

Review

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Review

CNTs as Excellent Adjuvants for Anticancer Therapeutic Strategies and Cancer Diagnosis: A Plethora of Laboratory Studies versus Few Clinical Trials and Still No Clinical Application

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Abstract: Encouraging discoveries and excellent advances in the fight against cancer have led to innovative therapies such as photothermal therapy (PTT), photodynamic therapy (PDT), drug targeting (DT), gene therapy (GT), immunotherapy (IT) and therapies which combine these treatments with conventional chemotherapy (CT). Anyway, 2,041,910 new cancer cases and 618,120 cancer deaths have been estimated in the United States for the year 2025. The low survival rate (<50%) and poor prognosis of several cancers, despite aggressive treatments, are due to therapy-induced secondary tumorigenesis and the emergence of drug resistance. Moreover, serious adverse effects and/or great pain, usually arise during treatments and/or in survived individuals, thus lower the overall effectiveness of these cures. Although prevention is of paramount importance, novel anticancer approaches are urgently needed to address these issues. In the field of anticancer nanomedicine, carbon nanotubes (CNTs) could be of exceptional help, due to their intrinsic unprecedented optical, thermal, electronic, mechanic features, easy functionalization and large surface allowing excellent drug loading to serve as drug carriers, as well to engineer multifunctional platforms associable with diverse treatments for both anticancer therapy and diagnosis. The present review debates the most relevant advancements about the adjuvant role which CNTs could have in cancer diagnosis and therapy, when associated with PTT, PDT, DT, GT, CT and IT. Numerous sensing strategies utilizing CNTs-based electrochemical, colorimetric, plasmonic biosensors, as well as immunosensors for cancer diagnosis have been disserted in detail, never forgetting the still not fully clarified toxicological aspects existing by their use in biomedicine. The unsolved challenges to be faced to make possible the translation of CNTs-based material in clinic have been discussed, to push scientists to focus on the development of advanced synthetic methods and purification work-up procedures, thus achieving more perfect CNTs for their safer real-life clinical use.

Keywords: carbon nanotubes (CNTs); CNT-improved anticancer therapies; CNT-improved cancer diagnosis; photothermal therapy (PTT); photodynamic therapy (PDT); gene therapy (GT); immunotherapy (IT)

1. Introduction: Carbon Nanotubes in Cancer Therapy

Cancer is considered the nastiest disorder of the past decades, whose rate of mortality is at worrying levels [1]. It represents the major societal, public health, and economic problem in our century. According to International Agency for Research on Cancer (IARC) and GLOBOCAN, which estimated the incidence and mortality worldwide for 36 cancers in 185 countries in 2022, cancer caused almost 16.8-22.8% of deaths deriving from noncommunicable diseases (NCDs) [2], where NCDs are intended as diseases that are not transmissible directly from one person to another. NCDs killed at least 43 million people in 2021, equal to 75% of non-pandemic-related deaths globally (World

Health Organization (WHO), Noncommunicable diseases, available online at <https://www.who.int/news-room/fact-sheets/detail/noncommunicable-diseases>. Accessed on 26 May 2025). Specifically, NCDs include Parkinson's disease, autoimmune diseases, strokes, heart diseases, diabetes, chronic kidney disease, osteoarthritis, osteoporosis, Alzheimer's disease, cataracts, and cancer among others (Non-communicable disease – Wikipedia, available online at https://en.wikipedia.org/wiki/Non-communicable_disease. Accessed on 26 May 2025). Three out of ten global premature deaths from NCDs are caused by cancer and the 30% of individuals prematurely dead were aged 30–69 years [3]. Globally, 20 million new cases of cancer, including nonmelanoma skin cancers (NMSCs) have been observed in the year 2022, which were responsible of 9.7 million deaths[3]. Lung cancer was the most frequently diagnosed cancer in 2022, followed by cancers of the female breast (11.6%), colorectum (9.6%), prostate (7.3%), and stomach (4.9%). Lung cancer was also the leading cause of cancer death, with an estimated 1.8 million deaths (18.7%), followed by colorectal (9.3%), liver (7.8%), female breast (6.9%), and stomach (6.8%) cancers [2]. Only in the United States, 2,041,910 new cancer cases and 618,120 cancer deaths have been estimated for this year (2025). Demographics-based predictions indicate that the number of new cases of cancer will reach 35 million worldwide by 2050. Cancer represents a notable obstacle to life expectancy, causing substantial societal and macroeconomic costs, which can depend on cancer types, geography, and gender [4]. This detrimental condition is often characterised by an uncontrolled development of cancer cells without inhibition, which invasively spread to other organs and tissues in whole the body [5]. As a direct consequence, the physiological functions of normal cells are impaired and the health and quality of life of the affected individuals are totally compromised [6,7]. Investments in prevention, mainly regarding the tentative to limit/reduce the key risk factors for cancer, such as smoking, overweight, obesity and infection, could avert millions of future cancer diagnoses and save many lives worldwide, leading to a massive economic and societal improvement in countries over the approaching years [2]. Meanwhile, the current state of cancer therapy has greatly improved owing to the rapid development of treatment modalities, such as surgery, chemotherapy, radiotherapy, endocrine therapy, immunotherapy, phototherapy, and gene therapy. Unfortunately, these promising therapeutic strategies have still to face numerous obstacles that limit their widespread use, particularly for the treatment of metastatic cancer and cancer cells which have acquired resistance [8,9]. On this scenario, consisting of a severe and lethal disease currently difficult to treat and a limited or poorly efficient arsenal to limit its global damage, new strategies to counteract cancer are urgently needed. In the recent year bio nanoparticles (NPs) including single, centre/shell, and multi-material composite nanomaterials, in terms of inorganic/organic, inorganic/inorganic, organic/organic, and organic/inorganic composites [10–19] have become a cutting-edge cancer treatment option [20–23]. Among nanomaterials, carbon nanotubes (CNTs), discovered in 1991 by Ijima[24], belong to the fullerene (carbon allotropes and C60) and graphene family. In their pristine form, they are made of sp²-bonded carbon atoms, which confer them a structure encompassing planar graphite sheets, which roll up forming tubes with a diameter of a few nanometres and a variable length, decidedly greater in size (microns) than diameters [25]. As reported, CNTs comprise multiwalled carbon nanotubes (MWCNTs), which contain numerous concentric tubes sharing a common axis (Figure 1a), and single-walled carbon nanotubes (SWCNTs), which are made of a single graphene sheet 'rolled' into a tube (Figure 1b) [25,26]. Also, two models of MWCNTs are reported including the Russian Doll model, in which sheets of graphite are arranged in concentric SWCNTs cylinders (Figure 1a, left), and the Parchment model, in which a single sheet of graphite is rolled in around itself, resembling a scroll of parchment or a rolled up newspaper (Figure 1a, right).

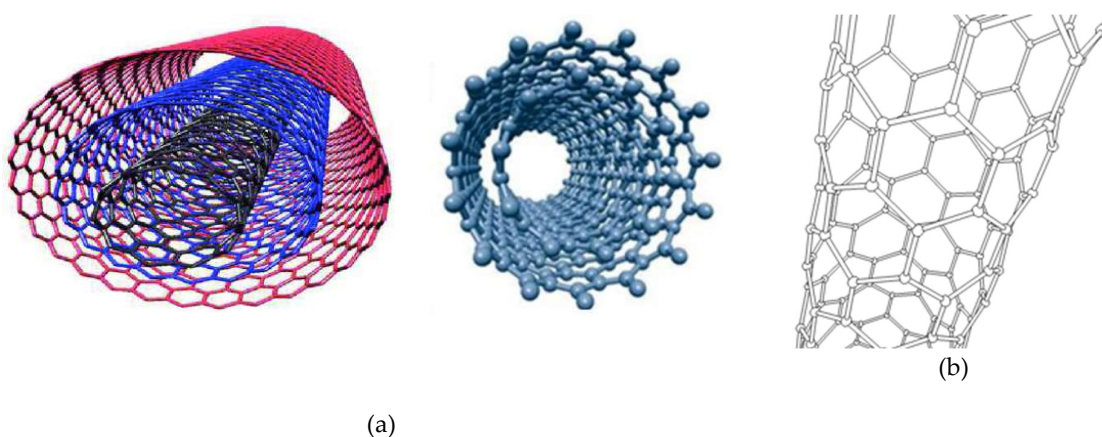


Figure 1. (a) Russian Doll model (left) [27] and (b) Parchment model of a MWCNT (right) [28]. (c) Structure of a SWCNT. The left image in panel (a) has been reproduced by an article available under the Creative Commons CC-BY-NC-ND license which permits non-commercial use of the work as published, while the right image in panel (a) and that in panel (b) are by an unknown author, and are licensed under CC BY-SA 3.0 which not requires permissions [25,26].

Depending on their length, internal diameter, chirality or rotational conformations different CNTs can exist [26]. CNTs are characterized by great surface area, mechanical resistance, thermal conductivity, photoluminescence, transparency, constructional durability and remarkable electrical conductivity[29]. Nowadays, CNTs have a wide-ranging of possible application and are increasingly inserted in several nanocomposites, such as thin-film transistors, transparent conducting electrodes, photovoltaics, supercapacitors, printed electronics, e-readers, flexible displays, conductive and/or waterproof paper, catalyst supports, nano-porous filters, coatings. CNTs are also considered in the I/R optics industry, and in the food industry for nanomaterial-improved food packaging [5,25,26]. Also, CNTs find several applications in nanomedicine as biosensors, DNA based sensors, piezoelectric and gas sensor [30], and are applied for nanomaterials-improved tissue engineering and energy storage medical treatments[25]. CNTs have great potential in nanomedicine for disease diagnosis and are capable to transport various biomolecules, such as proteins, DNA, RNA, immune-active compounds, and lectins, thus being promising new drug delivery systems and gene therapy [31–33]. Additionally, cationic CNTs have demonstrated interesting antibacterial and antifungal activity [25,30]. SWNTs have influenced a significant amount of activity in both research and industry across the world. Furthermore, the increasing application of SWNTs have stimulated considerable investment in ameliorating manufacturing methods, characterization and purification techniques, to obtain more precise CNTs without defects and impurities, thus reducing their toxicity and improve the development of safer applications. The main recognized drawback of CNTs consists of their scarce solubility in most solvents, which limits their dispersibility in water and their use in nanocomposites manufacture [29]. To address this issue, several surface modifications have been carried out to increase their hydrophilicity and decrease toxicity[29,34,35]. In addition to advanced synthetic methods, highly efficient purification procedures and post-synthesis chemical modifications, preventive behavioural conducts have already been developed to limit CNTs' toxic effects[25]. Anyway, more and incessant studies should be implemented to make their large-scale production safer and to allow a no-risk extensive utilization. Despite CNTs can be considered the most relevant and valuable nanomaterial ever used until now, the knowledge of their nanotoxicology is still limited. Nowadays, CNTs are receiving substantial interest in cancer therapy as theragnostic [36], photodynamic therapy (PDT) sensitizer [37], and are very promising materials for uses in photothermal treatment (PTT), due to their significant absorption in near-infrared (NIR) areas [5]. CNTs are advantageous for cancer diagnosis and imaging because they can convert laser energy into acoustic signals and display strong resonant Raman scattering and photoluminescence within the NIR range [38]. Many authors have reported that CNTs can enter different kind of cells due to their

needle-like structure, which promotes efficient tumour penetration [39]. CNTs can also easily reach the different constituents of tumour microenvironment (TME), thus irreversibly disturb the living conditions of tumour cells, and allow the arrival of therapeutic compounds to various intracellular destinations, including nuclei, mitochondria, cytoplasm, for synergistic anticancer effects[5]. CNTs are evolving in many biomedical devices, including genetic engineered ones, and are applied in imaging, and bio sensing to treat and diagnose various cancers [40–42]. Due to their exclusive structural characteristics, CNTs demonstrated great hydrophobicity in water, which confer them intrinsic cytotoxic properties to tumour cells [43]. Functionalization and/modification of pristine CNTs with various chemical groups or biomolecules, not only has demonstrated to ameliorate their hydrophilicity, solubility, dispersibility and to reduce toxicity to normal cells, but has shown to be crucial in bettering their effectiveness for widespread and safer use in cancer therapy[44]. Anyway, other several factors, including their chemical composition, target cells and environmental reactions, can affect the CNTs' biological effects[45].

Table S1, in Supplementary Materials, reports some structural properties demonstrated by several modified and/or activated CNTs, compared with those of unmodified ones (first row).

In this paper, we have debated the most relevant advancements about the adjuvant role which CNTs could have in cancer diagnosis and therapy, when associated with photothermal therapy (PTT), photodynamic therapy (PDT), drug targeting (DT), gene therapy (GT), chemotherapy (CT) and immunotherapy (IT). Numerous sensing strategies utilizing CNTs-based electrochemical, colorimetric, plasmonic biosensors, as well as immunosensors for cancer diagnosis have been disserted in detail, never forgetting the still not fully clarified toxicological aspects existing by their use in biomedicine. The unsolved challenges to be faced to make possible the translation of CNTs-based material in clinic anticancer practice, have been discussed, revealing the existence of only few clinical trials and no clinical use. The main scope of this review was to provide scientists with a huge, updated information on the high potential of CNTs if applied in cancer therapy, to push them to focus on the development of advanced synthetic methods and purification work-up procedures, thus achieving more perfect CNTs for their safer real-life clinical use.

2. Current Synthetic Methods to Obtain CNTs

Table S2 (Supplementary Materials) includes the main conventional methods extensively explained in our previous works [25,26,46]. Other less used methods to produce CNTs have been extensively described in Alfei et al. 2022 [26] and listed in Alfei et al. 2025[25,46]. Collectively, all available methods described in Table S2 (Supplementary Materials), use precursor gases such as CH₄ and C₂H₂, which derive from coal and petroleum raw materials and Fe, Co and Ni NPs as catalysts. Therefore, the major limitations of CVD processes consist in the use of not renewable carbon source and in the large emission of waste gases such as toluene[47], thus worsening energy shortages and environmental pollution[48]. Aiming at developing new environmentally friendly, safe, inexpensive methods to produce CNTs, renewable raw materials as precursors to prepare CNTs, such as biomass, seem to be a promising option to the use of unrennewable gases[46]. Table S3 (Supplementary Materials) collects several examples of CNTs prepared using biomass as an eco-sustainable source of carbon [46]. Operating conditions, types of catalysts, methods as well as the CNTs types obtainable and their physical characteristics have been also reported in Table S3 [46].

3. Carbon Nanotubes as Delivery Systems for Anticancer Drugs

CNTs have been extensively experimented as nano-vectors for the transport and delivery of various agents, including contrast media and drugs. Drug-based, nucleotide-based, and plasmid-based CNTs complexes have been developed and have been tested in vitro and in vivo, against several typologies of cancer. Also, being intrinsically cytotoxic mainly for their needle-like structure, which allows their easy penetration of cells membranes, CNTs associated with existing cytotoxic agents in drugs/carrier complexes, were essayed to assess possible synergistic effects. Hybrid

nanocomposites based on CNTs have demonstrated great potential for realizing cancer combination treatments, when associated with different current available techniques, including photothermal therapy (PTT) and sonodynamic therapy (SDT). CNTs are studied for both early detection of tumorigenic cells and for treating enhanced metastatic cancers. Cirillo et al., realized the pH dependent selective delivery of MTX to H1299 cancer cells, thus lowering the damage to healthy MRC-5 cells [41], while MWCNTs loaded with Pt-NPs and polybenzimidazole (PBI) caused cycle arrest in breast cancer stem cells (CSCs), thus diminishing drug resistance and impeding DNA repair [49]. CNTs embedded with Span, PEG, folic acid (FA) and paclitaxel were capable to easy penetrate breast cancer cells, thus inhibiting their development, inducing tumor cells death [50]. Behazadpour, et al., reported that the administration of 250 mg/mL of MWCNTs loaded with poly pyrrole to C540 Male Balb/c mice reduced tumor cells viability under 9%, under multi-step ultrasonic irradiation. 8.9% viable cells presenting 75% necrosis and 50% tumor volume reduction (TVR) were observed after 10 days of SDT[51]. 6.4 times higher accumulation of CREKA peptide in tumor tissues were observed when the peptide was administered to mice embedded in MWCNTs, with xenograft eradication after 4 cycles of illumination [52]. Also, higher toxicity to tumor cells were observed when PEG-O-CNTs, O-CNTs, as well as pure CNTs were administered to both tumor cells and female mice, under continuous-wave NIR laser diode (808 nm) for 10 min, with higher TVR in animal group administered with PEG-O-CNTs [53].Table 1 schematically summarizes the above-mentioned papers where various CNTs-based nanomaterials have been used as drug carriers in vitro and in vivo to treat several forms of cancer in association with PTT and SDT.

Table 1. Examples of in vitro and in vivo applications of CNTs-based nanocarriers associated with different types of anticancer therapies.

CNTs	Measure s	Cell Line/Anima l Models	Drug(s)	Concentration/Dos e	Results	Ref s
MWCNT s	L = 110– 980 nm	H1299	MTX	7.72×10^{-5} – 1.51×10^{-3} mg/mL	pH dependant selective delivery of MTX to H1299 Low damage to healthy MRC-5	[41]
MWCNT s	L = 330 nm Ø = 30 nm	BM-MSCs, MDA-MB- 231 CD44high, CD24low CSCs	Pt-NPs, PBI	100 µM	Cause cell cycle arrest, diminish drug resistance Impede DNA repair in breast CSCs.	[49]

CNTs	N.R.	Kunming mice	Span, PEG, FA Paclitaxel	350 mg/kg	Penetrate breast tumours Inhibited development, induced tumour cells death	[50]
SWCNTs	N.R.	HT-29	Porphyrin , PEG	25 mg/mL	No antitumour activity	[54]
MWCNTs	Ø = 17.1±3.0 nm	C540 Male Balb/c mice	Poly pyrrole	250 mg/mL, 10 mg/kg	Concentration-dependent cytotoxicity under multi-step ultrasonic irradiation (8.9% cell viability for 250 mg/mL) 75% Necrosis and 50% TVR after 10 days of SDT	[51]
MWCNTs	Ø = 10 nm L = 5–15 µm	Male Balb/c mice	CREKA	MWNTs-PEG 2 mg/kg CMWNTs-PEG 4 mg/kg	6.4 Times ↑ accumulation in tumour tissue Xenograft eradicated after 4 cycles of illumination	[52]
MWCNTs	Ø = 5–20 nm L = 1–10 µm	HepG2, HeLa cells C57BL/6J female mice	PEG-O-CNTs	0–1000 mL 1 mg/mL injected in tumour at 200 µL/cm ³	↑Cell toxicity of PEG-O-CNTs, O-CNTs, pure CNTs Continuous-wave NIR laser diode (808 nm) for 10 min ↑TVR in animal group (PEG-O-CNTs)	[53]

L = Lenth; Ø = diameter; PEG-O-CNTs = oxidised CNTs conjugated to polyethylene glycol (PEG); FA = folic acid; Pt-NPs; PBI = polybenzimidazole; MTX = methotrexate; SDT = sonodynamic therapy; TVR = tumour volume reduction; PTT = photothermal treatment; ↑ = higher; CSCs = cancer stem cells; BM-MSCs = bone marrow mesenchymal stem cells (BM-MSCs); Pt-NPs = platinum nanoparticles; CREKA peptide = linear polypeptide composed of 5 amino acids arranged by Cys-Arg-Glu-Lys-Ala. 3.1. Carbon Nanotubes-Based Drug Delivery Systems for Target Anticancer Therapy.

Cancer diseases are usually treated using chemotherapy associated with surgery and radiation, with good achievements in terms of tumor mass reduction and its expansion limitation, but nonspecific drug release and toxic side effects to normal cells can promote drug resistance and restrain the therapeutic window[55]. Moreover, other limitations are the susceptibility of drugs towards enzyme degradation and denaturation, which can alter their in vivo efficacy (Figure 2).

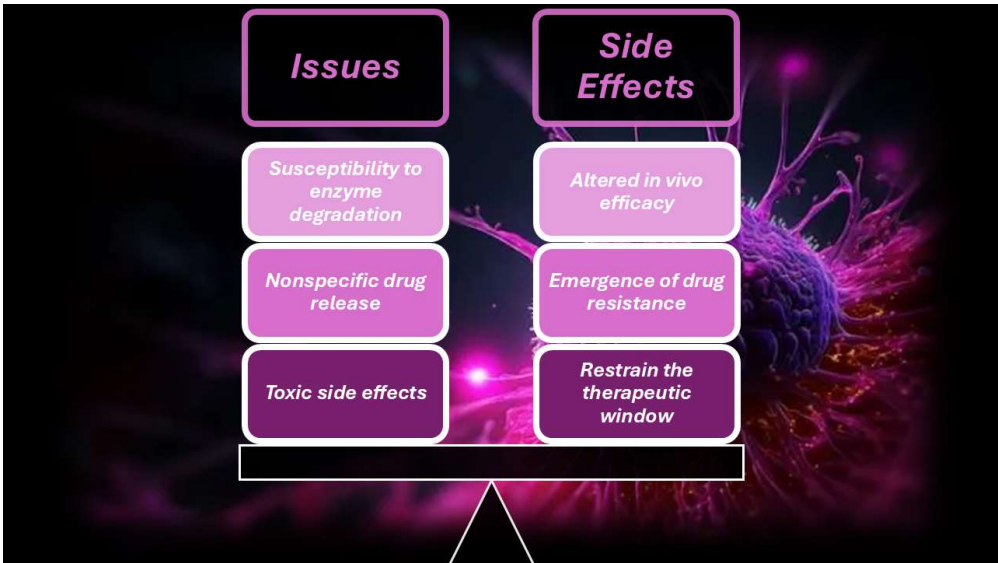


Figure 2. Limitations and associated undesired side effects of chemotherapy for treating tumors.

To limit such issues and improve therapeutic efficacy of chemotherapeutics, while reducing their toxicity towards normal cells and enhancing the patient’s compliance with chemotherapy, new techniques for specific drug targeting have emerged[56,57]. Specifically, nanotechnology provided several nanomaterial-based delivery systems (DSs), which demonstrated high potential to solve most of these issues. Nanomaterial-based DSs can accumulate near tumor sites, thanks to enhanced permeation and retention (EPR) effects, due to more penetrable blood vessels and the missing lymphatic drainage in tumor tissue[58]. When in vivo administered, drugs enveloped into nanomaterials result protected from the degradative environment, while the nanocarriers could be capable to provide drugs release in a controlled and protracted mode [59]. Among nanomaterials, CNTs possess unparalleled properties, such as large surface area and high aspect ratio, thus allowing high loading capacity and being ideal candidates for efficient drug delivery[60]. Also, CNTs showed the capability to realize pH-dependent sustained drug release, enhanced cellular internalization, as well as demonstrated high stability and possibility of post-synthesis modification to further enhance their original properties [60]. Table 2 summarizes the most relevant functionalizing moieties, which have been used to functionalize/modify pristine CNTs and the applications of obtained CNTs derivatives in cancer therapy.

Table 2. Functionalization of CNTs through various molecules and their applications as anticancer agents.

CNTs	Functionalizing molecules	Effectiveness	Tumor model	Biocompatibility test	Refs.
SWNTs	PEG	↑Solubility, prevent particle aggregation, ↓side effects	Gastric cancer	↓Toxicity towards normal tissue	[61]
SWNTs/MWNTs	PEI	↑Solubility, homogeneity, dispersity, ↓particle size ↑Positive charge to interact with siRNA	Cervical cancer	N/A	[62]
SWNTs	HA	↑Stability in serum, overcome MDR Target CD44-overexpressing cancer cells	Ovarian cancer	No cytotoxicity to normal tissue No drop in mice weight	[63]
MWNTs	Chitosan	↑Solubility, ↑cell-penetrating ability, ↓toxicity	Breast cancer	↓Cytotoxicity	[64]
MWNTs	PLGA	↑Dispersity, ↓toxicity, provide, tune the temporal release Provide attachment sites for drugs	Osteosarcoma	↓Cytotoxicity in normal cells	[65]
SWNTs	CD33 mAB	Recognize and specifically target the GNM-CD33 ⁺ cells	Glioblastoma	N/A	[66]
SWNTs	IGF1R mAB/HER2 mAB	Target IGF1R and HER2 surface receptors	Breast cancer	↓↓↓In vitro toxicity in normal cells	[67]
SWNTs	RGD peptide	Target α _v β ₃ -expressing cancer cells	MM	↓Toxicity in vitro	[68]
SWNTs	EGF receptor	Active targeting ability, ↑uptake of drugs	HSC, NSC	↓↓↓Toxicity in vitro and in vivo	[69]

MDR = Multidrug resistance; MM= malignant melanoma; HSC = head squamous carcinoma; NSC = neck squamous carcinoma; enhanced, improved, high, higher; low, lower, decreased, reduced; PEG = polyethylene glycol; PEI =polyethylene amine; IGF-1R = insulin-like growth factor receptor 1R; WS = water solubility; HER2 = human endothelial receptor 2; RGD = arginyl-glycyl-aspartic acid; EGF = epidermal grow factor.The following Figure 3 schematizes the structure of a -COOH polyethylene glycol (PEG) and polyethylene imine (PEI) modified SWCNTs-based nanocarrier, developed by Yang et al., and the doxorubicin (DOX)-loading process [70].



Figure 3. (A) Schematic representation of -COOH modified SWCNT-PEG-PEI nanocarriers and the DOX-loading process [70]. The image has been reproduced by a paper published in diamond open access under the CC BY 4.0 licence (<https://creativecommons.org/licenses/by/4.0/>, accessed on 24 May 2025), which permits you to copy and redistribute the material in any medium or format for any purpose, even commercially.

Yang et al. observed that SWCNT@PEG@PEI@COOH DSs displayed higher anti-cancer effects against MCF-7 cells and better drug delivery under acidic conditions, than CNT@COOH and CNT@PEG. Flow cytometry and fluorescence experiments evidenced improved internalization of SWCNT@PEG@PEI@COOH DSs and enhanced tumor cell death via apoptotic mechanisms, due to their high dispersibility and greater affinity towards cancer cells [70]. Similarly, Liu et al. prepared PEG-functionalized SWCNTs loaded with DOX, which demonstrated a loading capacity of ~400% by weight, and a controlled release rate of cargo [71]. SWCNTs generally exhibited a higher drug loading capacity than MWCNTs[72], while short SWCNTs and MWCNTs showed higher and faster loading capacity than long CNTs[73]. Furthermore, aromatic-peptides showed high binding affinity to SWCNTs, due to their interaction with the π electrons of their surface[74]. A foremost advantage of using CNTs-based nanocomposites to develop cancer drug DSs (CDDSs) consists of their susceptibility to release drugs specifically in the acidic environment existing in tumor tissues, thus allowing selective passive targeting of the tumor site, with reduction of side effects to normal cells. Ph-sensitive release of anticancer drugs from CNTs CDDSs increased drug residence time in circulation and selectivity, decreased administration frequency, with possible major compliance of cancer-affected patients to therapy, while preserved the optimum drug concentration[75]. In this regard, Cao et al. demonstrated that PEI and hyaluronic acid (HA) modified MWCNTs designed for the targeted delivery of DOX to cancer cells overexpressing CD44 receptors, showed a drug loading capacity of 72% and a higher release rate in acidic pH (5.8 pH in cancer) than in physiological conditions (pH 7.4 in normal cells)[76]. Therefore, PEI@MWCNT@HA@DOX nanocomposite showed good biocompatibility in the tested concentration range, while exercised substantial cytotoxic effects to cancer cells (Figure 4).

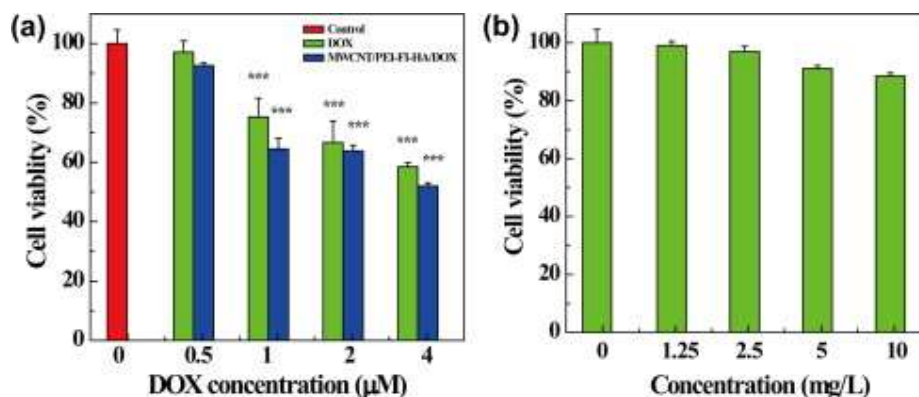


Figure 4. (A) MTT viability assay of HeLa cells treated with free DOX and MWCNT/PEI-FI-HA/DOX complexes at the DOX concentrations of 0–4 μ M for 24 h, and (B) DOX-free MWCNT/PEI-FI-HA at corresponding DOX concentrations of the complexes between 1.25 and 10 mg/L. Reprinted with permission from Carbohydrate Research, Copyright 2015, Elsevier [76]. License number 6018200526333 released on 29 April 2025 by Elsevier and Copyright Clearance Centre.

Similar results were reported by Gu et al., for SWCNTs modified with benzoic acid via hydrazine bonding (HBA) loaded with DOX when tested on HepG2 cells[77]. The release to cancer cells occurred at the pH 5.5 of tumor microenvironment (TME). Additionally, after 60 h of incubation, SWCNT@HBA@DOX complex demonstrated a higher release of DOX than SWCNT@DOX composite (73% drug release vs only 50%). Also, higher cytotoxic effects were observed for SWCNT@HBA@DOX than for SWCNT@ DOX complex, due to enhanced cellular internalization. Since cancer cells require high amounts of folic acid for DNA synthesis and rapid proliferation, they overexpress folate receptors. Conjugation of DDSs with folic acid (FA) is a common approach for targeting cancer cells[78]. Lu et al. functionalized a magnetic nanocomposite composed of MWCNTs and Fe₂O₃ nanoparticles (IONPs) with poly (acrylic acid) (PAA) by free radical polymerization (FRP) and the obtained nanocomposite was further conjugated to FA and loaded with DOX[79]. When utilized on U87 human glioblastoma cells, the nanocomposite exhibited a dual-targeting effect via both magnetic field and ligand-receptor interaction[79]. The nanocomposite demonstrated higher efficiency than DOX alone, due to easier hydrogen bonding interactions and π - π stacking, while exhibited enhanced cytotoxic effects[79]. The higher effectiveness of DOX when delivered by the CNTs-based DDS derived also by its efficient internalization inside the cells and transport to the nucleus, which permitted the intracellular release of DOX[79]. The following Tables 3 and 4 collect various reported experiments which used CNTs for the target delivery of different anticancer drugs and genetic material. Specifically, Table 3 summarizes several in vitro and in vivo relevant experiments on the delivery of anti-tumor drugs and/or nucleic acids using modified CNTs-based nanocarriers, while Table 4 collects results about the cytotoxicity to different tumors and biocompatibility results concerning several modified CNTs-based anticancer DDSs.

Table 3. In vitro and in vivo experiments which used modified CNTs as drug and gene delivery systems for various chemotherapeutics and nucleic acids.

Drug/gen e	Modified	Cancer type	In vitro	In vivo	CNTs	Therapeutic Outcome	Refs
Pt (IV) drug, PD	PL-PEG F	Testicular cancer	Ntera-2	N.R.	SWCNT	25 Folds \uparrow cytotoxicity	[80]
CP	Covalent amide linkage	Head, neck cancer	HNCC	N.R.	SWCNT	\uparrow Effective than free CP and siRNA conjugate	[69]
CA	Interior filling	Bladder cancer	EJ28	N.R.	SWCNT MWCN T	Controlled release properties	[81]
OP	PEG600 F	Colorectal cancer	HT29	N.R.	MWCN T	Delayed cytotoxic activity	[82]
DOX	PEGylation F	N.R.	Ascites Sarcoma 180	N.R.	SWCNT	\uparrow Retention of drug in situ \downarrow Effect on other tissues	[83]

PTX	Not-F	Breast cancer	MCF-7	N.R.	MWCN T	↑ Cytotoxicity	[84]
	P-gp F	Leukaemia	K562	N.R.	SWCNT	↑Cellular uptake 23 folds ↑ cytotoxicity	[85]
	PEGylated F	Lung cancers	A549, NCI-H460	N.R.	SWCNT	↑ PTX activity	[86]
		Ovarian cancer	OVCAR 3	N.R.	SWCNT	Chemosensitizer, ↑cell death	[87]
	Poly citric acid F	Lung, ovary cancer	A549, SKOV3	N.R.	MWCN T	↑Potency	[88]
SB-T-1214, TAXO	PEGylated F	N.R.	N.R.	Murine 4T1	SWCNT	↓↓↓Tumour growth 10 folds ↑ uptake	[89]
	Biotin F	Lung cancer, leukaemia	L1210FR, L1210 W138	N.R.	SWCNT	↑ Target due to biotin	[90]
	Folic acid F	Breast cancer	MCF-7 cells	N.R.	MWCN T	↑Cytotoxic than free GEM	[91]
GEM	PEGylated F	Pancreatic cancer	N.R.	BxPC-3-B/c		↓Metastatic lymph nodes	
HIF-1α siRNA	Cationic F	N.R.	N.R.	MiaPaCa 2/HRE MM	SWCNT	↓↓↓Tumour growth	[92]
Anti-EGFP siRNA	PEI and Pyridinium F	Lung cancer	H1299	N.R.	MWCN T	↑Cytotoxicity	[93]
Cyclin A2 siRNA	Amine (NH ₃ ⁺) F	Leukaemia	K562	N.R.	SWCNT	↓Cell proliferation and apoptosis	[94]
PLK 1 siRNA	NH ₃ ⁺ F	Lung cancer	Calu6	LCXGM	MWCN T	↓↓↓Tumour	[95]
siNEG siRNA	Amidation with PUT SPERD, SPER	Lung cancer	A549	N.R.	SWCNT MWCN T	↓Tumour proliferation	[96]
Bcl91 siRNA	Aptamer and Pyr-PEI F	Breast cancer	MDA-MB-231	N.R.	SWCNT	Targeted silencing activity	[97]

GAPDH siRNA	PEI F	Cervical cancer	HeLa	N.R.	SWCNT	Induced cell death	[62]
siTOX, siNEG siRNAs	Cationic DD F	Cervical, lung cancer	HeLa, A549	N.R.	MWCN T	↓Toxicity, effective delivery Gene silencing capabilities	[98]
PLK-1 & siTOX siRNAs	NH ₂ -F or cationic liposomes	Breast, cervical, lung, melanoma , prostate, renal, kidney cancers	Calu6, A549, DU145 C-33A, MCF-7, HEK293 HeLa, NIH 3T3 B16F10	LCXGM	MWCN T	Apoptosis, ↓↓↓tumour	[99]
PLK1 siRNA	NH ⁴⁺ and Guanidium DD	Cervical cancer	HeLa	N.R.	MWCN T	Significant silencing of PLK-1	[100]
hTERT siRNA+ NIR TPP	PEI F	Prostate cancer	PC-3	TBM	SWCNT	Severe apoptosis, ↓↓↓ tumour	[101]
pDNA	Cationic glyP F	Cervical cancer	HeLa	N.R.	SWCNT	Biocompatible, ↑ transfection *	[102]
p53 plasmids	Ethylenediamin e F	Breast cancer	MCF-7	N.R.	SWCNT	Induced apoptosis	[103]
pDNA	Ammonium F	Cervical cancer	HeLa, CHO	N.R.	SWCNT & MWCN T	First reported gene carrier	[104]
GFP gene	PAMAM	Cervical cancer	HeLa	N.R.	MWCN T	↑ GFP gene transfection	[105]
dsDNA	Multi-F	Epithelial cancer	Mouse ovarian epithelia l	N.R.	SWCNT	Targeted drug delivery	[106]
miRNA	PEI-g-GNR	Cervical cancer	HeLa	N.R.	MWCN T	New nonviral vector for in situ detection	[107]
dsODN- NF-kB	Carbodiimide F	Cervical cancer	HeLa	N.R.	SWCNT	Targeted successful deliver	[108]
ANT c- myc ODNs	PAMAM	Breast, liver cancer	MCF-7, MDA-MB-435, HepG2	N.R.	MWCN T	↑↑↑Gene efficiency	[109]

MUC-1 Apt	Carboxylate F	Breast, lung cancer	MCF-7, Calu-6	N.R.	MWCN T	Aptamer successful integration	[110]
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↑ = High, higher, improved, enhanced; ↓ = low, lower, decreased; reduced; ↑↑↑ = strongly enhanced; ↓↓↓ = strongly reduced; suppressed; N.R. = not reported; CP = Cisplatin; CA = carboplatin; OP = oxaliplatin; DOX = doxorubicin; PTX = paclitaxel; TAXO = taxoid; GEM = gemcitabine (GEM); EGPF = enhanced green fluorescent protein; CyA2 = cyclin A2; PLK 1 = polo-like kinase 1; GAPDH = glycerinaldehyd-3-phosphat-dehydrogenase; hTERT = human telomerase reverse transcriptase; pDNA = plasmid DNA; GFP = green fluorescent protein; dsDNA = double stranded; dsODN = double-stranded oligodeoxynucleotides; NF-κB = nuclear factor ‘kappa-light-chain-enhancer’ of activated B-cells; MUC-1 = mucin-1 protein; ANT = antisense; F = functionalization; * = lipofectamine 2000; PAMAM = poly-amidoamine; glyP = glycol-polymer; PEI= poly-ethylenimine; DD = dendron; Pyr = pyrazine; SPER = spermine; SPERD = spermidine; PUT = putrescine; PD = prodrug; LCXGM = xenografted mice; B/c = BALB/c mouse; TBM = tumour bearing mice; MM = mouse model.

Table 4. Improved biocompatibility and anticancer efficiency of several modified CNTs-based carriers used to deliver in vitro and in vivo anticancer drugs and nucleic acids to treat various tumours.

CNTs	Functionalised molecules	Effectiveness	Tumour model	Biocompatibility test	Refs.
MWCNTs	FA	Targeted and pH sensible release of DOX	N.R.	N.R.	[111]
SWCNTs	PEG-10–10%PEI/pDNA	Allowed dosage 58-fold ↓ conventional IC ₅₀ ↑WS, ↑biological fluids dispersity	AGS and L929 *	↓Toxicity	[61]
SWCNTs	PEG-PCL and PEG-PCL-PEI	Faster acidic DOX release; ↑anticancer efficacy, ↑WS	MCF-7 *	↑Biocompatibility	[112]
SWCNTs	ALG, CHI, FA, DOX	Precise magnetic field-dependent CNTs vibration causing cancer cells membranes destruction and DOX release	Lung cancer *	↑Biocompatibility	[113]
SWCNTs	COOH-, PEG-PEI- DOX	Faster acidic DOX release, ↑Antitumor activity ↑Dispersibility, ↑Affinity for tumour cells.	MCF-7 *	↓Toxicity to normal cells	[70]
SWCNTs	PEI-BET-DOX and SUR siRNA	↑↑↑pH-responsive lysosomal escape of siRNA ↑Antitumor properties	HeLa, A549 *,**	↓Toxicity to normal cells	[114]
SWCNTs	Chim/PEI/5-FU/CNT	↑↑↑Tissue penetrability, ↑↑↑DL, CT allowed ↓Invasion and proliferation, apoptosis in 5-FU-RCCs	Gastric cancer *	↑Biocompatibility	[115]
SWCNTs	II-NCC	↑↑↑SWCNTs water dispersion; ↑ Anticancer effects of CAP	Caco-2 *	↑Biocompatibility	[116]

MWNTs	iRGD-PEI-MWNT-SS-CD/pAT ₂	↑WS and complexation capacity by PEI, ↓↓↓ Tumour growth Target smart release in TRE by SS	A549 *,**	↑Biocompatibility [117]
SWCNTs	PEG and PEI	↑WS, ↑ Cytotoxicity	AMJ13 *	↓Cytotoxicity to normal cells [118]

* In vitro experiments; ** in vivo experiments; DL = drug loading; SS = cystamine; FA = folic acid; PEG = polyethylene glycol; PEI = polyethylene imine; pDNA = plasmid DNA; PCL = polycaprolactone; ALG = sodium alginate; CHI = chitosan; DOX = doxorubicin; BET = betaine; SUR = surviving; siRNA = silencing RNA; Chim = chimera; 5-FU = 5-fluorouracil; II-NCC = N, carboxy cellulose II; iRGD = tumour-homing peptide; CD = candesartan; pAT₂ = plasmid AT₂; CAP = capecitabine; ↑ = improving, improved, augmented, high, higher; ↓ = low, lower, decreased; reduced; ↑↑↑ strongly enhanced, increased; suppression; WS = water solubility; TRE = tumour reducing environment; CT = combination therapy; RCCs = resistant cancer cells. 3.2.1. Carbon Nanotubes-Based Drug Delivery Systems for the Tumor Microenvironment (TME)- and Cite Specific-Target Cancer Therapy.

To effectively manage and precisely remove malignant tumours, a true understanding of cancer features and the interchanges between tumour cells and their surrounding microenvironment is necessary. As extensively demonstrated in the previous sections, many anticancer strategies focus on the target delivery of chemotherapeutics to tumour cells, to enhance their selectivity and anticancer efficiency, while reducing toxic effects to normal cells. The emergence of cancer nanomedicine has allowed huge advancements in the treatment of cancer and in the targeted cancer therapy and several types of nanomaterial-based drug delivery systems have been developed for anticancer target delivery. Among nanomaterials, carbon nanomaterials including CNTs, and mainly functionalized CNTs, have demonstrated to be versatile multifunctional platforms very promising for innovative cancer treatment (Figure 5).

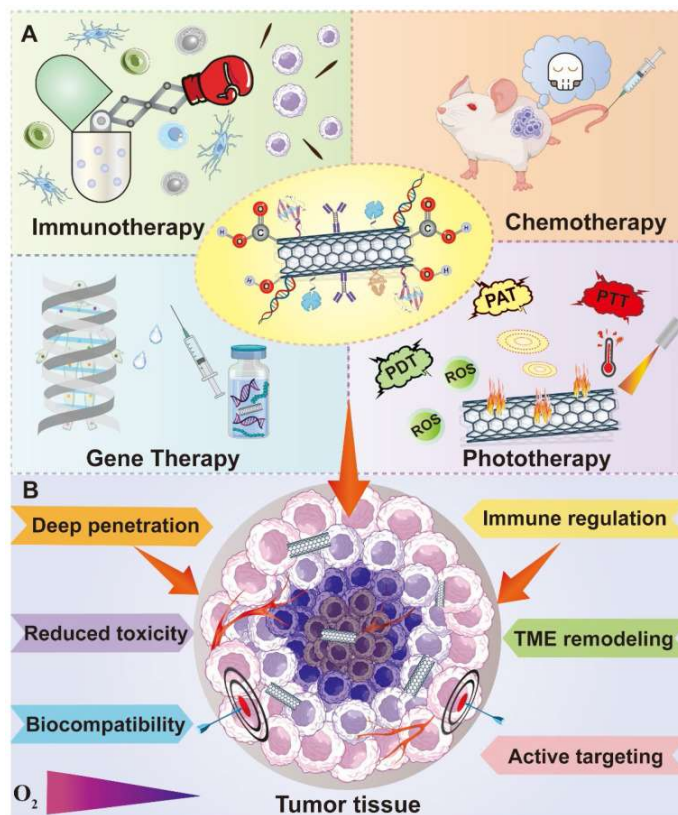


Figure 5. Advantages of CNT functionalization and multiple applications of functionalized CNTs in cancer treatment by different therapeutic approaches. PAT = photo-acoustic tomography; PTT = photothermal therapy; PDT = photodynamic therapy; ROS = reactive oxygen species; TME = tumour microenvironment. The image has been reproduced from an open access article distributed under the terms of the Creative Commons Attribution License CC BY 4.0 licence (<https://creativecommons.org/licenses/by/4.0/>, accessed on 26 May 2025). See also <http://ivyspring.com/terms> for full terms and conditions, accessed on 25 May 2025 [119].

As schematized in Figure 5, CNTs have been successfully experimented in association with many treatment modalities like chemotherapy, gene therapy, phototherapy and immunotherapy, and resulted efficient tools to develop CNTs-based nanocarriers, capable to deliver various anticancer agents to the intracellular sites of interest in cancer cells, such as nucleus, mitochondria, cytoplasm and other organelles, thus realizing direct tumoricidal effects [8,120]. A promising current approach in cancer therapy regards targeting the components present in the tumour microenvironment (TME) where tumour cells live, thus detrimentally impacting on cells survival by impairing their living environment[121,122]. TME is a very complicated system stuffed with various types of cells and full of extracellular matrix (ECM), characterized by altered vasculature, higher acidity and interstitial pressure, abundant glutathione (GSH) levels, poor blood perfusion, abnormal metabolism and hypoxia[119]. Collectively, TME is essential for tumour establishment and progression and represents the obstacle that limits the efficacy of numerous cancer treatment approaches[123]. Anyway, despite the immunosuppressive properties of TME promoting the tumour cells resistance to immunotherapy and despite it produces a lot of impediments to effective cancer treatment, TME can be also exploited as therapeutic target, where abnormalities can be exploited for the development of new selective anticancer strategies. In this context, many innovative anticancer approaches, such as ECM modulation, angiogenesis and cancer stem cells (CSCs) inhibition, immunoregulation, and TME-responsive controlled drug delivery, aiming at remodelling and impairing TME, have been widely studied (Figure 5). CSCs inhibition strategy has been experimented by Faraj et al., in the attempt to address the depressing failures of many currently adopted treatments in the cure of breast

cancer. They designed multimodal nanoplatforms based on SWCNTs to achieve noninvasive imaging and specific targeting towards breast CSCs[124]. To this end, SWCNTs were modified with PEG and, since CD44 is a surface marker of breast CSCs, PEGylated SWCNTs were conjugated with CD44 antibodies, thus realizing active targetability toward breast CSCs. Magnetic resonance imaging (MRI), single photon emission computed tomography (SPECT) and NIR fluorescence techniques evidenced an enhanced selective tumor targeting phenomenon in MDA-MB-231 tumor-bearing mice. Moreover, it was observed that PEG@SWCNTs@anti CD44 distributed specifically in the tumor sites where CD44 receptors are abundant, thus further confirming an elevated targetability to CSCs. As previously reported, the immunosuppressive power of TME, often prevents the immune system from effective tumor eradication. Aiming at addressing this problem, Hassan et al. employed MWCNTs as DDSs for the co-delivering of immunoadjuvants such as CpG, anti-CD40 Ig, and OVA antigen, for enhanced immunotherapy. The covalent conjunction of OVA and CpG remarkably elevated the responses of OVA-specific T cells both in vitro and in C57BL/6 mice, while the subsequent loading of anti-CD40 Ig amplified the antitumor immune reactions. Using MWCNTs the co-loading ability of the three ingredients was improved translating into a significant inhibition of tumor growth and metastasis in OVA-expressing B16F10 melanoma model. Also, Tang et al. engineered a co-delivery platform based on PEI-functionalized MWCNTs to inhibit angiogenesis in lung cancer TME for its treatment[117]. Integrin $\alpha_v\beta_3$ is strictly correlated with angiogenesis, thus representing a therapeutic target for anticancer therapy. RGD peptides are capable to bind with integrin $\alpha_v\beta_3$, thus being a targeting ligand for anticancer drug delivery[125]. Fu and co-workers connected iRGD peptide and candesartan, an angiotensin receptor blocker, to PEI-modified MWCNTs, and assembled the achieved nanocomposite with plasmid angiotensin II type 2 receptor (pAT₂) through electrostatic interaction. A multifunctional DDS was achieved, which demonstrated to be capable of successfully delivering candesartan and pAT₂ into tumor cells, thus significantly inhibiting tumor growth and neovascularization in A549 lung cancer model. Table 5 and 6 report several examples of CNTs based DDSs which demonstrated capability to deliver the transported material to specific intracellular sites and TME constituents.

Table 5. Some examples of application of CNTs-based DDSs to deliver the transported material to specific intracellular sites and TME constituents.

CNTs	TT/modulation	DDS	TM	Model	Effectiveness	Refs.
SWNTs	Cytoplasm	SWNT-PS/siRNA	PTT, GT	Hela	↑ Gene transport capacity, ↑ antitumor activity Controlled gene release	[126]
	Cytoplasm	SWNT-CY7-IGF-1Ra	PTT, IT	ASPC-1, BXPC-3 PANC-1 SW1990 *	Precise tumor-targeting therapy, ↑ body weight ↑ Survival rate of tumor-bearing mice	[127]
	Mitochondria	SWNTs-PL-PEG-NH ₂	TAT	H22 liver cancer	Selective destruction of tumor mitochondria Tumor cell apoptosis	[128]
	CSCs	N.R.	IT	MDA-MB-231 breast cancer	Realize active targetability toward breast CSCs	[124]
MWNTs	Antigen-presenting cells	MWNTs-CpG- α CD40-OVA	IT	B16F10 melanoma	↑ Co-loading ability of OVA, CpG and anti-CD40 Ig	[129]

					↓Tumor growth and metastasis	
Tumor vasculature	iRGD-PEI-MWNT-SS-CD/pAT ₂	CT	A549 lung cancer	↓↓↓Tumor growth and neovascularization	[117]	

TT = Therapeutic target; DDS = drug delivery system; GT = gene therapy; IT = immunotherapy; CT = chemotherapy; * pancreatic cancer cells; CD40 = clusters of differentiation 40; CpG = cytidine-phosphate-guanosine; CSCs = cancer stem cells; IGF-1Ra = insulin-like growth factor-1Ra; MWNTs = multi-wall carbon nanotubes; OVA = ovalbumin; pAT₂ = plasmid angiotensin II type 2 receptor; PEI = polyetherimide; PEG = polyethylene glycol; PL = peptide lipid; PTT = photothermal therapy; SWNTs = single-wall carbon nanotubes; SS = disulphide bond; ↑ = high, higher, improved, enhanced; ↓ = low, lower reduced; ↓↓↓ = suppression, inhibition.

Table 6. Application of CNT-based DDSs in targeting various intracellular therapeutic sites and tumour microenvironment (TME) components.

TS	DDS	TM	Tumour model	Effectiveness	Biocompatibili ty	Ref s.
Cytoplasm	MWNTs/SOR/siRNA	GT+CT	Liver	↑SOR release, ↑ siRNA stability, ↑anticancer effects	N/A	[130]
Cytoplasm	SWNT-HIF-1α/siRNA	GT	Pancreas	Transfected tumour, activated RNAi response, ↓↓↓TG	Minimal harm to NCs	[92]
Cytoplasm	Ox-DWNT/siRNA	GT	Prostate	Prevent survivin creation and induce apoptosis by the release of siRNA into cytoplasm	Good biocompatibility	[131]
Nucleus	PEG/SWNTs/DOX	PTT+CT	Breast	↑Delivery effectiveness, ↑DOX localization/accumulation inside nucleus, ↑↑ killing of cancer cells	N/A	[132]
Nucleus	SWNTs-carrier	CT	Colorectal	Targeted therapy, regulated medication release	Good biocompatibility	[133]
Nucleus	f-SWNTs/p53	GT	Breast	↑Transport and targeting gene into the nucleus, apoptosis	N/A	[103]
Mito	MWNT-Rho (PtBzt+BP)	CT	Ovarian	Still ↓ selective	Still too much side effects	[134]
Mito	PEG/CNTs/ABT737	CT	Lung	↑Cell targeting to mitochondria, apoptosis	↓Toxic to NHFB NCs than A549	[135]

Mito	P-D-CS-CNTs	PTT	Bladder	↑Mito targeting with damage, ROS burst, cancer cells death	N/A	[136]
Mito	PL-PEG-SWNT	PAT	Breast	Mito malfunction, mito outer membrane permeabilization	↓Toxicity, no harm to EI	[137]
ECM	MWNTs	PTT	Epidermoid	↓↓↓Tumours, ↓volume, collagen destruction, cell damage	N/A	[138]
CCSs	SWNT-Raw, SWNT-COOH	CT	Osteosarcoma	Activated TGFβR1, ↓↓↓ its signalling, ↓ OSCs population	No toxicity to normal cells	[139]
TV	DOX/CD-CNT, CUR/CD-CNT	PTT+CT	Hepatocellular	↑ DEE and achieved sustained release of both drugs	No damage to normal cells	[140]
TV	iRGD-PEI-MWNT-SS-CD/pAT2	CT	Lung	↑CU, TE, ↓↓↓ angiogenesis, ↓↓↓ tumour growth	No damage or inflammation No damage to tissues §	[117]
PD-1/PD-L1	Rg3-CNT	IT	Breast	Inhibit PD-1/PD-L1 axis and the TNBC cell growth	N/A	[141]
IMs	MWNTs-DOX and MWNTs-CpG	IT+CT+P TT	Melanoma	↓↓↓Tumour growth, ↑CD4+, CD8+ T cells, ↑TAM shifting ↓Tregs in TME	No-toxicity to the organs*	[142]

Mito = Mitochondria; Rg3 = ginsenoside component of puffed ginseng with anti-cancer activities; RGD = arginyl-glycyl-aspartic acid; ECM = extracellular matrix; CCSs = cancer stem cells; IMs = immune cells; TV = tumour vasculature; TME = tumour microenvironment; DDS = drug delivery system; SOR = sorafenib; HIF-1α = hypoxia-inducible factor-1 alpha; TM = therapeutic modality; PTT = photothermal therapy; PAT = photoacoustic therapy; Ox-DWNT = oxidized double walled carbon nanotubes; CT = chemotherapy; GT = gene therapy; f-SWNTs-p53 = functionalized SWCNTs/plasmid 53 complexes; PEG = polyethylene glycol; Rho/(PtBzt+BP) = Rhodamine-110/Platinum (IV) prodrug+3-bromopyruvate; ABT737 = selective small molecule B-cell lymphoma 2 (Bcl-2) Homology 3 (BH3) mimetic; CS = chitosan; PL = phospholipid; NCs = normal cells; CUR = curcumin; DOX = doxorubicin; PEI = polyethylene imine; CD = candesartan; pAT₂ = plasmid AT₂; SS =cystamine; ↑ = high, higher, improved, enhanced; ↓ = low, lower reduced; ↓↓↓ = suppression, inhibition; EI = epithelial injury; * = liver, spleen, kidneys, heart and lungs in mice; §= blood, hepatic or kidney function in mice; TG= tumour growth; DEE = drug entrapment efficiency; CU = cellular uptake; TE = transfection efficiency; TGFβR1 = TGFβ type I receptor; TAM = tumour-associated macrophage.

3.2.2. Carbon Nanotubes-Based Drug Delivery Systems for the Tumor Microenvironment (TME)-Responsive Release of Chemotherapeutics

The enhanced permeability and retention (EPR) effect is a concept [143,144]establishing that small molecules, typically liposomes, nanoparticles, and macromolecular drugs tend to accumulate in tumour tissue much more than they do in normal tissues [145,146]. The conventional drug delivery systems (DDSs) work mainly exploiting the EPR effect and receptor-mediated endocytosis.

Unfortunately, they have habitually to afford many obstacles, due to the biological intricacy of the tumour microenvironment (TME), where cancer cells reside [147]. Anyway, the acid and hypoxic characteristics of TME and the high concentrations of glutathione (GSH) and H₂O₂ allow to conceive TME-responsive drug release tactics. Activated by several stimuli deriving by the conditions existing in TME, the drugs transported by these appositely engineered DDSs, can be released in a controlled or sustained mode, thus realizing various therapeutic effects and specifically penetrating deeply in the tumour tissue. Moreover, the reaction occurring between drugs-loaded DDSs and these sensitive factors can lessen tumour hypoxia and acidity, thus creating an environment hostile to tumour cells and more suitable for better therapeutic results. In this context, Yang et al. ideated oxidized MWCNTs gifted with large inner diameter, which allowed them to entrap inside cisplatin and to load doxorubicin (DOX) on the surface[148]. To hamper the early release of cisplatin, polyethylene glycol (PEG) and folic acid (FA) were also employed. With this approach, the carried cargos were endowed with a pH-sensitive release profile, and was released under pH = 6.5, which is just the weak acidic condition existing in TME. When tested on MCF-7 breast cancer cells, the nanocomposite demonstrated a more marked cytotoxicity in the pH = 6.5 condition than in pH = 7.4, confirming its best capacity to specifically kill tumour cells in acidic TME. Wang et al. engineered MWCNTs-based nanocomposites to be employed in magnetic resonance imaging (MRI)-guided and TME-responsive phototherapy[149]. A uniform MnO₂ sheet merged with Ce6 photosensitizer were modified with MWCNTs achieving a nano-system for enhanced phototherapy. MnO₂ was used because it was capable to rapidly deplete GSH, through redox reaction of Mn⁴⁺ into Mn²⁺ ions and to decompose H₂O₂ present in the TME, to give ¹O₂. By this approach, the photothermal effect engendered by MWCNTs was encouraged, tumour hypoxia was downed, and Ce6-mediated PDT was eased. ROS-mediated cell death increased via chemo-dynamic therapy and Ce6 release was sped up. Overall, this multifaceted MWCNTs-based platform could represent a promising strategy to achieve synergistic cancer diagnosis and therapy. Also, Qin et al. developed a CNTs-based nanoplatfrom encapsulated into a certain thermos-/pH-sensitive nanogel, thus obtaining a DDS capable of near infrared (NIR)-triggered, TME-responsive drug release[150]. Upon loading with DOX, this nano-system showed a quicker release rate of DOX at 40 C than at 25 C, and at pH = 5.0 than pH = 7.4, which indicated combinational outcomes, due to CNTs-mediated photothermal effects under NIR irradiation and TME-responsive drug release. The following Table 7 collects the above-mentioned case studies and several other ones.

Table 7. Application of modified CNTs as nanocarrier for the tumour microenvironment (TME)-responsive release of anticancer drugs.

CNTs	Functionalizing molecules	Effects	TM	Biocompatibility	Refs.
MWNT	DOX, Cis, PEG, FA	Stopped Cisplatin release, pH-sensitive release, synergic antitumor impact	L929, MCF-7	↑Biocompatibility	[148]

MWNT	MnO ₂ , PEG, Ce6	MnO ₂ breakdown of H ₂ O ₂ produced ROS thus ↑Ce6-mediated PDT PEG ↑WD, ↑cell death by ROS, ↑Ce6 release, MRI-guided PDT and PTT Synergic ↓tumour growth, in vitro and in vivo tumour removal	HeLa		[149]
SWCNTs	DOX-CS/PNIPAAm	↑Release at 40 C than 25 C, at pH 5.0 than pH 7.4 ↑ dual stimuli DOX release (NIR+AE of TME)	HeLa		[150]
SWNTs	PEG	↑WS, prevent PA	Gastric cancer	↓Toxicity to NTs	[61]
SWNTs/MWNTs	PEI	↑WS, homogeneity and dispersity, ↓particle size, ↑PC, ↑interaction with siRNA	Cervical cancer	N/A	[62]
SWNTs	HA	↑SS, ↑target CD44-overexpressing cancer cells, overcome MDR	Ovarian cancer	No cytotoxicity to NTs	[63]
MWNTs	CS	↑Water solubility and cell-penetrating ability	Breast cancer	↓Cytotoxicity	[64]
MWNTs	PLGA	↑Dispersity, ↑attachment sites, tuned temporal release	Osteosarcoma	↓Cytotoxicity in NC	[65]
SWNTs	CD133 mAB	Recognize and specifically target the GNM-CD33 ⁺ cells	Glioblastoma	N/A	[66]
SWNTs	IGF1R mAB and HER2 mAB	Target IGF1R and HER2 surface receptors	Breast cancer	Negligible toxicity in NCs	[67]
SWNTs	RGD peptide	Target α _v β ₃ -expressing cancer cells	MM	↓Toxicity in vitro	[68]
SWNTs	EGF receptor	Active targeting ability and ↑uptake of drugs	HC, NC	Negligible toxicity	[69]

DOX = Doxorubicine; Cis = cisplatin; PEG = polyethylene glycol; FA = folic acid; MnO₂ = manganese dioxide; Ce6 = Chlorin e6; CS/PNIPAAm = chitosan-poly (N-isopropyl acrylamide; TME = tumour microenvironment; DDDS = dual-drug delivery system; MRI = magnetic resonance image; AE = acidic environment; ↑ = improving, improved, augmented, high, higher; ↓ = low, lower, decreased; reduced; HA = hyaluronic acid; PA = particles aggregation; NTs = normal tissues; PC = positive charge; NCs = normal cells; PLGA = polylactic-co-glycolic acid; CD33 = cluster of differentiation-133; IGF1R = insulin-like growth factor 1 receptor; HER2 = Human endothelial receptor 2; RGD = arginyl-glycyl-aspartic acid; MM = malignant melanoma; HC = head carcinoma; NC = neck carcinoma; WD = water dispersibility; WS = water solubility; SS = serum stability; MDR = multi drug resistance; N/A = not acquired; TM = tumour model.

4. Carbon Nanotubes Application in Anticancer Phototherapy

4.1. Anticancer Photothermal Therapy

As an unconventional anticancer therapeutic technique, photothermal therapy (PTT) made part of a family of minimally invasive strategies, that are based on the use of photosensitizers. Specifically, PTT kills cancer cells thermically, by creating local heat using an optical absorption mediator, that is a photosensitizer, capable of adsorbing electromagnetic energy (EME) and transforming it into heat[151]. By generating a condition of hyperthermia, tumor eradication is significantly improved by boosting immune activation and causing immunity towards metastatic cancer cells for long time[152]. Unfortunately, conventional photosensitizers are affected by several drawbacks, including undesired adverse effects on skin, limited target to cancer cells, and scarce therapeutic effects in hypoxic TME. In this regard, CNTs represent excellent next-generation photosensitizer agents for a more effective PTT. They possess better photophysical properties and the ability to target cancer cells and accumulate in the tumor site. They are gifted with a broad electromagnetic absorbance spectrum and the capability to convert near-infrared I and II windows, which match the optical transmission window of biological tissues[153]. Moreover, the use of CNT-based tumor-targeting conjugates, in combination with PTT treatment, can result in more precise and efficient tumor elimination. Using well dispersible PEG-wrapped CNTs containing 80% (*w/w*) PEG on the CNTs surface, Sobhani et al. demonstrated the effectiveness of PTT treatments against HeLa and HepG2 cells[53]. Moreover, when PEG@CNTs were evaluated against melanoma, using a tumor-bearing mice model exposed to a continuous-wave near-infrared laser diode for 10 min, a more significant reduction in tumor size was observed in mice receiving also PEG@CNTs associated with laser irradiation, than in mice receiving only the laser radiation. Similarly, Zhu et al., avoiding the use of surfactants, prepared biocompatible, normal cell-friendly, and not toxic hybrid complexes, encompassing MWCNTs and gold nano-stars (MWCNTs@Au@NSs) [154]. MWCNTs@Au@NSs were experimented associated to PTT, on B16F10 mouse melanoma cells, observing that under 808 nm radiation, a photothermal effect 12.4% and 2.4 times higher than NSs and Au@NSs, respectively was produced, which caused enhanced cancer cells death. Also, by merging PTT with immune-stimulation by means of annexin A5 (ANXA5)-conjugated SWCNTs and anti-CTLA-4 check point inhibitor, McKernan et al. tried to treat metastatic breast cancer cells[155]. The designed combined anticancer therapy caused a significant increase in the survival rate of mice, in the number of CD4⁺ helpers and of CD8⁺ cytotoxic T cells, while SWCNTs had no toxic effects during the experiment. Moreover, even if the targeted and controlled release of genes remains a major challenge, Zhao et al. showed the synergistic effect of SWCNTs/MWCNTs-based PTT and gene therapy in exerting anti-tumor activity[126]. SWCNTs and MWCNTs were coated with peptide lipid and sucrose laurate to form a bifunctional DDS with improved photothermal effects and temperature sensitivity, which was loaded with siRNA. The complex silenced the survivin expression, thus effectively repressing tumor growth, while exhibiting photothermal effects under NIR exposure. Peptide lipid and sucrose laurate facilitated the phase transition of lipids, thus enabling the systemic delivery of siRNA to the tumor site. SWCNTs produced by the CoMoCAT[®] method possess high nanotube chirality, display an absorption band at 980 nm, and photothermal activity, which was widely used for selective photo-tissue

interaction[155,156]. In this regard, with the aim to target folate receptors on surface of cancer cells, Zhou et al. coupled CoMoCAT®-SWCNTs with folic acid (FA)[157]. In vitro and in vivo experiments demonstrated that the conjugate significantly reduced the photothermal destruction of normal cells, while significantly enhancing the photothermal death of tumor cells. Fibrin is a final product of coagulation response, which is highly concentrated at the vasculature injury site[158]. Fibrin existing in the tumor vessels can serve as therapeutic target for drug delivery, due to its easy accessibility, ubiquitous presence, and high expression. On these considerations, Zhang et al. engineered fibrin-targeting CREKA peptide-conjugated PEG@MWCNT nanocomposites for the PTT of cancer cells[159]. Exposure of a tumor-bearing mice model to MWCNT@PEG significantly increased the temperature of the tumor site, after 24 hours NIR radiation. It was shown that, upon illumination, IR783-labeled MWCNT@PEG accumulation in the tumor site was 6.4 times higher than in the control group[159]. MWCNT@PEG completely suppressed tumor xenografts after four illuminations cycles. Overall, MWCNT@PEG demonstrated significant tumor targeting and photothermal therapeutic effectiveness. Suo et al. proposed MWCNTs coupled to a Pgp-specific antibody (Pab) for photothermal P-Glycoprotein (Pgp)-mediated extirpation of MDR ovarian cancer cells[159]. Results demonstrated a remarkably improved internalization of the Pab-MWCNTs in 3T3-MDR1 than in 3T3 cells at different time periods. Under NIR irradiation, Pab-MWCNTs demonstrated higher dose-dependent specific photokilling in 3T3-MDR1 than in 3T3 cells[159]. Due to the higher temperatures to cancer cells reachable by plasmon phenomenon, differently modified MWCNTs demonstrated higher cytotoxicity to cancer cells. The highest temperatures for MWCNT-COO, MWCNT-COOPt, and MWCNT-Pt were 43.4°C, 45.8°C, and 46.2°C, respectively, while those of MWCNT-COO, MWCNT-COOAu, and MWCNT-Au were 44.1 °C, 46 °C, and 46.9 °C, respectively [160,161].

4.2. Anticancer Photodynamic Therapy

Another unconventional anticancer therapeutic technique, included in the family of noninvasive strategies against cancer and based on the use of photosensitizers, is the low toxic photodynamic treatment (PDT). PDT utilizes a combination of light, chemical photosensitizers and molecular oxygen, to induce cell death[56]. Briefly, upon the topical or systemic administration of a photosensitizer to tumor cells, it can be activated by light, at specific wavelength (NIR)[56]. An energy transfer cascade occurs, that induces ROS overproduction in the presence of oxygen, resulting in selective cytotoxicity against cancerous cells[162]. Racheal et al. engineered zinc phthalocyanine@spermine@SWCNTs and compared them to mono carboxy phenoxy phthalocyanine not containing zinc and zinc mono carboxy phenoxy phthalocyanine-conjugated spermine deprived of the SWCNTs. Photophysical properties and PDT efficiency towards MCF-7 breast cancer cell lines were assessed[163]. Results demonstrated that both ZnMCPPc-spermine and ZnMCPPc-spermine-SWCNT possessed photophysical characteristics superior to those of zinc-free phthalocyanine, with > 50% improvements in triplet and singlet oxygen quantum yields. In vitro cytotoxicity experiment carried out using MCF-7 cancer cells evidenced that the PDT carried out using ZnMCPPc-spermine and ZnMCPPc-spermine-SWCNT caused 97% and 95% cell viability reduction, respectively at 40 mM. Also, the same Racheal with colleague Nyokong assessed the PDT outcomes of a Zn-phthalocyanine modified with SWCNTs in association with ascorbic acid against MCF-7 tumor cells[164]. ZnMCPPc, ZnMCPPc@AA, ZnMCPPc@SWCNT, and ZnMCPPc@AA@SWCNT were evaluated for their photophysical properties (PPPs) and PDT performance (PDTP). ZnMCPPc@SWCNTs demonstrated enhanced PPPs, prolonged lifetimes and improved singlet oxygen quantum yields, with respect for ZnMCPPc alone. ZnMCPPc@SWCNT showed excellent PDTP against MCF-7 cells, leading to 77% cell viability reduction. Sundaram et al. studied the PDTP of SWCNTs-modified hyaluronic acid (HA) and chlorin e6 (Ce6) nanocomposites (NCs) on colon cancer cells[165]. HA coating significantly enhanced the dispersibility of NCs. The as-synthesized SWCNT@HA@Ce6 NCs proved higher anticancer effects on Caco-2 cells, than free Ce6. Also, they showed improved capacity to deliver photosensitizers, as well as stronger apoptotic activity in colon tumor cells, due to a high surface area and strong binding capacity [166]. Nano-bio-composites

associated with PDT (both 5 and 10 J/cm² laser irradiated) triggered both early and late-stage apoptotic death at major extent in cancer cells than in control ones. Specifically, exposure to 10 J/cm² (41.9, 6.65) caused more early-stage apoptosis than 5 J/cm² (36, 6.4). Similarly, survived cells were in lower percentages when exposed to 10 J/cm² (53.4, 6.8) than to 5 J/cm² (58.27, 5.9). Moreover, SWCNT@HA@Ce6 NCs under 10 J/cm² laser irradiation showed higher apoptotic effects than those displayed by free Ce6 and empty SWCNTs. It was reported by Shi et al., that HA-conjugated CNTs NCs were endowed with tumor targeting capability and improved solubility [167]. CNTs were modified with hematoporphyrin monomethyl ether (HMME) PDT agent achieving CNT@HA@HMME NCs, which were capable to combine local selective PDT with exterior near-infrared PTT, thus exerting improved therapeutic efficacy and reduced toxicity to normal cells in the treatment of cancer, by a synergistic effect [167]. Overall, HMME@HA@CNTs could perform both PDT and PTT treatment simultaneously in future enhanced tumor therapy.

4.3. Enhanced Anticancer Phototherapies by CNTs-Improved Drug Delivery

Light-induced hyperthermia in PTT and PDT frequently needs high-power intensity, which may damage nontarget normal cells[168]. To address this issue, Daquan Wang designed a nanoplatfrom (NP) by coating cut MWCNTs (C-MWNTs) with poly-N-vinyl pyrrole (PVPy) and binding folic acid (FA)-polyethylene glycol (PEG)-SH with thiolene achieving MWCNT@PVPy@S@PEG@FA nanocomposites (NCs). NCs possessed high drug-carrier ratio, pH-sensitive release capability of doxorubicin (DOX), and broad-spectrum anticancer effects. These features depended mainly on their exceptional photothermal conversion efficiency, capacity of high loading of DOX and of its target deliver, due to the presence of PVPy shell[169]. Similarly, Oh et al. used DOX-loaded SWCNTs to exert NIR cancer PTT chemotherapy[132]. Conversely, FA and methotrexate (MTX) were linked to the surface of -COOH modified MWCNTs (COOH-MWCNT) by using ethylenediamine (ED) as coupling agent, achieving MWCNT@ED@FA and MWCNT@ED@MTX NCs, demonstrating the NIR- or IR-laser promoted highest rate of MCF-7 cancer cell death. Cell death was high also at low doses of MTX when the laser light and MTX in MWCNT@ED@MTX or the laser light and MWCNT@ED@FA worked together [170]. Thermal tumour suppression triggered by CNTs was exploited by Znanag et al. to treat cancer cells by chemotherapy and PTT using an association of MWCNTs, gemcitabine and lentinan, an anticancer drug and an immune-stimulator, respectively[171]. Under 808 nm laser radiation, the proposed NC was more efficient in penetrating and suppressing cancer cells and less toxic than each system ingredient when used alone. Also, CNT-based NCs give the possibility to combine therapeutic and diagnosis activities. CNTs can be associated with other nanoparticles of different origin to produce synergistic therapeutic and diagnostic effects[172]. Karimi et al. investigated the cytotoxic effects of MWCNT@MTX and MWCNT@MTX@PEI@FA under 808 nm laser radiation, without observing noticeable differences[173]. DOX or CpG entrapped onto MWCNTs significantly increased MWCNTs water dispersibility. When melanoma-bearing mice were exposed to such modified MWCNTs under 808 nm NIR laser radiation at a dose of 1 W/cm² for 5 min, suppression of tumour growth and improved number of lymph draining CD4⁺ and CD8⁺ T cells were observed in the spleen, with increased antitumour efficacy[142]. PTT using targeted SWCNTs associated with immune system activation using a checkpoint inhibitor was proposed as an innovative treatment for the management of metastatic breast cancer. Selective NIR photothermal ablation of the primary orthotope at an energy and power level of 175 J/cm² and 1 W/cm², respectively, enhanced the anti-cytotoxic T lymphocyte-dependent abs copal response, translating into an increase in survival rate (55%), 100 days after tumour vaccination. The Annexin A5 functionalised SWCNTs, SWCNTs@ANXA5 NC was administered systemically before PTT[155]. Biocompatible 3D CNT@MXene@DOX microspheres were engineered displaying nonpareil photothermal effects and photothermal stability under 650 and 808 nm NIR laser radiation. As-manufactured microspheres were endowed with a maximum drug loading capacity of 85.6% for DOX. Also, 3D CNT@MXene microspheres effectively created singlet oxygen, due to TiO₂

photosensitized present on their surface. In-vitro experiments showed that 3D CNT@MXene@DOX successfully inhibited HeLa cancer cell proliferation[174].

4.4. Application of CNTs for Enhanced Anticancer Combined PTT and PDT

Single walled carbon nano-horns (SWCNHs) are a particular typology of CNTs, characterized by high photothermal conversion efficiency (PCE), which were studied to develop nanoplatforms suitable for different phototherapies thus enabling to engineer nanocomposites for combined PTT and PDT. In all relevant articles where PDT and PTT were experimented in combination, it was evidenced an increase of the pace at which tumours were reduced in animal models. Additionally, to further improve this impact, such combination can also be associated with another treatment, such as chemotherapy. In this context, Gao and co-authors developed a SWCNH@indocyanine green (ICG) theragnostic nano-system, which was experimented for diagnosis and combined PTT and PDT in breast tumour cells at low laser power, observing enhanced synergistic antitumour effects. Particularly, SWCNH@ICG induced ROS overproduction and hyperthermia effects, under light radiation. The mechanism which led to kill the tumour and slows its growth is shown in Figure 6[175].

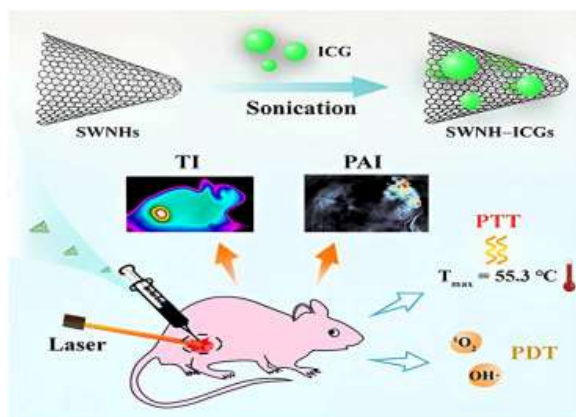


Figure 6. Thermal/photoacoustic imaging-guided PTT and PDT synergistic therapy nanoplatform developed by Gao et al. This image is published [175] and it has been reproduced under licence number 6027091475870 provided by John Wiley and Sons and Copyright Clearance Centre on 13 May 2025 (available online at [Rightslink® by Copyright Clearance Center](#), accessed on 13 May 2025).

The same authors created a similar platform, this time containing hypericin (Hyps). Upon laser radiation at 590 and 808 nm, this nanocomposite functioned as dual agent for simultaneous PDT and PTT against 4T1 cells implanted subcutaneously in mice as tumour models. Optimum outcomes in terms of anticancer impact by induced hyperthermia and ROS overproduction were observed [176]. Yang et al., immobilized Ce6 and Gd³⁺ on the surface of polymer-coated SWCNHs. Authors investigated its effects when experimented in combined PDT and PTT against tumours associated to the immune system's response [177]. High tumour targeting, enhanced tumour penetration efficiency and immune adjuvant effects were evidenced [177]. The possible synergistic effects of PDT and PTT were investigated by Yin et al., using a CNTs-based nanocomposite comprising Ce6 and MnO₂ to prevent tumour hypoxia, under radiation treatments at both 660 and 808 nm to activate both therapies. Despite tumour growth was inhibited, also by each therapy used independently, the PS, PA, MnO₂, and Ce6-MnO₂@CNTs (CMCs) compounds produced the best outcomes, when applied in simultaneous PDT and PTT, by synergistic effects[178]. Zhang et al., in addition to combine PDT with PTT, associated PDT and PTT with CT developing a nanoplatform including both SWCNTs and carbon quantum dots (CQDs). To provide outstanding selectivity toward tumour cells, water solubility of nanoplatform was improved using PEG, it was gifted with magnetic characteristics to

allow IMR using Fe₃O₄ NPs and DOX was loaded thus engineering SWCNTs@PEGFe₃O₄@CQDs@DOX@Apt nanocomposite. Despite all groups who receive radiation outperform those who receive only CT treatment, when the combination PTT, PDT and CT was used, tumour was suppressed, thus establishing its superiority for treating tumours[179]. Similarly, Wang et al. created a multifunctional nanomaterial, namely CNT@MnO₂-PEG@Ce6 for PDT, PTT, and MRI therapy. It comprised CNTs, MnO₂ NPs layers, PEG and Ce6 as photosensitiser (PS). When this nanoplatform was used for PDT and MRI, Ce6 and Mn²⁺ were rapidly released, due to the low pH and high GSH concentration of the TME, respectively. When CNT@MnO₂-PEG@Ce6 was applied for realizing a PDT and PTT combined treatment, by radiating it using simultaneously two-wavelength sources at 660 and 808 nm, a synergistic PDT/PTT effect was observed. which significantly decreased the tumour-development rate[149]. Also, Marangon et al. engineered a nanocomposite for treating SKOV3 ovarian cancer cells, using a combination of PDT and PTT. It was based on MWCNTs and the m-tetra hydroxyphenyl chlorin (m-THPC) as photosensitiser. The photothermal and photodynamic cytotoxic effects of m-THPC@MWCNTs complexes were investigated at the cellular level by several methods including viability tests, analysis of apoptosis-related proteins, genomic analysis of 84 genes involved in OS etc. The combination of PDT and PTT therapy resulted in cancer cells death, by the activation of various signalling pathways and by suppressing the cell's defence against OS[180]. Then, as previously reported, a new PDT agent, namely HMME, was adsorbed onto HA-modified CNTs (HA@CNTs), thus creating HMME@HA@CNTs nanocomposite, endowed with high aqueous solubility, neutral pH, and tumour-targeting activity. When this new nanocomposite was used in combination with PTT and PDT to treat tumour in vivo and in vitro, improved anticancer efficacy and low toxicity to normal organs was observed, due to the capacity of HMME@HA@CNTs NPs to combine local selective PDT with exterior NIR PTT, thus achieving a synergistic impact [167]. The following Tables 8 and 9 collect some relevant case studies reporting on the application of CNTs in anticancer PTT and PDT, while Table 10 summarizes reports on enhanced anticancer phototherapy by CNTs-based drug delivery. Lastly, Table 11 collects case studies on the application of CNTs to realize nanoplatforms to perform combination of PTT, PDT and CT.

Table 8. Application of modified CNTs in anticancer photothermal therapy (PTT).

CNTs	Functional molecules	Effectiveness	Tumour model	Observations	Refs.
MWCNT	Ag NPs	↑ NIR LA at 670 nm	Melanoma B16/F10 cells	↑CNT/Ag optical absorption to CNTs and Ag NPs	[181]
MWCNT	PEG	↑ MWCNTs water dispersibility	Melanoma	↓↓↓Tumour size in the cancerous mice	[53]
MWCNT	-COO, -COOPt, -Pt NPs	-COO ↑ CNTs solubility, Pt NPs ↑ LA at 1064nm	Cancer PC3 cells	PP ↑cytotoxicity due to ↑temperature of CCs	[161]
MWCNT	-COO, -COOAu, -Au NPs	-COO ↑ CNTs solubility, Au NPs ↑ LA at 1064nm	MCF7 cells	PP ↑cytotoxicity due to ↑temperature of CCs	[160]
MWCNT	MTX, PEI, FA	MTX-based ACA, FA-based TP, PEI-based hydrophilicity	MCF7 cells	↑Synergistic effect of MTX and PTT (808 nm laser) MWCNT-MTX-PEI-FA are absorbed selectively	[173]

MWCNT	PS MAA	Antibody against prostate-specific MA	Prostate cancer LNCaP cells	> 80% cell ablation with a single 30-s (a 532-nm)	[182]
MWCNT	Au NSs	↑Biocompatibility, ↑photothermal efficiency	Melanoma B16F10 mice cells	MWCNT/Au NSs have 12.4% ↑ PE (808 nm)	[154]
MWCNT	PEG	↑Photothermal transduction (NIR)	Benign prostatic hyperplasia	↓Prostate size, apoptosis in PEC	[183]
SWCNTs and MWCNTs	PL, SL	BDS with ↑ PE, TS	HeLa, MRC-5 cells	Muted surviving expression, ↓↓↓TG PE under 808 nm NIR light	[126]
MWCNT	TiO ₂ NPs	Ability to absorb and scatter the 808 nm NIR light	Melanoma	Selective tumour death, ↓HR, ↑necrosis	[184]
SWCNT	anti-IGF-1R, CY7	WS, BS, LT SWCNT-CY7-IGF-1R	PDCA	Target to PDCA for dye imaging-guided PTT	[127]

↓↓↓ = highly lowered, suppression; ↑ = improving, improved, augmented, high, higher; ↓ = low, lower, decreased; reduced; PEG = polyethylene glycol; LA = laser absorption; MTX = methotrexate; PEI =polyethylene amine; FA = folic acid; PS MAA = prostate-specific membrane antigen antibody; Au NSs = gold nano stars; PL = peptide lipid; SL = sucrose laurate; anti-IGF-1R = insulin-like growth factor receptor anti-IGF-1R antibody; CY7 = imaging agent; TiO₂ NPs = titanium oxide nanoparticles; WS = water soluble; BS = bio stable; LT = low toxic; BDS = bifunctional delivery system; PDAC = pancreatic ductal adenocarcinoma; PE = photothermal effects; TS = temperature sensitivity; BPH = benign prostatic hyperplasia; PEC = prostatic epithelial cells; PTT = photothermal therapy; PP = plasmon phenomenon; CC = cancer cells; ACA = anticancer activity; TP = targeting power; TG tumour growth; HR = heat resistance.

Table 9. Application of modified CNTs in anticancer photodynamic therapy (PDT).

CNTs	Functionalized molecules	Effectiveness	Tumour model	Biocompatibility test	Refs.
SWCNT	HA, Ce6	↑SWCNTs stability, WD (HA), Ce6-based ACA by PDT	Colon cancer cells	Colon cancer cells death, ↑PDT capacity	[165]
SWCNT	ZnMCP	>50% ↑ singlet and triplet oxygen quantum yields	MCF-7 breast cells	64% ↓ CV at 40 M (C1), 97-95% ↓CV (C2,3)	[163]
MWCNT	OEG, DOX, FA	OEG used to link DOX with CNT DOX used as anticancer drug FA act as the system's targeted ligand	HeLa, L929, A549	↑Killing of cancer cells when exposed to light.	[185]
SWCNT	AA, ZnMCP	↑ Lifetimes, quantum yields, singlet oxygen quantum yields	MCF-7	77% cytotoxicity by combined therapy	[164]

ACA = anticancer activity; ↑ = improving, improved, augmented, high, higher; ↓ = low, lower, decreased; reduced, reduction; HA = Hyaluronic acid; Ce6 = chlorine e6; ZnMCPP = zinc mono carboxy phenoxy phthalocyanine; OEG = ethylene glycol oligomers; DOX = doxorubicin; FA = folic acid; AA = ascorbic acid; WD = water dispersity; C1, C2 and C3 = complexes 1, 2 and 3; CV = cell viability.

Table 10. Applications of CNTs-based drug delivery systems for enhanced phototherapy.

CNTs	FM	Effectiveness §	TM	Biocompatibility	NIR laser	Refs.
MWCNTs	PVPy-S-PEG-FA, DOX	↑ATA, ↑WD, ↑BCT, ↑DL, ↑PC of CNTs by PVPy, pH-sensitive	HeLa	↑Biocompatibility	808	[169]
MWCNTs	ED	↑ATA, ↑AA of MTX, ↑CD by SE, MWCNT-ED-MTX = MWCNT-ED-FA*	MCF7	N.R.	808	[173]
MWCNTs	Ge, Le	↑AA by LH, ↑biocompatibility, ↑HDP, ↑PC	MCF-7	↓Danger to NC	808	[171]
MWCNTs	MTX, PEI, FA	↑ATA, ↑WD by PEI, ↑AA of MTX by SE with PPT	MCF-7	N.R.	808	[173]
SWCNTs	ANXA5	↑Stimulation of IS, AE by SWCNT-ANXA5 SE	EMT6 BCCs	N.R.	980	[155]
CNTs	MXene	↓CNT aggregation, DOX DL = 85.6%, ↓↓↓HeLa cells by PTT+DOX SE	HeLa	N.R.	650/808	[174]

FM = Functionalizing molecules; TM = tumour model; PVPy = poly-N-vinyl pyrrole; S = ; PEG = polyethylene glycol; ED = ethylenediamine; Ge = gemcitabine; Le = lentinan; FA = folic acid; MTX = methotrexate; PEI = polyethyleneimine; ANXA5 = annexin 5 is a phospholipase A2 and protein kinase C inhibitory protein with calcium channel activity and a potential role in cellular signal transduction, inflammation, growth and differentiation; MXene = tuneable family of 2D carbides and nitrides; ↑ = enhancing, enhanced, improving, improved, augmented, high, higher; ↓ = low, lower, decreased; reduced, reduction; BCT = blood circulation time; WD = water dispersibility; ATA = active targeting ability; BCCs = breast cancer cells; AA = anticancer activity; PC = photothermal characteristics; HDP = hydrophilicity; IS = immune system; DL = drug loading; * for anticancer effects; CD = cell death; SE = synergistic effects; LH = localized hypothermia; § = under NIR radiation; AE = abscopal effect; ↓↓↓ = suppression; NCs = normal cells.

Table 11. Application of modified CNTs in combined PTT and PDT for cancer treatment.

CNTs	FM	Effectiveness	TM	Biocompatibility	Refs.
SWCNHs	ICG	↑WD, ↑photostability, ↑AA by PTT+PDT SE	4T1 BCCs *	↑Photostability, ↑biocompatibility.	[175]
SWCNHs	Ce6, Gd ³⁺	↑IS, ↑ATA, ↑TP, ↑ISR by PTT+PDT SE in AMMs	4T1	N.R.	[177]
SWCNHs	Hyp	↑WS ↓4T1 growth by PDT+PTT SE	4T1 *	↑Photostability, ↑biocompatibility	[176]
MWCNT	MnO ₂ , Ce6	O ₂ +H ⁺ production by MnO ₂ ↓TM, ↑potential for IG PDT+PTT SE	HeLa *	↑Biocompatibility	[178]

SWCNT	PEG, Fe ₃ O ₄ , CQDs, DOX, sgc8c	↑AA by PTT+PDT+CT by SE	HeLa mice	↑Biocompatibility	[179]
MWCNT	MnO ₂ , PEG, Ce6	MnO ₂ create 1O ₂ , ↑PTT, ↓hypoxia, ↑PDT, ↑WD, ↓↓ TD by MRI-guided PDT+PTT SE	HeLa *	↑Biocompatibility	[149]
MWCNT	mTHPC	mTHPC acted as PS, ↑AA by PDT+PTT SE	SKOV3 OCCs	N.R.	[186]
SWCNT	HA, HMME	↑WS, HMME = PDT agent, ↑AA by PDT+PTT SE	B16F10 MMCs	↑Biocompatibility	[167]

SWCNHs = single walled carbon nano horns; ICG = indocyanine green; Ce6 = chlorine e6 photosensitizer; Gd³⁺ = gadolinio; Hyp = hypericin; PEG = polyethylene glycol; CQDs = carbon quantum dots; DOX = doxorubicin; sgc8c = aptamer; m-THPC = m-tetrahydroxy phenyl chlorin; HA = hyaluronic acid; HMME = hematoporphyrin monomethyl ether; FMs = functionalizing molecules; TM = tumor model; TD = tumor development; ↑ = enhancing, enhanced, improving, improved, augmented, high, higher; ↓ = low, lower, decreased; reduced, reduction; WD = water dispersibility; ATA = active targeting ability; TP = tumour penetration; BCCs = breast cancer cells; AA = anticancer activity; ISR = immune system response; SE = synergistic effects; ↓↓↓ = suppression; WS = water solubility; OCC = ovarian cancer cells; CT = chemotherapy; PS photosensitizer; MMCs = mice melanoma cells; AMMs = advanced metastatic malignancies; * in vivo and in vitro experiments; IG = imaging guided; MRI = magnetic resonance imaging; N.R. = not reported.5. Carbon Nanotubes Application in Anticancer Gene Therapy.

Gene therapy (GT) is a novel engineered therapeutic approach which aims at using genes to cure several diseases, including cancer, by replacing defective genome of diseased cells with healthy one [187]. In GT, the opportunely selected healthy genes are inserted into the selected cells, including cancer ones, by transfection, to repair defects in their genome or compensate for the cells’ deficiencies, due to uncontrolled mutations inexplicably occurred [187]. GT and its synergistic combination with CT are gaining strong interest in cancer treatment. Unfortunately, insufficient endosomal escape of genes/nanocarriers complexes, due to an inadequate buffer capacity of the carrier cause lysosomal early degradation [56,187]. This event causes poor transfection capability and strongly limits the therapeutic applicability in vivo of this approach. The use of CNTs-based carriers, opportunely modified to enhance their buffer capacity, can promote endosomal effects and survival of the gene complex, thus ameliorating significantly the transfection efficiency. The use of suicide genes is among the most efficacious approaches to realize efficient anticancer gene therapy. It consists of using therapeutic transgenes to express toxic products from a toxic gene or to convert a nontoxic prodrug into a toxic one, or both, to fight the effects of cancer disease. This strategy was essayed to treat several cancers, such as breast[188], liver, colon[189], prostate[190], glioma[191], and lung cancer [192] also when cancer cells have acquired chemo-resistance [193]. Additionally, this approach has demonstrated to enhance the efficacy of radiation therapy [194]. Furthermore, combination therapy usually is more efficient than monotherapy, due to its capability to elude the cell cycle arrest caused by chemical drugs. Several studies were developed based on the above-mentioned considerations. Cao et al. engineered a novel pH-responsive SWCNTs-carrier functionalized with PEI-betaine (PB) (SPB), improved with BR2 peptide and loaded with DOX and survivin siRNA (SPB@BR2@DOX@sur/siRNA), for the co-delivery of an anticancer drug and a silencing gene and realizing efficient anticancer gene therapy[114]. When administered to both HeLa and A549 cancer cells and 293T normal cells, the nanocomposite was selectively internalized into cancer cells, while it did not enter normal cells[114]. The nanocomposite caused less survivin expression and a higher apoptotic index than Lipofectamine 2000, due to release of siRNA/DOX into the A549 cell cytoplasm

and nuclei, without lysosomal degradation. In comparison to SPB@BR2@siRNA or SPB@BR2@DOX separate treatments, that in association using SPB@BR2@DOX@sur/siRNA demonstrated synergistic effects, causing significant reduction in the volume of tumor both in A549 cells and in nude mice[114]. Based on the capability of iC9 suicide gene to induce apoptotic death in MCF-7 human breast cancer cells, Dargah et al. used pyridine modified MWCNTs (pyr@MWCNTs) as carriers to transfect iC9, achieving the pyr@MWCNTs@iC9 complex [195]. Upon its administration, MCF-7 cells were exterminated, and when associated with chemotherapy, it evaded cell cycle arrest. Also, gene regulation and anticancer therapy were attempted by Zhang et al. experimenting the chitosan modified fluorescent carbon nanoparticle (FCN)-based siRNA conjugate (Ch@FCN@siRNA)[196]. The core-shell nanocomposite comprised a core made of Ch@FCN and a shell of siRNA. siRNA down-regulated the key regulator of mitosis, namely polo-like kinase-1 (PLK1) expression. Notable, only a concentration of FCN 30 times lower than that of AuNPs was sufficient to transfect the same amount of siRNA. The nanocomposite, in vitro treatment of A375 and MCF-7 tumor cells, was better performant of the commercial Lipofectamine 2000 inducing 31.9% and 20.33% apoptosis, respectively. Also, its intravenous administration to mice bearing the A375 tumor cells reduced tumor volume by 11-fold compared with control groups. As reported in previous sections, Guo et al. used MWCNTs as efficient siRNA vectors. Precisely, the authors modified MWCNTs with NH_3 groups, which were cationic at physiological pH, obtaining positively charged MWCNTs- NH_4^+ tubes, which were used for siRNA delivery against PLK-1 cancer cells in mice [95]. Upon the administration of siRNS@MWCNTs- NH_4^+ complex lung cancer xenografts were eradicated [95]. Another study by Anderson et al. used SWCNTs for the targeted delivery of siRNA to pancreatic cancer cells for anticancer gene therapy [197]. In this contribution, the prepared SWCNTs@siRNA complex was tested in vitro on pancreatic cancer cells, observing high siRNA transfection efficiency, successful internalization of the nano complex in cancer cells and low toxicity versus normal cells[197]. The release of siRNA from the nano complex resulted in the downregulation of the target oncogene [197].

6. Carbon Nanotubes Application in Anticancer Immunotherapy

Immunotherapy is a therapeutic approach, which is finalized to improve the patient's capability to fight several types of diseases, including cancer, by either the modification or amplification of the immune system by exploiting antigenic targets [198]. This therapeutic strategy has proven to be efficacy to enhance therapeutic effects against chronic infections and cancer. Another anticancer strategy consists of blocking regulatory systems which may interfere with the immunotherapeutic effects. In the body, dendritic cells (DCs) represent a crucial connexion between innate and adaptive immunity. They are the most potent specialized antigen presenting cells (APCs), playing a pivotal role in anti-infection and anti-tumour responses. Unfortunately, cancer cells can overpower both the immune system and the functionality of DCs, limiting the efficacy of DCs-based antitumor immunotherapy. Hence, it could be of paramount help improving the antitumor immune response by controlling DCs functionality and disabling immune tolerance. Also, cytotoxic T lymphocytes (CTLs), also known as CD8^+ T and CD4^+ T cells, are crucial cells of the adaptive immune system. They play a determinant role in the defence against pathogens such as viruses, bacteria, and tumours [199]. An inadequate infiltration of CD8^+ T cells in the immunosuppressive TME, results in a decreased antitumor response. When CD^+ T cells are even absent, the body will lack antitumor immune function. Conversely, the over-presence of CD8^+ T cells can trigger excessive immune responses, leading to immune-mediated tissue damage or pathological reactions [200]. Thus, enhancing CD8^+ T cell infiltration in TME, as well as promoting their correct functional activity are pivotal strategies in tumour treatment. Carbon-based nanoparticles, including CNTs could be supreme platforms for tumour detection and immunotherapy. Several research indicates that polymer modified CNTs can treat tumours by acting as immune adjuvants, to promote the maturation of dendritic cells (DCs), the CD8^+ T cells infiltration in TME, and to release anti-tumour factors [201]. Furthermore, MWCNTs conjugated with peptides can promote cytokine secretion, stimulating T cell differentiation and proliferation[202].

Carbon-based nanomaterials (CNMs) and CNTs can augment antitumor immunity of patients, via multiple and several mechanisms of immune system modulation. The most recognized comprises first the pickup of the CNT–antigen conjugate by DCs, which transfer the antigen peptides to naive T cells for activation. To this work, a multitude of unique receptors exists on the surface of DCs, which naturally serve as recognition sites for activating specific immune cells. The movement of antigens to specific compartments for presentation in DCs is critical. In DCs, the lysosome-dependent pathway causes the antigen breaking down into antigenic peptides (APs) inside lysosomes. APs are then loaded onto Class II major histocompatibility complex (MHC-II) molecules for presentation to CD4⁺ helper T cells. On the other hand, MHC-I molecules display cytosolic antigens to activate CD8⁺ T cells and trigger cytotoxic T lymphocyte (CTL) responses[203]. Among cytokines, TNF- α provide chemical signals to the cancer cells, causing inflammation and cell death[204]. Activated T cells and natural killer (NK) cells release IFN- γ , which activates macrophages and improves antigen presentation[205]. Other cytokines, such as IL-15 and IL-12 activate and stimulate proliferation and expansion of NK cells and other antitumor immune cells, including CD8⁺ T cells [206] (Figure 7).

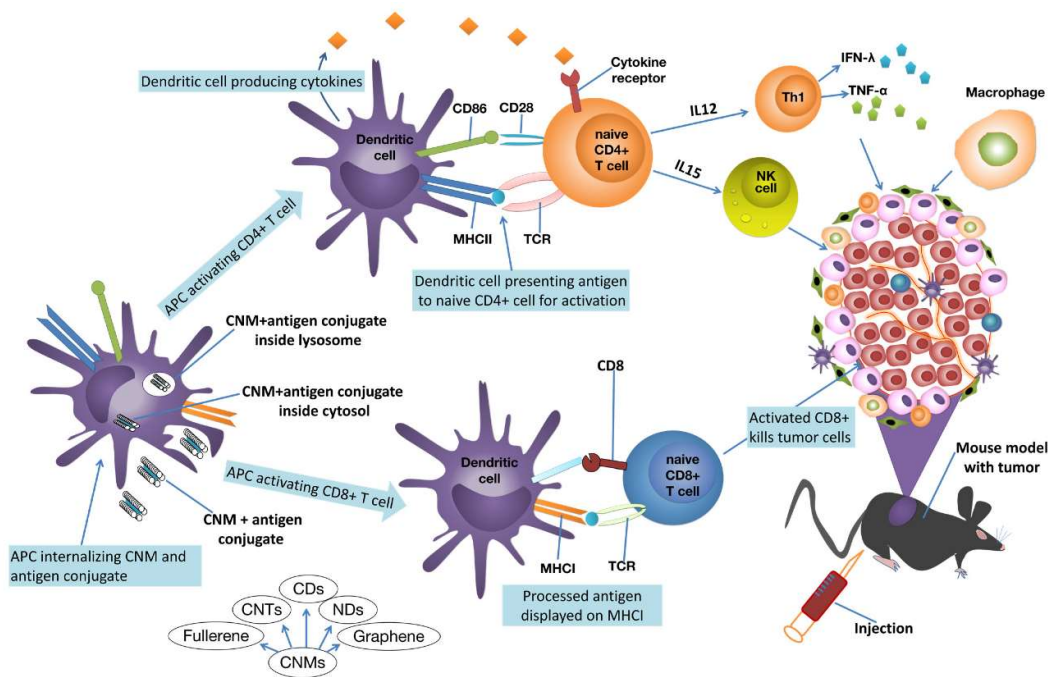


Figure 7. Schematic representation of the immunomodulatory effects of CNMs in cancer therapy [207]. The image has been reproduced by an open access article [207]. Reproduction is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>. The Creative Commons Public Domain Dedication waiver (<http://creativecommons.org/publicdomain/zero/1.0/>) applies to the data made available in this article, unless otherwise stated in a credit line to the data. APCs = antigen-presenting cells; DCs = dendritic cells; CNM = carbon nanomaterial; CNTs = carbon nanotubes; CDs = carbon dots; NDs = nanodots; MHC-II = histocompatibility complex II; CD4⁺, CD8⁺ = helper T cells; MHC-I = histocompatibility complex I; CTL = cytotoxic T lymphocyte NK = natural killer.

The subsequent Table 12 collects studies where CNTs were utilized to synthesize CNTs-based nanoplateforms for immune oncotherapy.

Table 12. CNTs application in anticancer immunotherapy.

CNTs	FM	effectiveness	TM	Biocompatibility	Refs.
MWCNTs	COOH	↑WS, ↑CSA, ↑CP, ↑MA	H22 HCCs	N.F.	[208]
MWCNTs	Rg3	Puffed ginseng, ACA, Apoptosis, ↓PD-L1 expression	TNBC	N.F.	[141]
SWCNTs	CpG	ACA in gliomas, ↓CCP, ↓invasion/migration	HCCsT116	↑Mice survival rate	[209]
MWCNTs	DOX/ CpG	↑WD; no change in MWCNTs PS, ↓TG, ↑CD4+, ↑CD8+, ↑T cells	B16 MCs	Biocompatible to normal cells	[142]
SWCNTs	COOH/OVA	Primary immunization/mice, ↑anti-ovalbumin antibody response	AMBCs	↓Toxicity, ↓inflammation	[210]
MWCNTs	PDAEMA	↑Design flexibility, ↑control on SARs, ↑SE, ↑CU	B16-F10	↑Biocompatibility	[211]

FM = Functionalizing molecules; Rg3 = Ginsenoside Rg3; CpG = complex; DOX = doxorubicin; OVA = Ovalbumin; PDAEMA = poly(2-dimethylaminoethylmethacrylate); WS = water solubility; WD = water dispersibility; ↑ = enhancing, enhanced, improving, improved, augmented, high, higher; ↓ = low, lower, decreased; reduced, reduction; ACA = anticancer activity; PS = physical structure; SAR = structure activity relationships; HCCs = hepatocarcinoma cells; TNBCs = triple-negative breast cancer; HCCsT116 = human colon cancer cell line T116; MCs = melanoma cells; AMBCs = activated mouse B cells; TM = tumor model; CSA = component system activation; CP = cytokines production; MA = macrophages activation; CCP = cancer cells proliferation; TG = tumor growth; SE = silencing efficiency; CU = cellular uptake; N.F. = not found.

Immuno-based oncotherapy was experimented by Xia et al. in vitro and in vivo, using MWCNTs-based nano-delivery systems loaded with unmethylated CpG moieties, an oligodeoxynucleotide, and H3R6 polypeptide (MHR-CpG) for treating prostate cancer [212]. Authors observed enhanced biocompatibility, endosomal TLR9 targeting and improved immunogenicity of CpG in both the humoral and the cellular immune pathways. An increase in the expression of CD4⁺ T-cells, CD8⁺ T-cells, TNF-, and IL-6 were detected. When tested in vivo in RM-1 prostate tumor-bearing mice, the nanocomposite demonstrated to be capable of delivering the immune therapeutics to both the tumor site and to lymph nodes, thus inhibiting prostate cancer growth. Hassan et al. used antigen-bearing MWCNTs to deliver immunoadjuvants such as cytosine-phosphate-guanine oligodeoxynucleotide (CpG), anti-CD40 Ig (CD40), ovalbumin (OVA) antigen to trigger immune response against OVA-expressing cancer cells. When tested in vitro and in vivo, the MWCNTs-based nanoplatfrom caused a dramatically high OVA-specific T cell responses in vitro and in C57BL/6 mice. Co-loaded OVA antigen, CpG, and anti-CD40 Ig prevented the proliferation of OVA-expressing B16F10 melanoma cells in pseudo-metastatic subcutaneous or lung tumour models[129]. Furthermore, SWCNTs realized the efficient delivery of CpG in CX3CR1GFP mouse models, without toxicity to normal cells, and increased the production of proinflammatory cytokines by primary monocytes. Surprisingly, a single intracranial injection of low-dose CNT-CpG removed intracranial GL261 gliomas in half of tumour-bearing animals, via activation of NK and CD8⁺ cells, and protected the surviving mice from the recurrence of intracranial cancer.[213]. Oxidised MWCNTs (ox-MWCNTs) bearing COOH groups were prepared by Radzi et al. and tested for treating breast cancer in EMT6 model mice, associated with PTT. The combined therapy resulted in full cancer eradication and in a substantial increase of mice median survival rate. Additionally, MWCNTs-based nanocomposite increased the infiltration and maturation of DCs, CD8⁺, CD4⁺ T cells, macrophages and NKs in tumours treated with ox-MWCNTs–hypothermia combination therapy [214].

Wilm's tumour protein (WT1) is a protein upregulated in many human leukaemia and cancers. Villa et al. covalently attached WT1 ligands onto soluble SWCNT architectures achieving SWCNT-peptide nanoplatforms, which were speedily absorbed by dendritic cells and macrophages in vitro. Immunization of BALB/c mice with SWCNT-peptide nanocomposite and immunological adjuvants provoked specific IgG responses versus the peptide [215].

Fadel et al. utilized SWCNT bundles in the presentation of T-cell-activating antibodies to evoke immune responses in target tumours. SWCNTs bundles delivered anti-CD3 T-cell-stimulating antibodies, with high local concentrations, resulting in powerful activation of T cells, thus demonstrating that SWCNT bundles constitute a unique model for the effective activation of lymphocytes, with implications for fundamental science and clinical immunotherapy[216].

Functionalized bundled SWCNTs (*fb*-SWNTs) have demonstrated to be efficient antigen-presenting substrates and were used by Fadel et al. to absorb T cell antigens and CD3 and CD28 costimulatory ligands. The as achieved nanoplatforms were used to treat splenocytes obtained from the spleens of C57BL/6 mice. The adsorption of T-cell-stimulating antibodies improved both the kinetics and amount of T cell activation, thus supporting the utilization of chemically processed nanotube bundles in clinical applications requiring the presentation of artificial antigen[217].

Later, the same authors developed a simple yet robust technique of noncovalently attaching the T cell stimulus (MHC-I) to CNT substrates, for avoiding undesired denaturation effects. They used the achieved nanocomposite to treat OT1 mice, observing increased antigen-specific T cell responses which was 3-fold higher than in control [218].

Also, Burkert et al. developed AuNPs-bearing nitrogen-doped nitrogen nanotube cups (Au-NCNCs), which demonstrated capability in entrap and target deliver of paclitaxel in the tumour site. Au-NCNCs altered the TME, reduced tumour growth rate, and counteract immunosuppressive macrophages [219].

Anyway, it is dutiful remembering that the needle-like shape of CNTs, which allows them to penetrate cellular membranes, causing damage, can also trigger inflammatory responses, that may lead to harm in both animals and humans. MWCNTs revealed high phagocytic activity towards undifferentiated HL60 cells and cytotoxic effects on differentiated HL60 cells [220].

Due to a lack of critical clinical evidence, the exact mechanisms by which CNTs can harm humans and animals remains unclear. On the other hand, many studies suggest that proper modifications can lower the hazardous effects of CNTs making them eligible for applications in the biomedical field. Scientists in different field concerning CNTs should conduct extensive research in collaboration to address concerns surrounding CNTs safety and to enhance their credibility. Looking ahead, researchers are expected to develop innovative synthetic methods or create novel composite materials to improve cancer treatment outcomes and enhance human health.

7. Carbon Nanotubes Application in Cancer Diagnosis

Cancer diagnosis aims at examining and detecting the etiologies and related symptoms concerning various types of cancer, using modern technologies. Due to their nonpareil properties, CNTs are increasingly attracting interest also in this field. The application of CNTs in various cancer imaging, such as Raman imaging, nuclear magnetic resonance imaging (NMRI), ultrasonography (US), photoacoustic imaging (PAI), radionuclide imaging (RNI), near-infrared fluorescence imaging (NIR-FI), as well as their use to engineer cancer nano-biosensors, as schematized in Figure 8 and summarized in Table 13, are the topics of this section.

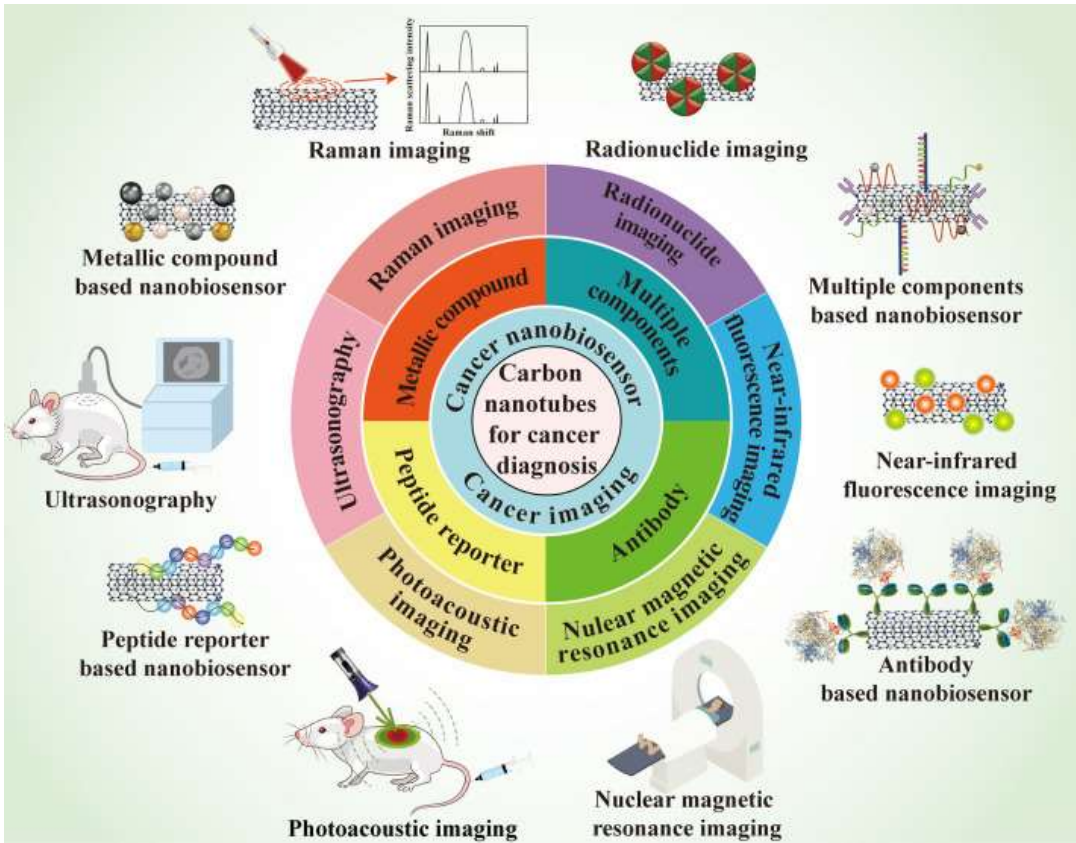


Figure 8. An overview of the contributions of CNTs to cancer diagnosis. The image has been reproduced by an open access article [221]. Reproduction is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>. The Creative Commons Public Domain Dedication waiver (<http://creativecommons.org/publicdomain/zero/1.0/>) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

Table 13. Applications of CNTs in cancer diagnosis.

CNTs	DM	Effect	Refs.
SWCNTs	RI	Spots of RSs were visible in colon-26 cells after 5-days administration of o-SWNTs-PEG	[222]
SWCNTs	RI	↑SERS-E suitable for labeling and fast RI of biological samples	[223]
SWCNTs	MRI	Excellent MRI functions for tumor diagnosis	[224]
MWCNTs	MRI	↑Longitudinal proton relaxation process, T1 enhanced MRI effect	[225]
MWCNTs	US	↑Contrast in ultrasonic imaging	[226]
MWCNTs	US	Strong, long-lived and ↑-quality ultrasound signal after sonication treatment	[227]
SWCNTs	PAI	Clear and stable PA signals	[228]
MWCNTs	PAI	Precise target to the tumor site in vivo, ↑PA imaging effect	[229]
MWCNTs	RNI	Easy and direct SPECT/CT imaging	[230]
SWCNTs	RNI	↑Tumor tissue accumulation for the subsequent radionuclide imaging	[231]

SWCNTs	NIR-FI	↑Signal-to-noise performance, ↑specificity to ovarian tumors and tumor nodules	[232]
SWCNTs	NIR-FI	↑Resolution intravital tumor vessel images through the thick skin in live mice	[233]
MWCNTs	NBSs	↑Biosensing ability, wide linear range for detecting miR-21, ↓↓↓detection limit	[234]
SWCNTs	NBSs	Broad detection range, ↓detection limit, ↑specificity to only OPN in prostate cancer	[235]
MWCNTs	NBSs	Simple detection of CDK1, report the enzymatic activity of CDK1 for cancer diagnosis	[236]
MWCNTs	NBSs	↑Selectivity/sensitivity to HPV-18, early, rapid, easy, and accurate diagnosis of cervical cancer	[237]

RI = Raman imaging; NBSs = nano-biosensors; NIR-FI = near infrared fluorescence imaging; CNTs in cancer imaging; RNI = radionucleotide imaging; US = ultrasonography; PAI = photoacoustic imaging; MRI = magnetic resonance imaging; DM = diagnostic method; RS = Raman signal; SERS-E = surface-enhanced Raman scattering effect; SPECT = single photon emission computed tomography; CT = computed tomography; ↑ = strong, enhancing, enhanced, improving, improved, augmented, high, higher; ↓ = low, lower, decreased; reduced, reduction.

CNTs can provide multifunctional bio-probes with several unprecedented properties, including strong absorbance in NIR, good resonance Raman scattering, and high modifiability, thus representing excellent materials suitable for cancer imaging, with research and clinical prospective in cancer diagnosis.

7.1. Raman Imaging

The radial breathing model (RBM) and tangent G-module (TGM) are unique vibrational features of CNTs, which can be detected by a Raman microscope [238]. SWCNTs doped with oxygen, bearing epoxide groups and modified with PEG (o-SWNTs@PEG) were engineered by Sekiyama et al. The achieved nanocomposites were created to serve as over-thousand-nanometer (OTN)-NIR fluorescent probes, to investigate the time-dependent change in OTN-NIR fluorescence images of colon-26 cancer cells[222]. Upon administration of the probes to colon-26 cancer cells, their distribution in cells was studied using Raman microscopy, observing Raman signals on the 5th day from first administration. Since noble metals on the surface of CNTs improve their Raman signals[239], Wang et al. enriched the surface of PEG@CNTs with gold or silver, thus improving the Raman scattering (SERS) effect of pristine CNTs. The use of noble metal modified CNTs rather than that of not modified PEG@CNTs allowed to acquire the Raman images under NIR radiation in remarkably minor time [223].

7.2. Nuclear Magnetic Resonance Imaging

Nuclear magnetic resonance imaging (NMRI) is a non-invasive imaging technique, which does not use ionizing radiation, thus being not risky for the human body. It provides the original 3D cross-section images of a tissue or organ and is of paramount help in medical imaging [240]. Since CNTs can be used as T_2 spin dephasing contrast agents, they are utilized to enhance nuclear magnetic resonance imaging (NMRI) in clinics [241]. Yan et al. by the non-covalent association of the NGR (asparagine-glycine-arginine) peptide with DOX and CNTs bearing NMR contrast agent, namely Gd-DTPA, prepared a new nanocomposite theragnostic, for both detecting the tumor by NMRI and anticancer therapy [224]. Zhang et al. synthesized a multimodal nanoplatform based on MWCNTs (FA@GdN@CQDs-MWNTs/DOX) using gadolinium NPs (GdN), magneto fluorescent carbon quantum dots (CQDs), and folic acid (FA) [225]. In vitro targeting NMRI experiment revealed that

FA-@dN@CQDs-MWNTs worked as excellent T1 contrasting agent overstating the longitudinal proton relaxation process. As confirmation, in vivo essays showed that the NMR signal at tumor site was positively enhanced after intravenous administration of FA@GdN@CQDs-MWNTs.

7.3. Ultrasonography

Ultrasonography is a low-cost and intrinsically safe diagnostic imaging technique [242]. CNTs are nanomaterials particularly eligible for ultrasonic imaging, since during ultrasonography they are capable to produce high signals that can be detected by a contrast-enhanced ultrasound imager. Saghatchi et al. prepared multi-functionalized (*mf*) MWCNTs by their modification with both magnetic Fe₃O₄ and gold NPs (*mf*-MWCNT@AuNPs for simultaneous cancer imaging and therapy [226]. This nanocomposite, when applied at various concentrations, exhibited notable high contrast. Delogu et al. modified MWCNTs using azomethine ylides to improve their biocompatibility and applied the achieved nanocomposite (ox-MWNT-NH₃⁺) in ultrasonography[227], observing a strong, long-lived and high-quality ultrasound signal, after sonication treatment.

7.4. Photoacoustic Imaging

In photoacoustic imaging (PAI), upon the irradiation of cells, tissues and/or organs by pulsed laser, the luminous energy is adsorbed, converted into ultrasonic by the thermal expansion of tissues and organs, and detected by sensors[243]. Since CNTs possess strong NIR absorption and deep tissue penetration, they could serve as ideal contrast agents for PAI, whose signal can be recorded by a photoacoustic microscopy[244]. Using PAI technology in cell imaging, Avti et al. detected, localized and quantified the content of CNTs in different tissue samples, thanks to signals clearly understandable and stable, due to the potent NIR absorbance of CNTs[228]. Wang et al. engineered a multifunctional MWCNTs-based probe encompassing the RGD peptide, a silica-coating, and Au nanorods (RGD@sGNR@MWNTs) endowed with active targeting ability, to be used for the in vivo PAI of gastric cancer[229]. After intravenous administration of RGD@sGNR@MWNTs probe, the treated mice were analyzed by an optoacoustic imaging system, evidencing that RGD@sGNR@MWNTs precisely targeted the tumor site and provided strong photoacoustic imaging effects.

7.5. Radionuclide Imaging

Radionuclide imaging (RNI) is an imaging technology which exploits radioactive isotopes such as ¹¹¹In, ¹³¹I, ⁶⁴Cu, and ⁸⁶Y, which upon injection into the body are adsorbed by human tissues and organs, thus functioning as radiation source in vivo emitting γ -rays during their decay process [245,246]. Such rays can be detected by nuclear detection devices which provide the distribution density of the radioactive isotopes in vivo. RNI is endowed with deep tissue penetration, negligible limitation and high sensitivity[247]. The organ biodistribution of various functionalized (*f*)-MWCNTs in vivo was studied by Wang et al., who used MWCNTs radio-labelled with ¹¹¹In to empower easy in vivo single photon emission computed tomography/computed tomography (SPECT/CT) imaging[230]. Zhao et al. modified SWCNTs with polydopamine (PDA) and PEG, achieving a nanoplatform (SWNTs@PDA-PEG), which was further labelled with ¹³¹I, for the subsequent RNI[231]. After in vivo administration, the tumor tissue distribution of ¹³¹I@SWNTs@PDA-PEG was detected by a gamma counter.

7.6. Near-Infrared Fluorescence Imaging

Near-infrared fluorescence (NIR-F) imaging, having a NIR biological wavelength window in the range of 780-1700 nm, is an appealing and fast-progressing imaging technique, extremely useful for cancer diagnosis. CNTs possess clear optical absorption and intrinsic fluorescence in the above-mentioned range, thus being promising materials for application in NIR-F imaging during cancer diagnosis, whose images are usually acquired using an in- vivo imager [248,249]. Since SWCNTs

possess higher absorbing properties, stronger optical absorption, better E11 optical transitions, and less photobleaching than MWCNTs, they are more appropriate for NIR imaging than the others [250]. Ghosh et al. developed a M13-stabilized SWCNTs probe, which exhibited in vivo precise target to tumor nodules expressing secreted protein, acidic and rich in cysteines (SPARCs) [232]. Second-window NIR light (NIR-II) was used in this study, as the fluorescence source to avoid optical scattering and gain deeper tissue penetration effect during NIR-F imaging. The diagnostic result achieved using this NIR2-emitting M13@SWCNTs probe evidenced outstanding signal-to-noise performance and high specificity towards in situ ovarian tumor and tumor nodules present on the surfaces of other peritoneal organs. Welsher et al.[233] combined SWCNTs phospholipid-PEG (PL@PEG) achieving a biocompatible nanoplatform (SWCNTs@PL@PEG) with low toxicity levels and enhanced stability. Upon its administration in live mice and the application of an InGaAs camera, high-resolution intravital tumor vessel images were acquired.

7.7. CNTs in Nano Biosensors

Biosensors are studied starting from the year 1962[251], and recently, they are considered very attractive tools, mainly due to their easy application, fast response, and low cost[252]. Nano-biosensors (NBSs) are biosensors, consisting of both biological recognition elements and nanomaterials, sized 1-100 nm[253,254]. The unparalleled electrical conductivity, excellent electrocatalytic properties, high stability, slow oxidation kinetics and good modifiability of CNTs, render them a class of nanomaterial with great potential for applications as NBs in cancer diagnosis[255].

7.8. CNTs Combination with Metallic Nanoparticles

Gold and silver nanoparticles (NPs) (AuNPs and AgNPs) are used to modify CNTs, to allow the construction of NBs. Rawashdeh et al. reported on AuNPs@MWCNTs which worked well as NBs, demonstrating easy and improved detection of micro ribonucleic acid named miR-21, which is of paramount importance in the early diagnosis of pancreatic cancer [234]. AuNPs@MWCNTs had a limit of detection (LOD) as low as 3.68 femtomolar (fM) using the source measure unit (SMU).

7.9. CNTs Combination with Antibody

Nano-immunosensors (NISs) based on antibodies are a particular type of NBSs used in cancer diagnosis. Osteopontin (OPN) is a biomarker which is used to detect prostate cancer cells, and its levels are important to predict the survival time of patients bearing prostate cancer [256,257]. To overcome the issues which affect the traditional method used for the measurement of OPN (ELISA assay), Sharma et al. covalently combined OPN monoclonal antibodies onto the SWCNTs@COOH surface, achieving a NIS to determine OPN for prostate cancer diagnosis [235]. AntiOPN@SWCNTs@COOH exhibited high specificity to only OPN, broad detection range and low LOD.

7.10. CNTs Combination with Peptide Reporter

Detecting CDK1 peptide in tissues and organs could be of great help in tumors early diagnosis. Anyway, diagnostic methods using traditional BSs for the detection and quantification of activities of CDK1 poses great difficulties, due to the lack of sensitivity and specificity [258]. Aiming at overcoming these issues, functionalized MWCNTs further modified with a fluorescent peptide reporter specific to CDK1 were developed for the sensitive and fluorescence-based quantification of CDK1 activity via fluorescence imaging [236]. The as prepared NBS provided both easy detection of CDK1, reported the enzymatic activity of CDK1 and detected the CDK/cyclin activity in numerous cancer types, usually associated with poor prognosis.

7.11. CNTs Combination with Multiple Modifications

Multiple modifications on CNTs can further improve the diagnostic effects obtained using the NBSs engineered via single modification. Mahmoodi et al. developed a rGO@MWCNTs/L@Cys@AuNPs multifunctional nanocomposite and enriched its surface with a single strand DNA (ssDNA) probe, as an electrochemical DNA-biosensor to diagnose HPV-18 [237]. rGO@MWCNTs/L@Cys@AuNPs@ssDNA demonstrated high selectivity and sensitivity to HPV-18, towards both the extracted DNA from HPV-18 patients and the synthetic one. Overall, this constructed rGO@MWCNTs/L@Cys@AuNPs@ssDNA NBS represents an essential tool for the early, easy, rapid, and accurate diagnosis of HPV-18. Table 14 summarizes other relevant case studies on the application of CNTs as sensors for early diagnosis of cancer.

Table 14. Carbon nanotube-based cancer detection techniques.

CNTs	Cell line/Biomarkers	Linear Range	LoD	Techniques	Ref.
SWCNTs	PSA	N.R.	250 pg/mL	Electrochemical	[259]
SWCNTs	PSA	0.4–40 pg/mL	4 pg/mL	Immune sensing	[260]
MWCNTs	AFP	0.02–2.0 ng/mL	8.0 pg/mL	Immune sensing	[261]
MWCNTs	CEA	0.5–15.0 and 15.0–200 ng/mL	0.01 ng/mL	Immune sensing	[262]
MWCNTs	AFP	0.1–15.0 and 15.0–200.0 ng/mL	0.08 ng/mL	Immune sensing	[263]
MWCNTs	CA 19-9	12.5–270.0 U/mL	8.3 U/mL	Immune sensing	[264]
MWCNTs	hCG	Up to 600 mIU/mL	14.6 mIU/mL	Electrochemical	[265]
MWCNTs	hCG	0.8–500 mIU/mL	0.3 mIU/mL	Electrochemical	[266]
CNTs	PSA	1–100 ng/mL	1.0 ng/mL	Electrochemical	[267]
MWCNTs	CA 125	1.0–30 and 30–150 U/mL	0.36 U/mL	Electrochemical	[268]
CNTs	AFP	1–55 ng/mL	0.6 ng/mL	Immune sensing	[269]
MWCNTs	CA19-9	0–1000 U/mL	N.R.	Electrochemical	[270]
CNTs	GP73	0–80 ng/mL	58.1 pg/mL	Immune sensing	[271]
CNTs	AFP	0–64 ng/mL	47.1 pg/mL	Immune sensing	[271]
CNTs	AKT2 gene	1 pM–1 μ M	2 fM	Electrochemical	[272]

CNTs	Cell line/Biomarkers	Linear Range	LoD	Techniques	Ref.
CNTs	CA 125	0.001–0.1 ng/mL/0.1–30 ng/mL	0.5 pg/mL	Electrochemical	[273]
CNTs	Cyfra 21-1	0.1–10,000 ng/mL	0.5 ng/mL	Fluorescence	[274]
CNTs	HepG2	10–10 ⁵ cells/mL	5 cells/mL	Electrochemical	[275]

N.R. = Not reported.

8. Clinical Translatability and Challenges Concerning CNTs

The clinical translatability (CT) of carbon nanomaterials (CNMs), including CNTs, is a relevant issue which must be considered, but many problems, such as their biodegradability and metabolism remain unsolved and should be addressed before their successful translation in clinical practice. CNMs are unprecedented effective tools with nonpareil properties in cancer theragnostic. Anyway, the gap between laboratory research and clinical transformation still needs long time a more in deep research. CNMs toxicity, mainly dependent on their sizes, structure and shape, surface chemistry, and on the type of cells target[276], should be taken into great consideration when they have been developed for clinical biomedical applications. Research should be focused on finding methods to prepare ideal CNMs structures without defects, impurities and low cytotoxicity mainly tuning size and designing suitable decorations. To quicken their translation in clinical practice, their stability in physiological conditions, cellular uptake, biodistribution and accumulation in different tissues and organs should be ascertained in advance. Their transformation and metabolic pathway in vivo, as well as their acute and latent toxicity should be clarified [277]. To obtain the desired theragnostic goals, the delivery efficiency of CNMs of therapeutic agents specifically at tumor site is of paramount importance. A suitable CNMs size is pivotal for realizing enhanced permeability and retention (EPR)-mediated passive tumor targeting [278]. Conversely, to achieve active targeting, a deep study of the receptors over-expressed specifically on tumor cells surface is mandatory. Once CNMs-based nanoplatfroms capable of efficient therapeutics delivery are realized, the endogenous or exogenous stimuli should be fully exploited to accomplish controlled drug release. Unfortunately, many obstacles still exist hampering the achievement of these goals, such complicated designing, boring synthesis, scarce synergistic functions, poor integrated efficiency, and ambiguous biological response. Also, while the anti-tumor properties of CNMs have been extensively demonstrated [279], their antineoplastic mechanism and their tumor proliferation and metastasis regulation machinery have not been systematically clarified. In this regard, before the direct clinical application of CNMs as anticancer drugs, their metabolic pathway, biological safety and how they can be used as anticancer agents for cancer treatment to overcome the limitations of conventional chemotherapeutics should be studied and clarified. Moreover, because of hydrophobicity and structural stability, CNMs suffer of low solubility and dispersibility in aqueous media, poor miscibility and tendency to self-aggregate, which mostly limit their biological applications [280]. When CNTs are applied as drug delivery systems (DDSs), due to their stability, free CNTs survive in the body for long time after drug dissociation, thus triggering secondary damage [281]. Several are the problems which impede CNTs clinical application. They comprise the following:

1. the mechanisms of action of CNTs on normal cells and tissues are not fully unveiled
2. potential toxic effects can occur, due to their special structure
3. the mechanisms of CNT-induced toxicity are not fully clarified
4. the biodegradability of raw CNTs is too low because their hydrophobic prevents enzymes from approaching them, thus impeding enzymatic degradation



To promote CNTs translatability in clinic practice, more labors is necessary to explore the best functional molecules for the CNTs modification to improve their biocompatibility, to avoid the toxic and risk factors, to enhance their solubility and create defect sites, which offer desired binding sites for enzymes and promoting enzymatic degradation [282]. Collectively, although there is no clinical practice that have ever reported the application of CNMs as DDSs in cancer theragnostic, there are some clinical trials cases that assessed the employment of carbon nanoparticles (CNPs) in lymphatic monitoring during colorectal cancer surgeries, lymph nodes collection in advanced gastric cancer, and lymph node biopsy of papillary thyroid carcinoma, confirming the unprecedented potential and promising future of CNMs and CNTs in cancer theragnostic [283–285]. The following Table 15 and 16 summarize other two clinical trials carried out to evaluate the possible applications of CNTs-based nanocomposites in anticancer treatments and cancer diagnosis.


Table 15. Clinical trials assessing the possible application of CNTs in anticancer therapy.

Trial Number	Sponsor	Device/Intervention	Problem	Participants	Study Status	Ending year	Ref
NCT01773850 *	UNC Lineberger Comprehensive Cancer Center	CNT x-ray source array for SDT of BN	BN	54	Completed	2018	[286]
NCT01420588 **	Anhui Medical University	Of-AUNPs/CNTs-based chemical nano sensors	PCGLs	1000	Completed	2020	[287,288]

SDT = Stationary digital tomosynthesis; BN = breast neoplasm; PCGLs = pre-cancerous gastric lesions; AuNPs = gold nanoparticles; CNTs = carbon nanotubes; *Of* = organically functionalized; * study title: Stationary carbon nanotube X-ray digital breast tomosynthesis scanner; ** study title: Diagnosis of gastric lesions from exhaled breath and saliva.

Table 16. Study type assessment.

Trial Number	Sponsor	Study Type	Ethics Approval	Status	Ref
NCT01773850 *	UNC Lineberger Comprehensive Cancer Center	OCPS		Completed	[286]
NCT01420588 **	Anhui Medical University	OCPS		Completed	[287,288]

OCPS = Observational cohort prospective study * study title: Stationary carbon nanotube X-ray digital breast tomosynthesis scanner; ** study title: Diagnosis of gastric lesions from exhaled breath and saliva;  = positive ethical approval.

Other clinical trials regarded the application of CNTs to treat dental caries, Parkinson (available online at <https://clinicaltrials.gov/study/NCT02873585>, <https://clinicaltrials.gov/study/NCT01246336>. Accessed on 26 May 2025) and cervical pain[289]. The assessment of the clinical observational studies reported in Table 15 focused on the potential for misleading, selection of participants, measurement of exposures and outcomes, handling of missing data, and selective reporting of results. This comprehensive approach ensured a thorough examination of factors that could influence the results
















of the study, from the allocation of participants to the reporting of the outcomes. Each domain was rated as “low risk ,” “high risk ,” or “some concerns ,” based on specific criteria[290,291]. This assessment processes emphasized the importance of transparency and rigor. Using the Cochrane Risk of Bias, two reviewers independently assessed the risk of bias (Table 17).

Table 17. Risk of bias in randomized studies as assessed by the Cochrane Collaboration’s “Risk of Bias” tool.

Trial Number	RSG	AC	BPP	BOA	IOD	SR	Ref
NCT01773850							[286]
NCT01420588							[287,288]

RSG = Random sequence generation; AC = allocation concealment; BPP = blinding of Participants and personnel; BOA = blinding of outcomes assessment; IOD = incomplete outcome data; SR = selective reporting.

Supplementary Materials: The following supporting information can be downloaded at the website of this paper posted on Preprints.org. Table S1. Structural properties of various modified/activated CNTs vs pristine CNTs (first row). Table S2. Main methods to synthesize carbon nanotubes (CNTs). Table S3. Most relevant research articles on CNT synthesis from biomass by different methods, operating parameters and main properties.

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References

1. Siegel, R.L.; Miller, K.D.; Goding Sauer, A.; Fedewa, S.A.; Butterly, L.F.; Anderson, J.C.; Cercek, A.; Smith, R.A.; Jemal, A. Colorectal Cancer Statistics, 2020. *CA Cancer J Clin* 2020, 70, doi:10.3322/caac.21601.
2. Bray, F.; Laversanne, M.; Sung, H.; Ferlay, J.; Siegel, R.L.; Soerjomataram, I.; Jemal, A. Global Cancer Statistics 2022: GLOBOCAN Estimates of Incidence and Mortality Worldwide for 36 Cancers in 185 Countries. *CA Cancer J Clin* 2024, 74, 229–263, doi:10.3322/caac.21834.
3. Bray, F.; Laversanne, M.; Weiderpass, E.; Soerjomataram, I. The Ever-increasing Importance of Cancer as a Leading Cause of Premature Death Worldwide. *Cancer* 2021, 127, 3029–3030, doi:10.1002/cncr.33587.
4. Chen, S.; Cao, Z.; Prettnner, K.; Kuhn, M.; Yang, J.; Jiao, L.; Wang, Z.; Li, W.; Geldsetzer, P.; Bärnighausen, T.; et al. Estimates and Projections of the Global Economic Cost of 29 Cancers in 204 Countries and Territories from 2020 to 2050. *JAMA Oncol* 2023, 9, doi:10.1001/jamaoncol.2022.7826.
5. Naief, M.F.; Mohammed, S.N.; Mayouf, H.J.; Mohammed, A.M. A Review of the Role of Carbon Nanotubes for Cancer Treatment Based on Photothermal and Photodynamic Therapy Techniques. *J Organomet Chem* 2023, 999.
6. Klochkov, S.G.; Neganova, M.E.; Nikolenko, V.N.; Chen, K.; Somasundaram, S.G.; Kirkland, C.E.; Aliev, G. Implications of Nanotechnology for the Treatment of Cancer: Recent Advances. *Semin Cancer Biol* 2021, 69.
7. Chen, K.; Cao, X.; Li, M.; Su, Y.; Li, H.; Xie, M.; Zhang, Z.; Gao, H.; Xu, X.; Han, Y.; et al. A TRAIL-Delivered Lipoprotein-Bioinspired Nanovector Engineering Stem Cell-Based Platform for Inhibition of Lung Metastasis of Melanoma. *Theranostics* 2019, 9, doi:10.7150/thno.31157.

8. Wang, W.; Chen, K.; Su, Y.; Zhang, J.; Li, M.; Zhou, J. Lysosome-Independent Intracellular Drug/Gene Codelivery by Lipoprotein-Derived Nanovector for Synergistic Apoptosis-Inducing Cancer-Targeted Therapy. *Biomacromolecules* 2018, 19, doi:10.1021/acs.biomac.7b01549.
9. Ganesh, K.; Massagué, J. Targeting Metastatic Cancer. *Nat Med* 2021, 27.
10. Shakibaie, M.; Hajighasemi, E.; Adeli-Sardou, M.; Doostmohammadi, M.; Forootanfar, H. Antimicrobial and Anti-Biofilm Activities of Bi Subnitrate and BiNPs Produced by *Delftia* Sp. SFG against Clinical Isolates of *Staphylococcus Aureus*, *Pseudomonas Aeruginosa*, and *Proteus Mirabilis*. *IET Nanobiotechnol* 2019, 13, doi:10.1049/iet-nbt.2018.5102.
11. Mohammed, S.N.; Al-Rawi, K.F.; Mohammed, A.M. Kinetic Evaluation and Study of Gold-Based Nanoparticles and Multi-Walled Carbon Nanotubes as an Alkaline Phosphatase Inhibitor in Serum and Pure Form. *Carbon Letters* 2023, 33, doi:10.1007/s42823-023-00509-z.
12. faiad naief, M.; Mishaal Mohammed, A.; Khalaf, Y.H. Kinetic and Thermodynamic Study of ALP Enzyme in the Presence and Absence MWCNTs and Pt-NPs Nanocomposites. *Results Chem* 2023, 5, doi:10.1016/j.rechem.2023.100844.
13. Naief, M.F.; Mohammed, S.N.; Ahmed, Y.N.; Mohammed, A.M.; Mohammed, S.N.; Mohammed, S.N. Novel Preparation Method of Fullerene and Its Ability to Detect H₂S and NO₂ Gases. *Results Chem* 2023, 5, doi:10.1016/j.rechem.2023.100924.
14. Naief, M.F.; Mohammed, S.N.; Ahmed, Y.N.; Mohammed, A.M. Synthesis and Characterisation of MWCNTCOOH and Investigation of Its Potential as Gas Sensor. *Inorg Chem Commun* 2023, 157, doi:10.1016/j.inoche.2023.111338.
15. Latif, Z.A.A.; Mohammed, A.M.; Abbass, N.M. Synthesis and Characterization of Polymer Nanocomposites from Methyl Acrylate and Metal Chloride and Their Application. *Polymer Bulletin* 2020, 77, doi:10.1007/s00289-019-03047-9.
16. Abdullah, O.H.; Mohammed, A.M. Green Synthesis of Aunps from the Leaf Extract of *Prosopis Farcta* for Antibacterial and Anti-Cancer Applications. *Dig J Nanomater Biostruct* 2020, 15, doi:10.15251/djnb.2020.153.943.
17. Nasir, G.A.; Najm, M.A.; Hussein, A.L. The Effect of Silver Nanoparticles on Braf Gene Expression. *Systematic Reviews in Pharmacy* 2020, 11, doi:10.31838/srp.2020.10.85.
18. Mohammed, A.M.; Saud, W.M.; Ali, M.M. Green Synthesis of Fe₂O₃ Nanoparticles Using *Olea Europaea* Leaf Extract and Their Antibacterial Activity. *Dig J Nanomater Biostruct* 2020, 15, doi:10.15251/djnb.2020.151.175.
19. Jwied, D.H.; Nayef, U.M.; Mutlak, F.A.H. Synthesis of C:Se Nanoparticles Ablated on Porous Silicon for Sensing NO₂ and NH₃ Gases. *Optik (Stuttg)* 2021, 241, doi:10.1016/j.ijleo.2021.167013.
20. Altaee, M.F.; Yaaqoob, L.A.; Kamona, Z.K. Evaluation of the Biological Activity of Nickel Oxide Nanoparticles as Antibacterial and Anticancer Agents. *Iraqi Journal of Science* 2020, 61, doi:10.24996/ijis.2020.61.11.12.
21. Ghdeeb, N.J.; AbdulMajeed, A.M.; Mohammed, A.H. Role of Extracted Nano-Metal Oxides From Factory Wastes In Medical Applications. *Iraqi Journal of Science* 2023, 64, doi:10.24996/ijis.2023.64.4.12.
22. Al-Ali, A.A.A.; Alsalami, K.A.S.; Athbi, A.M. Cytotoxic Effects of CeO₂ NPs and β -Carotene and Their Ability to Induce Apoptosis in Human Breast Normal and Cancer Cell Lines. *Iraqi Journal of Science* 2022, 63, doi:10.24996/ijis.2022.63.3.2.
23. Salih, A.M.A.; Abdo, G.M. Water Conservation in the Arab Region. In *Water, Energy and Food Sustainability in the Middle East: The Sustainability Triangle*; 2017.
24. Iijima, S. Helical Microtubules of Graphitic Carbon. *Nature* 1991, 354, 56–58, doi:10.1038/354056a0.
25. Alfei, S.; Schito, G.C. Antimicrobial Nanotubes: From Synthesis and Promising Antimicrobial Upshots to Unanticipated Toxicities, Strategies to Limit Them, and Regulatory Issues. *Nanomaterials* 2025, 15, 633, doi:10.3390/nano15080633.
26. Alfei, S.; Schito, G.C. Nanotubes: Carbon-Based Fibers and Bacterial Nano-Conduits Both Arousing a Global Interest and Conflicting Opinions. *Fibers* 2022, 10, 75, doi:10.3390/fib10090075.
27. Hussain, A.; Haider, Q.; Rehman, A.; Ahmad, H.; Baili, J.; Aljahdaly, N.H.; Hassan, A. A Thermal Conductivity Model for Hybrid Heat and Mass Transfer Investigation of Single and Multi-Wall Carbon

- Nano-Tubes Flow Induced by a Spinning Body. *Case Studies in Thermal Engineering* 2021, 28, 101449, doi:10.1016/j.csite.2021.101449.
28. Sobamowo, M.G.; Akanmu, J.O.; Adeleye, O.A.; Akingbade, S.A.; Yinusa, A.A. Coupled Effects of Magnetic Field, Number of Walls, Geometric Imperfection, Temperature Change, and Boundary Conditions on Nonlocal Nonlinear Vibration of Carbon Nanotubes Resting on Elastic Foundations. *Forces in Mechanics* 2021, 3, 100010, doi:10.1016/j.finmec.2021.100010.
 29. Abd-Elsalam, K.A. Carbon Nanomaterials: 30 Years of Research in Agroecosystems. In *Carbon Nanomaterials for Agri-Food and Environmental Applications*; Elsevier, 2020; pp. 1–18.
 30. Anzar, N.; Hasan, R.; Tyagi, M.; Yadav, N.; Narang, J. Carbon Nanotube - A Review on Synthesis, Properties and Plethora of Applications in the Field of Biomedical Science. *Sensors International* 2020, 1, 100003, doi:10.1016/j.sintl.2020.100003.
 31. Negri, V.; Pacheco-Torres, J.; Calle, D.; López-Larrubia, P. Carbon Nanotubes in Biomedicine. *Top Curr Chem* 2020, 378, 15, doi:10.1007/s41061-019-0278-8.
 32. Lamberti, M.; Pedata, P.; Sannolo, N.; Porto, S.; De Rosa, A.; Caraglia, M. Carbon Nanotubes: Properties, Biomedical Applications, Advantages and Risks in Patients and Occupationally-Exposed Workers. *Int J Immunopathol Pharmacol* 2015, 28, 4–13, doi:10.1177/0394632015572559.
 33. Kumar, S.; Rani, R.; Dilbaghi, N.; Tankeshwar, K.; Kim, K.-H. Carbon Nanotubes: A Novel Material for Multifaceted Applications in Human Healthcare. *Chem Soc Rev* 2017, 46, 158–196, doi:10.1039/C6CS00517A.
 34. Su, Q.; Gan, L.; Liu, J.; Yang, X. Carbon Dots Derived from Pea for Specifically Binding with *Cryptococcus Neoformans*. *Anal Biochem* 2020, 589, 113476, doi:10.1016/j.ab.2019.113476.
 35. Salari, S.; Jafari, S.M. Application of Nanofluids for Thermal Processing of Food Products. *Trends Food Sci Technol* 2020, 97, 100–113, doi:10.1016/j.tifs.2020.01.004.
 36. Chen, D.; Dougherty, C.A.; Zhu, K.; Hong, H. Theranostic Applications of Carbon Nanomaterials in Cancer: Focus on Imaging and Cargo Delivery. *Journal of Controlled Release* 2015, 210.
 37. Law, S.K.; Leung, A.W.N.; Xu, C.; Au, D.C.T. Is It Possible for Curcumin to Conjugate with a Carbon Nanotube in Photodynamic Therapy? *Photodiagnosis Photodyn Ther* 2022, 40.
 38. Deshmukh, M.A.; Jeon, J.Y.; Ha, T.J. Carbon Nanotubes: An Effective Platform for Biomedical Electronics. *Biosens Bioelectron* 2020, 150, doi:10.1016/j.bios.2019.111919.
 39. Costa, P.M.; Bourgoignon, M.; Wang, J.T.W.; Al-Jamal, K.T. Functionalized Carbon Nanotubes: From Intracellular Uptake and Cell-Related Toxicity to Systemic Brain Delivery. *Journal of Controlled Release* 2016, 241.
 40. Vashist, A.; Kaushik, A.; Vashist, A.; Sagar, V.; Ghosal, A.; Gupta, Y.K.; Ahmad, S.; Nair, M. Advances in Carbon Nanotubes–Hydrogel Hybrids in Nanomedicine for Therapeutics. *Adv Healthc Mater* 2018, 7.
 41. Cirillo, G.; Vittorio, O.; Kunhardt, D.; Valli, E.; Voli, F.; Farfalla, A.; Curcio, M.; Spizzirri, U.G.; Hampel, S. Combining Carbon Nanotubes and Chitosan for the Vectorization of Methotrexate to Lung Cancer Cells. *Materials* 2019, 12, doi:10.3390/ma12182889.
 42. Zhang, J.; Liu, Z.; Zhou, S.; Teng, Y.; Zhang, X.; Li, J. Novel Span-PEG Multifunctional Ultrasound Contrast Agent Based on CNTs as a Magnetic Targeting Factor and a Drug Carrier. *ACS Omega* 2020, 5, doi:10.1021/acsomega.0c03325.
 43. Garriga, R.; Herrero-Contiente, T.; Palos, M.; Cebolla, V.L.; Osada, J.; Muñoz, E.; Rodríguez-Yoldi, M.J. Toxicity of Carbon Nanomaterials and Their Potential Application as Drug Delivery Systems: In Vitro Studies in Caco-2 and MCF-7 Cell Lines. *Nanomaterials* 2020, 10, doi:10.3390/nano10081617.
 44. Corletto, A.; Shapter, J.G. Nanoscale Patterning of Carbon Nanotubes: Techniques, Applications, and Future. *Advanced Science* 2021, 8.
 45. Azizi-Lalabadi, M.; Hashemi, H.; Feng, J.; Jafari, S.M. Carbon Nanomaterials against Pathogens; the Antimicrobial Activity of Carbon Nanotubes, Graphene/Graphene Oxide, Fullerenes, and Their Nanocomposites. *Adv Colloid Interface Sci* 2020, 284, 102250, doi:10.1016/j.cis.2020.102250.
 46. Alfei, S.; Zuccari, G. Carbon-Nanotube-Based Nanocomposites in Environmental Remediation: An Overview of Typologies and Applications and an Analysis of Their Paradoxical Double-Sided Effects. *J Xenobiot* 2025, 15, 76, doi:10.3390/jox15030076.

47. Griffiths, O.G.; O'Byrne, J.P.; Torrente-Murciano, L.; Jones, M.D.; Mattia, D.; McManus, M.C. Identifying the Largest Environmental Life Cycle Impacts during Carbon Nanotube Synthesis via Chemical Vapour Deposition. *J Clean Prod* 2013, 42, doi:10.1016/j.jclepro.2012.10.040.
48. Wang, Z.; Shen, D.; Wu, C.; Gu, S. State-of-the-Art on the Production and Application of Carbon Nanomaterials from Biomass. *Green Chemistry* 2018, 20, doi:10.1039/c8gc01748d.
49. Berber, M.R.; Elkhenany, H.; Hafez, I.H.; El-Badawy, A.; Essawy, M.; El-Badri, N. Efficient Tailoring of Platinum Nanoparticles Supported on Multiwalled Carbon Nanotubes for Cancer Therapy. *Nanomedicine* 2020, 15, doi:10.2217/nmm-2019-0445.
50. Zhang, J.; Song, L.; Zhou, S.; Hu, M.; Jiao, Y.; Teng, Y.; Wang, Y.; Zhang, X. Enhanced Ultrasound Imaging and Anti-Tumor: In Vivo Properties of Span-Polyethylene Glycol with Folic Acid-Carbon Nanotube-Paclitaxel Multifunctional Microbubbles. *RSC Adv* 2019, 9, doi:10.1039/c9ra06437k.
51. Behzadpour, N.; Ranjbar, A.; Azarpira, N.; Sattarahmady, N. Development of a Composite of Polypyrrole-Coated Carbon Nanotubes as a Sonosensitizer for Treatment of Melanoma Cancer Under Multi-Step Ultrasound Irradiation. *Ultrasound Med Biol* 2020, 46, 2322–2334, doi:10.1016/J.ULTRASMEDBIO.2020.05.003.
52. Zhang, B.; Wang, H.; Shen, S.; She, X.; Shi, W.; Chen, J.; Zhang, Q.; Hu, Y.; Pang, Z.; Jiang, X. Fibrin-Targeting Peptide CREKA-Conjugated Multi-Walled Carbon Nanotubes for Self-Amplified Photothermal Therapy of Tumor. *Biomaterials* 2016, 79, 46–55, doi:10.1016/J.BIOMATERIALS.2015.11.061.
53. Sobhani, Z.; Behnam, M.A.; Emami, F.; Dehghanian, A.; Jamhiri, I. Photothermal Therapy of Melanoma Tumor Using Multiwalled Carbon Nanotubes. *Int J Nanomedicine* 2017, 12, doi:10.2147/IJN.S134661.
54. Bosca, F.; Corazzari, I.; Foglietta, F.; Canaparo, R.; Durando, G.; Pastero, L.; Arpicco, S.; Dosio, F.; Zonari, D.; Cravotto, G.; et al. SWCNT-Porphyrin Nano-Hybrids Selectively Activated by Ultrasound: An Interesting Model for Sonodynamic Applications. *RSC Adv* 2020, 10, doi:10.1039/d0ra03944f.
55. Singh, R.; Kumar, S. Cancer Targeting and Diagnosis: Recent Trends with Carbon Nanotubes. *Nanomaterials* 2022, 12.
56. Faraji Dizaji, B.; Khoshbakht, S.; Farboudi, A.; Azarbaijan, M.H.; Irani, M. Far-Reaching Advances in the Role of Carbon Nanotubes in Cancer Therapy. *Life Sci* 2020, 257, 118059, doi:10.1016/J.LFS.2020.118059.
57. Son, K.H.; Hong, J.H.; Lee, J.W. Carbon Nanotubes as Cancer Therapeutic Carriers and Mediators. *Int J Nanomedicine* 2016, 11.
58. Zhang, Y.R.; Lin, R.; Li, H.J.; He, W. ling; Du, J.Z.; Wang, J. Strategies to Improve Tumor Penetration of Nanomedicines through Nanoparticle Design. *Wiley Interdiscip Rev Nanomed Nanobiotechnol* 2019, 11.
59. Zhang, A.; Meng, K.; Liu, Y.; Pan, Y.; Qu, W.; Chen, D.; Xie, S. Absorption, Distribution, Metabolism, and Excretion of Nanocarriers in Vivo and Their Influences. *Adv Colloid Interface Sci* 2020, 284.
60. Zare, H.; Ahmadi, S.; Ghasemi, A.; Ghanbari, M.; Rabiee, N.; Bagherzadeh, M.; Karimi, M.; Webster, T.J.; Hamblin, M.R.; Mostafavi, E. Carbon Nanotubes: Smart Drug/Gene Delivery Carriers. *Int J Nanomedicine* 2021, 16.
61. Taghavi, S.; Nia, A.H.; Abnous, K.; Ramezani, M. Polyethylenimine-Functionalized Carbon Nanotubes Tagged with AS1411 Aptamer for Combination Gene and Drug Delivery into Human Gastric Cancer Cells. *Int J Pharm* 2017, 516, doi:10.1016/j.ijpharm.2016.11.027.
62. Huang, Y.P.; Lin, I.J.; Chen, C.C.; Hsu, Y.C.; Chang, C.C.; Lee, M.J. Delivery of Small Interfering RNAs in Human Cervical Cancer Cells by Polyethylenimine-Functionalized Carbon Nanotubes. *Nanoscale Res Lett* 2013, 8, doi:10.1186/1556-276X-8-267.
63. Bhirde, A.A.; Chikkaveeraiah, B. V.; Srivatsan, A.; Niu, G.; Jin, A.J.; Kapoor, A.; Wang, Z.; Patel, S.; Patel, V.; Gorbach, A.M.; et al. Targeted Therapeutic Nanotubes Influence the Viscoelasticity of Cancer Cells to Overcome Drug Resistance. *ACS Nano* 2014, 8, doi:10.1021/nn501223q.
64. Dong, X.; Liu, L.; Zhu, D.; Zhang, H.; Leng, X. Transactivator of Transcription (TAT) Peptide– Chitosan Functionalized Multiwalled Carbon Nanotubes as a Potential Drug Delivery Vehicle for Cancer Therapy. *Int J Nanomedicine* 2015, 10, doi:10.2147/IJN.S81762.
65. Cheng, Q.; Blais, M.O.; Harris, G.M.; Jabbarzadeh, E. Correction: PLGA-Carbon Nanotube Conjugates for Intercellular Delivery of Caspase-3 into Osteosarcoma Cells (PLoS ONE (2013) 8, 12, DOI: 10.1371/Journal.Pone.0081947). *PLoS One* 2014, 9.

66. Wang, C.H.; Chiou, S.H.; Chou, C.P.; Chen, Y.C.; Huang, Y.J.; Peng, C.A. Photothermolysis of Glioblastoma Stem-like Cells Targeted by Carbon Nanotubes Conjugated with CD133 Monoclonal Antibody. *Nanomedicine* 2011, 7, 69–79, doi:10.1016/J.NANO.2010.06.010.
67. Shao, N.; Lu, S.; Wickstrom, E.; Panchapakesan, B. Integrated Molecular Targeting of IGF1R and HER2 Surface Receptors and Destruction of Breast Cancer Cells Using Single Wall Carbon Nanotubes. *Nanotechnology* 2007, 18, doi:10.1088/0957-4484/18/31/315101.
68. Koh, B.; Park, S.B.; Yoon, E.; Yoo, H.M.; Lee, D.; Heo, J.N.; Ahn, S. AV β 3-Targeted Delivery of Camptothecin-Encapsulated Carbon Nanotube-Cyclic RGD in 2D and 3D Cancer Cell Culture. *J Pharm Sci* 2019, 108, 3704–3712, doi:10.1016/J.XPHS.2019.07.011.
69. Bhirde, A.A.; Patel, V.; Gavard, J.; Zhang, G.; Sousa, A.A.; Masedunskas, A.; Leapman, R.D.; Weigert, R.; Gutkind, J.S.; Rusling, J.F. Targeted Killing of Cancer Cells in Vivo and in Vitro with EGF-Directed Carbon Nanotube-Based Drug Delivery. *ACS Nano* 2009, 3, doi:10.1021/nn800551s.
70. Yang, S.; Wang, Z.; Ping, Y.; Miao, Y.; Xiao, Y.; Qu, L.; Zhang, L.; Hu, Y.; Wang, J. PEG/PEI-Functionalized Single-Walled Carbon Nanotubes as Delivery Carriers for Doxorubicin: Synthesis, Characterization, and in Vitro Evaluation. *Beilstein Journal of Nanotechnology* 2020, 11, doi:10.3762/BJNANO.11.155.
71. Liu, Z.; Sun, X.; Nakayama-Ratchford, N.; Dai, H. Supramolecular Chemistry on Water- Soluble Carbon Nanotubes for Drug Loading and Delivery. *ACS Nano* 2007, 1, doi:10.1021/nn700040t.
72. Rahamathulla, M.; Bhosale, R.R.; Osmani, R.A.M.; Mahima, K.C.; Johnson, A.P.; Hani, U.; Ghazwani, M.; Begum, M.Y.; Alshehri, S.; Ghoneim, M.M.; et al. Carbon Nanotubes: Current Perspectives on Diverse Applications in Targeted Drug Delivery and Therapies. *Materials* 2021, 14.
73. Mahajan, S.; Patharkar, A.; Kuche, K.; Maheshwari, R.; Deb, P.K.; Kalia, K.; Tekade, R.K. Functionalized Carbon Nanotubes as Emerging Delivery System for the Treatment of Cancer. *Int J Pharm* 2018, 548, 540–558, doi:10.1016/J.IJPHARM.2018.07.027.
74. Barzegar, A.; Mansouri, A.; Azamat, J. Molecular Dynamics Simulation of Non-Covalent Single-Walled Carbon Nanotube Functionalization with Surfactant Peptides. *J Mol Graph Model* 2016, 64, 75–84, doi:10.1016/J.JMGM.2016.01.003.
75. Hasnain, M.S.; Ahmad, S.A.; Hoda, M.N.; Rishishwar, S.; Rishishwar, P.; Nayak, A.K. Stimuli-Responsive Carbon Nanotubes for Targeted Drug Delivery. *Stimuli Responsive Polymeric Nanocarriers for Drug Delivery Applications: Volume 2: Advanced Nanocarriers for Therapeutics* 2019, 321–344, doi:10.1016/B978-0-08-101995-5.00015-5.
76. Cao, X.; Tao, L.; Wen, S.; Hou, W.; Shi, X. Hyaluronic Acid-Modified Multiwalled Carbon Nanotubes for Targeted Delivery of Doxorubicin into Cancer Cells. *Carbohydr Res* 2015, 405, 70–77, doi:10.1016/J.CARRES.2014.06.030.
77. Gu, Y.J.; Cheng, J.; Jin, J.; Cheng, S.H.; Wong, W.T. Development and Evaluation of PH-Responsive Single-Walled Carbon Nanotube-Doxorubicin Complexes in Cancer Cells. *Int J Nanomedicine* 2011, 6, doi:10.2147/ijn.s25162.
78. Fernández, M.; Javaid, F.; Chudasama, V. Advances in Targeting the Folate Receptor in the Treatment/Imaging of Cancers. *Chem Sci* 2018, 9.
79. Lu, Y.J.; Wei, K.C.; Ma, C.C.M.; Yang, S.Y.; Chen, J.P. Dual Targeted Delivery of Doxorubicin to Cancer Cells Using Folate-Conjugated Magnetic Multi-Walled Carbon Nanotubes. *Colloids Surf B Biointerfaces* 2012, 89, 1–9, doi:10.1016/J.COLSURFB.2011.08.001.
80. Feazell, R.P.; Nakayama-Ratchford, N.; Dai, H.; Lippard, S.J. Soluble Single-Walled Carbon Nanotubes as Longboat Delivery Systems for Platinum(IV) Anticancer Drug Design. *J Am Chem Soc* 2007, 129, doi:10.1021/ja073231f.
81. Hampel, S.; Kunze, D.; Haase, D.; Krämer, K.; Rauschenbach, M.; Ritschel, M.; Leonhardt, A.; Thomas, J.; Oswald, S.; Hoffmann, V.; et al. Carbon Nanotubes Filled with a Chemotherapeutic Agent: A Nanocarrier Mediates Inhibition of Tumor Cell Growth. *Nanomedicine* 2008, 3, doi:10.2217/17435889.3.2.175.
82. Wu, L.; Man, C.; Wang, H.; Lu, X.; Ma, Q.; Cai, Y.; Ma, W. PEGylated Multi-Walled Carbon Nanotubes for Encapsulation and Sustained Release of Oxaliplatin. *Pharm Res* 2013, 30, 412–423, doi:10.1007/s11095-012-0883-5.

83. Liu, H.; Xu, H.; Wang, Y.; He, Z.; Li, S. Effect of Intratumoral Injection on the Biodistribution and Therapeutic Potential of Novel Chemophor EL-Modified Single-Walled Nanotube Loading Doxorubicin. *Drug Dev Ind Pharm* 2012, 38, doi:10.3109/03639045.2011.637050.
84. Ali-Boucetta, H.; Al-Jamal, K.T.; McCarthy, D.; Prato, M.; Bianco, A.; Kostarelos, K. Multiwalled Carbon Nanotube-Doxorubicin Supramolecular Complexes for Cancer Therapeutics. *Chemical Communications* 2002, 8, doi:10.1039/b712350g.
85. Li, R.; Wu, R.; Zhao, L.; Wu, M.; Yang, L.; Zou, H. P-Glycoprotein Antibody Functionalized Carbon Nanotube Overcomes the Multidrug Resistance of Human Leukemia Cells. *ACS Nano* 2010, 4, doi:10.1021/nn9011225.
86. Arya, N.; Arora, A.; Vasu, K.S.; Sood, A.K.; Katti, D.S. Combination of Single Walled Carbon Nanotubes/Graphene Oxide with Paclitaxel: A Reactive Oxygen Species Mediated Synergism for Treatment of Lung Cancer. *Nanoscale* 2013, 5, doi:10.1039/c3nr33190c.
87. Zhang, X.; Meng, L.; Lu, Q.; Fei, Z.; Dyson, P.J. Targeted Delivery and Controlled Release of Doxorubicin to Cancer Cells Using Modified Single Wall Carbon Nanotubes. *Biomaterials* 2009, 30, doi:10.1016/j.biomaterials.2009.07.025.
88. Atyabi, F.; Sobhani, Adeli, M.; Dinarvand, R.; Ghahremani Increased Paclitaxel Cytotoxicity against Cancer Cell Lines Using a Novel Functionalized Carbon Nanotube. *Int J Nanomedicine* 2011, doi:10.2147/ijn.s17336.
89. Liu, Z.; Chen, K.; Davis, C.; Sherlock, S.; Cao, Q.; Chen, X.; Dai, H. Drug Delivery with Carbon Nanotubes for in Vivo Cancer Treatment. *Cancer Res* 2008, 68, doi:10.1158/0008-5472.CAN-08-1468.
90. Chen, J.; Chen, S.; Zhao, X.; Kuznetsova, L. V.; Wong, S.S.; Ojima, I. Functionalized Single-Walled Carbon Nanotubes as Rationally Designed Vehicles for Tumor-Targeted Drug Delivery. *J Am Chem Soc* 2008, 130, doi:10.1021/ja805570f.
91. Razzazan, A.; Atyabi, F.; Kazemi, B.; Dinarvand, R. In Vivo Drug Delivery of Gemcitabine with PEGylated Single-Walled Carbon Nanotubes. *Materials Science and Engineering C* 2016, 62, doi:10.1016/j.msec.2016.01.076.
92. Bartholomeusz, G.; Cherukuri, P.; Kingston, J.; Cognet, L.; Lemos, R.; Leeuw, T.K.; Gumbiner-Russo, L.; Weisman, R.B.; Powis, G. In Vivo Therapeutic Silencing of Hypoxia-Inducible Factor 1 Alpha (HIF-1 α) Using Single-Walled Carbon Nanotubes Noncovalently Coated with SiRNA. *Nano Res* 2009, 2, doi:10.1007/s12274-009-9026-7.
93. Varkouhi, A.K.; Foillard, S.; Lammers, T.; Schiffelers, R.M.; Doris, E.; Hennink, W.E.; Storm, G. SiRNA Delivery with Functionalized Carbon Nanotubes. *Int J Pharm* 2011, 416, doi:10.1016/j.ijpharm.2011.02.009.
94. Wang, X.; Ren, J.; Qu, X. Targeted RNA Interference of Cyclin A2 Mediated by Functionalized Single-Walled Carbon Nanotubes Induces Proliferation Arrest and Apoptosis in Chronic Myelogenous Leukemia K562 Cells. *ChemMedChem* 2008, 3, doi:10.1002/cmdc.200700329.
95. Guo, C.; Al-Jamal, W.T.; Toma, F.M.; Bianco, A.; Prato, M.; Al-Jamal, K.T.; Kostarelos, K. Design of Cationic Multiwalled Carbon Nanotubes as Efficient SiRNA Vectors for Lung Cancer Xenograft Eradication. *Bioconj Chem* 2015, 26, doi:10.1021/acs.bioconjchem.5b00249.
96. Singh, P.; Samorì, C.; Toma, F.M.; Bussy, C.; Nunes, A.; Al-Jamal, K.T.; Ménard-Moyon, C.; Prato, M.; Kostarelos, K.; Bianco, A. Polyamine Functionalized Carbon Nanotubes: Synthesis, Characterization, Cytotoxicity and SiRNA Binding. *J Mater Chem* 2011, 21, doi:10.1039/c0jm04064a.
97. Mohammadi, M.; Salmasi, Z.; Hashemi, M.; Mosaffa, F.; Abnous, K.; Ramezani, M. Single-Walled Carbon Nanotubes Functionalized with Aptamer and Piperazine-Polyethylenimine Derivative for Targeted SiRNA Delivery into Breast Cancer Cells. *Int J Pharm* 2015, 485, doi:10.1016/j.ijpharm.2015.02.031.
98. Al-Jamal, K.T.; Toma, F.M.; Yilmazer, A.; Ali-Boucetta, H.; Nunes, A.; Herrero, M.A.; Tian, B.; Eddaoui, A.; Al-Jamal, W.; Bianco, A.; et al. Enhanced Cellular Internalization and Gene Silencing with a Series of Cationic Dendron-Multiwalled Carbon Nanotube:SiRNA Complexes. *FASEB Journal* 2010, 24, doi:10.1096/fj.09-141036.
99. Podesta, J.E.; Al-Jamal, K.T.; Herrero, M.A.; Tian, B.; Ali-Boucetta, H.; Hegde, V.; Bianco, A.; Prato, M.; Kostarelos, K. Antitumor Activity and Prolonged Survival by Carbon-Nanotube-Mediated Therapeutic SiRNA Silencing in a Human Lung Xenograft Model. *Small* 2009, 5, doi:10.1002/sml.200801572.

100. Battigelli, A.; Wang, J.T.W.; Russier, J.; Da Ros, T.; Kostarelos, K.; Al-Jamal, K.T.; Prato, M.; Bianco, A. Ammonium and Guanidinium Dendron-Carbon Nanotubes by Amidation and Click Chemistry and Their Use for siRNA Delivery. *Small* 2013, 9, doi:10.1002/smll.201300264.
101. Wang, L.; Shi, J.; Zhang, H.; Li, H.; Gao, Y.; Wang, Z.; Wang, H.; Li, L.; Zhang, C.; Chen, C.; et al. RETRACTED: Synergistic Anticancer Effect of RNAi and Photothermal Therapy Mediated by Functionalized Single-Walled Carbon Nanotubes. *Biomaterials* 2013, 34, 262–274, doi:10.1016/j.biomaterials.2012.09.037.
102. Ahmed, M.; Jiang, X.; Deng, Z.; Narain, R. Cationic Glyco-Functionalized Single-Walled Carbon Nanotubes as Efficient Gene Delivery Vehicles. *Bioconjug Chem* 2009, 20, doi:10.1021/bc900229v.
103. Biris, A.; Karmakar, Braton; Dervishi; Ghosh; Mahmood; Saeed; Mustafa; Casciano, D.; Radominska-Pandya; et al. Ethylenediamine Functionalized-Single-Walled Nanotube (f-SWNT)-Assisted in Vitro Delivery of the Oncogene Suppressor P53 Gene to Breast Cancer MCF-7 Cells. *Int J Nanomedicine* 2011, doi:10.2147/ijnn.s17684.
104. Pantarotto, D.; Singh, R.; McCarthy, D.; Erhardt, M.; Briand, J.P.; Prato, M.; Kostarelos, K.; Bianco, A. Functionalized Carbon Nanotubes for Plasmid DNA Gene Delivery. *Angewandte Chemie - International Edition* 2004, 43, doi:10.1002/anie.200460437.
105. Qin, W.; Yang, K.; Tang, H.; Tan, L.; Xie, Q.; Ma, M.; Zhang, Y.; Yao, S. Improved GFP Gene Transfection Mediated by Polyamidoamine Dendrimer-Functionalized Multi-Walled Carbon Nanotubes with High Biocompatibility. *Colloids Surf B Biointerfaces* 2011, 84, doi:10.1016/j.colsurfb.2011.01.001.
106. Yang, X.; Zhang, Z.; Liu, Z.; Ma, Y.; Yang, R.; Chen, Y. Multi-Functionalized Single-Walled Carbon Nanotubes as Tumor Cell Targeting Biological Transporters. *Journal of Nanoparticle Research* 2008, 10, doi:10.1007/s11051-007-9316-5.
107. Dong, H.; Ding, L.; Yan, F.; Ji, H.; Ju, H. The Use of Polyethylenimine-Grafted Graphene Nanoribbon for Cellular Delivery of Locked Nucleic Acid Modified Molecular Beacon for Recognition of MicroRNA. *Biomaterials* 2011, 32, doi:10.1016/j.biomaterials.2011.02.001.
108. Crinelli, R.; Carloni, E.; Menotta, M.; Giacomini, E.; Bianchi, M.; Ambrosi, G.; Giorgi, L.; Magnani, M. Oxidized Ultrashort Nanotubes as Carbon Scaffolds for the Construction of Cell-Penetrating NF-KB Decoy Molecules. *ACS Nano* 2010, 4, doi:10.1021/nn100057c.
109. Pan, B.; Cui, D.; Xu, P.; Ozkan, C.; Feng, G.; Ozkan, M.; Huang, T.; Chu, B.; Li, Q.; He, R.; et al. Synthesis and Characterization of Polyamidoamine Dendrimer-Coated Multi-Walled Carbon Nanotubes and Their Application in Gene Delivery Systems. *Nanotechnology* 2009, 20, doi:10.1088/0957-4484/20/12/125101.
110. Van Den Bossche, J.; Al-Jamal, W.T.; Tian, B.; Nunes, A.; Fabbro, C.; Bianco, A.; Prato, M.; Kostarelos, K. Efficient Receptor-Independent Intracellular Translocation of Aptamers Mediated by Conjugation to Carbon Nanotubes. *Chemical Communications* 2010, 46, doi:10.1039/c0cc02092c.
111. Yan, Y.; Wang, R.; Hu, Y.; Sun, R.; Song, T.; Shi, X.; Yin, S. Stacking of Doxorubicin on Folic Acid-Targeted Multiwalled Carbon Nanotubes for in Vivo Chemotherapy of Tumors. *Drug Deliv* 2018, 25, 1607–1616, doi:10.1080/10717544.2018.1501120.
112. Li, J.; Du, Y.; Su, H.; Cheng, S.; Zhou, Y.; Jin, Y.; Qi, X.R. Interfacial Properties and Micellization of Triblock Poly(Ethylene Glycol)-Poly(ϵ -Caprolactone)-Polyethyleneimine Copolymers. *Acta Pharm Sin B* 2020, 10, 1122–1133, doi:10.1016/j.apsb.2020.01.006.
113. Sohrabi, N.; Alihosseini, A.; Pirouzfard, V.; Pedram, M.Z. Analysis of Dynamics Targeting CNT-Based Drug Delivery through Lung Cancer Cells: Design, Simulation, and Computational Approach. *Membranes (Basel)* 2020, 10, doi:10.3390/membranes10100283.
114. Cao, Y.; Huang, H.Y.; Chen, L.Q.; Du, H.H.; Cui, J.H.; Zhang, L.W.; Lee, B.J.; Cao, Q.R. Enhanced Lysosomal Escape of PH-Responsive Polyethylenimine-Betaine Functionalized Carbon Nanotube for the Codelivery of Survivin Small Interfering RNA and Doxorubicin. *ACS Appl Mater Interfaces* 2019, 11, doi:10.1021/acsami.8b20810.
115. Chen, W.; Yang, S.; Wei, X.; Yang, Z.; Liu, D.; Pu, X.; He, S.; Zhang, Y. Construction of Aptamer-SiRNA Chimera/PEI/5-FU/Carbon Nanotube/Collagen Membranes for the Treatment of Peritoneal Dissemination of Drug-Resistant Gastric Cancer. *Adv Healthc Mater* 2020, 9, doi:10.1002/adhm.202001153.

116. González-Domínguez, J.M.; Grasa, L.; Frontiñán-Rubio, J.; Abás, E.; Domínguez-Alfaro, A.; Mesonero, J.E.; Criado, A.; Ansón-Casaos, A. Intrinsic and Selective Activity of Functionalized Carbon Nanotube/Nanocellulose Platforms against Colon Cancer Cells. *Colloids Surf B Biointerfaces* 2022, 212, 112363, doi:10.1016/J.COLSURFB.2022.112363.
117. Su, Y.; Hu, Y.; Wang, Y.; Xu, X.; Yuan, Y.; Li, Y.; Wang, Z.; Chen, K.; Zhang, F.; Ding, X.; et al. A Precision-Guided MWNT Mediated Reawakening the Sunk Synergy in RAS for Anti-Angiogenesis Lung Cancer Therapy. *Biomaterials* 2017, 139, 75–90, doi:10.1016/J.BIOMATERIALS.2017.05.046.
118. Jwameer, M.R.; Salman, S.A.; Noori, F.T.M.; Sulaiman, G.M.; Jabir, M.S.; Khalil, K.A.A.; Ahmed, E.M.; Soliman, M.T.A. Antiproliferative Activity of PEG-PEI-SWCNTs against AMJ13 Breast Cancer Cells. *J Nanomater* 2023, 2023, doi:10.1155/2023/2855788.
119. Tang, L.; Li, J.; Pan, T.; Yin, Y.; Mei, Y.; Xiao, Q.; Wang, R.; Yan, Z.; Wang, W. Versatile Carbon Nanoplatforams for Cancer Treatment and Diagnosis: Strategies, Applications and Future Perspectives. *Theranostics* 2022, 12.
120. Harisa, G.I.; Faris, T.M. Direct Drug Targeting into Intracellular Compartments: Issues, Limitations, and Future Outlook. *Journal of Membrane Biology* 2019, 252.
121. Tang, L.; Mei, Y.; Shen, Y.; He, S.; Xiao, Q.; Yin, Y.; Xu, Y.; Shao, J.; Wang, W.; Cai, Z. Nanoparticle-Mediated Targeted Drug Delivery to Remodel Tumor Microenvironment for Cancer Therapy. *Int J Nanomedicine* 2021, 16.
122. Li, M.; Zhang, F.; Su, Y.; Zhou, J.; Wang, W. Nanoparticles Designed to Regulate Tumor Microenvironment for Cancer Therapy. *Life Sci* 2018, 201.
123. Kaymak, I.; Williams, K.S.; Cantor, J.R.; Jones, R.G. Immunometabolic Interplay in the Tumor Microenvironment. *Cancer Cell* 2021, 39.
124. Al Faraj, A.; Shaik, A.S.; Al Sayed, B.; Halwani, R.; Al Jammaz, I. Specific Targeting and Noninvasive Imaging of Breast Cancer Stem Cells Using Single-Walled Carbon Nanotubes as Novel Multimodality Nanoprobos. *Nanomedicine* 2016, 11, doi:10.2217/nnm.15.182.
125. Fu, S.; Zhao, Y.; Sun, J.; Yang, T.; Zhi, D.; Zhang, E.; Zhong, F.; Zhen, Y.; Zhang, S.; Zhang, S. Integrin $\alpha\text{v}\beta\text{3}$ -Targeted Liposomal Drug Delivery System for Enhanced Lung Cancer Therapy. *Colloids Surf B Biointerfaces* 2021, 201, 111623, doi:10.1016/j.colsurfb.2021.111623.
126. Zhao, Y.; Zhao, T.; Cao, Y.; Sun, J.; Zhou, Q.; Chen, H.; Guo, S.; Wang, Y.; Zhen, Y.; Liang, X.J.; et al. Temperature-Sensitive Lipid-Coated Carbon Nanotubes for Synergistic Photothermal Therapy and Gene Therapy. *ACS Nano* 2021, 15, doi:10.1021/acsnano.0c08790.
127. Lu, G.H.; Shang, W.T.; Deng, H.; Han, Z.Y.; Hu, M.; Liang, X.Y.; Fang, C.H.; Zhu, X.H.; Fan, Y.F.; Tian, J. Targeting Carbon Nanotubes Based on IGF-1R for Photothermal Therapy of Orthotopic Pancreatic Cancer Guided by Optical Imaging. *Biomaterials* 2019, 195, doi:10.1016/j.biomaterials.2018.12.025.
128. Wen, L.; Ding, W.; Yang, S.; Xing, D. Microwave Pumped High-Efficient Thermoacoustic Tumor Therapy with Single Wall Carbon Nanotubes. *Biomaterials* 2016, 75, doi:10.1016/j.biomaterials.2015.10.028.
129. Hassan, H.A.F.M.; Smyth, L.; Wang, J.T.W.; Costa, P.M.; Ratnasothy, K.; Diebold, S.S.; Lombardi, G.; Al-Jamal, K.T. Dual Stimulation of Antigen Presenting Cells Using Carbon Nanotube-Based Vaccine Delivery System for Cancer Immunotherapy. *Biomaterials* 2016, 104, doi:10.1016/j.biomaterials.2016.07.005.
130. Wen, Z.; Feng, Y.; Hu, Y.; Lian, L.; Huang, H.; Guo, L.; Chen, S.; Yang, Q.; Zhang, M.; Wan, L.; et al. Multiwalled Carbon Nanotubes Co-Delivering Sorafenib and Epidermal Growth Factor Receptor Signa Enhanced Tumor-Suppressing Effect on Liver Cancer. *Aging* 2021, 13, doi:10.18632/aging.103905.
131. Neves, V.; Heister, E.; Costa, S.; Tilmaciu, C.; Flahaut, E.; Soula, B.; Coley, H.M.; McFadden, J.; Silva, S.R.P. Design of Double-Walled Carbon Nanotubes for Biomedical Applications. *Nanotechnology* 2012, 23, doi:10.1088/0957-4484/23/36/365102.
132. Oh, Y.; Jin, J.O.; Oh, J. Photothermal-Triggered Control of Sub-Cellular Drug Accumulation Using Doxorubicin-Loaded Single-Walled Carbon Nanotubes for the Effective Killing of Human Breast Cancer Cells. *Nanotechnology* 2017, 28, doi:10.1088/1361-6528/aa5d7d.
133. Lee, P.C.; Chiou, Y.C.; Wong, J.M.; Peng, C.L.; Shieh, M.J. Targeting Colorectal Cancer Cells with Single-Walled Carbon Nanotubes Conjugated to Anticancer Agent SN-38 and EGFR Antibody. *Biomaterials* 2013, 34, doi:10.1016/j.biomaterials.2013.07.067.

134. Yoong, S.L.; Wong, B.S.; Zhou, Q.L.; Chin, C.F.; Li, J.; Venkatesan, T.; Ho, H.K.; Yu, V.; Ang, W.H.; Pastorin, G. Enhanced Cytotoxicity to Cancer Cells by Mitochondria-Targeting MWCNTs Containing Platinum(IV) Prodrug of Cisplatin. *Biomaterials* 2014, 35, doi:10.1016/j.biomaterials.2013.09.036.
135. Kim, S.W.; Kyung Lee, Y.; Yeon Lee, J.; Hee Hong, J.; Khang, D. PEGylated Anticancer-Carbon Nanotubes Complex Targeting Mitochondria of Lung Cancer Cells. *Nanotechnology* 2017, 28, doi:10.1088/1361-6528/aa8c31.
136. Wang, M.; Ruan, L.; Zheng, T.; Wang, D.; Zhou, M.; Lu, H.; Gao, J.; Chen, J.; Hu, Y. A Surface Convertible Nanoplatfrom with Enhanced Mitochondrial Targeting for Tumor Photothermal Therapy. *Colloids Surf B Biointerfaces* 2020, 189, doi:10.1016/j.colsurfb.2020.110854.
137. Zhou, F.; Wu, S.; Yuan, Y.; Chen, W.R.; Xing, D. Mitochondria-Targeting Photoacoustic Therapy Using Single-Walled Carbon Nanotubes. *Small* 2012, 8, doi:10.1002/smll.201101892.
138. Marangon, I.; Silva, A.A.K.; Guilbert, T.; Kolosnjaj-Tabi, J.; Marchiol, C.; Natkhunarajah, S.; Chamming's, F.; Ménard-Moyon, C.; Bianco, A.; Gennisson, J.L.; et al. Tumor Stiffening, a Key Determinant of Tumor Progression, Is Reversed by Nanomaterial-Induced Photothermal Therapy. *Theranostics* 2017, 7, doi:10.7150/thno.17574.
139. Miao, Y.; Zhang, H.; Pan, Y.; Ren, J.; Ye, M.; Xia, F.; Huang, R.; Lin, Z.; Jiang, S.; Zhang, Y.; et al. Single-Walled Carbon Nanotube: One Specific Inhibitor of Cancer Stem Cells in Osteosarcoma upon Downregulation of the TGF β 1 Signaling. *Biomaterials* 2017, 149, doi:10.1016/j.biomaterials.2017.09.032.
140. Das, M.; Nariya, P.; Joshi, A.; Vohra, A.; Devkar, R.; Seshadri, S.; Thakore, S. Carbon Nanotube Embedded Cyclodextrin Polymer Derived Injectable Nanocarrier: A Multiple Faceted Platform for Stimulation of Multi-Drug Resistance Reversal. *Carbohydr Polym* 2020, 247, doi:10.1016/j.carbpol.2020.116751.
141. Luo, X.; Wang, H.; Ji, D. Carbon Nanotubes (CNT)-Loaded Ginsenosides Rb3 Suppresses the PD-1/PD-L1 Pathway in Triple-Negative Breast Cancer. *Aging* 2021, 13, doi:10.18632/aging.203131.
142. Wang, X.; Li, B.; Jing, H.; Dong, X.; Leng, X. MWCNT-Mediated Combinatorial Photothermal Ablation and Chemo-Immunotherapy Strategy for the Treatment of Melanoma. *J Mater Chem B* 2020, 8, doi:10.1039/c9tb02238d.
143. Nichols, J.W.; Bae, Y.H. EPR: Evidence and Fallacy. *Journal of Controlled Release* 2014, 190, 451–464, doi:10.1016/J.JCONREL.2014.03.057.
144. Danhier, F. To Exploit the Tumor Microenvironment: Since the EPR Effect Fails in the Clinic, What Is the Future of Nanomedicine? *Journal of Controlled Release* 2016, 244, 108–121, doi:10.1016/J.JCONREL.2016.11.015.
145. Vasey, P.A.; Kaye, S.B.; Morrison, R.; Twelves, C.; Wilson, P.; Duncan, R.; Thomson, A.H.; Murray, L.S.; Hilditch, T.E.; Murray, T.; et al. Phase I Clinical and Pharmacokinetic Study of PK1 [N-(2-Hydroxypropyl)Methacrylamide Copolymer Doxorubicin]: First Member of a New Class of Chemotherapeutic Agents-Drug-Polymer Conjugates. Cancer Research Campaign Phase I/II Committee. *Clin Cancer Res* 1999, 5, 83–94.
146. Matsumura, Y.; Maeda, H. A New Concept for Macromolecular Therapeutics in Cancer Chemotherapy: Mechanism of Tumorotropic Accumulation of Proteins and the Antitumor Agent Smancs. *Cancer Res* 1986, 46, 6387–6392.
147. Thomas, R.G.; Surendran, S.P.; Jeong, Y.Y. Tumor Microenvironment-Stimuli Responsive Nanoparticles for Anticancer Therapy. *Front Mol Biosci* 2020, 7.
148. Yang, T.; Wu, Z.; Wang, P.; Mu, T.; Qin, H.; Zhu, Z.; Wang, J.; Sui, L. A Large-Inner-Diameter Multi-Walled Carbon Nanotube-Based Dual-Drug Delivery System with PH-Sensitive Release Properties. *J Mater Sci Mater Med* 2017, 28, doi:10.1007/s10856-017-5920-9.
149. Wang, D.; Zhang, N.; Jing, X.; Zhang, Y.; Xu, Y.; Meng, L. A Tumor-Microenvironment Fully Responsive Nano-Platform for MRI-Guided Photodynamic and Photothermal Synergistic Therapy. *J Mater Chem B* 2020, 8.
150. Qin, Y.; Chen, J.; Bi, Y.; Xu, X.; Zhou, H.; Gao, J.; Hu, Y.; Zhao, Y.; Chai, Z. Near-Infrared Light Remote-Controlled Intracellular Anti-Cancer Drug Delivery Using Thermo/PH Sensitive Nanovehicle. *Acta Biomater* 2015, 17, 201–209, doi:10.1016/J.ACTBIO.2015.01.026.

151. Doughty, A.C.V.; Hoover, A.R.; Layton, E.; Murray, C.K.; Howard, E.W.; Chen, W.R. Nanomaterial Applications in Photothermal Therapy for Cancer. *Materials* 2019, 12.
152. Moy, A.J.; Tunnell, J.W. Combinatorial Immunotherapy and Nanoparticle Mediated Hyperthermia. *Adv Drug Deliv Rev* 2017, 114, 175–183, doi:10.1016/J.ADDR.2017.06.008.
153. Sundaram, P.; Abrahamse, H. Phototherapy Combined with Carbon Nanomaterials (1d and 2d) and Their Applications in Cancer Therapy. *Materials* 2020, 13.
154. Zhu, Y.; Sun, Q.; Liu, Y.; Ma, T.; Su, L.; Liu, S.; Shi, X.; Han, D.; Liang, F. Decorating Gold Nanostars with Multiwalled Carbon Nanotubes for Photothermal Therapy. *R Soc Open Sci* 2018, 5, doi:10.1098/rsos.180159.
155. McKernan, P.; Virani, N.A.; Faria, G.N.F.; Karch, C.G.; Prada Silvy, R.; Resasco, D.E.; Thompson, L.F.; Harrison, R.G. Targeted Single-Walled Carbon Nanotubes for Photothermal Therapy Combined with Immune Checkpoint Inhibition for the Treatment of Metastatic Breast Cancer. *Nanoscale Res Lett* 2021, 16, doi:10.1186/s11671-020-03459-x.
156. Bachilo, S.M.; Balzano, L.; Herrera, J.E.; Pompeo, F.; Resasco, D.E.; Weisman, R.B. Narrow (n,m)-Distribution of Single-Walled Carbon Nanotubes Grown Using a Solid Supported Catalyst. *J Am Chem Soc* 2003, 125, doi:10.1021/ja036622c.
157. Zhou, F.; Xing, D.; Ou, Z.; Wu, B.; Resasco, D.E.; Chen, W.R. Cancer Photothermal Therapy in the Near-Infrared Region by Using Single-Walled Carbon Nanotubes. *J Biomed Opt* 2009, 14, doi:10.1117/1.3078803.
158. Vilar, R.; Fish, R.J.; Casini, A.; Neerman-Arbez, M. Fibrin(Ogen) in Human Disease: Both Friend and Foe. *Haematologica* 2020, 105.
159. Zhang, B.; Wang, H.; Shen, S.; She, X.; Shi, W.; Chen, J.; Zhang, Q.; Hu, Y.; Pang, Z.; Jiang, X. Fibrin-Targeting Peptide CREKA-Conjugated Multi-Walled Carbon Nanotubes for Self-Amplified Photothermal Therapy of Tumor. *Biomaterials* 2016, 79, 46–55, doi:10.1016/J.BIOMATERIALS.2015.11.061.
160. Naser Mohammed, S.; Mishaal Mohammed, A.; Al-Rawi, K.F. Novel Combination of Multi-Walled Carbon Nanotubes and Gold Nanocomposite for Photothermal Therapy in Human Breast Cancer Model. *Steroids* 2022, 186, 109091, doi:10.1016/J.STEROIDS.2022.109091.
161. Naief, M.F.; Khalaf, Y.H.; Mohammed, A.M. Novel Photothermal Therapy Using Multi-Walled Carbon Nanotubes and Platinum Nanocomposite for Human Prostate Cancer PC3 Cell Line. *J Organomet Chem* 2022, 975, 122422, doi:10.1016/J.JORGANCHEM.2022.122422.
162. Doix, B.; Trempelec, N.; Riant, O.; Feron, O. Low Photosensitizer Dose and Early Radiotherapy Enhance Antitumor Immune Response of Photodynamic Therapy-Based Dendritic Cell Vaccination. *Front Oncol* 2019, 9, doi:10.3389/fonc.2019.00811.
163. Ogbodu, R.O.; Limson, J.L.; Prinsloo, E.; Nyokong, T. Photophysical Properties and Photodynamic Therapy Effect of Zinc Phthalocyanine-Spermine-Single Walled Carbon Nanotube Conjugate on MCF-7 Breast Cancer Cell Line. *Synth Met* 2015, 204, 122–132, doi:10.1016/J.SYNTHMET.2015.03.011.
164. Ogbodu, R.O.; Nyokong, T. The Effect of Ascorbic Acid on the Photophysical Properties and Photodynamic Therapy Activities of Zinc Phthalocyanine-Single Walled Carbon Nanotube Conjugate on MCF-7 Cancer Cells. *Spectrochim Acta A Mol Biomol Spectrosc* 2015, 151, 174–183, doi:10.1016/J.SAA.2015.06.063.
165. Sundaram, P.; Abrahamse, H. Effective Photodynamic Therapy for Colon Cancer Cells Using Chlorin E6 Coated Hyaluronic Acid-Based Carbon Nanotubes. *Int J Mol Sci* 2020, 21, doi:10.3390/ijms21134745.
166. Son, K.H.; Hong, J.H.; Lee, J.W. Carbon Nanotubes as Cancer Therapeutic Carriers and Mediators. *Int J Nanomedicine* 2016, 11.
167. Shi, J.; Ma, R.; Wang, L.; Zhang, J.; Liu, R.; Li, L.; Liu, Y.; Hou, L.; Yu, X.; Gao, J.; et al. The Application of Hyaluronic Acid-Derivatized Carbon Nanotubes in Hematoporphyrin Monomethyl Ether-Based Photodynamic Therapy for in Vivo and in Vitro Cancer Treatment. *Int J Nanomedicine* 2013, 8, doi:10.2147/IJN.S45407.
168. Dheyab, M.A.; Aziz, A.A.; Khaniabadi, P.M.; Jameel, M.S.; Oladzadabbasabadi, N.; Rahman, A.A.; Braim, F.S.; Mehrdel, B. Gold Nanoparticles-Based Photothermal Therapy for Breast Cancer. *Photodiagnosis Photodyn Ther* 2023, 42, 103312, doi:10.1016/J.PDPDT.2023.103312.
169. Wang, D.; Ren, Y.; Shao, Y.; Yu, D.; Meng, L. Facile Preparation of Doxorubicin-Loaded and Folic Acid-Conjugated Carbon Nanotubes@Poly(N-Vinyl Pyrrole) for Targeted Synergistic Chemo-Photothermal Cancer Treatment. *Bioconjug Chem* 2017, 28, doi:10.1021/acs.bioconjugchem.7b00515.

170. Karimi, A.; Erfan, M.; Mortazavi, S.A.; Ghorbani-Bidkorbeh, F.; Kobarfard, F.; Shirazi, F.H. Functionalisation of Carbon Nanotubes by Methotrexate and Study of Synchronous Photothermal Effect of Carbon Nanotube and Anticancer Drug on Cancer Cell Death. *IET Nanobiotechnol* 2019, 13, doi:10.1049/iet-nbt.2018.5085.
171. Zhang, P.; Yi, W.; Hou, J.; Yoo, S.; Jin, W.; Yang, Q. A carbon Nanotube-Gemcitabine-Lentivirus Three-Component Composite for Chemo-Photothermal Synergistic Therapy of Cancer. *Int J Nanomedicine* 2018, 13, doi:10.2147/IJN.S165232.
172. Saliev, T.; Akhmetova, A.; Kulsharova, G. Multifunctional Hybrid Nanoparticles for Theranostics. *Core-Shell Nanostructures for Drug Delivery and Theranostics: Challenges, Strategies and Prospects for Novel Carrier Systems* 2018, 177–244, doi:10.1016/B978-0-08-102198-9.00007-7.
173. Karimi, A.; Erfan, M.; Mortazavi, S.A.; Ghorbani-Bidkorbeh, F.; Landi, B.; Kobarfard, F.; Shirazi, F.H. The Photothermal Effect of Targeted Methotrexate-Functionalized Multi-Walled Carbon Nanotubes on MCF7 Cells. *Iranian Journal of Pharmaceutical Research* 2019, 18, doi:10.22037/ijpr.2020.14484.12423.
174. Gao, W.; Zhang, W.; Yu, H.; Xing, W.; Yang, X.; Zhang, Y.; Liang, C. 3D CNT/MXene Microspheres for Combined Photothermal/Photodynamic/Chemo for Cancer Treatment. *Front Bioeng Biotechnol* 2022, 10, doi:10.3389/fbioe.2022.996177.
175. Gao, C.; Dong, P.; Lin, Z.; Guo, X.; Jiang, B.P.; Ji, S.; Liang, H.; Shen, X.C. Near-Infrared Light Responsive Imaging-Guided Photothermal and Photodynamic Synergistic Therapy Nanoplatfrom Based on Carbon Nanohorns for Efficient Cancer Treatment. *Chemistry - A European Journal* 2018, 24, doi:10.1002/chem.201802611.
176. Gao, C.; Jian, J.; Lin, Z.; Yu, Y.X.; Jiang, B.P.; Chen, H.; Shen, X.C. Hypericin-Loaded Carbon Nanohorn Hybrid for Combined Photodynamic and Photothermal Therapy in Vivo. *Langmuir* 2019, 35, doi:10.1021/acs.langmuir.9b00624.
177. Yang, J.; Hou, M.; Sun, W.; Wu, Q.; Xu, J.; Xiong, L.; Chai, Y.; Liu, Y.; Yu, M.; Wang, H.; et al. Sequential PDT and PTT Using Dual-Modal Single-Walled Carbon Nanohorns Synergistically Promote Systemic Immune Responses against Tumor Metastasis and Relapse. *Advanced Science* 2020, 7, doi:10.1002/advs.202001088.
178. Yin, Z.; Chen, D.; Zou, J.; Shao, J.; Tang, H.; Xu, H.; Si, W.; Dong, X. Tumor Microenvironment Responsive Oxygen-Self-Generating Nanoplatfrom for Dual-Imaging Guided Photodynamic and Photothermal Therapy. *ChemistrySelect* 2018, 3, doi:10.1002/slct.201800498.
179. Zhang, M.; Wang, W.; Cui, Y.; Chu, X.; Sun, B.; Zhou, N.; Shen, J. Magnetofluorescent Fe₃O₄/Carbon Quantum Dots Coated Single-Walled Carbon Nanotubes as Dual-Modal Targeted Imaging and Chemo/Photodynamic/Photothermal Triple-Modal Therapeutic Agents. *Chemical Engineering Journal* 2018, 338, doi:10.1016/j.cej.2018.01.081.
180. Marangon, I.; Ménard-Moyon, C.; Silva, A.K.A.; Bianco, A.; Luciani, N.; Gazeau, F. Synergic Mechanisms of Photothermal and Photodynamic Therapies Mediated by Photosensitizer/Carbon Nanotube Complexes. *Carbon N Y* 2016, 97, 110–123, doi:10.1016/j.CARBON.2015.08.023.
181. Behnam, M.A.; Emami, F.; Sobhani, Z.; Koohi-Hosseinabadi, O.; Dehghanian, A.R.; Zebarjad, S.M.; Moghim, M.H.; Oryan, A. Novel Combination of Silver Nanoparticles and Carbon Nanotubes for Plasmonic Photo Thermal Therapy in Melanoma Cancer Model. *Adv Pharm Bull* 2018, 8, doi:10.15171/apb.2018.006.
182. Lee, S.S.; Roche, P.J.R.; Giannopoulos, P.N.; Mitmaker, E.J.; Tamila, M.; Paliouras, M.; Trifiro, M.A. Prostate-Specific Membrane Antigen-Directed Nanoparticle Targeting for Extreme Nearfield Ablation of Prostate Cancer Cells. *Tumor Biology* 2017, 39, doi:10.1177/1010428317695943.
183. Koohi Hosseinabadi, O.; Behnam, M.A.; Khoradmehr, A.; Emami, F.; Sobhani, Z.; Dehghanian, A.R.; Dehghani Firoozabadi, A.; Rahmanifar, F.; Vafaei, H.; Tamadon, A.; et al. Benign Prostatic Hyperplasia Treatment Using Plasmonic Nanoparticles Irradiated by Laser in a Rat Model. *Biomedicine & Pharmacotherapy* 2020, 127, 110118, doi:10.1016/j.BIOPHA.2020.110118.
184. Asrar, A.; Sobhani, Z.; Behnam, M.A. Melanoma Cancer Therapy Using PEGylated Nanoparticles and Semiconductor Laser. *Adv Pharm Bull* 2022, 12, doi:10.34172/apb.2022.055.

185. Fan, J.P.; Xie, C.F.; Luo, Y.S.; Song, F.T.; Liu, X.Y.; Zhu, J.H.; Zhang, X.H. Solubilities of Betulin and Betulinic Acid in Sodium Hydroxide Aqueous Solutions of Varied Mole Fraction at Temperatures from 283.2 K to 323.2 K. *J Chem Thermodyn* 2013, *67*, 1–4, doi:10.1016/J.JCT.2013.07.014.
186. Marangon, I.; Ménard-Moyon, C.; Silva, A.K.A.; Bianco, A.; Luciani, N.; Gazeau, F. Synergic Mechanisms of Photothermal and Photodynamic Therapies Mediated by Photosensitizer/Carbon Nanotube Complexes. *Carbon N Y* 2016, *97*, doi:10.1016/j.carbon.2015.08.023.
187. Alfei, S. Cationic Materials for Gene Therapy: A Look Back to the Birth and Development of 2,2-Bis-(Hydroxymethyl)Propanoic Acid-Based Dendrimer Scaffolds. *Int J Mol Sci* 2023, *24*, 16006, doi:10.3390/ijms242116006.
188. Piña, M.J.; Girotti, A.; Serrano, S.; Muñoz, R.; Rodríguez-Cabello, J.C.; Arias, F.J. A Double Safety Lock Tumor-Specific Device for Suicide Gene Therapy in Breast Cancer. *Cancer Lett* 2020, *470*, doi:10.1016/j.canlet.2019.11.031.
189. Cáceres, B.; Ramirez, A.; Carrillo, E.; Jimenez, G.; Griñán-Lisón, C.; López-Ruiz, E.; Jiménez-Martínez, Y.; Marchal, J.A.; Boulaiz, H. Deciphering the Mechanism of Action Involved in Enhanced Suicide Gene Colon Cancer Cell Killer Effect Mediated by Gef and Apoptin. *Cancers (Basel)* 2019, *11*, doi:10.3390/cancers11020264.
190. Lin, M.C.; Wang, M.; Chou, M.C.; Chao, C.N.; Fang, C.Y.; Chen, P.L.; Chang, D.; Shen, C.H. Gene Therapy for Castration-Resistant Prostate Cancer Cells Using JC Polyomavirus-like Particles Packaged with a PSA Promoter Driven-Suicide Gene. *Cancer Gene Ther* 2019, *26*, doi:10.1038/s41417-019-0083-0.
191. Hossain, J.A.; Marchini, A.; Fehse, B.; Bjerkvig, R.; Miletic, H. Suicide Gene Therapy for the Treatment of High-Grade Glioma: Past Lessons, Present Trends, and Future Prospects. *Neurooncol Adv* 2020, *2*, doi:10.1093/noonadv/daa013.
192. Chao, C.N.; Lin, M.C.; Fang, C.Y.; Chen, P.L.; Chang, D.; Shen, C.H.; Wang, M. Gene Therapy for Human Lung Adenocarcinoma Using a Suicide Gene Driven by a Lung-Specific Promoter Delivered by JC Virus-like Particles. *PLoS One* 2016, *11*, doi:10.1371/journal.pone.0157865.
193. Michaelsen, S.R.; Christensen, C.L.; Sehested, M.; Cramer, F.; Poulsen, T.T.; Patterson, A. V.; Poulsen, H.S. Single Agent- and Combination Treatment with Two Targeted Suicide Gene Therapy Systems Is Effective in Chemoresistant Small Cell Lung Cancer Cells. *Journal of Gene Medicine* 2012, *14*, doi:10.1002/jgm.2630.
194. Shi, Q.; Sutariya, V.; Gupta, S.V.; Bhatia, D. GADD45 α -Targeted Suicide Gene Therapy Driven by Synthetic CARG Promoter E9NS Sensitizes NSCLC Cells to Cisplatin, Resveratrol, and Radiation Regardless of P53 Status. *Onco Targets Ther* 2019, *12*, doi:10.2147/OTT.S192061.
195. Mohseni-Dargah, M.; Akbari-Birgani, S.; Madadi, Z.; Saghatchi, F.; Kaboudin, B. Carbon Nanotube-Delivered IC9 Suicide Gene Therapy for Killing Breast Cancer Cells in Vitro. *Nanomedicine* 2019, *14*, doi:10.2217/nnm-2018-0342.
196. Zhang, L.; Zheng, W.; Tang, R.; Wang, N.; Zhang, W.; Jiang, X. Gene Regulation with Carbon-Based SiRNA Conjugates for Cancer Therapy. *Biomaterials* 2016, *104*, 269–278, doi:10.1016/J.BIOMATERIALS.2016.07.015.
197. Anderson, T.; Hu, R.; Yang, C.; Yoon, H.S.; Yong, K.T. Pancreatic Cancer Gene Therapy Using an SiRNA-Functionalized Single Walled Carbon Nanotubes (SWNTs) Nanoplex. *Biomater Sci* 2014, *2*, doi:10.1039/c4bm00019f.
198. Koury, J.; Lucero, M.; Cato, C.; Chang, L.; Geiger, J.; Henry, D.; Hernandez, J.; Hung, F.; Kaur, P.; Teskey, G.; et al. Immunotherapies: Exploiting the Immune System for Cancer Treatment. *J Immunol Res* 2018, *2018*, 1–16, doi:10.1155/2018/9585614.
199. Zhang, N.; Bevan, M.J. CD8⁺ T Cells: Foot Soldiers of the Immune System. *Immunity* 2011, *35*.
200. Singer, M.; Wang, C.; Cong, L.; Marjanovic, N.D.; Kowalczyk, M.S.; Zhang, H.; Nyman, J.; Sakuishi, K.; Kurtulus, S.; Gennert, D.; et al. A Distinct Gene Module for Dysfunction Uncoupled from Activation in Tumor-Infiltrating T Cells. *Cell* 2016, *166*, doi:10.1016/j.cell.2016.08.052.
201. Wang, C.; Xu, L.; Liang, C.; Xiang, J.; Peng, R.; Liu, Z. Immunological Responses Triggered by Photothermal Therapy with Carbon Nanotubes in Combination with Anti-CTLA-4 Therapy to Inhibit Cancer Metastasis. *Advanced Materials* 2014, *26*, doi:10.1002/adma.201402996.

202. Li, Y.; Xu, Z.; Qi, Z.; Huang, X.; Li, M.; Liu, S.; Yan, Y.; Gao, M. Application of Carbon Nanomaterials to Enhancing Tumor Immunotherapy: Current Advances and Prospects. *Int J Nanomedicine* 2024, Volume 19, 10899–10915, doi:10.2147/IJN.S480799.
203. Wang, F.; Ullah, A.; Fan, X.; Xu, Z.; Zong, R.; Wang, X.; Chen, G. Delivery of Nanoparticle Antigens to Antigen-Presenting Cells: From Extracellular Specific Targeting to Intracellular Responsive Presentation. *Journal of Controlled Release* 2021, 333.
204. Jang, D.I.; Lee, A.H.; Shin, H.Y.; Song, H.R.; Park, J.H.; Kang, T.B.; Lee, S.R.; Yang, S.H. The Role of Tumor Necrosis Factor Alpha (Tnf- α) in Autoimmune Disease and Current Tnf- α Inhibitors in Therapeutics. *Int J Mol Sci* 2021, 22.
205. Tau, G.; Rothman, P. Biologic Functions of the IFN- γ Receptors. *Allergy: European Journal of Allergy and Clinical Immunology* 1999, 54.
206. Paul, S.; Lal, G. The Molecular Mechanism of Natural Killer Cells Function and Its Importance in Cancer Immunotherapy. *Front Immunol* 2017, 8.
207. Patrick, B.; Akhtar, T.; Kousar, R.; Huang, C.C.; Li, X.G. Carbon Nanomaterials: Emerging Roles in Immuno-Oncology. *Int J Mol Sci* 2023, 24.
208. Meng, J.; Yang, M.; Jia, F.; Kong, H.; Zhang, W.; Wang, C.; Xing, J.; Xie, S.; Xu, H. Subcutaneous Injection of Water-Soluble Multi-Walled Carbon Nanotubes in Tumor-Bearing Mice Boosts the Host Immune Activity. *Nanotechnology* 2010, 21, doi:10.1088/0957-4484/21/14/145104.
209. Jin, H.; Gao, S.; Song, D.; Liu, Y.; Chen, X. Intratumorally CpG Immunotherapy with Carbon Nanotubes Inhibits Local Tumor Growth and Liver Metastasis by Suppressing the Epithelial-Mesenchymal Transition of Colon Cancer Cells. *Anticancer Drugs* 2021, 32, doi:10.1097/CAD.0000000000001000.
210. Dutt, T.S.; Saxena, R.K. Enhanced Antibody Response to Ovalbumin Coupled to Poly-Dispersed Acid Functionalized Single Walled Carbon Nanotubes. *Immunol Lett* 2020, 217, doi:10.1016/j.imlet.2019.11.003.
211. Li, D.; Ahmed, M.; Khan, A.; Xu, L.; Walters, A.A.; Ballesteros, B.; Al-Jamal, K.T. Tailoring the Architecture of Cationic Polymer Brush-Modified Carbon Nanotubes for Efficient siRNA Delivery in Cancer Immunotherapy. *ACS Appl Mater Interfaces* 2021, 13, doi:10.1021/acsami.1c02627.
212. Xia, Q.; Gong, C.; Gu, F.; Wang, Z.; Hu, C.; Zhang, L.; Qiang, L.; Ding, X.; Gao, S.; Gao, Y. Functionalized Multi-Walled Carbon Nanotubes for Targeting Delivery of Immunostimulatory CpG Oligonucleotides against Prostate Cancer. *J Biomed Nanotechnol* 2018, 14, doi:10.1166/jbn.2018.2605.
213. Zhao, D.; Alizadeh, D.; Zhang, L.; Liu, W.; Farrukh, O.; Manuel, E.; Diamond, D.J.; Badie, B. Carbon Nanotubes Enhance CpG Uptake and Potentiate Antiglioma Immunity. *Clinical Cancer Research* 2011, 17, doi:10.1158/1078-0432.CCR-10-2444.
214. Radzi, M.R.M.; Johari, N.A.; Zawawi, W.F.A.W.M.; Zawawi, N.A.; Latiff, N.A.; Malek, N.A.N.N.; Wahab, A.A.; Salim, M.I.; Jemon, K. In Vivo Evaluation of Oxidized Multiwalled-Carbon Nanotubes-Mediated Hyperthermia Treatment for Breast Cancer. *Biomaterials Advances* 2022, 134, 112586, doi:10.1016/J.MSEC.2021.112586.
215. Villa, C.H.; Dao, T.; Ahearn, I.; Fehrenbacher, N.; Casey, E.; Rey, D.A.; Korontsvit, T.; Zakhaleva, V.; Batt, C.A.; Philips, M.R.; et al. Single-Walled Carbon Nanotubes Deliver Peptide Antigen into Dendritic Cells and Enhance IgG Responses to Tumor-Associated Antigens. In Proceedings of the ACS Nano; 2011; Vol. 5.
216. Fadel, T.R.; Steenblock, E.R.; Stern, E.; Li, N.; Wang, X.; Haller, G.L.; Pfefferle, L.D.; Fahmy, T.M. Enhanced Cellular Activation with Single Walled Carbon Nanotube Bundles Presenting Antibody Stimuli. *Nano Lett* 2008, 8, doi:10.1021/nl080332i.
217. Fadel, T.R.; Look, M.; Staffier, P.A.; Haller, G.L.; Pfefferle, L.D.; Fahmy, T.M. Clustering of Stimuli on Single-Walled Carbon Nanotube Bundles Enhances Cellular Activation. *Langmuir* 2010, 26, doi:10.1021/la902068z.
218. Fadel, T.R.; Li, N.; Shah, S.; Look, M.; Pfefferle, L.D.; Haller, G.L.; Justesen, S.; Wilson, C.J.; Fahmy, T.M. Adsorption of Multimeric T Cell Antigens on Carbon Nanotubes: Effect on Protein Structure and Antigen-Specific T Cell Stimulation. *Small* 2013, 9, doi:10.1002/smll.201201684.
219. Burkert, S.C.; He, X.; Shurin, G. V.; Nefedova, Y.; Kagan, V.E.; Shurin, M.R.; Star, A. Nitrogen-Doped Carbon Nanotube Cups for Cancer Therapy. *ACS Appl Nano Mater* 2022, 5.

220. Zhang, Y.; Ali, S.F.; Dervishi, E.; Xu, Y.; Li, Z.; Casciano, D.; Biris, A.S. Cytotoxicity Effects of Graphene and Single-Wall Carbon Nanotubes in Neural Phaeochromocytoma-Derived Pc12 Cells. *ACS Nano* 2010, 4, doi:10.1021/nn1007176.
221. Tang, L.; Xiao, Q.; Mei, Y.; He, S.; Zhang, Z.; Wang, R.; Wang, W. Insights on Functionalized Carbon Nanotubes for Cancer Theranostics. *J Nanobiotechnology* 2021, 19.
222. Sekiyama, S.; Umezawa, M.; Iizumi, Y.; Ube, T.; Okazaki, T.; Kamimura, M.; Soga, K. Delayed Increase in Near-Infrared Fluorescence in Cultured Murine Cancer Cells Labeled with Oxygen-Doped Single-Walled Carbon Nanotubes. *Langmuir* 2019, 35, doi:10.1021/acs.langmuir.8b03789.
223. Wang, X.; Wang, C.; Cheng, L.; Lee, S.T.; Liu, Z. Noble Metal Coated Single-Walled Carbon Nanotubes for Applications in Surface Enhanced Raman Scattering Imaging and Photothermal Therapy. *J Am Chem Soc* 2012, 134, doi:10.1021/ja300140c.
224. Yan, C.; Chen, C.; Hou, L.; Zhang, H.; Che, Y.; Qi, Y.; Zhang, X.; Cheng, J.; Zhang, Z. Single-Walled Carbon Nanotube-Loaded Doxorubicin and Gd-DTPA for Targeted Drug Delivery and Magnetic Resonance Imaging. *J Drug Target* 2017, 25, doi:10.1080/1061186X.2016.1221958.
225. Zhang, M.; Wang, W.; Cui, Y.; Zhou, N.; Shen, J. Magnetofluorescent Carbon Quantum Dot Decorated Multiwalled Carbon Nanotubes for Dual-Modal Targeted Imaging in Chemo-Photothermal Synergistic Therapy. *ACS Biomater Sci Eng* 2018, 4, doi:10.1021/acsbiomaterials.7b00531.
226. Saghatchi, F.; Mohseni-Dargah, M.; Akbari-Birgani, S.; Saghatchi, S.; Kaboudin, B. Cancer Therapy and Imaging Through Functionalized Carbon Nanotubes Decorated with Magnetite and Gold Nanoparticles as a Multimodal Tool. *Appl Biochem Biotechnol* 2020, 191, doi:10.1007/s12010-020-03280-3.
227. Delogu, L.G.; Vidili, G.; Venturelli, E.; Ménard-Moyon, C.; Zoroddu, M.A.; Pilo, G.; Nicolussi, P.; Ligios, C.; Bedognetti, D.; Sgarrella, F.; et al. Functionalized Multiwalled Carbon Nanotubes as Ultrasound Contrast Agents. *Proc Natl Acad Sci U S A* 2012, 109, doi:10.1073/pnas.1208312109.
228. Avti, P.K.; Hu, S.; Favazza, C.; Mikos, A.G.; Jansen, J.A.; Shroyer, K.R.; Wang, L. V.; Sitharaman, B. Detection, Mapping, and Quantification of Single Walled Carbon Nanotubes in Histological Specimens with Photoacoustic Microscopy. *PLoS One* 2012, 7, doi:10.1371/journal.pone.0035064.
229. Wang, C.; Bao, C.; Liang, S.; Fu, H.; Wang, K.; Deng, M.; Liao, Q.; Cui, D. RGD-Conjugated Silica-Coated Gold Nanorods on the Surface of Carbon Nanotubes for Targeted Photoacoustic Imaging of Gastric Cancer. *Nanoscale Res Lett* 2014, 9, doi:10.1186/1556-276X-9-264.
230. Wang, J.T.W.; Fabbro, C.; Venturelli, E.; Ménard-Moyon, C.; Chaloin, O.; Da Ros, T.; Methven, L.; Nunes, A.; Sosabowski, J.K.; Mather, S.J.; et al. The Relationship between the Diameter of Chemically-Functionalized Multi-Walled Carbon Nanotubes and Their Organ Biodistribution Profiles in Vivo. *Biomaterials* 2014, 35, doi:10.1016/j.biomaterials.2014.07.054.
231. Zhao, H.; Chao, Y.; Liu, J.; Huang, J.; Pan, J.; Guo, W.; Wu, J.; Sheng, M.; Yang, K.; Wang, J.; et al. Polydopamine Coated Single-Walled Carbon Nanotubes as a Versatile Platform with Radionuclide Labeling for Multimodal Tumor Imaging and Therapy. *Theranostics* 2016, 6, doi:10.7150/thno.16047.
232. Ghosh, D.; Bagley, A.F.; Na, Y.J.; Birrer, M.J.; Bhatia, S.N.; Belcher, A.M. Deep, Noninvasive Imaging and Surgical Guidance of Submillimeter Tumors Using Targeted M13-Stabilized Single-Walled Carbon Nanotubes. *Proc Natl Acad Sci U S A* 2014, 111, doi:10.1073/pnas.1400821111.
233. Welsher, K.; Liu, Z.; Sherlock, S.P.; Robinson, J.T.; Chen, Z.; Daranciang, D.; Dai, H. A Route to Brightly Fluorescent Carbon Nanotubes for Near-Infrared Imaging in Mice. *Nat Nanotechnol* 2009, 4, doi:10.1038/nnano.2009.294.
234. Rawashdeh, I.; Al-Fandi, M.G.; Makableh, Y.; Harahsha, T. Developing a Nano-Biosensor for Early Detection of Pancreatic Cancer. *Sensor Review* 2021, 41, doi:10.1108/SR-01-2020-0004.
235. Sharma, A.; Hong, S.; Singh, R.; Jang, J. Single-Walled Carbon Nanotube Based Transparent Immunosensor for Detection of a Prostate Cancer Biomarker Osteopontin. *Anal Chim Acta* 2015, 869, doi:10.1016/j.aca.2015.02.010.
236. Tilmaciu, C.M.; Dinesh, B.; Pellerano, M.; Diot, S.; Guidetti, M.; Voltaire, J.; Bianco, A.; Ménard-Moyon, C.; Josserand, V.; Morris, M.C. Nanobiosensor Reports on CDK1 Kinase Activity in Tumor Xenografts in Mice. *Small* 2021, 17, doi:10.1002/smll.202007177.

237. Mahmoodi, P.; Rezayi, M.; Rasouli, E.; Avan, A.; Gholami, M.; Ghayour Mobarhan, M.; Karimi, E.; Alias, Y. Early-Stage Cervical Cancer Diagnosis Based on an Ultra-Sensitive Electrochemical DNA Nanobiosensor for HPV-18 Detection in Real Samples. *J Nanobiotechnology* 2020, 18, doi:10.1186/s12951-020-0577-9.
238. Liu, Z.; Tabakman, S.; Welsher, K.; Dai, H. Carbon Nanotubes in Biology and Medicine: In Vitro and in Vivo Detection, Imaging and Drug Delivery. *Nano Res* 2009, 2, 85–120, doi:10.1007/s12274-009-9009-8.
239. Chen, Y.C.; Young, R.J.; MacPherson, J. V.; Wilson, N.R. Silver-Decorated Carbon Nanotube Networks as SERS Substrates. *Journal of Raman Spectroscopy* 2011, 42, doi:10.1002/jrs.2862.
240. Dai, Y.; Su, J.; Wu, K.; Ma, W.; Wang, B.; Li, M.; Sun, P.; Shen, Q.; Wang, Q.; Fan, Q. Multifunctional Thermosensitive Liposomes Based on Natural Phase-Change Material: Near-Infrared Light-Triggered Drug Release and Multimodal Imaging-Guided Cancer Combination Therapy. *ACS Appl Mater Interfaces* 2019, 11, doi:10.1021/acsami.8b22748.
241. Ghaghada, K.B.; Starosolski, Z.A.; Bhayana, S.; Stupin, I.; Patel, C. V.; Bhavane, R.C.; Gao, H.; Bednov, A.; Yallampalli, C.; Belfort, M.; et al. Pre-Clinical Evaluation of a Nanoparticle-Based Blood-Pool Contrast Agent for MR Imaging of the Placenta. *Placenta* 2017, 57, 60–70, doi:10.1016/j.PLACENTA.2017.06.008.
242. Sanginario, A.; Miccoli, B.; Demarchi, D. Carbon Nanotubes as an Effective Opportunity for Cancer Diagnosis and Treatment. *Biosensors (Basel)* 2017, 7.
243. Ji, Z.; Ding, W.; Ye, F.; Lou, C.; Xing, D. Shape-Adapting Thermoacoustic Imaging System Based on Flexible Multi-Element Transducer. *Appl Phys Lett* 2015, 107, doi:10.1063/1.4929881.
244. De La Zerda, A.; Zavaleta, C.; Keren, S.; Vaithilingam, S.; Bodapati, S.; Liu, Z.; Levi, J.; Smith, B.R.; Ma, T.J.; Oralkan, O.; et al. Carbon Nanotubes as Photoacoustic Molecular Imaging Agents in Living Mice. *Nat Nanotechnol* 2008, 3, doi:10.1038/nnano.2008.231.
245. Tao, Y.; Sun, Y.; Shi, K.; Pei, P.; Ge, F.; Yang, K.; Liu, T. Versatile Labeling of Multiple Radionuclides onto a Nanoscale Metal-Organic Framework for Tumor Imaging and Radioisotope Therapy. *Biomater Sci* 2021, 9, doi:10.1039/d0bm02225j.
246. Hernandez, R.; Grudzinski, J.J.; Aluicio-Sarduy, E.; Massey, C.F.; Pinchuk, A.N.; Bitton, A.N.; Patel, R.; Zhang, R.; Rao, A. V.; Iyer, G.; et al. 177Lu-NM600 Targeted Radionuclide Therapy Extends Survival in Syngeneic Murine Models of Triple-Negative Breast Cancer. *Journal of Nuclear Medicine* 2020, 61, doi:10.2967/jnumed.119.236265.
247. Gong, H.; Peng, R.; Liu, Z. Carbon Nanotubes for Biomedical Imaging: The Recent Advances. *Adv Drug Deliv Rev* 2013, 65, 1951–1963, doi:10.1016/j.ADDR.2013.10.002.
248. Mandal, A.K.; Wu, X.; Ferreira, J.S.; Kim, M.; Powell, L.R.; Kwon, H.; Groc, L.; Wang, Y.H.; Cognet, L. Fluorescent Sp3 Defect-Tailored Carbon Nanotubes Enable NIR-II Single Particle Imaging in Live Brain Slices at Ultra-Low Excitation Doses. *Sci Rep* 2020, 10, doi:10.1038/s41598-020-62201-w.
249. Dang, X.; Bardhan, N.M.; Qi, J.; Gu, L.; Eze, N.A.; Lin, C.W.; Kataria, S.; Hammond, P.T.; Belcher, A.M. Deep-Tissue Optical Imaging of near Cellular-Sized Features. *Sci Rep* 2019, 9, doi:10.1038/s41598-019-39502-w.
250. Silindir-Gunay, M.; Sarcan, E.T.; Ozer, A.Y. Near-Infrared Imaging of Diseases: A Nanocarrier Approach. *Drug Dev Res* 2019, 80, doi:10.1002/ddr.21532.
251. Clark, L.C.; Lyons, C. ELECTRODE SYSTEMS FOR CONTINUOUS MONITORING IN CARDIOVASCULAR SURGERY. *Ann N Y Acad Sci* 1962, 102, doi:10.1111/j.1749-6632.1962.tb13623.x.
252. Gupta, S.; Murthy, C.N.; Prabha, C.R. Recent Advances in Carbon Nanotube Based Electrochemical Biosensors. *Int J Biol Macromol* 2018, 108, 687–703, doi:10.1016/j.IJBIOMAC.2017.12.038.
253. Moon, D.; Cha, Y.K.; Kim, S. ong; Cho, S.; Ko, H.J.; Park, T.H. FET-Based Nanobiosensors for the Detection of Smell and Taste. *Sci China Life Sci* 2020, 63.
254. Bellan, L.M.; Wu, D.; Langer, R.S. Current Trends in Nanobiosensor Technology. *Wiley Interdiscip Rev Nanomed Nanobiotechnol* 2011, 3.
255. Yoosefian, M.; Etminan, N. Leucine/Pd-Loaded (5,5) Single-Walled Carbon Nanotube Matrix as a Novel Nanobiosensors for in Silico Detection of Protein. *Amino Acids* 2018, 50, doi:10.1007/s00726-018-2552-4.
256. Popovics, P.; Awadallah, W.N.; Kohrt, S.E.; Case, T.C.; Miller, N.L.; Ricke, E.A.; Huang, W.; Ramirez-Solano, M.; Liu, Q.; Vezina, C.M.; et al. Prostatic Osteopontin Expression Is Associated with Symptomatic Benign Prostatic Hyperplasia. *Prostate* 2020, 80, doi:10.1002/pros.23986.

257. Wisniewski, T.; Winiecki, J.; Makarewicz, R.; Zekanowska, E. The Effect of Radiotherapy and Hormone Therapy on Osteopontin Concentrations in Prostate Cancer Patients. *J BUON* 2020, 25, 527–530.
258. Hiraoka, D.; Hosoda, E.; Chiba, K.; Kishimoto, T. SGK Phosphorylates Cdc25 and Myt1 to Trigger Cyclin B-Cdk1 Activation at the Meiotic G2/M Transition. *Journal of Cell Biology* 2019, 218, doi:10.1083/JCB.201812122.
259. Li, C.; Curreli, M.; Lin, H.; Lei, B.; Ishikawa, F.N.; Datar, R.; Cote, R.J.; Thompson, M.E.; Zhou, C. Complementary Detection of Prostate-Specific Antigen Using In 2O3 Nanowires and Carbon Nanotubes. *J Am Chem Soc* 2005, 127, doi:10.1021/ja053761g.
260. Yu, X.; Munge, B.; Patel, V.; Jensen, G.; Bhird, A.; Gong, J.D.; Kim, S.N.; Gillespie, J.; Gutkind, J.S.; Papadimitrakopoulos, F.; et al. Carbon Nanotube Amplification Strategies for Highly Sensitive Immunodetection of Cancer Biomarkers. *J Am Chem Soc* 2006, 128, doi:10.1021/ja062117e.
261. Bi, S.; Zhou, H.; Zhang, S. Multilayers Enzyme-Coated Carbon Nanotubes as Biolabel for Ultrasensitive Chemiluminescence Immunoassay of Cancer Biomarker. *Biosens Bioelectron* 2009, 24, doi:10.1016/j.bios.2009.03.002.
262. Ou, C.; Yuan, R.; Chai, Y.; Tang, M.; Chai, R.; He, X. A Novel Amperometric Immunosensor Based on Layer-by-Layer Assembly of Gold Nanoparticles-Multi-Walled Carbon Nanotubes-Thionine Multilayer Films on Polyelectrolyte Surface. *Anal Chim Acta* 2007, 603, doi:10.1016/j.aca.2007.08.052.
263. Ou, C.; Chen, S.; Yuan, R.; Chai, Y.; Zhong, X. Layer-by-Layer Self-Assembled Multilayer Films of Multi-Walled Carbon Nanotubes and Platinum-Prussian Blue Hybrid Nanoparticles for the Fabrication of Amperometric Immunosensor. *Journal of Electroanalytical Chemistry* 2008, 624, doi:10.1016/j.jelechem.2008.09.017.
264. Ding, Y.; Liu, J.; Jin, X.; Lu, H.; Shen, G.; Yu, R. Poly-L-Lysine/Hydroxyapatite/Carbon Nanotube Hybrid Nanocomposite Applied for Piezoelectric Immunoassay of Carbohydrate Antigen 19-9. *Analyst* 2008, 133, doi:10.1039/b713824e.
265. Sánchez, S.; Roldán, M.; Pérez, S.; Fàbregas, E. Toward a Fast, Easy, and Versatile Immobilization of Biomolecules into Carbon Nanotube/Polysulfone-Based Biosensors for the Detection of HCG Hormone. *Anal Chem* 2008, 80, doi:10.1021/ac7025282.
266. Li, N.; Yuan, R.; Chai, Y.; Chen, S.; An, H. Sensitive Immunoassay of Human Chorionic Gonadotrophin Based on Multi-Walled Carbon Nanotube-Chitosan Matrix. *Bioprocess Biosyst Eng* 2008, 31, doi:10.1007/s00449-008-0201-0.
267. Kim, J.P.; Lee, B.Y.; Lee, J.; Hong, S.; Sim, S.J. Enhancement of Sensitivity and Specificity by Surface Modification of Carbon Nanotubes in Diagnosis of Prostate Cancer Based on Carbon Nanotube Field Effect Transistors. *Biosens Bioelectron* 2009, 24, doi:10.1016/j.bios.2009.04.048.
268. Chen, S.; Yuan, R.; Chai, Y.; Min, L.; Li, W.; Xu, Y. Electrochemical Sensing Platform Based on Tris(2,2'-Bipyridyl)Cobalt(III) and Multiwall Carbon Nanotubes-Nafion Composite for Immunoassay of Carcinoma Antigen-125. *Electrochim Acta* 2009, 54, 7242–7247, doi:10.1016/J.ELECTACTA.2009.07.035.
269. Lin, J.; He, C.; Zhang, L.; Zhang, S. Sensitive Amperometric Immunosensor for α -Fetoprotein Based on Carbon Nanotube/Gold Nanoparticle Doped Chitosan Film. *Anal Biochem* 2009, 384, 130–135, doi:10.1016/J.AB.2008.09.033.
270. Jin, W.; Zhang, R.; Dong, C.; Jiang, T.; Tian, Y.; Yang, Q.; Yi, W.; Hou, J. A Simple MWCNTs@paper Biosensor for CA19-9 Detection and Its Long-Term Preservation by Vacuum Freeze Drying. *Int J Biol Macromol* 2020, 144, 995–1003, doi:10.1016/J.IJBIOMAC.2019.09.176.
271. Hu, D.; Yang, L.; Deng, S.; Hao, Y.; Zhang, K.; Wang, X.; Liu, Y.; Liu, H.; Chen, Y.; Xie, M. Development of Nanosensor by Bioorthogonal Reaction for Multi-Detection of the Biomarkers of Hepatocellular Carcinoma. *Sens Actuators B Chem* 2021, 334, 129653, doi:10.1016/J.SNB.2021.129653.
272. Ma, S.; Zhang, Y.; Ren, Q.; Wang, X.; Zhu, J.; Yin, F.; Li, Z.; Zhang, M. Tetrahedral DNA Nanostructure Based Biosensor for High-Performance Detection of Circulating Tumor DNA Using All-Carbon Nanotube Transistor. *Biosens Bioelectron* 2022, 197, 113785, doi:10.1016/J.BIOS.2021.113785.
273. Biswas, S.; Lan, Q.; Xie, Y.; Sun, X.; Wang, Y. Label-Free Electrochemical Immunosensor for Ultrasensitive Detection of Carbohydrate Antigen 125 Based on Antibody-Immobilized Biocompatible MOF-808/CNT. *ACS Appl Mater Interfaces* 2021, 13, doi:10.1021/acsami.0c14946.

274. Song, C.K.; Oh, E.; Kang, M.S.; Shin, B.S.; Han, S.Y.; Jung, M.; Lee, E.S.; Yoon, S.Y.; Sung, M.M.; Ng, W.B.; et al. Fluorescence-Based Immunosensor Using Three-Dimensional CNT Network Structure for Sensitive and Reproducible Detection of Oral Squamous Cell Carcinoma Biomarker. *Anal Chim Acta* 2018, 1027, 101–108, doi:10.1016/J.ACA.2018.04.025.
275. Liu, Y.; Zhu, F.; Dan, W.; Fu, Y.; Liu, S. Construction of Carbon Nanotube Based Nanoarchitectures for Selective Impedimetric Detection of Cancer Cells in Whole Blood. *Analyst* 2014, 139, doi:10.1039/c4an00758a.
276. Liao, C.; Li, Y.; Tjong, S.C. Graphene Nanomaterials: Synthesis, Biocompatibility, and Cytotoxicity. *Int J Mol Sci* 2018, 19.
277. Devasena, T.; Francis, A.P.; Ramaprabhu, S. Toxicity of Graphene: An Update. In *Reviews of Environmental Contamination and Toxicology*; 2021; Vol. 259.
278. Zhao, C.; Song, X.; Liu, Y.; Fu, Y.; Ye, L.; Wang, N.; Wang, F.; Li, L.; Mohammadniaei, M.; Zhang, M.; et al. Synthesis of Graphene Quantum Dots and Their Applications in Drug Delivery. *J Nanobiotechnology* 2020, 18.
279. Ye, L.; Kollie, L.; Liu, X.; Guo, W.; Ying, X.; Zhu, J.; Yang, S.; Yu, M. Antitumor Activity and Potential Mechanism of Novel Fullerene Derivative Nanoparticles. *Molecules* 2021, 26.
280. Malhotra, N.; Audira, G.; Castillo, A.L.; Siregar, P.; Ruallo, J.M.S.; Roldan, M.J.; Chen, J.R.; Lee, J.S.; Ger, T.R.; Hsiao, C. Der An Update Report on the Biosafety and Potential Toxicity of Fullerene-Based Nanomaterials toward Aquatic Animals. *Oxid Med Cell Longev* 2021, 2021.
281. Dong, J.; Ma, Q. Advances in Mechanisms and Signaling Pathways of Carbon Nanotube Toxicity. *Nanotoxicology* 2015, 9.
282. Chen, M.; Qin, X.; Zeng, G. Biodegradation of Carbon Nanotubes, Graphene, and Their Derivatives. *Trends Biotechnol* 2017, 35, 836–846.
283. Li, J.; Deng, X.; Wang, L.; Liu, J.; Xu, K. Clinical Application of Carbon Nanoparticles in Lymphatic Mapping during Colorectal Cancer Surgeries: A Systematic Review and Meta-Analysis. *Digestive and Liver Disease* 2020, 52, 1445–1454.
284. Xu, X.F.; Gu, J. The Application of Carbon Nanoparticles in the Lymph Node Biopsy of CN0 Papillary Thyroid Carcinoma: A Randomized Controlled Clinical Trial. *Asian J Surg* 2017, 40, 345–349, doi:10.1016/J.AJSUR.2015.11.004.
285. Li, Z.; Ao, S.; Bu, Z.; Wu, A.; Wu, X.; Shan, F.; Ji, X.; Zhang, Y.; Xing, Z.; Ji, J. Clinical Study of Harvesting Lymph Nodes with Carbon Nanoparticles in Advanced Gastric Cancer: A Prospective Randomized Trial. *World J Surg Oncol* 2016, 14, doi:10.1186/s12957-016-0835-3.
286. Qian, X.; Tucker, A.; Gidcumb, E.; Shan, J.; Yang, G.; Calderon-Colon, X.; Sultana, S.; Lu, J.; Zhou, O.; Spronk, D.; et al. High Resolution Stationary Digital Breast Tomosynthesis Using Distributed Carbon Nanotube X-Ray Source Array. *Med Phys* 2012, 39, doi:10.1118/1.3694667.
287. Amal, H.; Leja, M.; Funka, K.; Skapars, R.; Sivins, A.; Ancans, G.; Liepniece-Karele, I.; Kikuste, I.; Lasina, I.; Haick, H. Detection of Precancerous Gastric Lesions and Gastric Cancer through Exhaled Breath. *Gut* 2016, 65, doi:10.1136/gutjnl-2014-308536.
288. Xu, Z.Q.; Broza, Y.Y.; Ionsecu, R.; Tisch, U.; Ding, L.; Liu, H.; Song, Q.; Pan, Y.Y.; Xiong, F.X.; Gu, K.S.; et al. A Nanomaterial-Based Breath Test for Distinguishing Gastric Cancer from Benign Gastric Conditions. *Br J Cancer* 2013, 108, doi:10.1038/bjc.2013.44.
289. Bittar, R.; Alves, N.G.P.; Bertoldo, C.; Brugnera, C.; Oiticica, J. Efficacy of Carbon Microcoils in Relieving Cervicogenic Dizziness. *Int Arch Otorhinolaryngol* 2017, 21, doi:10.1055/s-0036-1592418.
290. Sterne, J.A.; Hernán, M.A.; Reeves, B.C.; Savović, J.; Berkman, N.D.; Viswanathan, M.; Henry, D.; Altman, D.G.; Ansari, M.T.; Boutron, I.; et al. ROBINS-I: A Tool for Assessing Risk of Bias in Non-Randomised Studies of Interventions. *BMJ (Online)* 2016, 355, doi:10.1136/bmj.i4919.
291. Higgins, J.P.T.; Altman, D.G.; Gøtzsche, P.C.; Jüni, P.; Moher, D.; Oxman, A.D.; Savović, J.; Schulz, K.F.; Weeks, L.; Sterne, J.A.C. The Cochrane Collaboration's Tool for Assessing Risk of Bias in Randomised Trials. *BMJ (Online)* 2011, 343, doi:10.1136/bmj.d5928.

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