

Review

Potential Utilization of Bacterial Consortium of Symbionts Marine Sponges in Removing Pollutants Global Trends, A Review

Ismail Marzuki ^{1*}, Rosmiati Rosmiati ², Akhmad Mustafa ², Sahabuddin Sahabuddin ², Tarunamulia ², Endang Susianingsih ², Erfan Andi Hendrajat ², Andi Sahrijanna ², Muslimin Muslimin², Erna Ratnawati ², Kamariah Kamariah ², Khaerun Nisaa ², Susila Herlambang ³, Sri Gunawan ⁴, Idum Satia Santi ⁴, Bambang Heri Isnawan ⁵, Ernawati Syahrudin Kaseng ⁶, Early Septiningsih ⁷, Ruzkiah Asaf ⁷, Admi Athirah ⁷ and Basri Basri⁸

¹ Department of Chemical Engineering, Fajar University, Makassar 90231, South Sulawesi, Indonesia; ismailmz@unifa.ac.id (I.M)

² Research Center for Fishery National Research and Innovation Agency, Cibinong 16911, West Java, Indonesia; rosm005@brin.go.id (R.R); andi.akhmad.mustafa@brin.go.id (A.M); saha006@brin.go.id (S.S) tarunamulia@brin.go.id (T.M); enda055@brin.go.id (E.S); erfa001@brin.go.id (E.H); andi058@brin.go.id (A.S); musliminsyari2@gmail.com (M.M); erna010@brin.go.id (E.R); kama004@brin.go.id (K.K), nisauicha27@gmail.com (K.N)

³ Soil Science Department of agriculture faculty Universitas Pembangunan Nasional Veteran Yogyakarta; susilaherlambang@upnyk.ac.id (S.H)

⁴ Department of Agrotechnology, Institut Pertanian Stiper, Yogyakarta 55283, DI Yogyakarta; sriegun@instiperjogja.ac.id (S.G); idum@instiperjogja.ac.id (I.S)

⁵ Department of Agrotechnology, Universitas Muhammadiyah Yogyakarta, Bantul 55183, DI Yogyakarta; bambanghi@umy.ac.id (B.I)

⁶ Agricultural Technology Education Department, Faculty of Engineering, Makassar State University; ernawatisyahrudin71@unm.ac.id (E.K)

⁷ Research Center for Conservation of Marine and Inland Water Resources, National Research and Innovation Agency, Cibinong 16911, West Java, Indonesia; earl002@brin.go.id (Early.S); ruzk001@brin.go.id (R.A); admi001@brin.go.id (A.A)

⁸ Institute of Health Science (STIK) Makassar, South Sulawesi; basrikesmas@gmail.com (B.B)

* Correspondence: ismailmz@unifa.ac.id (I.M)

Abstract: Toxic materials in waste generally contain several components of the global trending pollutant category, especially PAHs and heavy metals. Bioremediation technology for waste management that utilizes microorganisms (bacteria) has not been fully capable of breaking down these toxic materials into simple and environmentally friendly chemical products. This study examines the potential application of a consortium of marine sponge symbionts with high performance and efficiency in removing PAHs and heavy metal contaminants. The method is carried out through a review of several related research articles by the author and published by other researchers. The results of the study concluded that the development of GTP bioremediation technology could be carried out to increase the efficiency of remediation. Several types of marine sponge symbiont bacteria, hydrocarbonoclastic (R-1), metaloclastic (R-2), and metallohydro-carbonoclastic (R-3), have the potential to be applied to improve waste removal performance. A consortium of crystalline bacterial preparations is required to mobilize to GTP-exposed sites rapidly. Bacterial symbionts of marine sponges can be traced mainly to sea sponges whose body surface is covered with mucus.

Keywords: removal; PAHs; heavy metals; marine sponges; bacterial consortium

1. Introduction

Global trending pollutant (GTP) is a term applied to several types of pollutant materials (heavy metals, aromatic hydrocarbons, microplastics, medical waste, pesticide residues) that pose many complex problems to the global environment [1-7]. The problem of environmental quality is especially felt by developing countries [3,8]. The issue of GTP in

this decade has been very much discussed, not only by environmental observers and activists as well as scientists in the field of environmental management but also by several world leaders who have voiced the need for real action in reducing fossil fuel consumption [9-11]. The global policy of reducing carbon consumption is a tangible manifestation of environmental quality in an emergency [7,12]. It is very reasonable because the rate of increase in global trends of pollutants increases massively, far beyond the natural recovery ability of nature to reduce these pollutants [13-15]. Reducing carbon in the environment is not enough because various types of GTP and the adverse environmental effects are also different [7,16].

The aquatic environment is a giant container that is most vulnerable to being affected by GTP contaminants [17]. It is because the topography of the area is generally low. Almost all types of GTP contaminants are found in particulates and residues and can be dissolved in air, soil and water bodies [4,7]. These materials eventually empty into the aquatic environment of rivers, lakes, swamps and the sea [18,19]. In this environment, these toxic pollutants form parasitic accumulations of almost all marine organisms, especially fish, sponges, algae, plankton, phytoplankton and other types of biota [20,21].

At the same time, much research has been carried out related to the management of toxic pollutants in order to achieve the dream of creating a green environment [22,23]. The results of this research have given birth to many findings, technologies, innovations and methods related to removing the toxic nature of contaminants [24,25]. Degradation, reduction, destruction, and absorption are recommended and can be applied to decompose carcinogenic pollutants in the environment [26-28]. This method can be called bioremediation technology if it involves the role of living organisms, including the contribution of microorganisms as biodegradators that can decompose or may be able to eliminate the toxic properties of GTP contaminants [29-31].

The principle of bioremediation technology is the development of biological methods that can be applied in a gradual combination with physical, chemical, and biological methods, depending on the characteristics of the pollutant being degraded [32,33]. Bioremediation technology that uses the role of microorganisms as a decomposer component is generally good and is more often applied to wastewater treatment [34,35]. Sources of microorganisms that can carry out bioremediation functions can be obtained in an aquatic environment contaminated with pollutants, for example, in port areas, offshore oil processing industries or marine areas around locations that have experienced oil spills [36-38]. Pollutant degrading microorganisms can also be found in the soil, especially on land with a history of contamination due to oil spills and agricultural land exposed to pesticides [3,39]. Toxic pollutant-degrading microorganisms can also be found in other organisms that form a symbiotic relationship, for example, in sponges and mangroves [40,41].

Isolation, screening and screening methods can be applied to obtain potential microorganisms as biomaterials for degrading pollutants [42,43]. In general, three microorganisms have the potential as biomaterials for degrading pollutants, namely bacteria, fungi and fungi [44-46]. These three groups of microorganisms have different abilities and degradation mechanisms for pollutants [47]. The application of bacteria in bio-remediation mainly removes hydrocarbon contaminants, especially polyaromatic hydrocarbons (PAHs) and heavy metal pollutants. At the same time, the use of fungi and fungi in pollutant bioremediation has also been carried out but is still limited or not as popular as bacteria [48-50].

The types of bacteria identified have bioremediation capabilities against PAHs, but the level of bioremediation produced is still low, mainly if one type of bacteria is used [51,52]. It is because bioremediation bacteria are generally resistant to acidic environments. At the same time, it is known that one of the hydrocarbon component bioremediation products is simple organic compounds resulting from oxidation reactions in the form of alcohols, aldehydes, ketones and possibly carboxylate group compounds [18,53,54]. Carboxylic compounds of bacterial bioremediation products can change the conditions of bacterial habitat (media) in an acidic environment so that the degradation activity of bac-

terial cells decreases or may die in bulk [51,55]. This condition is called the limiting factor for the performance of bacterial bioremediation [56].

In the aquatic environment, especially in marine ecosystems, there are several types of biota, such as sponges which are known to be often used as objects for biomonitoring of pollution of hydrocarbon components and heavy metals, even some types of sponges are used as references or bioindicators in analyzing the level of PAHs and heavy metal contaminants [54,55]. Further exploration related to biomonitoring and bioindicators of this type of GTP contamination, it is known that this sponge can perform the degradation of hydrocarbon components and biosorption of heavy metals [7,56-58]. It is based on research results that show the ability of sponges to live and thrive in an environment contaminated with GTP pollutants [7,59]. The development of knowledge about the degradation and adsorption function of marine sponges against pollutants was revealed after discovering that these sponges can have a mutualistic symbiosis with microorganisms, especially bacteria [60-62].

The bacterial-sponge symbiosis model is intensified when the sponge habitat is exposed to hydrocarbon or heavy metal pollutants or perhaps both, whereas the sponge tries to survive in the extreme conditions of its growing environment [63-65]. At the same time, symbiont bacteria also use these conditions to produce a mucus substance that behaves as an enzyme, which is then spread on the surface of the sponge body to avoid the toxic nature of the pollutant [66-68]. Internal sponges also independently stimulate themselves to have immunity against all forms of predators and changes in their habitat by producing metabolic substances [53, 69,70].

The types and populations of sponges are huge, so the right sponge selection is needed by tracing, including symbiont bacteria that have the potential and ability to degrade and adsorb. It can be done by selecting sponges in their habitat, especially those with dark colours or smooth body surfaces because they are coated with mucus [20,21,71-73]. Bacterial symbionts from the selected sponge were then isolated to obtain a single isolate [74,75]. Phenotypic analysis of sponge symbiont bacteria through biochemical tests using standard reagents needs to be carried out to ensure that these symbiotic bacteria can perform the functions of PAH degradation and heavy metal adsorption [6,76]. Bacterial symbionts are potential if they react positively with several biochemical reagents, especially Methyl Red, Voges-Proskauer, citrates, lactose, catalase, nitrate reduction and indole reagents [77,78]. Genotypic analysis of bacterial symbionts is important to obtain complete information related to bacterial species, strain and number of DNA base pairs, and it is also possible to carry out genotypic analysis of these symbiotic bacteria using 16S rRNA sequences [36,79-81].

The degradation function of sponge symbiont bacteria for the target of qualitative analysis can be done by spotting several bacterial cells on media containing hydrocarbon components such as pyrene [82-84]. Bacteria that can adapt to the environment exposed to PAHs are characterized by their activity after being incubated for ± 24 hours. This situation indicates that bacteria can carry out the function of biodegradation of polyaromatic hydrocarbon pollutants [85,86]. A preliminary qualitative test can determine the adsorption function of sponge symbiont bacteria by inserting ± 1 mL of bacterial suspension into a medium containing heavy metal contaminants after being incubated for ± 24 hours, then measuring the optical density (OD600) of the interaction medium. If there is an increase in increased turbidity or absorption indicates that the symbiont bacteria could perform the adsorption function on the heavy metals tested [87,88].

The mechanism of bioremediation of hydrocarbon components is slightly different between aliphatic and aromatic hydrocarbons [89,90]. The mechanism of degradation of hydrocarbon components in general by microorganisms, especially sponge symbiont bacteria, through oxidation reactions or biochemical reactions at the molecular level [69,91]. The entry of metabolic substances or enzymes (dioxygenase) produced by bacteria into the structure of hydrocarbon molecules that act as substrates so that these molecules un-

dergo destruction, causing the molecules to break down into molecules that produce organic compound products containing hydroxyl functional groups, which then turn into keto-enol compounds [92-94].

The oxidation reaction continues for the initial component and the degradation product of the first product of the keto-enol complex, which is further oxidized to produce organic compounds of the aldehyde, carboxylic and possibly ketone groups [95,96]. The oxidation reaction process ideally continues until simple organic molecules are formed in the form of substances that can enter the metabolic cycle [97]. The oxidation reaction of hydrocarbon components can run if the ideal conditions required for the degradation of bacteria are met [98]. Generally, the degradation performance of bacteria decreases when the oxidation results produce a carboxylic acid product [99]. Another factor that can inhibit bacterial degradation of the substrate (reactant) of the hydrocarbon component is the low solubility of the hydrocarbon component, making it difficult for bacteria to penetrate [100,101].

The rate of bacterial biodegradation of hydrocarbon contaminants varies in the range of 35-97%. Several factors cause this, for example, the type of bacteria used and the type of hydrocarbon pollutant (aliphatic or PAHs) [102,103]. Interaction time, degradation method, the concentration of hydrocarbon components as reactants, number of bacterial cells, presence or absence of nutrition, oxygen injection (aeration), the scale of experiments carried out and other factors [104,105]. Variations in the level of bacterial degradation of hydrocarbon components indicate that multiple factors influence bioremediation [106]. Two things always occur in the biodegradation of hydrocarbon components (subtracts) by bacteria (degradators), namely: (1) Biodegradation of hydrocarbon pollutants by bacteria through an oxidation reaction pathway involving enzymes produced by bacteria in response to the presence of toxic substances in their growth habitat [107,108]. (2) A decrease in the performance of bacterial degradation when degradation products are formed in the form of carboxylic compounds. These two things cause the degradation of hydrocarbon components, especially PAHs, to be incomplete or unable to degrade the substrate 100% [109]. Pyrene biodegradation generally stops at the stage where the benzene component is formed [110,111]. Under these conditions, it was assumed that all bacterial cells had died [112].

These data make it possible to modify the biodegradation of hydrocarbon components by using a consortium of bacteria with diverse species or a consortium of microorganisms (a mix of bacteria and fungi) [51,92,113]. The first modification is a consortium of certain species of bacteria (X), which has high degradation activity, is expected to work at the beginning of contact and is combined with bacteria (Y) which have a slow adaptation rate to continue the degradation process when the cells of the X-species bacteria have died [114]. The second modification, a consortium of X-species bacteria that performs in degradation combined with fungi of type (Z), is more tolerant of acids that can degrade hydrocarbons in acidic media [115,116]. This method is one of the potential alternatives for developing bioremediation technology. It is hoped that all hydrocarbon components will be wholly or 100% degraded and produce the final product of simple organic compounds, in the form of salicylic acid and the like, which are environmentally friendly [117,118].

2. Global Trends Pollutant Bioremediation Analysis Instrument

Bioremediation of global pollutant trends is important by applying new technologies and innovations to improve remediation efficiency so that the natural balance, especially in marine ecosystems, can be maintained [119]. Bacterial consortia in bioremediation is a new approach and are needed in the future. Analysis of the performance and efficiency of bacterial remediation against PAHs generally uses instruments such as Gas chromatography-mass spectrometry (GC-MS) [3,57], Fourier-transform infrared spectroscopy (FTIR), Scanning electron microscopy (SEM), Energy dispersive X-ray spectroscopy (EDS) and X-ray diffraction analysis (XRD). In contrast, remediation of heavy metals by bacteria

generally uses the Atomic Absorption Spectroscopy (AAS) analytical instruments; Inductively Coupled Plasma (ICP) can be combined with Optical Emission spectroscopy or mass spectrometry (MS) [120-124]. The combination of analytical instruments in pollutant bioremediation can be carried out, especially for the bioremediation of pollutants containing two or more types [122].

Waste generated in the petroleum processing industry in the form of sludge which generally contains hydrocarbon component pollutants, both aliphatic and aromatic, also contains heavy metal toxic materials, so a combination of analytical instruments is often used to obtain data related to performance, efficiency, mechanism, remediation products, including models and changes to the pollutant material during remediation [125,126]. Researchers often use the combination of analytical instruments in the bioremediation of petroleum sludge waste by a single bacterium/bacterial consortium, such as GC-MS, FTIR and AAS [20,57,121]. SEM, EDS and XRD instruments usually observe changes in surface shape or structure of bacteria-heavy metal complexes through extracellular bonding [120-123].

3. Bacterial performance in pollutant bioremediation

Bioremediation innovation by using a consortium of bacteria in pollutant remediation is a global trend, especially for PAHs and heavy metal pollutants, which aims to improve remediation efficiency and performance so that the final product of remediation is in the form of simple organic compounds, environmentally friendly and no longer causes health effects on all living things around them [51,91,110].

The relationship between single species biodegradator remediation performance on PAH components and bacterial cell growth based on interaction time is presented in Figure 1.

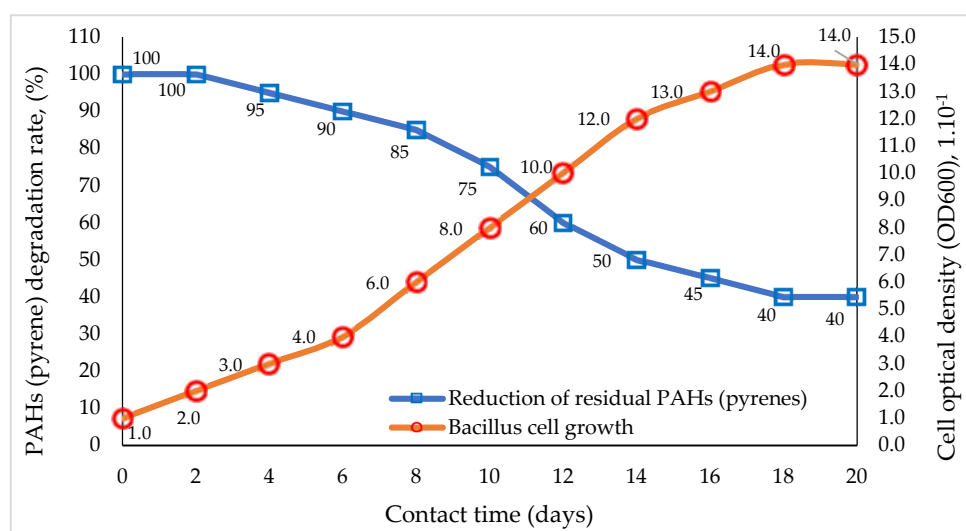


Figure 1. Comparison of the percentage of PAHs (Pyrene) bioremediation with bacterial cell growth rates based on interaction time.

Bioremediation of PAHs using one type of microorganism (bacteria) is often inefficient and has low performance in degrading PAHs substrate [48,106]. It happens because the biodegradation of PAHs by bacteria takes place in several stages, generally starting with an oxidation reaction and then destroying the substrate's molecular structure. The substrate degradation process, both biostimulation and bioaugmentation methods [127,128], using bacteria, continues until transition products are obtained in the form of acidic compounds (carboxylic) [8-11,33,56]. At this stage, the performance of bacteria often drops significantly due to their inability to tolerate an acidic medium [28,77]. As a result of this process, bacterial cells cannot continue the process of cell division, so there is no

more prolonged regeneration of bacterial cells that grow to continue the degradation process [56,124]. The degradation step is the formation of carboxylic acid compounds, where most bacteria cannot tolerate acidic conditions, so this step in the bioremediation method is often called the rate-limiting step of degradation [11,23,41].

The reduction of the PAHs component (pyrene) by *Bacillus* bacteria (Fig. 1) increased with increasing interaction time, followed by an increase in the growth of *Bacillus* cells [38,51]. The growth of *Bacillus* cells appeared to be stagnant on the 18th to 20th day of interaction (Fig.1). In this condition, the growth activity of *Bacillus* cells was considered non-existent, so the biodegradation process of the pyrene substrate also stopped, while the pyrene residue remained $\pm 40\%$ of the initial amount [30,129].

Figure 1 above illustrates the weaknesses or limitations of using one type of micro-organism species (bacteria) in the biodegradation process. This illustration shows the performance of *Bacillus* in pyrene degradation for 18 days [6,109]. Bacteria use this time duration to carry out their degradation mission of pyrene through the adaptation phase of the interaction environment, cell growth and multiplication, and stationary and cell-cell death phases [111,113]. The performance of bacterial degradation results in the gradual destruction of the pyrene molecular structure following the growth and development of bacterial cells, resulting in transitional organic compound products until the bacteria reach the degradation stage of the production of carboxylic acid components [44,129]. The concentration of $\pm 40\%$ remaining pyrene at the end of the biodegradation process, if it enters the environment, is still high and does not guarantee safety for living creatures around it [4,98].

Bioremediation technology continues to develop today. One of the developments and advances in bioremediation technology is the innovation of using a consortium of bacteria to remediate toxic and carcinogenic PAHs [22,105,124]. The study of the performance and efficiency of biodegradation in the application of bacterial consortia or groups of tolerant bacteria to the toxicity of PAHs (hydrocarbonoclastic) [68,113] is presented in Figure 2.

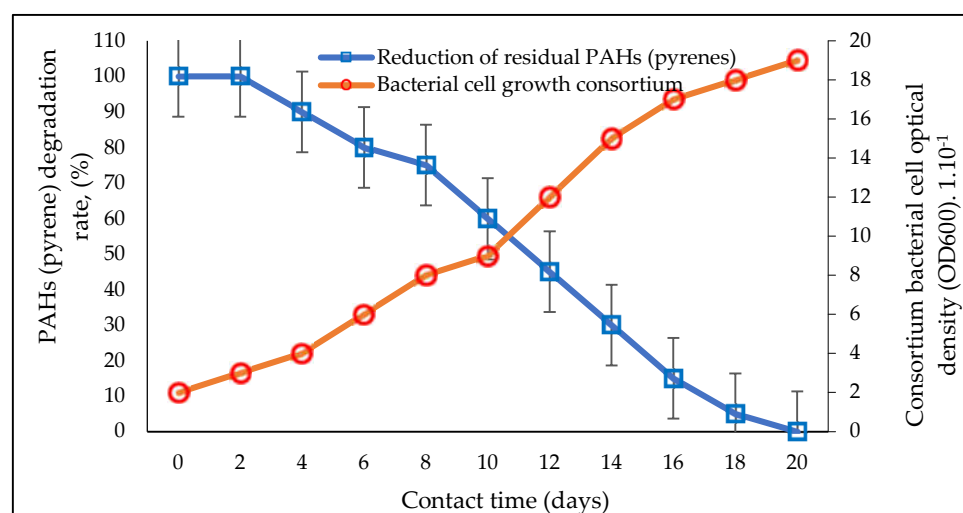


Figure 2. Comparison of the percentage of PAHs (Pyrene) bioremediation with the bacterial cell growth rate of the consortium based on interaction time.

Studies on using hydrocarbonoclastic biodegradators in the bioremediation of PAHs can give better results than using single species bacteria [81,106]. The application of consortium bacteria in the bioremediation of PAHs is considered to have higher performance and substrate degradation power and is more efficient because it is estimated that the bioremediation performance of PAHs increases to 100% (Fig. 2) [27,60,126]. The time needed by the consortium bacteria to degrade potential PAHs is less than 20 days, which

can be seen from the consortium bacterial cells that still show growth [51,92]. This condition indicates that bacterial cells can still carry out cell division and degradation of hydrocarbon components as an energy source [5,31].

Table 1. Results of a recent study on the application, performance and efficiency of bacteria in the remediation of polycyclic aromatic hydrocarbons (PAHs) and heavy metal pollutants.

Type of contaminant	Bioremediation method	Test System	interaction duration	Removal Efficiency	Conclusion	References
Pyrene (±10 mg/kg)	Biosurfactants (Biodegradation)	soil microorganism	10 days	±60%	The biodegradation process can occur due to the ability of rhamnolipids to convert carbon into energy sources	[111]
Phenanthrene (±1.0 mg/L)	Biodegradation	using soil adsorption reactor	±90%	more than 50 days	No significant effect of the observed biodegradation efficiency of surfactants	[54]
PAHs (±574 mg/kg)	Biodegradation	Soil microorganism	84 days	72-77%	The formation of surfactants marks the ongoing biodegradation process	[74]
Pyrene (±100 mg/L)	Biodegradation	<i>Sphingobacterium</i> sp. strain 21	30 days	±38,29%	The biodegradation performance of pyrene increases at the contact period of 6-20 days	[71]
Pyrene, phenanthrene and others (±6 mg/kg)	Biosurfactants (Biodegradation)	Soil microorganism	±35 days	58-72%	Biosurfactants (<i>rhamnolipids</i>) can only carry out biodegradation until the 7th day, then the PAHs biodegradation process does not appear until the 35th day	[85]
Phenanthrene (±1.0 mg/L)	Biodegradation	flake model	14 days	60%	<i>Rhamnolipids</i> as surfactants can increase the efficiency of biodegradation at a concentration of 100 mg/L	[127]
PAHs (±1.5 mg/g)	Biostimulation	Soil microorganism	56 days	±99%	Biostimulant effect can increase biodegradation kinetics	[94]
Pyrene (100 mg/L)	Biodegradation using vial reactor	<i>Alcaligenes faecalis</i> strain Cu4-1	25 days	97.65%	The products of pyrene biodegradation by the two types of bacteria are relatively different, indicating that there are different metabolic pathways that are influenced by these types of bacteria	[64]
Petroleum refinery waste (±144 g/kg)	Combined biostimulation and bioaugmentation	<i>Bacillus Cereus</i> strain MER-8	120 days	57-75%	modification of the method by applying a combination of biostimulation and bioaugmentation to increase remediation efficiency	[128]
Alkanes (initial concentration not determined)	bioaugmentation	<i>Microorganisms in vial activated microcosm consortium</i>	85 days	35-66%	The use of the adapted microcosm consortium is able to degrade the hydrocarbon component as a substrate to produce biosurfactants	[95]
Pb(II) and Cd(II)	bioadsorption	<i>Burkholderia fungorum</i>	7 days	50 mg/L and	<i>B. fungorum</i> strain FM-2 is tolerant to Pb(II) and Cd(II) or can	[129]

		FM-2		400 mg/L	carry out the function of bioadsorption of heavy metals	
Pollutant Cd and Hg	Bioadsorption	<i>Pseudomonas</i> sp., <i>Salinobacter</i> sp., <i>Streptomyces</i> sp., <i>Roseobacter</i> sp., <i>Vibrio</i> sp., <i>Saccharomonospora</i> sp. and others isolated from marine sponge <i>Fasciospongia cavernosa</i>	7 days	preliminary test	This sponge symbiotic bacteria is able to survive in habitats contaminated with heavy metals mercury and cadmium	[130]
Pb(II) and Cd(II)	remediation in oxidation method	Natural adsorbent available in aquatic habitat	2 days	9.03 mg/g and 8,85 mg/g	Natural adsorbent found in the aquatic environment in remediating Pb(II) and Cd(II) pollutants. The isotherm data was processed using the Langmuir approach, showing that lead remediation is endothermic and cadmium is exothermic	[28]
Ions Co, Pb, Cu, Zn	Bio-adsorption	<i>Rumex crispus</i> L	7 days	83.5-91.0%	The findings revealed that the heavy metal absorption mechanism occurs on the surface of the bio-sorbent to form a metal-biosorbent complex	[60]

Studies conducted regarding the bioremediation of hydrocarbon components using microorganisms applying different methods, such as biodegradation, biostimulation, bioaugmentation or a combination of the two methods (Table 1), show that none of the experiments has succeeded in degrading hydrocarbon pollutants with efficiency reaching 100% [94,128]. The study of heavy metal bioremediation by microorganisms also showed that no single type of bacteria could absorb heavy metal contaminants in waste with 100% efficiency [52,116,130].

The results of this search indicate the need to develop bioremediation technology for hydrocarbon pollutants (PAHs) and heavy metals. One of the innovations and remediation engineering is the use of several types of bacteria that can bioremediate PAHs and heavy metals to achieve 100% remediation efficiency [37,60,66].

Table 2. Various studies on the biodegradation performance of sponge symbiont bacteria on hydrocarbon components.

Types of hydrocarbon contaminants	Sponge symbiont bacterial species	Type of sea sponge	interaction duration	Removal Efficiency	Conclusion	References
Pyrene (± 100 mg/L)	<i>Bacillus licheniformis</i> strain ATCC 9789 (BI)	<i>Auleta</i> sp.	30 days	±39,00	Performance and biodegradation kinetics increased during the contact period of 10-25 days, then slowed down to day 30	[71]
PAHs (Anthracene and pyrene)	<i>Bacillus pumilus</i> strain GLB197 <i>Pseudomonas stutzeri</i> strain SLG510A3-8	<i>Niphates</i> sp. <i>Hyrtios erectus</i>	25 days	Anthracene (21.89%) Pyrene (7.71 %)	The consortium of three types of bacteria isolated from sea sponges can carry out the function of biodegradation of pyrene and anthracene components, but the performance is less significant, presumably due to competition for carbon as an energy source	[113]
PAHs	<i>Acinetobacter calcoaceticus</i> strain SLFDA 976 <i>Pseudomonas</i> sp. strain Hi1 <i>Bacillus subtilis</i> strain BAB-1684 <i>Pseudomonas stutzeri</i> strain RCH2 <i>Bacillus flexus</i> strain PHCD-20	<i>Clathria (Thalysias) reinwardtii</i> <i>Auleta</i> sp. <i>Clathria reinwardtii</i> <i>Callyspongia</i> sp. <i>Hyrtios erectus</i>	Preliminary test on PAHs contaminated media	observation (qualitative)	All types of sponge symbiont bacteria showed activity on media exposed to PAHs	[78]
Naphthalene	<i>Bacillus</i> sp. <i>Acinetobacter Calcoaceticus</i>	<i>Neopetrosia</i> sp. <i>Callyspongia Aerizusa</i>	25 days	±51.37% ±37.26%	Both types of spongy symbiont bacteria can degrade naphthalene, characterized by several parameters, namely increased acidity of the interaction medium, increased optical density (OD600), smells of fermentation and gas bubbles are formed	[58]
Pyrene	SpAB1 and SpAB2 SpBB1 and SpBB2 SpCB1 and SpCB2	<i>Hyrtios erectus</i> (SpA) <i>Clathria (Thalysias) reinwardtii</i> (SpB) <i>Niphates</i> sp. (SpC)	Preliminary test on pyrene contaminated media	The activity of the two isolates is weak Both isolates did not show activity Both isolates showed strong activity	The activity of isolates against pyrene generally came from sponges whose body surface was covered with mucus. This mucus is thought to have an enzyme character	[76]

	SpDB1 and SpDB2	<i>Callyspongia</i> sp. (SpD)		Both isolates showed moderate activity		
PAHs Naphthalene and Anthracene	Isolate Sp6.B2	<i>Auletta</i> sp.	20 days	There is biodegradation activity	The biodegradation activity of Sp6.B2 isolates against naphthalene and anthracene appeared to be more dominant than Sp8.B1 isolates. GC-MS and FTIR detect new organic compounds of alcohol, aldehyde and carboxylic acid groups	[124]
	Isolate Sp8.B1	<i>Callyspongia Aerizusa</i>				
Aliphatic Components	<i>Bacillus cohnii</i> strain DSM 6307 <i>Bacillus pumilus</i> strain GLB197	<i>Niphates</i> sp.	25 days	Average 48.11%		[31]
petroleum sludge	<i>BacillusFlexus</i> strain PHCDB20.	<i>Callyspongia</i> sp.	35 days	Identified 18 types of aliphatic comp. and 2 aromatic comps.	All hydrocarbon components in the degraded sludge are characterized by a decrease in abundance	[13]

Sea sponges generally have a mutualistic symbiosis with microorganisms, especially bacteria [31,113]. Research on bioremediation of waste containing hydrocarbon components (aliphatic, aromatic) using several types of marine sponge symbiont bacteria shows the ability of these bacteria to degrade hydrocarbon components (Table 2) [12,100]. The search results above show that there are 3 (three) groups of symbiotic sponge bacteria (*Bacillus*, *Pseudomonas* and *Acinetobacter*) [38,52,127]. These bacteria showed biodegradation performance against hydrocarbon components [116]. Research findings related to tracking the remediation performance of sponge symbiont bacteria against pollutants containing hydrocarbon components are that these bacteria are generally isolated from sponges whose body surface is covered with mucus or dark-coloured sponges [73,130]. It has to do with the dynamics experienced by sponges suspected of being exposed to pollutants in their habitat, thus stimulating themselves to survive in that environment by producing mucus substances [13,20,131].

Table 3. Various studies on the bio-adsorption performance of sponge symbiont bacteria against heavy metal contaminants.

Types of hydrocarbon contaminants	Sponge symbiont bacterial species	Type of sea sponge	interaction duration	Removal Efficiency	Conclusion	References
Chromium (VI) Manganese (VII)	<i>Acinetobacter calco-aceticus</i> strain PHCDB14	<i>Callyspongia aerizusa</i>	15 days	±63.21% ±66.80%	Both types of pollutants are absorbed maximum at a contact period of 3 days	[65]
Cr, Zn, Cu, Fe, Co, Mn, Ag and Cd	<i>Bacillus cohnii</i> strains DSM 6307 <i>Pseudomonas stutzeri</i> RCH2	<i>Niphates</i> sp. <i>Clathria (Tha-lysi- as) rein-wardtii</i>	16 days	Heavy metal pollutant removal efficiency varies	All types of heavy metals tested can be absorbed by the symbiont bacteria isolate <i>Niphates</i> sp. and <i>Clathria (Tha-lysi- as) rein-wardtii</i> with varying biosorption performance. Optimum biosorption occurs at a contact period of 4 days	[77]
Cd ²⁺ and As ³⁺	Isolate Sp6.B2 Isolate Sp8.B1	<i>Auletta</i> sp. <i>Callyspongia Aerizusa</i>	20 days	83.19%, and 82.24% 99.89%, and 99.89%	Optimum biosorption occurred at a contact duration of 5 days, then weakened until the 20th day of the contact period	[124]
Cr(VI) and Cd(II)	<i>Bacillus pumilus</i> strain GLB197 <i>Pseudomonas stutzeri</i> strain SLG510A3-8	<i>Niphates</i> sp. <i>Hyrtios erectus</i>	15 days	56.30% and 61.23% 52.74% and 57.80%.	The optimum bioadsorption of these two types of sponge symbiont bacteria against the two types of heavy metal pollutants tested occurred in the range of 3-6 days of contact. Bio-sorption takes place optimally at a contact duration of 3-6 days. Another supporting indicator is the increase in optical density (OD600), gas bubbles detected in the interaction medium	[32]
As ³⁺ and Hg ²⁺	<i>Bacillus licheniformis</i> strain ATCC 9789	<i>Auletta</i> sp.	16 days	99.95%, and 88.49%,		[131]

Similar research has also been carried out to analyze the ability and performance of remediation of sponge symbiont bacteria against heavy metal pollutants (Table 3). The search results provide information that several types of bacteria isolated from sea sponges whose body surface is coated with mucus can also carry out the adsorption function of several kinds of heavy metal pollutants [32,58,77,132]. The bioremediation ability varies, but in general, the sponge symbiont bacteria are efficient and have high performance against heavy metal adsorption [6,25,133].

The adsorption pattern of sponge symbionts on several types of heavy metal pollutants is different from the biodegradation of PAHs, which tends to be directly proportional to the duration of the interaction [6,28,109]. The adsorption of heavy metal pollutants by bacteria occurs extracellularly through the binding of heavy metal ions by extracellular polymers produced by bacterial cells, which act as negatively charged biosorbents

on the cell surface to bind and form complexes with positively charged heavy metal ions [59,127].

Analysis of the bioremediation pattern of marine sponge symbiont bacteria against several kinds of heavy metals showed that the optimal adsorption of heavy metal pollutants by bacteria generally occurred at a contact duration of 2-6 days or when the bacteria had passed the adaptation phase in a new environment exposed to heavy metal toxins [20,130]. Bacterial remediation activity decreased until it reached a contact time of 20 days. This biosorption pattern is illustrated in Figure 3.

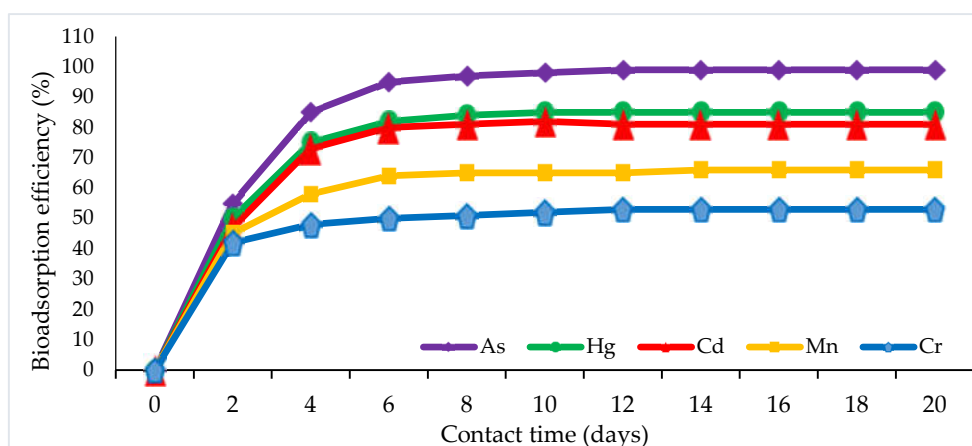


Figure 3. The pattern of adsorption of sponge symbiont bacteria on some heavy metal pollutants

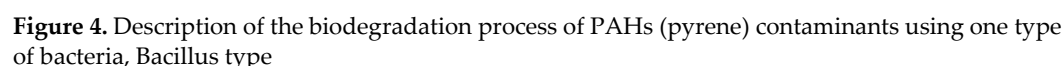
This phenomenon provides information that the adsorption model is an extracellular ionic bond between the positive pole of heavy metals and the negative pole of the bacteria on the surface so that adsorption can occur very quickly when the negative surface of the bacteria is active [60,65,132,134]. The contact duration of 6 days is considered the general period required for the fishery to reach the saturation stage, where most of the negative poles of the bacterial surface have formed ionic bonds with heavy metal ions [35,124]. Under these conditions, the bacterial cells can no longer continue their adsorption activity, and the process towards the division phase and cell growth is declared to have stopped [28,107].

4. Process and mechanism of pollutant bioremediation by marine sponge symbiont bacteria

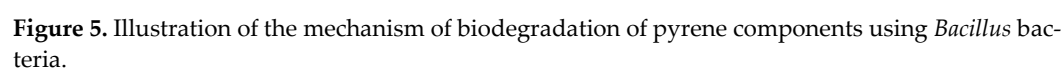
The degradation process of PAH components by bacteria differs from the heavy metal adsorption process [65,131]. The same thing also differs in the degradation mechanism of PAHs with the mechanism of heavy metal adsorption by bacteria as a bioremediator, although using the same bacterial species. Different types of pollutants (PAHs and heavy metals) as a material to remove [16,24,25]. The search and analysis of several types of research on bioremediation using bacteria raise several assumptions that can be scientifically justified [17,42,43].

4.1. Processes and mechanisms of biodegradation of PAHs

The Pyrene biodegradation process using *Bacillus pumilus* strain GLB197 isolated from marine sponge *Niphates* sp. (Table 2) [124] is illustrated in Figure 4. The illustration illustrates that the degradation performance in using one type of bacteria cannot provide significant degradation results, or the degradation takes place incompletely [9,11,31,135].



The process of pyrene degradation by bacillus is similar to the bioremediation mechanism that occurs, and the difference is that the bioremediation mechanism takes place at the molecular level or metabolic substances (micromolecules). In contrast, the remediation process occurs at the component or macromolecular level [11,34,118]. The degradation mechanism of PAHs (pyrene) by bacteria is known as a cycle. Namely, one cycle shows a series of changes in the molecular structure of PAHs from pyrene (4 aromatic benzene rings) to phenanthrene (3 aromatic rings). The following cycle changes phenanthrene to naphthalene (2 aromatic rings), see imaginary line (Fig. 5), and so on until the conversion of benzene (1 aromatic ring) into simple non-aromatic organic compounds that are environmentally friendly is achieved [83,117].



The mechanism of pyrene degradation by *Bacillus pumilus* strain GLB197 [113] is similar to the degradation pathway of PAHs by *Cyclocloasticus* sp. [110,112,116] and adopted

the mechanism of pyrene degradation by *Mycobacterium* sp. [91,99,132], combined with genomic analysis and experimental analysis (Fig. 5). The mechanism of pyrene degradation is described through an oxidation reaction in four stages of change or one cycle, i.e., starting with the change to produce the cis/trans transition product 4,5-dihydrodiol-pyrene (Fig. 5(a)), then converted to 4,5-dihydroxy pyrene (Fig. 5(b)), then to 4,5-dicarboxylic acid-phenanthrene (Fig. 5(c)) and finally to the product phenanthrene-4-carboxylate transition (Fig. 5(d)) [17,46,64]. The fourth stage of the first cycle is the first vulnerable point for bacteria because, at that stage, a carboxylic acid transition product is formed, which causes the acidity of the media to increase [110,112,113]. In this condition, the bacterial cells are susceptible, so the potential cell activity decreases drastically and can even experience mass death [91,138]. The mechanism of conversion of pyrene to the transition product of phenanthrene can be called destruction, namely the destruction of the pyrene molecular structure or an open aromatic ring [117,136].

4.2. Process and mechanism of heavy metal bioadsorption

The process of heavy metal adsorption using bacteria is similar to the process of forming a heavy metal complex (X) with ethylene diamine tetra acetate (EDTA) molecules forming an X-EDTA complex [55,136]. The mechanism of heavy metal adsorption is forming extracellular ionic bonds on the surface of bacterial cells that are negatively charged with positively charged heavy metals [16,118]. The process and mechanism of adsorption of heavy metal ions last for a shorter duration than the degradation of PAHs [25,128]. The adsorption process continues until the saturation point is reached [65'96]. The mechanism and changes in heavy metal adsorption by bacterial cells can be observed using analytical instruments such as SEM, EDS and XRD while determining the adsorption efficiency can use AAS or ICP [125,139,140].

5. Parameters of Pollutant Bioremediation

Several changes can be observed either directly by observation or by using analytical instruments as the performance of microorganisms (bacteria) in the bioremediation process of PAHs and heavy metal pollutants [141]. These changes are indicators and parameters that can be used as the basis for the occurrence of pollutant bioremediation activities by bacteria [27,95].

5.1. Biodegradation of PAHs

The biodegradation parameters of PAHs, which indicate the presence of degradation activity as a performance of bacteria, include: (1) The growth of bacterial cells can be seen from the increase in the optical density of the interaction medium (OD600) [5,53]. The increase in OD600 medium as an indicator of bacterial cells has passed the adaptation phase and heading to the phase of enlargement and division cells [16,33]. At this stage, the degradation activity of PAHs has taken place. (2) The increase in the acidic properties of the media is a manifestation of the work of bacterial cell degradation that has succeeded in destroying the molecular structure of PAHs, forming several new components, one of which is the formation of carboxylic compounds, which results in increased media acidity [110,112]. At this stage, it is called a vulnerable period for bacteria which can result in cells not being able to enlarge and divide, and maybe even the bacterial cells are threatened with mass death [103,134]. (3) The increase in the temperature of the interaction media due to the formation of new compounds resulting from degradation. This parameter has only increased by a few points, generally in the 0.4 – 1.2 °C [23,37]. (4) The emergence of gas bubbles is suspected that this bacterium is an aerobic group that requires oxygen in carrying out PAH remediation. The oxygen demand of the interaction media is carried out by aeration using a shaker or can be injected directly (5). The smell of fermentation is a characteristic feature of the enzymatic reaction that occurs at the stage of destruction of the molecular structure of PAHs [17,46,68]. The existing enzymes are produced by bacteria as a response of cells to defend themselves in extreme environments exposed to PAHs

[66,67] and (6) New peaks were identified on the GC-MS chromatogram [120,121]. The peaks recorded with varying abundance are authentic evidence for all the previously described narratives (points 1-5) [8,126]. (The detection of functional groups of organic compounds from the FTIR chromatogram strengthens the GC-MS data that the degradation products produce organic compounds, one of which is a carboxylic component containing carbonyl and hydroxyl groups [122,142,143].

5.2. Heavy Metal Bioadsorption

Parameters of heavy metal adsorption by bacteria that can be observed and measured include increased media turbidity, a limited change in the pH range of 0.2 - 0.4, temperature increase in a narrow range of 0.3-0.8 °C, gas bubbles appearing, and the smell of fermentation is very weak [3,50]. Changes during the adsorption process are not strong enough to be described in detail as in the biodegradation of PAHs [87,106]. It is because the remediation occurs in the form of adsorption with the extracellular ionic bonding model of the negative part of the bacterial surface to the positive charge of heavy metals [7,37,61]. Therefore, at a certain contact duration, a saturation point can occur in the media, where this saturation point cannot be observed directly. The saturation point is known if the adsorption efficiency is determined by running every two days of contact using the AAS instrument [20,60]. In this case, the saturation point is assumed to occur at the contact period of 6 - 20 days (Fig. 3).

6. Development and Formulation of Remediator Bacteria Consortium

The performance and efficiency of PAHs pollutant biodegradation can be improved by making experimental modifications, especially using consortium bacteria. Similarly, the capacity and level of bacterial adsorption to heavy metal contaminants with the modification of a consortium of bacterial biodegradators [35,92,130].

6.1. Hydrocarbonoclastic bacteria

The application of hydrocarbonoclastic bacteria (R-1) in the biodegradation of PAHs is one of the modifications that is considered to improve the performance and efficiency of bioremediation [34,95]. One of the efforts to improve the performance and efficiency of biodegradation is made by using several types of bacteria, for example, *Bacillus pumilus*, *Pseudomonas stutzeri* and *Acinetobacter calcoaceticus* which are known [9,127], all of which can biodegrade PAHs (hydrocarbonoclastic bacteria) [99,144]. These bacteria were formulated as a consortium bacterial suspension and interacted with pollutant PAHs, e.g. pyrene (Fig.6) [112,145].

This modification is believed to increase the degradability of PAHs because each type of bacteria can take on the role of degradation in parallel so that it can complete one cycle of conversion of pyrene to phenanthrene in a possibly shorter duration of time [71,90]. The study of the potential of the R-1 consortium bacteria was intended to remediate waste containing hydrocarbon components, especially PAHs, with high remediation performance and efficiency [38,145].

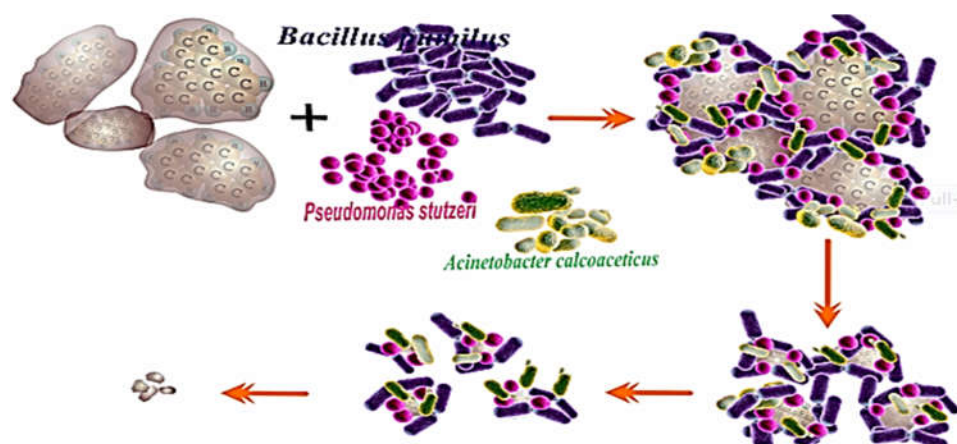


Figure 6. The process of biodegradation of PAH contaminants uses a consortium of hydrocarbonoclastic bacteria. An illustration

The degradability of bacteria was significantly increased, resulting in a high-efficiency degradation. Thus, all pyrene could be degraded to produce the final product in the form of simple non-aromatic organic compounds that are environmentally friendly (Fig. 6) [44,117]. The problem of limiting factors in the PAHs degradation process when it comes to the formation of transition products of carboxylic acid compounds is also assumed to be minimized so that bacterial cells can work continuously to convert carbon elements into energy [82,110].

6.2. Metalloclastic bacteria

Performance, capacity and efficiency of heavy metal adsorption by using several types of bacteria formulated in the form of a consortium of bacteria adsorb heavy metals or bacteria of the metaloclastic category (R-2) [9,131]. The increase in the capacity and efficiency of heavy metal pollutant adsorption occurs due to the abundance of negative poles from the bacterial cell surface, allowing the formation of ionic bonds that occur very much at the surface at almost the same time [28,77].

The difference in adsorption efficiency due to differences in the reactivity and affinity of each heavy metal, as well as the presence of several types of bacterial cells in the consortium formulation, allows the barriers that suppress the abundance of ionic bond formation to be overcome because each type of bacteria has a specific adsorption model [6,20]. The positive charge of each heavy metal also affects the bonds formed. The adaptability factor of bacterial cells to the environment exposed to heavy metals greatly determines the adsorption process. In contrast, the external influence of adsorption, such as the provision of nutrients, and the presence of aeration, does not positively impact the number of ionic bonds formed [35,113]. The consortium of bacteria-coded R-2 has high adsorption power and efficiency, which is intended to be applied in the remediation of waste exposed to heavy metals [77,131].

6.3. Metallo-hydrocarbonoclastic bacteria

In addition to containing aliphatic and aromatic hydrocarbon components, Sludge waste originates from petroleum processing and contains several types of heavy metals [65,96]. This condition requires using bacteria that have a biodegradation function and an adsorption function [60]. Several studies have shown that several types of sponge symbiont bacteria can degrade PAH components and adsorb heavy metals, although these ability tests were carried out separately [6,45,146]. Groups of bacteria with multiple abilities are called Metallo-hydrocarbonoclastic (R-3) bacteria.

Several types of research have been conducted; it was found that several sponge symbiont bacteria can be included in the group of bacteria coded R-3, namely a collection of

bacteria that can biodegrade PAHs and also have the ability to adsorb heavy metals [34,109].

This study concludes the findings in the form of recommendations and suggestions for developing bioremediation technology for GTP, especially PAHs and heavy metals are important to produce and improve remediation efficiency [19,110]. The bacterial consortium of marine sponge symbionts, both hydrocarbonoclastic (R-1), metaloclastic (R-2) or Metallo-hydrocarbonoclastic (R-3) bacteria, have the potential to be applied to increase remediation efficiency in various types of waste [51,81,92]. Screening is important to find and categorize bacteria (R-1; R-2; R-3) with different abilities in GTP remediation [1,7,19]. Bacterial formulation codes R-1, R-2, and R-3 can also be developed in the future into crystalline preparations so that these bacteria are easily mobilized for rapid culture at sites exposed to GTP [2,7,147]. The search for marine sponge symbiont bacteria for bioremediation with high performance and efficiency can be traced only to marine sponges whose body surface is covered with mucus or dark in colour [59,88].

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References

1. Gad, A.K.; Midway, S.R. Relationship of Microplastics to Body Size for Two Estuarine Fishes. *Microplastics* **2022**, *1*, pp. 211–220. <https://doi.org/10.3390/microplastics1010014>.
2. Akoto, O.; Azuure, A.A.; Adotey, K.D. Pesticide residues in water, sediment and fish from Tono Reservoir and their health risk implications. *Springer Plus*, **2016**, *5*, p. 1849. <https://doi.org/10.1186/s40064-016-3544-z>
3. Dimitrov, N.; Kratočil Krehula, L.; Ptiček Siročić, A.; Hrnjak-Murčić, Z. Analysis of recycled PET bottles products by pyrolysis-gas chromatography. *Polym. Degrad. Stab.* **2013**, *98*, pp. 972–979. <https://doi.org/10.1016/j.polymdegradstab.2013.02.013>.
4. Ye, J.; Song, Y.; Liu, Y.; Zhong, Y. Assessment of medical waste generation, associated environmental impact, and management issues after the outbreak of COVID-19: A case study of the Hubei Province in China. *PLoS ONE* **2022**, *17*, pp. 1–17. <http://dx.doi.org/10.1371/journal.pone.0259207>
5. Abraham, N.A.; Offiong, N.A.O.; Ukafia, O.P.; Akpan, P.E. Source Apportionment of Polycyclic Aromatic Hydrocarbons (PAHs) in a Tropical Estuarine Epipelagic Sediment and Its Associated Bacterial Degrading Potentials. *Current Journal of Applied Science and Technology*, **2018**, *32*(1), pp. 1–11. <https://doi.org/10.9734/CJAST/2019/42891>
6. Gran, S.A.; Ramos, Z.J.; Fuentes, E.; Bravo, D.; Pérez, D.J.M. Effect of co-contamination by PAHs and heavy metals on bacterial communities of diesel contaminated soils of south shetland islands, antarctica. *Microorganisms*, **2020**, *8*, pp. 1–17. <https://doi.org/10.3390/microorganisms8111749>

7. Marzuki, I.; Septiningsih, E.; Kaseng, E.S.; Herlinah, H.; Sahrijanna, A.; Sahabuddin, S.; Asaf, R.; Athirah, A.; Isnawan, B.H.; Samidjo, G.S.; Rumagia, F.;; Nisaa, K. Investigation of Global Trends of Pollutants in Marine Ecosystems around Barrang Caddi Island, Spermonde Archipelago Cluster: An Ecological Approach. *Toxics*, **2022**, *10*, p. 301. <https://doi.org/10.3390/toxics10060301>
8. Asgari, A.; Nabizadeh, R.; Mahvi, A.H.; Nasser, S.; Dehghani, M.H.; Nazmara, S.; Yaghmaeian, K. Biodegradation of total petroleum hydrocarbons from acidic sludge produced by re-refinery industries of waste oil using invessel composting. *J Environ Heal Sci Eng*. **2017**, *15*(3), pp. 1–9. <https://doi.org/10.1186/s40201-017-0267-1>
9. Medic', A.; Lješević, M.; Inui, H.; Stojanović, K.; Karadžić, I.; Beškoski, V.; Koji, I. Efficient biodegradation of petroleum: N - alkanes and polycyclic aromatic hydrocarbons by polyextremophilic *Pseudomonas aeruginosa* sanai with multidegradative capacity. *RSC Adv*. **2020**, *10*(24), pp. 14060-14070. <https://doi.org/10.1039/C9RA10371F>
10. Al-Dhabaan, F.A. Morphological, biochemical and molecular identification of petroleum hydrocarbons biodegradation bacteria isolated from oil polluted soil in Dhahran, Saud Arabia. *Saudi J Biol Sci*. **2019**, *26*(6), pp. 1247-1252. <https://doi.org/10.1016/j.sjbs.2018.05.029>
11. Akinde, S.B.; Iwuozor, C.C. Alkane Degradative Potentials of Bacteria Isolated From the Deep Atlantic Ocean of the Gulf of Guinea. *J. of Bioremediation and Biodegradation*, **2012**, *03*(1), pp. 1–6. <https://doi.org/10.4172/2155-6199.1000135>
12. Khabouchi, I.; Khadhar, S.; Chaouachi, D.; Chekirbene, A.; Doumenq, P. Study of organic pollution in superficial sediments of Meliane river catchment area : aliphatic and polycyclic aro-matic hydrocarbons Study of organic pollution in superficial sediments of Meliane river catchment area: aliphatic and polycyclic aromatic hydrocarbons. *Environ Monit Assess*. **2020**, *April*, pp. 282-290. <https://doi.org/10.1007/s10661-020-8213-6>
13. Marzuki, I.; Noor, A.; La-Nafie, N.; Djide, M.N. Sponge Role In Alleviating Oil Pollution Through Sludge Reduction, A Preliminary Approach. *Int J Appl Chem*. **2015**, *11*(4), pp. 427–41.
14. Marzuki, I.; Pratama, I.; Heryani, H.I.; Paserangi, I.; Kamaruddin, M.; Chaerul, M.; Ahmad, R. The Identification and Distribution Components of Polycyclic Aromatic Hydrocarbon Contaminants at the Port of Paotere, Makassar, South Sulawesi. In: The 1st International Conference on Biotechnology and Food Sciences, 11 Sept. 2020, Surabaya, Indonesia, *IOP Conf Series: Earth and Env. Sci*. **2021**. 679, p. 012017. <https://doi.org/10.1088/1755-1315/679/1/012017>
15. Zhang, B.; Zhang, L.; Zhang, X. Bioremediation of petroleum hydrocarbon-contaminated soil by petroleum-degrading bacteria immobilized on biochar. *RSC Adv*., **2019**, *9*(60), pp. 35304-311. <https://doi.org/10.1039/C9RA06726D>
16. Košnář, Z.; Částková, T.; Wiesnerová, L.; Praus, L.; Jablonský, I.; Koudela, M.; Tlustoš, P. Comparing the removal of polycyclic aromatic hydrocarbons in soil after different bioremediation approaches in relation to the extracellular enzyme activities. *J. of Env. Sci*. **2019**, *76*, pp. 249-258. <https://doi.org/10.1016/j.jes.2018.05.007>
17. Sibero, M.T.; Igarashi, Y.; Radjasa, O.K.; Sabdono, A.; Trianto, A.; Zilda, D.S.; Wijaya, Y.J. Sponge-associated fungi from a mangrove habitat in Indonesia: species composition, antimicrobial activity, enzyme screening and bioactive profiling. *Int Aquat Res*. **2019**, *11*(2), pp. 173–86. <https://doi.org/10.1007/s40071-019-0227-8>
18. Bendouz, M.; Dionne, D.; Tran, L.H.; Coudert, L.; Mercier, G.; Blais, J.F. Polycyclic Aromatic Hydrocarbon Oxidation from Concentrates Issued from an Attrition Process of Polluted Soil Using the Fenton Reagent and Permanganate. *Water, Air, and Soil Pollution*. **2017**, *228*(3), pp. 114-127. <https://doi.org/10.1007/s11270-017-3292-x>
19. Bojes, H.K.; Pope, P.G. Characterization of EPA's 16 priority pollutant polycyclic aromatic hydrocarbons (PAHs) in tank bottom solids and associated contaminated soils at oil exploration and production sites in Texas. *Regul Toxicol Pharmacol*. **2007**, *47*(3), pp. 288-95. <https://doi.org/10.1016/j.yrtph.2006.11.007>
20. Abdel-monem, N.M.; Abdel-azeem, A.M.; Ghareeb, D.A.; Nabil-adam, A. Pretreatment Hepatoprotective Effect of the Marine Fungus Derived from Sponge on Hepatic Toxicity Induced by Heavy Metals in Rats. *Biomed Res Int*. **2013**, pp. 1–15. <https://doi.org/10.1155/2013/510879>
21. Baquiran, J.I.P.; Nada, M.A.L.; Posadas, N.; Manogan, D.P.; Cabaitan, P.C.; Conaco, C. Population structure and microbial community diversity of two common tetillid sponges in a tropical reef lagoon. *PeerJ*. **2020**, *4*, pp. 1-25. <https://doi.org/10.7717/peerj.9017>
22. Al-Hawash, A.B.; Dragh, M.A.; Li, S.; Alhujaily, A.; Abbood, H.A.; Zhang, X.; Ma, F. Principles of microbial degradation of petroleum hydrocarbons in the environment. *Egypt. J. Aquat. Res.*, **2018**, *44*(2), pp. 71-76. <https://doi.org/10.1016/j.ejar.2018.06.001>
23. Igiri, B.E.; Okoduwa, S.I.R.; Idoko, G.O.; Akabuogu, E.P.; Adeyi, A.O.; Ejiogu, I.K.; Toxicity and Bioremediation of Heavy Metals Contaminated Ecosystem from Tannery Wastewater: A Review. *J. Toxicol*. **2018**, *2018*, p. 2568038. <https://doi.org/10.1155/2018/2568038>
24. Košnář, Z.; Částková, T.; Wiesnerová, L.; Praus, L.; Jablonský, I.; Koudela, M.; Tlustoš, P. Comparing the removal of polycyclic aromatic hydrocarbons in soil after different bioremediation approaches in relation to the extracellular enzyme activities. *J Environ Sci*. **2018**, *XX*, pp. 1-10. <https://doi.org/10.1016/j.jes.2018.05.007>
25. Zaimee, M.Z.A.; Sarjadi, M.S.; Rahman, M.L. Heavy metals removal from water by efficient adsorbents. *Water*. **2021**, *13*, p. 2659. <https://doi.org/10.3390/w13192659>
26. Bisht, S.; Pandey, P.; Bhargava, B.; Sharma, S.; Kumar, V.; Krishan, D. Bioremediation of polyaromatic hydrocarbons (PAHs) using rhizosphere technology. *Braz. J. Microbiol*. **2015**, *46*, pp. 7–21 <https://doi.org/10.1590/S1517-838246120131354>
27. Dutta, K.; Shityakov, S. New Trends in Bioremediation Technologies Toward Environment-Friendly Society: A Mini-Review. *Front. Bioeng. Biotechnol.*, **2021**, *9*, p. 666858. <https://doi.org/10.3389/fbioe.2021.666858>

28. Khan, M.N.; Ullah, H.; Naeem, S.; Uddin, J.; Hamid, Y.; Ahmad, W.; Ding, J. Remediation of Emerging Heavy Metals from Water Using Natural Adsorbent: Adsorption Performance and Mechanistic Insights. *Sustainability*. **2021**, *13*, p. 8817. <https://doi.org/10.3390/su13168817>
29. Duran, R.; Cravo, -L.C. Role of environmental factors and microorganisms in determining the fate of polycyclic aromatic hydrocarbons in the marine environment. *FEMS Microbiol Rev.*, **2016**, *40*(6), pp. 814-830. <https://doi.org/10.1093/femsre/fuw031>. 814–830
30. Hou, L.; Majumder, E.L.W. Potential for and distribution of enzymatic biodegradation of polystyrene by environmental microorganisms. *Materials (Basel)*, **2021**, *14*(3), pp. 1-20. <https://doi.org/10.3390/ma14030503>
31. Marzuki, I.; Chaerul, M.; Erniati, E.; Asmeati, A.; Paserangi, I. Biodegradation of aliphatic waste components of oil sludge used micro symbiont of Sponge *Niphates* sp. The 3rd International Conference on Marine Science, "Towards Sustainable Marine Resources and Environment" 4 September 2019, Bogor City, Indonesia, *IOP Conf. Ser.: Earth Environ. Sci.* **2020**, *429*, p.012056. <https://doi.org/10.1088/1755-1315/429/1/012056>
32. Marzuki, I.; Daris, L.; Nisaa, K.; Emelda, A. The power of biodegradation and bio-adsorption of bacteria symbiont sponges sea on waste contaminated of polycyclic aromatic hydrocarbons and heavy metals. In: Intern. Conf. on Fisheries and Marine, 13-14 July 2020, North Maluku, Indonesia, *IOP Conf. Ser.: Earth and Env Sci.* **2020**, *584*, p. 012013. <https://doi.org/10.1088/1755-1315/584/1/012013>
33. Nikel, P.I.; Silva-Rocha, R.; Benedetti, I.; De-Lorenzo, V. The private life of environmental bacteria: Pollutant biodegradation at the single cell level. *Environmental Microbiology*, **2014**, *16*(3), pp. 628–642. <https://doi.org/10.1111/1462-2920.12360>
34. Bezza, F.A.; Chirwa, E.M.N. Biosurfactant-enhanced bioremediation of aged polycyclic aromatic hydrocarbons (PAHs) in creosote contaminated soil Fisseha. *Chemosphere*. **2016**, *144*, pp. 635-644. <https://doi.org/10.1016/j.chemosphere.2015.08.027>
35. Kui-Ma, X.; Ling-Wu, L.; Fam, H. Heavy metal ions affecting the removal of polycyclic aromatic hydrocarbons by fungi with heavy-metal resistance. *Appl. Microbiol. Biotechnol.*, **2014**, *98*(23), pp. 9817-27. <https://doi.org/10.1007/s00253-014-5905-2>
36. Yu, Y.; Zhang, Y.; Zhao, N.; Guo, J.; Xu, W.; Ma, M.; Li, X. Remediation of crude oil-polluted soil by the bacterial rhizosphere community of suaeda salsa revealed by 16S rRNA genes. *Int. J. Environ. Res. Public Health*, **2020**, *17*(5), pp. 1-18. <https://doi.org/10.3390/ijerph17051471>
37. Liu, S.H.; Zeng, G.-M.; Niu, Q.-Y.; Liu, Y.; Zhou, L.; Jiang, L.-H.; Tang, Z.-F.; Xu, P.; Zhang, C.; Cheng, M. Bioremediation mechanisms of combined pollution of PAHs and heavy metals by bacteria and fungi: A mini review. *Bioresour. Technol.*, **2017**, *224*, pp. 25-33. <https://doi.org/10.1016/j.biortech.2016.11.095>
38. Elenga-Wilson, P.S.; Kayath, C.A.; Mokemiabeka, N.S.; Nzaou, S.A.E.; Nguimbi, E.; Ahombo, G. Profiling of Indigenous Biosurfactant-Producing Bacillus Isolates in the Bioremediation of Soil Contaminated by Petroleum Products and Olive Oil. *Int. J. Microbiol. Hindawi*, **2021**, *221*, p. 9565930. <https://doi.org/10.1155/2021/9565930>
39. Essumang, D.K.; Togoh, G.K.; Chokky, L. Pesticide residues in the water and Fish (lagoon tilapia) samples from lagoons in Ghana. *Bull. Chem. Soc. Ethiop.* **2009**, *23*, pp. 19–27. <https://doi.org/10.4314/bcse.v23i1.21294>
40. Li, Q.; Liu, J.; Gadd, G.M. Fungal bioremediation of soil co-contaminated with petroleum hydrocarbons and toxic metals. *Appl. Microbiol. Biotechnol.*, **2020**, *104*(21), pp. 8999-9008. <https://doi.org/10.1007/s00253-020-10854-y>
41. Medaura, M.C.; Guivernau, M.; Moreno-Ventas, X.; Prenafeta-Boldú, F.X.; Viñas, M. Bioaugmentation of Native Fungi, an Efficient Strategy for the Bioremediation of an Aged Industrially Polluted Soil With Heavy Hydrocarbons. *Front. Microbiol.* **2021**, *12*, pp. 1–18. <https://doi.org/10.3389/fmicb.2021.626436>
42. Yetti, E.; Thontowi, A.; Yopi, Y.; Lisdiyanti, P. Screening of Marine bacteria capable of degrading various polyaromatic hydrocarbons. *Squalen Bull Mar Fish Postharvest Biotechnol.* **2015**, *10*(3), pp. 121-127. <https://doi.org/10.15578/squalen.v10i3.123>
43. Kamaruddin, M.; Marzuki, I.; Burhan, A.; Ahmad, R. Screening acetylcholinesterase inhibitors from marine-derived actinomycetes by simple chromatography. In Proceedings of the 1st International Conference on Biotechnology and Food Sciences, 2021, 11 September 2020, Surabaya, Indonesia, *IOP Conf Ser. Earth Env. Sci.* **2021**, *679*, p. 012011. <https://doi.org/10.1088/1755-1315/679/1/012011>
44. Agrawal, N.; Verma, P.; Shahi, S.K. Degradation of polycyclic aromatic hydrocarbons (phenanthrene and pyrene) by the ligninolytic fungi *Ganoderma lucidum* isolated from the hardwood stump. *Bioresour. Bioprocess.* **2018**, *5*, p. 11, <https://doi.org/10.1186/s40643-018-0197-5>.
45. Atagana, H.I. Biodegradation of PAHs by fungi in contaminated-soil containing cadmium and nickel ions. *Afr. J. Biotechnol.* **2009**, *8*, pp. 5780–5789. <https://doi.org/10.5897/AJB2009.000-9465>.
46. Cao, H.; Wang, C.; Liu, H.; Jia, W.; Sun, H. Enzyme activities during Benzo[a]pyrene degradation by the fungus *Lasiodiplodia theobromae* isolated from a polluted soil. *Sci. Rep.* **2020**, *10*(1), pp. 1–11. <https://doi.org/10.1038/s41598-020-57692-6>.
47. Armus, R.; Selry, C.; Marzuki, I.; Hasan, H.; Syamsia; Sapar, A. Investigation of Potential Marine Bacterial Isolates in Biodegradation Methods on Hydrocarbon Contamination. 2nd Workshop on Engineering, Education, Applied Sciences and Technology (WEAST) Oct. 5, 2020. Makassar, Indonesia. *IOP Publishing, J. of Physics: Conf. Series*, **2021**, *1899*, p. 012006 <https://doi.org/doi:10.1088/1742-6596/1899/1/012006>
48. Obire, O.; Aleruchi, O.; Wemedo, S. Fungi in Biodegradation of Polycyclic Aromatic Hydrocarbons in Oilfield Wastewater. *Acta Sci Microbiol.* **2020**, *3*(4), pp. 220-224. <https://doi.org/10.31080/ASML.2020.03.0572>
49. Omoni, V.T.; Lag-B.A.J.; Ibeto, C.N.; Semple, K.T. Effects of biological pre-treatment of lignocellulosic waste with white-rot fungi on the stimulation of 14C-phenanthrene catabolism in soils. *Int. Biodeterior Biodegrad*, **2021**, *165*, p. 105324. Available from: <https://doi.org/10.1016/j.ibiod.2021.105324>

50. Saraswath, A.; Hallberg, R. Degradation of pyrene by indigenous fungi from a former gasworks site. *FEMS Microbiol. Lett.* **2002**, *210*, pp. 227–232. <https://doi.org/10.1111/j.1574-6968.2002.tb11185.x>.
51. Nedoroda, V.; Trokhymenko, G.; Khrapko, T.; Koliehova, A. Analysis of Petroleum Biodegradation by a Bacterial Consortium of *Bacillus amyloliquefaciens* ssp. *Plantarum* and *Bacillus subtilis*. *J. Ecol. Eng.*, **2021**, *22*(11), pp. 36–42. <https://doi.org/10.12911/22998993/143017>
52. Bello-, A.M.; Adeleke, R.; Swanevelde, D.; Thantsha, M. Draft Genome Sequence of *Pseudomonas* sp. Strain 10-1B, a Polycyclic Aromatic Hydrocarbon Degradator in Contaminated Soil. *Genome Announc*, **2015**, *3*(3), p. e00325-15. <https://doi.org/10.1128/genomeA.00325-15>.
53. Brzeszcz, J.; Kaszycki, P. Aerobic bacteria degrading both n-alkanes and aromatic hydrocarbons: an undervalued strategy for metabolic diversity and flexibility. *Biodegradation*, **2018**, *29*(4), pp. 359–407. <https://doi.org/10.1007/s10532-018-9837-x>
54. Crampon, M.; Cébron, A.; Portet-Koltalo, F.; Uroz, S.; Le Derf, F.; Bodilis, J. Low effect of phenanthrene bioaccessibility on its biodegradation in di_ensely contaminated soil. *Environ. Pollut.* **2017**, *225*, pp. 663–673. <https://doi.org/10.1016/j.envpol.2017.03.053>
55. Dadrasnia, A.; Usman, M.M.; Lim, K.T.; Farahiyah, F.H.; Rodzhan, N.Sb.M.; Karim, S.H.A.; Ismail, S. Bio-Enhancement of Petroleum Hydrocarbon Polluted Soil Using Newly Isolated Bacteria. *Polycyclic Aromatic Compounds*, **2020**, *40*, pp. 484–493. <https://doi.org/10.1080/10406638.2018.1454966>
56. Guo, J.; Wen, X. Performance and kinetics of benzo(a)pyrene biodegradation in contaminated water and soil and improvement of soil properties by biosurfactant amendment. *Ecotoxicol Environ Saf.* **2021**, *207*, p. 111292. <https://doi.org/10.1016/j.ecoenv.2020.111292>
57. Hermabessiere, L.; Himber, C.; Boricaud, B.; Kazour, M.; Amara, R.; Cassone, A.-L.; Laurentie, M.; Paul-Pont, I.; Soudant, P.; Dehaut, A.; Duflos, G. Optimization, performance, and application of a pyrolysis-GC/MS method for the identification of microplastics. *Anal. Bioanal. Chem.* **2018**, *410*, pp. 6663–6676. <https://doi.org/10.1007/s00216-018-1279-0>.
58. Marzuki, I.; Sinardi, S.; Pratama, I.; Chaerul, M.; Paserangi, I.; Kamaruddin, M.; Performance of sea sponges micro symbionts as a biomaterial in biodegradation naphthalene waste of modified. In The 5th International Seminar on Sustainable Urban Development 5 August 2020, Jakarta, Indonesia, *IOP Conf Ser: Earth and Env Sci*, **2021**, *737*, p. 012016. <https://doi.org/10.1088/1755-1315/737/1/012016>
59. Freeman, C.J.; Easson, C.G.; Fiore, C.L.; Thacker, R.W. Sponge–Microbe Interactions on Coral Reefs: Multiple Evolutionary Solutions to a Complex Environment. *Front. Mar. Sci.* **2021**, *8*, pp. 1–24. <https://doi.org/10.3389/fmars.2021.705053>.
60. Zahirnejad, M.; Ziarati, P.; Asgarpanah, J. The Efficiency of Bio-adsorption of Heavy Metals from Pharmaceutical Effluent by *Rumex crispus* L. Seed. *J Pharm Heal Sci.* **2017**, *5*(3), pp. 231–43. https://journals.iau.ir/article_535243.html
61. Abass, O.K.; Zhuo, M.; Zhang, K. Concomitant degradation of complex organics and metals recovery from fracking wastewater: Roles of nano zerovalent iron. *Chem. Eng. J.* **2017**, *328*, pp. 159–171. <https://doi.org/10.1016/j.cej.2017.07.030>.
62. Lavy, A.; Keren, R.; Haber, M.; Schwartz, I.; Ilan, M. Implementing sponge physiological and genomic information to enhance the diversity of its culturable associated bacteria. *FEMS Microbiol Ecol.* **2014**, *87*(2), pp. 486–502. <https://doi.org/10.1111/1574-6941.12240>.
63. Maldonado, M.; López, -A.M.; Busch, K.; Slaby, B.M.; Bayer, K.; Beazley, L.; Hentschel, U.; Kenchington, E.; Rapp, H.T. A Microbial Nitrogen Engine Modulated by Bacteriosyncytia in Hexactinellid Sponges: Ecological Implications for Deep-Sea Communities. *Front Mar Sci.* **2021**, *8*, pp. 1–35. <https://doi.org/10.3389/fmars.2021.638505>
64. Marzuki, I.; Enryani, H.I.; Nafie, N.-L.; Dali, S.; Study Biodegradation of Aromatics Pyrene Using Bacterial Isolates from the Sea and micro symbionts Sponges. *Int J Appl Chem.* **2017**, *13*(3), 707–20.
65. Marzuki, I. The Bio-adsorption Pattern Bacteria Symbiont Sponge Marine Against Contaminants Chromium and Manganese In The Waste Modification of Laboratory Scale. *Indo Chim Acta.* **2020**, *13*(1), pp. 1–9. <https://doi.org/10.20956/ica.v13i1.9972>
66. Cajthaml, T.; Erbanová, P.; Kollmann, A.; Novotný, C.; Sasek, V.; Mougin, C. Degradation of PAHs by ligninolytic enzymes of *Irpex lacteus*. *Folia Microbiol (Praha).* **2008**, *53*(4), pp. 289–94. <https://doi.org/10.1007/s12223-008-0045-7>.
67. Kadri, T.; Rouissi, T.; Brar, S.K.; Cledon, M.; Sarma, S.; Verma, M. Biodegradation of polycyclic aromatic hydrocarbons (PAHs) by fungal enzymes: A review. *J. of env. Sci.*, **2017**, *51*, pp. 52–74. <http://dx.doi.org/10.1016/j.jes.2016.08.023>
68. Wang, M.; Zhang, X.; Huang, Y.; Zhou, F.; Zhao, Q.; Zhang, W.; Chen, G.; Gao, Y. Isolation of Petroleum Degradators and Petroleum-Degradation Characteristics of Crude Enzymes from *Providencia rettgeri* L1. *Polish J. Environ Studies.* **2022**, *31*(4), pp. 1–8. <https://doi.org/10.15244/pjoes/146467>
69. Ohowa, B.; Kiteresi, L.I.; Wanjeri, V.W.; Mwamburi, S.M.; Tunje, S.L. Sponges as simple biomonitoring tools for trace element pollution in marine environments: insights from a Kenyan study focused on the leaf sponge *Phyllospongia foliascens*. *African J. of Marine Sci.* **2021**, *43*(4), pp. 533–538. <https://doi.org/10.2989/1814232X.2021.1989487>
70. Marzuki, I.; Gusty, S.; Armus, R.; Sapar, A.; Asaf, R.; Athirah, A.; Jaya, J. Secondary Metabolite Analysis and Anti-Bacteria and Fungal Activities of Marine Sponge Methanol Extract Based on Coral Cover. In: *The 6th International Conference on Basic Sciences*, 4–5 Nov.2020, Ambon City, Indonesia. *IAP Conf Proc.* **2021**, *2360*(1), p. 1–9. <https://doi.org/10.1063/5.0059500>
71. Marzuki, I.; Nisaa, K.; Asaf, R.; Paena, M.; Athirah, A.; Susianingsih, E.; Nurhidayah, N.; Kadriah, I.A.K.; Kamaruddin, K.; Sahabuddin, S.; Nurbaya, N.; Septiningsih, E Herlinah, H.; Hendrajat, E.A.; Suwardi, S.; Ramlan, A. Comparison of Pyrene Biodegradation Using Two Types of Marine Bacterial Isolates. *Sustainability*, **2022**, *14*(16), p. 9890. <https://doi.org/10.3390/su14169890>
72. Rua, C.P.J.; Oliveira, L.S.; De-Froes, A.; Tschoeke, D.A.; Soares, A.C.; Leomil, L.; Gregoracci, G.B.; Coutinho, R.; Hajdu, E.; Thompson, C.C.; Berlinck, R.G.S.; Thompson, F.L. Microbial and Functional Biodiversity Patterns in Sponges that Accumulate

- Bromopyrrole Alkaloids Suggest Horizontal Gene Transfer of Halogenase Genes. *Microb. Ecol. J.* **2018**, *76*, pp. 825–838. <https://doi.org/10.1007/s00248-018-1172-6>.
73. Marzuki, I.; Daris, L.; Yunus, S.; Riana, A.D. Selection and characterization of potential bacteria for polycyclic aromatic biodegradation of hydrocarbons in sea sponges from Spermonde Islands, Indonesia. *AACL Bioflux.* **2020**, *13*(6), pp. 3493–506.
 74. Kim, H.-W.; Jo, J.H.; Kim, Ye.-B.; Le, T.K.; Cho, C.-W.; Yun, C.-H.; Chi, W.S.; Yeom, S.-J. et al. Biodegradation of polystyrene by bacteria from the soil in common environments. *J. Hazard. Mater.*, **2021**, *416*, p. 126239. <https://doi.org/10.1016/j.jhazmat.2021.126239>
 75. Fu, X.; Wang, H.; Bai, Y.; Xue, J.; Gao, Y.; Hu, S.; Wu, T.; Sum, J. Systematic degradation mechanism and pathways analysis of the immobilized bacteria: Permeability and biodegradation, kinetic and molecular simulation. *Environ Sci Ecotechnology.* **2020**, *2*, p. 100028. <https://doi.org/10.1016/j.esec.2020.100028>
 76. Marzuki, I.; Kamaruddin, M.; Ahmad, R. Identification of marine sponges-symbiotic bacteria and their application in degrading polycyclic aromatic hydrocarbons. *Biodiversitas*, **2021**, *22*(3), pp. 1481–88. <https://doi.org/10.13057/biodiv/d220352>
 77. Marzuki, I.; Ahmad, R.; Kamaruddin, M.; Asaf, R.; Armus, R.; Siswanti, I. Performance of cultured marine sponges-symbiotic bacteria as a heavy metal bio-adsorption. *Biodiversitas*. **2021**, *22*(12), pp. 5536–43. <https://doi.org/10.13057/biodiv/d221237>
 78. Marzuki, I.; Ali, M.Y.; Syarif, H.U.; Erniati.E.; Gusty, S.; Ritnawati, R.; Daris, L.; Nisaa, K. Investigation of Biodegradable Bacteria as Bio indicators of the Presence of PAHs Contaminants in Marine Waters in the Marine Tourism Area of Makassar City. In: 6th International Conference on Tropical Coastal Region Eco-Development, 27-28 Oct. 2020, Indonesia, *IOP Conf Ser: Earth and Env. Sci.* **2021**, *750*, p. 012006. <https://doi.org/10.1088/1755-1315/750/1/012006>.
 79. Liu, Y.F.; Mbadinga, S.M.; Gu, J.D.; Mu, B.Z. Type II chaperonin gene as a complementary barcode for 16S rRNA gene in study of Archaea diversity of petroleum reservoirs. *Int Biodeterior Biodegrad.* **2017**, *123*, pp. 113–20. <http://dx.doi.org/10.1016/j.ibiod.2017.04.015>
 80. Su, X.M.; Bamba, A.M.; Zhang, S.; Zhang, Y.G.; Hashmi, M.Z.; Lin, H.J.; Ding, L.X. Revealing potential functions of VBNC bacteria in polycyclic aromatic hydrocarbons biodegradation. *Lett Appl Microbiol.* **2018**, *66*(4), pp. 277–283. <https://doi.org/10.1111/lam.12853>; P.Mid: 29350767
 81. Syakti, A.D.; Yani, M.; Hidayati, N.V.; Siregar, A.S.; Doumenq, P.; Sudiana, I.M. The bioremediation potential of hydrocarbon-oclastic bacteria isolated from a mangrove contaminated by petroleum hydrocarbons on the cilacap coast, Indonesia. *Bioremediat J.* **2013**, *17*(1), pp. 11–20. <https://doi.org/10.1080/10889868.2012.731446>
 82. Ahmad, M.; Wang, P.; Li, J.L.; Wang, R.; Duan, L.; Luo, X.; Irfan, M.; Peng, Z.; Yin, L.; Li, W.-J. Impacts of bio-stimulants on pyrene degradation, prokaryotic community compositions, and functions. *Environ. Pollut.* **2021**, *289*, p. 117863. <https://doi.org/10.1016/j.envpol.2021.117863>.
 83. Fang, H.; Shi, Y.; Zhou, M.; Niu, Q. Influence of n-Hexadecane and Naphthalene on Anaerobic Digestion: Kinetic Simulation, DOM Variation and Microbial Community Assessment. *GEESD, IOP, Conf. Series: Earth and Environmental Science*, **2020**, *555*, p. 012038. <https://doi.org/10.1088/1755-1315/555/1/012038>
 84. Galitskaya, P.; Biktasheva, L.; Blagodatsky, S.; Selivanovskaya, S. Response of bacterial and fungal communities to high petroleum pollution in different soils. *Sci. Rep.*, **2021**, *11*(1), pp. 1–18. <https://doi.org/10.1038/s41598-020-80631-4>
 85. Lu, L.; Zhang, J.; Peng, C. Shift of Soil Polycyclic Aromatic Hydrocarbons (PAHs) dissipation pattern and microbial community composition due to rhamnolipid supplementation. *Water Air Soil Pollut.* **2019**, *230*, p.107. <https://doi.org/10.1007/s11270-019-4118-9>
 86. Mao, J.; Guan, W.; Fungal degradation of polycyclic aromatic hydrocarbons (PAHs) by *Scopulariopsis brevicaulis* and its application in bioremediation of PAH-contaminated soil. *Acta Agric. Scand Sect. B Soil Plant Sci.* **2016**, *66*(5), pp. 399–405. <https://doi.org/10.1080/09064710.2015.1137629>.
 87. Sandhu, M.; Paul, A.T. Metagenomic Analysis for Taxonomic and Functional Potential of Polyaromatic Hydrocarbons (PAHs) and Polychlorinated Biphenyl (PCB) Degrading Bacterial Communities in Steel Industrial Soil. *PLoS ON*, *17*(4), p. e0266808. <https://doi.org/10.1371/journal.pone.0266808>
 88. Tziveleka, L.-A.; Ioannou, E.; Tsiourvas, D.; Berillis, P.; Foufa, E.; Roussis, V. Collagen from the marine sponges *Axinella canabina* and *Suberites carnosus*: Isolation and morphological, biochemical, and biophysical characterization. *Mar Drugs*. **2017**, *15*(6), p. 152. <https://doi.org/10.3390/md15060152>
 89. Marzuki, I.; Nisaa, K.; Asaf, R.; Armus, R.; Kamaruddin, M.; Sapor, A.; Emelda, A. Biodegradation mechanism of naphthalene using marine sponge symbiotic bacteria. In 2nd International Conference on Fisheries and Marine 15 July 2021, Khairun University, Ternate, Indonesia, *IOP Conf Ser Earth Env Sci.* **2021**, *890*, p. 012020. <https://doi.org/10.1088/1755-1315/890/1/012020>
 90. Chulalaksananukul, S.; Gadd, G.M.; Sangvanich, P.; Sihanonth, P.; Piapukiew, J.; Vangnai, A.S. Biodegradation of benzo(a)pyrene by a newly isolated *Fusarium* sp. *FEMS Microbiol. Lett.* **2006**, *262*, pp. 99–106. <https://doi.org/10.1111/j.1574-6968.2006.00375.x>.
 91. Govarthanan, M.; Fuzisawa, S.; Hosogai, T.; Chang, Y.C. Biodegradation of aliphatic and aromatic hydrocarbons using the filamentous fungus *Penicillium* sp. CHY-2 and characterization of its manganese peroxidase activity. *RSC Adv.*, **2017**, *7*(34), pp. 20716–23. <https://doi.org/10.1039/C6RA28687A>
 92. Laothamteep, N.; Kawano, H.; Vejarano, F.; Minakuchi, C.S.; Shintani, M.; Nojiri, H.; Pinyakong, O. Effects of environmental factors and coexisting substrates on PAH degradation and transcriptomic responses of the defined bacterial consortium OPK. *Environ Pollut.* **2021**, *277*, p. 116769. <https://doi.org/10.1016/j.envpol.2021.116769>
 93. Fu, W.; Xu, M.; Sun, K.; Cao, L.H.; Dai, C.; Jia, Y. Biodegradation of phenanthrene by endophytic fungus *Phomopsis liquidambari* in vitro and in vivo. *Chemosphere* **2018**, *203*, pp. 160–169. <https://doi.org/10.1016/j.chemosphere.2018.03.164>.

94. Haleyur, N.; Shahsavari, E.; Jain, S.S.; Koshlaf, E.; Ravindran, V.B.; Morrison, P.D.; Osborn, A.M.; Ball, A.S. Influence of bioaugmentation and biostimulation on PAH degradation in aged contaminated soils: Response and dynamics of the bacterial community. *J. Environ. Manag.* **2019**, *238*, pp. 49–58. <https://doi.org/10.1016/j.jenvman.2019.02.115>
95. Guerra, A.B.; Oliveira, J.S.; Silva-Portela, R.C.B.; Araújo, W.; Carlos, A.C.; Vasconcelos, A.T.R.; Freitas, A.T.; Domingos, Y.S.; de Farias, M.F.; Fernandes, G.J.T.; et al. Metagenome enrichment approach used for selection of oil-degrading bacteria consortia for drill cutting residue bioremediation. *Environ. Pollut.* **2018**, *235*, pp. 869–880. <https://doi.org/10.1016/j.envpol.2018.01.014>
96. Ja'nczuk, B.; Szymczyk, K.; Zdziennicka, A. Adsorption Properties of Hydrocarbon and Fluorocarbon Surfactants Ternary Mixture at the Water-Air Interface. *Molecules*, **2021**, *26*, p. 4313. <https://doi.org/10.3390/molecules26144313>
97. Lasota, J.; Łyszczarz, Y.; Kempf, P.; Kempf, M.; Błońska, E. Effect of Species Composition on Polycyclic Aromatic Hydrocarbon (PAH) Accumulation in Urban Forest Soils of Krakow. *Water. Air. Soil Pollut.*, **2021**, *232*(2), p. 74. <https://doi.org/10.1007/s11270-021-05043-0>
98. Lu, C.; Hong, Y.; Liu, J.; Gao, Y.; Ma, Z.; Yang B.; Ling, W.; Waigi, M.G. A PAH-degrading bacterial community enriched with contaminated agricultural soil and its utility for microbial bioremediation. *Environmental Pollution*, **2019**, *251*, pp. 773–782. <https://doi.org/10.1016/j.envpol.2019.05.044>
99. Lundstedt, S. *Analysis of PAHs and their transformation products in contaminated soil and remedial processes*. Netherlands: Akademisk avhandling Som, Department of Env. and Toxicological Chemistry, **2003**, University of Amsterdam. <http://www.diva-portal.org/smash/record.jsf?pid=diva2%3A143820&dsid=3258>
100. Nzila, A. Current status of the degradation of aliphatic and aromatic petroleum hydrocarbons by thermophilic microbes and future perspectives. *Int. J. Environ. Res. Public Health*, **2018**, *15*(12), pp. 1–23. <https://doi.org/10.3390/ijerph15122782>
101. Pandey, P.; Kapley, A.; Brar, S.K. Editorial: Biodegradation of High Molecular Weight Polyaromatic Hydrocarbons in Different Environments. *Front. Microbiol.*, **2021**, *12*, pp. 2020–2022. <https://doi.org/10.3389/fmicb.2021.704897>
102. Sawulski, P.; Boots, B.; Clipson, N.; Doyle, E. Differential degradation of polycyclic aromatic hydrocarbon mixtures by indigenous microbial assemblages in soil. *Lett Appl Microbiol.* **2015**, *61*(2), pp. 199–207. <https://doi.org/10.1111/lam.12446>
103. Scheuch, G.A.; Zuñiga, J.R.; Fuentes, E.; Bravo, D.; Donoso, J.M.P. Effect of co-contamination by PAHs and heavy metals on bacterial communities of diesel contaminated soils of south shetland islands, antarctica. *Microorganisms*. **2020**, *8*(11), pp. 1–17. <https://doi.org/10.3390/microorganisms8111749>
104. Shehu, U.; Ahmad, F.A.; Yusuf, F.; Muhammad, F.; Yakasai, H.M. Isolation and Identification of Anthracene Utilizing *Proteus vulgaris* from Oil Spill Contaminated Soil at NNPC Depot Kano State Nigeria. *J. Adv. Biol. Biotechnol.* **2021**, *24*, pp. 46–53. <https://doi.org/10.9734/jabb/2021/v24i1030246>
105. Smulek, W.; Sydow, M.; Matejuk, Z.J.; Kaczorek, E. Bacteria involved in biodegradation of creosote PAH – A case study of longterm contaminated industrial area. *Ecotoxicol Environ Saf.* **2020**, *187*, p. 109843. <https://doi.org/10.1016/j.ecoenv.2019.109843>
106. Souza, H.M.de-L.; Barreto, L.R.; da Mota, A.J.; de Oliveira, L.A.; Barroso, H.dos-S.; Zanutto, S.P. Tolerance to polycyclic aromatic hydrocarbons (PAHs) by filamentous fungi isolated from contaminated sediment in the Amazon region. *Acta Sci. - Biol. Sci.*, **2017**, *39*(4), pp. 481–488. <https://doi.org/10.4025/actascibiols.v39i4.34709>
107. Ray, M.; Kumar, V.; Banerjee, C.; Gupta, P.; Singh, S.; Singh, A. Investigation of biosurfactants produced by three indigenous bacterial strains, their growth kinetics and their anthracene and fluorene tolerance. *Ecotoxicol Environ Saf.* **2021**, *208*, p. 111621. <https://doi.org/10.1016/j.ecoenv.2020.111621>
108. Sahu, L. Presence of Hydrocarbon Degrading Bacteria in Contaminated Soil Collected From Various Fuel Station in Bhilai, Chhattisgarh. *Int. J. Res. Appl. Sci. Eng. Technol.*, **2021**, *9*(11), pp. 1802–1804. <https://doi.org/10.22214/ijraset.2021.39107>
109. Vaezzadeh, V.; Zakaria, M.P.; Bong, C.W.; Masood, N.; Magam, M.S.; Alkhadher, S. Mangrove Oyster (*Crassostrea belcheri*) as a Biomonitor Species for Bioavailability of Polycyclic Aromatic Hydrocarbons (PAHs) from Sediment of the West Coast of Peninsular Malaysia. *Polycycl Aromat Compd.* **2019**, *39*(5), pp. 470–85. <https://doi.org/10.1080/10406638.2017.1348366>
110. Wang, B.; Lai, Q.; Cui, Z.; Tan, T.; Shao, Z. A pyrene-degrading consortium from deep-sea sediment of the West Pacific and its key member *Cycloclasticus* sp. P1. *Environ Microbiol J.* **2008**, *10*(8), pp. 1948–63. <https://doi.org/10.1111/j.1462-2920.2008.01611.x>
111. Wolf, D.C.; Gan, J. Influence of rhamnolipid biosurfactant and Brij-35 synthetic surfactant on ¹⁴C-pyrene mineralization in soil. *Environ. Pollut.* **2018**, *243*, pp. 1846–1853. <https://doi.org/10.1016/j.envpol.2018.10.031>
112. Wang, W.; Wang, L.; Shao, Z. Polycyclic Aromatic Hydrocarbon (PAH) Degradation Pathways of the Obligate Marine PAH Degrader *Cycloclasticus* sp. Strain P1. *App. and Env. Microbiology*, **2018**, *84*, (21), pp. 1261–18. <https://doi.org/10.1128/AEM.01261-18>
113. Marzuki, I.; Asaf, R.; Paena, M.; Athirah, A.; Nisaa, K.; Ahmad, R.; Kamaruddin, M. Anthracene and Pyrene Biodegradation Performance of Marine Sponge Symbiont Bacteria Consortium. *Molecules*. **2021**, *26*(22), p.6851. <https://doi.org/10.3390/molecules26226851>
114. Zhang, Y.; Qin, F.; Qiao, J.; Li, G.; Shen, C.; Huang, T.; Hu, Z. Draft Genome Sequence of *Rhodococcus* sp. Strain P14, a Biodegrader of High-Molecular-Weight Polycyclic Aromatic Hydrocarbons. *J Bacteriol.* **2012**, *194*(13), p. 3546. <https://doi.org/10.1128/JB.00555-12>
115. Vila, J.; Tauler, M.; Grifoll, M. Bacterial PAH degradation in marine and terrestrial habitats. *Curr Opin Biotechnol.* **2015**, *33*, pp. 95–102. <https://doi.org/10.1016/j.copbio.2015.01.006>
116. Tawniczak, T.; Woźniak-Karczewska, M.; Loibner, A.P.; Heipieper, H.J.; Chrzanowski, T. Microbial degradation of hydrocarbons-basic principles for bioremediation: A review. *Molecules*, **2020**, *25*(4), pp. 1–19. <https://doi.org/10.3390/molecules25040856>
117. Miao, L.L.; Qu, J.; Liu, Z.P. Hydroxylation at Multiple Positions Initiated the Biodegradation of Indeno[1,2,3-cd]Pyrene in *Rhodococcus aetherivorans* IcdP1. *Front Microbiol.* **2020**, *11*, pp. 1–13. <https://doi.org/10.3389/fmicb.2020.568381>

118. Bodor, A.; Bounedjouma, N.; Feigl, G.; Duzs, A.; Laczi, K.; Szilágyi, A.; Rákhely, G.; Perei, K. Exploitation of extracellular organic matter from *Micrococcus luteus* to enhance ex situ bioremediation of soils polluted with used lubricants. *J. Hazard. Mater.*, **2021**, *417*, p. 125996. <https://doi.org/10.1016/j.jhazmat.2021.125996>
119. Zakaria, H.Y.; Hassan, A.K.M.; El-Naggar, H.A.; Abo-Senna, F.M. Biomass determination based on the individual volume of the dominant copepod species in the Western Egyptian Mediterranean Coast. *Egypt J Aquat Res*, **2018**, *44*(2), pp. 89–99. <https://doi.org/10.1016/j.ejar.2018.05.002>
120. Çoban-Yıldız, Y.; Chiavari, G.; Fabbri, D.; Gaines, A.F.; Galletti, G.; Tuğrul, S. The chemical composition of Black Sea suspended particulate organic matter: Pyrolysis-GC/MS as a complementary tool to traditional oceanographic analyses. *Mar. Chem.* **2000**, *69*, pp. 55–67. [https://doi.org/10.1016/S0304-4203\(99\)00093-6](https://doi.org/10.1016/S0304-4203(99)00093-6)
121. Gomiero, A.; Øysæd, K.B.; Palmas, L.; Skogerbø, G. Application of GC/MS-pyrolysis to estimate the levels of microplastics in a drinking water supply system. *J. Hazard. Mater.* **2021**, *416*, p. 125708. <https://doi.org/10.1016/j.jhazmat.2021.125708>
122. Käßler, A.; Fischer, M.; Scholz-Böttcher, B.M.; Oberbeckmann, S.; Labrenz, M.; Fischer, D.; Eichhorn, K.-J.; Voit, B. Comparison of μ -ATR-FTIR spectroscopy and py-GCMS as identification tools for microplastic particles and fibers isolated from river sediments. *Anal. Bioanal. Chem.* **2018**, *410*, pp. 5313–5327. <https://doi.org/10.1007/s00216-018-1185-5>
123. Spini, G.; Spina, F.; Poli, A.; Bliieux, A.-L.; Regnier, T.; Gramellini, C.; Varese, G.C.; Puglisi, E. Molecular and Microbiological Insights on the Enrichment Procedures for the Isolation of Petroleum Degrading Bacteria and Fungi. *Front. Microbiol.*, **2018**, *9*, p. 2543. <https://doi.org/10.3389/fmicb.2018.02543>
124. Marzuki, I.; Asdar, A.M.; Hardimas, H. Performance Analysis of bio-sorption of Heavy Metal and Biodegradation PAH of Isolates Marine Sponges Symbiont Bacteria. *Indo Chim Acta.* **2021**, *13*(2), pp. 1–10. <https://doi.org/10.20956/ica.v14i3.18333>
125. Tsaboula, A.; Papadakis, E.; Vryzas, Z.; Kotopoulou, A.; Kintzikoglou, K.; Papadopoulou, -M.E. Assessment and Management of Pesticide Pollution at a River Basin Level Part II: Optimization of Pesticide Monitoring Networks on Surface Aquatic Ecosystems by Data Analysis Methods. *Sci. Total Environ.* **2019**, *653*, pp. 1597–1611. <https://doi.org/10.1016/j.scitotenv.2018.10.270>
126. Pokhrel, B.K.; Paudel K.P. Assessing the efficiency of alternative best management practices to reduce nonpoint source pollution in a rural watershed located in Louisiana, USA. *Water (Switzerland)*. **2019**, *11*(8), p.1714. <https://doi.org/10.3390/w11081714>
127. Ma, Z.; Liu, J.; Dick, R.P.; Li, H.; Shen, D.; Gao, Y.; Waigi, M.G.; Ling, W. Rhamnolipid influences biosorption and biodegradation of phenanthrene by phenanthrene-degrading strain *Pseudomonas* sp. Ph6. *Environ. Pollut.* **2018**, *240*, pp. 359–367. <https://doi.org/10.1016/j.envpol.2018.04.125>
128. Roy, A.; Dutta, A.; Pal, S.; Gupta, A.; Sarkar, J.; Chatterjee, A.; Saha, A.; Sarkar, P.; Sar, P.; Kazy, S.K. Biostimulation and bio-augmentation of native microbial community accelerated bioremediation of oil refinery sludge. *Bioresour. Technol.* **2018**, *253*, pp. 22–32. <https://doi.org/10.1016/j.biortech.2018.01.004>
129. Liu, X.; Hu, X.; Cao, Y.; Jing, W.P.; Yu, H.J.; Guo, P.; Huang, L. Biodegradation of Phenanthrene and Heavy Metal Removal by Acid-Tolerant *Burkholderia fungorum* FM-2. *Front. Microbiol.* **2019**, *10*, pp. 1–13. <https://doi.org/10.3389/fmicb.2019.00408>
130. Selvin, J.; Shanmugha Priya, S.; Seghal-Kiran, G.; Thangavelu, T.; Sapna-Bai, N. Sponge-associated marine bacteria as indicators of heavy metal pollution. *Microbiol. Res.*, **2009**, *164*(3), pp. 352–363. <https://doi.org/10.1016/j.micres.2007.05.005>
131. Anggela, A.; Marzuki, I. Bioadsorption capacity of marine sponge symbiotic bacteria against heavy metal Contaminants, *Ko-valen: Jurnal Riset Kimia*, **2021**, *7*(1), PP. 12–22, [In Indonesia] <https://doi.org/10.22487/kovalen.2021.v7.i1.15439>
132. Knobloch, S.; Jóhannsson, R.; Marteinsson, V. Bacterial diversity in the marine sponge *Halichondria panicea* from Icelandic waters and host-specificity of its dominant symbiont *candidatus Halichondriabacter symbioticus*. *FEMS Microbiology Ecology*, **2018**, *95*(1), pp. 1–13. <https://doi.org/10.1093/femsec/fiy220>
133. Ziss, E.; Friesl, -H.W.; Noller, C.; Watzinger, A.; Hood, -N.R. Heavy Metal City-Zen. Exploring the potential risk of heavy metal contamination of food crop plants in urban gardening contexts using a citizen science approach. *EGU Gen. Assem. Conf. Abstr.* **2020**, *13*(15), p. 8626. <https://doi.org/10.3390/su13158626>
134. White, J.R.; Patel, J.; Ottesen, A.; Arce, G.; Blackwelder, P.; Lopez, J.V. Pyrosequencing of Bacterial Symbionts within *Axinella corrugata* Sponges: Diversity and Seasonal Variability. *Plos ONE*, **2012**, *7*(6), pp. 1–12. <https://doi.org/10.1371/journal.pone.0038204>
135. Parhamfar, M.; Abtahia, H.; Godinib, K.; Saeedi, R.; Sartaje, M.; Villaseñor, J.; Couloung, F.; Kumarg, V.; Soltanighiash, T.; Radi, E.G.; Koolivand, A. Biodegradation of heavy oily sludge by a two-step inoculation composting process using synergistic effect of indigenous isolated bacteria. *Process Biochem.* **2020**, *91*, pp. 223–230. <https://doi.org/10.1016/j.procbio.2019.12.014>
136. Arroyo, A.; Provoste, F.; Rodríguez, M.; Prieto, A.L. A mechanistic model to assess the fate of naphthalene and Benzo(A)etilene in a chilean wwtp. *Processes*. **2021**, *9*(8), p. 1313. <https://doi.org/10.3390/pr9081313>
137. Okoro, C.C. Biosurfactant-enhanced remediation of hydrocarbon contaminated mangrove swamp. *Nature and Science*, **2010**, *8*(8), pp. 152–162. <https://doi.org/10.4314/ijbcs.v3i1.42736>
138. Liu, W.J.; Duan, X.D.; Wu, L.P.; Masakorala, K. Biosurfactant Production by *Pseudomonas aeruginosa* SNP0614 and its Effect on Biodegradation of Petroleum. *Applied Biochemistry and Microbiology*, **2018**, *54*(2), pp. 155–162. <https://doi.org/10.1134/S0003683818020060>
139. Tenea, A.G.; Vasile, G.G.; Dinu, C.; Gheorgh, S.; Pascu, L.F.; Mureseanu, M.; Ene, C. Behavior of Cd accumulation in *sinapis alba* L. In the presence of essential elements (Ca, Mg, Fe, Zn, Mn, Cu, Ni). *Rev. Chim.* **2020**, *71*, 378–389. <https://doi.org/10.37358/RC.20.6.8204>
140. Alaboudi, A.A.; Ahmed, B.; Brodie, G. Annals of Agricultural Sciences Phytoremediation of Pb and Cd contaminated soils by using sun flower (*Helianthus annuus*) plant. *Ann. Agric. Sci.* **2018**, *63*, pp. 123–127. <https://doi.org/10.1016/j.aoas.2018.05.007>

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141. Isikhuemhen, O.S.; Mikiashvili, N.A.; Senwo, Z.N.; Ohimain, E.I. Biodegradation and Sugar Release from Canola Plant Biomass by Selected White Rot Fungi. *Advances in Biological Chemistry*, **2014**, 04(06), pp. 395–406. <https://doi.org/10.4236/abc.2014.46045>
 142. Al-Nasrawi, H. Role of Fungi in Bioremediation. *Adv. Biotechnol. Microbiol.*, **2019**, 12(4), pp. 77-81. <https://doi.org/10.19080/AIBM.2019.12.555841>
 143. Fomina, M.; Charnock, J.M.; Hillier, S.; Alvarez, R.; Livens, F.; Gadd, G.M. Role of fungi in the biogeochemical fate of depleted uranium. *Curr. Biol.*, **2008**, 18(9), pp. 375-377. <https://doi.org/10.1016/j.cub.2008.03.011>
 144. Morganti, T.M.; Ribes, M.; Yahel, G.; Coma, R. Size Is the Major Determinant of Pumping Rates in Marine Sponges. *Front Physiol.* **2019**, 10, p. 1474. <https://doi.org/10.3389/fphys.2019.01474>
 145. Keller-Costa, T.; Jousset, A.; Van-Overbeek, L.; Van-Elsas, J. D.; Costa, R. The freshwater sponge *Ephydatia fluviatilis* harbours diverse *Pseudomonas* species (*Gammaproteobacteria*, *Pseudomonadales*) with broad-spectrum antimicrobial activity. *PLoS ONE*, **2014**, 9(2), p. e88429. <https://doi.org/10.1371/journal.pone.0088429>
 146. Shimoda, T.; Suryati, E.; Ahmad, T. valuation in a Shrimp Aquaculture System Using Mangroves, Oysters, and Seaweed as Biofilters Based on the Concentrations of Nutrients and Chlorophyll. *JARQ* **2006**, 40, pp. 189–193. <https://doi.org/10.6090/jarq.40.189>.
 147. Cecotti, M.; Coppotelli, B.M.; Mora, V.C.; Viera, M.; Morelli, I.S. Efficiency of surfactant-enhanced bioremediation of aged polycyclic aromatic hydrocarbon-contaminated soil: Link with bioavailability and the dynamics of the bacterial community. *Sci. Total Environ.* **2018**, 634, pp. 224–234. <https://doi.org/10.1016/j.scitotenv.2018.03.303>