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Remieri

Overview and New Insights into the Metabolic Syndrome: Risk Factors and Emerging Variables in the Development of Type 2 Diabetes and Cerebrocardiovascular Disease

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Abstract: Metabolic syndrome (MetS) is considered a metabolic disorder that has been steadily increasing globally and seems to parallel the increasing prevalence of obesity. It consists of a cluster of risk factors, which traditionally includes obesity and hyperlipidemia, hyperinsulinemia, hypertension, and hyperglycemia. These four core risk factors are associated with insulin resistance (IR) and importantly, the MetS is known to increase the risk for developing cerebrocardiovascular disease and type 2 diabetes mellitus. The MetS had its early origins in IR and syndrome X. It has undergone numerous name changes with additional risk factors and variables being added over the years; however, it still remains the MetS worldwide for the past three decades. This overview continues to add novel insights to the MetS and suggests that leptin resistance with hyperleptinemia, aberrant mitochondrial stress and reactive oxygen species (ROS), impaired folate-mediated one-carbon metabolism with hyperhomocysteinemia, vascular stiffening, microalbuminuria, and visceral adipose tissues extracellular vesicle exosomes be added to the list of associated variables. Notably, the role of a dysfunctional and activated endothelium and deficient nitric oxide bioavailability along with a dysfunctional and attenuated endothelial glycocalyx, vascular inflammation, systemic metainflammation, and the important role ROS and reactive species interactome are discussed. With new insights and knowledge regarding the MetS, comes the possibility of new findings through further research.

Keywords: Endothelial dysfunction; Exosomes; Hyperinsulinemia; Hyperglycemia; Hyperlipidemia; Hypertension; Insulin Resistance; Leptin Resistance; Metainflammation; miRNAs

1. Introduction

Metabolic syndrome (MetS) is a cluster of multiple risk factors and variables that are associated with an increased risk for the development of atherosclerotic cardiovascular disease and type 2 diabetes mellitus (T2DM) [1,2]. In this review cardiovascular disease will also include cerebrovascular disease or more specifically, the two combined as cerebrocardiovascular disease (CCVD), since the vasculature system, heart, and brain are all involved in the clinical utility of the MetS. The core underlying risk factors include: 1) obesity (central or visceral obesity), systemic insulin resistance (IR) with hyperinsulinemia, hyperamylinemia, hyperleptinemia, and leptin resistance (LR); 2) hyperlipidemia with atherogenic dyslipidemia typified by elevated very-low-density lipoproteins or triglycerides, elevated small dense low-density lipoproteins-cholesterol, and decreased high-density lipoproteins-cholesterol; 3) essential hypertension (HTN); 4) hyperglycemia with or without manifest T2DM [1,2].

Since the early days in 1923 (Kylin) [3] and 1939 (Himsworth and Kerr) [4], it was clinically obvious to many that certain CCVD risk factors and variables tended to cluster or co-occur and that this clustering phenomenon shared multiple underlying causes, pathobiological mechanisms, and features. Following Reaven's definition of syndrome X in 1988 [5] the National Cholesterol Education Panel Adult Treatment Panel III (NCEP ATP III) identified and characterized the MetS with clinical utility and comprised five clinical and laboratory screening criteria: 1) abdominal obesity

by waist circumference (males >100 cm/>40 inches and females >88cm/>35 inches reflecting increased visceral adiposity and possible IR; 2) triglycerides \geq 150 mg/dL; 3) HDL-cholesterol <40 mg/dL in males and <50 mg/dL females reflecting atherogenic dyslipidemia and hyperlipidemia; 4) blood pressure \geq 130/ \geq 85 mm Hg reflecting HTN; 5) fasting glucose \geq 110 mg/dL and the newer American Diabetes Association cut point of \geq 100 mg/dL in 2002. Importantly, any three of the previously listed five criteria qualify individuals for the diagnosis of the MetS [1]. Importantly, note that in the NCEPT ATP III guidelines increased triglycerides (2) are separated from low HDL-cholesterol (3) reflecting atherogenic dyslipidemia and that is why NCEP ATP III differs somewhat from the earlier mentioned four core risk factors in the preceding paragraph.

Since the MetS was first described, it has been described by many different major global organizations/associations and given many different names with multiple risk factors and variables added to what could now be described as the MetS reloaded. In its simplest form, the MetS consists of multiple clinically relevant risk factors and associated variables as they intersect, for the development of atherosclerotic CCVD and T2DM (Figure 1).

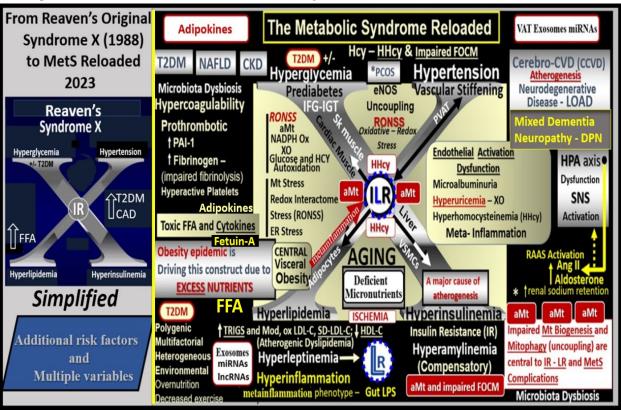


Figure 1. From Reaven's simplified syndrome X to the complicated metabolic syndrome (MetS) reloaded. This image illustrates the simple Reaven's Syndrome X (far left) to the complex MetS reloaded and its multiple risk factors. The central "X" in both figures honors Jerry Reaven who initially coined the term Syndrome X in 1988 and championed the concept that the resistance to insulin-mediated glucose disposal - insulin resistance (IR) was a characteristic finding in individuals with type 2 diabetes mellitus (T2DM) and cardiovascular disease (CVD). According to Reaven, the letter "X" was originally chosen because at the time coronary artery disease (CAD) and/or cerebrocardiovascular disease (CCVD) was a relatively unknown risk of human insulin resistance. Syndrome X was later termed the MetS by National Cholesterol Education Program Adult Treatment Panel III and Grundy et al, 2002 and 2004 respectively. There are four arms to this letter X and each arm of the letter X has a designated condition to further illustrate the H phenomenon, representing a "hyper" - "H" state e.g., Hyperlipidemia (lower left arm), Hyperinsulinemia and hyperamylinemia (lower right arm), Hypertension (essential) (upper right arm), and Hyperglycemia with or without manifest T2DM (upper left arm) in the MetS reloaded. Importantly, visceral or central obesity is thought to be a major driver of this syndrome and is related to the emergent science of visceral adipose tissue (VAT) with adipocyte-derived adipokines and macrophage-derived cytokines/chemokines.

Additionally, it is important to note the novel adipocyte and macrophage-derived exosomes with their novel signaling microRNAs (miRNAs) and long-non-coding RNAs (lncRNAs), which are capable of both paracrine and endocrine (inter-organ and long-distant) signaling in addition to the signaling via peripheral cytokine/chemokine adipokine network due to excessive meta-inflammation within VAT depots. Note that insulin resistance (IR) and leptin resistance (LR or Lr) (ILR) are placed centrally within the letter X. Importantly, note the red bold arrow connecting central visceral obesity to the central ILR within the letter X that is related to increased meta-inflammation. Also, note how the aberrant mitochondria (aMt) (a constant finding in obese and diabetic models) and hyperhomocyteinemia (HHcy) are flanking the central insulin and leptin resistance (ILR) that reflects impaired folate-mediated one-carbon metabolism (FOCM). Additionally, note the important role of angiotensin II; CAD = coronary artery disease; CKD = chronic kidney disease; CCVD = cerebrocardiovascular disease; DPN = diabetic peripheral neuropathy; eNOS = endothelial nitric oxide synthase; <math>ER = endoplasmicreticulum; FFA = free fatty acids; Hcy = homocysteine; ILR = insulin/leptin resistance; LOAD = late-onset Alzheimer's disease; lncRNAs = long non-cording RNAs; HPA = hypothalamic-pituitary-adrenal axis; LPS =lipopolysaccharide; LR = leptin resistance - selective Lr; miRNAs = micro ribonucleic acids; Mt = mitochondria; NADPHOx = nicotinamide adenine dinucleotide phosphate oxidase; NAFLD = non-alcoholic fatty liver disease; $NO = nitric \ oxide; \ Non-HDL-C = non-high-density \ lipoprotein-cholesterol; \ O2 = oxygen; \ oxLDL-C = oxidized$ low density lipoprotein-cholesterol; PAI-1 = plasminogen activator inhibitor -1; PCOS = polycystic ovary syndrome; RAAS = renin-angiotensin-aldosterone-system; RNA = ribonucleic acid; RONSS = reactive oxygen, nitrogen, sulfur species; RSI = reactive species interactome;; sdLDL-C = small dense low-density lipoproteincholesterol; SNS = sympathetic nervous system; Trigs = triglycerides; XO = xanthine oxidase.

The late Jerry Reaven initially coined the term Syndrome X and championed the concept that the resistance to insulin-mediated glucose disposal – IR was a characteristic of patients with T2DM and cardiovascular disease (CVD) [5]. Subsequently, the Syndrome X concept was later termed the MetS by the NCEP-ATIII and Grundy et al, respectively (2002, 2004) [1,2]. There are four arms to this letter X and each arm of the letter X has a designated condition to further illustrate the H phenomenon, representing a "Hyper" state e.g., Hyperlipidemia – atherogenic dyslipidemia nonhigh-density lipoprotein-cholesterol, Hyperinsulinemia and Hyperamylinemia, HTN, and Hyperglycemia. Importantly, central or visceral obesity is thought to be a major driver of this syndrome and relates to the emerging science of visceral adipose tissue (VAT). VAT with its novel exosomal and microRNA signaling in addition to the signaling via peripheral cytokine/chemokine (pCC) network and adipokines results in metainflammation that continues to be an intriguing emerging science [6]. Hyperleptinemia and LR, hyperinflammation, hyperinsulinemia/hyperamylinemia, pancreatic islet β-cell dysfunction and apoptosis, HTN, and vascular stiffening are extremely important when evaluating individuals for MetS. Hyperglycemia including impaired fasting glucose (IFG) or impaired glucose tolerance (IGT) (reflecting a prediabetic state) with or without the presence of manifest T2DM is also very important. Also, aberrant mitochondria (aMt) that are leaky allow for increased cellular toxicity and pathologic remodeling due to excessive reactive oxygen species (ROS) that are present in multiple organs in those individuals with MetS. Elevated homocysteine (HCY) and hyperhomocysteinemia (HHCY) is present in the MetS and contributes to Mt dysfunction and/or damage due to impaired folate-mediated one-carbon metabolism (FOCM). Additionally, endothelial cell activation and dysfunction, endothelial nitric oxide synthase (eNOS) uncoupling with decreased nitric oxide (NO) bioavailability, and an attenuated and/or loss of the endothelial glycocalyx (ecGCx) are all very important to the MetS that contribute to and result from EC activation and dysfunction. Interestingly, the World Health Organization has now included microalbuminuria in its list of predisposing criteria for the presence of MetS (Figure 1) [7].

Each of the four arms of the letter X (Hyperlipidemia, Hyperinsulinemia, Hypertension, and Hyperglycemia) is important, and note that insulin and leptin resistance can impact each of the tissues and the clinical disease states (Figure 1). Further, note that the MetS constitutes a milieu conducive to tissue oxidative-redox stress. Importantly, oxidative-redox stress incorporates the sum

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of reactive oxygen, nitrogen, and sulfur species (RONSS) stress and the cellular RONSS reactive species interactome (RSI) along with chronic low-grade (sterile) inflammation (hereafter termed metainflammation). Metainflammation is mediated by proinflammatory macrophages located within the adipose tissue that forms crown-like structures (CLS) with VAT being greater than the subcutaneous adipose tissue. Also, the gut (primarily the colon) is capable of generating multiple proinflammatory pathogen-associated molecular patterns (PAMPS) and lipopolysaccharides (LPS) when there is associated obesity-related gut microbiome dysbiosis (Figure 1). metainflammation results in oxidative-redox stress and oxidative-redox stress results in metainflammation creating a bidirectional vicious cycle. The multiple metabolic toxicities associated with the MetS including the RONSS interactome and endothelial activation/dysfunction combine to weave a complicated mosaic fabric, which is also known to be associated with the activation of the renin-angiotensin-aldosterone-system (RAAS) and the sympathetic nervous system (SNS) (Figure 1). Additionally, CCVD and T2DM may be associated with neurodegenerative diseases such as vascular dementia (VaD), late-onset Alzheimer's disease (LOAD), mixed cooccurance dementia, Parkinson's disease, and diabetic encephalopathy - cognopathy [8-11]. Importantly, the combination of CCVD and chronic kidney disease together comprise the heartbrain-kidney (HBK) axis in the MetS reloaded that becomes involved when there is systemic vascular stiffening (particularly in the aortic, renal, and carotid arterial vessels) associated with MetS, HTN, and T2DM [12-15].

It is important to note that hyperinsulinemia is initially protective and a compensatory response to nutrient excess and obesity to control glucose elevation. However, if hyperinsulinemia remains chronic as in the MetS reloaded there is a 'price to pay' by becoming a central commonality along with the development of aMt and the development of RONSS interactome and its detrimental role in each of the four arms of the syndrome X and its clinical disease states and syndromes to result in increased atherogenesis, accelerated atherosclerosis – atheroscleropathy, and the development of CCVD (Figure 2) [13,14,16].

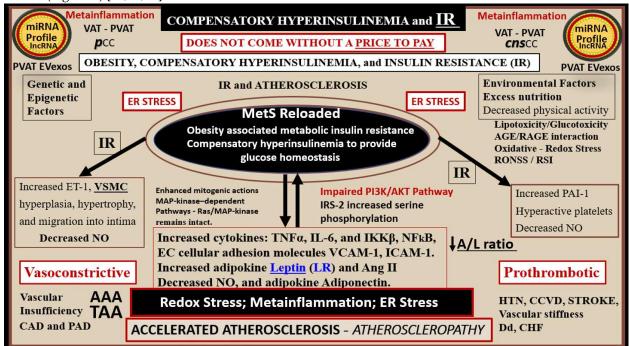


Figure 2. Insulin resistance (IR) and compensatory hyperinsulinemia do not come without a price to pay. IR plays a central role in the MetS and results in an impaired PI3K/AKT pathway due to impaired IRS-2 function (IRS-2 increased serine phosphorylation) and decreased insulin-stimulated NO synthesis and impaired vasodilation. However, the elevated insulin can still signal through the mitogenic MAPkinase-dependent pathway and contribute to arterial vessel wall remodeling and accelerated atherosclerosis (atheroscleropathy) due to pro-atherosclerotic, pro-thrombotic, and vasoconstrictive effects). The increased redox stress and inflammatory vicious cycle that drive IR are

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promoted by an associated increased peripheral and central nervous cytokines and chemokines (pCC and cnsCC respectively) and the IR promoting effects from adipose-derived miRNAs via increased extracellular vesicle exosomes (EVexos) and their microRNA profiles along with the ongoing metainflammation. $AAA = abdominal \ aortic \ aneurysm; \ Ang \ II = angiotensin \ II; \ CAD = coronary \ artery \ disease; CHF = congestive heart failure; ET-1 = endothelin-1; ER = endoplasmic reticulum; HTN = hypertension; ICAM-1= Intercellular Adhesion Molecule 1; IKK<math>\beta$ = inhibitor of nuclear factor kappa-B kinase subunit beta; IL-6 = interleukin-6; $lncRNA = long \ non-coding \ ribonucleic \ acid; \ LR = leptin \ resistance; \ miRNA = micro \ ribonucleic \ acid; \ NFkB = Nuclear \ factor \ kappa-light-chain-enhancer \ of \ activated \ B \ cells; \ NO = nitric \ oxide; \ PAD = peripheral \ arterial \ disease; \ PAI-1 = plasminogen \ activator \ inhibitor-1; \ ROS = reactive \ oxygen \ species; \ RNA = ribonucleic \ acid; \ RONSS = reactive \ oxygen, \ nitrogen, \ sulfur \ species; \ RSI = reactive \ species \ interactome; \ TAA = thoracic \ aortic \ aneurysm; \ TNF<math>\alpha$ = tumor necrosis \ factor \ alpha; \ VCAM-1 = \ vascular \ cell \ adhesion \ molecule-1; \ VSMC = \ vascular \ smooth \ muscle \ cell(s).

The addition of aMt adds even further strength to understanding the important role of the MetS reloaded since Mt dysfunction plays a key role in systemic IR and brain insulin resistance (BIR), HTN, and T2DM. Notably, even Reaven added the risk factors hyperuricemia and increased plasminogen activator-1 (PAI-1) and these were incorporated into the insulin resistance syndrome in 1993 [17,18]. HHcy along with elevated uric acid (UA) hyperuricemia [19] could be added to the "H" phenomenon, especially, since HHcy is an independent risk factor for CCVD, HTN, and vascular stiffening in addition to being a biomarker for impaired FOCM [20]. It is important to note the inclusion of toxic free fatty acids (FFAs) to the left of the hyperlipidemia arm and just above the obesity epidemic (silver box) and present in the earlier simplistic syndrome X (Figure 1). While these toxic, short-chain, saturated FFA are detrimental and promote accelerated atherosclerosis, it is important to note that the long-chain n-3 polyunsaturated fatty acids may be beneficial and antiatherogenic, anti-inflammatory and antithrombotic.

Notably, hyperleptinemia is included and LR (encircled lower mid-figure 1) and that WAT and VAT are important to the development of hyperleptinemia and LR. Finally, EC activation – dysfunction and eNOS uncoupling with decreased bioavailable NO are extremely important to the development and progression of the MetS reloaded and its increased risk for the development of CCVD and T2DM (Figure 1) [14,16]. Interestingly, note that the WHO is one of the only organizations that includes microalbuminuria in its definition of the MetS and that microalbuminuria may reflect endothelial activation and dysfunction with decreased NO and increased glomerular capillary permeability [7].

Indeed, the MetS has a rich history along with the various key obesity models that have been created to study the role of obesity. MetS, systemic IR and BIR, T2DM, and leptin - LR along with the discovery of the *ob/ob* or the *fa/fa* gene and the discovery of leptin in 1994. This important discovery resulted in a paradigm shift in regards to the metabolic, secretory and endocrine roles played by adipose tissue in addition to a better understanding of the MetS [21,22].

The following timelines of discovery provide the background knowledge that was necessary to develop our understanding and enabled the current concept of the MetS that has been important for obesity and T2DM research that is ongoing to date and its implications with obesity (Figure 3) [1, 2, 3, 4. 5, 17, 18, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32]

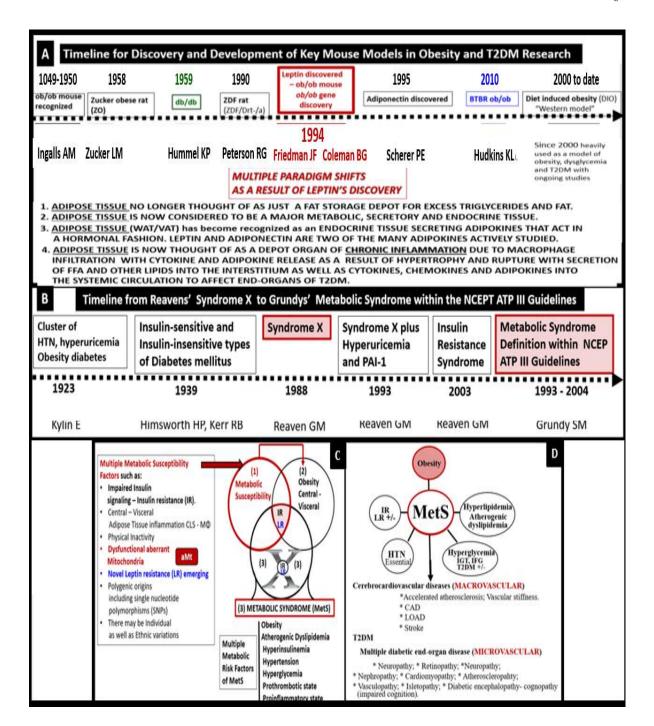


Figure 3. Panel A demonstrates the timeline for the development of key obesity and T2DM models from 1948 to date that have resulted in multiple paradigm shifts in regards to the metabolic syndrome (MetS). Panel B demonstrates the timeline of Reaven's syndrome X to the NCEP ATP III guidelines for the MetS and Grundy's emphasis on the Met S. These two combined timelines allow one to compare the importance of obesity, insulin and leptin resistance to the timeline for the development of the MetS. Panel C illustrates that the MetS is associated with multiple variables and the importance of an individual's metabolic susceptibility factors (boxed-in factors) to better understand why only some obese individuals develop MetS. Panel D illustrates that the MetS is associated with multiple variables and an increased risk for cerebrocardiovascular disease (CCVD) via macrovascular disease, while diabetic end-organ disease is associated with type 2 diabetes mellitus (T2DM) and microvascular disease. Panel A is adapted and modified with permission by CC 4.0 [32]. FFA = free fatty acids; VAT = visceral adipose tissue; WAT = white adipose tissue.

Importantly, Grundy et al. have previously discussed the importance of the MetS as being a multiplex cardiovascular risk, which incorporates the interplay of obesity, metabolic susceptibility, and multiple metabolic risk factors of the MetS (Figure 3C and D) [2]. Further, Grundy has emphasized metabolic susceptibility in order to support the clinical utility of the MetS, in that, when an individual develops excess central-VAT body fat (obesity) the MetS develops. This emphasis is in addition to the previous five NCEPT ATP criteria previously discussed [33].

Alberti et al, (2009) held a joint international task force meeting in an attempt to harmonize the definition and discuss the various risk factors for the diagnosis of the MetS. The task force emphasized the importance of identifying those with the MetS so that they could receive the appropriate treatment by their health care providers to improve these risk factors and the increased risk of CCVD (two times higher) and T2DM (five-fold higher) as compared to non-MetS individuals. Further, they made the decision to retain an elevated waist circumference as a risk factor with the realization that the cut points may be different for different ethnic groups, which would be determined later by other task forces [34].

Throughout this overview, the use of various preclinical obese, insulin- and leptin-resistant preclinical diabetic rodent models will be utilized in order to provide a better understanding of how the multiple risk factor and variables of the MetS result in cellular and organ ultrastructural remodeling and functional abnormalities. Recently, Panchal and Brown (2011) have discussed the use of rodent models to study the MetS and concluded that the ideal model to study the parallels between rodents and humans is the diet-induced obesity (DIO) model fed a high fat and high sucrose glucose diet [35]. While our group has studied multiple obesity models including DIO models, most of our transmission electron microscopy (TEM) studies have utilized the *db/db* leptin receptor-deficient mouse models with obesity, elevated leptin, LR, IR, and diabetes to identify cellular remodeling changes in multiple organ systems.

The addition of novel emerging risk factors and variables may allow for a better understanding of the MetS and earlier recognition for individuals and their healthcare providers to prevent the development of CVVD and T2DM. By providing new knowledge and novel insights, it is hoped that further research in this exciting field of study may be stimulated in order to decrease the risk of developing CCVD and T2DM along with their complications in these at-risk individuals.

The specific aim of this overview is to add novel risk factors, emerging variables, and possible biomarkers to the existing traditional risk factors associated with the MetS. In the following sections (2. through 5.), each of the four traditional risk factor arms of the letter X will be discussed in greater detail. Additionally, the more novel non-traditional emerging risk factors and variables such as endothelial cell activation/dysfunction, endothelial glycocalyx remodeling (section 6.) and metainflammation including gut microbiota dysbiosis, adipokines, cytokines/chemokines, adiposederived extracellular exosomes, micro RNAs, and long-non-coding RNAs (Section 7.) (Figure 1).

2. MetS Reloaded, Hyperlipidemia – Atherogenic Dyslipidemia

The late Russell Ross was a pioneer and a giant in the understanding, development, and progression of atherosclerosis acting as an inflammatory disease of the arterial vessel wall [36]. Ross felt that this injury to the arterial vessel wall was largely due to elevated levels of cholesterol and particularly LDL-C, modified or oxidized LDL-C, and now we know that the small-dense LDL-C is extremely proatherogenic and proinflammatory. Importantly, the toxic lipids of atherogenic dyslipidemia that are associated with the MetS act as an injury to the intima ECs and the subintimal space. Further, these atherogenic lipids will serve as an injury to the endothelium of the arterial vessel wall with an ensuing response to injury wound healing remodeling, which will result in accelerated atherosclerosis in the MetS and T2DM [14,16].

Hyperlipidemia or dyslipidemia in the MetS (Figure 1) is characterized by an atherogenic dyslipidemia, which consists of elevated triglycerides and atherogenic modified LDL-C; oxLDL-C; small dense LDL-C; decreased atheroprotective HDL-C [37,38]. Additionally, non-HDL-C is now included to demonstrate not only increased VLDL-triglycerides but also remaining remanent atherogenic lipoproteins including lipoprotein (a) (Lp(a)), LDL-C, intermediate-density lipoprotein,

and VLDL-C along with elevated toxic, short chain, saturated FFAs, which play an important role in atherogenic dyslipidemia but are often not included in its definition [37Wilson]. For example, the elevated triglycerides (triacylglycerols) in atherogenic dyslipidemia associated with the MetS are lowered by long-chain n-3 polyunsaturated fatty acids (long chain n-3 PUFA (LC n-3PUVA) including ω -3 fatty acids) due to a decrease in VLDL via a decreased sterol receptor element-binding protein-1c and decreasing β-Oxidation in mitochondria and peroxisomes [38]. Additionally, LC n-3 PUFAs are known to significantly decrease the risk of fatal coronary heart disease (CHD) [39]. The exact mechanisms through which long chain n-3 PUFA has an effect on CHD are not completely established but it is thought that they might include decreased fasting and postprandial triacylglycerol (triglyceride) levels, decreased arrhythmias, modulation of platelet aggregation affecting thrombus formation, and decreased synthesis of pro-inflammatory molecules [39]. The mechanistic relation between long chain n-3 PUFA and inflammation has attracted great interest, and in vitro studies have demonstrated that these specific fatty acids decrease endothelial activation and may affect eicosanoid metabolism and result in improved resolution of inflammation [38]. Indeed, there are a multitude of studies that support the positive role of long chain n-3 PUFA in reducing the risks of ASCVD and CHD that has been extensively reviewed by deRoos et al. [38].

Metainflammation is currently recognized as a significant process in the development and progression of not only atherosclerosis but also CCVD and CHD. As put forth by Ross [36], the instigation of inflammation may well provide the link between hyperlipidemia - atherogenic dyslipidemia and atherogenesis since, inflammatory pathways are known to promote thrombosis, the most serious complication of atherosclerosis responsible for both myocardial infarctions and most transient ischemic attacks and strokes. Importantly, we may be facing the dawn of a new era in regards to lipid-lowering with newer therapies emerging such as monoclonal antibodies, antisense oligonucleotides small interfering ribonucleic acids (RNAs), and even the possibilities of vaccination that are emerging as novel treatments with LDL-C still being a primary target, non-HDL-C which includes all of the atherogenic lipoproteins, which includes LDL-C, VLDLs (which transports endogenous triglycerides, <u>phospholipids</u>, <u>cholesterol</u>, and <u>cholesteryl esters</u>) as a secondary target, and the remanent lipoprotein Lp(a)) [39]. The mechanisms of these novel emerging therapies are beyond the scope of this overview; however, for those who have a greater interest, a full discussion can be better appreciated [40].

Before leaving this section, it is important to address the elevation in triglycerides as part of the MetS atherogenic dyslipidemia. Evidence has accumulated that modifying or lowering triglycerides and increasing HDL-C levels with specific drug therapy will reduce the risk of CHD independently of statin therapy or LDL-C lowering. While at first, this may seem like a bold statement; however, therapy with gemfibrozil in the Veterans Administration HDL Intervention Trial (VA-HIT) led to triglyceride-lowering of 31%, HDL-C raising of 6%, and no change in LDL-C. The individuals in this trial consisted of US military veterans with known coronary artery disease at the outset who experienced a 24% reduction in coronary events (both nonfatal myocardial infarction and coronary death) during follow-up [41,42]. Also, it is appropriate to close this section with a quote from the late Elliott P Joslin who was one of the great clinicians and diabetologists of his time and presented the following quote to the American College of Physicians in 1927 as follows:

"I believe the chief cause of premature development of arteriosclerosis in diabetes, save for advancing age, is due to an excess of fat, an excess of fat in the body, obesity, an excess of fat in the diet, and an excess of fat in the blood. With an excess of fat diabetes begins and from an excess of fat diabetics die, formerly of coma, recently of arteriosclerosis" [43].

Joslin's role of an excess of fat, obesity, and hyperlipidemia was "spot on" and still remains so to this day. The important role of excess omental – visceral adipose depots along with the excess subcutaneous fat dopots still remain an important tissue to the excess leptin production, LR, and chronic sterile metainflammation that is so very important for the development of accelerated atherosclerosis, ASCVD, and CCVD [16]. The role of fat is especially important in obese models that are available to study such as the diet-induced obesity (DIO). Western models due to high fat, high sucrose, and fructose feeding, the leptin receptor-deficient db/db models, the novel leptin-

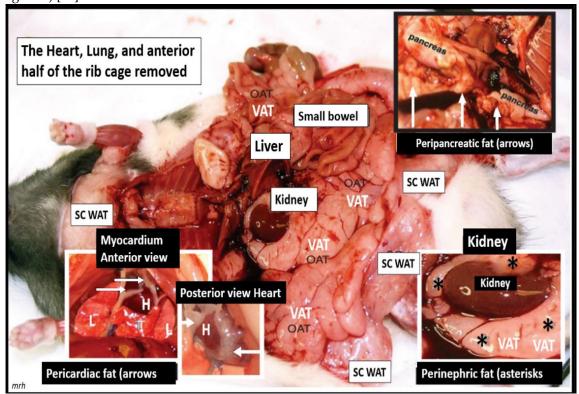


Figure 4. Necropsy of a male Zucker fa/fa or ob/ob at nine-weeks of age. Gross findings at necropsy of obesity in the nine-week-old adolescent male Zucker obese ob/ob rat model. This image depicts the massive accumulation of omental adipose tissue (OAT) or visceral adipose tissue (VAT) in the Zucker fa/fa or ob/ob model. Also, note there is an accumulation of excessive subcutaneous (SC) adipose tissue - white adipose tissue (WAT) depot. Inserts depict the accumulation of peripancreatic VAT fat (white arrows) (upper-right), pericardiac VAT fat (white arrows) (lower-left), and extremely excessive perinephric VAT fat (asterisks) (lower-right). This image is adapted with permission from open access [44]. H = heart; L = collapsed lung; mrh = necroscopy-photographer initials melvin ray hayden; OAT = Omental adipose tissue; T = Thymus.

3. MetS Reloaded, Insulin, Hyperinsulinemia, IR, Leptin, Hyperleptinemia, and LR

Both pancreatic islet β -cell-derived insulin and adipose tissue-derived leptin hormones in obesity (both white adipose and visceral adipose tissue) act centrally. Both of these hormones are important in regulating food intake and adiposity in rodent models and humans [32,45]. Additionally, glucose homeostasis is closely regulated not only by insulin but also assisted by leptin. In genetic rodent models that are leptin-deficient such as the Zucker obese ob/ob-fa/fa rat or mouse models; the novel BTBR ob/ob mouse models or leptin-resistant models such as occurs in the db/db mouse models secondary to a point mutation in the leptin receptor gene, which causes deficient leptin signaling with hyperleptinemia and LR are important rodent models to study the effects of hyperphagia, obesity, hyperglycemia due to impaired glucose tolerance – prediabetes in ob/ob models, and manifest T2DM in db/db models (Figure 3A). Indeed, hyperphagia and obesity are both hallmark phenotypes in the ob/ob and db/db models with dysglycemia; however, only the db/db model and the BTBR ob/ob models are defined as being overtly diabetic [32,46–50]. The importance of IR from an epidemiologic standpoint is that IR affects nearly one-third of the US population and is known to precede the development of T2DM [13,14,16,51]. Additionally, IR and compensatory hyperinsulinemia are associated with multiple abnormalities associated with the MetS. These

abnormalities include impaired insulin signaling of the phosphoinositide 3-kinase/protein kinase B (PI3Kinase/AKT) pathway that is associated with serine hyperphosphorylation of the insulin receptor substrate 2 (IRS2) signaling pathway, decreased insulin-stimulated NO production with decreased vasodilation and proconstrictive, prothrombotic, and pro-atherosclerotic effects on the arterial vessels with accelerated atherosclerosis. Also, there is an associated increased redox stress, endoplasmic reticulum stress, and promotion of metainflammation as previously discussed and depicted in Section 1. (Figure 2).

Leptin, hyperleptinemia, and LR are known to be present in human obesity [32,52–55]. Both peripheral leptin resistance and central leptin resistance may be present under pathophysiological conditions such as inflammation, IR, hyperlipidemia, hypertension, atherosclerosis, and obesity of the MetS [56,57]. The adipocyte-derived peptide leptin (adipokine/hormone) in humans has been linked to adiposity and insulin resistance [45,52–54]. The rate of insulin-mediated glucose uptake has been found to be significantly associated with leptin levels even after adjustment for the percentage of body fat in middle-aged men and women [55]. Recent research suggests that leptin may be an important factor linking obesity, the MetS, and CCVD [56]. It is known that under physiologic conditions that leptin may be an important factor in regulating blood pressure and volume. However, during conditions of leptin resistance and hyperleptinemia as occurs in obesity of the MetS, this endocrine hormone and adipokine may function pathologically in regards to the development of HTN and CCVD [57]. Thus, leptin and LR seem to be emerging novel players that are playing a central role in the MetS, in that, LR is not only associated with obesity but also contributes to obesity and independently affects IR [52–55,57].

3.1. MetS Reloaded, Compensatory Hyperinsulinemia, Insulin Resistance (IR) and Compensatory Hyperamylinemia

As previously mentioned, the importance of IR from an epidemiologic standpoint is that IR affects nearly one-third of the US population and is known to precede the development of T2DM [13,14,16,51]. Also, in regards to the MetS, it has been recently estimated that the MetS prevalence was more than one-third in the US population for all sociodemographic groups and these numbers were increased from 1988 [58,59].

Hyperinsulinemia and hyperamylinemia are the result of islet β -cells compensatory response to overcome cellular insulin resistance to glucose uptake in the MetS. MetS affects approximately 47 million or greater Americans in the US [60,61]. It is thought that approximately 20% will develop overt T2DM, while the remaining 80% would be capable of developing compensatory hyperinsulinemia and hyperamylinemia. This compensation for the underlying IR is due to increased β -Cell secretion of both insulin and amylin at least for a period of time through the processes of pancreatic β -cell expansion, hypertrophy, increased β -cell endoplasmic reticulum (ER) stress, and hyperplasia [62,63]. However, the compensatory hyperinsulinemia (37.6 million = 80% of 47million) that develops in the remaining 80% to prevent the development of T2DM does not come without a 'price to pay', in that, this chronic compensatory hyperinsulinemia places these patients at risk for HTN, accelerated atherosclerosis - atherogenesis, and subsequent CCVD due to atherosclerotic cerebral vascular and coronary heart disease (CCVD) (Figs. 1, 2) [5,14,63,64]. Additionally, this continued ongoing islet β -cell stress of producing more and more insulin over time would place the β -cell at risk for what is commonly referred to as β -cell fatigue and would develop β-cell ER stress and excess gluco-lipotoxicity that could cause aberrant Mt formation with decreased β -cell energy from decreased Mt-derived ATP with decreased insulin secretion and eventually β -cell failure due to β -cell exhaustion (apoptosis) as in the young db/db obese, insulin and leptin resistant diabetic models [62,63] (Figure 5).

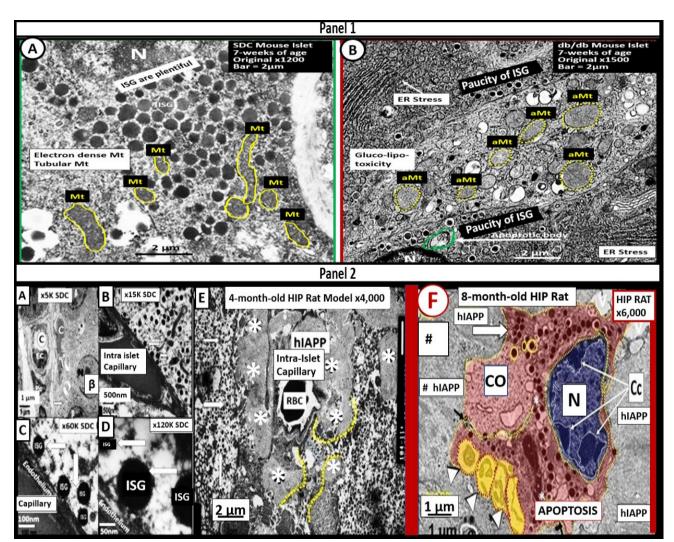


Figure 5. Ultrastructural remodeling of pancreatic islets and islet Beta-cells (β-cells) in obese and type 2 diabetes mellitus (T2DM) rodent models. **Panel 1** depicts the paucity of insulin secretory granules (ISGs), endoplasmic reticulum (ER) stress, and an increase in aberrant mitochondria (aMt) in the obese, diabetic db/db mouse (B) as compared to controls (A). **Panel 2** depicts the loss of ISGs and the excessive deposition of islet amyloid in the human islet amyloid polypeptide (HIP) rat model (E) that results in β-cell apoptosis (F) as compared to controls (A-D). Images are reproduced and modified with permission by CC 4.0 [58 Immediate and long term compl Covid may be T2DM].

Because of this increased compensation by producing more insulin and amylin there would also be an increased risk of islet amyloid deposition due to the amyloidogenic hyperamylinemia and hyperinsulinemia, which are both common to IR of the MetS [62,63]. Incidentally, there has been an increased risk for those older obese individuals who are developing T2DM during this COVID-19 pandemic and who also experienced endothelial cell activation and dysfunction [59,64]. Over time β -cell failure will develop due to β -cell apoptosis and T2DM will develop as a result of glucolipotoxicity and islet ER and redox stress (Figure 5) [59,62,63].

3.2. MetS Reloaded, Insulin Resistance, Compensatory Hyperamylinemia, and Islet Amyloid Deposition

Amylin (IAPP) is a 37 amino acid β -cell-derived hormone that is co-synthesized and copackaged in the endoplasmic reticulum within the insulin secretory granule (ISG) of the Golgi apparatus, which is then co-secreted within ISG along with insulin from the pancreatic islet β -cells into the systemic circulation and deposited within the pancreatic islets as islet amyloid polypeptide (IAPP) [59,65–67]. In situations of insulin resistance, as occurs in MetS with associated compensatory hyperinsulinemia, the β -cells will also synthesize and secrete greater amounts of

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amyloidogenic amylin, which will result in hyperamylinemia (<u>Figure 1</u>), with subsequent islet amyloid deposition within the pancreatic islets that contribute to the development of T2DM (Figure 5E) [59,62,63,66–73].

Importantly, Cooper has found that the amylin-hyperamylinemia (known to be present in IR of the MetS reloaded) is capable of activating renin and the renin-angiotensin-aldosterone system (RAAS) [74]. There are changes in endoplasmic reticulum stress (Figure 5), which can activate the apoptotic machinery [75] and aMt in obese insulin-resistant diabetic db/db models and impaired insulin secretory granule uptake by islet capillaries and β -cell apoptosis in a genetic transfection with the human islet amyloid polypeptide gene in the Sprague Dawley rat that is called the human islet amyloid polypeptide (hIAPP) rat model of diabetes (Figure 5) [59, 62, 66, 67, 68. 69].

Indeed, islet amyloid polypeptide (hIAPP) – islet amyloid is a definite hallmark remodeling change found within pancreatic islets and is known to be present in a substantial portion of human individuals with T2DM [76–79]. In the past two decades, there has been a better understanding of the role of hyperamylinemia and pancreatic islet amyloid in T2DM since its presence was initially described by Eugene Opie in 1900-1901 [80,81].

Importantly, amylin is also known to activate the renin-angiotensin-aldosterone system (RAAS) [62,63,74]. Because amylin (IAPP) is an amyloidogenic protein, it is important to note the hIAPP amyloid (through protein misfolding) also acts as an amyloid niche in the brain. This niche is thought to instigate and contribute to amyloid β (A β) deposition and progression in the brain that will accelerate cognitive decline, late-onset Alzheimer's disease (LOAD), and neurodegeneration [82–86].

Mitochondria (Mt) are vital to the normal function of pancreatic islet beta cells since the synthesis and secretion of the islet insulin secretory granules require a constant supply of energy in the form of adenosine triphosphate (ATP) [13,59]. Islet glucose and lipids serve as energy nutrients to the islet beta cells which sense rising glucose to activate insulin secretion. As the surrounding systemic glucose levels rise within the islet, there will be an increased activity of the mitochondrial electron transport chain (ETC) to produce ATP, which results in the following sequence of events: The rise in the ATP/ADP ratio is responsible for the closing of the ATP-sensitive potassium (K+) channel, which results in the depolarization of the islet beta-cell plasma membrane and serves as a signal to open the voltage-gated calcium channels and this, in turn, is responsible for the exocytosis of insulin secretory granules and insulin [13,59]. If there is Mt dysfunction with decreased production of ATP, there will be less ATP generated and this will interfere with the proper secretion of insulin secretory granules and dysfunction of insulin secretion. Additionally, if the beta-cell mitochondria are further damaged due to increased RONSS and the RSI then the abnormally remodeled aberrant Mt (aMt) with loss of Mt matrix electron density and crista become leaky allowing cytochrome c to be released with activation of beta-cell caspases, which will result in beta-cell loss due to apoptosis (Figure 5E) (Figure 6) [13,59,87].

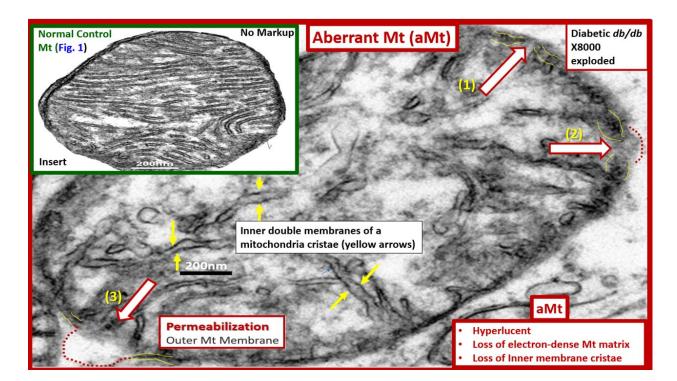


Figure 6. Aberrant Mitochondria (aMt) in the MetS Reloaded. This image depicts an exploded aberrant mitochondrion with an intact scale bar from a 20-week-old obese, insulin-resistant, diabetic *db/db* female preclinical mouse model. aMt are a common phenotype found in multiple different cells and organs in models of obesity, insulin resistance, metabolic syndrome, and type 2 diabetes mellitus as compared to control healthy electron dense C57BL/6J models (insert upper left). Thinning and interruption with permeabilization of the outer aMt membrane consist of 1) thinning of the inner and outer membranes (open red arrow); 2) interruption of the inner and outer membranes with partial permeabilization (open red arrow and red dashed line); 3) complete loss of the inner and outer membranes with permeabilization of the outer membrane (open red arrow, dashed red lines, and asterisk). The permeabilization of the aMt outer membrane allows leakage of multiple mitochondrial-derived toxicities including mitochondrial-derived reactive oxygen species (Mt-ROS), cytochrome c, and MtDNA. Original magnification x2000; scale bar = 200nm. This image is reproduced and adapted with permission by CC 4.0 [13].

Also, when both aMt and ER stress are found to co-occur within the pancreatic islet beta-cells there will be apoptotic signals from ER stress and the unfolded protein response (UPR) and cytochrome c leakage from aMt that signals caspases for the beta-cell to undergo apoptosis [59,63]. Anello et al, were able to demonstrate that human islet beta-cells showed an impaired insulin secretory response to glucose and an associated marked alteration of Mt function and morphology that was associated with an increased expression of uncoupling protein 2 (UCP2). UCP2 was thought to be due to increased glucose overload that was associated with a lower ATP, decreased ATP/ADP ratio, and a consequent reduction of insulin release [88]. Also, they found that mitochondria numbers were similar between the control and T2DM in beta cells; however, the volume of the Mt were statically increased and hyperlucent with loss of electron-dense mitochondria matrix and loss of cristae in their figure 5 [88], which this author has found in figure 5 and also found in the preclinical human islet amyloid polypeptide rat models, commonly referred to as the HIP model. Importantly, the accumulation of aMt and impaired mitophagy in islet beta-cells are associated with impaired synthesis of insulin secretory granules (paucity of ISGs figure 5) and deficient ATP production and thus a decrease in insulin secretion and the eventual loss of beta cells via apoptosis (Figure 5F).

Further, it has been recently proposed that there exists a critical interdependence between aMt dysfunction and T2DM that is found in the pancreatic islet β cells [89,90]. Indeed, T2DM is a

multifactorial and polygenic disease, which is associated with mitochondrial dysfunction due to aMt remodeling phenotypes. This central defect of aMt contributes to both insulin resistance and β -cell dysfunction and eventually β -cell failure via apoptosis in T2DM. Therefore, it is important to note that Mt dysfunction (aMt) and T2DM may be bidirectional. (Figure 7).

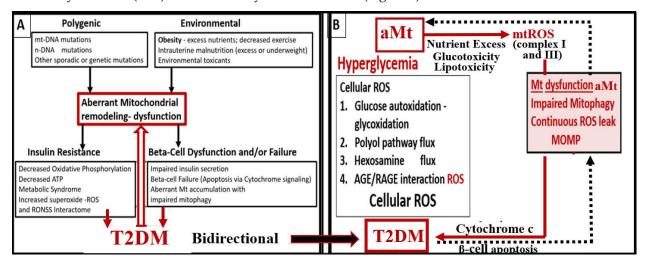


Figure 7. Type 2 diabetes mellitus (T2DM) is a multifactorial and polygenic disease in addition to having dysfunctional aberrant mitochondria (aMt) capable of bidirectional signaling. **Panel A** depicts that aberrant mitochondrial (aMt) phenotypes are a central and critical defect that leads to T2DM via multifactorial polygenic and environmental factors. **Panel B** demonstrates that aMt dysfunction with increased mitochondrial reactive oxygen species (mtROS) and T2DM are capable of interacting via bidirectional mechanisms. *ATP = adenosine triphosphate; AGE= advanced glycation end product; mt-DNA = mitochondrial deoxyribonucleic acid; MOMP = mitochondria outer membrane permeabilization; n-DNA = nuclear DNA; RAGE = receptor for AGEs; ROS = reactive oxygen species; RONSS = reactive oxygen, nitrogen, sulfur species.*

3.3. MetS Reloaded, Leptin, Hyperleptinemia, and LR

The discovery of leptin in 1994 provided a paradigm shift in how the adipose tissue is currently viewed (Figure 3A) [21,22]. The adipose tissue is no longer thought to be just an organ or tissue for fat and triglyceride storage, but as a metabolic, secretory, and endocrine organ. Adipose tissue is now considered to be an endocrine organ that is capable of secreting important adipocytokines/hormones such as leptin and adiponectin, which play an important endocrine function. Obesity results in hyperleptinemia and LR and decreased levels of adiponectin that are emerging as important players in the MetS reloaded as previously discussed in Section 3. [10,43– 48,50–55]. Leptin resistance occurs in the db/db mouse model and the Zucker fa/fa rat model due to a deficiency of the leptin receptor; however, in the BTBR ob/ob models there is only IR and LR does not occur due to leptin deficiency. LR is known to lead to glucose intolerance at least in the Zucker obese fa/fa rat model and occurs mainly due to hepatic glucose overproduction, and this occurs even prior to developing obesity [91]. Importantly, leptin is thought to be a predictive marker of MetS in humans [92]. Further, leptin and adiponectin are two of the most abundant adipokines secreted from adipocytes that have been implicated in the development of T2DM and CCVD-and are associated with MetS, VAT, IR, β-cell dysfunction, inflammation, arterial stiffness, and subclinical atherosclerosis at least in children [93]. In chronic obesity, it is commonly accepted that leptin elevated, while adiponectin becomes decreased and results in a decreased adiponectin/leptin ratio which predisposes to accelerated atherosclerosis and increased risk of CCVD [94]. Additionally, hyperleptinemia and hypoadiponectinemia are thought to be therapeutic targets of the MetS (Figure 8) [93,94].

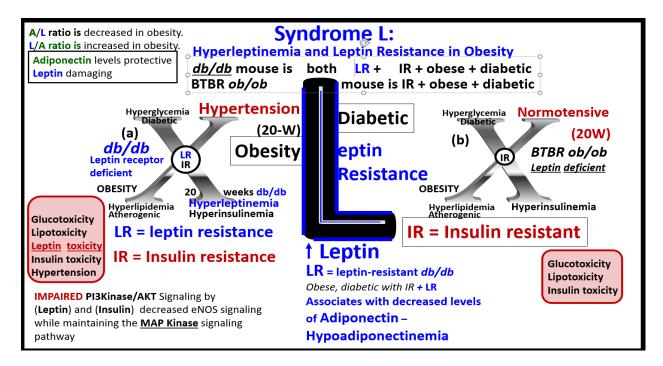


Figure 8. Syndrome L – hyperleptinemia and selective leptin resistance (LR) are emerging and novel risk factors in obesity, insulin resistance (IR), metabolic syndrome (MetS), and type 2 diabetes mellitus (T2DM). This figure compares the leptin receptor deficiency db/db with both leptin resistance (LR) panel (a) to the leptin deficiency BTBR ob/ob mouse models with IR that develops obesity and diabetes panel (b). LR is known to lead to glucose intolerance at least in the Zucker obese fa/fa rat models, db/db mouse models, and occurs mainly due to hepatic glucose overproduction that may occur even prior to developing obesity. LR is an emerging and novel finding in the MetS reloaded as previously depicted in figure 1. Importantly, its relationship to the MetS reloaded has increased over recent years. Leptin resistance is associated with decreased adiponectin and is proinflammatory, proatherosclerotic, and is associated with decreased insulin sensitivity. $AKT = \underline{protein\ kinase\ B};$ $MAP\ Kinase = mitogen-activated\ protein\ kinase;\ PI3\ Kinase = phosphoinositide\ 3-kinase$.

Notably, Leptin is also an emerging trigger for stroke [82,83,85,86,95–99] and is associated with obstructive sleep apnea [100].

Our global society now has one of the oldest populations in history [10,101] and it is well accepted that the prevalence of MetS increases with aging [52,53,99,101]. Interestingly, the prevalence of MetS increased 5-fold in aging women and only 2-fold in men [102]. Even though resistance to leptin is known to occur in aging [103], one must keep in mind that Leptin and Lr still remain incompletely understood and that we must continue to closely watch future research on leptin and how it applies to human clinical medicine. For example, it has recently been shared that leptin resistance during aging may be independent of fat mass in preclinical rodent modes [104]. Thus, in relation to the MetS, we must be cautious that we examine not only for age but also for gender differences [104]. Since leptin and adiponectin are the are two most abundantly produced adipokines secreted from adipocytes that play an important role in glucose homeostasis, inflammation, and accelerated atherosclerosis in chronic obesity, it is important to elaborate further on the important role of obesity-related adipokines/hormones, cytokines/chemokines, and toxic saturated FFAs [104,105].

4. MetS Reloaded, Essential Hypertension, and Vascular Stiffening

HTN may be defined as the elevation of blood pressure of unknown cause (in contrast to secondary hypertension with a known cause) and it usually clusters with other cardiovascular risk factors such as aging, obesity, IR, LR, diabetes, and hyperlipidemia in the MetS reloaded [106].

The MetS reloaded is associated with HTN and is one of the core features along with obesity, hyperlipidemia, hyperinsulinemia, IR, LR, and hyperglycemia (Figure 1). HTN is strongly associated with hyperinsulinemia, IR and LR (Figure 9) [106–110].

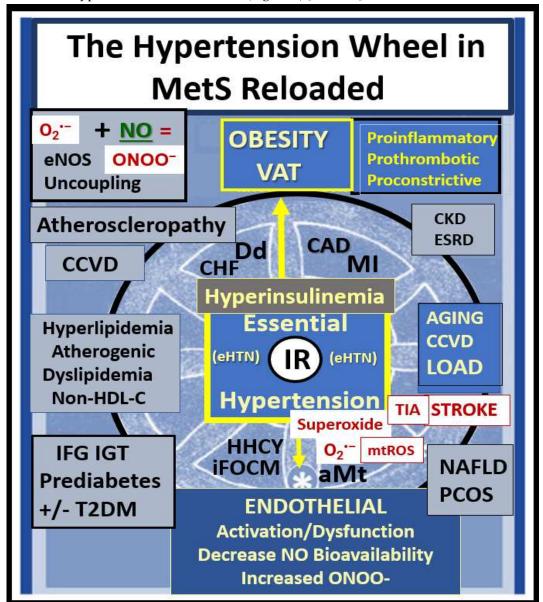


Figure 9. The essential hypertension (eHTN) wheel in the metabolic syndrome (MetS) reloaded. This wheel depicts that insulin resistance (IR) is a central core feature of the wheel as it is in the MetS reloaded. IR is central to the development of eHTN and IFG-IGT-T2DM, along with multiple metabolic and clinical conditions surrounding the outer portions of this hypertension wheel. Obesity is placed at the top of the wheel because it is believed to be the driving force behind the subsequent clinical end-organ remodeling and disease and the development of eHTN. Endothelial activation and dysfunction result in increased peroxynitrite (ONOO-) and decreased nitric oxide (NO) bioavailability. Peroxynitrite is generated by the reaction between superoxide and NO. One can note the multiple diseases that are associated with eHTN and the wheel depicts the interconnectedness between eHTN and the multiple disease states including vascular stiffening. Thus, IR, eHTN, and T2DM are not to be underestimated. Atheroscleropathy is a term that may be used when discussing accelerated atherosclerosis and macrovascular disease in those individuals with T2DM and the MetS reloaded. The wheel was chosen as a background icon because it goes round and round and over time it just keeps on turning and results in vascular stiffening and endorgan damage in the heart-brain-kidney axis that has high capillary flow with low resistance and increased vulnerability to the increased pulse wave velocity associated with vascular stiffening, HTN,

and microvascular disease. CAD = coronary artery disease; CCVD = cerebro-cardiovascular disease; CHF = congestive heart failure; CKD = chronic kidney disease; Dd = diastolic dysfunction; eNOS = endothelial nitric oxide synthase; ESRD = end-stage renal disease; IFG = impaired fasting glucose; IGT = impaired glucose tolerance; LOAD = late-onset Alzheimer's disease; MI = myocardial infarction; mtROS = mitochondrial reactive oxygen species; $O2^{\bullet -} = superoxide$.

Leptin's effect on HTN is mainly through its vascular, renal, and sympathetic nervous system actions [108]. The pathophysiology for the development of HTN in the MetS reloaded is very complicated and consists of a complex network of interconnected pathways. While some of these pathways still remain incompletely understood the following figure provides further insight as to how this complex network is interconnected with the various components of the MetS reloaded (Figure 10) [65,104,106–114].

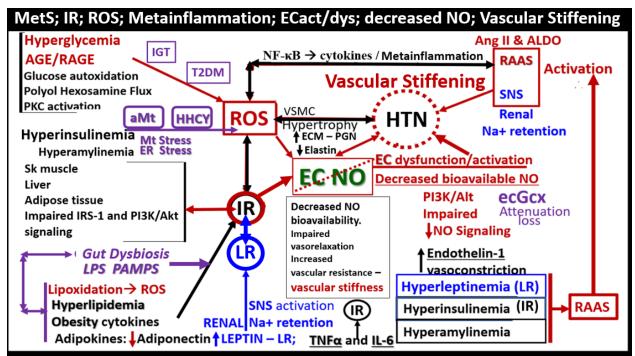


Figure 10. Pathophysiology of essential hypertension (HTN) in the MetS reloaded. This figure demonstrates the complex interconnected pathways between insulin resistance (IR encircled in red color) and HTN. IR is the cornerstone of the MetS reloaded and this schematic depicts how it is related to the development of HTN (dashed red circle). Note the important role of endothelial derived nitric oxide (NO) (green), which is essential for proper vascular relaxation and homeostasis and how it is negated (red-dashed diagonal line) in the development of HTN. Importantly, decreased bioavailable NO results in decreased vasorelaxation and increased vascular resistance with a resulting increase in vascular stiffening and elevated blood pressure (BP). Next, note the devastating role that reactive oxygen species (ROS) has on NO and endothelial cell dysfunction and activation resulting in decreased vasorelaxation, increased vascular (Vasc) resistance, blood pressure elevation, vascular stiffness, and the role they each play in the development of HTN. Activation of the renin-angiotensin-aldosterone system (RAAS) is also very important to the development of HTN as well as the activation of the sympathetic nervous system (SNS). Importantly, note the involvement of the Leptin – Leptin resistance (LR) pathway in blue coloring. Also, note the emerging roles of the endothelial glycocalyx (ecGCx), aberrant mitochondria (aMt), hyperhomocysteinemia (HHCY), and gut dysbiosis, which are related to inflammation via lipopolysaccharide (LPS) as these are emerging roles not only the MetS reloaded but also in the development of HTN (colored in purple). The sum of these interacting pathways and functional changes are associated with vascular wall remodeling and vascular stiffening. This image adds additional information regarding some of the possible mechanisms in the development of HTN presented in figure nine because it focuses more on the involved possible mechanistic pathways in the development of HTN. Not shown is the

relationship between the sodium/potassium ATPase enzyme that is redox-sensitive and the excessive redox stress in the MetS is currently thought to also promote a salt-sensitive HTN due to the sodium/potassium ATPase enzyme inhibition. While some of these mechanisms may seem redundant, they are actually complementary to figure nine. The inverse relationship between leptin and adiponectin also plays an important role, because when leptin is elevated adiponectin is decreased, and thus, there is a decrease in the A/L ratio that is associated with the loss of adiponectin's protective vascular role. Importantly, systolic HTN predominates in the MetS reloaded. AGE/RAGE = advanced glycation end products and receptor for AGE; Akt = protein kinase B; ALDO = aldosterone; Ang II = angiotensin II; ECM = extracellular matrix; IGT = impaired glucose tolerance; IL-6 = interleukin-6; LPS = lipopolysaccharide; Na + = sodium; NF-κB = nuclear factor kappa B; PAMP = pathogen associated molecular pattern; PGN = proteoglycan; PI3K = Phosphoinositide 3-kinase; PKC = protein kinase C; Sk = skeletal; T2DM = type 2 diabetes mellitus; TNFα = tumor necrosis factor-alpha; VSMC = vascular smooth muscle cell.

In regards to vascular-arterial stiffening, Lopes-Vincente et al, determined that vascular or arterial stiffening as measured by pulse wave velocity in the carotid-femoral segments was already present in a cohort of individuals with newly diagnosed MetS as compared to those without MetS. Further, they found that aging and the number of risk factors (greater than the three required to diagnose MetS) were associated with increased vascular stiffness [115]. Also, Schillaci et al. were able to show that the MetS influences arterial functional properties that are related to cardiovascular risk and provide a pathophysiological framework for understanding the associations between MetS and vascular stiffness related to cardiovascular morbidity and mortality [116]. These findings are not surprising since the pathophysiology for the development of HTN in the MetS was associated with so many interacting pathways (Figs. 9, 10) involved with alterations in vascular functions and the known association with vascular remodeling as in atherosclerosis discussed in section 2. Importantly, the Framingham Heart Study has confirmed that vascular stiffness is an independent predictor of cardiovascular morbidity and mortality in the general population, hypertensive individuals, and the elderly [117]. Additionally, vascular stiffness is known to be a consequence of pathophysiological alterations involving ECs, vascular smooth muscle cells (VSMCs), extracellular matrix (ECM), and elastin (Figs. 9,10) [118-120]. Additionally, Yan et al. have proposed that HTN in the MetS is a form of salt-sensitive HTN that is largely based on the dysfunctional Na/K-ATPase enzyme that is redox sensitive. Also, there is known to be a lack of IR in the kidney and there is increased renal sodium reabsorption due to hyperinsulinemia in the MetS that leads to sodium retention and the resultant salt-sensitive HTN [121,122].

Notably, obesity and the hypothalamic-pituitary-adrenal (HPA) axis dysfunction are implicated in RAAS activation in the MetS reloaded. Previously, in figure 1 the yellow line that extends downward from boxed-in HPA axis dysfunction to the arrow indicating RAAS activation of Ang II and aldosterone. In turn, this is responsible for the sympathetic nervous system activation. Previously, our group has found considerable remodeling within the adrenal gland zona glomerulosa cells of the diet-induced obesity Western models at 12 weeks. The remodeling changes consisted of zona glomerulosa hypertrophy, hyperplasia, capillary dilation, and endoplasmic reticulum stress [10].

5. MetS Reloaded, Hyperglycemia, Mitochondrial Dysfunction, and Importance of the Reactive Oxygen Species Interactome (RSI)

There is a very strong overlap between IFG and IGT in the MetS reloaded and this points to the important role of hyperglycemia in the upper left-hand side of the MetS reloaded (Figure 1) [123]. T2DM is a multifactorial polygenic disease that may be characterized as a chronic metabolic-endocrine disorder that associates with insulin resistance or relative lack of insulin or insulin deficiency and thus, hyperglycemia and is a constant finding in IFG-IGT and T2DM. The resulting glucotoxic state promotes oxidative stress and chronic inflammation [123]. Importantly, T2DM is a risk factor for macrovascular (accelerated atherosclerosis and vascular stiffness) [14] and microvascular end-organ pathologies such as retinopathy [124], neuropathy [125], nephropathy [126],

vasculopathy-intimopathy [14], isletopathy [127], accelerated atherosclerosis - atheroscleropathy [14], cardiomyopathy [128], and diabetic encephalopathy-cognopathy (cognitive dysfunction), which is associated with an increased risk for age-related neurodegenerative diseases such late-onset Alzheimer's disease (LOAD) [8]. Importantly, macrovascular and microvascular diseases are also at the crossroads of T2DM and eHTN [129]

IFG-IGT and T2DM result in hyperglycemia, which is associated with increased ROS production via the following six pathways that increase ROS: 1) glucose autoxidation; 2) polyol flux; 3) hexosamine flux; 4) advanced glycation end product(s) (AGE); 5) increased expression of the receptor for AGEs (RAGE) and AGE/RAGE interactions; 6) increased protein kinase C and mitochondrial overproduction of ROS in macrovascular endothelial cells by the increasing FFA flux and oxidation [121,122]. Additionally, the MetS reloaded is associated with leaky aMt, which leak superoxide and contributes to the overall ROS, which interacts with RSI consisting of RONSS to result in excessive oxidative stress. This cumulative increase in oxidative stress is responsible for the inactivation of two critical antiatherosclerotic enzymes eNOS and prostacyclin synthase and defective angiogenesis in response to ischemia. This oxidative stress also activates a number of proinflammatory pathways which enables ROS to induce inflammation via increased stimulated nuclear factor-kappa B (NF- κB) and downstream cytokines/chemokines, and inflammation to induce ROS, which creates a vicious cycle. These pathways are also responsible for long-lasting epigenetic changes that continue to drive the persistent expression of proinflammatory genes even after glycemia is normalized and thus important in creating hyperglycemic memory [129,130]. There are several lines of evidence, which indicate that all of the six previous listed causes for increased ROS in hyperglycemia are activated by a single important upstream event and that is the mitochondrial excess production of mitochondria ROS (mtROS) superoxide (*O₂-) [128–131]. Importantly, aMt leak superoxide or mitochondria ROS (mtROS), which would induce deoxyribonucleic acid (DNA) strand breaks and activate nuclear poly (ADP-ribose) polymerase resulting in decreased glyceraldehyde 3-phosphate dehydrogenase (GAPDH) and subsequently increase the previous causes for increased oxidative stress in the MetS reloaded (pathways1-6) [13,118,129-131].

Over the past 17 years, the identification of aMt have been a constant recurrent unifying finding utilizing TEM in multiple organs including the endothelium [13, 47, 48, 65t, 126] identified in our obese, MetS, IR, LR, IGT, and diabetic preclinical rodent models with hyperglycemia. The organs involved with aMt include the kidney [13,44,126,132,133], skeletal muscle [13,44,133], cardiac muscle [13,44,86,112,134,135], liver [112], aorta [13,136,137,139], VSMCs [13,44,131,132], pancreatic islets [44, 59. 62, 66, 79] and brain [4,7,8,10,11,47–50,138,139]. Additionally, there are two recent topical reviews of Mt as unifying organelles and metabolic hubs in multiple organs of obesity, IR, MetS, and T2DM for those who wish to view additional ultrastructural images of normal Mt and aMt [13] and the role of the Mt in the MetS [89].

Brownlee et al, have previously noted that the upstream aMt and the increased mtROS may be extremely important due to the additive damage from hyperglycemia-induced ROS [130,131]. This is especially important when there are excessive mtROS due to leaky aMt. Indeed, Mt remodeling to an aberrant and leaky phenotype (aMt) with Mt dysfunction is a common thread that weaves across the multiple organ systems in obesity, IR, LR, the MetS reloaded, and the T2DM mosaic fabric of disease. aMt contribute to the overall increased oxidative stress and will interact with RONSS and the RSI to further increase and contribute to the overall oxidative stress in the MetS and T2DM. Since FOCM is known to be compartmentalized to the mitochondria, cytosol, and nucleus, it is important to next examine the role of impaired FOCM in the MetS [20].

5.1. MetS Reloaded and Impaired Folate-Mediated One-Carbon Metabolism (FOCM) in the Metabolic Syndrome Reloaded

FOCM is a complicated metabolic network of interdependent biosynthetic pathways and cycles that are known to be compartmentalized in the cytoplasm, mitochondria, and nucleus (Figure 11) [20].

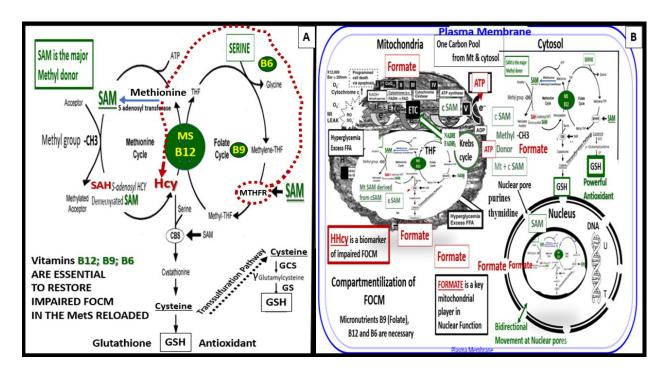


Figure 11. Folate-Mediated One-Carbon Metabolism (FOCM). FOCM involves multiple complex interacting cycles within the cytosol, mitochondria, and nucleus in the metabolic syndrome (MetS) reloaded. **Panel A** illustrates both the folate and methionine interdependent cycles and supports the importance of the methyl donor S-adenosylmethionine (SAM) as well as demonstrating the importance of the essential B12, B9, and B6 vitamins. **Panel B** illustrates the compartmentalization of FOCM in the cytosol, mitochondria, and nucleus. Additionally, note the importance of formate being transferred from the mitochondria to the nucleus, as well as *S*-adenosylmethionine (SAM) via nuclear pores. This figure is modified and adapted with permission by CC 4.0 [20].

Importantly, methionine and tetrahydrofolate (THF) are derived primarily through the dietary intake to supply the methionine and folate cycles and that the enzyme methionine synthase (MS) and its essential cofactor vitamin B12 are placed in a central position of the interconnected folate and methionine cycles (Figure 12). Also, vitamin B12 (cobalamin) is of the utmost importance for the demethylation of HCY to methionine [12,140]. If B12 or MS are deficient or impaired there will be elevations of the clinical biomarkers of impaired FOCM with resulting elevations of HCY with ensuing HHCY, and elevations of methylmalonic acid. Additionally, vitamin B12 is also important in the production of succinyl Coenzyme A that is necessary for the proper function of TCA/Krebs cycle. Folic acid (folate or vitamin B9) is important for the FOCM along with B12 to properly supply Succinyl-CoA, while glycolysis provides oxaloacetate from pyruvate to the tricarboxylic acid TCA cycle to provide NADH and FADH2 to the ETC to produce ATP by the mitochondria [20,140]. Also, note that when there is an intact FOCM metabolic state, Hcy can be either demethylated to methionine via the methionine synthase with its essential cofactor vitamin B12 or enter the cystathionine beta synthase pathway via an intact vitamin B6 pathway [20]. The ETC generates ATP via complexes I-V to generate the energy currency of ATP. Importantly, if there are excess nutrients in the form of glucose, sucrose, fructose, and fat delivered to the ETC at complex I, there will be a generation of excessive ROS in the form of superoxide (*O₂-) or mtROS. An excess of mtROS will then interact with extra-mitochondrial or cytosolic nitrogen, and sulfur to form RONSS, which induces the RSI, wherein ROS-induced ROS release (RIRR) comes into play and will accelerate RONSS RSI and redox stress. Importantly, note that if the methyltetrahydrofolate reductase (MTHFR) enzyme (encircled with a red-dashed line) becomes dysfunctional, HCY in the methionine cycle will also become elevated as depicted by the red-dashed line from MTHF to HCY (Figure 12 panel A) [20].

Also, formate plays a central and formidable role in providing proper nucleus function and maintenace of its cellular structure in homeostatic conditions. Serine, glycine, methionine, and choline are necessary substrates to provide formate in order to provide proper mitochondrial function to the nucleus to produce purines, thymidylate and methionine to fulfil their role in the nucleus. Formate is primarily produced within the mitochondria and then secreted to enter the nucleus via nuclear pores in health to provide FOCM to the nucleus for proper chromatin, histone modeling and normal function [20]. Deoxythymidine monophosphate synthesis occurs in the cytosol, nucleus, and mitochondria, whereas purine synthesis and methionine synthesis take place within the cytosol. Mitochondrial FOCM generates formate for cytosolic and nuclear FOCM and biosynthetic precursors for mtDNA synthesis and mitochondrial protein translation. Thymidylate synthase converts deoxyuridine monophosphate to deoxythymidine monophosphate (dTMP) in a 5,10-methylene-THF-dependent reaction. It is important to note that mitochondrial SAM (Mt SAM) is derived from cytosolic SAM. Additionally, the Krebs cycle also resides within the mitochondria and provides reduced nicotinamide adenine dinucleotide and the reduced flavin adenine dinucleotide to the ETC for ATP production [20].

Folate (folic acid, essential vitamin B9) belongs to a family of enzyme cofactors that carry chemically activated 1-carbon units to formate, formaldehyde, and methanol. Folate 1-carbons are necessary for the synthesis of purine, thymidylate, and the remethylation of HCY to methionine. Further, methionine is an essential amino acid that is utilized in protein synthesis that can also be adenosylated to S-adenosylmethioinine (SAM), which is responsible for the methylation of protein cytosine bases of DNA (important for nuclear chromatin structure), neurotransmitters, phospholipids and other protein molecules (Figure 11) [20,140].

FOCM (a metabolic network of interdependent biosynthetic pathways) is important for physiologic maintenance and proper cellular homeostasis due to its role in purine and thymidylate because it is compartmentalized in the nucleus, mitochondria, and cytosol. However, it becomes impaired in the hyperglycemia (IFG, IGT, and T2DM) of the MetS reloaded [20,140-144]. Additionally, Hcy is known to be elevated in T2DM and is associated with HHCY, which may be considered a clinical biomarker for impaired FOCM [20, 140, 142, 143, 144, 145]. HHCY would also add to the existing six factors for increased ROS production due to hyperglycemia and the aMt [13,130,131,136]. Hyperglycemia induced ROS could interact with not only the aMt (mt ROS) but also the HHCY induced ROS, which could subsequently interact with RONSS of the RSI and these multiplicative ROSs could form a situation, in which ROS becomes self-perpetuating that is a condition of ROS-induced ROS release creating a vicious cycle of excessive oxidative stress to cells and tissues in IFG, IGT, and T2DM [20,146]. The folate and methionine cycles and the transsulfuration pathway are all extremely important not only in the mitochondria but also in the cytosol and nucleus [20]. In order for the normal functioning of the FOCM, there must be an adequate supply of water-soluble B vitamins including B12, B9 (folic acid or folate), and B6 (pyridoxal 5'-phosphate) as they relate to FOCM (Figs. 1, 11).

Interestingly, an intact properly functioning FOCM metabolic pathway with adequate amounts of B vitamins is now being considered to play the role of providing antioxidant roles at least in stroke [147]. It is known that folate (B9) is capable of aiding in the role to prevent complete BH4 oxidant stress to this cofactor necessary for the production of endothelial-derived NO. Additionally, the FOCM metabolic pathway is necessary to convert the potent oxidant of HYC – HHCY via the cystathionine beta-synthase enzyme (CBS) with adequate amounts of vitamin B6 and serine to generate glutathione via the conversion of cystathionine to cysteine to GSH a potent cellular derived intracellular antioxidant (Figure 11). Also, cysteine may enter the transsulfuration pathway. Indeed, the FOCM pathways are far reaching and play multiple important roles in providing proper cellular homeostasis [20]. FOCM is important to metabolize HCY (a potent oxidant) to methionine, which when elevated is a known risk for developing conditions such as stroke and cellular redox stress. In preclinical animal models and even human individuals, when increasing the B-vitamins it is possible to lower elevated Hcy; however, utilization of nutrient therapies such as B vitamins to improve the ischemia associated with acute stroke has yet to be convincingly demonstrated [147].

Impaired FOCM and associated HHCY are associated with multiple common clinical diseases including the developmental anomalies of neural tube defects; CCVDs including stroke and cognitive decline, and intestinal malignancies (Figure 12) [20,146].

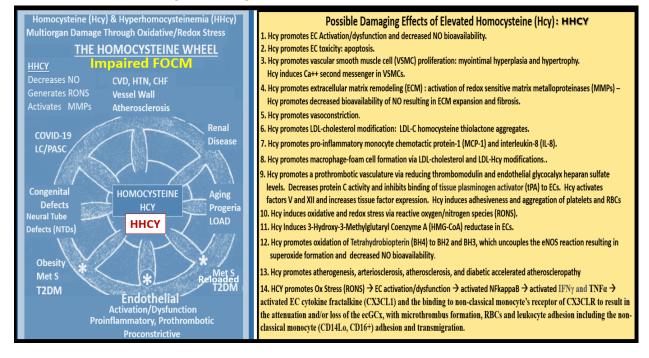


Figure 12. Impaired folate-mediated one-carbon metabolism (FOCM) and associated hyperhomocyteinemia (HHCY) are associated with multiorgan damage and multiple clinical diseases via the possible damaging effects of excessive redox stress. Once impaired FOCM develops and goes unchecked this homocysteine wheel begins turning and just keeps on turning causing damage to various organ and tissue systems identified and results in multiple clinical disease states on the lefthand side of this image. Once impaired FOCM has developed and HHCY is established, HCY is capable of undergoing autoxidation, formation of Hcy mixed disulfides, interaction of Hcy thiolactones and protein homocysteinylation reactions, which result in damage and dysfunction to proteins, lipids, and nucleic acids to result in at least the 14 damaging effects listed above on the righthand side of this image. This image is provided with permission by CC 4.0 [17]. CHF = congestive heart failure; COVID-19 = coronavirus disease-19; CVD = cebro-cardiovascular disease; EC = endothelial cell; $ecGCx = endothelial\ cell\ glycocalyx;\ eNOS = endothelial\ nitric\ oxide\ synthase;\ HTN = hypertension;\ IFN\gamma =$ interferon gamma; NO = nitric oxide; LDL = low-density lipoprotein cholesterol; MetS = metabolic syndrome; $MMPs = matrix\ metalloproteinases;\ NFkappaB = nuclear\ factor\ kappa\ B;NTD = neural\ tube\ defects;\ LC/PASC$ = Long COVID/post-acute sequelae of SARS-CoV-2; LOAD = late-onset Alzheimer's disease; Ox = oxidative stress; RBCs = red blood cells; RONS = reactive oxygen nitrogen species; T2DM = type 2 diabetes mellitus. $TNF\alpha$ = tumor necrosis factor alpha.

In the general population, mild to moderate HHcy occurs in approximately 5–7% [148Welch]. Importantly, there are known to be genetic mutations in human individuals involving the methyltetrahydrofolate (MTHFR) gene, which will result in the impairment of its function and thus allow HCY to accumulate and result in HHCY. This is especially true if there are deficiencies of the essential B vitamins (B9 -folate, B12, and B6) (Figure 12). The most common human genetic variant of the MTHFR gene to date is the primary homozygous C677T (T677T) and the secondary compound heterozygous A1298C + C677T. This genetic variant occurs in 10-15% of the general population and is likely to also occur in similar percentages in the MetS population [149–151].

6. MetS Reloaded and Endothelial Activation (ECact) and Dysfunction (ECdys)

The thin protective monolayer of cells that line the luminal side of blood vessels are known collectively as the endothelium and are known to a play an important and complex role in vascular

biology. The endothelium and its endothelial cell(s) ECs are key to vascular hemostasis, tone, leukocyte recruitment, hormone trafficking, and fluid movement from the blood to the interstitial space [152]. Endothelial cell activation (ECact) may be defined by the endothelial expression of cellsurface adhesion molecules, such as VCAM-1, ICAM-1, and E-selectin. EC dysfunction (ECdys) may be defined as the decreased synthesis, release, and/or activity of endothelium-derived NO that results in decreased bioavailable NO [65,152,153]. Notably, ECact and ECdys are tightly related and share a bidirectional relationship and lead to vascular disease and atherosclerosis by inducing a proconstrictive, vascular smooth muscle cell proliferation, and a proinflammatory state with leukocyte adhesion, platelet aggregation, lipid oxidation, and matrix metalloproteinase (MMP) activation [152]. ECact is typically induced by 1) proinflammatory cytokines, such as TNF- α and IL-6; 2) turbulent blood flow such as occurs at bifurcations and branch points of arteries; 3) AGEs, which are elevated in hyperglycemia and aging, and 4) inflammatory stressors such as metainflammation and plasma membranes peptides of gram-negative bacteria such as LPS. These four functions are each important mediators of ECact via the activation of the nuclear transcription factor of EC NF-κB and are known to be mediators of ECact. The following TEM images are examples of brain EC (BEC) activation (Figure 13) [6, 8, 10,14. 16. 19, 20, 47. 48. 49, 49. 45, 46, 48, 59, 65].

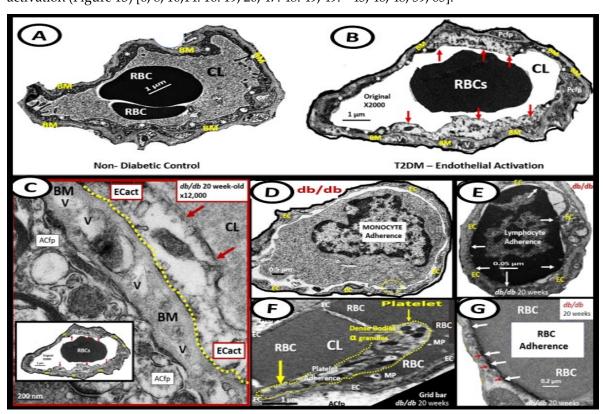


Figure 13. Examples of transmission electron microscopic (TEM) images for endothelial cell activation (ECact). **Panel B** depicts the thickened electron-lucent areas (red arrows) of ECact as compared to control models in panel A. **Panel C** depicts basement membrane (BM) thickening with increased vacuoles (V) and vesicles (v). **Panels D and E** depict monocyte (D) and lymphocyte (E), platelet (F) and red blood cell (RBC) adhesion (G) to the activated ECs respectively. Original magnification x2000; scale bar = 1μ m and are modified with permission CC by 4.0 [65]. Images (panels C-G) are reproduced and modified with permission by CC 4.0 [10,47,48,65]. Magnifications and scale bars vary. ACfp = astrocyte foot processes; Cl, capillary lumen; EC = endothelial cells; ECact = endothelial cell activation; MP = microparticle of the platelet.

Since 2012, author has observed multiple endothelial (systemic and brain endothelial cell (BEC)) ultrastructural transmission electron microscopic (TEM) remodeling changes (Table 1) [10,13,47–50,65,136,154–156].

 Vascular endothelial cell thickening that may partially be due to increased transcytosis with increased transcytotic vesicles, macropinocytotic vesicles with complications that are increased BEC permeability and pericapillary edema.

nitrogen, sulfur species; ROS = reactive oxygen species; RSI = reactive species interactome; W = Western mice.

- 2. Plasma membrane ruffling with associated microparticle, microvesicle, and extracellular vesicle exosome formation
- 3. Plasma membrane microparticles-microvesicles and extracellular vesicle exosome (EVexo) formation.
- Increased aberrant mitochondria (aMt) with loss of electron dense Mt matrix and crista. aMt are leaky and increase
 mitochondria-derived reactive oxygen species (mtROS). Increased BEC Redox stress increased RONSS RSI
 interactome.
- 5. Oxidative stress due to mtROS, increased NADPH Ox and decreased antioxidant reserve due to aMt.
- 6. Increased endoplasmic reticulum (ER) stress with widened and swollen ER.
- 7. Increased Transcytosis. Increased transcytotic vesicles and macropinocytosis.
- 8. Electron lucency of ECs (due to increased vesicles vacuoles i.e. macropinocytosis along with increased transcytosis of transcytotic vesicles.
- Attenuation and/or loss of the endothelial glycocalyx (ecGCx) that have only been identified in obese, diabetic BTBR ob/ob models to date.
- 10. Increased inflammatory receptors I-CAM and V-CAM (proinflammatory). Plus, the adage that ROS begets inflammation and Inflammation begets ROS. This can be detected in activated BECs by increased WBC adherence to the endothelium.
- 11. Prothrombotic by multiple mechanisms, visualized by observing RBC and platelets adhering to the endothelium.
- 12. Basement membrane thickening (vacuolation/vesiculation) may or may not be present; however, if it is present it strengthens the case for endothelial activation.
- 13. EC stiffening. Apical luminal rearrangement of actin fibers in DIO W, db/db, and ECMR -/- models. Quantitation in all three models.
- 14. EC contraction shortening. DIO W, db/db, and ECMR -/- Quantitation in DIO and db/db models.

ECdys is defined as the decreased synthesis and activity of endothelium-derived nitric oxide (NO), which is frequently referred to as decreased NO bioavailability [65,152]. We now know that ECact and ECdys are closely linked in a response to molecular and structural injury to the arterial vessel wall such that a decrease in the bioavailability or loss of NO can lead to ECact and in a like manner ECact can cause ECdys and thus, the close linkage between these two processes [13,65,152]. Additionally, we now know that NO has a multitude of positive effects on the vascular wall, which includes its anti-inflammatory, antithrombotic, anti-atherosclerotic effects, and vasodilation properties [65,136,141,152]. This places endothelium-derived NO at the very central core for maintaining vascular homeostasis. If there is decreased bioavailable NO, vascular homeostasis will be attenuated and/or lost. Further, it is also known that there are shared events between ECact and ECdys that are closely linked [12,65,146]. Importantly, each of the four arms of the MetS reloaded (obesity - hyperlipidemia, hyperinsulinemia - IR, HTN, and hyperglycemia) is known to be associated with decreased bioavailability of NO (ECdys) either directly or indirectly that may result in ECact [14,65,116,147]. IR results in decreased signaling of the eNOS enzyme and its activation. Thus, with this combined of IR and excessive nutrient supply via excess glucose, sucrose, or fructose will increase mtROS via the electron transport chain. Likewise in manifest T2DM there will be an associated increase in aMt that will leak mtROS – superoxide and there will be excessive oxidation of the eNOS enzyme cofactor tetrahydrobiopterin (BH4) that ultimately uncouple the eNOS production of NO (eNOS enzyme uncoupling) that will decrease endothelial-derived NO bioavailability [13, 14 IRS, 16, 59, 65, 140, 152]. eNOS is the endothelial constitutive and rate-limiting enzyme that is responsible for the conversion of L-arginine to NO and L-citrulline. eNOS requires the cofactor tetrahydrobiopterin (BH4) [14,16,59,65,140,152,157]. Reduced BH4 or the substrate L-arginine result in the uncoupling of the eNOS enzyme, which results in decreased bioavailable of NO and increased

production of superoxide and ECdys. Causes for this uncoupling include increased superoxide and peroxynitrite (ONOO-), glucotoxicity, small dense LDL-C, oxidized LDL-C, HHCY, increased mtROS, decreased L-arginine, increased asymmetric dimethylarginine (ADMA), and highly sensitive C reactive protein plus others that promote oxidative stress to the endothelium [14,16]. In physiologic homeostasis the endothelium is a net producer of NO; however, in obesity, IR, and especially T2DM the endothelium may become a net producer of superoxide unless blood glucose is optimally controlled. When there is an excessive production of superoxide as in T2DM there develops a decrease in the ratio of NO/ROS [157–160]. Indeed, reduced endothelial NOS enzyme activity and eNOS uncoupling result in decreased NO bioavailability and may be considered as the main factors underlying endothelial dysfunction that occurs in the MetS reloaded (Figure 14) [14,65,158–160]. The mitochondria and production of NO are definitely closely related in the MetS reloaded and that is why the aMt are placed centrally and flanked with IR and LR (Figure 1) [160].

In T2DM as discussed previously there is an increase in aMt, which suggests that the excess glucose may be responsible for the phenotypic ultrastructure of the aMt and causes increased production of mtROS and superoxide. Also, the remodeled aMt that are frequently found in T2DM are leaky and allow an even greater release of mtROS (Figure 6). This increase in mtROS will in turn produce eNOS uncoupling and decreased NO bioavailability to vascular tissues [161,162]. IR and LR that are central to the MetS reloaded is coupled to the function of the Mt and also Mt dysfunction is coupled to IR and LR. This complex interaction is reflected by a finding from Peterson et al, as they found that IR in the elderly population was associated with decreased mitochondrial ATP production, a reduction of Mt RNA, and decreased activity of oxidative phosphorylation complexes [163]. The association of IR, LR, MetS, and T2DM are related to the function of the Mt; however, it is still not completely known as to whether Mt dysfunction results from or causes IR [11,13,14,160– 167]. In our studies in obese, insulin-resistant, and diabetic models we have found the following abnormalities in Mt such as impaired Mt biogenesis, Mt fragmentation, increased fission, and impaired mitophagy with the accumulation of aMt. The constant and persistent finding in these models of aberrant Mt seemed to be a constant and unifying theme of our ultrastructure findings, which strongly suggested impaired mitophagy [13,14,43,48,124,132–135]. Indeed, Mt dysfunction is implicated in the development of IR, LR, and T2DM and plays a very significate role in the development of diabetic complications.

6.1. MetS Reloaded and the Endothelial Glycocalyx

The endothelial glycocalyx is often referred to as the endothelial surface coating layer and consists of a unique extracellular matrix (ECM) that acts as an initial barrier between the vascular wall and its luminal contents. Lanthanum nitrate perfusion fixation staining of the ecGCx has recently been found in our laboratory to be both reliable and reproducible as well as others who are utilizing this technique (Figure 14) [45,168–171].

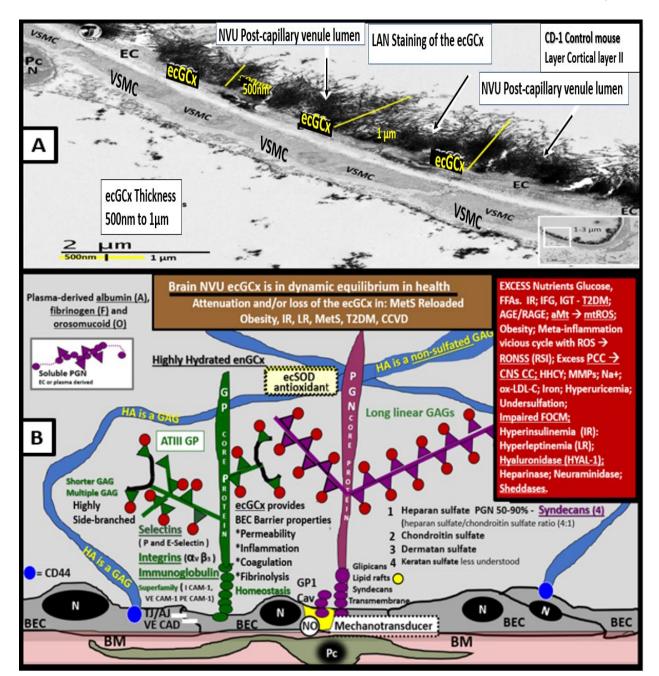


Figure 14. Electron microscopic view of lanthanum nitrate staining of the endothelial glycocalyx (ecGCx) and an illustration of the (ecGCx) in the metabolic syndrome reloaded. demonstrates the normal Lanthanum nitrate (LAN) perfusion fixation staining of the endothelial glycocalyx (ecGCx) in a post-capillary venule. This exploded image demonstrates the normal staining of the ecGCx in a healthy control male CD-1 mouse brain from the frontal cortical grey matter layer III with the original scale bar of 2µm intact. Note the intense electron-dense staining of lanthanum nitrate of the apical brain endothelial cell(s) (BECs) for the ecGCx such that one cannot make out the structural content of the ecGCx that is revealed in the following panel B illustration. Note the boxed-in insert in the lower right-hand corner (scale bar = 2µm) with white outline, which is the original image, from which the exploded image in panel A is derived. Scale bar (black) = 2µm and yellow scale bars = 500nm. Panel B illustrates the various proteoglycans (PGNs) (purple), glycoproteins (GPs) (green), hyaluronan (HA) (blue), glycosaminoglycans (GAGs) purple and green triangles), and their sulfation (red circles). Panel A is an original image recently acquired by author that has not been previously published and is presented only for its educational purposes. Panel B is a modified and adapted image with permission by CC 4.0 [6,10,13,45,65]. A = albumin; AGE/RAGE = advanced glycation end BM = basement membrane; CAD = cadherin; CAM = cellular adhesion molecule;

CD44 = cluster of differentiation 44; EC = endothelial cell(s); ecSOD = extracellular superoxide dismutase; F = fibrinogen; FGF2 = fibroblast Growth Factor 2; FOCM = folate-mediated one-carbon metabolism; GCx = glycocalyx; ICAM-1 = intercellular adhesion molecule; Ox LDL = oxidized low-density lipoprotein; LPL = lipoprotein lipase; MMPs = matrix metalloproteinases; N = nucleus; Na+ = sodium; O = orosomucoids; Pc = vascular mural cell pericyte(s); PECAM- 1= platelet endothelial cell adhesion molecule-1. RONS = reactive oxygen species; VEC = vascular endothelial cell(s); TFPI = tissue factor pathway inhibitor; TJ/AJ = tight and adherens junctions; VCAM = Vascular cell adhesion protein; VE CAD = vascular endothelial cadherins; VEGF = vascular endothelial growth factor; XOR = xanthine oxioreductase.

The ecGCx is vasculoprotective and acts as the first barrier of a tripartite BBB, including *i*) ecGCx; ii) Brain endothelial cell(s) (BEC) and its basement membrane (BM) of the NVU; iii) BEC BM and astrocyte foot processes or end feet (ACfps) of the extravascular compartment [172] of the NVU. The neurovascular unit (NVU) consists of 1) ecGCx; 2) BEC; 3) abluminal BM, pericytes (Pcs) and pericyte foot processes (Pcfps) within the BM, and ACfps in addition to the paracellular tight and adherens junctions (TJ/AJs) between each BEC consisting of occludins, claudins, and junctional adhesion molecules in addition to vascular endothelial cadherins (VE-Cadherins) [20,23,65,173]. The BECs are very specialized and their ecGCx are thicker and more continuous when compared to the heart and lung capillary ECs [169]. The BECs are responsible for the synthesis of not only the luminal ecGCx (with contributions from plasma albumin, orosomucoids, fibrinogen, glycolipids, and glycoproteins) but also the synthesis of the abluminal ECM of the BM [16,23,65,173,174]. Additionally, BECs are primarily responsible for the synthesis of the TJ/AJ, JAMs, and VE-Cadherins that form the paracellular barrier of the BBB. The ecGCx is primarily synthesized by BECs with some contributions by plasma albumin, orosomucoids, fibrinogen, glycolipids and glycoproteins [10,13,20,23,65,173]. The ecGCx is anchored to the BEC luminal plasma membranes by highly sulfated proteoglycans (syndecans and glycipans), glycoproteins (including selectins such as various cellular adhesion molecules and integrins), and non-sulfated hyaluronan (a glycosaminoglycan) via BEC cluster of differentiation 44 (CD44). Hyaluronan may also be free-floating (unbound) or attached to the assembly proteins such as the BEC hyaluronan synthases, or form hyalurononhyaluronon stable complexes. Further, the ecGCx is also anchored via the proteoglycan (glypican) to the caveolae and this plays a key role in mechanotransduction of BEC luminal fluid shear stressinduced synthesis of essential endothelial cell-derived NO via glypican caveolae interactions located within the BEC lipid rafts. Importantly, the ecGCx has a net negative charge largely due to the sulfation of glycosaminoglycan side chains, which allow for strong electrostatic binding to the polyvalent cation lanthanum nitrate (La(3+) nitrate) (LAN). This feature of the ecGCx allows it to bind strongly to lanthanum nitrate for its identification by TEM when perfusion fixation is performed in animal models (Figure 15) [23,65,169–171].

Importantly, impaired FOCM with hyperhomocyteinemia, oxidative stress, and meta-inflammation can be damaging to the ecGCx and contribute to endothelial cell activation and dysfunction with detrimental effects on the vascular tissue that predispose to increased vascular inflammation and a prothrombotic state and ischemia, which is also an inducer of ecGCx loss. Additionally, aberrant mitochondria (aMt) within the vascular endothelial cells may also be associated with the attenuation and/or loss of the ecGCx along with systemic aMt that are associated with increased oxidative stress and the RONSS interactome, impaired FOCM, and HHCY in the MetS reloaded [14,20,141].

The ecGCx as indentified with LAN stains so strongly and electron-dense, it does not allow one to appreciate the multiple proteins and binding sites to the BECs, therefore an illustration was provided to gain a better concept of its composition (Figure 15B).

Recently, our lab has studied the brains of the leptin-deficient BTBR ob/ob mouse, which has the phenotype of obesity with IR and diabetes at 20 weeks of age and compared them to their heterozygote littermates BTBRob+/- [45]. Hudkins et al, developed this genetic model in order to study diabetic nephropathy (DN) in the CE Alpers Lab [31]. They found that this model resulted in pathologic remodeling changes in the kidney, which closely resembled human diabetic nephropathy at 22 weeks of age. We were able to demonstrate in this same obese, insulin-resistant diabetic model

that there was an attenuation and/or loss of the ecGCx while there was no significant remodeling of the Pcs, ACs, or the TJ/AJs, which suggested that the attenuation and/or loss of the ecGCx was an early event in this model (Figure 15) [23].

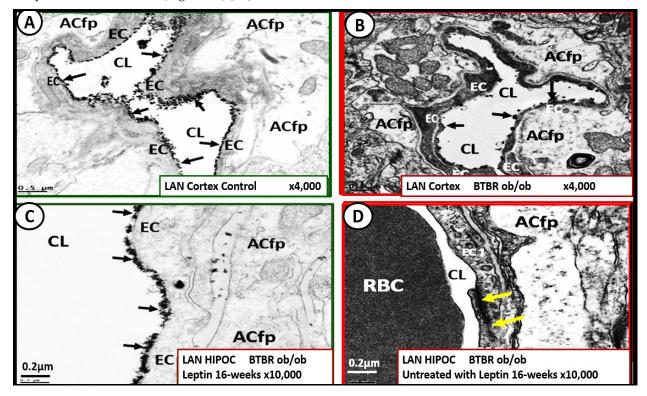


Figure 15. Leptin replacement in the obese diabetic BTBR ob/ob protects the brain endothelial.

cell glycocalyx (ecGCx) in cortical layer III and hippocampus. This suggests that the presence of functioning leptin is important in maintaining a proper ecGCx covering of endothelial cells and that hyperleptinemia and selective leptin resistance (LR) could interfere with a healthy ecGCx as well as insulin resistance (IR). Panel A demonstrates the continuous decoration of the ecGCx with lanthanum nitrate (LAN) staining in the heterozygous non-diabetic control model cortical layer III (arrows). Panel B depicts the marked attenuation and/or loss of the ecGCx in the obese diabetic BTBR ob/ob model cortical layer III and note when the ecGCx was present it was clumped (arrows) and discontinuous as compared to the control in Panel A. Panel C also demonstrates the continuous decoration of the ecGCx in hippocampus CA-1 regions of the BTBR ob/ob models that were treated with intraperitoneal leptin for 16-weeks and stained with LAN (arrows) and the ecGCx is comparable to the control model in panel A and that the ecGCx is continuous. Panel D depicts the complete loss of the ecGCx by LAN staining in the hippocampus CA-1 regions of the BTBR ob/ob and note that the tight and adherens junction (TJ/AJ) remain intact (yellow arrows). The loss of the first barrier (ecGCx) of the tripartite blood-brain barrier (BBB) may result in increased permeability. Images provided by CC 4.0 [23]. Magnification x4000; scale bar = 0.5 µm in panels A and B. Magnification x10,000; scale bar = $0.2 \mu m$ in panels C and D. ACfp = astrocyte foot process; Cl = capillary lumen; BEC = brain endothelial cell; HIP and HC = hippocampus CA-1 regions.

Importantly, the absence of leptin results in the attenuation and even loss of the ecGCx, which may be an early change in BTBR *ob/ob* models. We also know that adiponectin is important in maintaining the function of the EC and hyperlipidemia is associated with decreased adiponectin that is known to be present in BTBR *ob/ob* models [175]. Thus, a decrease in adiponectin in BTBR *ob/ob* models may also be playing an additional role in ecGCx remodeling with its attenuation and or loss.

When the NVU ecGCx becomes dysfunctional, attenuated and/or lost via shedding as in the obese, insulin-resistant BTBR *ob/ob* model there would be a decrease in bioavailable NO to signal pericytes in the capillary NVU as well as increased neurovascular unit permeability [20,65].

Additionally, we have been able to also demonstrate ACfp retraction in the obese, insulin resistant, leptin resistant diabetic *db/db* model [8,23,47,167]. Each of these remodeling changes could result in a regional loss of communication between neurons and the neurovascular unit (NVU) and result in the loss of neurovascular coupling. The attenuation and/or loss of the ecGCx could additionally result in the loss of regional mechanotransduction and result in endothelial dysfunction and possibly the loss of neurovascular coupling, which would result in decreased regional cerebral blood flow and ischemia [176–178].

Since ecGCx loss appears to be an early event in the BTBR *ob/ob* models, this could add to the oxidative stress in brain ECs and contribute to an increase in NADPH oxidase-derived ROS. An intact ecGCx and Mt of brain ECs are of great importance in maintaining the NVU blood-brain barrier (BBB) homeostasis. The attenuation and/or loss of the ecGCx in T2DM can have multiple pathophysiological outcomes, which include increased vascular permeability, edema formation, increased adhesion of circulating inflammatory cells, accelerated inflammatory processes, activation of the coagulation cascade, and platelet aggregation [13,20,23,166,168–173]. Excess nutrients such as increased glucose and fatty acids that occur in obesity, MetS, IR, LR, and T2DM will result in leaky aMt phenotypes and an increase in mtROS as previously observed in the brains of the obese, insulin-resistant diabetic *db/db* models in addition to the increase in NADPH oxidase (Figure 16) [44,46–48].

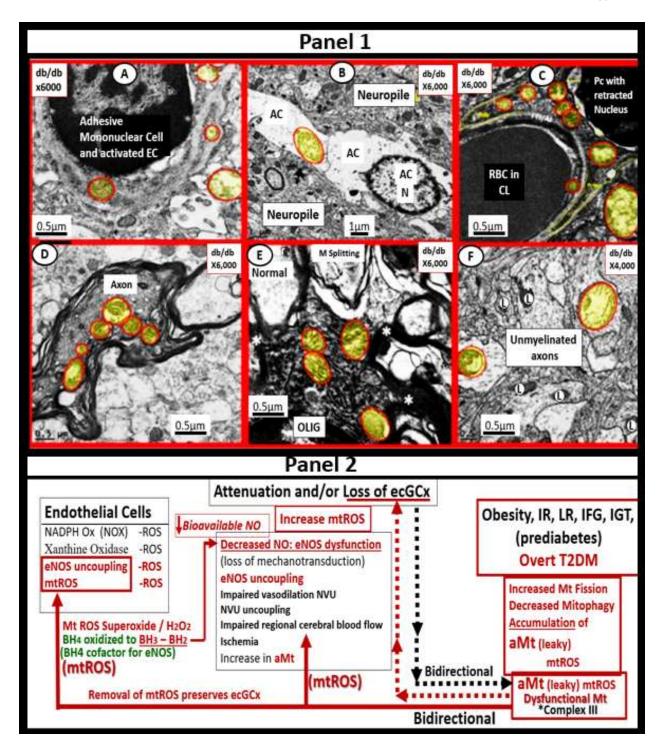


Figure 16. Aberrant mitochondria (aMt) are present in multiple cell types in the brain and emerging evidence suggest a bidirectional relationship between aMt and the endothelial glycocalyx (ecGCx). Panel 1 depicts aMt in multiple cell types in the brain (pseudo-colored yellow with red outlines). This is in contrast to the smaller electron-dense Mt found in healthy control models in brain endothelial cells. These aMt are hyperlucent and lose their electron dense mitochondrial matrix and cristae in brain endothelial cells (BECs) (figure A), astrocytes (AC) (figure B), Pericytes (Pc) and foot processes (figure C), myelinated neuronal axons (figure D), oligodendrocyte (OLIG) (figure E), and unmyelinated axons (figure F). These modified images are provided by CC 4.0 [10, 47, 48, 49. 50]. Magnifications and scale bars are included in each panel. Panel 2 illustrates that there may be a bidirectional relationship between aMt and dysfunction, attenuation, and/or loss (shedding) of the brain BECs ecGCx. Note that this illustration establishes the role of oxidant stress—reactive oxygen species (ROS) in BECs and specifically mitochondrial ROS (mtROS) (left-hand box). Further, it establishes the important role of the dysfunctional attenuated and/or shedding loss of the ecGCx in

the brain (center box). Next, it establishes that obesity, IR, LR, impaired fasting glucose (IFG), impaired glucose tolerance (IGT), and overt T2DM are related to increased Mt fission, decreased mitophagy, and the accumulation of leaky aMt that leak mtROS (right-hand box). Further, this figure depicts that leaky aMt may be responsible for the attenuation and/or loss of the ecGCx (red-dashed arrows) and that, in turn, may result in the loss of the ecGCx that may contribute to an increase in aMt (black-dashed arrows). Importantly, mtROS superoxide or hydrogen peroxide (H2O2) could oxidize the essential tetrahydrobiopterin (BH4) cofactor that is absolutely essential for the eNOS enzyme to produce nitric oxide (NO) and result in eNOS uncoupling, which would ultimately result in decreased bioavailable NO. This bidirectional interaction could result in a vicious cycle, which could be interrupted by preventing the accumulation of aMt (improved mitophagy) or preventing the dysfunction, attenuation, and/or loss (shedding) of the ecGCx. Asterisk = Myelin (M); L = lysosome; RBC = red blood cell.

Additionally, these aMt will lead to dysfunction and or loss of the ecGCx and to a further increase of aMt and mtROS. Increased EC mtROS could result in contributing to even further ecGCx shedding or impairment of its regeneration [179–181]. Thus, this sequence of events between the aMt, mtROS, and ecGCx shedding could be bidirectional, such that a vicious cycle might develop since one abnormality can lead to the other. Decreasing mtROS from leaky aMt results in improved homeostasis [48,130] and in a similar fashion restoring the ecGCx could improve homeostasis and function [164–166]. Importantly, there may exist a bidirectional role between the accumulation of aMt (due to impaired mitophagy) and the dysfunction or loss of the ecGCx in ECs and specifically the BECs (Figure 16 panel 2) [13,48,50,123,164–166].

Certainly, more research may be necessary to confirm this bidirectional relationship between aMt and ecGCx attenuation or loss via shedding. However, it remains a very intriguing association and presents an emerging opportunity to further unlock some of the mysteries associated with aMt, obesity, IR, LR, hyperglycemia of IFG, IGT, T2DM, and the MetS reloaded in addition to associated complications of diabetic end-organ disease. Additionally, this concept may allow for future interventions to interrupt this bidirectional vicious cycle by utilizing SGLT2i empagliflozin [50] or the specific Mt carbonic anhydrase inhibition with topiramate [138], uncoupling proteins such as UCP2 [182], and the emerging role of Mt transfer [183–185].

In order to aid in the restoration of the dysfunctional, attenuated or loss of the ecGCx as occurs in sepsis, COVID-19, neuroinflammation and neurodegenerative disease. and T2DM one would need to make provisions for an adequate supply of sulfur compounds and/or thiols [168]. From this standpoint sulodexide (SDX), a heparin sulfate-like compound resistant to degradation by heparanase, accelerated ecGCx regeneration in vitro and in vivo [186], sodium thiosulfate (STS) [65,168,187] and n-acetyl-cysteine (NAC) [188] are approved and available for clinical use and have been recently utilized in the treatment of the T2DM and sepsis, glia-mediated neuroinflammation in neurodegenerative disease, and COVID-19 pandemic

7. MetS Reloaded and Metainflammation

Historically, it is important to note that Hotamisligil et al were the first group to demonstrate that the adipose-derived cytokine tumor necrosis factor alpha (TNF α) provided the first link between the adipose tissue and IR and eventually linked adipose tissue metainflammation to the MetS [189,190].

Chronic low-grade sterile inflammation – metainflammation may possibly represent a triggering factor in the origin of the metabolic syndrome [191,192]. Predisposing factors such as overnutrition, physical inactivity, obesity, and aging would result in cytokine hypersecretion via visceral adiposity and adipocyte-derived adipokines associated with crown-like structures and increased adipose cytokine secreting macrophages, which could eventually lead to IR and T2DM in genetically or metabolically predisposed individuals such as discussed in section 1. by Grundy et al, regarding metabolic_susceptibility and obesity (Figs. 1, 3) [2,33]. Additionally, gut microbiota dysbiosis, which is associated with excess nutrients, high fat, sucrose, and fructose Western DIO, obesity, MetS, IR, LR, and prediabetes and manifest T2DM in preclinical diabetic *db/db and* BTBR *ob/ob* models. Gut

dysbiosis is known to result in leaky gut-derived metainflammation as a result of metabolic endotoxemia due to the leakage of LPS into the systemic circulation [191-193]. Mechanisms that produce metabolic endotoxemia are thought to be due to gut microbiome dysbiosis. This dysbiosis is associated with a leaky gut due to the dysfunction, attenuation, and/or loss of the intestinal lining epithelial TJ/AJ that allows LPS and other PAMPs derived from cellular membranes. These released PAMPS and LPS (derived from cellular epithelial membranes of the dysbiotic intestinal gramnegative bacteria) result in increased peripheral cytokines/chemokines that signal TLR-4 receptors. Once signaled TLR-4 signal NF-kB activation and downstream cytokines/chemokines to result in metabolic endotoxemia and metainflammation. This gut-derived endotoxemia metainflammation are capable of further activation of the macrophages within the VAT to become inflamed with crown-like structures that secrete inflammatory cytokines/chemokines to cause VATderived obesity-related metainflammation [191–193]. Importantly, there may exist a triangulation of metabolic functions, gut microbiota, and the innate immune system (Figure 17) [194–197].

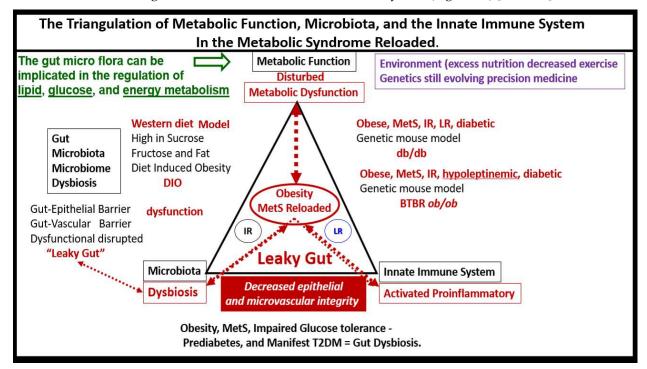


Figure 17. The triangulation of metabolic function, gut microbiota, and the innate immune system. This illustration demonstrates that microbiota dysbiosis, metabolic dysfunction, and an activated proinflammatory innate immune system are bidirectionally associated with obesity and the metabolic syndrome reloaded. Importantly, this triangulation results in a dysfunctional gut-epithelial and gut-vascular barrier that results in a leaky gut to allow increased metainflammation and impaired gut-liver, gut- brain, gut-heart, and gut-kidney homeostasis with resulting impairment of the gut to the brain-heart-kidney axis .

Notably, author fondly remembers when Schmidt et al, presented their data regarding the importance of metainflammation in the development of T2DM via decreased albumin, increased fibrinogen, orosomucoids, and sialic acid in 1999 at the American Diabetes Association [198] and the follow-up study in 2003, which demonstrated elevations in interleukin-6, and C-reactive protein [199]. Thus, the combination of obesity and gut dysbiosis associated with obesity are important causative factors in the development of metabolic endotoxemia and metainflammation in the setting of the MetS reloaded and manifest T2DM [192,193,200].

In addition to the important role of IR, hyperleptinemia and selective leptin resistance (LR) play an important role in the development of metainflammation in the MetS [45,191–193,199,200]. Further, the adipokine/hormone leptin is known to be a proinflammatory cytokine/adipokine hormone and in selective leptin resistance, hyperleptinemia could further enhance

metainflammation. Selective leptin resistance (LR) implies that some of the signaling effects of leptin are impaired, while some such as those providing for heightened effect on inflammation could be additive to the proinflammatory effects due to IR [201–205]. Enhanced VAT-derived cytokine expression that includes the perivascular adipose tissue (PVAT) is another plausible mechanism for the metainflammation/MetS reloaded relationship (Figs.18, 19) [6,112].

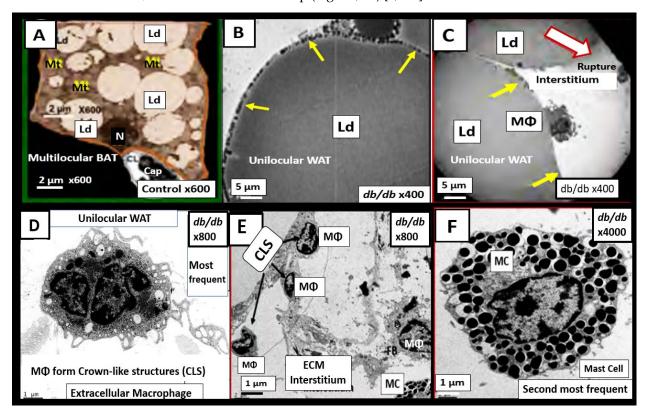


Figure 18. From multilocular brown adipose tissue (BAT) in controls to unilocular white adipose tissue in the obese, insulin resistant (IR), selective leptin resistant (LR), diabetic db/db models in the upper one-third of the descending aorta perivascular adipose tissue (PVAT) - visceral adipose tissue (VAT) of the tunica adiposa. Panel A demonstrates the normal multilocular BAT in the PVAT – VAT with multiple lipid droplet(s) (Ld) and electron-dense mitochondria of the descending thoracic aorta in control models. Panel B depicts the markedly expanded unilocular WAT with engorgement of triglycerides to form the huge lipid droplet droplets (Ld) with marked thinning of the adipocyte plasma membrane and the compressed mitochondria (yellow arrows). Panel C depicts the inflammatory macrophage (MΦ) cytoadherence to the unilocular hypertrophic engorged adipocyte to the point of rupture (red open arrow) and note the absence of the mitochondria at the site of rupture. Panel D depicts the most prominent inflammatory cell macrophage (MΦ) found within the PVAT of the diabetic db/db models. Panel E depicts the formation of macrophage crown-like structures (CLS) (arrows) in the diabetic db/db models. Panel F depicts the second most prominent inflammatory cell the mast cell (MC) with its prominent electron-dense secretory granules. These modified images are presented with permission by CC 4.0 [6,139]. Magnifications and scale bars vary and included in each image. $ECM = extracellular \ matrix; Mt = mitochondria; N = nucleus.$

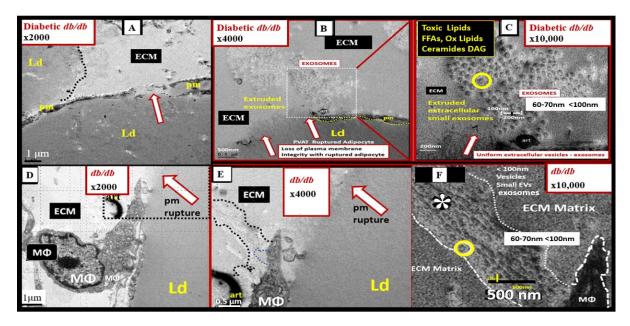


Figure 19. Adipocyte plasma membrane thinning with rupture of plasma membrane (pm) with extrusions of the lipid droplet (Ld) contents and macrophage (M Φ) extrusions of extracellular vesicles (small exosome-like vesicles 60-70nm in diameter) in the obese, insulin-resistant, leptin-resistant, diabetic db/db perivascular adipose tissue (PVAT) – visceral adipose tissue (VAT) of the tunica adiposa in the upper one-third of the descending thoracic aorta. Panel A demonstrates the extreme thinning and loss of the plasma membrane integrity (dashed line) that is associated with crown-like structures of macrophage cytoadherence. Panels B and C depict the complete loss of the pm with rupture and extrusion of the lipid droplet contents into the extracellular matrix (ECM) interstitium of the PVAT -VAT. Importantly, note that these extruded lipids appear as extracellular exosome-like vesicles (encircled) in panel C, in that, their diameter is ~ 60-70 nm, and also note the scale bar that is placed centrally to suggest their diameter meets the criteria for small exosomes (< 100nm) being extruded from the unilocular ruptured adipocytes in the PVAT. Importantly, the control models that consisted of brown adipose tissue and the models treated with empagliflozin did not demonstrate any crown-like structures (CLS) or ruptured adipocytes as in the diabetic db/db models. **Panels D, E**, and F depict the extrusion of extracellular exosome-like vesicles that are less than 100 nm and are therefore considered to be small macrophage-derived EVexosomes (asterisk and encircled) and similar to the morphology of the adipocyte EVexosome-like vesicles in panel C. These modified images are presented with permission by CC 4.0 [6,139]. Magnification and scale bars vary and are present in each panel.

Note the extruded contents of the ruptured adipocytes and macrophages respectively in the *db/db* models (Figure 20C and F). These extruded vesicles appeared to be very uniform with variable electron dense staining and vesicle-like morphology. These ruptured adipocytes and macrophage vesicles are thought to be small exosomes due to their nanosized diameter (60-70nm) and exosomes are known to range from 50-100nm in diameter making these vesicles within the size range of small exosomes [6].

7.1. MetS Reloaded and Adipokines/hormones, Cytokines/chemokines, and Toxic Free Fatty Acids (FFA)

In obesity, the adipose tissue is an active endocrine and paracrine organ that releases several bioactive mediators including adipokines, cytokines/chemokines and toxic saturated free fatty acids (FFA) that influence not only body weight homeostasis but also inflammation, coagulation, fibrinolysis, as well as insulin and leptin resistance [206]. As one views the vast amount of adipose tissue (both SAT and VAT) in the obese Zucker model in figure 4, one can almost visualize the adipokines, cytokines/chemokines, and toxic FFAs spewing forth from these expanded; excessive adipose depots. Indeed, adipokines, cytokines/chemokines, and FFAs each have various functions in obesity and effects on clinical CCVD (Figure 20) [206–210],

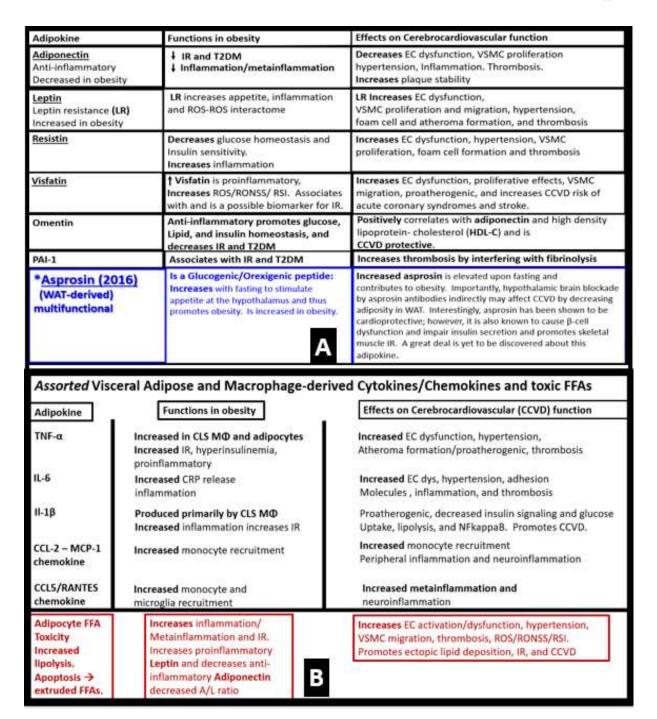


Figure 20. Increased adipokines, assorted cytokines, chemokines, and toxic free fatty acids (FFAs) effects on cerebrocardiovascular function and disease. **Panel A (upper panel)** depicts multiple adipokines, their functions in obesity, and effects influencing cerebrocardiovascular disease. **Panel B (lower panel)** depicts assorted visceral adipose and adipose-derived macrophage-derived cytokines/chemokines and toxic FFAs. $CCL2 = chemokine \ ligand \ 2$; $CCL5 = chemokine \ ligand \ 5$; $CCVD = cerebrocardiovascular \ disease$; $CLS = crown-like \ structures$; $EC = endothelial \ cell$; $FFA = saturated \ free \ fatty \ acids$; $IL-1\beta = interleukin \ 1-\beta$; IL-6 = interleukin-6; $IR = insulin \ resistance$; $LR = leptin \ resistance$; $MCP-1 = monocyte \ chemotactic \ protein-1$; $M\Phi = macrophage$; $PAI-1 = plasminogen \ activator \ inhibitor \ -1$; $RANTES = regulated \ on \ activation$, $normal \ T \ cell \ expressed \ and \ secreted$; $ROS = reactive \ oxygen \ species$; $RONSS = reactive \ oxygen$, nitrogen, $sulfur \ species$; $T2DM = type \ 2 \ diabetes \ mellitus$; $TNF\alpha = tumor \ necrosis \ alpha$; $VSMC = vascular \ smooth \ muscle \ cell$.

For those who may wish to see an even more complete list of adipokines please see the following reference [211].

7.2. PVAT Adipocyte and Macrophage-Derived Exosome microRNAs (miRNAs) and long-non-coding RNAs (lncRNAs)

PVAT adipocyte-derived extracellular vesicle small exosomes (PVAT-dEVexos) and PVAT macrophage-derived extracellular vesicle small exosomes (PVATMΦ-dEVexos) are capable of delivering a multitude of proteins, lipids, nucleic acids (RNA and DNA) via micro RNAs (miRNAs) and long-non-coding RNAs (lncRNAs) (longer than 200 nucleotides). These RNAs are capable of signal transduction and even the exchange of genetic information [6]. Thus, the obese PVAT adipose-derived exosomes are an important source of circulating exosomes that are capable of regulating gene expression of adjacent cells (paracrine function) or distant cells and tissues (endocrine function) via their miRNAs in an inter-organ signal transduction fashion. exosome-like vesicles are known to be related to the development of IR via the toll-like receptor (TLR4)/Toll-interleukin-1 receptor (TIR) domain-containing adaptor protein inducing interferon-β (Trif) –(TLR4/TRIF) pathway [212]. In regards to inflammation, these exosomes are capable of preventing or promoting macrophage polarization by inhibiting Kruppel-like factor 4 and thus, these M-1-like macrophages are capable of increasing systemic inflammation and contribute to metainflammation and metabolic abnormalities induced by obesity in the MetS reloaded [213]. Indeed, miRNAs found within adipocyte and $M\Phi$ -derived exosomes are not only novel but also an exciting and emerging field of research that will continue to grow and change over time [6,214,215].

Importantly, Kwan et al, have recently examined the role of miRNAs that are generated by the obese adipocytes within the VAT-PVAT depots [216]. They have now identified multiple miRNAs that are capable of being shed from these obese adipocytes that have both paracrine cell-cell signaling mechanism to interstitial macrophages and mast cells within these depots as well as providing an endocrine and inter-organ signaling mechanisms to affect other organs such as the liver, cerebrocardiovascular tissues, and pancreatic islet β-cells to promote insulin resistance and endorgan dysfunction. These miRNAs include the following: decreased miR-148 and miR-4269; increased miR-23b and miR-4429, which activates the TGF-β and Wnt/β-catenin signaling pathways that promote fibrosis; increased miR-155 and miR-223 and decreased miR-34a promote macrophage polarization to an M1-like phenotype; increased miR-34a promotes hepatic steatosis, glucose intolerance and IR; decreased miR141-3p; increased miR-222 and miR-27a promotes peripheral IR; increased miR-221-3p promotes vascular remodeling [216]. Further, Żbikowski et al, have identified key miRNAs generated from obese VAT-PVAT adipocytes and polarized M1-like macrophages that are capable of paracrine, endocrine, and inter-organ signaling that contributes to peripheral IR. They found the following increased miRNAs that are important in promoting IR as follows: miR-27a, miR-29a, miR-141-3p, miR-155, miR-222. Additionally increased miR-34a and miR-155 also promote the polarization of MΦs to M1-like macrophages [217]. Various miRNA profiles are beginning to emerge that implicate the PVAT adipocytes and macrophages to result in IR. Moreover, in due time, there will be certain miRNA profiles that will emerge to better define the MetS reloaded. Interestingly, the exosomal miRNA numbers may be somewhat analogous to the letters of the alphabet, in that they will eventually allow us to spell words with these numbers and eventually write sentences, paragraphs, and stories in regards to the various miRNAs profiles that are still emerging. Further, investigations of the mechanisms of action of adipocyte and $M\Phi$ exosomal miRNAs and their role in the MetS, CCVD and T2DM will be of considerable value for a more thorough understanding of the pathobiological mechanisms of the MetS and its increased risk for CCVD and T2DM. Indeed, there are cell-specific miRNAs secreted by the VAT-PVAT adipocytes and activated, CLS, M1-like M Φ s in obesity (Figure 21) [216–219].

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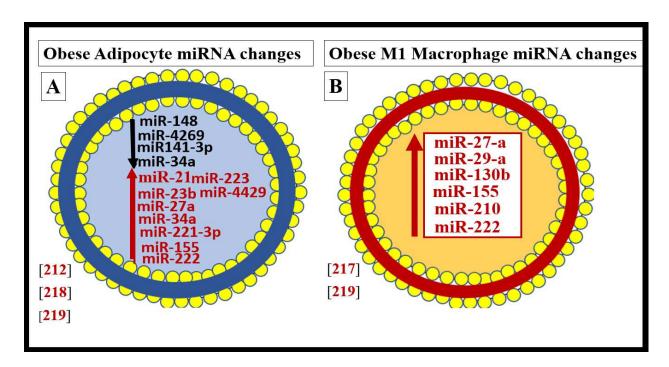


Figure 21. Cell specific miRNAs profiles in adipocytes and activated, M1-like macrophages ($M\Phi s$) in the obese PVAT - VAT. **Panel A** depicts the decreased miRNAs (black lettering and black arrow) and the increased miRNAs (red lettering and red arrow) in obese visceral adipose tissue including perivascular adipose tissue (PVAT). **Panel B** depicts the elevated miRNAs (red lettering and red arrow) in the activated M1-like macrophages in the PVAT - VAT [6,217–219].

Currently, lncRNAs are even more in their infancy in regards to the MetS; however, they are emerging rapidly and will soon contribute to our understanding, since they like miRNAs are transported via adipocyte and $M\Phi$ extracellular vesicle exosomes that may impact neuroglia activation and function including impaired cognition and neurodegeneration [220,221] as well as other organs throughout the body. These lncRNAs have the capability to control gene transcription, pre-mRNA processing, the transport of mature mRNAs to specific cellular compartments, the regulation of mRNA stability, protein translation and turnover [221]. Additionally, lncRNAs are known to be involved in nuclear chromatin remodeling, which imply the possibility of post-natal epigenetic mechanisms. Of interest, we have observed in our TEM studies of microglia and oligodendrocytes in the obese, IR, hyperleptinemic, diabetic db/db models chromatin remodeling consisting of chromatin condensation [48,49].

Thus, there are three possible mechanisms that may be in play when evaluating the obese PVAT - VAT depots containing hypertrophic; prone to rupture unilocular WAT adipocytes and polarized M1-macrophages of the CLS. The first possible mechanism i) the engorged unilocular adipocytes of the WAT PVAT-VAT rupture and shed their toxic contents of FFA, oxidized lipids, ceramides, and exosomes to evoke an innate immune response. This innate immune response will result in an accumulation of macrophages forming CLS and incite both mast cell and MΦ activation to result in metainflammation with the production of both local and systemic damaging cytokines and The second possible mechanism ii) the obese adipocytes and activated proinflammatory polarized M1-like macrophages secrete miRNAs that promote peripheral insulin resistance that result in remodeling changes to end-organs affected by both obesity and T2DM. While the third possible mechanism iii) the elevated proinflammatory adipokine/hormone leptin and the associated decrease in the protective anti-inflammatory adipokine/hormone adiponectin will result in the activation of innate PVAT immune cells such that they could develop excessive damaging cytokine/chemokine release that would be taken up by the peripheral circulation and promote systemic metainflammation, which we know interferes with insulin sensitivity and promotes IR and thus ties LR to IR in the MetS reloaded. These three mechanisms may act

individually; however, they are most likely functioning in concert and synergistically. Importantly, these three scenarios complement the findings proposed by Chadt et al, in that they proposed three hypotheses to help bridge the gap between epidemiology and pathobiological mechanisms in obesity. Their three hypotheses included: 1). The inflammation hypothesis or metainflammation hypothesis; 2). The adipose tissue expandability or overflow hypothesis, which predicts that obesity may result in increased ectopic lipid deposition and IR in insulin-sensitive tissues such as the skeletal muscle, cardiac muscle, liver, islet pancreatic β -cells, and the PVAT (VAT itself; 3). The adipokine hypothesis, which refers to the WAT cells in the SAT to function as an endocrine organ with excessively increased leptin and decreased adiponectin via both autocrine, paracrine, and endocrine mechanisms to allow for the metabolic impairment of insulin-sensitive target tissue to develop IR. Which associates with LR and eventually results in not only β -cell dysfunction but also β -cell failure due to both glucotoxicity and lipotoxicity – glucolipotoxicity with ensuing T2DM [222].

8. Conclusions

The MetS reloaded supports the concept that many novel risk factors and emerging variables now belong to the original MetS as proposed by NCEP (2002) Grundy et al. (2004) (Figure 1) [2,33] and multiple associations and organizations. This overview has attempted to validate novel emerging risk factors and variables as well as exploring each of the original and traditional four arms of the MetS reloaded including hyperlipidemia, hyperinsulinemia and hyperamylinemia, HTN (essential), and hyperglycemia in sections (2.); (3., 3.1., 3.2., 3.3.); (4.); (5.). While section (6. and 6.1.) explored the role of EC activation - dysfunction and the ecGCx, and section (7., 7.1., and 7.2.) examined the role of metainflammation in the MetS reloaded and the potential role of adipokines, cytokines/chemokines and toxic FFF, and PVAT-derived adipocyte and activated M Φ exosomes (Figure 1). It is hoped that the examination of the MetS reloaded will provide not only a better understanding of the original definition of metabolic syndrome but also will allow us to keep the concept of the MetS reloaded at the forefront of our research concepts and clinical practices. After all, new concepts may fuel the fire for new discoveries.

There has been controversy over the years since the MetS early definition (and whether or not its whole may be greater than the sum of its parts [223–225], The author concludes from preparing this overview the following: Even if an individual does not meet the criteria for the diagnosis of the MetS (due to missing just a few under-evaluated measurements or mg/dl deficit cut points), and if there are other risks factors within the MetS criteria that are elevated that these individual elevated risk factors should be treated aggressively to goal if at all possible. Further, clinicians should definitely evaluate and treat all T2DM and CCVD risk factors without regard to whether the individual meets the 3 out of the 5 criteria for the diagnosis of the metabolic syndrome [226]. For a balanced view of the MetS controversy written in 2007 that still holds true to date, it is suggested that one read a very elegantly written manuscript by Ferrannini [227].

The MetS may certainly be viewed as a risk syndrome that definitely provides a platform for individual indentification and multifactorial risk reduction in regards to its treatment profile including the all-important necessary life style changes especially in regards to dietary and exercise fitness. Sometimes, a simple acronym such as the (A-FLIGHT-UR) acronym is helpful to remember the multiple risk factors, variables, and emerging novel metabolic toxicities and signaling mechanisms that can be applied to develop individual risk reduction of each of the multiple abnormal risks and variables associated with the MetS reloaded (Table 2) [14].

Table 2. The A-FLIGHT-UR acronym may be utilized as an aid in remembering the multiple risk factors and metabolic toxicities associated with the MetS reloaded. This modified and updated table is provided with permission by CC 4.0 [14]. AGE = advanced glycation end products; ALE = advancel lipoxidation end products; FOCM = folate one-carbon metabolism; HDL-C = high density lipoprotein-cholesterol; HPA = hypothalamic pituitary adrenal; II-6 = interleukin-6; HDL = high density lipoprotein; LDL-C = low density lipoprotein-cholesterol; PKC = protein kinase C; RAGE = receptor or AGE; $TNF\alpha = tumor$ necrosis alpha.

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A	Aging; Amylin (hyperamylinemia) induces the RAAS; Angiotensin II; AGE/RAGE activation; ALE; Antioxidant reserve compromised; Angiogenesis (increased); Arteriogenesis (impaired); Accelerated atherosclerosis (Atheroscleropathy); Aberrant Mt (aMt); Adipose tissue and Adipose exosomes; Albuminuria.		
	A-FLIGHT; A term to describe which order teams play in golf.		
F	FFA (free fatty acids); FOCM impaired – iFOCM; Females lose their cardioprotection; I FEAR = Insulin resistance erases the female advantage in cerebro-CVD (CCVD).		
L	Lipotoxicity (atherogenic hyperlipidemia) related to obesity and metainflammation; contributes to metainflammation and increased leptin and Leptin Resistance Selective (Lr)		
	leptin resistance (<u>Lr</u>); decreased protective adiponectin; cytokine release especially IL-6 and TNF alpha.		
1	Insulin toxicity (hyperinsulinemia) endogenous; compensatory hyperinsulinemia and insulin resistance (IR) – central to the MetS reloaded.		
G	Glucotoxicity (compounds peripheral insulin resistance); both oxidative stress and reductive stress; results in glucose autoxidation, polyol/hexosamine flux, AGE/RAGE, activates PKC.		
Н	Hypertension (essential hypertension (eHTN) toxicity; hyperhomocysteinemia (HHCY), which is a marker of impaired folate-mediated one-carbon metabolism (iFOCM); relates strongly to vascular - arterial stiffening (AS or VS); HPA axis impaired.		
T	Triglyceride toxicity - atherogenic hyperlipidemia with increased sd LDL-C and decreased HDL-C; non-HDL; lipoxidation		
U	Uric acid toxicity - Hyperuricemia		
	ROS; RONS (reactive oxygen, nitrogen species); RONSS (reactive oxygen,		
R (ROS)	nitrogen, sulfur species); RSI (reactive species interactome); ROS induced ROS release (RIRR).		

Each of these metabolic risk factors and variables promotes ROS - RONSS either directly or indirectly and result in excessive oxidative and redox stress. Importantly, remember that ROS begets ROS, and ROS begets metainflammation, and metainflammation begets ROS and is capable of creating vicious cycles that may be referred to now as ROS induced ROS release (RIRR) [146].

Multiple efforts have been made to utilize supportive ultrastructural TEM images and illustrations to discuss ultrastructural cellular remodeling and functional changes in the visceral adipose tissue depots including the PVAT, pancreatic islets, brain, and depict aMt, and activated ECs in the obese, MetS, IR, LR, prediabetic, and diabetic preclinical rodent models. This overview has also shared the important role of identifying the MetS reloaded with its multiple traditional risk factors and the evolving novel risk factors and variables as early as possible. Additionally, the MetS has been embedded in controversy as to its importance and necessity since its initial definition. However, the term MetS and its definitions have persisted on a global scale for the past three decades and it is likely to continue to persist as the MetS into the future.

In summary, the clinical utility of the MetS (International Classification of Diseases, Tenth Revision Clinical Modification diagnostic code (ICD-10-CM E88.81) is important. The early diagnosis of the MetS (E88.81) allows both the health care providers and affected individuals to develop treatment plans for each of the specific risk factors and variable components and lifestyle changes. Further, the early identification and treatment of the MetS allow the opportunity to prevent the development of CCVD and T2DM and importantly their long-term complications with increased morbidity and mortality.

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References

- 1. National Cholesterol Education Program (NCEP) Expert Panel on Detection, Evaluation, and Treatment of High Blood Cholesterol in Adults: Executive summary of the third report of the National Cholesterol Education Program (NCEP) Expert Panel on Detection, Evaluation, and Treatment of High Blood Cholesterol in Adults (Adult Treatment Panel III). JAMA. 2002;285:2846–2497. doi:10.1001/jama.285.19.2486
- 2. Grundy SM, Brewer HB Jr, Cleeman JI, Smith SC Jr, Lenfant C, American Heart Association; National Heart, Lung, and Blood Institute. Definition of metabolic syndrome: Report of the National Heart, Lung, and Blood Institute/American Heart Association conference on scientific issues related to definition. Circulation. 2004;109(3):433-438. doi:10.1161/01.CIR.0000111245.75752.C6
- 3. Kylin E: Studien ueber das Hypertonie-Hyperglyka "mie-Hyperurika" miesyndrom. Zentralblatt fuer Innere Medizin. 1923, 44: 105-127.
- 4. Himsworth HP, Kerr RB. Insulin-sensitive and insulin-insensitive types of diabetes mellitus. Clin Sci. 1939;4:119–152
- 5. Reaven GM. Banting lecture 1988. Role of insulin resistance in human disease. Diabetes. 1988;37(12):1595-1607. doi:10.2337/diab.37.12.1595
- 6. Hayden MR. <u>Overview of Neuroglia Activation, Chronic Neuroinflammation, Remodeling, and Impaired Cognition Due to Perivascular Adipose Tissue-Derived Extracellular Vesicle Exosomes in Obesity and Diabetes.</u> Neuroglia. 2022;3(4):112-138. doi: https://doi.org/10.3390/neuroglia3040008
- 7. Alberti KG, Zimmet PZ. Definition, diagnosis and classification of diabetes mellitus and its complications. Part 1: diagnosis and classification of diabetes mellitus provisional report of a WHO consultation. Diabet Med.1998;15(7):539-553. doi: 10.1002/(SICI)1096-9136(199807)15:7<539::AID-DIA668>3.0.CO;2-S
- 8. Hayden MR, Banks WA, Shah GN, Gu Z, Sowers JR. Cardiorenal metabolic syndrome and diabetic cognopathy. Cardiorenal Med. 2013;3(4):265-82. doi: 10.1159/000357113
- 9. Firoz CK, Jabir NR, Khan MS, Mahmoud, M, Shakil S, Damanhouri GA, et al. An overview on the correlation of neurological disorders with cardiovascular disease. Saudi J Biol Sci. 2015; 22(1): 19–23. doi: 10.1016/j.sjbs.2014.09.003

40

- 10. Hayden MR. Type 2 Diabetes Mellitus Increases the Risk of Late-Onset Alzheimer's Disease: Ultrastructural Remodeling of the Neurovascular Unit and Diabetic Gliopathy. Brain Sci. 2019;9(10):262. doi: 10.3390/brainsci9100262
- 11. Paoletti FP, Simoni S, Parnetti L, Gaetani L. The Contribution of Small Vessel Disease to Neurodegeneration: Focus on Alzheimer's Disease, Parkinson's Disease and Multiple Sclerosis. Int J Mol Sci. 2021 May; 22(9): 4958. doi: 10.3390/ijms22094958
- 12. Jia G, Aroor AR, Sowers JR. Arterial Stiffness: A Nexus between Cardiac and Renal Disease. Cardiorenal Med. 2014 Apr;4(1):60-71. doi: 10.1159/000360867
- 13. Hayden MR. The Mighty Mitochondria Are Unifying Organelles and Metabolic Hubs in Multiple Organs of Obesity, Insulin Resistance, Metabolic Syndrome, and Type 2 Diabetes: An Observational Ultrastructure Study. Int J Mol Sci. 2022; 23(9):4820. https://doi.org/10.3390/ijms23094820
- 14. Hayden MR, Tyagi SC. Intimal redox stress: accelerated atherosclerosis in metabolic syndrome and type 2 diabetes mellitus. Atheroscleropathy. Cardiovasc Diabetol. 2002;1:3. doi: 10.1186/1475-2840-1-3
- 15. <u>Aroor AR, Das NA, Carpenter AJ, Habibi J, Jia G, Ramirez-Perez FI, Martinez-Lemus L, Manrique-Acevedo CM, Hayden MR, et al. Glycemic control by the SGLT2 inhibitor empagliflozin decreases aortic stiffness, renal resistivity index and kidney injury. Cardiovasc Diabetol. 2018; 17, 108 https://doi.org/10.1186/s12933-018-0750-8</u>
- 16. Hayden MR, Tyagi SC. Is type 2 diabetes mellitus a vascular disease (atheroscleropathy) with hyperglycemia a late manifestation? The role of NOS, NO, and redox stress. Cardiovasc Diabetol. 2003; 2: 2. doi: 10.1186/1475-2840-2-2
- 17. Reaven GM. Role of Insulin Resistance in Human Disease (Syndrome X): An Expanded Definition. Annu. Rev. Med. 1993;44:121-123. doi:10.1146/annurev.me.44.020193.001005
- 18. Reaven GM. The insulin resistance syndrome. Curr Atheroscler Rep. 2003;5(5):364-71. doi:10.1007/s11883-003-0007-0
- 19. Hayden MR, Tyagi SC. <u>Uric acid: A new look at an old risk marker for cardiovascular disease, metabolic syndrome, and type 2 diabetes mellitus: The urate redox shuttle.</u> Nutr Metab (Lond). 2004 Oct 19;1(1):10. doi: 10.1186/1743-7075-1-10
- 20. Hayden MR, Tyagi SC. Impaired Folate-Mediated One-Carbon Metabolism in Type 2 Diabetes, Late-Onset Alzheimer's Disease and Long COVID. Medicina. 2021;58(1):16. doi: 10.3390/medicina58010016
- 21. Friedman J. Leptin at 20: An overview. J. Endocrinol. 2014, 223, 1. doi:10.1530/JOE-14-0405
- 22. Friedman J. The long road to leptin. J. Clin. Investig. 2016, 126, 4727–4734. doi:10.1172/JCI91578
- 23. Ingalls AM Dickie, MM, Snell GD. Obese, a new mutation in the house mouse. J. Hered. 1950, 41, 317–318. doi:10.1093/oxfordjournals.jhered.a106073
- 24. Zucker LM, Zucker TF. Fatty, a new mutation in the rat. J. Hered. 1961, 52, 275–278.

- 25. Hummel KP, Dickie MM, Coleman DL. Diabetes, a new mutation in the mouse. Science 1966, 153, 1127–1128. doi:10.1126/science.153.3740.1127
- 26. Peterson RG, Shaw WN, Neel MA, Little LA, Eichberg J. Zucker Diabetic Fatty Rat as a Model for Non-insulin-dependent Diabetes Mellitus. Ilar J. 1990, 32, 16–19. doi:10.1093/ilar.32.3.16
- 27. Zhang Y, Proenca R, Maffei M, Barone M, Leopold L, Friedman JM. Positional cloning of the mouse obese gene and its human homologue. Nature 1994, 372, 425–432. doi:10.1038/372425a0
- 28. Scherer PE, Williams S, Fogliano M, Baldini G, Lodish HF. A novel serum protein similar to C1q, produced exclusively in adipocytes. J. Biol. Chem. 1995, 170, 26746–26749. doi:10.1074/jbc.270.45.26746
- 29. Trujillo ME, Scherer PE. Adiponectin—Journey from an adipocyte secretory protein to biomarker of the metabolic syndrome. J. Int. Med. 2005, 257, 167–175. doi:10.1111/j.1365-2796.2004.01426.x
- 30. Wang ZV, Scherer PE. Adiponectin, the past two decades. J. Mol. Cell Biol. 2016, 8, 93–100. doi:10.1093/jmcb/mjw011
- 31. Hudkins KL, Pichaiwong W, Wietecha T, Kowalewska J, Banas MC, Spencer MW, Mühlfeld A, Koelling M, Pippin JW, Shankland SJ et al. BTBR Ob/Ob mutant mice model progressive diabetic nephropathy. J. Am. Soc. Nephrol. 2010, 21, 1533–1542. doi: 10.1681/ASN.2009121290
- 32. Hayden MR, Banks WA. Deficient Leptin Cellular Signaling Plays a Key Role in Brain Ultrastructural Remodeling in Obesity and Type 2 Diabetes Mellitus. Int J Mol Sci. 2021;22(11):5427. doi: 10.3390/ijms22115427
- 33. Grundy SR. Metabolic syndrome: a multiplex cardiovascular risk factor. J Clin Endocrinol. 2007;92(2):399-404. doi: 10.1210/jc.2006-0513
- 34. Alberti KG, Eckel RH, Grundy SM, Zimmet PZ, Cleeman JI, Donato KA, Fruchart JC, James WP, Loria CM, Smith SC Jr; International Diabetes Federation Task Force on Epidemiology and Prevention; National Heart, Lung, and Blood Institute; American Heart Association; World Heart Federation; International Atherosclerosis Society; International Association for the Study of Obesity. Harmonizing the metabolic syndrome: a joint interim statement of the International Diabetes Federation Task Force on Epidemiology and Prevention; National Heart, Lung, and Blood Institute; American Heart Association; World Heart Federation; International Atherosclerosis Society; and International Association for the Study of Obesity. Circulation. 2009;120(16):1640-5. doi: 10.1161/CIRCULATIONAHA.109.192644
- 35. Panchal SK, Brown L. Rodent models for metabolic syndrome research. J Biomed Biotechnol. 2011;2011:351982. doi: 10.1155/2011/351982
- 36. Ross R. Atherosclerosis An Inflammatory Disease. N Engl J Med. 1999 Jan 14;340(2):115-26. doi: 10.1056/NEJM199901143400207
- 37. Wilson PWF, Grundy SM. The Metabolic Syndrome. A Practical Guide to Origins and Treatment: Part II. Circulation. 2003;108(13):1537-1540. doi: 10.1161/01.CIR.0000089506.12223.F

- 38. deRoos B, Mavrommatis Y, Brouwer IA. Long-chain n-3 polyunsaturated fatty acids: new insights into mechanisms relating to inflammation and coronary heart disease. Br J Pharmacol. 2009;158(2):413–428. doi: 10.1111/j.1476-5381.2009.00189.x
- 39. Libby P. Inflammation in atherosclerosis. Nature. 2002;420(6917):868-874. doi: 10.1038/nature01323
- 40. Tokgözoğlu L, Libby P. The dawn of a new era of targeted lipid-lowering therapies. Eur Heart J. 2022 Jan 20;ehab841. DOI: 10.1093/eurheartj/ehab841
- 41. Rubins HB Robins SJ, Collins D, et al. Gemfibrozil for the secondary prevention of coronary heart disease in men with low levels of high-density lipoprotein cholesterol.
- 42. Veterans Affairs High-Density Lipoprotein Cholesterol Intervention Trial Study Group. N Engl J Med. 1999;341; 410-418. doi: DOI: 10.1056/NEJM199908053410604
- 43. Joslin EP. Arteriosclerosis and Diabetes. Annals of Clinical Medicine. 1927;5:1061–1079.
- 44. Hayden MR, Joginpally T, Salam M, Sowers JR. Childhood and adolescent obesity in cardiorenal metabolic syndrome and Type 2 diabetes: a clinical vignette and ultrastructure study. Diabetes Manage. 2011;1(6):601–614. doi: 10.2217/dmt.11.50
- 45. Paz-Filho G, Mastronardi C, Wong ML, Licinio J. Leptin therapy, insulin sensitivity, and glucose homeostasis. Indian J Endocrinol Metab. 2012;16(Suppl 3): S549–S555. doi: 10.4103/2230-8210.105571
- 46. Aroor AR, Sowers JR, Bender SB, Hayden MR, Nistala R, DeMarco VG, et al. Dipeptidylpeptidase Inhibition Is Associated with Improvement in Blood Pressure and Diastolic Function in Insulin-Resistant Male Zucker Obese Rats. Endocrinology. 2013; 154(7):2501–2513. doi: https://doi.org/10.1210/en.2013-1096
- 47. Hayden MR, Grant DG, Aroor AR, Demarco VG. Ultrastructural Remodeling of The Neurovascular Unit in The Female Diabetic *db/db* Model—Part I: Astrocyte. Neuroglia. 2018;1(1):220-244. doi: https://doi.org/10.3390/neuroglia1010015
- 48. Hayden MR, Grant DG, Aroor AR, Demarco VG. Ultrastructural Remodeling of The Neurovascular Unit in the Female Diabetic db/db Model–Part II: Microglia and Mitochondria. Neuroglia. 2018;1(2):311-326. doi: https://doi.org/10.3390/neuroglia1020021
- 49. Hayden MR, Grant DG, Aroor AR, Demarco VG. Ultrastructural Remodeling of the Neurovascular Unit in the Female Diabetic *db/db* Model—Part III: Oligodendrocyte and Myelin. Neuroglia. 2018;1(2):351-364. doi: https://doi.org/10.3390/neuroglia1020024
- 50. Hayden MR, Grant DG, Aroor AR, DeMarco VG. Empagliflozin Ameliorates Type 2 Diabetes-Induced Ultrastructural Remodeling of the Neurovascular Unit and Neuroglia in the Female *db/db* Mouse. Brain Sci. 2019;9(3):57. doi: 10.3390/brainsci9030057

- 51. Limberg JK, Soares RN, Padilla J. Role of the Autonomic Nervous System in the Hemodynamic Response to Hyperinsulinemia—Implications for Obesity and Insulin Resistance. Curr Diab Rep. 2022; doi: https://doi.org/10.1007/s11892-022-01456-1
- 52. Zamboni M, Zoico E, Fantin F, Panourgia MP, Francesco VD, Tosoni P, Solerte B, Vettor R, Bosello O. Relation Between Leptin and the Metabolic Syndrome in Elderly Women. The Journals of Gerontology: Series A. 2004;59(4):M396–M400. doi: https://doi.org/10.1093/gerona/59.4.M396
- 53. Van Gaal LF, Wauters MA, Mertens IL, Considine RV, De Leeuw IH. Clinical endocrinology of human leptin. Int J Obes. 1999;23:(Suppl 1): M29-36
- 54. Margetic S, Cazzola C, Pegg GG, Hill RA. Leptin: a review of its peripheral actions and interactions. *Int J Obes.* 2002;26:1407-1433.
- 55. Dong M, Ren J. What fans the fire: insights into mechanisms of leptin in metabolic syndrome-associated heart diseases. Curr Pharm Des. 2014;20(4):652-8. doi: 10.2174/138161282004140213160930
- 56. Patel SB, Reams GP, Spear RM, Freeman RH, Villarreal D. Leptin: Linking obesity, the metabolic syndrome, and cardiovascular disease. Curr Hypertens Rep. 2008;10(2):131-137. doi: 10.1007/s11906-008-0025-y
- 57. Donahue RP, Prineas RJ, De Carlo Donahue R, et al. Is fasting leptin associated with insulin resistance among nondiabetic individuals? Diabetes Care. 1999;22:1092-1096. doi: 10.2337/diacare.22.7.1092
- 58. Moore JX, Chaudhary N, Akinyemiju T. Metabolic Syndrome Prevalence by Race/Ethnicity and Sex in the United States, National Health and Nutrition Examination Survey, 1988–2012. Prev Chronic Dis. 2017;14:160287. doi: http://dx.doi.org/10.5888/pcd14.160287external icon
- 59. Hayden MR. An Immediate and Long-Term Complication of COVID-19 May Be Type 2 Diabetes Mellitus: The Central Role of β-Cell Dysfunction, Apoptosis and Exploration of Possible Mechanisms. Cells. 2020;9(11):2475. doi: 10.3390/cells911247
- 60. Ford ES, Giles WH, Dietz WH. Prevalence of the metabolic syndrome among US adults: findings from the third National Health and Nutrition Examination Survey. 2002;287(3):356-359. doi: 10.1001/jama.287.3.356
- 61. Ford ES, Giles WH, Mokdad AH. Increasing prevalence of the metabolic syndrome among U.S. Adults. Diabetes Care. 2004;27(10):2444-9. doi: 10.2337/diacare.27.10.2444
- 62. Hayden MR. Islet amyloid, metabolic syndrome, and the natural progressive history of type 2 diabetes mellitus. JOP. 2002 Sep;3(5):126-38. PMID: 12221327
- 63. Hayden MR, Tyagi SC. Islet redox stress: the manifold toxicities of insulin resistance, metabolic syndrome and amylin derived islet amyloid in type 2 diabetes mellitus. JOP. 2002 Jul;3(4):86-108. PMID: 1211076
- 64. Reaven G. Metabolic syndrome: pathophysiology and implications for management of cardiovascular disease. Circulation. 2002;106(3):286-288. doi: 10.1161/01.cir.0000019884.36724.d9

- 65. Hayden MR. Endothelial activation and dysfunction in metabolic syndrome, type 2 diabetes and coronavirus disease 2019. J Int Med Res. 2020;48(7):300060520939746. doi: 10.1177/0300060520939746
- 66. Hayden MR, Tyagi, SC. "A" is for amylin and amyloid in type 2 diabetes mellitus. JOP. 2001;2(4):124-39. PMID: 11875249
- 67. Jaikaran ET, Clark A. Islet amyloid and type 2 diabetes: From molecular misfolding to islet pathophysiology. Biochim Biophys Acta. 2001 Nov 29;1537(3):179-203. doi: 10.1016/s0925-4439(01)00078-3
- 68. Hayden MR. Islet amyloid, metabolic syndrome, and the natural progressive history of type 2 diabetes mellitus. JOP. 2002;3(5):126-38. PMID: 12221327
- 69. Hayden MR, Tyagi, SC, Kerklo MM, Nicolls MR. Type 2 diabetes mellitus as a conformational disease. JOP. 2005;6:287–302. PMID: 16006679
- 70. Hayden MR, Sowers JR. Treating hypertension while protecting the vulnerable islet in the cardiometabolic syndrome. J Am Soc Hypertens. 2008;2(4):239-66. doi: 10.1016/j.jash.2007.12.002
- 71. Westwell-Roper CY, Chehroudi CA, Denroche HC, Courtade JA, Ehses JA, Verchere CB. IL-1 mediates amyloid-associated islet dysfunction and inflammation in human islet amyloid polypeptide transgenic mice. Diabetologia. 2015;58(3):575-585. doi: 10.1007/s00125-014-3447-x
- 72. D'Alessio DA, Verchere CB, Kahn SE, Hoagland V, Baskin DG, Palmiter RD, Ensinck JW. Pancreatic Expression and Secretion of Human Islet Amyloid Polypeptide in a Transgenic Mouse. Diabetes. 1994, 43, 1457–1461. 1994 Dec;43(12):1457-1461. doi: 10.2337/diab.43.12.1457
- 73. Haataja L, Gurlo T, Huang CJ, Butler PC. Islet amyloid in type 2 diabetes, and the toxic oligomer hypothesis. Endocr Rev. 2008;29(3):303-316. doi: 10.1210/er.2007-0037
- 74. Cooper ME, McNally PG, Phillips PA, Johnston CI: Amylin stimulates plasma renin concentrations in humans. Hypertension. 1995;26(3):460-464. Doi 10.1161/01.hyp.26.3.460
- 75. Fonseca, S.G.; Gromada, J.; Urano, F. Endoplasmic reticulum stress and pancreatic β-cell death. Trends Endocrinol. Metab. 2011, 22, 266–274, doi:10.1016/j.tem.2011.02.00
- 76. Hull RL, Westermark GT, Westermark P, Kahn SE. Islet amyloid: a critical entity in the pathogenesis of type 2 diabetes. J Clin Endocrinol Metab. 2004;89(8):3629-43. doi: 10.1210/jc.2004-0405
- 77. Kahn SE, Andrikopoulos S, Verchere CB. Islet amyloid: a long-recognized but underappreciated pathological feature of type 2 diabetes. Diabetes. 1999 Feb;48(2):241-53. doi: 10.2337/diabetes.48.2.241
- 78. Butler AE, Janson J, Bonner-Weir S, Ritzel R, Rizza R, Butler PC. 2003 β -Cell deficit and increased β -cell apoptosis in humans with type 2 diabetes. *Diabetes* 2003; 52(1):102–110. doi: 10.2337/diabetes.52.1.102
- 79. Hayden MR, Karuparthi PR, Manrique CM, Lastra G, Habibi J, Sowers JR. Longitudinal ultrastructure study of islet amyloid in the HIP rat model of type

- 2 diabetes mellitus. Exp Biol Med (Maywood), 2007;232(6):772-779. PMID: 17526769
- 80. Opie EL. PATHOLOGICAL CHANGES AFFECTING THE ISLANDS OF LANGERHANS OF THE PANCREAS. J Boston Soc Med Sci. 1900;4(10):251-260. PMID: 19971322
- 81. Opie EL. THE RELATION OE DIABETES MELLITUS TO LESIONS OF THE PANCREAS. HYALINE DEGENERATION OF THE ISLANDS OE LANGERHANS. J Exp Med. 1901 Mar 25;5(5):527-40. doi: 10.1084/jem.5.5.527
- 82. Biessels GJ, Despa F. Cognitive decline and dementia in diabetes mellitus: mechanisms and clinical implications. Nat Rev Endocrinol. 2018;14(10):591-604. doi: 10.1038/s41574-018-0048-7
- 83. Despa F, Goldstein LB. Amylin Dyshomeostasis Hypothesis: Small Vessel-Type Ischemic Stroke in the Setting of Type-2 Diabetes. Stroke. 2021;52(6):e244-e249. doi: 10.1161/STROKEAHA.121.034363
- 84. Ly H, Despa F. Hyperamylinemia as a risk factor for accelerated cognitive decline in diabetes. Expert Rev Proteomics. 2015;12(6):575-577. doi: 10.1586/14789450.2015.1104251
- 85. Ly H, Despa F. Diabetes-related Amylin Dyshomeostasis: A Contributing Factor to Cerebrovascular Pathology and Dementia. J Lipid Atheroscler. 2019;8(2):144-151. doi: 10.12997/jla.2019.8.2.144
- 86. Despa F, Goldstein LB. Amylin Dyshomeostasis Hypothesis: Small Vessel-Type Ischemic Stroke in the Setting of Type-2 Diabetes. Stroke. 2021;52(6):e244e249. doi: 10.1161/STROKEAHA.121.034363 83
- 87. Ma ZA, Zhao Z, Turk J. Mitochondrial Dysfunction and β-Cell Failure in Type 2 Diabetes Mellitus. Experimental Diabetes Research. 2012;2012:703538. doi: 10.1155/2012/703538
- 88. Anello M, Lupi R, Spampinato D, Piro S, Masini M, Boggi U, Del Prato S, Rabuazzo AM, Purrello F, Marchetti P. Functional and morphological alterations of mitochondria in pancreatic beta cells from type 2 diabetic patients. Diabetologia. 2005;48(2):282-289. doi: 10.1007/s00125-004-1627-9
- 89. Prasun P. Mitochondrial dysfunction in metabolic syndrome. Biochim Biophys Acta Mol Basis Dis. 2020;1866(10):165838. doi: 10.1016/j.bbadis.2020.165838
- 90. Weksler-Zangen S. Is Type 2 Diabetes a Primary Mitochondrial Disorder? Cells. 2022;11(10):1617. doi: 10.3390/cells11101617
- 91. Guzzardi MA, Guiducci L, Campani D, La Rosa F, Insilla AC, Bartoli A, Cabiati M, De Sena V, Del Ry S, Burchielli S, Bonino F, Iozzo P. Leptin resistance before and after obesity: evidence that tissue glucose uptake underlies adipocyte enlargement and liver steatosis/steatohepatitis in Zucker rats from early-life stages. Int J Obes (Lond). 2022;46(1):50-58. doi: 10.1038/s41366-021-00941-z
- 92. Ghadge AA, Khaire AA. Leptin as a predictive marker for metabolic syndrome. Cytokine. 2019;121:154735. doi: 10.1016/j.cyto.2019.154735
- 93. Kelly AS, Metzig AM, Schwarzenberg SJ, Norris AL, Fox CF, Steinberger J. Hyperleptinemia and Hypoadiponectinemia in Extreme Pediatric Obesity. Metab Syndr Relat Disord. 2012;10(2):123–127. doi: 10.1089/met.2011.0086

- 94. <u>Frühbeck G, Catalán V., Rodríguez A, , Gómez-Ambrosi J. Adiponectin-leptin ratio: A promising index to estimate adipose tissue dysfunction. Relation with obesity-associated cardiometabolic risk. <u>Adipocyte.</u> 2018;7(1): 57–62. doi: 10.1080/21623945.2017.1402151</u>
- 95. Fisman EZ, Tenenbaum A. Adiponectin: a manifold therapeutic target for metabolic syndrome, diabetes, and coronary disease? Cardiovasc Diabetol. 2014 Jun 23;13:103. doi: 10.1186/1475-2840-13-103
- 96. Ghadge AA, KhairAA, Kuvalekar AA. Adiponectin: A potential therapeutic target for metabolic syndrome. Cytokine & Growth Factor Reviews. 2018;39:151-158. doi: 10.1016/j.cytogfr.2018.01.004
- 97. Menon B, Krishnan R. Role of Leptin in Acute Ischemic Stroke. J Neurosci Rural Pract. 2018 Jul-Sep; 9(3): 376–380. doi: 10.4103/jnrp.jnrp_5_18
- 98. Söderberg S, Stegmayr B, Ahlbeck-Glader C, Slunga-Birgander L, Ahrén B, Olsson T. High leptin levels are associated with stroke. Cerebrovasc Dis. 2003;15(1-2):63-9. doi: 10.1159/000067128
- 99. Beltowski J. Leptin and atherosclerosis. Atherosclerosis. 2006 Nov;189(1):47-60. doi: 10.1016/j.atherosclerosis.2006.03.003
- 100.Berger S, Polotsky VY. Leptin and Leptin Resistance in the Pathogenesis of Obstructive Sleep Apnea: A Possible Link to Oxidative Stress and Cardiovascular Complications. Oxid Med Cell Longev. 2018;2018:5137947. doi: 10.1155/2018/5137947
- 101.United Nations Department of Economics and Social Affairs Populations Division Population Ageing and Sustainable Development. [(accessed on 19 January 2023)]; Available online: https://www.un.org/development/desa/pd/sites/www.un.org.development.desa.pd/files/wpp2022_summary_of_results.pdf
- 102. Vishram JKK, Borglykke A, Andreasen AH, Jeppesen J, Ibsen, H, Jorgensen T, et al. Impact of Age and Gender on the Prevalence and Prognostic Importance of the Metabolic Syndrome and Its Components in Europeans. The MORGAM Prospective Cohort Project. PLoS One. 2014;9(9):e107294. doi: 10.1371/journal.pone.0107294
- 103.Ma XH, Muzumdar R, Yang XM, Gabriely I, Berger R, Barzilai N. Aging Is Associated With Resistance to Effects of Leptin on Fat Distribution and Insulin Action. J Gerontol A Biol Sci Med Sci. 2002;57(6):B225-B231. doi: 10.1093/gerona/57.6.b225
- 104.Gabriely I, Ma XH, Yang XM, Rossetti L, Barzilai N. Leptin resistance during aging is independent of fat mass. *Diabetes* (2002) 51:1016–21. doi: 10.2337/diabetes.51.4.1016
- 105.Stern JH, Rutkowski JM, Scherer PE. Adiponectin, Leptin, and Fatty Acids in the Maintenance of Metabolic Homeostasis through Adipose Tissue Crosstalk. Cell Metab. 2016 May 10;23(5):770-84. doi: 10.1016/j.cmet.2016.04.011
- 106.Messerli FH, Williams B, Ritz E. Essential hypertension. Lancet. 2007;370(9587):591-603. doi: 10.1016/S0140-6736(07)61299-9

- 107. Sowers JR. Insulin resistance and hypertension. Am J Physiol Heart Circ Physiol. 2004;286(5):H1597-602. doi: 10.1152/ajpheart.00026.2004.
- 108.Addison A, Stas S, Hayden MR, James R. Sowers JR. Insulin Resistance and Blood Pressure. Curr Hypertens Rep. 2008;10(4):319-325. doi: 10.1007/s11906-008-0059-
- 109.Correia MLG, Rahmouni K. Role of leptin in the cardiovascular and endocrine complications of metabolic syndrome. Diabetes Obes Metab. 2006;8(6):603-10. doi: 10.1111/j.1463-1326.2005.00562.x
- 110.Haynes WG. Role of leptin in obesity-related hypertension. Exp Physiol. 2005;90(5):683-688. doi: 10.1113/expphysiol.2005.031237
- 111.Dikalov SI, Ungvari Z. Role of mitochondrial oxidative stress in hypertension. Am J Physiol Heart Circ Physiol. 2013;305(10):H1417-27. doi: 10.1152/ajpheart.00089.2013
- 112.Kim JA, Wei Y, Sowers JR. Role of mitochondrial dysfunction in insulin resistance. Circ Res. 2008;102(4):401-414. doi: 10.1161/CIRCRESAHA.107.165472
- 113. Nicolson GL. Metabolic Syndrome and Mitochondrial Function: Molecular Replacement and Antioxidant Supplements to Prevent Membrane Peroxidation and Restore Mitochondrial Function. J Cell Biochem. 2007;100(6):1352-1369. doi:10.1002/jcb.21247
- 114.van Guldener C, Nanayakkara PWB, Stehouwer CDA. Homocysteine and blood pressure. Curr Hypertens Rep. 2003;5(1):26-31. doi: 10.1007/s11906-003-0007-z
- 115.Lopes-Vicente WRP, Rodrigues S, Cepeda FX, Jordão CP, Costa-Hong V, Dutra-Marques ACB, et al. Arterial stiffness and its association with clustering of metabolic syndrome risk factors. Diabetol Metab Syndr. 2017;9:8t. doi: 10.1186/s13098-017-0286-1
- 116. Schillaci G, Pirro M, Vaudo G, Mannarino MR, Savarese G, Pucci G, Franklin SS, Mannarino E. Metabolic Syndrome Is Associated With Aortic Stiffness in Untreated Essential Hypertension. Hypertension. 2005 Jun;45(6):1078-82. doi: 10.1161/01.HYP.0000165313.84007.7d
- 117.Mitchell G F, Hwang SJ, Vasan RS, Larson MG, Pencina MJ, Hamburg NM, et al. Arterial stiffness and cardiovascular events: the Framingham Heart Study. Circulation. 2010;121:505–511. 10.1161/CIRCULATIONAHA.109.886655
- 118.Brillante DG, O'Sullivan AJ, Howes LG. Arterial stiffness in insulin resistance: the role of nitric oxide and angiotensin II receptors. Vasc Health Risk Manag. 2009;5(1):73–78. 10.2147/VHRM.S3784
- 119. Jia G, Aroor AR, DeMarco VG, Martinez-Lemus LA, Meininger GA, Sowers JR. Vascular stiffness in insulin resistance and obesity. Front Physiol. 2015;6:231. doi: 10.3389/fphys.2015.00231
- 120.Yan Y, Wang J, Chaudhry MA, et al. Metabolic Syndrome and Salt-sensitive Hypertension in Polygenic Obese TALLYHO/JngJ Mice: Role of Na/K-ATPase Signaling. Int J Mol Sci. 2019;20(14):3495. doi: 10.3390/ijms20143494
- 121.Fujita T. Aldosterone in salt-sensitive hypertension and metabolic syndrome. J Mol Med (Berl). 2008;86(6):729-34. doi: 10.1007/s00109-008-0343-1 –

- 122.Alexander CM, Landsman PB, Grundy SM. Metabolic Syndrome and Hyperglycemia: Congruence and Divergence. Am J Cardiol. 2006;98(7):982-5. doi: 10.1016/j.amjcard.2006.04.046
- 123.Oguntibeju OO. Type 2 diabetes mellitus, oxidative stress and inflammation: Examining the links. Int. J. Physiol. Pathophysiol. Pharmacol. 2019, *11*, 45–63.
- 124. Yang Y, Hayden MR, Sowers S, Bagree SV, Sowers JR. Retinal redox stress and remodeling in cardiometabolic syndrome and diabetes. Oxid. Med. Cell Longev. 2010;3(6):392-403. doi: 10.4161/oxim.3.6.14786
- 125.Hayden MR, Salam M, Sowers, JR. Reactive Oxygen Species and Diabetic Peripheral Neuropathy—A Closer Look, Chapter 149; Lahey, I., Ed.; Systems Biology of Reactive Oxygen Species and Antioxidants; Springer: Berlin/Heidelberg, Germany, 2014; pp. 3375–3400.
- 126.Hayden MR, Whaley-Connell A, Sowers JR. Renal redox stress and remodeling in metabolic syndrome, type 2 diabetes mellitus, and diabetic nephropathy: Paying homage to the podocyte. Am. J. Nephrol. 2005;25(6):553–569. doi: 10.1159/000088810
- 127. Hayden MR, Sowers JR. Isletopathy in Type 2 diabetes mellitus: Implications of islet RAS, islet fibrosis, islet amyloid, remodeling, and oxidative stress. Antiox Redox Signal. 2007;9(7):891-910. doi: 10.1089/ars.2007.1610
- 128.Jia G, Hill MA, Sowers JR. Diabetic Cardiomyopathy: An Update of Mechanisms Contributing to This Clinical Entity. Circ Res. 2018 Feb 16;122(4):624-638. doi: 10.1161/CIRCRESAHA.117.311586
- 129.Climie RE, van Sloten TT, Bruno RM, Taddei S, Empana JP, Stehouwer CDA, Sharman JE, Boutouyrie P, Laurent S. Macrovasculature and Microvasculature at the Crossroads Between Type 2 Diabetes Mellitus and Hypertension. Hypertension. 2019;73(6):1138-1149. doi: 10.1161/HYPERTENSIONAHA.118.11769
- 130.Brownlee M. The pathobiology of diabetic complications: a unifying mechanism. Diabetes. 2005;54(6):1615-1625. doi: 10.2337/diabetes.54.6.1615
- 131.Giacco F, Brownlee M. Oxidative stress and diabetic complications. Circ Res. 2010;107(9): 1058–1070. doi: 10.1161/CIRCRESAHA.110.223545
- 132.Hayden MR, Chowdhury NA, Cooper SA, Whaley-Connell A, Habibi J, Witte L, Wiedmeyer C, Manrique CM, Lastra G, Ferrario C, Stump C, Sowers JR. Proximal tubule microvilli remodeling and albuminuria in the Ren2 transgenic rat. Am J Physiol Renal Physiol. 2007;292(2):F861-7. doi: 10.1152/ajprenal.00252.2006
- 133.Hayden MR, Sowers JR. Childhood-Adolescent Obesity in the Cardiorenal Syndrome: Lessons from Animal Models. Cardiorenal Med. 2011; 1(2): 75–86. doi: 10.1159/000327022
- 134.Habibi J, Aroor AR, Sowers JR, Jia G, Hayden MR, DeMarco VG, et al. Sodium glucose transporter 2 (SGLT2) inhibition with empagliflozin improves cardiac diastolic function in a female rodent model of diabetes. Cardiovasc Diabetol. 2017; 16: 9. doi: 10.1186/s12933-016-0489-z

- 135.Hayden MR, Chowdhury N, Govindarajan G, Karuparthi PR, Habibi J, Sowers JR. Myocardial myocyte remodeling and fibrosis in the cardiometabolic syndrome. J Cardiometab Syndr. 2006 Fall;1(5):326-33. doi: 10.1111/j.1559-4564.2006.05626.x
- 136. <u>Hayden MR, Habibi J, TJoginpally</u> T, <u>Karuparthi PR, Sowers JR</u>. Ultrastructure Study of Transgenic Ren2 Rat Aorta Part 1: Endothelium and Intima. <u>Cardiorenal Med.</u> 2012;2(1): 66–82. doi: <u>10.1159/000335565</u>
- 137. Hayden MR. Ultrastructure study of the transgenic REN2 rat aorta part 2: media, external elastic lamina, and adventitia. Biomedical Reviews. 2019;30(111); doi: 10.14748/bmr.v30.6392
- 138.Salameh TS, Shah GN, Price TO, Hayden MR, Banks WA. Blood-Brain Barrier Disruption and Neurovascular Unit Dysfunction in Diabetic Mice: Protection with the Mitochondrial Carbonic Anhydrase Inhibitor Topiramate. J Pharmacol Exp Ther. 2016;359(3):452-459. doi: 10.1124/jpet.116.237057
- 139.Hayden MR. Empagliflozin Ameliorates Tunica Adiposa Expansion and Vascular Stiffening of the Descending Aorta in Female *db/db* Mice. Adipobiology. 2020; 10(2019):1 doi: 10.14748/adipo.v10.6539
- 140.Stover PJ. One-Carbon Metabolism–Genome Interactions in Folate-Associated Pathologies. J Nutr. 2009;139(12):2402–2405. doi: 10.3945/jn.109.113670
- 141.Hayden MR, Tyagi SC. Homocysteine and reactive oxygen species in metabolic syndrome, type 2 diabetes mellitus, and atheroscleropathy: The pleiotropic effects of folate supplementation. Nutr. J. 2004;3:4. doi: 10.1186/1475-2891-3-4
- 142. Finer S, Saravanan P, Hitman G, Yajnik C. The role of the one-carbon cycle in the developmental origins of Type 2 diabetes and obesity. Diabet. Med. 2014, *31*, 263–272. doi: 10.1111/dme.12390
- 143. Joshi MB, Baipadithaya G, Balakrishnan A, Hegde M, Vohra M, Ahamed R, Nagri SK, Ramachandra L, Satyamoorthy K. Elevated homocysteine levels in type 2 diabetes induce constitutive neutrophil extracellular traps. Sci. Rep. 2016;6:36362. doi: 10.1038/srep36362
- 144. Huang T, Ren J, Haung J, Li D. Association of homocysteine with type 2 diabetes: A meta-analysis implementing Mendelian randomization approach. BMC Genom. 2013;14:867. doi: 10.1186/1471-2164-14-867
- 145. Piazzolla G, Candigliota M, Fanelli M, Castrovilli A, Berardi E, Antonica G, Battaglia S, Solfrizzi V, Sabbà C, Tortorella C. Diabetol Metab Syndr. 2019;11:87. doi: 10.1186/s13098-019-0484-0
- 146.Zorov DB, Filburn CR, Klotz LO, Zweier JL, Sollott SJ. Reactive oxygen species (ROS)-induced ROS release: a new phenomenon accompanying induction of the mitochondrial permeability transition in cardiac myocytes. J Exp Med. 2000;192(7):1001-1914. doi: 10.1084/jem.192.7.1001
- 147.Burgess K, Calli Bennett C, Mosnier H, Kwatra N, Bethel F, Jadavji NM. The Antioxidant Role of One-Carbon Metabolism on Stroke. Antioxidants (Basil). 2020 Nov 17;9(11):1141. doi: 10.3390/antiox9111141
- 148.Welch GN, Loscalzo J. Homocysteine and atherothrombosis. N Engl J Med. 1998; 338:1042–50. doi: 10.1056/NEJM199804093381507

- 149.Arruda VR, von Zuben PM, Chiaparini LC, Annichino-Bizzacchi JM, Costa FF. The mutation Ala677 → Val in the methylene tetrahydrofolate reductase gene: a risk factor for arterial disease and venous thrombosis. Thromb Haemost. 1997;77:818–821. PMID: 9184384\
- 150.Feix A, Fritsche-Polanz R, Kletzmayr J, Vychytil A, Horl WH, Sunder-Plassmann G, Fodinger M. Increased prevalence of combined MTR and MTHFR genotypes among individuals with severely elevated total homocysteine plasma levels. Am J Kidney Dis. 2001;956–964. doi: 10.1053/ajkd.2001.28581
- 151.Woo KS, Chook P, Lolin YI, Sanderson JE, Metreweli C, Celermajer DS. Folic acid improves arterial endothelial function in adults with hyperhomocystinemia. J Am Coll Cardiol. 1999;34:2002–2006. doi: 10.1016/S0735-1097(99)00469-6
- 152.Liao JK. Linking endothelial dysfunction with endothelial cell activation. J Clin Invest. 2013; 123(2): 540–541. doi: 10.1172/JCI66843
- 153.Christ M, Bauersachs J, Liebetrau C, Heck M, Gunther A, Wehling M. Glucose Increases Endothelial-Dependent Superoxide Formation in Coronary Arteries by NAD(P)H Oxidase Activation: Attenuation by the 3-Hydroxy-3-Methylglutaryl Coenzyme A
- 154. <u>Padilla J, Ramirez-Perez FI, Habibi J, Bostick B, Aroor AR, Hayden MR, et al.</u> Regular exercise reduces endothelial cortical stiffness in Western diet-fed female mice. <u>Hypertension. 2016; 68(5): 1236–1244.</u>doi: 10.1161/HYPERTENSIONAHA.116.07954
- 155. <u>Jia</u> G, <u>Habibi</u> J, <u>Aroor</u> AR, <u>Martinez-Lemus</u> LA, <u>DeMarco</u> VG, <u>Ramirez-Perez</u> FI, et al. Endothelial Mineralocorticoid Receptor Mediates Diet Induced Aortic Stiffness in Females. <u>Circ Res. 2016;118(6): 935–943.</u> doi: <u>10.1161/CIRCRESAHA.115.308269</u>
- 156. Sowers JR, Habibi J, Jia G, Bostick B, Manrique-Acevedo C, Lastra G, et al. Endothelial sodium channel activation promotes cardiac stiffness and diastolic dysfunction in Western diet fed female mice. Metabolism. 2020;109:154223. doi: 10.1016/j.metabol.2020.154223
- 157.Stepp DW, Ou J, Ackerman AW, Welak S, Klick D, Pritchard KA Jr. Native LDL and minimally oxidized LDL differentially regulate superoxide anion in vascular endothelium in situ. Am J Physiol Heart Circ Physiol. 2002;283(2):H750–759. doi: 10.1152/ajpheart.00029.2002
- 158.Zou MH, Shi C, Cohen RA. Oxidation of the zinc-thiolate complex and uncoupling of endothelial nitric oxide synthase by peroxynitrite. J Clin Invest. 2002;109(6):817–826. doi: 10.1172/JCI14442
- 159. Verma S, Wang CH, Li SH, Dumont AS, Fedak PW, Badiwala MV, Dhillon B, Weisel RD, Li RK, Mickle DA, Stewart DJ. A self fulfilling prophecy: C-reactive protein attenuates nitric oxide production and inhibits angiogenesis. Circulation. 2002;106(8):913–919. doi: 10.1161/01.cir.0000029802.88087.5e
- 160.Guzik TJ, Mussa S, Gastaldi D, Sadowski J, Ratnatunga C, Pillai R, Channon KM. Mechanisms of increased vascular superoxide production in human diabetes

- mellitus: role of NAD(P)H oxidase and endothelial nitric oxide synthase. Circulation. 2002;105(14): 1656-1662. doi: 10.1161/01.cir.0000012748.58444.08
- 161.Aroor AR, Habibi J, Kandikattu HK, Hayden MR, Garro-Kacher M, Bender SB. et al. Dipeptidyl peptidase-4 (DPP-4) inhibition with linagliptin reduces western diet-induced myocardial TRAF3IP2 expression, inflammation and fibrosis in female mice. Cardiovasc Diabetol. 2017 May 5;16(1):61. doi: 10.1186/s12933-017-0544-4
- 162.Ren J, Pulakat L, Whaley-Connell A, Sowers, JR. Mitochondrial biogenesis in the metabolic syndrome and cardiovascular disease. J. Mol. Med. (Berl). 2010; 88: 993–1001. doi: 10.1007/s00109-010-0663-9
- 163.Petersen KF, Befroy D, Dufour S, Dziura J, Ariyan C, Rothman DL, et al. Mitochondrial dysfunction in the elderly: possible role in insulin resistance. Science. 2003; 300: 1140–1142. doi: 10.1126/science.1082889
- 164. Vasquez-Vivar J, Kalyanaraman B, Martasek P, Hogg N, Masters BS, Karoui H, Tordo P, Pritchard KA Jr. Superoxide generation by endothelial nitric oxide synthase: the influence of cofactors. Proc Natl Acad Sci U S A.1998;95:9220–9225. doi: 10.1073/pnas.95.16.9220
- 165.Litvinova L, Atochin DN, Fattakhov N, Vasilenko M, Zatolokin P, Kirienkova E. Nitric oxide and mitochondria in metabolic syndrome. Front Physiol. 2015; 6: 20. doi: 10.3389/fphys.2015.00020
- 166.Martin SD, McGee SL. (2014). The role of mitochondria in the aetiology of insulin resistance and type 2 diabetes. Biochim. Biophys. Acta. 2014;1840:1303–1312. doi: 10.1016/j.bbagen.2013.09.019
- 167.Hayden MR. Hypothesis: Astrocyte Foot Processes Detachment from the Neurovascular Unit in Female Diabetic Mice May Impair Modulation of Information Processing-Six Degrees of Separation. Brain Sci. 2019;9(4):83. doi: 10.3390/brainsci9040083
- 168.du Preez HN, Aldous C, Hayden MR, Kruger HG, Lin J. Pathogenesis of COVID-19 described through the lens of an undersulfated and degraded epithelial and endothelial glycocalyx. FASEB J. 2022;36(1):e22052. doi: 10.1096/fj.202101100RR
- 169.Ando Y, Okada H, Takemura G, Suzuki K, Takada C, Tomita H, Zaikokuji R, Hotta Y, Miyazaki N, Yano H, et al. Brain-Specific Ultrastructure of Capillary Endothelial Glycocalyx and Its Possible Contribution for Blood Brain Barrier. Sci Rep. 2018;8(1):17523. doi: 10.1038/s41598-018-35976-2
- 170.Sampei S, Okada H, Tomita H, Takada C, Suzuki K, Kinoshita T, Kobayashi R, Fukuda H, Kawasaki Y, Nishio A, et al. Endothelial Glycocalyx Disorders May Be Associated With Extended Inflammation During Endotoxemia in a Diabetic Mouse Model. Front Cell Dev Biol. 2021;9:623582. doi: 10.3389/fcell.2021.623582
- 171.Okada, H, Takemura G, Suzuki K, Oda K, Takada C, Hotta Y, et al. Three-dimensional ultrastructure of capillary endothelial glycocalyx under normal and experimental endotoxemic condition. Crit. Care. 2017;21(1):261. doi: 10.1186/s13054-017-1841-8

- 172.Kutuzov N, Flyvbjerg H, Lauritzen M. Contributions of the glycocalyx, endothelium, and extravascular compartment to the blood-brain barrier. Proc Natl Acad Sci. USA. 2018;115:E9429-E9438. doi: 10.1073/pnas.1802155115
- 173.Reitsma S, Slaaf DW, Vink H, van Zandvoort MA, oude Egbrink MG. The endothelial glycocalyx: composition, functions, and visualization. Pflugers Arch. 2007; 454:345–359. doi:10.1007/s00424-007-0212-8
- 174.Hayden MR, Sowers JR, Tyagi SC. The central role of vascular extracellular matrix and basement membrane remodeling in metabolic syndrome and type 2 diabetes: the matrix preloaded. Cardiovasc Diabetol. 2005;4:9. doi: 10.1186/1475-2840-4-9
- 175. Ericsson A, Tonelius P, Lal M, Sabirsh A, Böttcher G, William-Olsso L, et al. The effects of dual PPAR α/γ agonism compared with ACE inhibition in the BTBRob/ob mouse model of diabetes and diabetic nephropathy. Physiol Rep. 2017;5(5): e13186. doi: 10.14814/phy2.13186
- 176.Kolárová H, Ambruzová B, Šindlerová LŠ, Klinke A, Kubala L. Modulation of endothelial glycocalyx structure under inflammatory conditions. Mediators Inflamm. 2014:694312. doi: 10.1155/2014/694312
- 177.Dogné S, Flamion B, Caron N. Endothelial Glycocalyx as a Shield Against Diabetic Vascular Complications: Involvement of Hyaluronan and Hyaluronidases. Arterioscler Thromb Vasc Biol. 2018 Jul;38(7):1427-1439. doi: 10.1161/ATVBAHA.118.310839
- 178.Stackhouse TK, Mishra A. Neurovascular Coupling in Development and Disease: Focus on Astrocytes. Front Cell Dev Biol. 2021;9:702832. doi: 10.3389/fcell.2021.702832
- 179.Alves NG, Trujillo AN, Breslin JW, Yuan SY. Sphingosine-1-Phosphate (S1P) Reduces Hemorrhagic Shock and Resuscitation-Induced Microvascular Leakage by Protecting Endothelial Mitochondrial Integrity. Shock. 2019;52(4): 423–433. doi: 10.1097/SHK.0000000000001280 180.
- 180.Liu XY, Xu HX, Li JK, Zhang D, Ma XH, Huang LN, Lü JH, Wang XZ. Neferine Protects Endothelial Glycocalyx via Mitochondrial ROS in Lipopolysaccharide-Induced Acute Respiratory Distress Syndrome. Front Physiol. 2018 Feb 22;9:102. doi: 10.3389/fphys.2018.00102
- 181.Alhayaza R, Haque E, Karbasiafshar C, Sellke FW, Abid MR. The Relationship Between Reactive Oxygen Species and Endothelial Cell Metabolism. Front Chem. 2020;8:592688. doi:10.3389/fchem.2020.592688
- 182.Koziel A, Sobieraj I, Jarmuszkiewicz W. Increased activity of mitochondrial uncoupling protein 2 improves stress resistance in cultured endothelial cells exposed in vitro to high glucose levels. 2015;1;309(1):H147-56. doi: 10.1152/ajpheart.00759.2014
- 183.Islam MN, Das SR, Emin Mt, Wei M, Sun L, Westphalen K, Rowlands DJ, Quadri SK, Bhattacharya S, Bhattacharya J. Mitochondrial transfer from bone-marrow-derived stromal cells to pulmonary alveoli protects against acute lung injury. Nat Med. 2012 Apr 15;18(5):759-65. doi: 10.1038/nm.2736

- 184.Yen W, Cai B, Yang J, Zhang L, Zeng M, Tarbell JM, et al. Endothelial surface glycocalyx can regulate flow-induced nitric oxide production in microvessels *in vivo*. PLoS One. 2015;10(1):e0117133. doi: 10.1371/journal.pone.0117133
- 185. Valenti D, Vacca RA, Moro L, Atlante A. Mitochondria Can Cross Cell Boundaries: An Overview of the Biological Relevance, Pathophysiological Implications and Therapeutic Perspectives of Intercellular Mitochondrial Transfer. 2021;22(15):8312. doi: 10.3390/ijms22158312
- 186.Broekhuizen LN, Lemkes BA, Mooij HL, Meuwese MC, Verberne H, Holleman F, Schlingemann RO, Nieuwdorph M. Stroes ESG, Vink H. Effect of sulodexide on endothelial glycocalyx and vascular permeability in patients with type 2 diabetes mellitus. Diabetologia. 2010;53(12):2646-2655. doi:10.1007/s00125-010-1910-x
- 187.Lee M, McGeer EG, McGeer PL. Sodium thiosulfate attenuates glial-mediated neuroinflammation in degenerative neurological diseases. J Neuroinflammation. 2016;13:32. doi:10.1186/s12974-016-0488-8
- 188.du Preez HN, Aldous C, Kruger HG, Lin J. N-acetylcysteine and other sulfurdonors as a preventative and adjunct therapy for COVID-19. Advances in Pharmacological and Pharmaceutical Sciences Journal. FASEB J. 2022;36(1):e22052. doi: 10.1096/fj.202101100RR
- 189.Hotamisligil GS, Shargill NS, Spiegelman BM. Adipose expression of tumor necrosis factor-alpha: direct role in obesity-linked insulin resistance. Science. 1993 Jan 1;259(5091):87-91. doi: 10.1126/science.7678183
- 190.Hotamisligil GS, Spiegelman BM. Tumor necrosis factor alpha: a key component of the obesity-diabetes link. Diabetes.1994;43(11):1271-8. doi: 10.2337/diab.43.11.1271
- 191.Esposito K, Giugliano D. The metabolic syndrome and inflammation: association or causation? Nutr Metab Cardiovasc Dis. 2004 Oct;14(5):228-32. doi: 10.1016/s0939-4179.
- 192.Mohammad S, Thiemermann C. Role of Metabolic Endotoxemia in Systemic Inflammation and Potential Interventions. Front Immunol. 2021;11:594150. doi: 10.3389/fimmu.2020.594150
- 193.Singh H, Miyamoto S, Darshi M, et al. Gut Microbial Changes in Diabetic db/db Mice and Recovery of Microbial Diversity upon Pirfenidone Treatment. *Microorganisms*. 2020;8(9):1347. Published 2020 Sep 3. doi:10.3390/microorganisms8091347
- 194.Di Tommaso N, Santopaolo F, Gasbarrini A, Ponziani FR. The Gut–Vascular Barrier as a New Protagonist in Intestinal and Extraintestinal Diseases. Int. J. Mol. Sci. 2023, 24, 1470. https://doi.org/10.3390/ijms24021470
- 195.Tilg H, Kaser A. <u>Gut microbiome, obesity, and metabolic dysfunction.</u> J Clin Invest. 2011;121(6):2126-32. doi: 10.1172/JCI58109
- 196.Tilg H, Moschen AR. Adipocytokines: mediators linking adipose tissue, inflammation and immunity. Nat Rev Immunol. 2006;6(10):772–783. doi: 10.1038/nri1937.

- 197.Ley RE, Backhed F, Turnbaugh P, Lozupone CA, Knight RD, Gordon JI. Obesity alters gut microbial ecology. Proc Natl Acad Sci U S A. 2005;102(31):11070–11075. doi: 10.1073/pnas.0504978102
- 198. Schmidt MI, Duncan BB, Sharrett AR, Lindberg G, Savage PJ, Offenbacher S, Azambuja MI, Tracy RP, Heiss G. Markers of inflammation and prediction of diabetes mellitus in adults (Atherosclerosis Risk in Communities study): a cohort study. Lancet. 1999;353(9165):1649-52. doi: 10.1016/s0140-6736(99)01046-6
- 199.Duncan BB, Schmidt MI, Pankow JS, Ballantyne CM, Couper D, Vigo A, Hoogeveen R, Folsom AR, Heiss G. Low-grade systemic inflammation and the development of type 2 diabetes: the atherosclerosis risk in communities study. Atherosclerosis Risk in Communities Study. Diabetes. 2003;52(7):1799-805. doi: 10.2337/diabetes.52.7.1799
- 200. Scheithauer TPM, Rampanelli E, Nieuwdorp M, Vallance BA, Verchere CB, van Raalte DH, Herrema H. Gut Microbiota as a Trigger for Metabolic Inflammation in Obesity and Type 2 Diabetes. Front Immunol. 2020;11:571731. doi: 10.3389/fimmu.2020.571731. eCollection 2020753(04)80048-6
- 201.de Git CG, Adan RAH. Leptin resistance in diet-induced obesity: the role of hypothalamic inflammation. Obesity rev. 2015 Mar;16(3):207-24. doi: 10.1111/obr.12243
- 202. Pérez-Pérez A, Sánchez-Jiménez F, Vilariño-García T, Sánchez-Margalet V. Role of Leptin in Inflammation and Vice Versa. Int J Mol Sci. 2020 Aug 16;21(16):5887. doi:10.3390/ijms21165887
- 203.Kiernan K, MacIver NJ. The Role of the Adipokine Leptin in Immune Cell Function in Health and Disease. Front Immun. Front Immunol. 2021;11:622468. doi: 10.3389/fimmu.2020.622468
- 204.Otero M, Lago R, Gomez R, Dieguez C, Lago F, Gómez-Reino J, Gualillo O. Towards a pro-inflammatory and immunomodulatory emerging role of leptin. Rheumatology (Oxford). 2006 Aug;45(8):944-50. doi: 10.1093/rheumatology/kel157
- 205.Martin SS, Qasim A, Reilly MP. Leptin resistance: a possible interface of inflammation and metabolism in obesity-related cardiovascular disease. J Am Coll Cardiol. 2008;52(15):1201-1210. doi: 10.1016/j.jacc.2008.05.060
- 206.<u>Lau</u> DCW, Dhillon B, Yan H, Yan H, Szmitko PE, Verma S. Adipokines: molecular links between obesity and atherosclerosis. Am J Physiol Heart Circ Physiol. 2005;288(5):H2031-41. doi: 10.1152/ajpheart.01058.2004
- 207. Molica F, Morel S, Kwak BR, Rohner-Jeanrenaud F, Steffens S. Adipokines at the crossroad between obesity and cardiovascular disease. Thromb Haemost. 2015;113(3):553-566. doi: 10.1160/TH14-06-0513
- 208.Mohammadi M, Gozashti MH, Aghadavood M, Mehdizadeh MR, Hayatbakhsh MM. Clinical Significance of Serum IL-6 and TNF-α Levels in Patients with Metabolic Syndrome. Rep Biochem Mol Biol. 2017 Oct;6(1):74-79. PMID: 29090232

- 209.Bing C. Is interleukin-1β a culprit in macrophage-adipocyte crosstalk in obesity? Adipocyte. 2015;4(2): 149–152. Doi: 10.4161/21623945.2014.979661
- 210.<u>Henderson</u> GC. Plasma Free Fatty Acid Concentration as a Modifiable Risk Factor for Metabolic Disease. <u>Nutrients.</u> 2021; 13(8): 2590. doi: 10.3390/nu13082590
- 211.Busslera S, Penkea M, Flemminga G, Elhassand YS, Kratzschc J, Sergeyeva E et al. Novel Insights in the Metabolic Syndrome in Childhood and Adolescence. Horm Res Paediatr. 2017;88(3-4):181-193. doi: 10.1159/000479510
- 212.Gao X, Salomon C, Freeman DJ. Extracellular Vesicles from Adipose Tissue-A Potential Role in Obesity and Type 2 Diabetes? Front Endocrinol (Lausanne). 2017;8:202. doi:10.3389/fendo.2017.00202
- 213.Liu Y, Wang C, Wei M, Yang G, Yuan L. Multifaceted Roles of Adipose Tissue-Derived Exosomes in Physiological and Pathological Conditions. Front Physiol. 2021;12: 669429. doi: 10.3389/fphys.2021.669429
- 214.Deng ZB, Poliakov A, Hardy RW, Clements R, Liu C, Liu Y, et al. (2009). Adipose tissue exosome-like vesicles mediate activation of macrophage-induced insulin resistance. Diabetes. 2009;58(11):2498-505. doi: 10.2337/db09-0216 58, 2498–2505
- 215.Pan Y, Hui X, Hoo RLC, Ye D, Chan CYC, Feng T, et al. Adipocyte-secreted exosomal microRNA-34a inhibits M2 macrophage polarization to promote obesity-induced adipose inflammation. J. Clin. Invest. 2019;129:834–849. doi: 10.1172/JCI123069
- 216.Kwan HY, Chen M, Xu K, Chen B. The impact of obesity on adipocyte-derived extracellular vesicles. Cellular and Molecular Life Sciences. 2021;78:7275–7288. doi: https://doi.org/10.1007/s00018-021-03973-w
- 217. Żbikowski A, Błachnio-Zabielska A, Galli M, Zabielski P. Adipose-Derived Exosomes as Possible Players in the Development of Insulin Resistance. Int J Mol Sci. 2021;22(14):7427. doi: 10.3390/ijms22147427
- 218.Guglielmi V, D'Adamo M, Menghini R, Marina C, Paolo G, Massimo F, Paolo S. MicroRNA 21 is up-regulated in adipose tissue of obese diabetic subjects. Nutr Healthy Aging. 2017;4(2):141-145. doi:10.3233/NHA-160020
- 219.Tryggestad JB, Teague AM, Sparling DP, Jiang S, Chernausek SD. Macrophage-Derived microRNA-155 Increases in Obesity and Influences Adipocyte Metabolism by Targeting Peroxisome Proliferator-Activated Receptor Gamma. Obesity (Silver Spring). 2019;27(11):1856-1864. doi:10.1002/oby.22616
- 220.<u>Roberts</u> TC, <u>Morris</u> KV, <u>Wood</u> MJA. The role of long non-coding RNAs in neurodevelopment, brain function and neurological disease. <u>Philos Trans R Soc Lond B Biol Sci.</u> 2014; 369(1652): 20130507. doi: 10.1098/rstb.2013.0507
- 221.Riva P, Ratti A, Venturin M <u>The Long Non-Coding RNAs in Neurodegenerative Diseases: Novel Mechanisms of Pathogenesis.</u> Curr Alzheimer Res. 2016;13(11):1219-1231. doi: 10.2174/1567205013666160622112234
- 222.Chadt A, Scherneck S, Joost HG, Al-Hasanin H. Molecular links between Obesity and Diabetes: "Diabesity". Endotext [Internet]. South Dartmouth (MA): MDText.com, Inc.; 2000. 2018. WWW.ENDOTEXT.ORG. https://www.ncbi.nlm.nih.gov/books/NBK279051/ (last viewed on Jan 19, 2023).

- 223.Simons LA, Simons J, Friedlander Y, McCallum J. Is Prediction of Cardiovascular Disease and All-cause Mortality Genuinely Driven by the Metabolic Syndrome, and Independently from its Component Variables? The Dubbo Study. Heart, Lung and Circulation. 2011;20(4):214-219. doi: https://doi.org/10.1016/j.hlc.2010.12.005
- 224. Reilly MP, Rader DJ. The Metabolic Syndrome More Than the Sum of Its Parts? Circulation. 2003;108:1546–1551. https://doi.org/10.1161/01.CIR.0000088846.10655.EO
- 225.Reaven GM. The metabolic syndrome: is this diagnosis necessary? *The American Journal of Clinical Nutrition*. 2006;83(6):1237–1247. doi: https://doi.org/10.1093/ajcn/83.6.1237_
- 226.Kahn R, Buse J, Ferranninin E, Stern M. <u>American Diabetes Association</u>; <u>European Association for the Study of Diabetes</u>. The metabolic syndrome: time for a critical appraisal: joint statement from the American Diabetes Association and the European Association for the Study of Diabetes. Diabetes Care. 2005;28(9):2289-304. doi: 10.2337/diacare.28.9.2289
- 227.Ferrannini E. Metabolic syndrome: a solution in search of a problem. J Clin Endocrinol Metab. 2007;92(2):396-398. doi: 10.1210/jc.2006-0944

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