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Article

Polyphenol Degradation Kinetics of Specialty Coffee in Different Presentations

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Abstract

Polyphenols are chemical compounds found in plants, and coffee is an important source of them. The objective of the study was to evaluate the kinetics of polyphenol degradation in a blend of specialty coffee (green, roasted and roasted-ground beans), packaged in eight different packages, under accelerated storage conditions. The samples were stored at 40, 50 and 60 °C for 12, 8 and 4 days, respectively. The degradation kinetics was modelled by determining the reaction order, rate constants (k), activation energies (E_a) using the Arrhenius model, and half-life. The degradation followed zero-order kinetics. The rate constant (k) varied between 0.437 and 9.534 days $^{-1}$ (40-60 °C). The E_a ranged from 49.321 to 118.04 kJ*mol $^{-1}$. The predicted longest half-life for daily storage at 25 °C was obtained for: vacuum packed green beans (27.16 months), vacuum roasted beans (3.14 months) and ground roasted coffee in trilaminated foil with valve (40.21 months). Polyphenol retention decreased significantly with increasing temperature. For green bean, roasted bean and roasted ground coffee, the packaging that showed the best protection for the coffee was vacuum packaging and trilaminate with valve respectively, being crucial for preserving these bioactive compounds.

Keywords: specialty coffee; polyphenols; reaction order; decay constant; activation energy; accelerated storage; Arrhenius; half-life

1. Introduction

Coffee is one of the world's popular beverages, noted for its psychoactive effects, pleasant taste and an important source of polyphenols [1], compounds that possess several bioactive properties that can positively influence human health [2–14]. Specialty coffees are characterized by the uniqueness of their quality and origin, from the selection of the seedlings to the preparation of the beverage itself [15]. Flavor and aroma are the important properties for assessing coffee quality [16,17] and consumer preferences [18]. The composition of coffee consists mainly of caffeine, diterpene, kahweol, chlorogenic acid and phenols [19], these compounds including volatiles, lipids, phenolic and alkaloids contribute to the basic taste sensation of coffee beverages, providing bitterness, astringency, strength and body to the coffee brew [20–22]. During roasting, simultaneous chemical reactions occur that promote the degradation of proteins, sugars, trigonelline, and chlorogenic acids [23], and the formation of substances resulting from Mallard reactions and Strecker degradation, which influence both the flavor and aroma of coffee [24,25]. Melanoidin is obtained from non-enzymatic roasting, which is responsible for the antioxidant activity of coffee [26]. The chlorogenic acid content decreases during this stage, while the melanoidin content increases [27].



Polyphenols are compounds with structures that oxidize easily, which can lead to the degradation of their antioxidant capacities [28,29]. There are two main forms of polyphenol degradation: enzymatic and non-enzymatic methods [30,31]. Enzymatic degradation involves enzymes such as polyphenol oxidase, lipoxygenase, and peroxidase [32]. Non-enzymatic degradation includes Maillard reactions, which cause phenol oxidation [33]. However, these are not the only ways in which polyphenols can be destroyed, but rather the main reactions that take place in food.

Studies show the presence of phenolic acids in green coffee (chlorogenic acid) and roasted coffee (other phenolic acids) [34–36], scientifically recognizing that coffee changes its chemical composition during the roasting process [25,37]. Roasted and ground coffee is considered a stable product [25]. The phenolic compounds in coffee are directly or indirectly related to the quality of the coffee in the cup [38], based on the presence of chlorogenic acids (6-12%) [35], which are responsible for the pigmentation, aroma formation, and astringency of coffee [39]. However, the thermal degradation of chlorogenic acids during roasting results in phenolic substances that contribute to bitterness [36].

It has been shown that the amount of polyphenols in coffee can vary significantly depending on the type of coffee, the roasting process, the extraction method, and storage conditions. In addition, factors such as temperature and packaging material play a crucial role in the preservation of these bioactive compounds [40,41]. Indicate that the rate of degradation of chlorogenic acids increases significantly with temperature during the storage of roasted coffee. Likewise, [7,42] show that the matrix, type of processing, and packaging are determining variables in the preservation of antioxidants [23]. Demonstrated that the roasting process has a negative influence on bioactive compounds.

The quantification, analysis, and study of the stability of polyphenols in coffee are essential for understanding their antioxidant potential and health benefits. The kinetics of polyphenol loss in coffee have been predominantly characterized as a zero-order reaction, indicating that the rate of loss does not depend on the concentration of the reactant, an observation that is consistent with the results of previous studies that analyzed the stability of these compounds [43–45]. When comparing linear regressions and R² between zero-order and first-order models, the results play a fundamental role in validating which model best describes the kinetics of polyphenol loss.

This study focuses on how different packaging and storage conditions affect the stability of polyphenols in green, roasted, and ground coffee beans, using the Folin-Ciocalteu method for measurement. This method is known for its ability to quantify phenolic compounds present in many matrices, including beverages and plant extracts, and is particularly valued for its robustness and accuracy [46,47]. The objective of the study was to evaluate the kinetics of deterioration of total polyphenols in a blend of specialty coffee (Coffea arabica) in green beans, roasted beans, and ground roasted beans packaged in eight packages subjected to accelerated storage at temperatures of 40, 50, and 60 °C for 12, 8, and 4 days, respectively.

2. Materials and Methods

2.1. Materials

The sample consisted of 120 kg of a blend of specialty coffees (Blend Gourmet oro verde with 50% Geisha, 25% Caturra, and 25% Pache) with organic and quality certification, from the 2024 harvest of the Cooperativa Agraria Cafetalera Cenfrocafe in the province of Jaen, department of Cajamarca, Peru. Green coffee beans were purchased and divided into three parts: green beans, roasted beans, and ground roasted beans. The initial moisture content of the green beans was 10.5%. The samples were packaged in tocuyo bags (100% polycotton material with a light weight of 120 g/cm²), double-laminated aluminum foil (aluminum weight 92 g/m² and permeability: 5.75 g/m²/day) and vacuum-packed in Ecotac bags for green beans; vacuum-packed in Ecotac bags (O² Transfer Rate (OTR) 0.13 cc/m²/24 hours, 20x30 cm) for roasted coffee beans, and in pressed cardboard boxes (made of micro-corrugated, self-assembling brown kraft material, 10x10x7 cm), bilaminate bag without valve with window and zipper (14x22 cm, white, standard zipper closure), bilaminate bag (Kraft

paper and aluminum) with degassing valve and zipper (14x22 cm, Kraft color, standard zipper closure, weight 145 g/m²), and in a trilaminate bag (Kraft paper, aluminum, polyethylene) with degassing valve and zipper (25x12.3 cm) for roasted ground coffee. Each container held 250 g of sample. However, the cup profile of the coffee samples and the validation of their specialty status during accelerated storage were determined using the cupping method carried out by six Q Arabica Graders [48] (Figure 1).

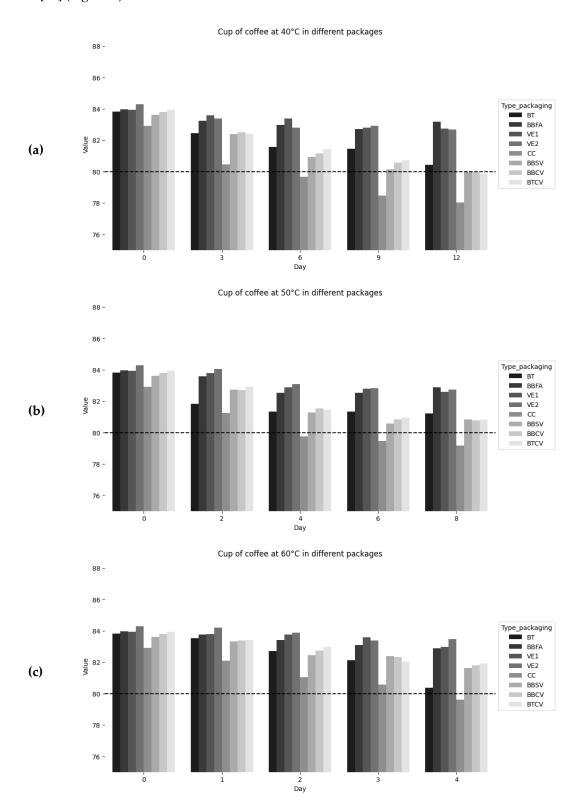


Figure 1. Cup scores for each presentation and packaging during accelerated storage at 40°C (a), 50°C (b), and 60°C (c). BT: tocuyo bag. BBFA: bilaminated aluminum foil bag. VE1: Ecotac vacuum bag with green beans. VE2:

Ecotac vacuum bag with roasted beans. CC: pressed cardboard box. BBSV: bilaminated bag without valve, with window and zipper. BBCV: bilaminated bag with degassing valve and zipper. BTCV: trilaminate bag with degassing valve and zipper.

2.2. Preparation of the Sample

A total of 120 kg of coffee was weighed using an electronic platform scale (T-Scale, TCS-K2, Taiwan), which was distributed into three parts: for green coffee beans, 45 units per 250 g package were used, distributed as follows: 15 tocuyo packages, 15 bilaminated foil packages, and 15 vacuumpacked Ecotec bags, which were packaged using a vacuum packer (Grondoy, EVE, China); to obtain roasted ground coffee, 15 kg of green coffee beans were placed in the roaster (Probat, P12-2, USA) at an initial temperature of 185 °C and a final temperature of 210 °C for 10.5 min under a medium roast. Once the process was complete, it was left to rest for 12 hours. For packaging, 15 Ecotec bags of 250 g were used with a vacuum packer (Grondoy, EVE, China). For the roasted ground coffee, the roasting was carried out as described above and then ground using an industrial grinder (Ditting, KFA1403, Switzerland). Fifteen kg of roasted coffee were placed in the hopper for 15 min with a grind setting of level 5, obtaining ground coffee. The roasted and ground coffee was then packaged in 60 250 g containers distributed in 15 pressed cardboard boxes, 15 bilaminate containers without a valve, with a window and zipper, 15 bilaminate containers with a degassing valve and zipper, and 15 trilaminate coffee containers with a degassing valve and zipper. The packaged samples were sealed using a sealing machine (Ditting, FR-900, Switzerland), with the exception of the pressed cardboard boxes and Ecotec vacuum bags.

2.3. Accelerated Storage

Once packaging was complete, each sample was coded and stored in three ovens (FAITHFUL, GX-45BE, China). Fifteen samples per container (120 containers) were placed in each oven at temperatures of 40, 50, and 60 °C over a period of 12 days for evaluation. Once the time for each temperature had elapsed, the samples were gradually removed from the ovens for roasting (in the case of green coffee bean samples) using a roaster (PROBAT-WERKE, BRZ, Germany). 110 g of green coffee was placed at an initial temperature of 185 °C and a final temperature of 190 °C for a period of 15 min. Before grinding (for green coffee samples and roasted coffee samples), the coffee was left to rest for 10 minutes and then ground using a grinder (Mahlkönig, BRZ2, Germany). 11.55 g of coffee was placed in the hopper and ground for 3 s, with the grind size set to level 6.

2.4. Total Polyphenol Content

The total polyphenol content of each extract and its replicates was determined using the Folin-Ciocalteu method following the methodology of [49] with some modifications. Gallic acid was used as the standard: 10 mg was dissolved in distilled water to a total volume of 25 mL, obtaining the stock solution. From this, the calibration curve was prepared in the range of 0–140 mg/L. For the extracts, 25 mg of ground coffee was weighed and mixed with 5 mL of ethanol/water (80:20 v/v). The mixture was stirred in an orbital shaker (Benchmark BT302) for 30 min at 3000 rpm and then centrifuged (PRO ANALYTICAL CR 4000) at 4000 rpm for 15 min. The supernatant was filtered using Whatman N° 42 paper. For analysis, 20 μ L of the extract was pipetted into microplates, followed by 800 μ L of distilled water, 40 μ L of 10% Folin-Ciocalteu reagent, and 160 μ L of 770 mM Na₂CO₃. Finally, the absorbance was measured at 765 nm using a Thermo ScientificTM Multiskan SkyHigh microplate reader with touchscreen.

2.5. Polyphenol Degradation Model

2.5.1. Chemical Kinetics

Equation (1) describes the main reaction of the polyphenol degradation model [50]:



Polyphenols + Oxygen
$$\stackrel{k}{\rightarrow}$$
 o-Quinone (1)

Where k is the pseudo-constant reaction rate (h^{-1}).

2.5.2. Determination of the Reaction Order

The reaction order of chemical kinetics (zero or one order) was determined for the loss of desirable compounds (Equations (2), (3), (4), and (5)) to represent the above reaction, according to the result defined by the highest value of the coefficient of determination (R²) of the regression constructed with the data obtained from the decay of total polyphenols over time, for each storage temperature and type of container.

Order 0:
$$\frac{d[c_{pt}]}{dt} = -k[C_{pt}]^n$$
 (2)

$$C_{pt} - C_{pt0} = -kt (3)$$

Order 1:
$$\frac{d[c_{pt}]}{dt} = -k[C_{pt}]^n$$
 (4)

$$\left[C_{pt}\right] = \left[C_{pt0}\right]e^{-kt} \tag{5}$$

Where C_{pt} and C_{pt0} are the total polyphenol content (mg/L) at a given time and at the start, respectively, t is time (s), and n is the reaction order (0 and 1).

Equations (3) and (5) were linearized, and graphs were plotted for $[C_{pt}]$ versus t (Equation (6)) and $ln[C_{pt}]$ versus t (Equation (7)). The rate constants k (1 or 2) were determined from the slopes of the linear graphs:

Order 0:
$$C_{pt} = C_{pt0} - k_1 t$$
 (6)

Order 1:
$$ln[C_{Pt}] = ln[C_{pt0}] - k_2t$$
 (7)

2.5.3. Calculation of Activation Energy (Ea) and Pre-Exponential Factor (Ao)

With the selected reaction order, the activation energy E_a (kJ/mol) and the pre-exponential factor A_0 (mg/L) were estimated using an Arrhenius-type fit via Equation (10), constructed with the slopes of the straight line graphs between $\ln(k)$ and (1/(T + 273.15)) for each temperature and packaging in the study, where R is the gas constant (8.31446 J/mol*K) and T is the constant air temperature in storage (°C).

$$k = A_0 e^{\left[-\left(\frac{E_a}{R}\right)\frac{1}{(T+273..15)}\right]}$$
(8)

$$ln[k] = ln \left[A_0 e^{\left[-\left(\frac{E_a}{R}\right) \frac{1}{(T+273..15)}\right]} \right]$$
(9)

$$ln[k] = ln[A_0] - \left[\left(\frac{E_a}{R} \right) \frac{1}{(T + 273..15)} \right]$$
(10)

2.5.4. Estimation of the Average Retention Time of Total Polyphenols

Using Equation (11), the half-life ($t_{1/2}$) of total polyphenol retention in specialty coffee samples was calculated, following a zero-order reaction.

Order 0:
$$t_{1/2} = \frac{c_{pt0} - c_{pt}}{k_1}$$
 (11)

2.6. Statistical Analysis

The experiments were performed in triplicate, and the results were expressed using descriptive statistics with means \pm standard deviations. In addition, data analysis was performed to determine whether there were significant differences in total polyphenol retention depending on the type of packaging, storage time, and temperature. For each storage temperature (40, 50, and 60 °C), a two-way analysis of variance (ANOVA) with replicates was applied, considering the type of packaging and storage time (days) as fixed factors. Subsequently, for multiple comparisons between the means of the different treatments (packages) at each measurement time, Tukey's honestly significant difference (HSD) test was used, with a confidence level of 95% (α = 0.05). The results of this test are presented in the results tables using letters as superscripts, where means within the same sampling day that share a letter are not significantly different. The analyses were performed using the statistical software R (version 4.3.0, R Core Team, 2023) and the software used was Python through Google Colaboratory.

3. Results and Discussion

3.1. Quantification of Total Polyphenols

Figure 2 shows the calibration curve for polyphenols with respect to absorbance, establishing the regression model y = 0.0038x + 0.0051 with an R^2 of 0.9781.

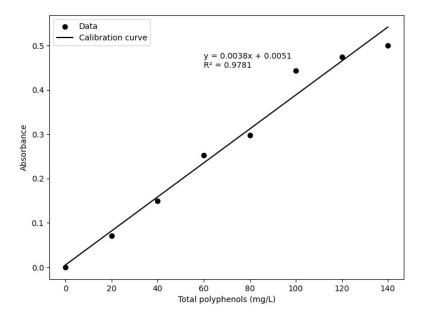


Figure 2. Calibration curve for total polyphenols.

Tables 1, 2, and 3 show the results of polyphenol degradation during accelerated storage of coffee in three states and packaged in different types of packaging.

Table 1. Total polyphenol values during storage at 40 °C.

	Total polyphenol content (mg/L)										
D		T 40°C / HR 65%									
Days		Green bean		Roasted bean		Ground roast		-			
	BT	BBFA	VE1	VE2	CC	BBSV	BBCV	BTCV			
0	113.25±0.02ª	117.61±0.01a	109.89±0.01a	106.11±0.00a	109.18±0.02a	113.31±0.09a	102.25±0.02a	107.87±0.01a			
3	109.89 ± 0.02^{a}	$104.71 {\pm} 0.00^{\rm b}$	107.78 ± 0.03^{a}	$105.15 {\pm} 0.01^{ab}$	$102.61 {\pm} 0.01^{c}$	$105.82 \!\pm\! 0.02^{\rm b}$	$101.90\pm0.00^{\circ}$	105.30 ± 0.03^{ab}			
6	106.20 ± 0.01^{a}	$104.01 \!\pm\! 0.01^a$	107.25 ± 0.02^a	$103.01 {\pm} 0.03^{ab}$	97.17±0.02d	100.15 ± 0.01^{c}	$101.20 {\pm} 0.05^{bc}$	104.89 ± 0.02^a			
9	100.50 ± 0.01^{b}	$102.75 {\pm} 0.02^a$	105.76 ± 0.01^a	101.64 ± 0.01^{b}	93.92±0.03d	97.78±0.01°	93.13±0.02d	104.10 ± 0.01^{a}			
12	98.31±0.01b	$91.03 \pm 0.02^{\circ}$	102.34 ± 0.02^a	81.03 ± 0.02^{e}	$92.63 \pm 0.02^{\circ}$	$92.69 \pm 0.02^{\circ}$	91.55±0.01°	101.29 ± 0.01^{a}			

BT: tocuyo bag. BBFA: bilaminated aluminum foil bag. VE1: Ecotac vacuum bag with green beans. VE2: Ecotac vacuum bag with roasted beans. CC: pressed cardboard box. BBSV: bilaminated bag without valve, with window and zipper. BBCV: bilaminated bag with degassing valve and zipper. BTCV: trilaminate bag with degassing valve and zipper. ±: Mean Standard Deviation for 6 repetitions. Different letters represent significant differences between treatments according to Tukey's test (p<0.05).

Table 2. Total polyphenol values during storage at 50 °C.

		Total polyphenol content (mg/L)									
Б		T 50°C / HR 65%									
Days	Green bean			Roasted bean		Ground roast		<u>.</u>			
	BT	BBFA	VE1	VE2	CC	BBSV	BBCV	BTCV			
0	113.25±0.02a	103.31±0.01c	107.78±0.03b	103.01±0.03c	100.15±0.01 ^d	109.27±0.01 ^b	102.43±0.04°	105.30±0.03°			
2	101.29±0.01°	$102.75 \!\pm\! 0.02^{bc}$	96.64±0.00e	$101.55 \pm 0.00^{\circ}$	96.29±0.02e	105.82 ± 0.02^a	$102.25 {\pm} 0.02^{bc}$	104.01 ± 0.03^{ab}			
4	$91.29\pm0.00^{\circ}$	98.57±0.02°	95.15±0.01d	94.97 ± 0.01 ^d	94.10±0.01d	91.82±0.02e	100.76 ± 0.02^{b}	102.25 ± 0.03^{a}			
6	88.66 ± 0.03 d	96.55±0.02b	93.57±0.01c	92.61±0.01c	92.63±0.02c	90.50 ± 0.03 ^d	93.39±0.00c	99.71±0.01a			
8	80.68 ± 0.02^{e}	91.99 ± 0.04^{b}	92.08±0.01 ^b	81.03±0.01e	91.55±0.01 ^b	86.82 ± 0.03^{d}	90.85±0.03°	92.78±0.03 ^b			

BT: tocuyo bag. BBFA: bilaminated aluminum foil bag. VE1: Ecotac vacuum bag with green beans. VE2: Ecotac vacuum bag with roasted beans. CC: pressed cardboard box. BBSV: bilaminated bag without valve, with window and zipper. BBCV: bilaminated bag with degassing valve and zipper. BTCV: trilaminate bag with degassing valve and zipper. ±: Mean Standard Deviation for 6 repetitions. Different letters represent significant differences between treatments according to Tukey's test (p<0.05).

Table 3. Total polyphenol values during storage at 60 °C.

			Т	otal polyphen	ol content (m	g/L)				
Davis		T 60°C / HR 65%								
Days		Green bean		Roasted bean		Ground roast				
	BT	BBFA	VE1	VE2	CC	BBSV	BBCV	BTCV		
0	113.25±0.02a	102.75 ± 0.02^{c}	107.78 ± 0.03^{b}	103.01±0.03c	99.01±0.03d	106.82±0.03b	102.69±0.03°	105.30±0.03bc		
1	106.20 ± 0.01^a	92.08 ± 0.04^{e}	95.24 ± 0.03 ^d	95.68 ± 0.01 ^d	$98.75 \pm 0.02^{\circ}$	105.82 ± 0.02^a	102.25 ± 0.02^{b}	94.54 ± 0.01 ^{de}		
2	101.38 ± 0.02^{a}	$84.45 \pm 0.00^{\text{f}}$	93.31±0.02c	86.73 ± 0.03^{ef}	$93.13 \!\pm\! 0.01^{cd}$	102.17 ± 0.04^{a}	$93.48 \pm 0.02^{\circ}$	83.31 ± 0.01^{f}		
3	81.55 ± 0.01 ^d	79.45 ± 0.03 ^d	89.54 ± 0.01 ^b	$83.57 \pm 0.05 c^d$	92.63 ± 0.02^{a}	91.38±0.01a	91.82 ± 0.06^{a}	79.01 ± 0.02 ^d		
4	$79.18 \pm 0.01^{\circ}$	71.99 ± 0.00^{d}	$76.46 \pm 0.01^{\circ}$	$76.82 \pm 0.00^{\circ}$	$79.97 \pm 0.00^{\circ}$	$75.59\pm0.00^{\circ}$	89.18 ± 0.04^{a}	76.55±0.01°		

BT: tocuyo bag. BBFA: bilaminated aluminum foil bag. VE1: Ecotac vacuum bag with green beans. VE2: Ecotac vacuum bag with roasted beans. CC: pressed cardboard box. BBSV: bilaminated bag without valve, with window and zipper. BBCV: bilaminated bag with degassing valve and zipper. BTCV: trilaminate bag with degassing valve and zipper. ±: Mean Standard Deviation for 6 repetitions. Different letters represent significant differences between treatments according to Tukey's test (p<0.05).

The results in Table 1 show that, as expected, there are no significant differences in polyphenol content on day 0 between the different types of packaging at the start of the experiment (all share the letter 'a'), but as time passes, the differences between packaging types become more evident. Green grain in BBFA packaging and vacuum packaging (VE1) generally showed greater polyphenol retention in the early days. Towards the end of storage (day 12), the trilaminate bag (BTCV) and

vacuum-packed green beans (VE1) were the packaging types that retained significantly more polyphenols, unlike the cardboard box (CC) and tocuyo bag (BT), which showed some of the greatest degradation.

The results in Table 2 show that the trilaminate bag (BTCV) is the packaging that preserves significantly more polyphenols over time (days 4, 6, and 8), followed by the bilaminate bag with aluminum foil (BBFA) for green beans and the bilaminate bag with valve (BBCV) for roasted ground coffee. On the other hand, the tocuyo bag (BT) and the vacuum-packed roasted beans (VE2) showed the worst results at the end of the period (day 8), being significantly different from the best packaging.

The results in Table 3 show that, at this extreme temperature, the degradation of polyphenols is very rapid and severe. The differences between packaging types are marked. The bilaminate bag with valve (BBCV) for roasted ground coffee showed significant superiority in polyphenol retention at the end of storage (day 4), being the only one that remained above 89 mg/L, while the bilaminated aluminum foil bag (BBFA) for green beans and the trilaminated bag (BTCV) showed some of the worst retention at this temperature, suggesting that their effectiveness may be compromised under extreme conditions.

Figure 3 shows descriptive statistics as a graphical summary of the decline in polyphenol content during accelerated storage for all coffee presentations, showing the temporal trends of polyphenols using a line graph for each temperature and evaluation day.

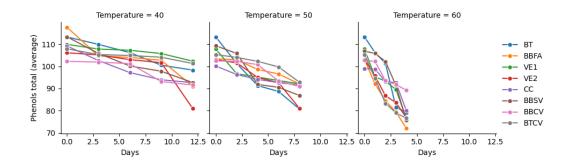


Figure 3. Temporal trends in total polyphenols in specialty coffee samples under accelerated storage conditions. BT: Tocuyo bag with green beans. BBFA: Bilaminated aluminum bag with green beans. VE1: Ecotac vacuum bag with green beans. VE2: Ecotac vacuum bag with roasted beans. CC: Pressed cardboard box with ground roasted coffee. BBSV: Bilaminated bag without valve, with window and zipper, with ground roasted coffee. BBCV: bilaminate bag with degassing valve and zipper with ground roasted coffee. BTCV: trilaminate bag with degassing valve and zipper with ground roasted coffee.

Furthermore, Figure 4 presents an overview of interactions using a heat map, in which it can be seen that areas with warm colors (reds and oranges) indicate the highest concentration of total polyphenols, while cool colors (yellows) show low values.

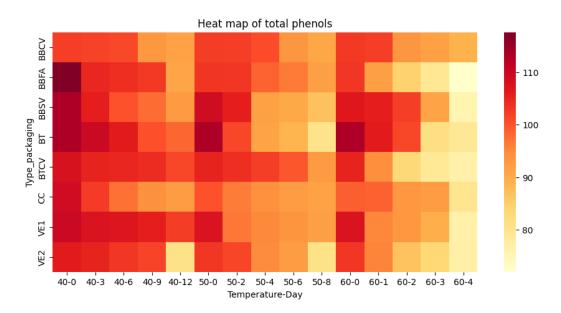


Figure 4. Heat map of total polyphenol degradation in specialty coffee during accelerated storage. BT: Tocuyo bag with green beans. BBFA: Bilaminated aluminum bag with green beans. VE1: Ecotac vacuum bag with green beans. VE2: Ecotac vacuum bag with roasted beans. CC: Pressed cardboard box with ground roasted coffee. BBSV: Bilaminated bag without valve, with window and zipper, with ground roasted coffee. BBCV: bilaminate bag with degassing valve and zipper with ground roasted coffee. BTCV: trilaminate bag with degassing valve and zipper with ground roasted coffee.

The total polyphenol content decreased significantly over time, affected by packaging material and storage conditions, particularly at high temperatures, with this decline being more pronounced in roasted beans and ground roasted beans compared to green beans. This phenomenon can be attributed to the thermal degradation of phenolic compounds during roasting and storage, thus affecting the antioxidant properties of coffee [13,40,47,51–57]. The Folin-Ciocalteu method, which was used to quantify polyphenols, has proven effective in determining the content of these compounds in various coffee preparations [58,59].

Table 1 shows the observed losses in polyphenol content at 40 °C. It is noteworthy that green coffee beans maintained a relatively high polyphenol content compared to the other processed formats, which is consistent with the literature suggesting that polyphenols are more abundant in unprocessed or minimally processed coffee [6,10,40]. This phenomenon may be related to the fact that green grain has less exposure to environmental factors such as light and air, which can accelerate the degradation of phenolic compounds [9].

At higher storage temperatures, as shown in Table 3 (50°C) and Table 4 (60°C), the polyphenol content decreases progressively in all types of packaging, although the impact varies depending on the type of packaging. Bilinamized bags, for example, act as effective barriers against the loss of important compounds, resulting in greater polyphenol retention compared to less efficient packaging, such as paper [5,46].

Table 4. Determination of the reaction order.

	Packaging		Temperature					
C 1 -		Orde	40 °C	50 °C	60 °C			
Sample		r	Regression (R ²)	Regression (R ²)	Regression (R ²)	Selection		
Green bean	ВТ	0	y = -1.309x + 113.48 (0.9858)	y = -3.8889x + 110.59 (0.9576)	y = -9.2778x + 114.87 (0.9341)	0		

_						. <u></u>	
			·· - 0.0124v + 4.722	y = -0.0406x +	y = -0.098x +		
		1	y = -0.0124x + 4.733 (0.9849)	4.7097	4.7533		
_			(0.9649)	(0.9706)	(0.9237)		
			1 927 11E 04	y = -1.4415x +	y = -7.4152x +		
		0	y = -1.837x + 115.04 (0.8546)	104.4	100.97		
	DDEA		(0.0540)	(0.9526)	(0.9835)	1	
	BBFA		0.010 . 4.7475	y = -0.0147x +	y = -0.0859x +	1	
		1	y = -0.018x + 4.7475	4.6494	4.6204		
			(0.8549)	(0.9486)	(0.9920)		
			0 F70·· + 110 02	y = -1.7237x +	y = -6.8333x +		
		0	y = -0.570x + 110.03	103.94	106.13		
	1751		(0.9303)	(0.763)	(0.9200)	0	
	VE1		0.005 . 4.5011	y = -0.0174x +	y = -0.0748x +	0	
		1	y = -0.005x + 4.7011	4.643	4.6704		
			(0.9261)	(0.7781)	(0.9119)		
			y = -1.7895x +	y = -2.6462x +	y = -6.4503x +		
		0	110.13	105.22	102.06		
Roasted	VE2		(0.6643)	(0.9109)	(0.9814)	0	
bean		1	y = -0.0191x +	y = -0.0286x +	y = -0.0722x +	0	
			4.7089	4.6609	4.6296		
			(0.6466)	(0.8929)	(0.9867)		
	CC		y = -1.3928x +	y = -1.0424x +	y = -4.4181x +		
		0	107.46	99.114	101.54		
			(0.9369)	(0.9356)	(0.8184)	1	
		1	y = -0.0139x +	y = -0.0109x +	y = -0.0491x +	1	
			4.6777	4.5964	4.6246		
-			(0.9457)	(0.9417)	(0.8003)		
		0	y = -1.6423x + 111.8	y = -3.0117x +	y = -7.6901x +		
			(0.9698)	108.89	111.73		
	BBSV		(0.5050)	(0.9044)	(0.8585)	1	
	۷ وطط		y = -0.016x + 4.7182	y = -0.0308x +	y = -0.0838x +	1	
		1	y = -0.010x + 4.7182 (0.9773)	4.6922	4.7276		
Ground			(0.5775)	(0.9108)	(0.8343)		
roast			y = -1.0058x +	y = -1.6009x +	y = -3.7456x +		
		0	104.04	104.34	103.38		
	BBCV		(0.8365)	(0.8713)	(0.9101)	0	
_	DDCV		y = -0.0104x +	y = -0.0165x +	y = -0.039x +	O	
		1	4.6461	4.6492	4.6394		
			(0.8353)	(0.8692)	(0.9151)		
			y = -0.4786x +	y = -1.4664x +	y = -7.3012x +		
		0	107.56	106.68	102.34		
	BTCV		(0.9192)	(0.8766)	(0.9264)	0	
	2101		y = -0.0046x +	y = -0.0148x +	y = -0.0817x +	Ŭ	
		1	4.6782	4.6713	4.6306		
			(0.9181)	(0.8651)	(0.9421)		

BT: tocuyo bag. BBFA: bilaminated aluminum foil bag. VE1: Ecotac vacuum bag with green beans. VE2: Ecotac vacuum bag with roasted beans. CC: pressed cardboard box. BBSV: bilaminated bag without valve, with window and zipper. BBCV: bilaminated bag with degassing valve and zipper. BTCV: trilaminate bag with degassing valve and zipper.

It is also interesting to note that packaging with degassing valves appears to provide additional protection against oxidation, facilitating better preservation of polyphenols in roasted ground coffee

[1,60]. The effectiveness of packaging could be associated with its oxygen permeability and moisture control, critical factors in the stability of phenolic content [5]. Vacuum packaging proved more efficient at retaining polyphenols in roasted coffee compared to other types of packaging such as aluminum foil bilaminate and trilaminate bags with degassing valves [61]. This suggests that oxygen permeability and the ability to limit exposure to moisture are key factors in the stability of bioactive compounds in packaged coffee [60,62].

In conclusion, differences in polyphenol content when using different packaging suggest a detailed study of the barrier properties of each material, such as limiting exposure to oxygen and heat, which could provide additional information for preserving the polyphenol content of coffee. Therefore, the choice of packaging and storage conditions can have a significant impact on coffee quality, as well as on the health benefits associated with its consumption.

3.2. Determination of the Reaction Order

Table 4 shows the comparison of the regressions and R² determined for the deterioration kinetics of polyphenols for zero and first order of the coffee samples under different storage conditions, which defined that the chemical reaction kinetics for the loss of phenolic compounds present in coffee exhibits a predominantly zero-order behavior.

The data obtained suggest that a zero-order model provides a better fit, indicating that the reaction of polyphenol loss under coffee storage conditions is constant over time, confirming that this phenomenon may be more related to physical factors such as temperature and exposure to oxygen than to the concentration of the polyphenols themselves [63–66]. These findings underscore the need for a consistent approach to coffee supply chain design, considering both bean quality and storage conditions to maximize the duration and effectiveness of its bioactive compounds and nutritional properties.

3.3. Determination of the Kinetic Constants of Deterioration (k)

Table 5 shows the Arrhenius equation determined for each coffee sample based on the values of the pre-exponential factor (A_0) and calculated activation energy (E_a), which can be used to predict the deterioration constants at different storage temperatures to which the different coffee presentations are subjected.

Table 5. E_a , A_0 and k values for the de	gradation of total	polyphenols in coffee i	or each sample and package.

Sample	Packaging	Ea (kJ/mol*K)	A_0	Arrhenius equation determined
	BT	85.085	2.06E+14	$k = 2.06E + 14e^{\left[-\left(\frac{85.085}{RT}\right)\right]}$
Green bean	BBFA	59.714	1.22E+10	$k = 1.22E + 10e^{\left[-\left(\frac{59.714}{RT}\right)\right]}$
	VE1	107.67	4.82E+17	$k = 4.82E + 17e^{\left[-\left(\frac{107.67}{RT}\right)\right]}$
Roasted bean	VE2	55.419	2.85E+09	$k = 2.85E + 09e^{\left[-\left(\frac{55.419}{RT}\right)\right]}$
	CC	49.321	1.75E+08	$k = 1.75E + 08e^{\left[-\left(\frac{49.321}{RT}\right)\right]}$
Ground	BBSV	66.850	2.16E+11	$k = 2.16E + 11e^{\left[-\left(\frac{66.850}{RT}\right)\right]}$
roast	BBCV	56.886	2.86E+09	$k = 2.86E + 09e^{\left[-\left(\frac{56.886}{RT}\right)\right]}$
	BTCV	118.04	2.09E+19	$k = 2.09E + 19e^{\left[-\left(\frac{118.04}{RT}\right)\right]}$

BT: tocuyo bag. BBFA: bilaminated aluminum foil bag. VE1: Ecotac vacuum bag with green beans. VE2: Ecotac vacuum bag with roasted beans. CC: pressed cardboard box. BBSV: bilaminated bag without valve, with window and zipper. BBCV: bilaminated bag with degassing valve and zipper. BTCV: trilaminate bag with degassing valve and zipper.

The total loss of polyphenols in coffee during storage is intrinsically related to several factors, including temperature, type of packaging, and the state of the bean (green, roasted, or ground). The

Arrhenius model, which relates the reaction rate to temperature through activation energy (E_a), is fundamental to understanding these phenomena. In particular, its application in assessing the stability of polyphenols at different temperatures is crucial for determining the viability of storage and the long-term quality of coffee [60,67]. For example, it has been reported that packaging that limits access to O_2 , such as vacuum packaging, tends to show lower E_a values, suggesting that reducing oxygen can inhibit the degradation of phenolic compounds, prolonging their effectiveness in terms of antioxidant properties [40,68]. Alternatively, packaging that allows gas exchange has shown greater loss of polyphenols at room temperature [69]. According to Arrhenius' theory, at higher temperatures, the E_a required for the loss of polyphenols tends to be lower, causing an acceleration in the degradation process.

Likewise, it was observed that optimized packaging, such as the Ecotac bag for green beans (E_a of 107.67 kJ/mol*K), has higher E_a values, suggesting greater resistance to polyphenol degradation compared to other packaging. This could be the result of the packaging's ability to limit the exposure of polyphenols to oxygen and moisture, two critical factors in the oxidation process [70,71]. On the other hand, the trilaminate bag packaging with degassing valve and zipper (BTCV) for roasted ground coffee showed an E_a value of 118.04 kJ/mol*K, which was the highest in the study. This result could indicate that the packaging design is particularly effective in retaining the quality of polyphenols, allowing for a gradual decrease in their oxidative degradation due to its sophisticated preservation methodology [72,73]. The high capacity of this packaging to minimize exposure to air and control O2 transfer is crucial for maintaining the stability of these compounds at higher temperatures [74]. In contrast, packaging with lower E_d values, such as pressed cardboard boxes with 49,321 kJ/mol*K for roasted and ground coffee, appear to be more susceptible to rapid polyphenol degradation. This phenomenon can be attributed to its design, which may allow more direct contact with O₂ and, therefore, an increase in the oxidation rate of polyphenols [75,76]. The permeability of cardboard can facilitate the diffusion of O2 and moisture into the packaging, accelerating the degradation process [77].

Furthermore, it can be seen that the order of E_a values among the different forms of coffee (green beans, roasted beans, and ground coffee) is also clear. Green beans, which are more susceptible to repeated oxidation processes under high temperature conditions, may undergo slow degradation in more preferred packaging that adequately protects the chemical integrity of polyphenols [78,79]. In general, differences in E_a values reflect not only the chemical nature of the phenolic compounds present, but also variations in the storage atmosphere generated by the type of packaging used [80,81].

3.4. Estimated Half-Life of Total Polyphenol Retention

Table 6 shows the calculated values of the kinetic constants of deterioration (k) and half-lives ($t_{1/2}$) (where half of the total polyphenol concentration is lost), following zero-order chemical kinetics, for coffee samples at the acceleration temperatures of the study (40, 50, and 60°C) and at the everyday storage temperatures of supermarkets and homes (15, 20, and 25°C).

Table 6. Half-life times $(t_{1/2})$ for the degradation of total polyphenols.

Campla	Paglaging C	torage temperature (°C)	le (days-1)	t _{1/2}
Sample	rackaging 5	torage temperature (°C)	k (day ^{s-1})	order zero (months)
		15	0.0790631	23.87
		20	0.1448005	13.04
	ВТ	25	0.2598701	7.26
Cusan haan	DI	30	1.3429945	1.41
Green bean		40	3.6884747	0.51
		60	9.5343346	0.20
	BBFA	15	0.1844677	10.63
	DDFA -	20	0.2820690	6.95

	_	25	0.4252168	4.61
		30	1.3465856	1.46
		40	2.7363711	0.72
		60	5.3289017	0.37
		15	0.0149565	122.45
		20	0.0321659	56.94
	VE1	25	0.0674234	27.16
	VE1	30	0.5388931	3.40
		40	1.9353609	0.95
		60	6.4372803	0.28
		15	0.2592307	6.82
		20	0.3844649	4.60
Roasted	MEO	25	0.5627154	3.14
bean	VE2	30	1.6402242	1.08
		40	3.1673386	0.56
		60	5.8794484	0.30
		15	0.2026485	8.98
		20	0.2877919	6.32
		25	0.4039308	4.50
	CC	30	1.0466383	1.74
		40	1.8799240	0.97
		60	3.2600315	0.56
		15	0.1668885	11.32
		20	0.2684727	7.03
	DDCI /	25	0.4250620	4.44
	BBSV	30	1.5448715	1.22
		40	3.4168621	0.55
Ground		60	7.2057031	0.26
roast		15	0.1413287	12.11
		20	0.2118019	8.08
	DD CL	25	0.3131404	5.47
	BBCV	30	0.9389597	1.82
		40	1.8450157	0.93
		60	3.4813704	0.49
		15	0.0085808	209.52
		20	0.0198655	90.50
	DECT.	25	0.0447145	40.21
	BTCV	30	0.4365327	4.12
		40	1.7730255	1.01
		60	6.6204522	0.27

BT: tocuyo bag. BBFA: bilaminated aluminum foil bag. VE1: Ecotac vacuum bag with green beans. VE2: Ecotac vacuum bag with roasted beans. CC: pressed cardboard box. BBSV: bilaminated bag without valve, with window and zipper. BBCV: bilaminated bag with degassing valve and zipper. BTCV: trilaminate bag with degassing valve and zipper.

The data in Table 6 show that green coffee in Ecotac bags (VE1) exhibits a $t_{1/2}$ of 122.45 months at 15 °C, as opposed to 0.20 months at 60 °C. This phenomenon of accelerated degradation can be attributed to high temperatures, which potentially promote undesirable chemical reactions that lead to the loss of bioactive compounds such as polyphenols. The literature supports this observation, suggesting that proper temperature management during storage is essential for preserving phenolic compounds in coffee [40,55,82]. Bilaminate aluminum foil packaging (BBFA) and tocuyo bags (BT) demonstrate different levels of effectiveness in terms of polyphenol retention. For example, green

coffee beans in BBFA packaging have a shelf life of 10.63 months at 15 °C, which is significantly less than green coffee stored in vacuum packaging. These findings underscore how packaging material can affect the exposure of products to light and oxygen, factors that are known to contribute to the oxidation of polyphenols [14,60]. In the case of roasted coffee, it can be seen that the $t_{1/2}$ values are considerably lower than those for green coffee. For example, coffee roasted in an Ecotac bag (VE2) has a $t_{1/2}$ of only 6.82 months at 15 °C. The reduction in stability can be attributed to the roasting process, which on the one hand improves the sensory profile of coffee but, on the other, can lead to the degradation of phenolic compounds such as chlorogenic acids, which are vital for the antioxidant properties of coffee [68,82,83]. According to the evaluation of $t_{1/2}$ at 60 °C, both roasted and green coffee beans exhibit extremely low polyphenol retention times. This indicates that, as the temperature increases, not only is faster oxidation promoted, but the formation of compounds is also favored which, although they may be more volatile and less desirable, can contribute to changes in the organoleptic characteristics of the beverage [6,39].

Analysis of the rate constant (k) provides further context for these data. It can be observed that k increases considerably with rising temperature, confirming a direct relationship between temperature and the rate of polyphenol degradation. This pattern is consistent with various studies that highlight that increased temperature accelerates the kinetics of chemical oxidation reactions, which are responsible for the decline in coffee quality [39,68,83].

4. Conclusions

The evaluation of the kinetics of polyphenol deterioration in different forms of specialty coffee and its correlation with various packaging systems has provided fundamental information on the stability and preservation of these bioactive compounds under accelerated storage conditions. The polyphenol loss reaction fits a zero-order kinetic model, indicating that the decay of bioactive compounds under specialty coffee storage conditions is constant over time, suggesting that this phenomenon may be more related to external physical factors such as temperature and oxygen exposure than to the concentration of the polyphenols themselves. The degradation rate constants found reflect an inverse relationship between temperature and shelf life. The data indicate that packaging such as aluminum trilaminate bags and vacuum bags proved effective in controlled environments, while cardboard containers showed lower polyphenol retention. Therefore, this study highlights the complexity of the degradation kinetics of polyphenols in coffee and the critical influence of storage and packaging systems. These findings are highly relevant to the coffee industry, suggesting that the appropriate selection of packaging and optimal storage conditions can significantly extend the shelf life of polyphenols, thus contributing to the preservation of coffee quality in the market.

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