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Review

Red-Leafed Lettuces: Genetic Variation or Epigenetic Photomorphogenesis?

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Abstract: Red-leafed lettuces (RLLs) attract attention of both consumers and researchers due to their unique decorative and functional properties. The review considers modern concepts of the causes and mechanisms that control color shades and intensity of RLLs. They are also enriched in anthocyanins that have valuable antioxidant properties. Anthocyanins are plant pigments, flavonoid chromophores, which are largely responsible for the color of many fruits and flowers, and their antioxidant activity increases the value of RLLs for consumer health. For effective selection of RLLs, it is important to know to what extent the color of lettuces is determined by the genetic variation of genes controlling the biosynthesis of anthocyanins, and to what extent by the epigenetic regulation of their expression, for which it has been shown that it is regulated in response to changes in environmental factors such as light, humidity and temperature, as well as biotic and abiotic stresses and is an adaptive reaction of plants. A number of researchers note a change or loss of genes of the wild type during domestication (cultivation) of lettuces, which also leads to the loss of their color. Mutation of genes can lead to the creation of a green variety of lettuce, which can accumulate colorless flavonoids, also valuable for consumers. In addition, mechanisms are noted that allow to regulate the intensity of red coloring through the exposure of lettuces to the light with a different spectrum, up to its disappearance in the leaves of RLLs. Thus, by controlling the mechanism of leaf coloring, through the suppression or inactivation of genes that control the biosynthesis of color pigments and the degradation of chlorophyll, it is possible to create different shades within the same lettuce variety.

Keywords: antioxidants; anthocyanins; environmental factors; epigenetics; flavonoids; gene expression; light; phytochemicals; red-leafed lettuces

1. Introduction

The world's population increasingly needs food products that not only provide nutritional value, but also contribute to improved health, reduced disease and increased life expectancy ([1]; Baslam et al., 2013). Research from the Cancer Institute has shown a correlation between increased vegetable consumption and a reduced risk of chronic diseases such as cancer, cardiovascular disease, and age-related decline in activity in older adults ([2,3]; Hung et al., 2004; Kim et al., 2018). Micronutrient deficiencies are a major global public health problem in many countries, with some authors noting that infants and pregnant women are particularly at risk of not receiving essential nutrients ([2,4]; Hung et al., 2004; Soetan et al., 2010). Particular attention should be paid to the

nutrition of children, since sufficient amounts of macro- and micronutrients are necessary for their full development. It is believed that these health-promoting compounds are associated with adequate intake of macronutrients and bioactive compounds present in vegetables. Several studies have found that fruits and vegetables are rich sources of antioxidant nutrients and bioactive compounds, including flavonoids, which are involved in neurodegenerative processes ([5]; Huot, 2014). Dietary intake of vitamins E and C for 3 years maintains cognitive abilities and slows down their deterioration. It has been found that levels of vitamin E in food and human blood plasma can reduce the risk of developing Alzheimer's disease (AD). Dietary intake of vitamin C, carotenoids, and flavonoids also reduces the risk of AD. Dietary intake of flavonoids provides a 50% reduction in the risk of dementia ([2,6]; Hung et al., 2004; Shi et al., 2022).

Lettuce is one of the most well-known leafy vegetables in the world, with many uses in both cooking and science. In addition, lettuce is an excellent source of bioactive compounds such as polyphenols, carotenoids and chlorophyll with associated benefits for human health ([3]; Kim et al., 2018). There are many different varieties of lettuce, with colors ranging from green and yellow to deep red, which is due to the different concentrations of chlorophyll and anthocyanins in the leaves ([7,8]; Mou, 2005; López et al., 2014).

Lettuce (*Lactuca sativa* L., family Asteraceae) is one of the most popular vegetables, consumed fresh or in salad mixtures and has high production and economic value ([9,10]; Mampholo et al., 2016; Das et al., 2024). The crop is unpretentious and can be grown both in open and closed ground. In addition, lettuce can be grown in various traditional ways on soil and soilless methods in containers or on vertical rack systems ([11]; Assefa et al., 2021). Consumer interest in lettuce is growing due to its superior visual quality, low calorie content, low disease incidence, and high content of beneficial phytochemicals. According to a report by the Food and Agriculture Organization of the United Nations ([12]; Mekouar, 2023), the production of lettuce in the world increases annually, while it is noted that the main supplier is China (~56%), and in second and third place are the United States (~12%) and India (~4.3%), respectively. Cultivated lettuces come in different types, and can differ in the shape and size of the stalk, stem, and can form different heads: oily, crispy (iceberg or cabbage).

The nutritional or nutraceutical properties of lettuce vary depending on growing conditions and abiotic factors. However, a number of studies show that the variety, i.e. its genetic makeup, also greatly influences the properties of lettuce. For example, romaine and leaf lettuce varieties contain higher amounts of ascorbic acid, vitamin A, carotenoids, and folate, while crispheads contain relatively low amounts of these compounds ([11]; Assefa et al., 2021). Differences in the content of primary and secondary metabolites in red and green lettuce leaves were observed between head and leaf types of lettuce ([13]; Altunkaya et al., 2009). In addition to growing conditions, genetic variation influences the nutritional and phytochemical properties of lettuce ([3]; Kim et al., 2018).

Anthocyanins are a family of naturally occurring flavonoids responsible for variations in the color of leaves, fruits, and flowers of many plants, particularly the red, purple, and bluish colors of lettuce ([10]; Das et al., 2024). Red pigmented lettuce accumulates large amounts of anthocyanins. Anthocyanins are typically present in the form of anthocyanidin glycosides and acylated anthocyanins (Figure 1) ([13]; Altunkaya et al., 2009).

Anthocyanin pigments extracted from plants have been traditionally used as dyes and food colorings to treat various diseases ([14]; Khoo et al., 2017). The most common anthocyanidins in nature are pelargonidin, cyanidin, delphinidin, peonidin, petunidin, and malvidin. Hydroxylation and methylation of the B-ring control the color and stability of anthocyanins. Table 1 lists most of the anthocyanidins and other coloring substances that contribute to the increase in red color under the influence of abiotic factors. Blueness increases with the increase in the number of hydroxyl groups, while redness increases with the increase in methylation in the B-ring ([15,16]; Alappat and Alappat, 2020; Anum et al., 2024).

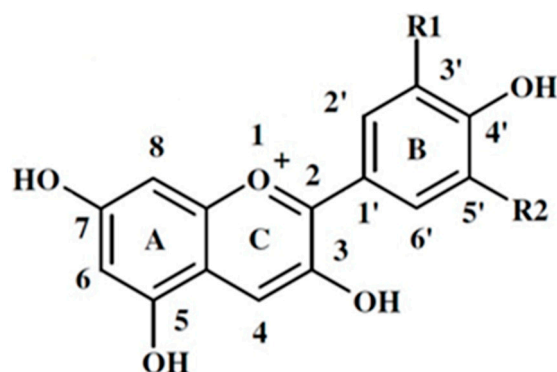


Figure 1. Basic structure of anthocyanidins and anthocyanins with carbon atom numbering.

Table 1. Anthocyanidins, which are precursors of all anthocyanin compounds.

Anthocyanidin	Side radicals of the B-ring		Color	Present in red leaf lettuce
	R ₁	R ₂		
Anthocyanidins				
Cyanidin (Cy)	HE	N	orange red	
Peonidin (Pn)	OSN ₃	N	red	
Pelargonidine (Pg)	N	N	orange	
Malvidin (Mv)	OSN ₃	OSN ₃	purple	
Delphinidin (Dp)	HE	HE	blue red	Cyanidin 3-0-(6'-0-malonylglucoside) – 97% of total
Petunidin (Pt)	OSN ₃	HE	purple	
Carajurin	H	H	-	anthocyanin content
Arrabidin	H	H	-	
5-methylcyanidin	HE	OCH ₃	orange red	([17]; Belwal et al., 2020) or 251-928
Peonidin	HE	HE	orange	mg/kg wet weight
Capensinidin	OH	OCH ₃	blue red	([18]; Colombo et al., 2021)
Eupinidin	OH	OCH ₃	blue red	
Pulchellidin	OH	OCH ₃	blue red	
6-hydroxydelphinidin	OH	OH	blue red	
Aurantidin	OH	OH	orange	
30-hydroxyarrabidine	H	H	blue red	
Tricetinidine	H	OH	red	
6-hydroxycyanidin	OH	OH	red	
Rosinidin	Red	Red	red	
other substances that color plants				
Quercetin				3-0-(6'-0-malonyl)glycoside ([11]; Assefa et al., 2021)
Tartaric acid				2,3 – di-O-caffeoyltartaric acid

There are many studies confirming the fact that anthocyanins participate in the prevention of cardiovascular diseases and have shown a connection between the level of anthocyanins in lettuce and the manifestation of antioxidant, antidiabetic, anticancer, anti-inflammatory, antimicrobial, antitumor, antimutagenic effects ([1,19–21]; Baslam et al., 2013; Petroni and Tonelli, 2011; Morais et al., 2016; Giampieri et al., 2023). However, quantitative data on the main health-promoting metabolites of red pigmented lettuce are clearly lacking.

It was found in early studies that red lettuce had higher total anthocyanin and phenolic content and antioxidant capacity than green lettuce ([22–24]; Ferreres et al., 1997; Stintzing et al., 2002; Zhao et al., 2005). The main difference between red and green lettuce was the anthocyanin content. Mulabagal et al. ([25]; 2010) compared *in vitro* biological activity of green and red lettuce and showed that the aqueous extract of red lettuce has a higher biological activity and contains more anthocyanins compared to green varieties. In addition, the presence of only one major anthocyanin was shown by HPLC analysis of the red lettuce extract, and it was characterized as cyanidin-3-O-6-malonyl- β -glucopyranoside. Subsequent glycosylation results in a red shift in the color of the anthocyanin with increased stability. While 3-glycosides increase stability, 5-glycosides tend to decrease stability. Moreover, attachment of 5-glycosides to anthocyanidin can also lead to the formation of colorless pseudobases since the loss of the hydroxyl group at position 5 makes the anthocyanin more susceptible to hydration reaction. Sugar residues in anthocyanins are often acylated with aromatic or aliphatic acids. Common aliphatic and aromatic acids involved in acylation reactions result in color changes (blue shift) and increased stability due to intra- and intermolecular copigmentation reactions. In addition to the biosynthetic genes involved in anthocyanin pigment formation, vacuolar pH and cell shape have a negative effect on anthocyanin pigments. For example, in petunia flowers, acidification of the vacuole causes a red color change, while mutations affecting pH result in a blue color change. Even with high pigment accumulation, plants may appear colorless due to the shape of their cells. This is due to differences in reflected light between conical and flat cells ([26]; Grotewold, 2006).

It was found that the variety, agricultural practices and growing conditions can change the content of phytochemicals, leaf color and quality of lettuce ([27,28]; Liu and Yang, 2012; Tsormpatsidis et al., 2008). The diversity of physiological characteristics of lettuce tissues, pigment forms and the characteristics of their distribution in cells contribute to the fact that phytochemicals and the antioxidant capacity of a given crop can differ even in the outer and inner leaves of plants. Thus, it was found in ([29]; Viacava et al., 2014) that total soluble solids (TSS) and color values of green and red lettuce cultivars were significantly dependent on leaf position on the plant: internal leaves showed significantly higher levels of PTB than average, and the outer leaves of lettuce are both red and green. The average RTV content in green varieties was significantly higher than that of red varieties. The color of the leaves can also vary depending on their position for each lettuce variety, which occurs as a result of the breakdown of chlorophyll and the accumulation of anthocyanins: the values indicating the lightness of the color gradually decreased from the inner to the outer leaf. Similar patterns were recorded for the indices indicating the color when the leaf color changes from green to red: the outer leaves of green varieties showed lower (more negative) values due to high chlorophyll content with a darker green color, while the outer leaves of red varieties showed higher values of this indicator due to the increased content of anthocyanins with a red color.

The influence of leaf position on antioxidant activity was demonstrated for the first time in ([30]; Ozgen and Sekerci, 2011). The nutrient content of several anthocyanin-rich red and green lettuce varieties was compared and quantified: green and red lettuce varieties showed different characteristics, mainly due to the presence of anthocyanins in red varieties. It was shown that in red lettuce leaves, the outer leaves have the highest content of phytonutrients and antioxidant properties. The results of the studies have practical and scientific value, including the use of the identified patterns in drawing up breeding programs aimed at creating new varieties and developing functional human nutrition systems, as well as for the pharmaceutical industry.

Thus, the food significance and nutritional value of red-leaved plants have been identified and studied quite well. The results of a number of studies ([9,16,31–33]; Mampholo et al., 2016; Anum et al. 2024; Liang et al., 2024; Bunning et al., 2019; Zhao et al., 2007) are schematically generalized in Figure 2, which shows that the individual development of each separate species or variety of lettuce is determined by the genetic characteristics of the individual, which are manifested in the activity of

the main biological processes of plants - respiration, photosynthesis, transpiration, metabolic processes and accumulation of nutrients in tissues and different parts of the plant. In addition, all processes are also under the influence of abiotic and biotic factors that determine their growth and development, the change of phenophases, the rate of seed maturation, and the quality of plant products for various purposes.

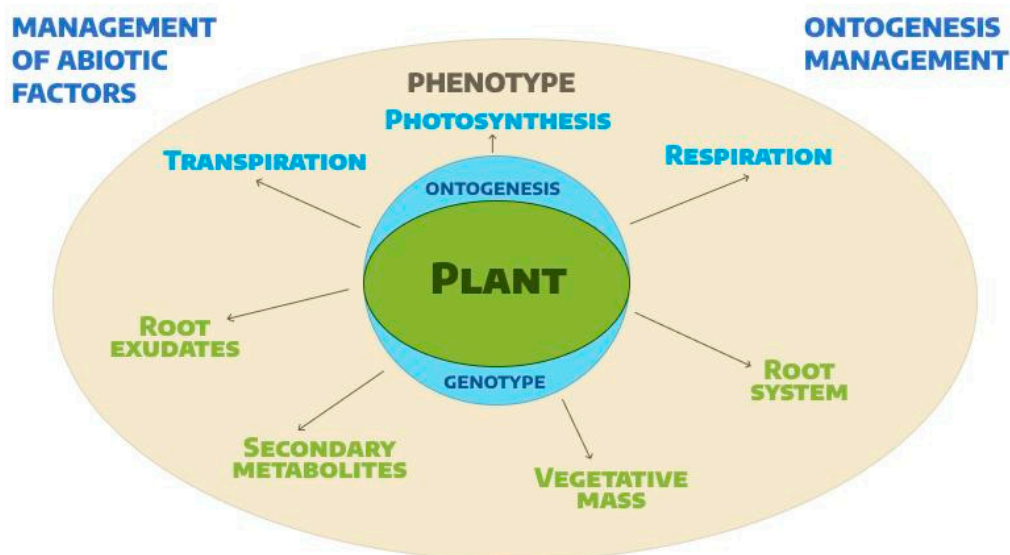


Figure 2. The diagram demonstrates the relationship between environmental conditions and individual characteristics of a plant at all stages of its development: the individual development of each crop of a certain variety or species is determined by the genetic characteristics of the individual, which are manifested in the activity of the main biological processes - respiration, photosynthesis, transpiration, metabolic processes and accumulation of nutrients in tissues and different parts of the plant. In addition, all processes are also under the influence of abiotic and biotic factors that determine the speed and rate of growth and development of plants, the change of phenophases, the rate of seed maturation, and the quality of plant raw materials for various purposes.

2. The Influence of Light on Leaf Color and Antioxidant Activity of Lettuce

It is known that light controls photosynthesis and regulates the morphology, physiology and phytochemical composition of plants. Plants use light as an energy source for photosynthesis, signaling and many other fundamental processes; therefore, primary and secondary metabolism are regulated by the quantity and quality of light entering the leaf blade ([10]; Das et al., 2024).

According to modern concepts, plants have at least five groups of photoreceptors that perceive information not only about lighting conditions and daylight hours, but also about ambient temperature, the presence of pathogens or competing neighbors, the direction of the gravity vector, and other factors ([34,35]; Pierik and de Wit, 2014; Lee et al., 2016). These receptors include red (RL) and far red (FRL) receptors – phytochromes; receptors that perceive ultraviolet A radiation, blue (BL) and green (GG) light – cryptochromes, phototropins, proteins of the ZEITLUPE family; as well as a receptor for ultraviolet B radiation (UVB) – the UVR8 protein ([36]; Briggs and Christie, 2002).

Using light-emitting diodes (LEDs), it is possible to create individual spectra, including a spectrum simulating sunlight, which allows for significant changes in the course of ontogenesis, growth and development of crops. Considering the fact that the spectrum of light can influence the physiological reactions of plants, it is already possible today to influence different stages of plant growth by means of several “fixed” spectra, which makes it possible for dynamic spectral manipulation, including changes in the mechanisms of functioning of light recipes throughout the growth cycle of lettuce Figure 3.

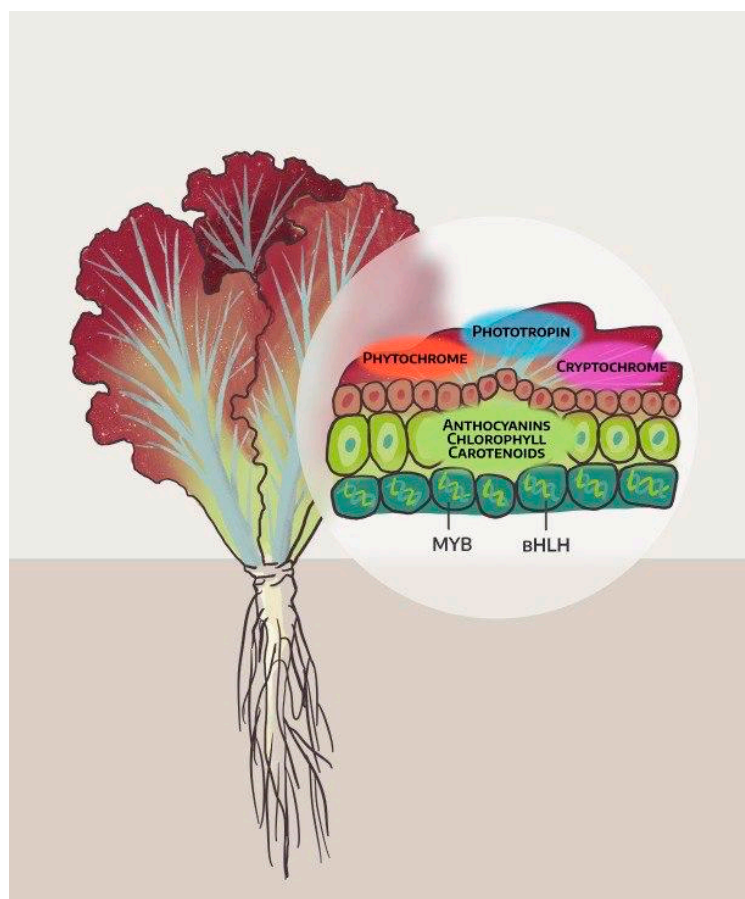


Figure 3. Light-sensitive system of red leaf lettuce, providing different productivity and product quality.

As has been established previously and confirmed by modern methods, plants use photosynthetic pigments in their leaves to capture the photosynthetic active radiation (PAR) to initiate the synthesis of sugar molecules. These photosynthetic pigments are present around the thylakoid membranes of chloroplasts and serve as the primary electron donors in the electron transport chain ([37]; Anderson, 1986). Specifically, photosynthetic pigments absorb light and transfer energy via resonance energy transfer to a specific chlorophyll pair in the reaction center of photosystem II P680 or photosystem I P700. In this light-dependent reaction, water molecules are broken down to generate ATP and NADPH and release oxygen molecules as a byproduct. It has been established that in a subsequent light-independent reaction in the chloroplast stroma, the energy from ATP and electrons from NADPH are used to convert carbon dioxide into glucose and other products via the Calvin cycle.

In ([27,38]; Liu and Yang, 2012; Spalholz et al., 2020) the results of studies on the assessment of growth, development and phytochemical composition of green red and green crops grown hydroponically under different exposition spectra are presented. One of spectra simulated the sun (SUN), and six others were conventional light spectra used in indoor growing systems: 5% ultraviolet-A (UV-A), 20% blue (B), 26% green (G), 26% red (R), and 23% far-red (FR) light as percentage photon flux density (PFD). Fluorescent white light (FL - 6500 K) was used as a control. Plants received $200 \pm 0.7 \mu\text{mol m}^{-2} \text{s}^{-1}$ of biologically active radiation (300-800 nm) for 18 h and were grown at a temperature of $20.0 \pm 0.2 \text{ }^\circ\text{C}$. The results indicate that the dry mass of plants under SUN treatment was not significantly different from that under red-blue light treatment at all harvest times: the dry mass of plants at day 17, the leaf area of lettuce grown under B+R conditions was 15–39% larger than that of those grown under blue and fluorescent white. The leaf area at day 42 was 39–78% larger in the 100B, SUN, and FL treatments than in the B+R treatment. The increase influence of LED-simulated solar resulted in bolting and flowering of lettuce in the SUN treatment. In addition, the authors noted that both total phenolic and anthocyanin concentrations in lettuce leaves were higher

in the B+R treatments than in the SUN, 100R, and 100B treatments. These studies further confirm the basic information on lettuce responses to LED-simulated solar spectrum compared to conventional B+R treatments and provide insight into lettuce growth change and morphology under different spectra.

As the authors note, amid the flood of studies on the effects of light spectra on lettuce growth, photoperiod is a relatively unexplored parameter in studies determining the effect of lighting on indoor leafy greens. This is likely due to the less pronounced effect of photoperiod on growth and nutrient accumulation in leaves compared to the quality and quantity of light received by the plant. It is noted that photoperiods of 14–18 hours of illumination are usually used when growing green crops, which is already an optimal level. Thus, any deviation from this established norm does not bring significant benefits during the vegetative growth of leafy greens. Despite the fact that all tested daylight integrals are only a fraction of full sunlight, leafy greens grow well in these so-called low-light conditions. Yudina et al. ([39]; 2023) developed a model to predict plant productivity to answer the question: does increasing light duration stimulate plant productivity without causing changes in photosynthesis and respiration. The results of the study showed that increasing light duration can stimulate dry weight accumulation and that this effect can also be caused by increasing the photoperiod with decreasing light intensity. Therefore, the authors showed that increasing light duration is an effective approach to stimulating lettuce production under artificial lighting.

As shown by the above studies, higher illumination can lead to higher biomass, but phytonutrient accumulation can be active at lower illumination. Furthermore, electric lighting can represent a significant portion of production costs, and therefore there is little incentive to use high PPFD or long photoperiod in a closed production agroecosystem to make such a system economically viable and environmentally sustainable ([10,40]; Das et al., 2024; Wong et al., 2020).

Every year, more and more research is being conducted in the field of photobiology and photomorphogenesis, but the answer to the question of producers by what mechanisms to obtain a redder or greener lettuce that would simultaneously have increased antioxidant capacity and nutritional value for the consumer has not been received. A number of authors (e.g., [28,41]; Tsormpatsidis et al., 2008; Samuolienė et al., 2011) noted that plant growth, including lettuce, occurs as a result of biomass accumulation following the trajectory of a sigmoid curve. The choice of the plant growth stage for experiments with the light spectrum can significantly affect the reaction, and vice versa, the light spectrum can be used to create the desired morphological characteristics. Lettuce seedlings should be compact plants with a good root system and a large leaf mass area. After rooting of the seedlings, when treated with light emitters, rapid leaf growth occurs to increase light capture ([16]; Anum et al., 2024).

Low light intensity affects the dormancy of the anthocyanin pathway in red-leaf lettuces, and conversely, high light intensity enhances the expression of genes associated with anthocyanin biosynthesis, including regulatory genes (*PAP1* and *PAP2*) and structural genes (*CHS*, *F3H*, *DFR*, and *LDOX*). The accumulation of anthocyanins was negatively affected by switching from fluorescent to LED lighting: the molecules decreased under white and red-blue LED illumination. Red light and UV-C radiation can inhibit the accumulation of anthocyanins or cause their degradation. At the same time, studies have shown that both blue and red light effectively stimulate the formation of anthocyanins in strawberry fruits ([42,43]; Inoue and Kinoshita, 2017; Fylladitakis, 2023).

Thus, lettuce responds positively to changes in the light spectrum and photoperiod, and it is noted that due to these parameters it is possible to change not only the aboveground biomass and morphometry, but also agronomically important traits, such as dry matter, the content of photosynthetic pigments, primarily anthocyanins, and antioxidant activity. In lettuce grown at an increased level of irradiation in the range from 250 to 300 $\mu\text{mol}/\text{m}^2$, an increase in the concentration of anthocyanins was obtained ([28,44]; Tsormpatsidis et al., 2008; Zhang et al., 2018). The effect of spectrum on growth and taste depends on the lettuce variety. By using only red light, it is possible to control the absorption of nutrients, and therefore the quality and taste of the grown produce. Using wavelengths of light emitters in the range from 620 nm to 700 nm has several positive effects on the nutritional value of lettuce: the concentration of ascorbic acid decreases, antioxidant properties increase, and the absorption of N, K, Ca and Mg by the plant root system is stimulated, which immediately affects the accumulation of sugars and other components. Both far-red and blue LEDs have promising prospects for use as supplemental lighting and for controlling quality and growth.

However, the influence of lettuce varietal characteristics on the efficiency of anthocyanin and other nutrient synthesis under modified artificial lighting conditions remains poorly understood ([16]; Anum et al. 2024), which means that studying the effect of photon flux density on the accumulation of anthocyanins is very promising and useful.

3. Effect of Temperature on Leaf Color and Antioxidant Activity of Lettuce

Plant growth and development are influenced by various environmental factors, including light, temperature, CO₂, water availability, and pathogens. External, environmental factors often trigger abiotic and biotic stress responses in plants, including the production of secondary metabolites, which play a key role in stress tolerance. However, additional production of secondary metabolites requires energy expenditure for synthetic processes, and these costs can lead to a decrease in plant growth and development ([11]; Assefa et al., 2021). Mutant plants responding to constitutive biotic stress showed a smaller size and weight of aboveground biomass due to the activation of adaptation reactions and resistance development through the synthesis of salicylic acid, including the production of antimicrobial secondary metabolites ([45]; Blokhina et al., 2003). This process is very important in the development of mechanisms for controlling plant growth processes and color. Accordingly, to increase the production of metabolites by a plant through responses to the environment, it is necessary to take into account most of the abiotic factors that affect plant growth and development, one of which is the ambient temperature of both roots and aboveground biomass. It was found that the optimal temperature for anthocyanin biosynthesis is in the range of 15-30 °C. However, plants can actively synthesize anthocyanins at lower and higher temperatures. For example, under low-temperature stress (4 °C) regulation of the expression of *FaMYB10* and *FaMYB1* genes was noted, which significantly increased the activity of structural genes and led to increased accumulation of anthocyanins in strawberry leaves ([16,46]; Anum et al., 2024; Noda et al., 1994).

Analyzing the data obtained by a number of scientists, it is possible to identify another mechanism for regulating the growth and amount of plant pigments, and minimizing crop losses - by changing the temperature of the root zone of plants (RZT) at different stages of their growth. This suggests that RZT control may be a relevant method, especially in vertical farms and greenhouse complexes where flood tables or a flooding system are used. For instance, Levine et al. ([47]; 2023) showed that exposure of red lettuce roots to low temperature significantly reduced leaf area, stem diameter, and above- and below-ground fresh weight. The results suggest that low-temperature root treatment triggers stress responses throughout the plant, resulting in reduced leaf and root growth.

However, in another study, heating the root zone by heating the solution in the root zone did not result in significant changes in plant biomass ([48]; He et al., 2013). Cooling the root zone to 20 °C increased the biomass of aeroponic lettuce compared to plants under ambient conditions (24–38 °C) in a tropical greenhouse ([49]; Paulsen, 1994).

The production of various plant metabolites is influenced by root zone temperature in many plants. Studies conducted in ([47,50]; Levine et al., 2023; Porter and Gawith, 1997) showed that human-preferred compounds, such as anthocyanin, phenolics, and sugars, increased significantly in red lettuce leaves when their roots were exposed to low temperature. Sugar accumulation was observed in spinach, cotton, and tomato when roots were exposed to low temperature. In contrast, exposure of roots to high temperature did not change the anthocyanin, phenolics, and sugars in leaves. However, increasing soil temperature using electric heating cables suppressed anthocyanin and sugar accumulation in lettuce leaves under field experimental conditions.

Root stress such as drought or salinity causes plant growth limitation followed by a reduction in leaf photosynthetic capacity (see Box 1 in [51]; Allen and Ort, 2001). Reduced plant growth and photosynthesis were observed when plants were exposed to low root temperatures ([52,53]; Akula and Ravishankar, 2011; Bumgarner et al., 2012). Low root temperature treatment resulted in decreased water uptake by lettuce, which resulted in a reduction in photosynthesis in the shoot growth zones. However, sugar content in both lettuce leaves and roots increased following low root temperature treatment, while nitrate concentrations, for example, changed significantly only in leaves where the temperature was reduced to 40% of that in plants under ambient conditions. Considering that photosynthetic nitrate uptake is likely not increased in leaves due to low root temperature stress, the suppression of nitrate transport from roots to leaves may be responsible for the decrease in nitrate concentration in the leaves themselves ([48]; He et al., 2013).

RZT can also significantly affect the plant metabolite contents in both leaves and roots ([53]; Bumgarner 2011). Comparing the results obtained in an experiment on lettuce whose root system was maintained at a solution temperature of 25 °C, the concentrations of 19 amino acids (isoleucine, serine, tyrosine, valine, methionine, leucine, cysteine, phenylalanine, threonine, homoserine, histidine, pyroglutamate, alanine, glutamate, glutamine, ornithine, glycine, β -alanine, and asparagine) were significantly increased in the roots under the 15 °C treatment. At the same time, the morphometry of the leaves was not significantly changed. In contrast, compared with the 25 °C treatment, the 35 °C treatment significantly affected the metabolite contents in both roots and leaves. The roots showed significant increases in seven sugars (glucose, fructose, sucrose, turanose, lactose, ribose, and maltose), 21 amino acids (arginine, phenylalanine, lysine, glycine, histidine, proline, cysteine, isoleucine, threonine, tyrosine, glutamine, valine, asparagine, pyroglutamate, β -alanine, 3-cyanoalanine, leucine, tryptophan, ornithine, serine, and methionine), and five metabolites in the tricarboxylic acid cycle (TCA) (isocitric acid, 2-xoglutaric acid, citric acid, fumaric acid, and malic acid).

In *Arabidopsis*, overexpression of a plasma membrane water channel protein mitigated the low root temperature-induced decline in hydraulic activity and plant growth, suggesting a role for water uptake in root temperature stress. Since drought stress triggers various secondary metabolic pathways, the accumulation of anthocyanins, phenolics, and sugar in leaves in the experiment may be a response to drought stress: a study in green crops showed that drought stress applied to roots can lead to accumulation of phenolics and sugar in hydroponically grown lettuce seedlings ([54,55]; Park et al., 2008; Boo et al., 2011).

It was noted in ([32,47]; Bunning et al., 2019; Levine et al., 2023) that photosynthesis impairment caused by abiotic stress often accompanies oxidative stress. Plants exposed to drought, salinity, and low temperature experience oxidative stress, which is followed by a decrease in photosynthetic capacity and oxygen emission, which is associated with the functioning of cofactors in photosystem II (PSII), and as a consequence, this leads to limitation of plant biomass production. Salinity stress induces oxidative stress in leaves, such as the production of reactive oxygen species (ROS), activation of antioxidant enzymes, and leaf chlorosis. As shown in ([56]; Rivas -San Vicente, 2011), low-temperature treatment of lettuce roots causes the accumulation of hydrogen peroxide in leaves, accompanied by lipid peroxidation. This finding indicated that low-temperature stress in the root zone induces oxidative stress in leaves, presumably by limiting photosynthesis. Plants cope with oxidative stress by producing antioxidant metabolites, including phenolic compounds such as anthocyanin. As a result of stress, anthocyanin and phenolic content in leaves increases when roots are exposed to low temperature, indicating an antioxidant role for these metabolites in response to elevated hydrogen peroxide levels in leaves.

It was found that lettuce exposed to low air temperature at a level of 10-15 degrees showed increased production of anthocyanins and polyphenols, which led to a change in the intense color of lettuce leaves, and also showed an increased ability to remove DPPH radicals. The *Arabidopsis* MYB transcription factor AtMYB60 functions as a transcriptional repressor of anthocyanin biosynthesis genes in lettuce ([32]; Bunning et al., 2019). Given that MYB family proteins are common regulators of the anthocyanin synthesis pathway in many plants ([57]; Chadwick et al., 2024), anthocyanin accumulation in lettuce leaves induced by low root zone temperature is also regulated by intrinsic MYB transcription factors. To enhance the levels of secondary metabolites, regulation of these transcription factors may be important for the production of crops with improved product quality.

It was shown in ([58,59]; Takahashi and Murata, 2008; Sairam et al., 2020) that treatment with a solution with a high RZT contributed to the improvement of pigment content, but negatively affected plant growth. In this experiment, plants were grown in five temperature variants of RZT: 25 °C; HT1 (12 days at 25 °C, then 4 days at 35 °C); HT2 (8 days at 25 °C, then 8 days at 35 °C); HT3 (4 days at 25 °C, then 8 days at 35 °C); and 35 °C at an air temperature of 22 °C. It was observed that an increase in the duration of RZT treatment at 35 °C led to a decrease in the dry weight of shoots and roots, and an increase in the content of chlorophyll, carotenoids, and anthocyanins. In addition, the chlorophyll content increased significantly when treated with RZT at 35 °C for more than 4 days, while the carotenoid and anthocyanin contents increased significantly when treated with RZT at 35 °C for more than 8 days. It was also found that RZT treatment at 35 °C compared with 25 °C limited plant growth in roots and leaves due to decreased root growth, but increased the pigment contents including

anthocyanin, chlorophyll, and carotenoids. This was probably due to the typical response to heat stress. The longer the period of exposure of the root zone to high temperature (35 °C), the more the anthocyanin, carotenoid and chlorophyll contents increased, while the shoot dry weight decreased, indicating gradual plant mortality before harvest. However, increasing the pigment levels may be an effective mechanism for controlling leaf color in lettuce. Plants typically accumulate ROS under stressful conditions, and plant stress tolerance is often associated with increased activity of antioxidant enzymes. The increase in anthocyanins and carotenoids in leaves at RZT 35 °C is due to the formation of ROS in leaves as a response to high-temperature stress and an increase in antioxidants to remove them.

In addition, the authors noted that carotenoids are produced from β -alanine via acetyl-CoA, but the leaves of plants in the 35 °C experiment were enriched in β -alanine, and it is possible that β -alanine metabolism influenced carotenoid accumulation. The root zone temperature significantly affected the concentrations of elements in both leaves and roots. Compared with the 25 °C treatment, plants in the 15 °C treatment experienced a significant decrease in Ca, Mn, As, and Cd in the roots, with a significant increase in P, K, Li, and Rb and a significant decrease in Cu and Cs in the leaves. Compared with the 25 °C treatment, the 35 °C treatment resulted in a significant increase in Fe, Zn, Mo, Cu, Ni, Co, Li, Se, and Cd and a significant decrease in Ca, S, Mn, As, and Sr in the roots. Simultaneously with 35 °C treatment, there was a significant decrease in P, K, Ca, Mg, S, Fe, Mn, Zn, Mo, Cu, Na, Ge, As, Se, Rb, Sr, and Cs in the leaves.

In summary, low and high temperatures in the root zone limit plant growth through various internal mechanisms, and growing at high temperatures in the root zone 4 days before harvesting can slightly reduce the production process, but lead to the accumulation of pigments, which in turn contributes to an increase in the added value of lettuce and color. However, in general, this direction and its practical application remain poorly understood and should be more studied.

4. Effect of pH on Leaf Color and Antioxidant Activity of Lettuce

Anthocyanins are potent antioxidants in plants and maintain optimal cellular redox equilibrium by neutralizing free radicals with their hydroxyl groups to reduce oxidative damage ([60]; Naing and Kim, 2021). Anthocyanins are derived from anthocyanidins, which are structurally based on the flavylium ion (2-phenylchromenylium). This flavylium ion is responsible for the red color of anthocyanins at low pH. At neutral pH, the flavylium cation undergoes deprotonation to form a resonance-stabilized quinoid base, which appears purple. With further increase in pH, the quinoid base can form anionic species that appear blue. The predicted values of the acid dissociation constant (pK_a) for these transitions are about 1–3 for the flavylium cation, 4–5 for the quinoid base, and 7.5–8 for the quinoid monoanion ([14]; Khoo et al., 2017). In addition, the specific color of the anthocyanin also depends on the substituents in the B-ring, with hydroxyl groups tending to shift the color toward blue, and methoxyl groups toward red. For example, delphinidin (with three hydroxyl groups) appears blue, while pelargonidin (with one hydroxyl group) appears red ([15]; Alappat and Alappat 2020).

Anthocyanins are water soluble. However, they exhibit a very interesting chemistry in aqueous solutions, and can present four main interconvertible species with different relative amounts at a given pH (Figure 4).

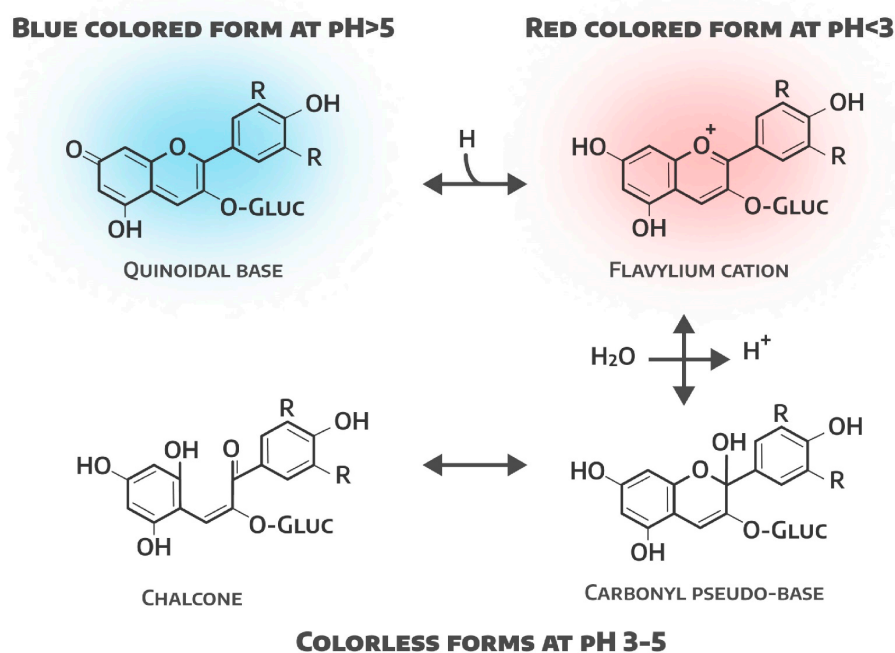


Figure 4. Conversion of anthocyanins into interchangeable forms at the different pH (adapted from Figure 4 in ([61]; Biswas and Mathur, 2017).

At low pH, the flavylium cation is most prominent and has a deep red color. As the pH increases, the anthocyanins are converted to colorless forms such as pseudobases and chalcones, and at pH > 5, the anthocyanin changes to a blue quinoid form. These anthocyanidins are further attached to sugars such as glucose, galactose, and rhamnose via an α/β linkage exclusively at position 3 of the aglycone. Alternatively, they can also be acylated with cinnamic acid, caffeic acid, ferulic acid, malic acid, oxalic acid, and succinic acid, to name a few ([62]; Jung et al., 2014).

It has been established that in an acidic environment, the color of red-leafed lettuces changes due to internal rearrangements of pigments, but in parallel with this, an oppositely directed process occurs, in which the antioxidant activity of the lettuce extract increases with increasing pH, which can be used in practice when growing red-leafed lettuces with given parameters of color and quality.

5. Bioactive Phytochemicals and Metabolites of Lettuce

Lettuce is an important dietary source of bioactive phytochemicals. Screening and identification of health-promoting metabolites and assessing relationships with phenotypic traits may help consumers adjust their preferences for lettuce plant types. Modern metabolomics analysis methods allow the investigation of key health-promoting individual metabolites and antioxidant potential of agricultural plants. Assefa et al. ([63]; 2018) using UPLC-DAD-QTOF/MS (TQ/MS) and UV-visible spectrometry instruments identified and quantified three anthocyanins, four hydroxycinnamoyl derivatives, two flavonols and one flavone in 113 samples of germplasm and commercial cultivars of lettuce at the mature stage. Total phenolic content (TPC) and 2,2-azino-bis-(3-ethylbenzothiazoline-6-sulfonic acid) (ABTS) radical absorbance potentials were estimated, and the relationships between biochemical and phenotypic traits of 113 lettuces were investigated. The metabolite contents varied significantly among the lettuce samples: cyanidin 3-O-(6-O- malonyl) glucoside (4.7-5013.6 $\mu\text{g/g DW}$), 2,3-di-O-caffeoyltartaric acid (337.1-19,957.2 $\mu\text{g/g DW}$), and quercetin 3-O-(6-O- malonyl) glucoside (45.4-31,121.0 $\mu\text{g/g DW}$) were the most dominant in the red pigmented lettuce samples, and hydroxycinnamoyl and flavonol derivatives were the most abundant anthocyanins, respectively. Lettuces with dark to very dark red pigmented leaves, round leaf shape, strong leaf waviness and very tight leaf cuts were found to have high levels of flavonoids and hydroxycinnamoyl derivatives. Key variables were identified that could make a major contribution to plant breeders developing varieties with improved bioactive compounds and to nutraceutical companies developing nutrient-dense foods and pharmaceutical formulations.

Assefa et al., ([63]; 2018) systematized the traits and quality of lettuces grown in the field and laboratory. Morphological traits were assessed based on the modified International Union for the Protection of New Varieties of Plants (UPOV) descriptors for lettuce: leaf length, width, and lettuce plant weight. Other qualitative morphological traits such as cotyledon color, plant growth type, leaf shape, leaf position, leaf blade (degree of edge waviness and density of marginal notches on the apical part), and intensity of red coloration of outer leaves were also investigated. About 76% of the samples contained red color at the cotyledon stage, while the remaining lettuce samples did not contain red color. Another important characteristic was the intensity of red coloration of the outer leaves. The intensity of red coloration was assessed on the outer leaf and ranked from 1 to 5 (very light, light, medium, dark, and very dark, respectively). Most of the samples had medium and light intensity, accounting for 37.2% and 31.9% of the total resources. The remaining 16.8%, 10.6%, and 3.5% of the samples had dark, very light, and very dark red color intensity, respectively. Total phenolic content (TPC) and antioxidant potential were quantified using spectrophotometry. As noted by the authors of the study, significant variations in the radical reduction potentials of TPC and ABTS were observed among genetic varieties ([63]; Assefa et al., 2018).

Llorach et al. ([64]; 2008) reported TPC contents ranging from 18.2 to 571.2 mg/100 g FW and ABTS antioxidant potential ranging from 61.3 to 647.8 mg TEAC/100 g FW for five lettuce and escarole "frisser" cultivars. In another study, TPC contents assessed in five lettuce cultivars ranged from 13,900 to 46,900 μg GAE/g DW; red leaf cultivars showed the highest amounts. Differences were found between green and red lettuce and escarole: caffeic acid derivatives were the main polyphenols in green varieties, while flavonols were observed in higher amounts in red varieties and escarole, and anthocyanins were present only in red-leafed varieties. Moreover, lettuce and escarole showed differences in flavonol composition, as quercetin derivatives were observed only in lettuce samples, while kaempferol derivatives were found only in escarole samples. In general, red-leafed vegetables showed higher levels of both flavonol and caffeic acid derivatives than green lettuce and escarole varieties. Regarding vitamin C, red-leafed lettuces showed higher contents than green salad vegetables, with the exception of the continental variety, which showed the highest levels.

Low phosphorus and low nitrogen often cause anthocyanin accumulation in plants. Under phosphorus and nitrogen limitation, plants adapt to environmental stress by modulating nutrient partitioning and metabolic pathways, including increased anthocyanin synthesis.

Various factors including genotype, cultivar, growth stage and other cultivation conditions influence the total phenolic content and antioxidant potential of lettuce ([3,8,65]; Kim et al., 2018; López et al., 2014; Pérez-López et al., 2014). The phenotypic properties of lettuce showed significant differences in antioxidant capacity and phenolic content. For example, the mean TPC and antioxidant potential of light and very light red pigmented samples were significantly lower than those of medium and dark or very dark samples. Broadly elliptical and round leaf-shaped samples were found to be superior to medium elliptical and broadly ovate leaf shapes. The mean TPC and ABTS values were also significantly higher in highly wavy leaf samples compared to those with medium and slightly wavy leaf margins. The high levels of antioxidants and total polyphenols in the lettuce samples significantly enhance their nutritional value.

Three anthocyanins (cyanidin 3-O-glucoside, cyanidin 3-O-(3-O-malonyl)glucoside, and cyanidin 3-O-(6-O-malonyl)glucoside), four hydroxycinnamoyl derivatives (3-O-caffeoylquinic acid, 5-O-caffeoylquinic acid, 3,5-di-O-caffeoylquinic acid, and 2,3-di-O-caffeoyltartaric acid), two flavonols (quercetin 3-O-glucuronide and quercetin 3-O-(6-O-malonyl)glucoside), and a flavone (luteolin 7-O-glucuronide) were identified and quantified in red pigmented lettuce samples in ([66]; Su et al., 2020). In addition, TPC and antioxidant activity were evaluated. A huge diversity of biochemical traits was observed among the lettuce samples. Cyanidin 3-O-(6-O-malonyl) glucoside, 2,3-di-O-caffeoyltartaric acid and quercetin 3-O-(6-O-malonyl) glucoside were the most dominant in the red pigmented lettuce samples among the metabolite groups.

The genetic potential was explored for the health-beneficial metabolites in baby leaves of 23 diverse lettuce cultivars in ([3]; Kim et al., 2018). The study revealed significant variations in metabolite composition and content among lettuce cultivars, primarily influenced by leaf color. Red-leaf cultivars were notably rich in carotenoids (especially all-E-lutein, all-E-violaxanthin, and all-E-lactucaxanthin), polyunsaturated fatty acids (mainly α -linolenic and linoleic acids), total phenolic content, and antioxidant potential. Cyanidin and other phenolic compounds emerged as the strongest

radical scavengers based on PCA analysis. Additionally, total folate content ranged from 6.51 to 9.73 $\mu\text{g/g}$ (DW), depending on the cultivar. These findings highlight red-leaf lettuce as a nutrient-rich food with a unique phytochemical profile.

Thus, several factors may contribute to the large variations in the phytochemical composition and color of lettuce, including the genotype of the cultivar. The intensity of red color, leaf shape, plant growth type, incision density, and leaf margin waviness have been found to influence the concentration of metabolites. Lettuce plants with high intensity of red color in leaves, round leaf shape, high leaf waviness, and very dense leaf incisions were found to accumulate the highest concentration of flavonoids and hydroxycinnamic acids. Overall, a number of studies have shown that red lettuce may be one of the main dietary sources of antioxidants such as caffeic acid and its derivatives.

6. Genetic Control of Anthocyanin in Lettuces

The molecular genetic basis of anthocyanin biosynthesis has been studied quite thoroughly, which has been greatly facilitated by mutants of various plant species with altered coloration. Anthocyanin biosynthesis, and consequently coloration, is affected by mutations in three types of genes: the first are genes that code for enzymes involved in the chain of biochemical transformations (structural genes); the second are genes that determine the transcription of structural genes at the right time in the right place (regulatory genes); and the third are genes of transporters that carry anthocyanins into vacuoles. It is known that anthocyanins are oxidized in the cytoplasm and form bronze-colored aggregates that are toxic to plant cells ([46]; Noda et al., 1994).

As shown in ([66]; Su et al., 2020), when grown under optimal conditions, wild lettuce typically develops green leaves with intermittent accumulation of anthocyanins near the leaf margin. Stress, such as drought, promotes anthocyanin accumulation, resulting in a noticeable red coloration of wild lettuce leaves. However, some lettuce cultivars have lost this stress response due to loss-of-function mutations in the gene encoding bHLH. On the other hand, some lettuce cultivars develop red leaves when grown under optimal conditions.

Recent studies have shown that light signals regulate anthocyanins through CONSTITUTIVE PHOTOMORPHOGENIC1 (COP1), which is a negative regulator and mediates the degradation of positive regulators of anthocyanins such as PAP1 and PAP2 in *Arabidopsis* or MdMYB1 in apple ([67,68]; Lee et al., 2012; Hong et al., 2016). Most experiments conducted to study light signal transmission are analyzed by excluding light. However, it was shown in ([44,69]; Zhang et al. 2018; Yang et al., 2018) that the COP1/SUPPRESSOR OF PHYA (SPA) complex may not be completely inactivated at low light intensity, suggesting the existence of some other light intensity-induced mechanism in anthocyanin accumulation.

Leaf color is an important factor influencing consumer acceptance of red leaf lettuce, and light significantly influences leaf color ([46]; Noda et al., 1994). It was shown in ([70,71]; de Pascual-Teresa and Sanchez-Ballesta, 2008; Khan and Abbas, 2023) that red leaf lettuce had greener leaf color when grown under controlled conditions at a light intensity of 40 $\text{mmol m}^{-2} \text{s}^{-1}$, whereas increasing the light intensity to 100 $\text{mmol m}^{-2} \text{s}^{-1}$ resulted in lettuce with a redder leaf color. This red color is known to be due to increased anthocyanin accumulation ([72]; Kumar et al., 2022). However, the molecular mechanisms governing light-induced anthocyanins in red leaf lettuce are still not fully understood.

Anthocyanin biosynthesis has been studied quite extensively in *Arabidopsis*, and more than 29 anthocyanin molecules have been identified in *Arabidopsis* ([71]; Khan and Abbas, 2023), which are regulated by strong light alone or in combination with low temperature exposure. However, the regulation of phytochemical biosynthesis and its accumulation, as well as the underlying molecular mechanisms of light-induced phytochemical biosynthesis, are poorly understood in lettuce.

Constitutive up-regulation of anthocyanin biosynthesis is mediated by a gain-of-function mutation in the gene encoding the MYB transcription factor and loss-of-function mutations in two negative regulators ([66,73]; Su et al., 2020; Khusnutdinov et al. 2021). Thus, green-leaf and red-leaf lettuce cultivars have been selected both during domestication and in modern breeding programs, demonstrating typical artificial disruptive selection, i.e., selection favoring extreme values over intermediate values ([6,31]; Shi et al., 2022; Liang et al., 2024). Varieties lacking anthocyanins may have reduced ability to withstand biotic and abiotic stress. On the other hand, varieties with high concentrations of anthocyanins in epidermal cells will block light penetration and, therefore, reduce

photosynthesis and plant growth. But such selection based on green and/or red vegetable leaves is carried out by humans, and this is artificial selection.

Using gene annotation and phylogenetic analysis, putative structural genes involved in anthocyanin synthesis and transport were identified (Table 2 adapted from Table 1 in ([44]; Zhang et al., 2018). NCBI GenBank accession numbers were MF579543–MF579560. The authors of the study identified nine key genes involved in anthocyanin biosynthesis: seven anthocyanin structural genes, including *CHS*, *CHI*, *F3H*, *F3'H*, *DFR*, *ANS*, and *3GT*, and two anthocyanin transport genes, *GST* and *MATE*. Besides of this, six anthocyanin regulatory genes were identified: five MYBs and one bHLH gene.

Table 2. Differentially expressed genes related to anthocyanin.

Gene name	Low-light FPKM	High-light FPKM	Log 2 Fold Change (hiht /low light)	FDR	Up/down
LsCHS					Up
Unigene12000_All	1,68	858.26	9.0	0	
CL4608.Contig2_All	11.36	97.28	3.1	5.33e ⁻²⁵⁹	
LsCHI					Up
Unigene10166_All	10.6	68,72	2.70	1,43 e ⁻⁹⁶	
LsF3H					Up
Unigene8465_All	14.21	408,44	4.85	0	
LsF'3H					Up
CL524.Contig1_All	9.08	193.8	4.42	0	
LsDFR					Up
Unigene2105_All	2,3	473	7.68	0	
LsANS					Up
CL1994.Contig1_All	4.25	269.69	5.99	0	
Ls3GT					Up
CL4808.Contig1_All	5.84	158.28	4.76	0	
CL4808.Contig2_All	3.31	84.6	4.66	0	
LsGST					Up
Unigene10814_All	1.84	245.41	7.06	0	
LsMATE					Up
Unigene12020_All	8.42	18.14	1.11	9.25 e ⁻¹⁵	
LsMYB					
Unigene12430_All	2.21	8.30	1.91	4.36 e ⁻¹⁰	
Unigene12294_All	3.14	37.14	3.56	5.02 e ⁻⁸⁵	Up
Unigene23058_All	0.11	7.25	6.04	5,68 e ⁻¹⁷	
Unigene24751_All	0.56	22.95	5.36	7.05 e ⁻⁸⁶	
CL6440.Contig1_All	0.72	4.60	2.66	9.14 e ⁻¹¹	
LsbHLH					Up
Unigene13011_All	3.64	21.95	2.59	3.37 e ⁻⁸⁴	
LsHY5					Up
Unigene19629_All	3.18	10.42	1.71	3.15 e ⁻⁵	

Anthocyanin biosynthesis in plants is mainly controlled by some structural anthocyanin biosynthesis genes, which are divided into two groups: early biosynthetic genes (EBGs, such as *CHS*, *CHI*, and *F30H*) and late biosynthetic genes (LBGs, such as *DFR*, *LDOX*, *UF 3 GT*, *UGT 75 C 1*, and *3 AT 1*) ([44]; Zhang et al., 2018). EBGs encode key enzymes for the synthesis of precursors common to flavonoids or other phenolics, while LBGs encode enzymes specific to anthocyanins ([17]; Belwal et al., 2020). Thus, it can be concluded that anthocyanin biosynthesis in plants is mainly regulated by the MYB-bHLH-WD40 (MBW) complex ([44,74]; (Zhang et al., 2018; Shi et al., 2023). The *HY5* gene was discovered, which can respond to light signals and regulate the structural genes of anthocyanins. These genes were significantly overexpressed ($\log_2FC = 2.7-9.0$) under high irradiance and were confirmed by quantitative real-time PCR ([44]; Zhang et al., 2018).

Although biosynthesis of anthocyanins is regulated mainly by the MYB-bHLH-WD40 (MBW) complex, it is also greatly influenced by various environmental factors (e.g., light, salinity, drought, cold) and phytohormones (e.g., jasmonate, abscisic acid, and auxin). Advances in understanding these regulatory networks highlight anthocyanins' potential applications in agriculture, horticulture, and the food industry ([73]; (Shi et al., 2023).

7. Practical Recommendations for Selection and Cultivation of Red-Leafed Lettuces

During lettuce growth, whether grown outdoors, indoors or in vertical farms, each leaf of the plant has different levels of exposure to environmental conditions (light, humidity, nutrient uptake and temperature), and the inner leaves are younger than the outer leaves. These factors may influence the distribution of phytochemicals and antioxidant capacity in the above-ground portion of the lettuce. Knowledge of the bioactive content and antioxidant capacity profile of lettuce plants may be of interest to both consumers and the food industry in order to select more suitable leaves or cultivars for salads or other ready-to-eat mixed vegetable dishes with high nutritional value.

The information presented in this review summarizes current research on the influence of various factors on certain plant species and discusses the use of LED lighting in horticulture, crop storage and disease control: it is in these areas that it will be especially important to consider the fact that the value, quality and weight of the grown products are the result of the work of the genotype and abiotic factors, which are either created and controlled by the agronomist or not controlled in the open ground, but used to manage the quality of the grown products. LED lamp control allows to combine significant reduction of energy losses and controlled plant growth, achieving improvement of two main parameters in one action.

The information presented in the review may be potentially useful for advertisers and health care providers to promote and develop a food culture of consuming red leafy plants, particularly lettuce, to increase the levels of pigments with nutritional value (e.g. beta-carotene and anthocyanins) in foods grown on vertical farms or in soil.

8. Conclusions

This review presents a comprehensive analysis of modern approaches to studying the uniqueness and value of red-leaf lettuces, highlighting their practical application and potential for advancing cultivation technologies. Red-leaf lettuces stand out due to their exceptional aesthetic and functional properties, offering a rich source of bioactive phytochemicals such as anthocyanins, flavonoids, and carotenoids. These compounds not only contribute to the vibrant red coloration but also play a significant role in improving human health by boosting immunity and reducing the risks of oxidative stress-related diseases. The ability to manipulate plant traits such as color, taste, size, and productivity by leveraging cultivation technologies underscores the versatility and adaptability of red-leaf lettuce as a crop. The summarized findings are presented in Table 3, which outlines the genetic and environmental factors influencing leaf coloration and phytochemical content.

The cultivation of red-leaf lettuces in controlled environments, such as vertical farms, offers significant advantages for meeting consumer and market demands. Technologies like LED lighting enable precise control over phenotypic traits, allowing growers to optimize color intensity, antioxidant content, and nutritional value. Low root-zone temperatures and carefully tailored light spectra have been shown to enhance the accumulation of anthocyanins and other valuable compounds, while maintaining acceptable levels of plant productivity. These insights are crucial for developing high-value products tailored to the preferences of consumers and the requirements of

industrial partners, such as LLC “Zelen,” which aims to commercialize red-leaf lettuce production in vertical farming systems.

Furthermore, the review emphasizes the importance of promoting a dietary culture that includes green crops, particularly red-leaf lettuces, due to their health benefits and rich phytochemical profiles. Consumers, breeders, and nutraceutical companies are encouraged to focus on key phenotypic traits—such as color intensity, leaf shape, and phenophase transitions—as well as abiotic conditions like temperature, pH, and light spectrum. These factors provide opportunities to enhance the genotype’s potential and produce crops with superior nutrient content and functional qualities.

The prospective studies should integrate these findings into practical agronomy to address both consumer preferences and market demands. As an example of such studies is the research that is planned to be conducted in the City Farming Laboratory of the Gastronomy Institute at Siberian Federal University (Krasnoyarsk, Russia; <https://www.sfu-kras.ru/en/news/26285>, assessed on December 15, 2024). This initiative aims to establish sustainable production technologies for red-leaf lettuces in vertical farming systems, creating innovative solutions for the restaurant industry and promoting functional foods in everyday diets. This approach combines cutting-edge cultivation practices with a deeper understanding of genetic and environmental factors, delivering high-quality products for modern agriculture and gastronomy.

Table 3. The main genetic and environmental factors influencing lettuce color.

Factor	Description of the factor	Effect on coloring	Mechanism of action	Reference
Genetic factors				
Mutations in <i>MYB/bHLH</i> genes	Mutations that result in gain or loss of gene function	Strengthening or weakening of red color	Changes in the transcriptional activity of genes, controlling anthocyanin biosynthesis	([66]; Su et al., 2020)
Gene changes and losses during domestication	Loss of function of wild-type genes during cultivation	Loss of red color, appearance of green varieties	Decreased or stopped synthesis of anthocyanins	([27]; Liu et al., 2012)
Genetic diversity of varieties	Differences between varieties and genotypes	Variation in the content of anthocyanins and phenolic compounds	Genetic predisposition to accumulation of phytochemicals	([64]; Llorach et al., 2008)
Environmental factors				
Light spectrum	Exposure to different light spectra (blue, red, UV-A)	Strengthening or weakening of red color	Regulation of anthocyanin biosynthesis gene expression through photomorphogenesis	([28]; Tsormpatsidis et al. 2008)
Root zone temperature	Changing the temperature of the nutrient solution (low or high)	Enhanced anthocyanin accumulation at low temperature; variable effect at high temperature	Induction of stress reactions, increase in antioxidant synthesis	([47]; Levine et al., 2023)
pH of the environment	Changing the acidity of the nutrient medium	Effect on antioxidant activity and possible color change	Modification of element bioavailability and enzyme activity	([16]; Anum et al. 2024)
Position of the leaf on the plant (ecological)	Inner, middle or outer leaves	Change in color intensity and pigment content	Differences in light exposure and tissue age	([28]; Tsormpatsidis et al. 2008)

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References

1. Baslam, M.; Morales, F.; Garmendia, I.; Goicoechea, N. Nutritional quality of outer and inner leaves of green and red pigmented lettuces (*Lactuca sativa* L.) consumed as salads. *Sci. Hort.* **2013**, *151*, 103-111. <https://doi.org/10.1016/j.scienta.2012.12.023>
2. Hung, H.-C.; Joshipura, K.J.; Jiang, R.; Hu, F.B.; Hunter, D.; Smith-Warner, S.A.; Colditz, G.A.; Rosner, B.; Spiegelman, D.; Willett, W.C. Fruit and Vegetable Intake and Risk of Major Chronic Disease. *J. Natl. Cancer Inst.* **2004**, *96*, 1577-1584. <https://doi.org/10.1093/jnci/djh296>
3. Kim, D.-E.; Shang, X.; Assefa, A.D.; Keum, Y.-S.; Saini, R.K. Metabolite profiling of green, green/red, and red lettuce cultivars: Variation in health beneficial compounds and antioxidant potential. *Food Res. Int.* **2018**, *105*, 361-370. <https://doi.org/10.1016/j.foodres.2017.11.028>
4. Soetan, K.; Olaiya, C.; Oyewole, O. The importance of mineral elements for humans, domestic animals and plants: A review. *Afr. J. Food Sci.* **2010**, *4*(5), 200-222.
5. Huot, B.; Yao, J.; Montgomery, B.L.; He, S.Y. Growth–Defense Tradeoffs in Plants: A Balancing Act to Optimize Fitness. *Mol. Plant* **2014**, *7*(8), 1267–1287. <https://doi.org/10.1093/mp/ssu049>
6. Shi, M.; Gu, J.; Wu, H.; Rauf, A.; Emran, T.B.; Khan, Z.; Mitra, S.; Aljohani, A.S.M.; Alhumaydhi, F.A.; Al-Awthan, Y.S.; et al. Phytochemicals, Nutrition, Metabolism, Bioavailability, and Health Benefits in Lettuce—A Comprehensive Review. *Antioxidants* **2022**, *11*, 1158. <https://doi.org/10.3390/antiox11061158>
7. Mou, B.Q. Genetic variation of beta-carotene and lutein contents in lettuce. *J. Am. Soc. Hort. Sci.* **2005**, *130*, 870–876. <https://doi.org/10.21273/JASHS.130.6.870>
8. López, A.; Javier, G.-A.; Fenoll, J.; Hellín, P.; Flores, P. Chemical composition and antioxidant capacity of lettuce: Comparative study of regular-sized (Romaine) and baby-sized (Little Gem and Mini Romaine) types. *J. Food Compos. Anal.* **2014**, *33*, 39-48. <https://doi.org/10.1016/j.jfca.2013.10.001>
9. Mampholo, B.M.; Maboko, M.M.; Soundy, P.; Sivakumar, D. Phytochemicals and Overall Quality of Leafy Lettuce (*Lactuca sativa* L.) Varieties Grown in Closed Hydroponic System. *J. Food Qual.* **2016**, *39*, 805-815. <https://doi.org/10.1111/jfq.12234>
10. Das, P.R.; Del Moro, D.S.; Givens, S.R.; Armstrong, S.P.; Walters, K.J. Propagation light intensity influences yield, morphology, and phytochemistry of purple-leaf butterhead lettuce (*Lactuca sativa*). *J. Agric. Food Res.* **2024**, *16*, 101210. <https://doi.org/10.1016/j.jafr.2024.101210>
11. Assefa, A.D.; Hur, O.-S.; Hahn, B.-S.; Kim, B.; Ro, N.-Y.; Rhee, J.-H. Nutritional Metabolites of Red Pigmented Lettuce (*Lactuca sativa*) Germplasm and Correlations with Selected Phenotypic Characters. *Foods* **2021**, *10*(10), 2504; <https://doi.org/10.3390/foods10102504>
12. Mekouar, M.A. Food and Agriculture Organization of the United Nations (FAO). *Yearb. Int. Environ. Law* **2023**, *34*(1), yvae031. <https://doi.org/10.1093/yiel/yvae031>
13. Altunkaya, A.; Becker, E.M.; Gökmen, V.; Skibsted, L.H. Antioxidant activity of lettuce extract (*Lactuca sativa*) and synergism with added phenolic antioxidants. *Food Chem.* **2009**, *115*, 163-168. <https://doi.org/10.1016/j.foodchem.2008.11.082>

14. Khoo, H.E.; Azlan, A.; Tang, S.T.; Lim, S.M. Anthocyanidins and anthocyanins: colored pigments as food, pharmaceutical ingredients, and the potential health benefits. *Food Nutrition Res.* **2017**, *61*, 1361779. <https://doi.org/10.1080/16546628.2017.1361779>
15. Alappat, B.; Alappat, J. Anthocyanin Pigments: Beyond Aesthetics. *Molecules* **2020**, *25*, 5500. <https://doi.org/10.3390/molecules25235500>
16. Anum, H.; Li, K.; Tabusam, J.; Saleh, S.A.A.; Cheng, R.-f.; Tong, Y.-x. Regulation of anthocyanin synthesis in red lettuce in plant factory conditions: A review. *Food Chem.* **2024**, *458*, 140111. <https://doi.org/10.1016/j.foodchem.2024.140111>
17. Belwal, T.; Singh, G.; Jeandet, P.; Pandey, A.; Giri, L.; Ramola, S.; Bhatt, I.D.; Venskutonis, P.R.; Georgiev, M.I.; Clément, C.; et al. Anthocyanins, multi-functional natural products of industrial relevance: Recent biotechnological advances. *Biotechnology Advances* **2020**, *43*, 107600. <https://doi.org/10.1016/j.biotechadv.2020.107600>
18. Colombo, R.; Ferron, L.; Papetti, A. Colored Corn: An Up-Date on Metabolites Extraction, Health Implication, and Potential Use. *Molecules* **2021**, *26*, 199. <https://doi.org/10.3390/molecules26010199>
19. Petroni, K.; Tonelli, C. Recent advances on the regulation of anthocyanin synthesis in reproductive organs. *Plant Sci.* **2011**, *181*, 219-229. <https://doi.org/10.1016/j.plantsci.2011.05.009>
20. Morais, C.A.; de Rosso, V.V.; Estadella, D.; Pisani, L.P. Anthocyanins as inflammatory modulators and the role of the gut microbiota. *J. Nutr. Biochem.* **2016**, *33*, 1-7. <https://doi.org/10.1016/j.jnutbio.2015.11.008>
21. Giampieri, F.; Cianciosi, D.; Alvarez-Suarez, J.M.; Quiles, J.L.; Forbes-Hernández, T.Y.; Navarro-Hortal, M.D.; Machì, M.; Casanova, R.d.J.P.; Espinosa, J.C.M.; Chen, X. Anthocyanins: What do we know until now? *J. Berry Res.* **2023**, *13*, 1-6. <https://doi.org/10.3233/JBR-220087>
22. Ferreres, F.; Gil, M.I.; Castañer, M.; Tomás-Barberán, F.A. Phenolic metabolites in red pigmented lettuce (*Lactuca sativa*). Changes with minimal processing and cold storage. *J. Agric. Food Chem.* **1997**, *45*, 4249-4254. <https://doi.org/10.1021/jf970399j>
23. Stintzing, F.C.; Stintzing, A.S.; Carle, R.; Frei, B.; Wrolstad, R.E. Color and Antioxidant Properties of Cyanidin-Based Anthocyanin Pigments. *J. Agric. Food Chem.* **2002**, *50*, 6172-6181. <https://doi.org/10.1021/jf0204811>
24. Zhao, J.; Davis, L.C.; Verpoorte, R. Elicitor signal transduction leading to production of plant secondary metabolites. *Biotechnol. Adv.* **2005**, *23*, 283-333. <https://doi.org/10.1016/j.biotechadv.2005.01.003>
25. Mulabagal, V.; Ngouajio, M.; Nair, A.; Zhang, Y.; Gottumukkala, A.L.; Nair, M.G. *In vitro* evaluation of red and green lettuce (*Lactuca sativa*) for functional food properties. *Food Chem.* **2010**, *118*, 300-306. <https://doi.org/10.1016/j.foodchem.2009.04.119>
26. Grotewold, E. The genetics and biochemistry of floral pigments. *Ann. Rev. Plant Biol.* **2006**, *57*, 761-780. <https://doi.org/10.1146/annurev.arplant.57.032905.105248>
27. Liu, W.K.; Yang, Q.C. Effects of short-term treatment with various light intensities and hydroponic solutions on nitrate concentration of lettuce. *Acta Agric. Scand. Sect. B - Soil Plant Sci.* **2012**, *62*, 109-113. <https://doi.org/10.1080/09064710.2011.580366>
28. Tsormpatsidis, E.; Henbest, R.G.C.; Davis, F.J.; Battey, N.H.; Hadley, P.; Wagstaffe, A. UV irradiance as a major influence on growth, development and secondary products of commercial importance in Lollo Rosso lettuce 'Revolution' grown under polyethylene films. *Environ. Exp. Bot.* **2008**, *63*, 232-239. <https://doi.org/10.1016/j.envexpbot.2007.12.002>
29. Viacava, G.E.; Gonzalez-Aguilar, G.; Roura, S.I. Determination of Phytochemicals and Antioxidant Activity in Butterhead Lettuce Related to Leaf Age and Position. *J. Food Biochem.* **2014**, *38*, 352-362. <https://doi.org/10.1111/jfbc.12060>
30. Ozgen, S.; Sekerci, S. Effect of leaf position on the distribution of phytochemicals and antioxidant capacity among green and red lettuce cultivars. *Span. J. Agric. Res.* **2011**, *9*, 801-809. <https://doi.org/10.5424/sjar/20110903-472-10>
31. Liang, Y.; Weng, X.; Ling, H.; Mustafa, G.; Yang, B.; Lu, N. Transcriptomic Insights into Molecular Response of Butter Lettuce to Different Light Wavelengths. *Plants* **2024**, *13*, 1582. <https://doi.org/10.3390/plants13121582>

32. Bunning, M.L.; Kendall, P.A.; Stone, M.B.; Stonaker, F.H.; Stushnoff, C. Effects of Seasonal Variation on Sensory Properties and Total Phenolic Content of 5 Lettuce Cultivars. *J. Food Sci.* **2010**, *75*, S156-S161. <https://doi.org/10.1111/j.1750-3841.2010.01533.x>
33. Zhao, X.; Iwamoto, T.; Carey, E.E. Antioxidant capacity of leafy vegetables as affected by high tunnel environment, fertilisation and growth stage. *J. Sci. Food Agric.* **2007**, *87*, 2692-2699. <https://doi.org/10.1002/jsfa.3032>
34. Pierik, R.; de Wit, M. Shade avoidance: phytochrome signalling and other aboveground neighbour detection cues. *J. Exp. Bot.* **2014**, *65*(11), 2815-2824. <https://doi.org/10.1093/jxb/ert389>
35. Lee, H.-J.; Ha, J.-H.; Kim, S.-G.; Choi, H.-K.; Kim, Z.H.; Han, Y.-J.; Kim, J.-I.; Oh, Y.; Fragoso, V.; Shin, K.; et al. Stem-piped light activates phytochrome B to trigger light responses in *Arabidopsis thaliana* roots. *Sci. Signal.* **2016**, *9*(452), ra106. <https://doi.org/10.1126/scisignal.aaf6530>
36. Briggs, W.R.; Christie, J.M. Phototropins 1 and 2: versatile plant blue-light receptors. *Trends Plant Sci.* **2002**, *7*, 204-210. [https://doi.org/10.1016/S1360-1385\(02\)02245-8](https://doi.org/10.1016/S1360-1385(02)02245-8)
37. Anderson, J.M. Photoregulation of the composition, function, and structure of thylakoid membranes. *Annu. Rev. Plant Physiol.* **1986**, *37*, 93-136. <https://doi.org/10.1146/annurev.pp.37.060186.000521>
38. Spalholz, H.; Perkins-Veazie, P.; Hernández, R. Impact of sun-simulated white light and varied blue:red spectrums on the growth, morphology, development, and phytochemical content of green- and red-leaf lettuce at different growth stages. *Sci. Hortic.* **2020**, *264*, 109195. <https://doi.org/10.1016/j.scienta.2020.109195>
39. Yudina, L.; Sukhova, E.; Gromova, E.; Mudrilov, M.; Zolin, Y.; Popova, A.; Nerush, V.; Pecherina, A.; Grishin, A.A.; Dorokhov, A.A.; et al. Effect of Duration of LED Lighting on Growth, Photosynthesis and Respiration in Lettuce. *Plants* **2023**, *12*, 442. <https://doi.org/10.3390/plants12030442>
40. Wong, C.E.; Teo, Z.W.N.; Shen, L.; Yu, H. Seeing the lights for leafy greens in indoor vertical farming. *Trends Food Sci. Technol.* **2020**, *106*, 48-63. <https://doi.org/10.1016/j.tifs.2020.09.031>
41. Samuolienė, G.; Brazaitytė, A.; Sirtautas, R.; Novičkovas, A.; Duchovskis, P. Supplementary red-LED lighting affects phytochemicals and nitrate of baby leaf lettuce. *J. Agric. Food Environ.* **2011**, *9*, 271-274. <https://doi.org/10.1134/S1021443709010038>
42. Inoue, S.-i.; Kinoshita, T. Blue Light Regulation of Stomatal Opening and the Plasma Membrane H⁺-ATPase. *Plant Physiol.* **2017**, *174*, 531-538. <https://doi.org/10.1104/pp.17.00166>
43. Fylladitakis, E.D. Controlled LED lighting for horticulture: a review. *Open J. Appl. Sci.* **2023**, *13*, 175-188. <https://doi.org/10.4236/ojapps.2023.132014>
44. Zhang, Y.; Xu, S.; Cheng, Y.; Peng, Z.; Han, J. Transcriptome profiling of anthocyanin-related genes reveals effects of light intensity on anthocyanin biosynthesis in red leaf lettuce. *PeerJ* **2018**, *6*, e4607. <https://doi.org/10.7717/peerj.4607>
45. Blokhina, O.V.; Virolainen, E.; Fagerstedt, K.V. Antioxidants, Oxidative Damage and Oxygen Deprivation Stress: a Review. *Ann. Bot.* **2003**, *91*, 179-194. <https://doi.org/10.1093/aob/mcf118>
46. Noda, K.-i.; Glover, B.J.; Linstead, P.; Martin, C. Flower color intensity depends on specialized cell shape controlled by a Myb-related transcription factor. *Nature* **1994**, *369*, 661-664. <https://doi.org/10.1038/369661a0>
47. Levine, C.P.; Hayashi, S.; Ohmori, Y.; Kusano, M.; Kobayashi, M.; Nishizawa, T.; Kurimoto, I.; Kawabata, S.; Yamori, W. Controlling root zone temperature improves plant growth and pigments in hydroponic lettuce. *Ann. Bot.* **2023**, *132*, 455-470. <https://doi.org/10.1093/aob/mcad127>
48. He, J.; Qin, L.; Lee, S.K. Root-zone CO₂ and root-zone temperature effects on photosynthesis and nitrogen metabolism of aeroponically grown lettuce (*Lactuca sativa* L.) in the tropics. *Photosynthetica* **2013**, *51*, 330-340. <https://doi.org/10.1007/s11099-013-0030-5>
49. Paulsen, G.M. High Temperature Responses of Crop Plants. In *Physiology and Determination of Crop Yield*; 1994; pp. 365-389.
50. Porter, J.R.; Gawith, M. Temperatures and the growth and development of wheat: a review. *European Journal of Agronomy* **1999**, *10*, 23-36. [https://doi.org/10.1016/S1161-0301\(98\)00047-1](https://doi.org/10.1016/S1161-0301(98)00047-1)
51. Allen, D.J.; Ort, D.R. Impacts of chilling temperatures on photosynthesis in warm-climate plants. *Trends Plant Sci.* **2001**, *6*, 36-42. [https://doi.org/10.1016/S1360-1385\(00\)01808-2](https://doi.org/10.1016/S1360-1385(00)01808-2)
52. Akula, R.; Ravishankar, G.A. Influence of abiotic stress signals on secondary metabolites in plants. *Plant Signal. Behav.* **2011**, *6*, 1720-1731. <https://doi.org/10.4161/psb.6.11.17613>

53. Bumgarner, N.R.; Scheerens, J.C.; Mullen, R.W.; Bennett, M.A.; Ling, P.P.; Kleinhenz, M.D. Root-zone temperature and nitrogen affect the yield and secondary metabolite concentration of fall- and spring-grown, high-density leaf lettuce. *J. Sci. Food Agric.* **2012**, *92*(1), 116-124. <https://doi.org/10.1002/jsfa.4549>
54. Park, J.-S.; Kim, J.-B.; Cho, K.-J.; Cheon, C.-I.; Sung, M.-K.; Choung, M.-G.; Roh, K.-H. *Arabidopsis* R2R3-MYB transcription factor AtMYB60 functions as a transcriptional repressor of anthocyanin biosynthesis in lettuce (*Lactuca sativa*). *Plant Cell Rep.* **2008**, *27*, 985-994. <https://doi.org/10.1007/s00299-008-0521-1>
55. Boo, H.-O.; Heo, B.-G.; Gorinstein, S.; Chon, S.-U. Positive effects of temperature and growth conditions on enzymatic and antioxidant status in lettuce plants. *Plant Sci.* **2011**, *181*, 479-484. <https://doi.org/10.1016/j.plantsci.2011.07.013>
56. Rivas-San Vicente, M.; Plasencia, J. Salicylic acid beyond defence: its role in plant growth and development. *J. Exp. Bot.* **2011**, *62*(10), 3321-3338. <https://doi.org/http://dx.doi.org/10.1093/jxb/err031>
57. Chadwick, M.; Swann, J.R.; Gawthrop, F.; Michelmore, R.; Scaglione, D.; Jose-Truco, M.; Wagstaff, C. Mapping taste and flavour traits to genetic markers in lettuce *Lactuca sativa*. *Food Chem. Mol. Sci.* **2024**, *9*, 100215. <https://doi.org/10.1016/j.fochms.2024.100215>
58. Takahashi, S.; Murata, N. How do environmental stresses accelerate photoinhibition? *Trends Plant Sci.* **2008**, *13*, 178-182. <https://doi.org/10.1016/j.tplants.2008.01.005>
59. Sairam, R.K.; Srivastava, G.C.; Saxena, D.C. Increased Antioxidant Activity under Elevated Temperatures: A Mechanism of Heat Stress Tolerance in Wheat Genotypes. *Biol. Plant.* **2000**, *43*, 245-251. <https://doi.org/10.1023/A:1002756311146>
60. Naing, A.H.; Kim, C.K. Abiotic stress-induced anthocyanins in plants: Their role in tolerance to abiotic stresses. *Physiol. Plant.* **2021**, *172*, 1711-1723. <https://doi.org/10.1111/ppl.13373>
61. Biswas, T.; Mathur, A. Plant Anthocyanins: Biosynthesis, Bioactivity and in vitro Production from tissue cultures. *Adv. Biotech. Micro.* **2017**, *5*(5), 555672. <https://doi.org/10.19080/AIBM.2017.05.5556672>
62. Jung, H.; Kwak, H.-K.; Hwang, K.T. Antioxidant and antiinflammatory activities of cyanidin-3-glucoside and cyanidin-3-rutinoside in hydrogen peroxide and lipopolysaccharide-treated RAW264.7 cells. *Food Sci. Biotechnol.* **2014**, *23*, 2053-2062. <https://doi.org/10.1007/s10068-014-0279-x>
63. Assefa, A.D.; Jeong, Y.-J.; Kim, D.-J.; Jeon, Y.-A.; Ok, H.-C.; Baek, H.-J.; Sung, J.-S. Characterization, identification, and quantification of phenolic compounds using UPLC-Q-TOF-MS and evaluation of antioxidant activity of 73 *Perilla frutescens* accessions. *Food Res. Int.* **2018**, *111*, 153-167. <https://doi.org/10.1016/j.foodres.2018.05.017>
64. Llorach, R.; Martínez-Sánchez, A.; Tomás-Barberán, F.A.; Gil, M.I.; Ferreres, F. Characterisation of polyphenols and antioxidant properties of five lettuce varieties and escarole. *Food Chem.* **2008**, *108*, 1028-1038. <https://doi.org/10.1016/j.foodchem.2007.11.032>
65. Pérez-López, U.; Pinzino, C.; Quartacci, M.F.; Ranieri, A.; Sgherri, C. Phenolic Composition and Related Antioxidant Properties in Differently Colored Lettuces: A Study by Electron Paramagnetic Resonance (EPR) Kinetics. *J. Agric. Food Chem.* **2014**, *62*, 12001-12007. <https://doi.org/10.1021/jf503260v>
66. Su, W.; Tao, R.; Liu, W.; Yu, C.; Yue, Z.; He, S.; Lavelle, D.; Zhang, W.; Zhang, L.; An, G.; et al. Characterization of four polymorphic genes controlling red leaf color in lettuce that have undergone disruptive selection since domestication. *Plant Biotechnol. J.* **2020**, *18*, 479-490. <https://doi.org/10.1111/pbi.13213>
67. Lee, S.H.; Chung, G.C.; Jang, J.Y.; Ahn, S.J.; Zwiazek, J.J. Overexpression of PIP2;5 Aquaporin Alleviates Effects of Low Root Temperature on Cell Hydraulic Conductivity and Growth in *Arabidopsis*. *Plant Physiol.* **2012**, *159*, 479-488. <https://doi.org/10.1104/pp.112.194506>
68. Hong, Y.; Yang, L.-w.; Li, M.-l.; Dai, S.-l. Comparative analyses of light-induced anthocyanin accumulation and gene expression between the ray florets and leaves in chrysanthemum. *Plant Physiol. Biochem.* **2016**, *103*, 120-132. <https://doi.org/10.1016/j.plaphy.2016.03.006>
69. Yang, X.; Wei, S.; Liu, B.; Guo, D.; Zheng, B.; Feng, L.; Liu, Y.; Tomás-Barberán, F.A.; Luo, L.; Huang, D. A novel integrated non-targeted metabolomic analysis reveals significant metabolite variations between different lettuce (*Lactuca sativa*, L.) varieties. *Hortic. Res.* **2018**, *5*, 33. <https://doi.org/10.1038/s41438-018-0050-1>

70. de Pascual-Teresa, S.; Sanchez-Ballesta, M.T. Anthocyanins: from plant to health. *Phytochem. Rev.* **2008**, *7*, 281-299. <https://doi.org/10.1007/s11101-007-9074-0>
71. Khan, R.A.; Abbas, N. Role of epigenetic and post-translational modifications in anthocyanin biosynthesis: A review. *Gene* **2023**, *887*, 147694. <https://doi.org/10.1016/j.gene.2023.147694>
72. Kumar, V.; Sugumaran, K.; Al-Roumi, A.; Shajan, A. De-novo transcriptome assembly and analysis of lettuce plants grown under red, blue or white light. *Scie. Rep.* **2022**, *12*, 22477. <https://doi.org/10.1038/s41598-022-26344-2>
73. Khusnutdinov, E.; Sukhareva, A.; Panfilova, M.; Mikhaylova, E. Anthocyanin Biosynthesis Genes as Model Genes for Genome Editing in Plants. *Int. J. Mol. Sci.* **2021**, *22*, 8752. <https://doi.org/10.3390/ijms22168752>
74. Shi, L.; Li, X.; Fu, Y.; Li, C. Environmental Stimuli and Phytohormones in Anthocyanin Biosynthesis: A Comprehensive Review. *Int. J. Mol. Sci.* **2023**, *24*(22), 16415. <https://doi.org/10.3390/ijms242216415>

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