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*Review*

# The Role of inflammation in the Cellular and Molecular Mechanisms of Cardiopulmonary Complications of Sickle Cell Disease

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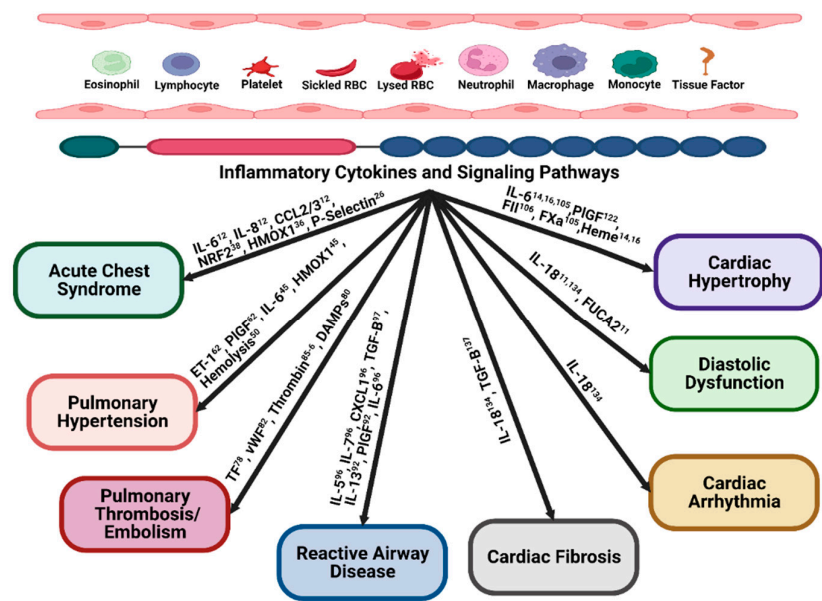
**Abstract:** Cardiopulmonary complications remain the major cause of mortality despite newer therapies and improvements in lifespan of patients with sickle cell disease (SCD). Inflammation has been identified as a major risk modifier in the pathogenesis of SCD associated cardiopulmonary complications in recent mechanistic and observational studies. In this review, we discuss recent cellular and molecular mechanisms of cardiopulmonary complications in SCD and summarize the most recent evidence from clinical and laboratory studies. We emphasize the role of inflammation in the onset and progression of these complications to better understand the underlying pathobiological processes. We also discuss future basic and translational research in addressing questions about the complex role of inflammation in the development of SCD cardiopulmonary complications, which may lead to promising therapies and reduce morbidity and mortality in this vulnerable population.

**Keywords:** sickle cell disease; cardiopulmonary complications; inflammation; acute chest syndrome; cardiac hypertrophy; cardiac fibrosis; diastolic dysfunction; pulmonary hypertension

## Introduction

Sickle cell disease (SCD) is the most common monogenic blood disorder, affecting approximately 100,000 Americans and millions more worldwide [1,2]. Cardiopulmonary complications are a major cause of morbidity and mortality in SCD, accounting for 32-70% of deaths [3–5]. Several pathophysiological processes, including anemia, hemolysis, endothelial dysfunction, and ventricular remodeling, may contribute to cardiopulmonary complications in SCD [3–5]. Although the etiology of cardiopulmonary complications in SCD is somewhat different from that in the general population, there are similarities in the cellular and molecular mechanisms that underlie the pathogenesis in both scenario, and that are beginning to gain prominence. Accumulating evidence has long identified chronic low-grade inflammation as a risk factor for the progression of myocardial infarction, ventricular hypertrophy, cardiac fibrosis, diastolic dysfunction, and pulmonary hypertension in the general population [6–10]. Recent mechanistic and observational studies on cardiopulmonary complications of SCD implicate inflammation as a major player in the onset and progression of cardiopulmonary complications in SCD [11–16]. These include several studies in animals and humans on the development of acute chest syndrome (ACS), cardiac hypertrophy, pulmonary hypertension (PH), cardiac fibrosis, and diastolic dysfunction [11–15]. These studies consistently suggested inflammatory pathways as a vital unifying mechanism that accompanies the structural and functional changes that occur at the onset and progression of these complications. Thus, integrating therapies that balance the pro-inflammatory and anti-inflammatory processes contributing to the chronic inflammatory state in SCD may provide opportunities for novel therapies that could be easily incorporated into the existing treatment options available to SCD patients. The recent advances in cellular and molecular mechanisms of cardiopulmonary complications of SCD, along with the complex interplay between inflammation and the unique

cardiac pathology of SCD such as acute chest syndrome, pulmonary hypertension, diastolic dysfunction, and cardiac hypertrophy, are described in this review (Figure 1).



**Figure 1.** A visual representation of the inflammatory mediators that may be involved in the cardiopulmonary complications of sickle cell disease. Interleukin 6 (IL-6), IL-5, IL-7, IL-18, IL-13, Tissue factor (TF), C-C motif chemokine ligand 2 (CCL2/3), Reactive oxygen species (ROS), nuclear-factor erythroid 2 like 2 (NRF2), heme-oxygenase 1 (HMOX1), Endothelin-1 (ET-1), Placental growth factor (PIGF), von Willebrand factor (VWF), Danger associated molecular patterns (DAMPs), Chemokine (C-X-C motif) ligand 1 (CXCL1), Factor II (FII), Factor Xa (Fxa) and Transforming growth factor (TGF-β).

**Inflammation and acute chest syndrome.**

Acute chest syndrome (ACS) is a pulmonary complication of SCD and the second leading cause of mortality and morbidity in both adults and children with SCD [17]. It is defined as the presence of fever and/or new respiratory symptoms such as cough, chest pain and presence of a new pulmonary infiltrate on chest X-ray [17]. Risk factors for ACS include younger age, severe SCD genotypes (SS or Sβ<sup>0</sup> thalassemia), lower fetal hemoglobin concentrations, inflammation, higher steady-state white blood cell counts, history of asthma, and tobacco smoke exposure [18,19]. The major causes known to trigger ACS include respiratory infection, pulmonary infarction or fat embolism, however no specific cause can be found in up to 30% of cases [18]. At a cellular level, an inciting trigger such as an infection permits increased adhesion of leukocytes (neutrophils) to the lung microvasculature, generation of cytokines, coupled with interactions with other cellular components such as platelets. This results in local hypoxemia and changes in rheology of the red blood cells (RBCs). This further facilitates interactions between RBCs, vascular endothelium and leukocytes resulting in increased oxidative stress, vaso-occlusion and tissue hypoxia. These events in turn result in additional recruitment of leukocytes and other cellular components to site thus amplifying the inflammatory cascade, resulting in a “vicious” cycle of lung injury and hypoxemia [20,21].

Evidence for heightened inflammation in the pulmonary microenvironment, during ACS comes from human studies which show that children with ACS have high levels of IL-6, IL-8, CCL2, and CCL3 in their sputum [12]. These cytokines, particularly CCL2 and CCL3, have been shown to recruit leukocytes, particularly neutrophils, via upregulation of platelet activating factor (PAF) and leukotriene B-4 (LTB4). The neutrophils firmly adhere to the endothelium and become activated as

assessed by shedding of CD62L and upregulation of CD11b [22]. Upregulation of CD11b in arrested leukocytes enables their interaction with GPIIb/IIIa expressed on platelets [23]. Arrested neutrophils can also interact with platelets via PSGL-1 on neutrophils binding to P-selectin on platelets. This is evidenced by autopsy studies which show presence of large neutrophil-platelet aggregates and platelet laden aggregates in pulmonary vasculature in patients with ACS [20,24]. Indeed, preclinical studies that inhibit P-selectin and GPIIb/IIIa interactions show fewer leukocyte-platelet aggregates [25] highlighting the importance of neutrophil and platelet heterotypic interactions in pathogenesis of ACS. Furthermore, a study by Ghosh et al., in Townes sickle cell mouse model showed that P-selectin in both platelet and endothelium compartments played a dominant role in promoting heme-induced ACS in SCD [26].

Hemolysis is a pathological feature of SCD that releases free hemoglobin and heme into the circulation due to RBC sickling and lysis, leading to the activation of inflammatory signaling pathways and vascular inflammation [27–30]. The release of free heme and cell-free hemoglobin also results in activation of neutrophils and generation of neutrophil extracellular traps (NETs) [31], iron-based generation of reactive oxidative species (ROS) with subsequent oxidization of membrane lipids [32], depletion of nitric oxide [33], and endothelial cytoskeleton remodeling resulting in barrier dysfunction [34]. Furthermore, plasma free heme and other markers of hemolysis have been associated with increased odds of developing ACS in children with SCD [35]. Additionally, a mutation in the heme-oxygenase 1 (HMOX1) short (GT)<sub>n</sub> repeat promoter that confers stronger inducibility of HMOX-1, the rate-limiting enzyme that degrades heme, was associated with a reduction in the rate of hospitalization for ACS in children with SCD [36]. These studies were validated in both the Townes and Berkeley SCD mouse models using extracellular heme as a trigger for ACS. Heme exposure causes respiratory failure due to rapid hypoxemia and death, mimicking some of the events associated with ACS in SCD patients [37]. Treatment of SCD mice with D3T (3H-1,2-dithiole-3-thione), an activator of nuclear-factor erythroid 2 like 2 (NRF2), which controls HMOX1 expression, reduced lethality in a model of heme-induced ACS in SS mice [38]. Additionally, treatment with hemopexin, the plasma heme binding protein, abrogates lung injury and mortality in Chlorine (Cl<sub>2</sub>) inhalation model of inducing ACS [39]. These studies suggest that therapies that target the product (heme) or molecular consequence(s) of hemolytic pathways may offer protection from ACS in SCD.

### Inflammation and pulmonary hypertension

Pulmonary hypertension (PH) is an independent risk factor for early death in SCD patients [40]. Its estimated prevalence, as assessed by right heart catheterization (RHC), is about 6-10% [41], although this estimate relied on an older definition used to diagnose PH. Per the most recent guidelines, PH is now defined as mean pulmonary artery pressure of >20 mm Hg in conjunction with pulmonary artery wedge pressure of ≤15 mm Hg and a pulmonary vascular resistance (PVR) of ≥3 Wood units (WU). A diagnosis of isolated postcapillary PH is made when PVR is <3 WU, whereas a PVR of ≥3 WU is supportive of pre-capillary PH [42]. In SCD, precapillary, post capillary and pulmonary thromboembolic PH or a combination can exist. Risk factors for PH include chronic intravascular hemolysis, pulmonary thrombosis or embolism, and heart failure [43,44].

The development of PH in SCD is complex and involves pulmonary vascular endothelial dysfunction, smooth muscle cell (SMC) proliferation and resistance to nitric oxide (NO) adventitial fibroblast accumulation, and inflammation. Interestingly, one of the unique features of PH in SCD is the presence of iron in pulmonary macrophages, a feature that is not seen in other forms of PH. An autopsy study of lung samples from SCD patients with PH and RV failure found peripheral monocytes and macrophages accumulating in the perivascular and alveolar regions of the lungs [45]. These macrophages had extensive iron accumulation concomitantly with the expression of HMOX1, ET-1, and IL-6 [45]. This suggests that, in pathological diseases with hemolysis such as SCD, circulating immune cells may be recruited into the lungs for heme degradation. However, this immune response may become maladaptive over time, as accumulated iron may contribute to oxidative stress, alter the redox balance, or induce transdifferentiation of resident lung macrophages

and other alveolar cells. This underscores an important role for intravascular hemolysis in the pathogenesis of PH [46–49]. SCD is characterized by increased stress erythropoiesis as a compensatory mechanism for anemia, which increases the number of reticulocytes and younger RBCs in circulation. During hemolysis, these young RBCs released a large amount of arginase into the plasma [50]. This plasma arginase consumes plasma L-arginine, the substrate required for NO production by endothelial cells, and, in conjunction with the consumption of endothelial NO by cell-free plasma Hb, reduces NO bioavailability [51,52]. The depletion of NO affects intracellular calcium signaling that leads to dephosphorylation of myosin protein preventing smooth muscle relaxation [53]. The depletion of NO also results in leukocyte recruitment via increased surface expression of leukocyte adhesion proteins such as E-selectin, VCAM and ICAM-1 [54,55] and results in smooth muscle proliferation and vascular remodeling [56]. Heme related generation of ROS decreases availability and or activation of soluble guanylyl cyclase or its regulators such as cytochrome b5 reductase 3 (CYB5R3) which can result in poor vasodilation of pulmonary vasculature increasing risk of pulmonary hypertension [57,58].

Cell free plasma hemoglobin and heme can also independently activate platelets and neutrophils via TLR4 signaling mechanism [31] resulting in further inflammation. In addition, cell free hemoglobin also generates ROS which furthers endothelial dysfunction and activates the coagulation system [59]. Chronic hemolysis also promotes transition of pulmonary endothelial cells to a mesenchymal or smooth muscle cell type and contributes to vascular remodeling [60]. Thus, heme exposure results in pathological endothelial activation, increased recruitment of leukocytes and depletion of protective mechanisms that preserve vascular integrity.

Mechanistically, endothelial dysfunction results in increased production of vasoconstrictors such as endothelin-1 (ET-1). Heme related endothelial dysfunction can deplete peroxisome proliferator-activated receptor  $\gamma$  (PPAR $\gamma$ ), which plays an active role in suppressing ET-1 production by regulating the level of microRNAs (miRs) such as miR-98 [55]. Lower levels of miR-98 associates with increased ET-1 production and endothelial dysfunction [52]. Exposure to heme also results in increased production of placenta growth factor (PlGF) by erythroid cells via the erythroid Krüppel-like factor (EKLf) [61] and NRF2-antioxidant response signaling [62]. PlGF is an angiogenic factor that activates endothelial cells to secrete ET-1 [63]. In an elegant study, overexpression of erythroid-specific PlGF in normal mice up to the levels seen in sickle cell mice resulted in an increase in the production of ET-1, which correlated with increased right ventricular pressure and pulmonary arteriolar thickening [64]. Elevated ET-1 and PlGF levels also correlate with severity of PH in patients with SCD [64]. PlGF was shown to activate expression of hypoxia-inducible factor 1 $\alpha$  (HIF-1 $\alpha$ ), independently of hypoxia, which in turn can stimulate expression of ET-1, which is involved with the development and severity of PH in SCD [63].

Indeed, these cellular and molecular mechanisms have informed the current therapeutics usually used in patients with pulmonary hypertension such as endothelin receptor (ETR) antagonists (Bosentan, Ambrisentan), those which prevents the degradation of cyclic guanosine monophosphate (cGMP) (Riociguat and Sildenafil), vasodilators (Epoprostenol) and anticoagulant (warfarin) among others. Clinical trials using hemopexin, a scavenger molecule that removes heme from circulation are underway in humans and have shown promising results in murine models [65]. Unfortunately, trials with ETR antagonists [66], Riociguat [67], and Sildenafil [68,69] were either limited by small sample size or adverse side effects respectively underscoring the need to better understand the pathology and larger clinical trials researching PH in SCD.

### **Inflammation and pulmonary thrombosis/ embolism**

Accumulating evidence from human studies discussed below suggests that inflammation is a risk factor for thrombosis. It is therefore not surprising that a retrospective study found that prevalence rates of venous thromboembolism (VTE) in adults with SCD was 25% and is associated with increased rates of recurrence and mortality [70,71]. Interestingly, the risk of pulmonary embolism (PE) is higher than risk of deep vein thrombosis (DVT) [71–73] suggesting that thrombosis may occur more ‘in situ’ in pulmonary vasculature of individuals with SCD. Risk factors for VTE

include elevated leukocyte count [74], severe phenotype as defined by > 3 hospitalizations annually for vaso-occlusive crisis, presence of SCD variant genotypes, elevated tricuspid regurgitation jet velocity (TRJV)  $\geq 2.5$  m/s [70], elevated body mass index, and prior splenectomy [70,75,76]. Even in those with lower hospitalizations, the cumulative incidence rate of VTE was at 6.8% compared to 1.6% in individuals who had similar number of hospitalizations for asthma exacerbation [71] suggesting that intrinsic pathology in addition to external risk factors play a role in development of PE and/or DVT.

Several cellular and molecular pathways are perturbed in SCD that leads to a prothrombotic state. Chronic hemolysis results in release of intracellular molecules known as danger (or damage)-associated molecular patterns (DAMPs) [77]. For example, one of the most studied DAMPs or alarmins, high-mobility group box 1 (HMGB1), is significantly elevated in the plasma of SCD patients and mice at baseline compared to controls [78,79]. VOC episodes further increased HMGB1 levels in SCD patients, or acute sickling induced following hypoxia-reoxygenation in mice [78]. Furthermore, treating the TLR4 reporter cell line with plasma from SCD patients increased TLR4 receptor activity, suggesting that HMGB1 contributes to TLR4 signaling in SCD [78]. Elevated circulating HMGB1 is associated with platelet nucleotide-binding domain, leucine-rich-containing family, pyrin domain-containing-3 (NLRP3) activation, which is mediated through the TLR4 and Bruton tyrosine kinase signaling pathways [79,80]. DAMPs have also been implicated in endothelial dysfunction [81], activation and recruitment of leukocytes [31,77] and inflammation [77] which shift the balance to a more prothrombotic state in SCD. The characteristic changes in RBC rheology contributes to the formation of venous thrombi that have denser fibrin network and a friable thrombus [82]. In addition, red cell derived microparticles contribute to thrombin generation via activation of Factor XI. Indeed, red cell derived microparticles associate with increased markers of coagulation activation [83]. Activated platelets promote inflammasome activation and generation of EVs which can lead to formation of heterotypic aggregate and occlusion of the lung's microvasculature [84]. DAMPs can also activate neutrophils and monocytes which can result in increased TF expression [85], NET formation, endothelial dysfunction and more inflammation [31] which have been linked to thrombus generation and propagation in non-SCD models [86]. Endothelial dysfunction from heme exposure results in the upregulation of adhesion molecules that attract neutrophils and platelets [31,34,81], and increased expression of TF and VWF which can contribute to pulmonary thrombosis [87].

The exact molecular mechanisms resulting in thrombus formation in lungs in SCD are not well studied and may involve mechanisms that either increases procoagulant proteins (such as TF, VWF, thrombin) [87,88], decreases anticoagulant proteins (like low protein C and S) [89,90], and/or decreases fibrinolysis [82]. There is some evidence that abrogation of TF using anti TF antibody reduces thrombus formation in sickle cell mouse model suggesting an important contribution of TF to thrombus generation in SCD. In addition, genetic or immunologic interventions that modulated expression of protein C and thrombin also diminished thrombus formation [91]. Data supporting the role of contact pathway leading to thrombosis in SCD is very sparse, however potential plausible sources include neutrophil nuclear content specifically nuclear DNA and histones which can initiate coagulation by activating Factor XII but also amplify thrombin-dependent factor XI activation [92], [86]. Partial support for this comes from observation that inducing neutropenia results in decreased thrombosis burden in an arterial thrombosis model [91]. Thus, several cellular and molecular mechanisms may be at play in pathogenesis of thrombosis.

### **Inflammation and reactive airway disease or airway hyper-activity (AHR)**

Reactive airway disease or AHR is common among adults and children with SCD [93,94] and can occur independent of clinical asthma. Studies show that up to 77% of children demonstrate a positive methacholine challenge test (MCT) [93,95]. Interestingly, the severity of AHR correlates with high LDH, suggestive of a critical role played by hemolysis and disease severity [96]. Indeed, one study did show that AHR was more common in those with HbSS genotype and was predictive of increased risk of ACS and vaso-occlusive crisis [97]. From a pathophysiology perspective, AHR is characterized by bronchial hypersensitivity to stimuli, airway and lung inflammation, abnormal

leukocyte recruitment, and airway and vessel wall thickening [98–100]. Chronic hemolysis and their byproducts may drive systemic inflammation and result in increased lung/airway inflammation. Indeed, in one study with SCD mice, even prior to sensitization with allergen, there was evidence of increased airway inflammation, increased lymphocytes in bronchoalveolar fluid (BAL), granulocyte-colony stimulating factor, interleukin 5 (IL-5), IL-7, and chemokine (C-X-C motif) ligand (CXCL)1 and lung T cell infiltration [101]. Mice exposed to specific allergen recapitulated specific features of asthma like phenotype including increased immunoglobulin E (IgE), IL-6, and IL-13 in serum and increased bronchial hyperresponsiveness to methacholine [101]. Another study corroborated the findings of increased IgE, and airway inflammation as evidenced by eosinophil infiltration, vessel wall thickening and increased concentrations of transforming growth factor beta (TGF- $\beta$ ) [102]. SCD mice with PIGF deficiency show decreased airway inflammation, leukotriene and IL-13 mediated immune responses suggesting an important role of PIGF signaling pathways in AHR [97]. Thus, multiple pathways are at work that makes individuals with SCD susceptible to allergens and environment pollutants.

### **Inflammatory mediators and cardiac hypertrophy**

Both concentric and eccentric hypertrophy had been reported in children and adults with SCD [103]. These changes in cardiac structure and function start in early childhood and worsen with age [104,105]. The enlargement of the heart begins as a compensatory myocardial remodeling in response to anemia [106]. The cardiomyocytes and capillary networks in the heart become enlarged to increase oxygen supply, leading to increase cardiac output at rest [106]. This elevated cardiac output also becomes exaggerated during exercise due to an increase in cardiac stroke volume in response to the increased oxygen consumption, indicating altered hemodynamics in SCD patients [106,107]. Restrictive cardiomyopathy can also co-exist with anemia-induced elevated cardiac output [104,105]. Morphological abnormalities of sickle RBCs such as polymerization and aberrant membrane transport properties, auto-oxidative ROS generation and ischemia-reperfusion injury, may also contribute to cardiac remodeling [108,109].

Studies have suggested that endothelial dysfunction and increased plasma markers of inflammation contribute to cardiac hypertrophy in SCD mouse models [110,111]. This may be due to the pre-activation of immune cells, including monocytes and endothelial cells in SCD [112–115]. There is also an increase in the mRNA and protein expression of pro-inflammatory markers such as TNF- $\alpha$ , IL-1, IL-6, MIP-1b and soluble endothelial adhesion molecules [114,116,117]. The heart, like the other organs is exposed to inflammatory insults from these circulating proinflammatory cytokines. Given that elevated systemic inflammation is associated with cardiac abnormalities in the general population [6], it is possible that the observed excessive systemic inflammation in individuals with SCD, could contribute to cardiac pathology in SCD.

In addition to pro-inflammatory cytokines, products of hemolysis such as heme and other DAMPs may further perturb the homeostatic state in the heart, thereby perpetuating the vicious cycle of inflammation and cardiac pathobiology described in the last paragraph. For instance, heme released into the circulation during hemolysis triggers several inflammatory pathways in SCD that contribute to organ damage [30,37,81]. In fact, a comparison of organ-specific expression patterns of HMOX1 in SCD mice treated with heme revealed that the heart has one of the highest expressions of HMOX1 [118]. This suggests that cardiac cells can uptake circulating heme and metabolize it. The potential problem with this process is that excess iron produced from heme breakdown and deposited in the heart can activate oxidative and apoptotic pathways. A recent study showed that heme-induced upregulation of HMOX1 promotes cardiac ferroptosis in SCD mice as well as the expression of cardiac hypertrophy genes [14], although T2\* cardiac magnetic resonance imaging measurement of cardiac iron showed that iron overload is rare even in chronically transfused SCD patients [119,120]. Another study showed that increasing circulating heme significantly elevated plasma IL-6 and the expression of cardiac hypertrophy markers in Townes sickle cell mice [16]. These studies underscore the importance of hemolysis in the pathogenesis of cardiac hypertrophy. Another way that inflammation may contribute to cardiac pathology is through a complex interaction with

the coagulation system. Increasing evidence in SCD has shown a link between vascular inflammation and hypercoagulation via activated intrinsic and extrinsic coagulation pathways, which may contribute to organ pathology [121,122]. A study by Sparkenbaugh et al. showed that short-term pharmacological inhibition of FXa in Berkeley sickle cell mice attenuated plasma IL-6 and cardiac hypertrophy [110]. Similarly, genetic inhibition of circulating FII in SCD mice improved right ventricle hypertrophy and dilatation, suggesting that increased thrombin generation or activity in SCD is a significant contributor to cardiac pathophysiology [111]. These studies suggest that perturbation in the coagulation system and the attendant vascular damage in SCD may contribute to inflammation, a key component of various mechanisms involved in cardiac dysfunction.

The inflammatory signaling pathway may also modify cardiac remodeling in SCD via the complex biological role played by PlGF. PlGF is an angiogenic cytokine that plays a role in the survival of endothelial cells and monocytes, and in cardiovascular health [115,123]. PlGF is crucial in the early inflammatory response needed for adaptive hypertrophic cardiac remodeling due to pressure overload [124,125]. Although studies have shown both beneficial and deleterious roles of PlGF in the heart [126–128], its expression is elevated in the plasma of SCD patients and linked with disease severity [64,114]. Furthermore, PlGF mRNA and protein expression were found to be elevated in the hearts of sickle cell mice at baseline and upon exposure to heme [129]. The role of PlGF in cardiopulmonary complications of SCD may be via an indirect effect on endothelial cells, fibroblasts, and monocytes in the heart, which are already primed for an exaggerated response due to a pro-inflammatory microenvironment mediated by cytokines such as IL-6. Nevertheless, no mechanistic studies have clearly delineated this intriguing role of PlGF in cardiac pathology in SCD by examining these cardiac cells individually.

### **Inflammation and diastolic dysfunction**

Diastolic dysfunction occurs in both children and adults with SCD. It has been associated with anemia, older age, higher creatinine levels, exercise impairment, increased LV mass, low sleep or waking oxygen saturation, and diffuse myocardial fibrosis [11,103,130–132]. Though the causative sequence is not well defined, diastolic dysfunction is also an independent risk factor for death in SCD patients [104,133]. Increased doppler echocardiography ratio of mitral valve inflow (E) velocities (E) over peak early diastolic annular velocity (E') > 8.2 has been shown to predict diastolic dysfunction in SCD patients [11,130,134]. Furthermore, diastolic dysfunction has been linked to the overexpression of interleukin-18 (IL-18), -L-fucosidase A2 (FUCA2), and thyroid hormone transporter (SLC16A2) in SCD patients' peripheral blood mononuclear cells (PBMCs) [11]. This finding was validated in mouse models, with results showing elevated expression of these genes in the myocardium of sickle cell mice compared to controls [11]. Although diastolic dysfunction in SCD involves multiple complex pathophysiological mechanisms, this report on IL-18 suggests the involvement of inflammation, which is chronic in SCD, either as a primary mechanism or as a secondary to cardiac remodeling due to the hyperdynamic state.

### **Inflammation and cardiac arrhythmia**

Cardiac arrhythmia (CA) is defined as a disruption in the normal activation of the heart or an irregular heartbeat rhythm that is either too slow (60 beats per minute) or too fast (>100 beats per minute) [135]. Some forms of CA, including sinus arrhythmia, are considered to be benign; however, the presence of structural heart defects such as LV dysfunction or genetic arrhythmia syndromes, including long or short QT syndrome, increases the severity of CA and could result in heart failure or sudden cardiac death [135,136]. Cardiac arrhythmias have been reported as the cause of death in 7.4% of in-hospital deaths in adult African Americans with sickle cell trait [137], and in 14% of SCD patients [138]. Cardiac arrhythmias have also been implicated in some sudden deaths recorded in SCD patients with a prolonged corrected QT interval (QTc), which is independently associated with an increased risk of death [107,139,140]. In a recent study, elevated expression of IL-18 in PBMCs of SCD patients was associated with longer QTc intervals and increased mortality risk [141]. Consequently, chronic inhibition of IL-18 binding protein in a sickle cell mouse model attenuated IL-

18-mediated ventricular tachycardia and improved diastolic function, suggesting a link between cardiac inflammation and arrhythmias, in SCD [141].

### Inflammation and cardiac fibrosis

Autopsy studies and studies in living SCD patients have shown the presence of both diffuse and transmural fibrosis [103,142,143]. The cellular and molecular mechanisms underlying cardiac fibrosis in SCD, are not completely understood. However, gene-expression profiles of heart tissue isolated from Berkeley sickle cell mice revealed elevated expression of genes involved in oxidative stress, angiogenesis, and TGF- $\beta$  signaling, which correlated with imaging and histology data demonstrating diffuse cardiac fibrosis and diastolic dysfunction [144]. Another study in SCD mice showed that sustained neutralization of the IL-18 binding protein ameliorated cardiac fibrosis [141] (Figure 2). In a small observational study of SCD patients, early initiation of disease-modifying therapy such as hydroxyurea and chronic transfusion was shown to prevent diffuse myocardial fibrosis and diastolic dysfunction [145]. Although the underlying specific mechanism(s) of how these therapies ameliorate the development of cardiac fibrosis remains unknown, there is a need for detailed mechanistic studies that elaborate on the link between inflammation and other pathways that may be involved in the development of cardiac fibrosis in SCD.

**Figure 2.** Summary of the inflammatory mechanisms associated with cardiac fibrosis and diastolic dysfunction in animals with SCD (with the relevant reports that describe them) addressed in the current review manuscript. TGF- $\beta$ : Transforming growth factor beta, IL-18: Interleukin-18, and FUCA2: -L-fucosidase A2. .

### Conclusions and future perspectives

Inflammation involves complex cellular and molecular pathways that have both beneficial and harmful effects, and it plays a major role in the pathogenesis of cardiopulmonary complications in SCD. Inflammation is also a common denominator in the development of ACS, cardiac fibrosis, PH, and diastolic dysfunction, as reviewed here (Table 1). Several studies have shown that chronic systemic inflammation as present in SCD contributes to a cascade of events that results in tissue injury, the generation of ROS, and endothelial dysfunction [146–148]. Additionally, inflammatory cells such as monocytes are in a state of pre-activation due to elevated levels of circulating inflammatory cytokines, which contribute to the chronic inflammation in SCD by amplifying the production and signaling of these cytokines [64,114,147]. Therefore, addressing cardiopulmonary manifestations of the disease by modulating inflammatory pathways using targeted therapies could offer a novel approach to reducing the risk of developing these complications. This would involve well-designed and detailed experiments that include investigating the role of genetic polymorphisms in the regulation of blood levels of circulating cytokines, inflammatory markers, and their signaling pathways in SCD. Additional investigation will also examine how genes encoding the production of inflammatory mediators are regulated and potentially silenced. Furthermore, a detailed examination of how the inflammatory SCD microenvironment modifies the functions of immune cells as well as the response of these immune cells. Identification of plasma inflammatory biomarkers with prognostic value for determining SCD patients at risk of cardiopulmonary complications may also be an additional direction of investigation that could be beneficial. The adoption and adaptation of advanced non-invasive imaging technology that provides both structural and functional information about the cardiopulmonary landscape in SCD patients may provide a novel approach to documenting cardiac remodeling in SCD patients. For example, hybrid positron emission tomography (PET) and magnetic resonance imaging (MRI) using the uptake of the radiotracer 18F-fluorodeoxyglucose (18F-FDG) have been used to identify and characterize vascular inflammation and different stages of atherosclerosis that are not yet detectable by other forms of structural imaging such as cardiac computed tomography and echocardiography [149,150]. This technique may be useful and could be adapted to SCD, where both inflammation and vascular dysfunction are present and linked with the development of cardiac hypertrophy, PH, and diastolic dysfunction. Because

inflammation may precede the development of cardiopulmonary complications in SCD patients, extending the current guidelines for screening, diagnosis, and management of cardiopulmonary complications in SCD patients [42], by adding biomarkers of inflammation linked with cardiopulmonary complications. This may improve risk stratification and help with better identification of patients needing more urgent therapy, thus preventing the progression to irreversible organ damage.

**Table 1.** Summary of inflammatory mediators and potential novel treatments in cardiopulmonary complications in sickle cell Disease.

| Disease complication                            | Major contributors   | Potential novel treatments that may target major inflammatory/anti-inflammatory pathways                | Citations  |
|---|--|---|--|
| 1. Acute Chest Syndrome (ACS)                   | Free heme, heme oxygenase (HMOX-1), neutrophils and platelet interactions, p-selectin                                | Glyco-protein Ibalpha inhibitor (CCP-224) [24]<br>D3T (3H-1,2-dithiole-3-thione) [37]<br>Hemopexin [38] | Anea [24], Jimenez [25], Ghosh [26], Bean [36], Ghosh [37], Ghosh [38], Alishlash [39]                               |
| 2. Pulmonary hypertension                       | Endothelial dysfunction, hemolysis, decreased NO, increased placenta growth factor (PIGF), PPAR alpha and PPAR gamma | Hemopexin [38]<br>BAY 54-6544 <sup>55</sup>   | Jang [53], Wood [55], Gonzales [58], Hsu [47], Morris [50] Perelman [109], Selvaraj [110], Potoka [56], Buehler [63] |
| 3. Pulmonary thrombosis                         | NETs, DAMPs, tissue factor upregulation, lower protein S and C endothelial dysfunction                               | Anti-TF antibody  | Sparkenbaugh [105]<br>Whelihan [85]<br>Faes [77]<br>Solovey [82]   |
| 4. Cardiac hypertrophy                          | ROS, endothelial dysfunction, hemolysis, hypercoagulation, PIGF, IL-6, heme,   | Rivaroxaban [104]   | Sparkenbaugh [104]<br>Bakeer [137]<br>Gbotosho [16]<br>Menon [14]<br>Arumugam [106]                                  |
| 5. Diastolic dysfunction and cardiac arrhythmia | IL-18, FUCA-2  | Anti-IL-18 binding protein [133]  | Duarte [11]<br>Gupta [134]   |

Overall, early diagnosis of cardiopulmonary complication through early recognition and application of molecular risks factors before irreversible organ damage occurs would contribute to a better quality of life, as SCD patients are now living longer due to the availability of disease-modifying therapies.

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## References

1. Piel F, Hay S, Gupta S, Weatherall D, Williams T. Global burden of sickle cell anaemia in children under five, 2010-2050: modelling based on demographics, excess mortality, and interventions. *PLoS Med* 2013;**10**:e1001484.
2. Piel F, Patil A, Howes R, Nyangiri O, Gething P, Dewi M, Temperley W, Williams T, Weatherall D, Haya S. Global epidemiology of sickle haemoglobin in neonates: a contemporary geostatistical model-based map and population estimates. *Lancet* 2013;**381**:142–51.
3. Fitzhugh C, Lauder N, Jonassaint J, Telen M, Zhao X, Wright E, Gilliam F, De Castro L. Cardiopulmonary complications leading to premature deaths in adult patients with sickle cell disease. *Am J Hematol* 2010;**85**:36–40.
4. Gladwin M. Cardiovascular complications and risk of death in sickle-cell disease. *Lancet* 2016;**387**:2565–74.
5. Sachdev V, Rosing DR, Thein SL. Cardiovascular complications of sickle cell disease. *Trends Cardiovasc Med* 2020;
6. Alfaddagh A, Martin SS, Leucker TM, Michos ED, Blaha MJ, Lowenstein CJ, Jones SR, Toth PP. Inflammation and cardiovascular disease: From mechanisms to therapeutics. *Am J Prev Cardiol* 2020;**4**:100130.
7. Pradhan AD, Aday AW, Rose LM, Ridker PM. Residual Inflammatory Risk on Treatment With PCSK9 Inhibition and Statin Therapy. *Circulation* 2018;**138**:141–49.
8. Fang L, Ellims AH, Beale AL, Taylor AJ, Murphy A, Dart AM. Systemic inflammation is associated with myocardial fibrosis, diastolic dysfunction, and cardiac hypertrophy in patients with hypertrophic cardiomyopathy. *Am J Transl Res* 2017;**9**:5063–73.
9. Xiao Z, Kong B, Yang H, Dai C, Fang J, Qin T, Huang H. Key Player in Cardiac Hypertrophy, Emphasizing the Role of Toll-Like Receptor 4. *Front Cardiovasc Med* 2020;**7**:579036.
10. Wang RR, Yuan TY, Wang JM, Chen YC, Zhao JL, Li MT, Fang LH, Du GH. Immunity and inflammation in pulmonary arterial hypertension: From pathophysiology mechanisms to treatment perspective. *Pharmacol Res* 2022;**180**:106238.
11. Duarte JD, Desai AA, Sysol JR, Abbasi T, Patel AR, Lang RM, Gupta A, Garcia JG, Gordeuk VR, Machado RF. Genome-Wide Analysis Identifies IL-18 and FUCA2 as Novel Genes Associated with Diastolic Function in African Americans with Sickle Cell Disease. *PloS one* 2016;**11**:e0163013.
12. Allali S, de Montalembert M, Rignault-Bricard R, Taylor M, Brice J, Brousse V, Talbot JM, Moulin F, Heilbronner C, Hermine O, Maciel TT. IL-6 levels are dramatically high in the sputum from children with sickle cell disease during acute chest syndrome. *Blood advances* 2020;**4**:6130–34.
13. Gbotosho OT, Kapetanaki MG, Ghosh S, Villanueva FS, Ofori-Acquah SF, Kato GJ. Heme Induces IL-6 and Cardiac Hypertrophy Genes Transcripts in Sickle Cell Mice. *Frontiers in Immunology* 2020;
14. Menon AV, Liu J, Tsai HP, Zeng L, Yang S, Asnani A, Kim J. Excess heme upregulates heme oxygenase 1 and promotes cardiac ferroptosis in mice with sickle cell disease. *Blood* 2022;**139**:936–41.
15. Karoor V, Swindle D, Pak DI, Strassheim D, Fini MA, Dempsey E, Stenmark KR, Hassell K, Nuss R, Buehler PW, Irwin DC. Evidence supporting a role for circulating macrophages in the regression of vascular remodeling following sub-chronic exposure to hemoglobin plus hypoxia. *Pulmonary circulation* 2021;**11**:20458940211056806.
16. Gbotosho OT, Kapetanaki MG, Ghosh S, Villanueva FS, Ofori-Acquah SF, Kato GJ. Heme Induces IL-6 and Cardiac Hypertrophy Genes Transcripts in Sickle Cell Mice. *Frontiers in immunology* 2020;**11**:1910.
17. Vichinsky EP, Styles LA, Colangelo LH, Wright EC, Castro O, Nickerson B. Acute chest syndrome in sickle cell disease: clinical presentation and course. Cooperative Study of Sickle Cell Disease. *Blood* 1997;**89**:1787–92.
18. Vichinsky EP, Neumayr LD, Earles AN, Williams R, Lennette ET, Dean D, Nickerson B, Orringer E, McKie V, Bellevue R, Daeschner C, Mancini EA. Causes and outcomes of the acute chest syndrome in sickle cell disease. National Acute Chest Syndrome Study Group. *The New England journal of medicine* 2000;**342**:1855–65.
19. Jain S, Bakshi N, Krishnamurti L. Acute Chest Syndrome in Children with Sickle Cell Disease. *Pediatr Allergy Immunol Pulmonol* 2017;**30**:191–201.
20. Bennwitz MF, Jimenez MA, Vats R, Tutuncuoglu E, Jonassaint J, Kato GJ, Gladwin MT, Sundt P. Lung vaso-occlusion in sickle cell disease mediated by arteriolar neutrophil-platelet microemboli. *JCI Insight* 2017;**2**:e89761.
21. Gladwin MT, Vichinsky E. Pulmonary complications of sickle cell disease. *N Engl J Med* 2008;**359**:2254–65.
22. Reichel CA, Rehberg M, Lerchenberger M, Berberich N, Bihari P, Khandoga AG, Zahler S, Krombach F. Ccl2 and Ccl3 mediate neutrophil recruitment via induction of protein synthesis and generation of lipid mediators. *Arterioscler Thromb Vasc Biol* 2009;**29**:1787–93.
23. Koltsova EK, Sundt P, Zarpellon A, Ouyang H, Mikulski Z, Zampolli A, Ruggeri ZM, Ley K. Genetic deletion of platelet glycoprotein Ib alpha but not its extracellular domain protects from atherosclerosis. *Thromb Haemost* 2014;**112**:1252–63.

24. Anea CB, Lyon M, Lee IA, Gonzales JN, Adeyemi A, Falls G, Kutlar A, Brittain JE. Pulmonary platelet thrombi and vascular pathology in acute chest syndrome in patients with sickle cell disease. *Am J Hematol* 2016;**91**:173-8
25. Jimenez MA, Novelli E, Shaw GD, Sundd P. Glycoprotein Ibalph inhibitor (CCP-224) prevents neutrophil-platelet aggregation in Sickle Cell Disease. *Blood Adv* 2017;**1**:1712-16
26. Ghosh S, Flage B, Weidert F, Ofori-Acquah SF. P-selectin plays a role in haem-induced acute lung injury in sickle mice. *British journal of haematology* 2019;**186**:329-33
27. Gbotosho OT, Kapetanaki MG, Kato GJ. The Worst Things in Life are Free: The Role of Free Heme in Sickle Cell Disease. *Frontiers in immunology* 2020;**11**:561917
28. Kato GJ, Steinberg MH, Gladwin MT. Intravascular hemolysis and the pathophysiology of sickle cell disease. *The Journal of clinical investigation* 2017;**127**:750-60
29. Gladwin MT, Kanas T, Kim-Shapiro DB. Hemolysis and cell-free hemoglobin drive an intrinsic mechanism for human disease. *J Clin Invest* 2012;**122**:1205-8
30. Gladwin MT, Ofori-Acquah SF. Erythroid DAMPs drive inflammation in SCD. *Blood* 2014;**123**:3689-90
31. Chen G, Zhang D, Fuchs TA, Manwani D, Wagner DD, Frenette PS. Heme-induced neutrophil extracellular traps contribute to the pathogenesis of sickle cell disease. *Blood* 2014;**123**:3818-27
32. Rank BH, Carlsson J, Hebbel RP. Abnormal redox status of membrane-protein thiols in sickle erythrocytes. *J Clin Invest* 1985;**75**:1531-7
33. Reiter CD, Wang X, Tanus-Santos JE, Hogg N, Cannon RO, 3rd, Schechter AN, Gladwin MT. Cell-free hemoglobin limits nitric oxide bioavailability in sickle-cell disease. *Nat Med* 2002;**8**:1383-9
34. Jones NM, Sysol JR, Singla S, Smith P, Sandusky GE, Wang H, Natarajan V, Dudek SM, Machado RF. Cortactin loss protects against hemin-induced acute lung injury in sickle cell disease. *Am J Physiol Lung Cell Mol Physiol* 2022;**322**:L890-L97
35. Adisa OA, Hu Y, Ghosh S, Aryee D, Osunkwo I, Ofori-Acquah SF. Association between plasma free haem and incidence of vaso-occlusive episodes and acute chest syndrome in children with sickle cell disease. *British journal of haematology* 2013;**162**:702-5
36. Bean CJ, Boulet SL, Ellingsen D, Pyle ME, Barron-Casella EA, Casella JF, Payne AB, Driggers J, Trau HA, Yang G, Jones K, Ofori-Acquah SF, Hooper WC, DeBaun MR. Heme oxygenase-1 gene promoter polymorphism is associated with reduced incidence of acute chest syndrome among children with sickle cell disease. *Blood* 2012;**120**:3822-8
37. Ghosh S, Adisa OA, Chappa P, Tan F, Jackson KA, Archer DR, Ofori-Acquah SF. Extracellular hemin crisis triggers acute chest syndrome in sickle mice. *The Journal of clinical investigation* 2013;**123**:4809-20
38. Ghosh S, Hazra R, Ihunnah CA, Weidert F, Flage B, Ofori-Acquah SF. Augmented NRF2 activation protects adult sickle mice from lethal acute chest syndrome. *British journal of haematology* 2018;**182**:271-75
39. Alishlash AS, Sapkota M, Ahmad I, Maclin K, Ahmed NA, Molyvdas A, Doran S, Albert CJ, Aggarwal S, Ford DA, Ambalavanan N, Jilling T, Matalon S. Chlorine inhalation induces acute chest syndrome in humanized sickle cell mouse model and ameliorated by postexposure hemopexin. *Redox Biol* 2021;**44**:102009
40. Platt OS, Brambilla DJ, Rosse WF, Milner PF, Castro O, Steinberg MH, Klug PP. Mortality in sickle cell disease. Life expectancy and risk factors for early death. *N Engl J Med* 1994;**330**:1639-44
41. Parent F, Bachir D, Inamo J, Lionnet F, Driss F, Loko G, Habibi A, Bennani S, Savale L, Adnot S, Maitre B, Yaici A, Hajji L, O'Callaghan DS, Clerson P, Girot R, Galacteros F, Simonneau G. A hemodynamic study of pulmonary hypertension in sickle cell disease. *N Engl J Med* 2011;**365**:44-53
42. Liem RI, Lanzkron S, T DC, DeCastro L, Desai AA, Ataga KI, Cohen RT, Haynes J, Osunkwo I, Lebensburger JD, Lash JP, Wun T, Verhovsek M, Ontala E, Blaylark R, Alahdab F, Katabi A, Mustafa RA. American Society of Hematology 2019 guidelines for sickle cell disease: cardiopulmonary and kidney disease. *Blood advances* 2019;**3**:3867-97
43. Adedeji MO, Cespedes J, Allen K, Subramony C, Hughson MD. Pulmonary thrombotic arteriopathy in patients with sickle cell disease. *Arch Pathol Lab Med* 2001;**125**:1436-41
44. Wood KC, Gladwin MT, Straub AC. Sickle cell disease: at the crossroads of pulmonary hypertension and diastolic heart failure. *Heart* 2020;**106**:562-68
45. Redinus K, Baek JH, Yalamanoglu A, Shin HKH, Moldova R, Harral JW, Swindle D, Pak D, Ferguson SK, Nuss R, Hassell K, Nozik-Grayck E, Palmer AF, Fini MA, Karoor V, Stenmark KR, Buehler PW, Irwin DC. An Hb-mediated circulating macrophage contributing to pulmonary vascular remodeling in sickle cell disease. *JCI Insight* 2019;**4**
46. Buehler PW, Swindle D, Pak DI, Fini MA, Hassell K, Nuss R, Wilkerson RB, D'Alessandro A, Irwin DC. Murine models of sickle cell disease and beta-thalassemia demonstrate pulmonary hypertension with distinctive features. *Pulm Circ* 2021;**11**:20458940211055996
47. Hsu LL, Champion HC, Campbell-Lee SA, Bivalacqua TJ, Mancini EA, Diwan BA, Schimel DM, Cochard AE, Wang X, Schechter AN, Noguchi CT, Gladwin MT. Hemolysis in sickle cell mice causes pulmonary hypertension due to global impairment in nitric oxide bioavailability. *Blood* 2007;**109**:3088-98

48. Gladwin MT, Barst RJ, Castro OL, Gordeuk VR, Hillery CA, Kato GJ, Kim-Shapiro DB, Machado R, Morris CR, Steinberg MH, Vichinsky EP. Pulmonary hypertension and NO in sickle cell. *Blood* 2010;**116**:852-4
49. Minneci PC, Deans KJ, Zhi H, Yuen PS, Star RA, Banks SM, Schechter AN, Natanson C, Gladwin MT, Solomon SB. Hemolysis-associated endothelial dysfunction mediated by accelerated NO inactivation by decompartmentalized oxyhemoglobin. *J Clin Invest* 2005;**115**:3409-17
50. Morris CR, Kato GJ, Poljakovic M, Wang X, Blackwelder WC, Sachdev V, Hazen SL, Vichinsky EP, Morris SM, Jr., Gladwin MT. Dysregulated arginine metabolism, hemolysis-associated pulmonary hypertension, and mortality in sickle cell disease. *JAMA* 2005;**294**:81-90
51. Reiter C, Wang X, Tanus-Santos J, Hogg N, Cannon Rr, Schechter A, MT G. Cell-free hemoglobin limits nitric oxide bioavailability in sickle-cell disease. *Nat Med* 2002;**8**:1383-9.
52. Palmer RM, Ferrige AG, Moncada S. Nitric oxide release accounts for the biological activity of endothelium-derived relaxing factor. *Nature* 1987;**327**:524-6
53. Lee MR, Li L, Kitazawa T. Cyclic GMP causes Ca<sup>2+</sup> desensitization in vascular smooth muscle by activating the myosin light chain phosphatase. *J Biol Chem* 1997;**272**:5063-8
54. Almeida CB, Souza LE, Leonardo FC, Costa FT, Werneck CC, Covas DT, Costa FF, Conran N. Acute hemolytic vascular inflammatory processes are prevented by nitric oxide replacement or a single dose of hydroxyurea. *Blood* 2015;**126**:711-20
55. Jang AJ, Chang SS, Park C, Lee CM, Benza RL, Passineau MJ, Ma J, Archer DR, Sutliff RL, Hart CM, Kang BY. PPARgamma increases HUWE1 to attenuate NF-kappaB/p65 and sickle cell disease with pulmonary hypertension. *Blood Adv* 2021;**5**:399-413
56. Zuckerbraun BS, Shiva S, Ifedigbo E, Mathier MA, Mollen KP, Rao J, Bauer PM, Choi JJ, Curtis E, Choi AM, Gladwin MT. Nitrite potently inhibits hypoxic and inflammatory pulmonary arterial hypertension and smooth muscle proliferation via xanthine oxidoreductase-dependent nitric oxide generation. *Circulation* 2010;**121**:98-109
57. Wood KC, Durgin BG, Schmidt HM, Hahn SA, Baust JJ, Bachman T, Vitturi DA, Ghosh S, Ofori-Acquah SF, Mora AL, Gladwin MT, Straub AC. Smooth muscle cytochrome b5 reductase 3 deficiency accelerates pulmonary hypertension development in sickle cell mice. *Blood advances* 2019;**3**:4104-16
58. Potoka KP, Wood KC, Baust JJ, Bueno M, Hahn SA, Vanderpool RR, Bachman T, Mallampalli GM, Osei-Hwedieh DO, Schrott V, Sun B, Bullock GC, Becker-Pelster EM, Wittwer M, Stampfuss J, Mathar I, Stasch JP, Truebel H, Sandner P, Mora AL, Straub AC, Gladwin MT. Nitric Oxide-Independent Soluble Guanylate Cyclase Activation Improves Vascular Function and Cardiac Remodeling in Sickle Cell Disease. *Am J Respir Cell Mol Biol* 2018;**58**:636-47
59. Ataga KI, Moore CG, Hillery CA, Jones S, Whinna HC, Strayhorn D, Sohler C, Hinderliter A, Parise LV, Orringer EP. Coagulation activation and inflammation in sickle cell disease-associated pulmonary hypertension. *Haematologica* 2008;**93**:20-6
60. Gonzales J, Holbert K, Czysk K, George J, Fernandes C, Fraidenburg DR. Hemin-Induced Endothelial Dysfunction and Endothelial to Mesenchymal Transition in the Pathogenesis of Pulmonary Hypertension Due to Chronic Hemolysis. *Int J Mol Sci* 2022;**23**
61. Wang X, Mendelsohn L, Rogers H, Leitman S, Raghavachari N, Yang Y, Yau Y, Tallack M, Perkins A, Taylor J, Noguchi C, Kato G. Heme-bound iron activates placenta growth factor in erythroid cells via erythroid Krüppel-like factor. *Blood* 2014;**124**:946-54.
62. Kapetanaki MG, Gbotosho OT, Sharma D, Weidert F, Ofori-Acquah SF, Kato GJ. Free heme regulates placenta growth factor through NRF2-antioxidant response signaling. *Free radical biology & medicine* 2019;**143**:300-08
63. Kalra VK, Zhang S, Malik P, Tahara SM. Placenta growth factor mediated gene regulation in sickle cell disease. *Blood reviews* 2018;**32**:61-70
64. Sundaram N, Tailor A, Mendelsohn L, Wansapura J, Wang X, Higashimoto T, Pauciulo MW, Gottliebson W, Kalra VK, Nichols WC, Kato GJ, Malik P. High levels of placenta growth factor in sickle cell disease promote pulmonary hypertension. *Blood* 2010;**116**:109-12
65. Buehler PW, Swindle D, Pak DI, Ferguson SK, Majka SM, Karoor V, Moldovan R, Sintas C, Black J, Gentinetta T, Buzzi RM, Vallelian F, Wassmer A, Edler M, Bain J, Schu D, Hassell K, Nuss R, Schaer DJ, Irwin DC. Hemopexin dosing improves cardiopulmonary dysfunction in murine sickle cell disease. *Free radical biology & medicine* 2021;**175**:95-107
66. Minniti CP, Machado RF, Coles WA, Sachdev V, Gladwin MT, Kato GJ. Endothelin receptor antagonists for pulmonary hypertension in adult patients with sickle cell disease. *British journal of haematology* 2009;**147**:737-43
67. Weir NA, Conrey A, Lewis D, Mehari A. Riociguat use in sickle cell related chronic thromboembolic pulmonary hypertension: A case series. *Pulm Circ* 2018;**8**:2045894018791802
68. Machado RF, Martyr S, Kato GJ, Barst RJ, Anthi A, Robinson MR, Hunter L, Coles W, Nichols J, Hunter C, Sachdev V, Castro O, Gladwin MT. Sildenafil therapy in patients with sickle cell disease and pulmonary hypertension. *Br J Haematol* 2005;**130**:445-53

69. Cramer-Bour C, Ruhl AP, Nouraie SM, Emeh RO, Ruopp NF, Thein SL, Weir NA, Klings ES. Long-term tolerability of phosphodiesterase-5 inhibitors in pulmonary hypertension of sickle cell disease. *Eur J Haematol* 2021;**107**:54-62
70. Naik RP, Streiff MB, Haywood C, Jr., Nelson JA, Lanzkron S. Venous thromboembolism in adults with sickle cell disease: a serious and under-recognized complication. *Am J Med* 2013;**126**:443-9
71. Brunson A, Lei A, Rosenberg AS, White RH, Keegan T, Wun T. Increased incidence of VTE in sickle cell disease patients: risk factors, recurrence and impact on mortality. *Br J Haematol* 2017;**178**:319-26
72. Stein PD, Beemath A, Meyers FA, Skaf E, Olson RE. Deep venous thrombosis and pulmonary embolism in hospitalized patients with sickle cell disease. *Am J Med* 2006;**119**:e7-11
73. Naik RP, Streiff MB, Haywood C, Jr., Segal JB, Lanzkron S. Venous thromboembolism incidence in the Cooperative Study of Sickle Cell Disease. *J Thromb Haemost* 2014;**12**:2010-6
74. Gollamudi J, Sarvepalli S, Vadaparti Binif A, Alin T, Little JA, Nayak L. Venous Thromboembolism in Sickle Cell Disease is Associated with Neutrophilia. *Hemoglobin* 2021;**45**:56-59
75. Srisuwananukorn A, Raslan R, Zhang X, Shah BN, Han J, Gowhari M, Molokie RE, Gordeuk VR, Saraf SL. Clinical, laboratory, and genetic risk factors for thrombosis in sickle cell disease. *Blood Adv* 2020;**4**:1978-86
76. Kumar R, Stanek J, Creary S, Dunn A, O'Brien SH. Prevalence and risk factors for venous thromboembolism in children with sickle cell disease: an administrative database study. *Blood Adv* 2018;**2**:285-91
77. Mendonca R, Silveira AA, Conran N. Red cell DAMPs and inflammation. *Inflamm Res* 2016;**65**:665-78
78. Xu H, Wandersee NJ, Guo Y, Jones DW, Holzhauer SL, Hanson MS, Machogu E, Brousseau DC, Hogg N, Densmore JC, Kaul S, Hillery CA, Pritchard KA, Jr. Sickle cell disease increases high mobility group box 1: a novel mechanism of inflammation. *Blood* 2014;**124**:3978-81
79. Vogel S, Arora T, Wang X, Mendelsohn L, Nichols J, Allen D, Shet AS, Combs CA, Quezada ZMN, Thein SL. The platelet NLRP3 inflammasome is upregulated in sickle cell disease via HMGB1/TLR4 and Bruton tyrosine kinase. *Blood advances* 2018;**2**:2672-80
80. Murthy P, Durco F, Miller-Ocuin JL, Takedai T, Shankar S, Liang X, Liu X, Cui X, Sachdev U, Rath D, Lotze MT, Zeh HJ, 3rd, Gawaz M, Weber AN, Vogel S. The NLRP3 inflammasome and bruton's tyrosine kinase in platelets co-regulate platelet activation, aggregation, and in vitro thrombus formation. *Biochemical and biophysical research communications* 2017;**483**:230-36
81. Belcher JD, Chen C, Nguyen J, Milbauer L, Abdulla F, Alayash AI, Smith A, Nath KA, Hebbel RP, Vercellotti GM. Heme triggers TLR4 signaling leading to endothelial cell activation and vaso-occlusion in murine sickle cell disease. *Blood* 2014;**123**:377-90
82. Faes C, Ilich A, Sotiaux A, Sparkenbaugh EM, Henderson MW, Buczek L, Beckman JD, Ellsworth P, Noubouossie DF, Bhoopat L, Piegore M, Renoux C, Bergmeier W, Park Y, Ataga KI, Cooley B, Wolberg AS, Key NS, Pawlinski R. Red blood cells modulate structure and dynamics of venous clot formation in sickle cell disease. *Blood* 2019;**133**:2529-41
83. van Beers EJ, Schaap MC, Berckmans RJ, Nieuwland R, Sturk A, van Doormaal FF, Meijers JC, Biemond BJ, group Cs. Circulating erythrocyte-derived microparticles are associated with coagulation activation in sickle cell disease. *Haematologica* 2009;**94**:1513-9
84. Vats R, Brzoska T, Bennewitz MF, Jimenez MA, Pradhan-Sundt T, Tutuncuoglu E, Jonassaint J, Gutierrez E, Watkins SC, Shiva S, Scott MJ, Morelli AE, Neal MD, Kato GJ, Gladwin MT, Sundt P. Platelet Extracellular Vesicles Drive Inflammasome-IL-1 $\beta$ -Dependent Lung Injury in Sickle Cell Disease. *Am J Respir Crit Care Med* 2020;**201**:33-46
85. Grover SP, Mackman N. Tissue Factor: An Essential Mediator of Hemostasis and Trigger of Thrombosis. *Arterioscler Thromb Vasc Biol* 2018;**38**:709-25
86. Noubouossie DF, Whelihan MF, Yu YB, Sparkenbaugh E, Pawlinski R, Monroe DM, Key NS. In vitro activation of coagulation by human neutrophil DNA and histone proteins but not neutrophil extracellular traps. *Blood* 2017;**129**:1021-29
87. Solovey A, Kollander R, Shet A, Milbauer LC, Choong S, Panoskaltis-Mortari A, Blazar BR, Kelm RJ, Jr., Hebbel RP. Endothelial cell expression of tissue factor in sickle mice is augmented by hypoxia/reoxygenation and inhibited by lovastatin. *Blood* 2004;**104**:840-6
88. Zhou Z, Han H, Cruz MA, Lopez JA, Dong JF, Guchhait P. Haemoglobin blocks von Willebrand factor proteolysis by ADAMTS-13: a mechanism associated with sickle cell disease. *Thromb Haemost* 2009;**101**:1070-7
89. Schnog JB, Mac Gillavry MR, van Zanten AP, Meijers JC, Rojer RA, Duits AJ, ten Cate H, Brandjes DP. Protein C and S and inflammation in sickle cell disease. *Am J Hematol* 2004;**76**:26-32
90. Whelihan MF, Lim MY, Mooberry MJ, Piegore MG, Ilich A, Wogu A, Cai J, Monroe DM, Ataga KI, Mann KG, Key NS. Thrombin generation and cell-dependent hypercoagulability in sickle cell disease. *J Thromb Haemost* 2016;**14**:1941-52
91. Gavins FN, Russell J, Senchenkova EL, De Almeida Paula L, Damazo AS, Esmon CT, Kirchhofer D, Hebbel RP, Granger DN. Mechanisms of enhanced thrombus formation in cerebral microvessels of mice expressing hemoglobin-S. *Blood* 2011;**117**:4125-33

92. Schulz C, Massberg S. Demystifying the prothrombotic role of NETs. *Blood* 2017;**129**:925-26
93. Leong MA, Dampier C, Varlotta L, Allen JL. Airway hyperreactivity in children with sickle cell disease. *J Pediatr* 1997;**131**:278-83
94. Vendramini EC, Vianna EO, De Lucena Ethngulo I, De Castro FB, Martinez JAB, Terra-Filho J. Lung function and airway hyperresponsiveness in adult patients with sickle cell disease. *Am J Med Sci* 2006;**332**:68-72
95. An P, Barron-Casella EA, Strunk RC, Hamilton RG, Casella JF, DeBaun MR. Elevation of IgE in children with sickle cell disease is associated with doctor diagnosis of asthma and increased morbidity. *J Allergy Clin Immunol* 2011;**127**:1440-6
96. Field JJ, Horst J, Strunk RC, White FV, DeBaun MR. Death due to asthma in two adolescents with sickle cell disease. *Pediatr Blood Cancer* 2011;**56**:454-7
97. Eiyimo Mwa Mpollo MS, Brandt EB, Shanmukhappa SK, Arumugam PI, Tiwari S, Loberg A, Pillis D, Rizvi T, Lindsey M, Jonck B, Carmeliet P, Kalra VK, Le Cras TD, Ratner N, Wills-Karp M, Hershey GK, Malik P. Placenta growth factor augments airway hyperresponsiveness via leukotrienes and IL-13. *The Journal of clinical investigation* 2016;**126**:571-84
98. Postma DS, Kerstjens HA. Characteristics of airway hyperresponsiveness in asthma and chronic obstructive pulmonary disease. *Am J Respir Crit Care Med* 1998;**158**:S187-92
99. Chapman DG, Irvin CG. Mechanisms of airway hyper-responsiveness in asthma: the past, present and yet to come. *Clin Exp Allergy* 2015;**45**:706-19
100. Wills-Karp M, Luyimbazi J, Xu X, Schofield B, Neben TY, Karp CL, Donaldson DD. Interleukin-13: central mediator of allergic asthma. *Science* 1998;**282**:2258-61
101. Andemariam B, Adami AJ, Singh A, McNamara JT, Secor ER, Guernsey LA, Thrall RS. The sickle cell mouse lung: proinflammatory and primed for allergic inflammation. *Transl Res* 2015;**166**:254-68
102. Nandedkar SD, Feroah TR, Hutchins W, Weihrauch D, Konduri KS, Wang J, Strunk RC, DeBaun MR, Hillery CA, Pritchard KA. Histopathology of experimentally induced asthma in a murine model of sickle cell disease. *Blood* 2008;**112**:2529-38
103. Niss O, Fleck R, Makue F, Alsaied T, Desai P, Towbin JA, Malik P, Taylor MD, Quinn CT. Association between diffuse myocardial fibrosis and diastolic dysfunction in sickle cell anemia. *Blood* 2017;**130**:205-13
104. Niss O, Quinn CT, Lane A, Daily J, Khoury PR, Bakeer N, Kimball TR, Towbin JA, Malik P, Taylor MD. Cardiomyopathy With Restrictive Physiology in Sickle Cell Disease. *JACC Cardiovasc Imaging* 2016;**9**:243-52
105. Muchtar E, Blauwet LA, Gertz MA. Restrictive Cardiomyopathy: Genetics, Pathogenesis, Clinical Manifestations, Diagnosis, and Therapy. *Circ Res* 2017;**121**:819-37
106. Varat MA, Adolph RJ, Fowler NO. Cardiovascular effects of anemia. *American heart journal* 1972;**83**:415-26
107. Hammoudi N, Lionnet F, Redheuil A, Montalescot G. Cardiovascular manifestations of sickle cell disease. *Eur Heart J* 2020;**41**:1365-73
108. James TN. Homage to James B. Herrick: a contemporary look at myocardial infarction and at sickle-cell heart disease: the 32nd Annual Herrick Lecture of the Council on Clinical Cardiology of the American Heart Association. *Circulation* 2000;**101**:1874-87
109. Gbotosho OT, Taylor M, Malik P. Cardiac pathophysiology in sickle cell disease. *J Thromb Thrombolysis* 2021;**52**:248-59
110. Sparkenbaugh EM, Chantrathamchart P, Chandarajoti K, Mackman N, Key NS, Pawlinski R. Thrombin-independent contribution of tissue factor to inflammation and cardiac hypertrophy in a mouse model of sickle cell disease. *Blood* 2016;**127**:1371-3
111. Arumugam PI, Mullins ES, Shanmukhappa SK, Monia BP, Loberg A, Shaw MA, Rizvi T, Wansapura J, Degen JL, Malik P. Genetic diminution of circulating prothrombin ameliorates multiorgan pathologies in sickle cell disease mice. *Blood* 2015;**126**:1844-55
112. Solovey A, Lin Y, Browne P, Choong S, Wayne E, Hebbel RP. Circulating activated endothelial cells in sickle cell anemia. *The New England journal of medicine* 1997;**337**:1584-90
113. Camus SM, De Moraes JA, Bonnin P, Abbyad P, Le Jeune S, Lionnet F, Loufrani L, Grimaud L, Lambry JC, Charue D, Kiger L, Renard JM, Larroque C, Le Clesiau H, Tedgui A, Bruneval P, Barja-Fidalgo C, Alexandrou A, Tharaux PL, Boulanger CM, Blanc-Brude OP. Circulating cell membrane microparticles transfer heme to endothelial cells and trigger vasoocclusions in sickle cell disease. *Blood* 2015;**125**:3805-14
114. Perelman N, Selvaraj SK, Batra S, Luck LR, Erdreich-Epstein A, Coates TD, Kalra VK, Malik P. Placenta growth factor activates monocytes and correlates with sickle cell disease severity. *Blood* 2003;**102**:1506-14
115. Selvaraj SK, Giri RK, Perelman N, Johnson C, Malik P, Kalra VK. Mechanism of monocyte activation and expression of proinflammatory cytochemokines by placenta growth factor. *Blood* 2003;**102**:1515-24
116. Antwi-Boasiako C, Donkor ES, Sey F, Dzudzor B, Dankwah GB, Otu KH, Doku A, Dale CA, Ekem I. Levels of Soluble Endothelium Adhesion Molecules and Complications among Sickle Cell Disease Patients in Ghana. *Diseases* 2018;**6**

117. Sarray S, Saleh LR, Lisa Saldanha F, Al-Habboubi HH, Mahdi N, Almawi WY. Serum IL-6, IL-10, and TNFalpha levels in pediatric sickle cell disease patients during vasoocclusive crisis and steady state condition. *Cytokine* 2015;**72**:43-7
118. Gbotosho OT, Ghosh S, Kapetanaki MG, Lin Y, Weidert F, Bullock GC, Ofori-Acquah SF, Kato GJ. Cardiac expression of HMOX1 and PGF in sickle cell mice and haem-treated wild type mice dominates organ expression profiles via Nrf2 (Nfe2l2). *British journal of haematology* 2019;**187**:666-75
119. Meloni A, Puliyl M, Pepe A, Berdoukas V, Coates TD, Wood JC. Cardiac iron overload in sickle-cell disease. *American journal of hematology* 2014;**89**:678-83
120. Tavares AHJ, Benites BD, Fertrin KY. Myocardial Iron Overload in Sickle Cell Disease: A Rare But Potentially Fatal Complication of Transfusion. *Transfus Med Rev* 2019;**33**:170-75
121. Sparkenbaugh E, Pawlinski R. Interplay between coagulation and vascular inflammation in sickle cell disease. *British journal of haematology* 2013;**162**:3-14
122. Chantrathammachart P, Mackman N, Sparkenbaugh E, Wang JG, Parise LV, Kirchhofer D, Key NS, Pawlinski R. Tissue factor promotes activation of coagulation and inflammation in a mouse model of sickle cell disease. *Blood* 2012;**120**:636-46
123. Dewerchin M, Carmeliet P. PlGF: a multitasking cytokine with disease-restricted activity. *Cold Spring Harbor perspectives in medicine* 2012;**2**
124. Carnevale D, Cifelli G, Mascio G, Madonna M, Sbroglio M, Perrino C, Persico MG, Frati G, Lembo G. Placental growth factor regulates cardiac inflammation through the tissue inhibitor of metalloproteinases-3/tumor necrosis factor-alpha-converting enzyme axis: crucial role for adaptive cardiac remodeling during cardiac pressure overload. *Circulation* 2011;**124**:1337-50
125. Accornero F, van Berlo JH, Benard MJ, Lorenz JN, Carmeliet P, Molkentin JD. Placental growth factor regulates cardiac adaptation and hypertrophy through a paracrine mechanism. *Circ Res* 2011;**109**:272-80
126. Iwama H, Uemura S, Naya N, Imagawa K, Takemoto Y, Asai O, Onoue K. Cardiac expression of placental growth factor predicts the improvement of chronic phase left ventricular function in patients with acute myocardial infarction. *J Am Coll Cardiol* 2006;**47**:1559-67
127. Kolakowski S, Jr., Berry MF, Atluri P, Grand T, Fisher O, Moise MA, Cohen J, Hsu V, Woo YJ. Placental growth factor provides a novel local angiogenic therapy for ischemic cardiomyopathy. *Journal of cardiac surgery* 2006;**21**:559-64
128. Matsui M, Uemura S, Takeda Y, Samejima K, Matsumoto T, Hasegawa A, Tsushima H, Hoshino E, Ueda T, Morimoto K, Okamoto K, Okada S, Onoue K, Okayama S, Kawata H, Kawakami R, Maruyama N, Akai Y, Iwano M, Shiiki H, Saito Y. Placental Growth Factor as a Predictor of Cardiovascular Events in Patients with CKD from the NARA-CKD Study. *Journal of the American Society of Nephrology : JASN* 2015;**26**:2871-81
129. Gbotosho OT, Ghosh S, Kapetanaki MG, Lin Y, Weidert F, Bullock GC, Ofori-Acquah SF, Kato GJ. Cardiac expression of HMOX1 and PGF in sickle cell mice and haem-treated wild type mice dominates organ expression profiles via Nrf2 (Nfe2l2). *British journal of haematology* 2019;**187**:666-75.
130. Gladwin MT, Sachdev V. Cardiovascular abnormalities in sickle cell disease. *J Am Coll Cardiol* 2012;**59**:1123-33
131. Johnson MC, Kirkham FJ, Redline S, Rosen CL, Yan Y, Roberts I, Gruenewald J, Marek J, DeBaun MR. Left ventricular hypertrophy and diastolic dysfunction in children with sickle cell disease are related to asleep and waking oxygen desaturation. *Blood* 2010;**116**:16-21
132. Alsaied T, Niss O, Powell AW, Fleck RJ, Cnota JF, Chin C, Malik P, Quinn CT, Taylor MD. Diastolic dysfunction is associated with exercise impairment in patients with sickle cell anemia. *Pediatric blood & cancer* 2018;**65**:e27113
133. Sachdev V, Machado RF, Shizukuda Y, Rao YN, Sidenko S, Ernst I, St Peter M, Coles WA, Rosing DR, Blackwelder WC, Castro O, Kato GJ, Gladwin MT. Diastolic dysfunction is an independent risk factor for death in patients with sickle cell disease. *J Am Coll Cardiol* 2007;**49**:472-9
134. Sachdev V, Kato GJ, Gibbs JS, Barst RJ, Machado RF, Nouraie M, Hassell KL, Little JA, Schraufnagel DE, Krishnamurti L, Novelli EM, Girgis RE, Morris CR, Rosenzweig EB, Badesch DB, Lanzkron S, Castro OL, Taylor JGt, Hannoush H, Goldsmith JC, Gladwin MT, Gordeuk VR, Walk PI. Echocardiographic markers of elevated pulmonary pressure and left ventricular diastolic dysfunction are associated with exercise intolerance in adults and adolescents with homozygous sickle cell anemia in the United States and United Kingdom. *Circulation* 2011;**124**:1452-60
135. Fu DG. Cardiac Arrhythmias: Diagnosis, Symptoms, and Treatments. *Cell Biochem Biophys* 2015;**73**:291-96
136. John RM, Tedrow UB, Koplan BA, Albert CM, Epstein LM, Sweeney MO, Miller AL, Michaud GF, Stevenson WG. Ventricular arrhythmias and sudden cardiac death. *Lancet* 2012;**380**:1520-9
137. Desai R, Bansod S, Patel U. Nationwide prevalence and trends in acute cardiovascular events and in-hospital mortality among adult African Americans with sickle cell trait. *Annals of hematology* 2020;**99**:2207-09

138. Fitzhugh CD, Lauder N, Jonassaint JC, Telen MJ, Zhao X, Wright EC, Gilliam FR, De Castro LM. Cardiopulmonary complications leading to premature deaths in adult patients with sickle cell disease. *American journal of hematology* 2010;**85**:36-40
139. Mancini EA, Culbertson DE, Yang YM, Gardner TM, Powell R, Haynes J, Jr., Shah AK, Mankad VN. Causes of death in sickle cell disease: an autopsy study. *British journal of haematology* 2003;**123**:359-65
140. Darbari D, Kple-Faget P, Kwagyan J, Rana S, Gordeuk V, Castro O. Circumstances of death in adult sickle cell disease patients. *Am J Hematol* 2006;**81**:858-63.
141. Gupta A, Fei YD, Kim TY, Xie A, Batai K, Greener I, Tang H, Ciftci-Yilmaz S, Juneman E, Indik JH, Shi G, Christensen J, Gupta G, Hillery C, Kansal MM, Parikh DS, Zhou T, Yuan JX, Kanthi Y, Bronk P, Koren G, Kittles R, Duarte JD, Garcia JGN, Machado RF, Dudley SC, Choi BR, Desai AA. IL-18 mediates sickle cell cardiomyopathy and ventricular arrhythmias. *Blood* 2021;**137**:1208-18
142. Desai AA, Patel AR, Ahmad H, Groth JV, Thiruvoipati T, Turner K, Yodwut C, Czobor P, Artz N, Machado RF, Garcia JGN, Lang RM. Mechanistic insights and characterization of sickle cell disease-associated cardiomyopathy. *Circ Cardiovasc Imaging* 2014;**7**:430-37
143. James TN, Riddick L, Massing GK. Sickle cells and sudden death: morphologic abnormalities of the cardiac conduction system. *J Lab Clin Med* 1994;**124**:507-20
144. Bakeer N, James J, Roy S, Wansapura J, Shanmukhappa SK, Lorenz JN, Osinska H, Backer K, Huby AC, Shrestha A, Niss O, Fleck R, Quinn CT, Taylor MD, Purevjav E, Aronow BJ, Towbin JA, Malik P. Sickle cell anemia mice develop a unique cardiomyopathy with restrictive physiology. *Proceedings of the National Academy of Sciences of the United States of America* 2016;**113**:E5182-91
145. Niss O, Detterich J, Wood JC, Coates TD, Malik P, Taylor MD, Quinn CT. Early initiation of disease-modifying therapy can impede or prevent diffuse myocardial fibrosis in sickle cell anemia. *Blood* 2022;**140**:1322-24
146. Platt OS. Sickle cell anemia as an inflammatory disease. *The Journal of clinical investigation* 2000;**106**:337-8
147. Belcher JD, Marker PH, Weber JP, Heibel RP, Vercellotti GM. Activated monocytes in sickle cell disease: potential role in the activation of vascular endothelium and vaso-occlusion. *Blood* 2000;**96**:2451-9
148. Pathare A, Al Kindi S, Alnaqdy AA, Daar S, Knox-Macaulay H, Dennison D. Cytokine profile of sickle cell disease in Oman. *American journal of hematology* 2004;**77**:323-8
149. Ogawa M, Nakamura S, Saito Y, Kosugi M, Magata Y. What can be seen by 18F-FDG PET in atherosclerosis imaging? The effect of foam cell formation on 18F-FDG uptake to macrophages in vitro. *J Nucl Med* 2012;**53**:55-8
150. Fernandez-Friera L, Fuster V, Lopez-Melgar B, Oliva B, Sanchez-Gonzalez J, Macias A, Perez-Asenjo B, Zamudio D, Alonso-Farto JC, Espana S, Mendiguren J, Bueno H, Garcia-Ruiz JM, Ibanez B, Fernandez-Ortiz A, Sanz J. Vascular Inflammation in Subclinical Atherosclerosis Detected by Hybrid PET/MRI. *J Am Coll Cardiol* 2019;**73**:1371-82.

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