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[Ari Rappoport](#) \*

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*Article*

# A Ceramide Theory of Multiple Sclerosis

**Ari Rappoport**

The Hebrew University of Jerusalem, Israel; ari.rappoport@mail.huji.ac.il

**Abstract:** This paper presents a new theory explaining the pathological mechanisms of multiple sclerosis (MS). MS is triggered by persistently activated immune cells, mainly B cells, as in reactivated Epstein-Barr virus infection. Activated immune cells release cytokines, the main one here being TNF $\alpha$ . A major role of TNF $\alpha$  is to support immune cell motility and tissue penetration, by promoting the breakdown of ceramide products that stabilize membranes (sphingomyelin (SPM) and galactosylceramide (GalCer)), by stimulating membrane breakdown via ceramide 1-phosphate-induced liberation of arachidonic acid, and by promoting the production of sphingosine 1-phosphate, a ceramide product that promotes immune motility. These ceramide products are expressed in endothelial cells, including the blood-brain barrier, and have a large presence in myelin. Thus, excessive cytokine release both allows immune penetration into the brain, and impairs existing myelin sheaths. SPM and GalCer are essential for myelin maintenance but not for its synthesis, explaining the dominant relapsing-remitting nature of the disease. The theory is supported by diverse evidence, and supports modern B cell-based treatment directions.

**Keywords:** multiple sclerosis; demyelination; ceramide; sphingomyelin; galactosylceramide

## 1. Introduction

Multiple sclerosis (MS) is a debilitating disease involving demyelination, impaired movement, vision, spinal cord functions (e.g., bladder control), and cognition [1]. It is the most prevalent chronic inflammatory disease of the central nervous system [1], affecting more than 2M people worldwide, with a prevalence of more than 1% in North America and Europe and 0.2% in East Asia and sub-Saharan Africa [2]. The efficacy of current MS therapies is moderate at best [1], and they do not stop disease progression.

MS is commonly perceived as an autoimmune disease mainly involving T cells [3]. However, the evidence for T cell autoimmunity, including for myelin-derived antigens, is circumstantial and weak [4,5]. There is epidemiological [6] and pathological [7] evidence supporting a major role for Epstein-Barr virus (EBV) infection, which affects B cells, in MS, and additional considerations (e.g., the efficacy of B cell depletion treatment) supporting a major role for B cells [3]. However, no convincing mechanisms have been described so far [8].

This paper presents a novel theory (T\*MS) as to how activated B cells induce the myelin damage seen in MS. The theory relies on well-known biological mechanisms, and is supported by diverse preclinical and clinical evidence.

**Theory overview.** T\*MS explains the mechanisms of MS in four simple steps. First, what causes the disease is strong persistent activation of B cells, as often happens with reactivated EBV infection. Second, activated B cells release cytokines, including tumor necrosis factor alpha (TNF $\alpha$ ), whose role is to promote immune cell migration and tissue penetration. Third, a major way through which TNF $\alpha$  attains its role is by acting on ceramide products. It induces the breakdown of sphingomyelin (SPM) and galactosylceramide (GalCer) and the liberation of arachidonic acid, thereby destabilizing membrane lipids. It also stimulates immune cell migration into blood and their penetration into tissues.

Finally, crucially, SPM and GalCer are essential for the maintenance of brain myelin sheaths, and their breakdown leads to demyelination. It does not impair myelin synthesis and non-terminal differentiation of myelin-producing cells, explaining the unique trajectory of MS lesions over time, specifically, the common relapsing-remitting course of the disease.

All aspects of this theory are supported by evidence. T\*MS explains the often dramatical improvements achieved by B cell depletion, the efficacy of fingolimod (an inhibitor of the signaling of one of the main ceramide products), and the relative lack of benefit of other treatment approaches.

## 2. Theory

### 2.1. Background

**Ceramide and its products.** Ceramides are lipids containing sphingosine and fatty acids. Ceramide participates in four main pathways [9]. First, it is used to generate sphingosine (via acid ceramidase, with the inverse reaction mediated by ceramide synthase). Sphingosine generates sphingosine 1-phosphate (S1P) via sphingosine kinases 1 and 2.

Second, ceramide and ATP form ceramide 1-phosphate (C1P) and ADP via ceramide kinase (CERK).

Third, ceramide and phosphocholine form sphingomyelin (SPM) via SPM synthase (SMS). SPM breakdown is mediated by sphingomyelin phosphodiesterase (SMase, sphingomyelinase), which has two main forms, acid (aSMase) and neutral (nASmase). SMase promotes the shedding of extracellular vesicles via SPM breakdown [10].

Finally, ceramide is the backbone of cerebrosides (mainly galactosylceramide (GalCer) and glucosylceramide (GlcCer), ceramide glycosylated with a galactose/glucose residue, respectively). GalCer is cleaved to ceramide and galactose by galactosylceramidase (GALC). The lysosomal enzymes saposinA and saposinC are coactivators of GALC to promote GalCer degradation [11], and this is assisted by cathepsinB, which mediates cleavage of prosaposin [12].

SPM and GalCer are major components in cell membranes, including myelin (see below), the blood-brain barrier (BBB), and endothelial cells in general, while S1P and C1P are major signaling molecules. S1P is abundant in the circulation and promotes immune cell migration, including exit from lymph nodes [13]. C1P promotes tissue destabilization and immune tissue penetration by stimulating cytosolic phospholipase A2, which liberates membrane arachidonic acid to destabilize membranes and produce prostaglandins (PGs) [14]. GalCer strongly promotes B cell proliferation, differentiation, and activation via the CD1d B cell receptor [15].

**Myelin.** Myelin sheaths are oligodendrocyte (OLG) processes that wrap around axons, mainly to support rapid electrical conduction [16]. In humans, about 70% of myelin is comprised of lipids, with cholesterol (>27%) and GalCer (>22%) being the main components. Sphingomyelin (>11%) and sulfatide (sulfated GalCer, 3.8%) are also important components [17]. GalCer and sulfatide are not essential for myelin formation, but are essential for its maintenance and stability [18]. OLG precursor cells (OPCs) and OLGs express GalCer and sulfatide only at terminal differentiation [19].

Remyelination is an ongoing process in adult brain [20]. New OLGs can be generated from a quiescent OPC pool to replace lost myelin.

**EBV.** EBV affects B cells. Transformed cells express high levels of TNFa, TNFb, and TGFb mRNA. Conversely, TNFa and TNFb enhance B cells after EBV [21]. EBV transformation increases galactosylation by 15-1225% in rheumatoid arthritis B cells [22].

EBV can infect astrocytes to enhance brain infiltration by peripheral blood lymphocytes [23]. It also affects epithelial cells [24], decreasing their exit from the cell cycle [25].

The expression of CD23 is greatly increased in the cell surface of EBV-transformed B cells [26]. CD23 stimulates growth and adhesion among B cells [27].

Humans are the only natural host for EBV [28]. Note that MS is a strictly human disease. It does not affect non-human primates [29].

**TNFa.** TNFa has two forms, soluble and transmembrane, which act on two receptors. It is released by macrophages, B cells, T cells, natural killer cells, dendritic cells, monocytes, microglia, astrocytes, and even neurons [30]. In particular, CD23, which marks activated B cells, induces TNFa release [31],

and there is high TNFa in EBV-positive peripheral T cell lymphomas [32,33]. Periodontitis lesions show very high levels of TNF, which are even higher in EBV-positive lesions [34].

TNFa activates all of the ceramide-based pathways involved in membrane destabilization. It activates SMase [35–38] decreasing myelin SPM [39]. It activates CERK (which induces C1P) [37], liberating arachidonic acid [40], and activates sphingosine kinase (to produce S1P) [41]. It increases the translation and activity of cathepsinB [42,43], which promotes GalCer degradation.

Myelin contains TNFa receptors (but mainly p75) [39]. TNF opposes myelin not only via ceramide products. Lipopolysaccharides kill OPCs by inducing TNFa release from microglia [44]. Reactive astrocytes inhibit OPC survival and differentiation via secreted TNFa that acts on OPC TNFR1 [45].

We need to keep in mind that TFN signaling is far from being completely understood. Although the new phrasing of its role presented in this paper (immune migration and tissue penetration) is definitely supported by the evidence, there are aspects of its signaling that are still puzzling. Specifically, TNF is known to promote both cell survival (by opposing apoptosis and necrosis) and death [46]. This may be because the survival effects act on immune cells while the negative effects are due to immune responses.

## 2.2. Multiple Sclerosis

**MS.** The above data about ceramide, myelin, TNFa, and EBV give rise to a simple theory of MS. MS occurs in situations where B cells (or other immune cells) are persistently activated, as is the case with some reactivated EBV infections. Activated immune cells induce TNFa release. TNFa promotes immune cell migration and tissue penetration, and a major tool through which this is done is by acting on ceramide products. TNFa promotes S1P, which stimulates immune migration, and promotes the degradation of SPM, GalCer, and arachidonic acid, all with the effect of degrading and destabilizing membranes. TNF's tissue penetration effects occur in the brain as well, promoting brain infiltration of immune cells. The main location of expression of SPM and GalCer in the brain is myelin, and their degradation impairs the proper maintenance of existing myelin sheaths.

**Disease types.** The most common MS type (about 85% of patients) is relapsing-remitting MS [1]. Here, relapses are followed by long (months to years) periods of relative quiet without new disease activity. Deficits that occur during attacks are fixed in about 60%. T\*MS explains this seemingly strange state as follows. The damage induced by TNFa is mainly limited to SPM and GalCer, which are needed for myelin maintenance, thereby causing lesions. Myelin synthesis is not impaired, explaining why the brain manages to execute substantial fixing processes. After causing a lesion, immune cells do not necessarily stay at the vicinity of the lesion, which can allow full recovery.

The second common disease type is primary progressive MS. Here, there is no remission after the initial symptoms. This happens when the initial disease drive (immune activation) is stronger, possibly involving several immune cell types, and with a wider spatial extent, which does not leave the brain time and space to recuperate.

Most (65%) relapsing-remitting patients switch at some point to a progressive course (termed secondary progressive). This can be explained by the presence of the problem continuously attracting immune cells into the brain and to existing lesions, even if they are small. Although immune activity can be helpful (e.g., by removing myelin debris), at some point, its persistent activation is likely to become toxic.

**Peripheral damage.** Myelin in the peripheral nervous system is damaged in MS, albeit to a lesser extent than central myelin [47]. T\*MS explains this by noting that peripheral myelin has less GalCer and much more SPM than central myelin [48].

**Pregnancy.** MS relapse rates diminish during pregnancy [49]. T\*MS explains this by noting that the first pregnancy trimester is characterized by a complete lack of TNFa. TNFa then increases to labor, with very high production at the onset of spontaneous abortions [50].

**Sex differences.** The prevalence of MS among women has increased in the last decades to be 2-3 higher than that in men [2]. Women have lower sphingolipids (including ceramide and SPM) between the ages 18-39yo. This is reversed at ages 56-70, but the MS age of onset is 31-33yo  $\pm 10$  [51].

**Vitamin D.** Vitamin D deficiency is one of the few established risk factors for MS [49]. Vitamin D decreases TNFa signaling [52].

**Altitude.** Living at higher altitudes is associated with an earlier age of onset of MS [53]. UV irradiation (which is higher at higher altitudes) is associated with aSMase activation [54], and ionizing radiation has been shown to induce rapid SPM hydrolysis to ceramide [55].

**Smoking.** Smoking is a well-established environmental risk factor in MS [49,56]. Smoking is associated with increased TNFa (but not always) [57], and activates nSMase in lung cell death [58]. In addition, smoking increases COX2 expression and PG synthesis (i.e., arachidonic acid liberation and membrane degradation) in human urinary bladder cancer [59]. On the other hand, active smoking was reported to decrease PG synthesis in human gut musoca [60]. If this is the case, then the high smoking rates in MS could be a form of self-treatment.

### 3. Evidence

**B cells and EBV in MS.** The involvement of B cells [3,61] and EBV [6,62] in MS pathogenesis has been reviewed at depth, so we will not repeat the evidence here. A recent result not included in these reviews is that expanded CSF T cells are specific to EBV-infected B cells [63].

**TNFa in MS.** There is strong evidence for TNFa involvement in MS. Increased blood TNFa preceded the exacerbation of symptoms in relapsing-remitting patients by at most two weeks [64]. The increase has normalized in many cases, with symptoms appearing only when it persisted. Blood mononuclear cells in relapsing but not stable patients showed higher TNFa and lymphotoxin mRNA [65]. Increased TNFa and IgG in peripheral mononuclear cells were reported, with increased soluble CD23 (B cell growth/activation marker) correlating with IgG [66]. TNFa production in intrathecal cells was reported, with CSF TNFa correlating with disease activity and with poor outcome [67]. MS lesions showed high TNFa, associated with astrocytes and macrophages [68,69]. Significantly high TNFa was reported in chronic active lesions [70].

TNFa was included in the 20 highest markers in a Belgium population GWAS [71].

Additional TNF data is discussed under treatment below.

**Ceramides in MS.** There is overwhelming evidence for ceramide abnormalities in MS. Increased ceramide and ceramide products (indicating degradation of ceramide products) were reported in patient CSF, serum and lymphocytes [72,73]. Ceramide accumulates in reactive astrocytes in active lesions [74]. Significantly increased ceramide metabolizing enzymes were found in plasma [75].

Serum aSMase activity was significantly higher in MS [76]. Patient CSF shows higher aSMase-enriched & total exosomes, significantly higher aSMase activity, and lower SPMs [77]. Increased aSMase activity was detected in active MS lesions, possibly driven by reactive astrocytes. Fingolimod, which acts on the S1P and C1P paths (see below), decreases this and ceramide-induced immune infiltration [78]. CSF SPM was identified as a fast, sensitive, fast, simple peripheral demyelination biomarker [79]. TNFa induced exosomal ceramide and SPM release in an OLG-related cell line [80].

Experimental autoimmune encephalomyelitis (EAE) is a popular MS model in which demyelination is induced by a stimulated immune response. EAE is blocked by inhibition of aSMase [81]. The cuprizone model of MS uses copper chelation to yield OLG death. In this model, aSMase deficiency enhances myelin repair [82]. Knockout of nSMase (but not aSMase) prevented oxidative stress-induced OLG death [83].

Plaques show decreased sulfatides and cerebroside [84]. Decreased cerebroside is an early white matter change [85]. Patients show decreased myelin sulfatides [86,87], and increased total cerebroside in serum [86]. Patient plasma extracellular vesicles contain sulfatides [88]. Sulfatide was decreased by 60% in plaque matter, and by 25% in adjacent normal-appearing white matter [89]. Normal-appearing white matter showed significantly reduced sulfatide [90]. Increased lysosomal

hydrolase activity, especially of sulfatide, was seen in plaques, which was more extensive in acute cases [91].

In chronic progressive MS patients, GalCer is undetectable in serum, indicating very strong ongoing breakdown. In relapsing-remitting patients, GalCer elevation positively correlated with relapses [92]. Similarly, plasma contained no GalCer in patients with unspecified disease type [93]. Plaques contained only 1% of GalCer, GlcCer, & sulfatide vs normal-appearing white matter. Periplaques showed intermediate amounts [94].

Patient CSF shows significantly increased cathepsinB (promoting GalCer degradation), in MS, chronic inflammatory demyelinating polyneuropathy, and Guillain-Barre syndrome (where the immune system targets muscles) [95].

CSF antibodies against sulfatide have been reported [96,97].

In normal-appearing white and grey matter in active patients, higher phospholipids and lower sphingolipids were found [98]. Normal lipid composition in normal-appearing white matter has also been reported [99].

C1P stimulates cPLA2, which liberates arachidonic acid for the synthesis of prostaglandins. MS patients show higher CSF [100] and leukocyte [101] prostaglandin E2. A sharp increase occurred in patients with active symptoms right before symptom onset [101]. COX inhibitors (which prevent prostaglandin E2 synthesis) are beneficial in MS [102].

Patient white matter and plaques showed increased sphingosine (the S1P path) [103].

In summary, ceramide definitely shows abnormalities in MS, with the evidence focusing mainly on increased degradation of SPM and GalCer, the two leading ceramide products in myelin, which are important for myelin maintenance.

**Risk factors.** The risk factors discussed above (smoking, vitamin D, altitude), and pregnancy protection, also constitute supporting evidence for T\*MS.

#### 4. Treatment

Classical MS treatments are described in the reviews cited above. Our focus in B cells, EBV, and TNFa.

**Depletion therapy.** The new focus on B cells is largely driven by the impressive success of B cell-based therapy by targeting CD20 [104].

CD20 depletion targets the mature naive and memory mature B cells, but does not affect immature and plasma cells. Although memory B cells and long-lived plasma cells are increased in MS CNS, it is possible that anti-CD20 therapy only depletes memory B cells [3]. This should decrease the effect of EBV-infected B cells.

**Ceramide-based.** Fingolimod is a structural analogue of sphingosine, phosphorylated by sphingosine kinase and acting on S1PR1, probably negatively regulating its signaling. It constitutes one of the more effective treatments in MS (relapsing-remitting, not progressive). Fingolimod blocks the exit of lymphocytes (not effector memory T cells) from lymph nodes [105], thereby reducing B and T cell migration and tissue penetration.

There are small molecule aSMase inhibitors (FIASMA) approved for treating other conditions [106]. Their use in MS needs to be examined. One such molecule has been shown to be beneficial in MS [107].

**TNF.** In apparent opposition to T\*MS, although successful in animal models, TNFa inhibitors were harmful in MS trials, and triggered demyelination when used for treating various diseases, mainly rheumatoid arthritis [30]. This is explained as follows. As noted above, TNF is capable of inducing completely opposite effects, both pro- and anti-survival [46]. The drugs used in trials and treatment are non-selective, and although their main effect is antagonism, they can also serve as agonists [30]. Selective inhibition of soluble TNF was protective in EAE, increasing myelination, while non-selective inhibition with RA drugs was not [108]. Non-selective inhibition increased memory B cells in rheumatoid factor positive RA patients [109]. These are the cells depleted by the beneficial B

cell depletion therapy. In addition, TNF knockout yields prolonged expansion of activated memory T cells, which exacerbates EAE [110].

Thus, it seems that the non-selective TNF $\alpha$  inhibitors that are approved and used for treating non-MS conditions act mainly to enhance memory B cells (which is not surprising, given that a major role of TNF $\alpha$  is to promote B cell function), which has a negative effect in MS.

**COX.** COX inhibitors, which oppose TNF-C1P action by preventing PG E2 synthesis, are beneficial in MS [102].

## 5. Discussion

This paper presented the first theory of MS that explains its pathological mechanisms. Contrary to the prevailing dominating T cell dogma, T\*MS posits that the main phenomenon in MS is driven by ceramide products. T\*MS shows how activated B cells (e.g., as a result of reactivated EBV) can cause demyelination by secreting cytokines, mainly TNF $\alpha$ , that break down myelin lipids essential for myelin maintenance. This novel account provides evidence for the causal role of B cell activation and EBV in MS etiology, and explains various perplexing properties of MS, the main one being why a relapsing-remitting disease course is so common.

**T cells, B cells.** T cells are involved in MS, and have been historically viewed as the primary participants in MS immunity and pathology, with the role of B cells being that of presenting antigens to T cells [3,111]. This can probably be attributed to the association of MS with an inflammatory T help profile, and the central role of T cells in models of demyelinating disease [61]. However, as cited above, the evidence for T cell autoimmunity in MS is actually weak [4,5], and no convincing mechanisms for T cell-induced myelin damage have been described. Recently, it has become clear that B cells play a major role, mainly due to the dramatic benefit of B cell depletion therapies [3,61]. Nonetheless, the precise role of B cells in MS, and the etiology of MS, are unknown at present [3,61]. The present paper closes this gap. People should stop describing MS as a T cell disease and start describing it as a B cell disease. T cells are certainly activated, but their role is secondary.

**Other theories.** In general, the demyelination mechanisms in MS are currently considered to be unknown [8]. The idea that myelin lipids are central in the disease has been raised [17], without the presentation of detailed mechanisms. A hypothesis in the direction of the present account has been presented in the previous century [112]. It identified TNF $\alpha$  as a major factor, but did not go further. Likewise, the possible role of immune cytokines, including TNF family members, in brain penetration by immune cells has been previously discussed [113].

**Autoimmunity.** The account here resolves one of the major puzzles of MS: the lack of evidence for antigens attacking myelin. MS is not a classical autoimmune disease in this sense, but it is an autoimmune disease in the literal sense, because the disease pathology is indeed induced by agents abnormally released by immune cells. These agents are not released as part of the final immune response, but mainly as part of the initial immune response, where immune cells destabilize membranes so that they could penetrate tissues to do their job (although memory B cells seem to be strongly involved).

**Cancer.** The account in this paper can be easily extended to provide a novel account of several types of cancer. B cell lymphomas invoke similar mechanisms to those described in this paper to disseminate themselves and penetrate tissue. Cancer has not been discussed in this paper in order to keep it focused. It will hopefully be described elsewhere.

**Strengths and weaknesses.** The main strength of the current theory is that it finally describes a convincing coherent biological account of what causes demyelination in MS. Its main weakness is that it does not fully explain the effects of non-selective TNF $\alpha$  inhibitors (although the fact that they enhance memory B cells does provide a good account).

T\*MS is not a complete theory of MS, because there are some important EBV-related questions that it does not answer (and does not purport to answer). First, it is not clear why EBV reactivation induces MS in some people, but does not do so in other people. Symptomatic EBV infection manifested as

infectious mononucleosis dramatically increases MS risk [49], but most patients do not get MS. Second, it is not clear whether reactivated EBV is a necessary condition for MS, or whether it is possible to get MS via other means (e.g., persistent non-EBV immune activation). Third, it is not clear why some people with EBV get cancers, while others get MS. Fourth, it is not clear why EBV specifically affects GalCer and SPM more than other ceramide products. The answer might be related to the fact that EBV cell entry is mediated by glycoproteins (the protein equivalent of cerebroside) [114]. It can be speculated that EBV hijacks the galactose stores in GalCer to expediate its cellular entry. Finally, it can be asked why the main damage is in the brain and not in other tissues (ignoring EBV lymphomas). This might be related to special properties of EBV, or to tissue lipid composition (the previous point).

**Theory predictions.** A bold prediction is that there should be a ceramide-based treatment that dramatically improves patient state and prevents relapse. Treatment that directly targets ceramide products might be more effective than TNF-based treatment, because although TNF $\alpha$  is the central focus in the untreated state, other immune-related cytokines can also probably drive the disease, certainly in its progressive form.

### List of Abbreviations

aSMase: acid sphingomyelin phosphodiesterase.  
 BBB: blood-brain barrier.  
 C1P: ceramide 1-phosphate.  
 COX: cyclooxygenase.  
 EBV: Epstein-Barr virus.  
 EAE: Experimental autoimmune encephalomyelitis.  
 GalCer: alpha-galactosylceramide (a cerebroside).  
 GlcCer: glucosylceramide (a cerebroside).  
 MS: multiple sclerosis.  
 nSMase: neutral sphingomyelin phosphodiesterase.  
 OLG: oligodendrocyte.  
 OPC: oligodendrocyte precursor cell.  
 RA: rheumatoid arthritis.  
 PG: prostaglandin.  
 S1P: sphingosine 1-phosphate.  
 SPM: sphingomyelin.  
 SMase: sphingomyelin phosphodiesterase.  
 sulfatide: sulfated GalCer.  
 TNF $\alpha$ : tumor necrosis factor alpha.

### References

1. Reich DS, Lucchinetti CF, Calabresi PA. Multiple Sclerosis. *The New England Journal of Medicine*. 2018;378:169–180.
2. Leray E, Moreau T, Fromont A, Edan G. Epidemiology of multiple sclerosis. *Revue neurologique*. 2016;172(1):3-13.
3. Baecher-Allan C, Kaskow BJ, Weiner HL. Multiple sclerosis: mechanisms and immunotherapy. *Neuron*. 2018;97(4):742-68.
4. Chaudhuri A, Behan PO. Multiple sclerosis is not an autoimmune disease. *Archives of neurology*. 2004;61 10:1610-2.
5. Wootla B, Eriguchi M, Rodriguez M. Is multiple sclerosis an autoimmune disease? *Autoimmune diseases*. 2012;2012(1):969657.
6. Bjornevik K, Münz C, Cohen JL, Ascherio A. Epstein-Barr virus as a leading cause of multiple sclerosis: mechanisms and implications. *Nature Reviews Neurology*. 2023;19(3):160-71.
7. Serafini B, Rosicarelli B, Franciotta D, Magliozzi R, Reynolds R, Cinque P, et al. Dysregulated Epstein-Barr virus infection in the multiple sclerosis brain. *The Journal of experimental medicine*. 2007;204(12):2899-912.

8. Lassmann H. Multiple sclerosis: lessons from molecular neuropathology. *Experimental neurology*. 2014;262:2-7.
9. Hannun YA, Obeid LM. Sphingolipids and their metabolism in physiology and disease. *Nature reviews Molecular cell biology*. 2018;19(3):175-91.
10. Van Niel G, d'Angelo G, Raposo G. Shedding light on the cell biology of extracellular vesicles. *Nature reviews Molecular cell biology*. 2018;19(4):213-28.
11. Harzer K, Paton BC, Christomanou H, Chatelut M, Levade T, Hiraiwa M, et al. Saposins (sap) A and C activate the degradation of galactosylceramide in living cells. *FEBS letters*. 1997;417(3):270-4.
12. Kim MJ, Jeong H, Krainc D. Lysosomal ceramides regulate cathepsin B-mediated processing of saposin C and glucocerebrosidase activity. *Human Molecular Genetics*. 2022.
13. Baeyens AA, Schwab SR. Finding a way out: SIP signaling and immune cell migration. *Annual review of immunology*. 2020;38(1):759-84.
14. Presa N, Gomez-Larrauri A, Rivera IG, Ordoñez M, Trueba M, Gomez-Muñoz A. Regulation of cell migration and inflammation by ceramide 1-phosphate. *Biochimica et Biophysica Acta (BBA)-molecular and cell biology of lipids*. 2016;1861(5):402-9.
15. Chen Q, Mosovsky KL, Ross AC. Retinoic acid and  $\alpha$ -galactosylceramide regulate the expression of costimulatory receptors and transcription factors responsible for B cell activation and differentiation. *Immunobiology*. 2013;218(12):1477-87.
16. Stadelmann C, Timmler S, Barrantes-Freer A, Simons M. Myelin in the central nervous system: structure, function, and pathology. *Physiological reviews*. 2019.
17. Podbielska M, Hogan E. Molecular and immunogenic features of myelin lipids: incitants or modulators of multiple sclerosis? *Multiple Sclerosis Journal*. 2009;15(9):1011-29.
18. Boggs JM. Role of galactosylceramide and sulfatide in oligodendrocytes and CNS myelin: formation of a glycosynapse. In: *Glycobiology of the Nervous System*. Springer; 2014. p. 263-91.
19. Jackman N, Ishii A, Bansal R. Oligodendrocyte development and myelin biogenesis: parsing out the roles of glycosphingolipids. *Physiology*. 2009;24(5):290-7.
20. Kremer D, Göttle P, Hartung HP, Küry P. Pushing forward: remyelination as the new frontier in CNS diseases. *Trends in neurosciences*. 2016;39(4):246-63.
21. Rochford R, Cannon MJ, Sabbe RE, Adusumilli K, Picchio G, Glynn JM, et al. Common and idiosyncratic patterns of cytokine gene expression by Epstein-Barr virus transformed human B cell lines. *Viral immunology*. 1997;10(4):183-95.
22. Wilson I, Platt F, Isenberg D, Rademacher T. Aberrant control of galactosyltransferase in peripheral B lymphocytes and Epstein-Barr virus transformed B lymphoblasts from patients with rheumatoid arthritis. *The Journal of Rheumatology*. 1993;20(8):1282-7.
23. Jakhmola S, Jha HC. Glial cell response to Epstein-Barr Virus infection: A plausible contribution to virus-associated inflammatory reactions in the brain. *Virology*. 2021;559:182-95.
24. Hutt-Fletcher LM. The long and complicated relationship between Epstein-Barr virus and epithelial cells. *Journal of virology*. 2017;91(1):10-1128.
25. Eichelberg MR, Welch R, Guidry JT, Ali A, Ohashi M, Makielski KR, et al. Epstein-Barr virus infection promotes epithelial cell growth by attenuating differentiation-dependent exit from the cell cycle. *MBio*. 2019;10(4):10-1128.
26. Kijimoto-Ochiai S, Noguchi A, Ohnishi T, Araki Y. Complex formation of CD23/surface immunoglobulin and CD23/CD81/MHC class II on an EBV-transformed human B cell line and inferable role of tetraspanin. *Microbiology and immunology*. 2004;48(5):417-26.
27. Gordon J. CD23 and B cell activation. *Clinical & Experimental Allergy*. 1992;22(2).
28. Fujiwara S, Nakamura H. Animal models for gammaherpesvirus infections: recent development in the analysis of virus-induced pathogenesis. *Pathogens*. 2020;9(2):116.
29. Bove RM. Why monkeys do not get multiple sclerosis (spontaneously) An evolutionary approach. *Evolution, Medicine, and Public Health*. 2018;2018(1):43-59.
30. Kemanetoglou E, Andreadou E. CNS demyelination with TNF- $\alpha$  blockers. *Current neurology and neuroscience reports*. 2017;17:1-15.

31. Lecoanet-Henchoz S, Gauchat JF, Aubry JP, Graber P, Life P, Paul-Eugene N, et al. CD23 regulates monocyte activation through a novel interaction with the adhesion molecules CD11b-CD18 and CD11c-CD18. *Immunity*. 1995;3(1):119-25.
32. and others. Upregulation of tumor necrosis factor- $\alpha$  gene by Epstein-Barr virus and activation of macrophages in Epstein-Barr virus-infected T cells in the pathogenesis of hemophagocytic syndrome. *The Journal of clinical investigation*. 1997;100(8):1969-79.
33. Ho J, Liang R, Srivastava G. Differential cytokine expression in EBV positive peripheral T cell lymphomas. *Molecular Pathology*. 1999;52(5):269.
34. Hernádi K, Gyöngyösi E, Mészáros B, Szakács L, Szalmás A, Csoma E, et al. Elevated tumor necrosis factor- $\alpha$  expression in periapical lesions infected by Epstein-Barr virus. *Journal of Endodontics*. 2013;39(4):456-60.
35. Inhibition of TSH-induced hydrogen peroxide production by TNF- $\alpha$  through a sphingomyelinase signaling pathway. *American Journal of Physiology-Endocrinology and Metabolism*. 1997;273(3):E638.
36. Clarke CJ, Truong TG, Hannun YA. Role for neutral sphingomyelinase-2 in tumor necrosis factor  $\alpha$ -stimulated expression of vascular cell adhesion molecule-1 (VCAM) and intercellular adhesion molecule-1 (ICAM) in lung epithelial cells: p38 MAPK is an upstream regulator of nSMase2. *Journal of Biological Chemistry*. 2007;282(2):1384-96.
37. Barth BM, Gustafson SJ, Hankins JL, Kaiser JM, Haakenson JK, Kester M, et al. Ceramide kinase regulates TNF $\alpha$ -stimulated NADPH oxidase activity and eicosanoid biosynthesis in neuroblastoma cells. *Cellular signalling*. 2012;24(6):1126-33.
38. Shamseddine AA, Airola MV, Hannun YA. Roles and regulation of neutral sphingomyelinase-2 in cellular and pathological processes. *Advances in biological regulation*. 2015;57:24-41.
39. Chakraborty G, Ziemba S, Drivas A, Ledeen R. Myelin contains neutral sphingomyelinase activity that is stimulated by tumor necrosis factor- $\alpha$ . *Journal of neuroscience research*. 1997;50(3):466-76.
40. Yang X, Sheng W, Ridgley D, Haidekker M, Sun G, Lee J. Astrocytes regulate  $\alpha$ -secretase-cleaved soluble amyloid precursor protein secretion in neuronal cells: Involvement of group IIA secretory phospholipase A2. *Neuroscience*. 2015;300:508-17.
41. Barr RK, Lynn HE, Moretti PA, Khew-Goodall Y, Pitson SM. Deactivation of sphingosine kinase 1 by protein phosphatase 2A. *Journal of Biological Chemistry*. 2008;283(50):34994-5002.
42. Guicciardi ME, Deussing J, Miyoshi H, Bronk SF, Svingen PA, Peters C, et al. Cathepsin B contributes to TNF- $\alpha$ -mediated hepatocyte apoptosis by promoting mitochondrial release of cytochrome c. *The Journal of clinical investigation*. 2000;106(9):1127-37.
43. Foghsgaard L, Wissing D, Mauch D, Lademann U, Bastholm L, Boes M, et al. Cathepsin B acts as a dominant execution protease in tumor cell apoptosis induced by tumor necrosis factor. *The Journal of cell biology*. 2001;153(5):999-1010.
44. Kim S, Steelman AJ, Koito H, Li J. Astrocytes promote TNF-mediated toxicity to oligodendrocyte precursors. *Journal of neurochemistry*. 2011;116(1):53-66.
45. Su Z, Yuan Y, Chen J, Zhu Y, Qiu Y, Zhu F, et al. Reactive astrocytes inhibit the survival and differentiation of oligodendrocyte precursor cells by secreted TNF- $\alpha$ . *Journal of neurotrauma*. 2011;28(6):1089-100.
46. Probert L. TNF and its receptors in the CNS: The essential, the desirable and the deleterious effects. *Neuroscience*. 2015;302:2-22.
47. Oudejans E, Luchicchi A, Strijbis EM, Geurts JJ, van Dam AM. Is MS affecting the CNS only? Lessons from clinic to myelin pathophysiology. *Neurology: Neuroimmunology & Neuroinflammation*. 2021;8(1):e914.
48. Morell P, editor. *Myelin*. 2nd ed. Springer Science & Business Media; 1984.
49. McKay KA, Jahanfar S, Duggan T, Tkachuk S, Tremlett H. Factors associated with onset, relapses or progression in multiple sclerosis: a systematic review. *Neurotoxicology*. 2017;61:189-212.
50. Daher S, Fonseca F, Ribeiro OG, Musatti CC, Gerbase-DeLima M. Tumor necrosis factor during pregnancy and at the onset of labor and spontaneous abortion. *European Journal of Obstetrics & Gynecology and Reproductive Biology*. 1999;83(1):77-9.
51. Muilwijk M, Callender N, Goorden S, Vaz FM, van Valkengoed IG. Sex differences in the association of sphingolipids with age in Dutch and South-Asian Surinamese living in Amsterdam, the Netherlands. *Biology of sex differences*. 2021;12:1-14.

52. Zwerina K, Baum W, Axmann R, Heiland GR, Distler JHW, Smolen JS, et al. Vitamin D receptor regulates TNF-mediated arthritis. *Annals of the Rheumatic Diseases*. 2011;70:1122-1129.
53. Tao C, Simpson S, Van Der Mei I, Blizzard L, Havrdova E, Horakova D, et al. Higher latitude is significantly associated with an earlier age of disease onset in multiple sclerosis. *Journal of Neurology, Neurosurgery & Psychiatry*. 2016;87(12):1343-9.
54. Komatsu M, Takahashi T, Abe T, Takahashi I, Ida H, Takada G. Evidence for the association of ultraviolet-C and H<sub>2</sub>O<sub>2</sub>-induced apoptosis with acid sphingomyelinase activation. *Biochimica et Biophysica Acta (BBA)-Molecular and Cell Biology of Lipids*. 2001;1533(1):47-54.
55. Ionizing radiation acts on cellular membranes to generate ceramide and initiate apoptosis. *The Journal of experimental medicine*. 1994;180(2):525-35.
56. Belbasis L, Bellou V, Evangelou E, Ioannidis JP, Tzoulaki I. Environmental risk factors and multiple sclerosis: an umbrella review of systematic reviews and meta-analyses. *The Lancet Neurology*. 2015;14(3):263-73.
57. Yanbaeva DG, Dentener MA, Creutzberg EC, Wesseling G, Wouters EF. Systemic effects of smoking. *Chest*. 2007;131(5):1557-66.
58. Levy M, Khan E, Careaga M, Goldkorn T. Neutral sphingomyelinase 2 is activated by cigarette smoke to augment ceramide-induced apoptosis in lung cell death. *American Journal of Physiology-Lung Cellular and Molecular Physiology*. 2009;297(1):L125-33.
59. Badawi AF, Habib SL, Mohammed MA, Abadi AA, Michael MS. Influence of cigarette smoking on prostaglandin synthesis and cyclooxygenase-2 gene expression in human urinary bladder cancer. *Cancer investigation*. 2002;20(5-6):651-6.
60. QUIMBY GF, BONNICE CA, BURSTEIN SH, EASTWOOD GL. Active smoking depresses prostaglandin synthesis in human gastric mucosa. *Annals of internal medicine*. 1986;104(5):616-9.
61. Cencioni MT, Mattosio M, Magliozzi R, Bar-Or A, Muraro PA. B cells in multiple sclerosis – from targeted depletion to immune reconstitution therapies. *Nature Reviews Neurology*. 2021;17:399-414.
62. Pender MP. The essential role of Epstein-Barr virus in the pathogenesis of multiple sclerosis. *The Neuroscientist*. 2011;17(4):351-67.
63. Gottlieb A, Pham HPT, Salterelli JG, Lindsey JW. Expanded T lymphocytes in the cerebrospinal fluid of multiple sclerosis patients are specific for Epstein-Barr-virus-infected B cells. *Proceedings of the National Academy of Sciences*. 2024;121(3):e2315857121.
64. Beck J, Rondot P, Catinot L, Falcoff E, Kirchner H, Wietzerbin J. Increased production of interferon gamma and tumor necrosis factor precedes clinical manifestation in multiple sclerosis: do cytokines trigger off exacerbations? *Acta Neurologica Scandinavica*. 1988;78(4):318-23.
65. Rieckmann P, Albrecht M, Kitze B, Weber T, Tumani H, Broocks A, et al. Cytokine mRNA levels in mononuclear blood cells from patients with multiple sclerosis. *Neurology*. 1994;44(8):1523-3.
66. Zaffaroni M, Stampino LG, Ghezzi A, Baldini SM, Zibetti A. In vitro cytokine, sCD23 and IgG secretion in multiple sclerosis. *Journal of neuroimmunology*. 1995;61(1):1-5.
67. Sharief MK, Hentges R. Association between tumor necrosis factor- $\alpha$  and disease progression in patients with multiple sclerosis. *New England Journal of Medicine*. 1991;325(7):467-72.
68. Hofman F, Hinton D, Johnson K, Merrill J. Tumor necrosis factor identified in multiple sclerosis brain. *The Journal of experimental medicine*. 1989;170(2):607-12.
69. Selmaj, K, Raine, CS, Cannella, B, Brosnan, CF. Identification of lymphotoxin and tumor necrosis factor in multiple sclerosis lesions. *The Journal of clinical investigation*. 1991;87(3):949-54.
70. Cannella B, Raine CS. The adhesion molecule and cytokine profile of multiple sclerosis lesions. *Annals of Neurology: Official Journal of the American Neurological Association and the Child Neurology Society*. 1995;37(4):424-35.
71. Goris A, Sawcer S, Vandenbroeck K, Carton H, Billiau A, Setakis E, et al. New candidate loci for multiple sclerosis susceptibility revealed by a whole genome association screen in a Belgian population. *Journal of neuroimmunology*. 2003;143(1-2):65-9.
72. Sela BA, Konat G, Offner H. Elevated ganglioside concentration in serum and peripheral blood lymphocytes from multiple sclerosis patients in remission. *Journal of the neurological sciences*. 1982;54(1):143-8.
73. Kułakowska A, Żendzian-Piotrowska M, Baranowski M, Konończuk T, Drozdowski W, Górski J, et al. Intrathecal increase of sphingosine 1-phosphate at early stage multiple sclerosis. *Neuroscience letters*. 2010;477(3):149-52.

74. Kim S, Steelman AJ, Zhang Y, Kinney HC, Li J. Aberrant upregulation of astroglial ceramide potentiates oligodendrocyte injury. *Brain Pathology*. 2012;22(1):41-57.
75. Kurz J, Brunkhorst R, Foerch C, Blum L, Henke M, Gabriel L, et al. The relevance of ceramides and their synthesizing enzymes for multiple sclerosis. *Clinical Science*. 2018;132(17):1963-76.
76. Leurs C, Pinheiro ML, Wiertz L, den Hoedt S, Mulder M, Eijlers A, et al. Acid sphingomyelinase: No potential as a biomarker for multiple sclerosis. *Multiple sclerosis and related disorders*. 2019;28:44-9.
77. Pieragostino D, Cicalini I, Lanuti P, Ercolino E, di Ioia M, Zucchelli M, et al. Enhanced release of acid sphingomyelinase-enriched exosomes generates a lipidomics signature in CSF of Multiple Sclerosis patients. *Scientific reports*. 2018;8(1):1-12.
78. van Doorn R, Nijland PG, Dekker N, Witte ME, Lopes-Pinheiro MA, van het Hof B, et al. Fingolimod attenuates ceramide-induced blood-brain barrier dysfunction in multiple sclerosis by targeting reactive astrocytes. *Acta neuropathologica*. 2012;124(3):397-410.
79. Capodivento G, Visigalli D, Garnerio M, Fancellu R, Ferrara MD, Basit A, et al. Sphingomyelin as a myelin biomarker in CSF of acquired demyelinating neuropathies. *Scientific reports*. 2017;7(1):1-9.
80. Podbielska M, Szulc ZM, Kurowska E, Hogan EL, Bielawski J, Bielawska A, et al. Cytokine-induced release of ceramide-enriched exosomes as a mediator of cell death signaling in an oligodendroglioma cell line. *Journal of lipid research*. 2016;57(11):2028-39.
81. Becker KA, Halmer R, Davies L, Henry BD, Ziobro-Henry R, Decker Y, et al. Blockade of experimental multiple sclerosis by inhibition of the acid sphingomyelinase/ceramide system. *Neurosignals*. 2017;25(1):88-97.
82. Chami M, Halmer R, Schnoeder L, Anne Becker K, Meier C, Fassbender K, et al. Acid sphingomyelinase deficiency enhances myelin repair after acute and chronic demyelination. *PloS one*. 2017;12(6):e0178622.
83. Jana A, Pahan K. Oxidative stress kills human primary oligodendrocytes via neutral sphingomyelinase: implications for multiple sclerosis. *Journal of Neuroimmune Pharmacology*. 2007;2(2):184-93.
84. Wilson R, Tocher DR. Lipid and fatty acid composition is altered in plaque tissue from multiple sclerosis brain compared with normal brain white matter. *Lipids*. 1991;26(1):9-15.
85. Gerstl B, Eng L, Tavaststjerna M, Smith J, Kruse S. LIPIDS AND PROTEINS IN MULTIPLE SCLEROSIS WHITE MATTER. *Journal of Neurochemistry*. 1970;17(5):677-89.
86. Clausen J, Hansen IB. Myelin constituents of human central nervous system: studies of phospholipid, glycolipid, and fatty acid pattern in normal and multiple sclerosis brains. *Acta Neurologica Scandinavica*. 1970;46(1):1-17.
87. Woelk H, Borri P. Lipid and fatty acid composition of myelin purified from normal and MS brains. *European neurology*. 1973;10(4):250-60.
88. Moyano AL, Li G, Boullerne AI, Feinstein DL, Hartman E, Skias D, et al. Sulfatides in extracellular vesicles isolated from plasma of multiple sclerosis patients. *Journal of neuroscience research*. 2016;94(12):1579-87.
89. Marbois BN, Faull KF, Fluharty AL, Raval-Fernandes S, Rome LH. Analysis of sulfatide from rat cerebellum and multiple sclerosis white matter by negative ion electrospray mass spectrometry. *Biochimica et biophysica acta*. 2000;1484 1:59-70.
90. Alling C, Svennerholm L, et al. Lipid alterations in apparently normal white matter in multiple sclerosis. *Brain research*. 1971;35(2):325-36.
91. Cuzner ML, Davison A. Changes in cerebral lysosomal enzyme activity and lipids in multiple sclerosis. *Journal of the neurological sciences*. 1973;19(1):29-36.
92. Lubetzki C, PharmD YT, PharmD AG, Lyon-Caen O, Lhermitte F, Zalc B. Galactosylceramide: a reliable serum index of demyelination in multiple sclerosis. *Annals of Neurology: Official Journal of the American Neurological Association and the Child Neurology Society*. 1989;26(3):407-9.
93. Baumann N, Lemonnier M, Jacque C, Marteau R, Harpin M, Lhermitte F. Plasma galactocerebrosides in multiple sclerosis. *Biomedicine/[publiee Pour l'AAICIG]*. 1975;23(9):387-90.
94. Yahara S, Kawamura N, Kishimoto Y, Saida T, Tourtellotte WW. A change in the cerebrosides and sulfatides in a demyelinating nervous system: development of the methodology and study of multiple sclerosis and Wallerian degeneration. *Journal of the neurological sciences*. 1982;54(2):303-15.
95. Nagai A, Murakawa Y, Terashima M, Shimode K, Umegae N, Takeuchi H, et al. Cystatin C and cathepsin B in CSF from patients with inflammatory neurologic diseases. *Neurology*. 2000;55(12):1828-32.

96. Kanter JL, Narayana S, Ho PP, Catz I, Warren KG, Sobel RA, et al. Lipid microarrays identify key mediators of autoimmune brain inflammation. *Nature medicine*. 2006;12(1):138-43.
97. Ilyas AA, Chen ZW, Cook SD. Antibodies to sulfatide in cerebrospinal fluid of patients with multiple sclerosis. *Journal of neuroimmunology*. 2003;139(1-2):76-80.
98. Wheeler D, Bandaru VVR, Calabresi PA, Nath A, Haughey NJ. A defect of sphingolipid metabolism modifies the properties of normal appearing white matter in multiple sclerosis. *Brain*. 2008;131(11):3092-102.
99. Suzuki K, Kamoshita S, Eto Y, Tourtellotte WW, Gonatas JO. Myelin in multiple sclerosis: composition of myelin from normal-appearing white matter. *Archives of Neurology*. 1973;28(5):293-7.
100. Mattsson N, Yaong M, Rosengren LE, Blennow K, Månsson JE, Andersen O, et al. Elevated cerebrospinal fluid levels of prostaglandin E2 and 15-(S)-hydroxyeicosatetraenoic acid in multiple sclerosis. *Journal of Internal Medicine*. 2009;265.
101. Dore-Duffy P, Donaldson JO, Koff T, Longo M, Perry WB. Prostaglandin release in multiple sclerosis: correlation with disease activity. *Neurology*. 1986;36:1587-1587.
102. Hoxha M, Spahiu E, Prendi E, Zappacosta B. A Systematic Review on the Role of Arachidonic Acid Pathway in Multiple Sclerosis. *CNS & neurological disorders drug targets*. 2020.
103. Miller LG, Young JA, Ray SK, Wang G, Purohit S, Banik NL, et al. Sphingosine toxicity in EAE and MS: evidence for ceramide generation via serine-palmitoyltransferase activation. *Neurochemical research*. 2017;42:2755-68.
104. Margoni M, Preziosa P, Filippi M, Rocca MA. Anti-CD20 therapies for multiple sclerosis: current status and future perspectives. *Journal of Neurology*. 2022;269(3):1316-34.
105. Pelletier D, Hafler DA. Fingolimod for multiple sclerosis. *New England Journal of Medicine*. 2012;366(4):339-47.
106. Skácel J, Slusher BS, Tsukamoto T. Small molecule inhibitors targeting biosynthesis of ceramide, the central hub of the sphingolipid network. *Journal of medicinal chemistry*. 2021;64(1):279-97.
107. Mostert J, Admiraal-Behloul F, Hoogduin J, Luyendijk J, Heersema D, Van Buchem M, et al. Effects of fluoxetine on disease activity in relapsing multiple sclerosis: a double-blind, placebo-controlled, exploratory study. *Journal of Neurology, Neurosurgery & Psychiatry*. 2008;79(9):1027-31.
108. Brambilla R, Ashbaugh JJ, Magliozzi R, Dellarole A, Karmally S, Szymkowski DE, et al. Inhibition of soluble tumour necrosis factor is therapeutic in experimental autoimmune encephalomyelitis and promotes axon preservation and remyelination. *Brain*. 2011;134(9):2736-54.
109. Roll P, Muhammad K, Schumann M, Kleinert S, Tony H. RF positivity has substantial influence on the peripheral memory B-cell compartment and its modulation by TNF inhibition. *Scandinavian journal of rheumatology*. 2012;41(3):180-5.
110. Kassiotis G, Kollias G. Uncoupling the proinflammatory from the immunosuppressive properties of tumor necrosis factor (TNF) at the p55 TNF receptor level: implications for pathogenesis and therapy of autoimmune demyelination. *The Journal of experimental medicine*. 2001;193(4):427-34.
111. Kuchroo VK, Weiner HL. How does Epstein-Barr virus trigger MS? *Immunity*. 2022;55(3):390-2.
112. Ledeen RW, Chakraborty G. Cytokines, signal transduction, and inflammatory demyelination: review and hypothesis. *Neurochemical research*. 1998;23:277-89.
113. Larochelle C, Alvarez JI, Prat A. How do immune cells overcome the blood-brain barrier in multiple sclerosis? *FEBS letters*. 2011;585(23):3770-80.
114. Shannon-Lowe C, Rowe M. Epstein Barr virus entry; kissing and conjugation. *Current opinion in virology*. 2014;4:78-84.

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