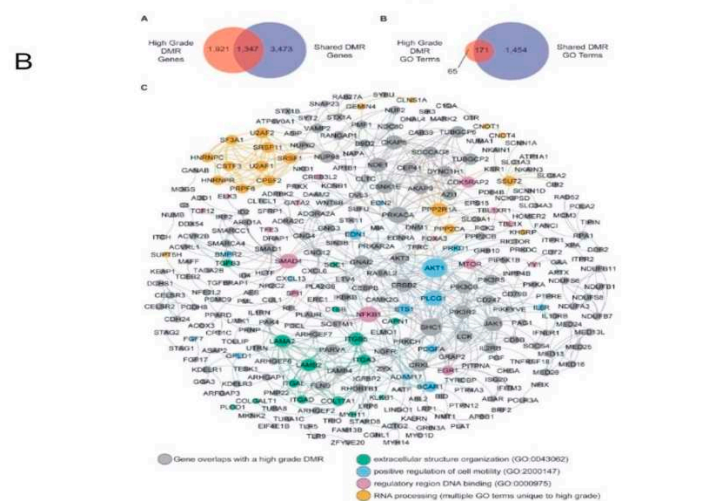
Sublist	Category	Term	RT	Genes	Count	%	P-Value	Benjamini
<input type="checkbox"/>	HPA_NORMAL_TISSUE_CELLTYPE	bronchus; ciliated cells (ciliary rootlets)	RT		14	4.9	1.4E-2	6.0E-1
<input type="checkbox"/>	HPA_NORMAL_TISSUE_CELLTYPE	liver; hepatocytes	RT		86	30.1	1.5E-2	6.0E-1
<input type="checkbox"/>	HPA_NORMAL_TISSUE_CELLTYPE	soft tissue 1; chondrocytes	RT		39	13.6	1.8E-2	6.0E-1
<input type="checkbox"/>	HPA_NORMAL_TISSUE_CELLTYPE	liver; cholangiocytes	RT		63	22.0	1.8E-2	6.0E-1
<input type="checkbox"/>	HPA_NORMAL_TISSUE_CELLTYPE	hippocampus; glial cells	RT		65	22.7	2.1E-2	6.0E-1
<input type="checkbox"/>	HPA_NORMAL_TISSUE_CELLTYPE	hippocampus; neuronal cells	RT		91	31.8	2.2E-2	6.0E-1
<input type="checkbox"/>	HPA_NORMAL_TISSUE_CELLTYPE	nasopharynx; ciliated cells (ciliary rootlets)	RT		11	3.8	2.6E-2	6.0E-1
<input type="checkbox"/>	HPA_NORMAL_TISSUE_CELLTYPE	duodenum; goblet cells	RT		11	3.8	3.1E-2	6.0E-1
<input type="checkbox"/>	HPA_NORMAL_TISSUE_CELLTYPE	bronchus; ciliated cells (cell body)	RT		23	8.0	3.1E-2	6.0E-1
<input type="checkbox"/>	HPA_NORMAL_TISSUE_CELLTYPE	cerebral cortex; neuropil	RT		75	26.2	3.2E-2	6.0E-1
<input type="checkbox"/>	HPA_NORMAL_TISSUE_CELLTYPE	endometrium 1; glandular cells	RT		97	33.9	3.6E-2	6.0E-1
<input type="checkbox"/>	HPA_NORMAL_TISSUE_CELLTYPE	rectum; endothelial cells	RT		8	2.8	4.1E-2	6.0E-1
<input type="checkbox"/>	HPA_NORMAL_TISSUE_CELLTYPE	rectum; peripheral nerve/ganglion	RT		8	2.8	4.6E-2	6.0E-1
<input type="checkbox"/>	HPA_NORMAL_TISSUE_CELLTYPE	cerebellum; cells in molecular layer	RT		67	23.4	4.6E-2	6.0E-1
<input type="checkbox"/>	HPA_NORMAL_TISSUE_CELLTYPE	thyroid gland; glandular cells	RT		100	35.0	4.7E-2	6.0E-1
<input type="checkbox"/>	HPA_NORMAL_TISSUE_CELLTYPE	cerebral cortex; neuronal cells	RT		95	33.2	4.7E-2	6.0E-1
<input type="checkbox"/>	HPA_NORMAL_TISSUE_CELLTYPE	skin 1; melanocytes	RT		77	26.9	5.9E-2	6.9E-1
<input type="checkbox"/>	HPA_NORMAL_TISSUE_CELLTYPE	nasopharynx; ciliated cells (cell body)	RT		21	7.3	6.4E-2	6.9E-1
<input type="checkbox"/>	HPA_NORMAL_TISSUE_CELLTYPE	adipose tissue; adipocytes	RT		59	20.6	7.3E-2	6.9E-1
<input type="checkbox"/>	HPA_NORMAL_TISSUE_CELLTYPE	colon; endothelial cells	RT		81	28.3	7.4E-2	6.9E-1
<input type="checkbox"/>	HPA_NORMAL_TISSUE_CELLTYPE	testis; sertoli cells	RT		37	12.9	7.7E-2	6.9E-1
<input type="checkbox"/>	HPA_NORMAL_TISSUE_CELLTYPE	fallopian tube; glandular cells	RT		82	28.7	7.9E-2	6.9E-1
<input type="checkbox"/>	HPA_NORMAL_TISSUE_CELLTYPE	pancreas; pancreatic endocrine cells	RT		80	28.8	8.2E-2	6.9E-1
<input type="checkbox"/>	HPA_NORMAL_TISSUE_CELLTYPE	ovary; ovarian stroma cells	RT		59	20.6	8.9E-2	6.9E-1
<input type="checkbox"/>	HPA_NORMAL_TISSUE_CELLTYPE	soft tissue 1; fibroblasts	RT		62	21.7	9.4E-2	6.9E-1
<input type="checkbox"/>	HPA_NORMAL_TISSUE_CELLTYPE	duodenum; endocrine cells	RT		11	3.8	9.6E-2	6.9E-1
<input type="checkbox"/>	HPA_NORMAL_TISSUE_CELLTYPE	cerebral cortex; endothelial cells	RT		67	23.4	9.8E-2	6.9E-1

Figure 7. Streptococcus pneumoniae Vs. Homo sapiens - tissue expressions.



INDRA GO-p-Value



Pathway Figures-p-Value

Figure 8. Gene ontology and pathway analysis through NDEx – Influenza virus Vs. Homo sapiens.

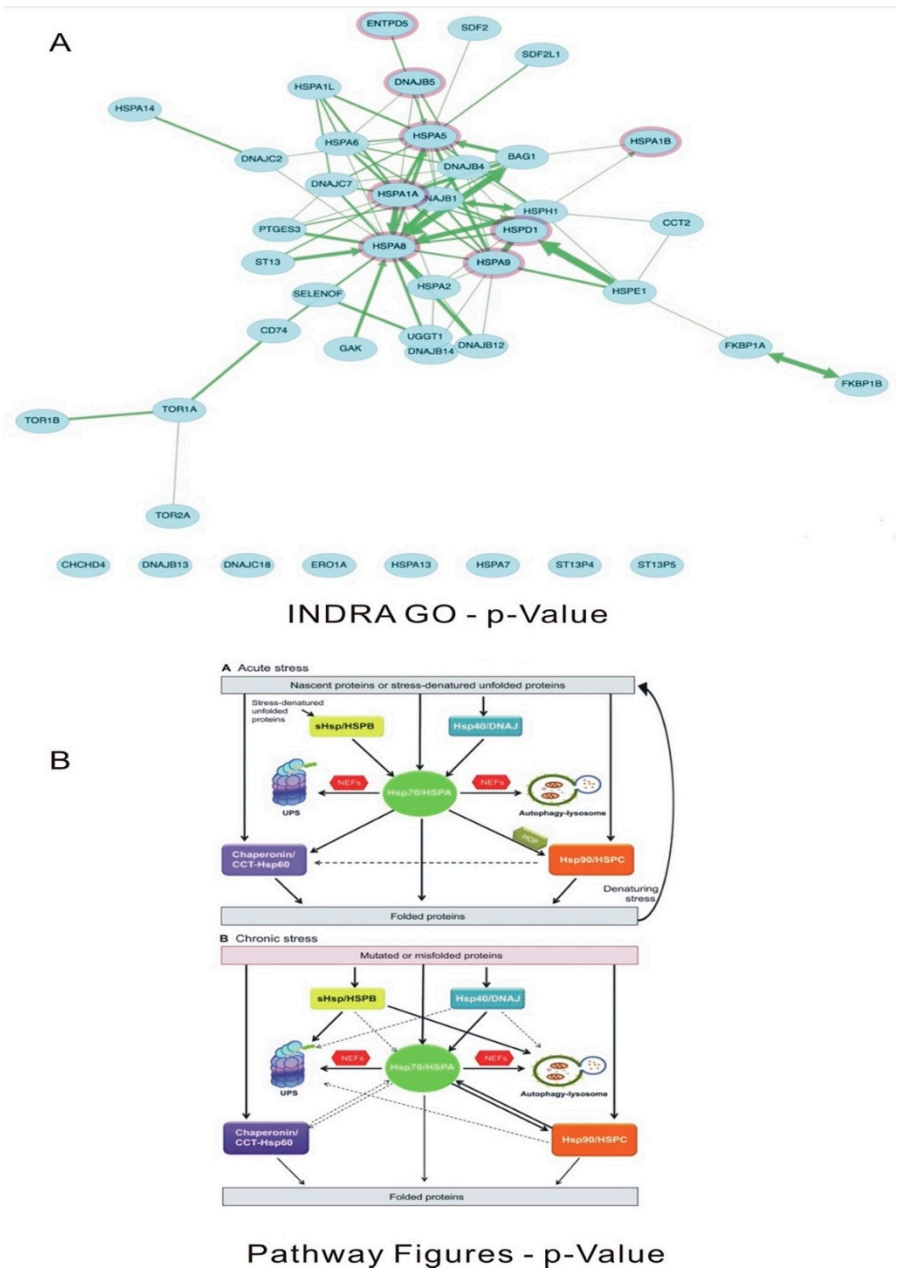


Figure 9. Gene ontology and pathway analysis through NDEx – *Streptococcus pneumoniae* Vs. *Homo sapiens*.

Discussion

In the comparison of the nucleotide sequences, there were many overlapping significant alignments between the influenza (H1N1) and *Streptococcus pneumoniae* and the human genome. This could be an evolutionary or a survival process where the organisms function and induce immunological processes by molecular mimicry. This can lead to the induction of cellular processes – cytoplasmic, nuclear, and various levels as seen through functional analysis with DAVID. These vaccinations are associated with various benefits, as reported by previous studies. These vaccinations are protective against diseases like Covid-19 through changes in adaptive immunity and are also reported to have cardioprotective effects though the exact mechanisms are not understood [9]. The countries which had high influenza or lower respiratory tract infections burden illnesses and countries which had higher influenza vaccinations, like South Korea, had significantly lower Covid 19 mortality during the pandemic [10]. Various pathways and genes are associated with the

overlapping gene. Predominantly these are inflammation, death signalling and pathways related to epigenetic modifications. Heat shock proteins are involved in normal and other cellular activities during stress, inflammation, etc. They are targets for cancer therapy, inflammation, myocardial ischemia, transplantation, and neurodegenerative diseases' activity modulation [11].

The PRKN gene is actively involved in the Parkin protein, which actively breaks down unnecessary proteins by tagging the damaged and excessive proteins with ubiquitin. Mutations in PRKN genes are well-established causes of early-onset parkinsonism [12]. TUBB, TUBA are microtubule related genes, SEPT genes are related to specific polymerisation during mitosis. The CHIP protein coded by the STUB1 gene binds and inhibits ATPase activity of the chaperone proteins HSC70 and HSP70 and prevent their forward reactions [13]. Alpha-synuclein is a protein encoded by SNCA and defects in this are associated with parkinsonism disease. GPR37 encodes [14] G-protein coupled receptor protein and it has been shown to interact with HSPA1A and Parkin(ligase). GPR37 are receptors for glial and neuroprotective factors [15].

As seen in Figure 8, extracellular structure organization ITGBF, LAMA2, LAMB2, ITG A3 and ITG B5; positive regulation of cell motility through Akt1, PLCG1 pathways; regulatory region DNA binding through NFKB1, mTOR, SMAD4, EGR1 pathways; and PRPF, SRSF 1/11, CSTF pathways associated with RNA processing are some of the actively involved in cellular pathways related genes associated with influenza-homo sapiens overlap associated genes. The AKT pathway has a significant role in interacting with oncogenes and also metabolic pathways [16]. Higher PLCG1 is associated with tumour growth and poor survival [17].

MAPKAPK2 is involved in various cellular pathways involved in stress and inflammation, nuclear export, and gene expression regulation. It plays an important role in tumour regulation [18,19]. ELAVL1 primarily couples mRNA stability with the 3' UTSs of interferon-stimulated genes [20]. The EGFR signalling pathway is one of the most important pathways in mammalian cells, which regulates a series of important events, including proliferation, migration, differentiation, and apoptosis, as well as those that regulate intercellular communication during development. EGFR is a major gene in the pathogenesis of lung cancer [21].

KRAS-related genes are involved in cellular growth, division, survival, and death. KRAS is also a target of active research for its regulatory molecular identification [22]. HSF1 is a major transcription factor for heat shock proteins. HSPA9 belongs to HSP 70 gene family. This plays a role in cellular proliferation, stress response, and maintenance of mitochondria. HSPA9 is active in regulating apoptosis also. HSP 27 was shown to be a substrate of this kinase in vivo [23]. GCCase catalyzes the cleavage of major glycolipid glucosylceramide (GL-1) into glucose and ceramide and the minor lipid glucosphingosine into sphingosine and water [24].

Further studies involving the protein encoded by these overlapping nucleotide sequences would provide more information about the changes or induction of the cellular functions. Similarly, other closely associated microorganisms like commensals can also be studied to better understand the cause-effect/associations of various disease pathology in humans, such as autoimmune disorders. Incidence of non-communicable diseases like coronary artery disease, diabetes, etc., is also interesting to study if there is any influence by the microbiological changes.

Conclusion

The study shows overlapping nucleotide sequences between the human genome and *Streptococcus pneumoniae* and Influenza (H1N1) virus genome sequences. These overlapping sequences are also associated with various functional annotations. Further evaluations can help to understand the modifications of the encoded proteins and metabolism during these infections.

Supplementary Materials: The following supporting information can be downloaded at the website of this paper posted on Preprints.org.

Author Contributions: MCA conceived the idea, performed the analysis in DAVID and NDEx, and wrote the paper. JW, JL, RH performed the Blastn analysis and derived the results in Blastn.

Funding: None.

Conflict of Interests: None.

References

1. Arokiaraj MC. Correlation of influenza vaccination and influenza incidence on COVID-19 severity and other perspectives. Available at SSRN 3572814. 2020 Apr 10.
2. Behrouzi B, Bhatt DL, Cannon CP, et al. Association of Influenza Vaccination With Cardiovascular Risk: A Meta-analysis. *JAMA Netw Open*. 2022;5(4):e228873. doi:10.1001/jamanetworkopen.2022.8873
3. Jaiswal V, Ang SP, Lnu K, Ishak A, Pokhrel NB, Chia JE, Hajra A, Biswas M, Matetic A, Dhatt R, Mamas MA. Effect of Pneumococcal Vaccine on Mortality and Cardiovascular Outcomes: A Systematic Review and Meta-Analysis. *J Clin Med*. 2022 Jun 30;11(13):3799. doi: 10.3390/jcm11133799. PMID: 35807082; PMCID: PMC9267914.
4. Hannah Chung and others, Influenza Vaccine Effectiveness Against All-Cause Mortality Following Laboratory-Confirmed Influenza in Older Adults, 2010–2011 to 2015–2016 Seasons in Ontario, Canada, *Clinical Infectious Diseases*, Volume 73, Issue 5, 1 September 2021, Pages e1191–e1199, <https://doi.org/10.1093/cid/ciaa1862>
5. Debisarun PA, Gössling KL, Bulut O, Kilic G, Zoodsma M, Liu Z, Oldenburg M, Rüchel N, Zhang B, Xu CJ, Struycken P. Induction of trained immunity by influenza vaccination-impact on COVID-19. *PLoS pathogens*. 2021 Oct 25;17(10):e1009928.
6. <https://guides.lib.berkeley.edu/ncbi/blast>.
7. Lobo, I. (2008) Basic Local Alignment Search Tool (BLAST). *Nature Education* 1(1):21.
8. DAVID Brad T Sherman and others, DAVID: a web server for functional enrichment analysis and functional annotation of gene lists (2021 update), *Nucleic Acids Research*, Volume 50, Issue W1, 5 July 2022, Pages W216–W221, <https://doi.org/10.1093/nar/gkac194>.
9. Wilcox CR, Islam N, Dambha-Miller H. Association between influenza vaccination and hospitalisation or all-cause mortality in people with covid-19: A retrospective cohort study. *BMJ Open Respiratory Research*. 2021;8(1). doi:10.1136/bmjresp-2020-000857.
10. Arokiaraj MC. Considering Interim Interventions to Control COVID-19 Associated Morbidity and Mortality-Perspectives. *Front Public Health*. 2020 Sep 22;8:444. doi: 10.3389/fpubh.2020.00444. PMID: 33072682; PMCID: PMC7537040.
11. Dubey A, Prajapati KS, Swamy M, Pachauri V. Heat shock proteins: a therapeutic target worth to consider. *Vet World*. 2015 Jan;8(1):46–51. doi: 10.14202/vetworld.2015.46–51. Epub 2015 Jan 13. PMID: 27046995; PMCID: PMC4777810.
12. Castelo Rueda MP, Raftopoulou A, Gögele M, Borsche M, Emmert D, Fuchsberger C, Hantikainen EM, Vukovic V, Klein C, Pramstaller PP, Pichler I and Hicks AA (2021). Frequency of Heterozygous Parkin (PRKN) Variants and Penetrance of Parkinson's Disease Risk Markers in the Population-Based CHRIS Cohort. *Front. Neurol*. 12:706145. doi: 10.3389/fneur.2021.706145
13. Zhang, S., Hu, Zw., Mao, Cy. *et al*. CHIP as a therapeutic target for neurological diseases. *Cell Death Dis* **11**, 727 (2020). <https://doi.org/10.1038/s41419-020-02953-5>
14. Guo Y, Sun Y, Song Z, Zheng W, Xiong W, Yang Y, Yuan L, Deng H. Genetic Analysis and Literature Review of SNCA Variants in Parkinson's Disease. *Front Aging Neurosci*. 2021 Aug 12;13:648151. doi: 10.3389/fnagi.2021.648151. PMID: 34456707; PMCID: PMC8397385.
15. Meyer RC, Giddens MM, Schaefer SA, Hall RA. GPR37 and GPR37L1 are receptors for the neuroprotective and glioprotective factors prosaptide and prosaposin. *Proceedings of the National Academy of Sciences*. 2013;110(23):9529–34. doi:10.1073/pnas.1219004110
16. Nitulescu G, Van De Venter M, Nitulescu G, Ungurianu A, Juzenas P, Peng Q, et al. The AKT pathway in oncology therapy and beyond (review). *International Journal of Oncology*. 2018; doi:10.3892/ijo.2018.4597
17. Li, T., Yang, Z., Li, H. *et al*. Phospholipase Cγ1 (PLCG1) overexpression is associated with tumor growth and poor survival in IDH wild-type lower-grade gliomas in adult patients. *Lab Invest* 102, 143–153 (2022). <https://doi.org/10.1038/s41374-021-00682-7>
18. Yang L, Liu B, Qiu F, Huang B, Li Y, Huang D, et al. The effect of functional *mapkapk2* copy number variation CNV-30450 on elevating nasopharyngeal carcinoma risk is modulated by EBV infection. *Carcinogenesis*. 2013;35(1):46–52. doi:10.1093/carcin/bgt314
19. Soni, S., Anand, P. & Padwad, Y.S. MAPKAPK2: the master regulator of RNA-binding proteins modulates transcript stability and tumor progression. *J Exp Clin Cancer Res* 38, 121 (2019). <https://doi.org/10.1186/s13046-019-1115-1>
20. Rothamel K, Arcos S, Kim B, Reasoner C, Lisy S, Mukherjee N, et al. ELAVL1 primarily couples mRNA stability with the 3' utrs of interferon-stimulated genes. *Cell Reports*. 2021;35(8):109178. doi:10.1016/j.celrep.2021.109178
21. Carcereny E, Morán T, Capdevila L, Cros S, Vilà L, de Los Llanos Gil M, Remón J, Rosell R. The epidermal growth factor receptor (EGFR) in lung cancer. *Transl Respir Med*. 2015 Feb 24;3:1. doi: 10.1186/s40247-015-0013-z. PMID: 25810955; PMCID: PMC4366432.

22. Huang, L., Guo, Z., Wang, F. *et al.* KRAS mutation: from undruggable to druggable in cancer. *Sig Transduct Target Ther* 6, 386 (2021). <https://doi.org/10.1038/s41392-021-00780-4>.
23. Liu T, Krysiak K, Shirai CL, Kim S, Shao J, Ndonwi M, et al. (2017) Knockdown of HSPA9 induces TP53-dependent apoptosis in human hematopoietic progenitor cells. *PLoS ONE* 12(2): e0170470. <https://doi.org/10.1371/journal.pone.0170470>
24. HSC 70 Wang Z, Li Y, Yang X, Zhao J, Cheng Y and Wang J (2020) Mechanism and Complex Roles of HSC70 in Viral Infections. *Front. Microbiol.*11:1577. doi: 10.3389/fmicb.2020.01577

Figures

1. Influenza H1N1 virus Vs. Homo sapiens – functional annotation chart with biogrid interactions by DAVID analysis.
2. Influenza H1N1 virus Vs. Homo sapiens – genes in pathway map.
3. Influenza H1N1 virus Vs. Homo sapiens – functional protein domains.
4. Influenza H1N1 virus Vs. Homo sapiens - tissue expressions.
5. Streptococcus pneumoniae Vs. Homo sapiens – functional protein domain.
6. Streptococcus pneumonia Vs. Homo sapiens – genes in pathway map.
7. Streptococcus pneumoniae Vs. Homo sapiens - tissue expressions.
8. Gene ontology and pathway analysis through NDEx – Influenza virus Vs. Homo sapiens.
9. Gene ontology and pathway analysis through NDEx – Streptococcus pneumoniae Vs. Homo sapiens.

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.