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Article

Zinc Bismuthate-Based Nanocomposites: Promising Antimicro-Bial Properties for Biomedical Applications

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Abstract

Background/Objectives The growing threat of antimicrobial resistance (AMR) has intensified the search for innovative antimicrobial materials. Herein, we synthesized and characterized zinc bismuthate-based nanocomposites (ZB1 and ZB4) and investigated their antimicrobial efficacy against two Gram-positive bacteria (Staphylococcus aureus and Corynebacterium diphtheriae), one Gramnegative bacterium (Klebsiella pneumoniae), and two fungal pathogens (Cryptococcus neoformans and Candida albicans). Toxicity was further assessed using Tenebrio molitor larvae as an alternative model. Methods: Nanocomposites were prepared via a hydrothermal process and characterized using X-ray diffraction (XRD), Raman spectroscopy, Zeta potential, and high-resolution transmission electron microscopy (HR-TEM). Antimicrobial activity was evaluated through disk diffusion assays coupled with minimum inhibitory concentration (MIC) and minimum fungicidal concentration (MFC) assessments. Results: ZB1 and ZB4 inhibited S. aureus $(10.7 \pm 1.2 \text{ mm})$ and $11.3 \pm 1.2 \text{ mm}$), C. diphtheriae $(10.7 \pm 1.1 \text{ mm})$ and $11.3 \pm 1.1 \text{ mm}$, and *C. neoformans* $(10.7 \pm 1.1 \text{ mm})$ and $11.7 \pm 2.9 \text{ mm}$, while no inhibition was observed for C. albicans and K. pneumoniae. From MIC and MFC evaluations, ZB1 was effective up to 1000 µg/mL, and ZB4 up to 500 µg/mL. Toxicity assays revealed survival rates of 70% for ZB1 and 60% for ZB4, with no statistically significant differences compared to the control (p = 0.156). Notably, ZB4 is a promising candidate due to its enhanced antimicrobial performance and moderate toxicity. Conclusions: Our findings highlight ZB1 and ZB4 nanocomposites as promising antimicrobial agents, offering new avenues in nanotechnology-driven healthcare solutions aimed at tackling antimicrobial resistance.

Keywords: zinc bismuthate nanocomposites; antimicrobial resistance; antimicrobial properties; zinc oxide nanoparticles; antimicrobial nanomaterials; nanotechnology; biocompatibility

1. Introduction

In recent decades, antimicrobial resistance (AMR) has become a major global health threat, contributing to high rates of morbidity and mortality, and highlighting the urgent need for innovative and effective containment strategies [1]. Despite substantial efforts, overcoming antimicrobial resistance (AMR) remains challenging, as the discovery and development of new antimicrobial agents are time-intensive, costly, and frequently hindered by issues of high toxicity and limited clinical efficacy [2].

The World Health Organization (WHO) has highlighted the growing prevalence of multidrugresistant microorganisms in water bodies, accelerating the spread of resistance [3]. In Brazil alone, an estimated 34,000 annual deaths are directly linked to AMR, with an additional 138,000 associated with related complications. Poor waste management practices, particularly the improper discharge of domestic and hospital effluents, introduce pathogenic bacteria and antibiotics into aquatic environments, fostering an ecosystem for the selection and horizontal transfer of resistance genes [4– 6].

In this context, the development of safe and efficient water purification and microbial control technologies has become increasingly critical. Nanotechnology, especially the application of metallic and metal oxide nanoparticles (NPs), offers a promising strategy to combat AMR. These nanomaterials, characterized by their high surface area and nanoscale effects, operate via multiple mechanisms: (i) generation of reactive oxygen species (ROS) or metal ion release, leading to microbial DNA and protein damage; (ii) disruption of microbial cell membranes; and (iii) interference with electron transport across membranes, compromising vital cellular functions [7].

Nevertheless, the clinical translation of conventional metallic NPs faces significant challenges. Gold nanoparticles are expensive, silver nanoparticles are unstable under light, and copper nanoparticles may exert toxicity by interfering with essential physiological processes [8]. In contrast, zinc oxide (ZnO) and bismuth (Bi) nanoparticles have emerged as safer, more stable, and effective alternatives, demonstrating potent antimicrobial properties against resistant strains and biofilms, while maintaining low toxicity and high chemical stability [9–11].

Zinc bismuthate-based materials have attracted increasing attention for their notable bactericidal activity [12,13] and promising applications in the photocatalytic degradation of organic pollutants in aquatic environments [14,15]. Their nanostructures also offer high surface areas and excellent electrochemical performance [16]. However, their antimicrobial potential remains largely unexplored.

Zinc oxide nanoparticles have been widely recognized for their antibacterial and antifungal activities against diverse Gram-positive and Gram-negative pathogens. Their bactericidal effect is primarily attributed to the generation of oxidative stress via ROS [17–19]. Recent evidence suggests that ROS generation can occur without light. The process is facilitated by structural defects such as oxygen vacancies (Vo) which distort the ZnO crystalline lattice and enhance electron transfer to adsorbed molecules, thus sustaining ROS production [20].

While zinc bismuthate/zinc oxide nanocomposites have been investigated for their photocatalytic efficiency [14,21], no studies to date have evaluated their antimicrobial efficacy. In this study, we report the hydrothermal synthesis of these nanocomposites and assess, for the first time, their antimicrobial performance and toxicity.

The hydrothermal method offers significant advantages for nanoparticle synthesis, including excellent reproducibility, cost-effectiveness, simple operation, and environmental friendliness [22–24]. It enables precise control over particle morphology and size, employing a high-pressure autoclave system typically composed of stainless steel with Teflon lining [25,26].

Thus, this work aims to synthesize, characterize, and evaluate the antibacterial and antifungal activities of zinc bismuthate/zinc oxide nanocomposites (ZB1 and ZB4) against *Staphylococcus aureus*, *Corynebacterium diphtheriae*, *Klebsiella pneumoniae*, *Cryptococcus neoformans*, and *Candida albicans*, while also investigating their toxicity profiles to establish their potential as safe and innovative antimicrobial agents.

2. Materials and Methods

2.1. Materials

Zinc Nitrate Hexahydrate (98%, Sigma-Aldrich), Bismuth Nitrate Pentahydrate (98%, Synth), Potassium Hydroxide (analytical purity, Dinâmica Química), Glacial Acetic Acid (99%, Neon). Additionally, other laboratory reagents such as Dimethyl Sulfoxide (DMSO, Sigma-Aldrich), Phosphate Buffered Saline (PBS) and Brain Heart Infusion (BHI) broth were used for the microbiological analyses. All reagents used in this work were obtained from commercial suppliers, and were used without additional purification, ensuring the reproducibility of the synthesis.

2.2. Hydrothermal Synthesis of Nanocomposites

The zinc bismuthate-based nanocomposite was synthesized via a hydrothermal method, according to the methodology adapted from Dilly Rajan et al. [27]. First, 2 mmol of Bi(NO₃)₃·5H₂O were dissolved in 3 mL of ultrapure water, to which 5 mL of glacial acetic acid (PA) were added, forming precursor solution A. Simultaneously, 1 mmol of Zn(NO₃)₂·6H₂O was dissolved in 5 mL of ultrapure water, yielding solution B.

The precursor solutions A and B were mixed under constant stirring. Then, a 10 mol·L-¹ aqueous solution of KOH was slowly added dropwise until the pH of the reaction medium reached 13. The resulting suspension was kept under moderate stirring for 2 hours and then transferred to a Teflon bottle, which was sealed and placed in a hydrothermal reactor (Welljoin brand). The system was then subjected to hydrothermal treatment at 180°C for 12 hours, using a muffle furnace (Jung brand) with a heating rate of 5°C/min. After the process, the reactor was cooled to room temperature, and the material obtained was carefully collected and washed repeatedly with ultrapure water until the supernatant presented neutral pH. Finally, the sample was dried in an oven at 80°C for 24 hours, deagglomerated and named ZB1.

2.3. Synthesis of Nanocomposites Using the Monowave 50 Reactor

The synthesis of zinc bismuthate-based nanocomposite was also performed using a more robust reactor, with better control of heating, pressure, time and temperature (Monowave 50, Anton Paar), where it was possible to optimize the synthesis method at 180 °C in only 1 hour. Thus, solution A was prepared by dissolving 2 mmol of Bi(NO₃)₃·5H₂O in 3 mL of ultrapure water, followed by the addition of 5 mL of glacial acetic acid (P.A.).

Simultaneously, solution B was prepared from 1 mmol of Zn(NO₃)₂·6H₂O in 6 mL of ultrapure water at room temperature. The precursor solutions (A and B) were then mixed under stirring and the pH of the mixture was adjusted to 13 by adding, dropwise, 10 mol·L·¹ aqueous KOH solution. The suspension was transferred to the reaction tube of the Monowave 50 system, where it was subjected to programmed hydrothermal treatment. After cooling to room temperature, the material was collected and subjected to successive washings with ultrapure water until the supernatant presented neutral pH. The sample was dried in an oven at 80°C for 24 hours, deagglomerated and labeled as ZB4.

2.4. Characterization

The crystal structure of the zinc bismuthate-based nanocomposites was investigated by X-ray diffraction (XRD) using a Bruker D8 Advance diffractometer under copper $k\alpha 1/k\alpha 2$ radiation (λ = 1.540598/1.544426Å), at 40 kV and a current of 40 mA. The diffraction patterns were obtained in a scanning range of 2θ = 10–80°, with a step of 0.02° and a scanning speed of 2°/min. In order to complement the structural analyses, Raman spectroscopy spectra were acquired using a T64000 spectrometer (Horiba/Jobin Yvon) using a 532 nm laser at a power of 90 mW, covering the spectral range from 100 a 1000 cm⁻¹. The morphological and structural characterization of the samples was performed by high-resolution transmission electron microscopy (HR-TEM), using the MEI



equipment, model Tecnai FEG G2F20, operating at an accelerating voltage of 200 kV. The microscope is equipped with EDS (Energy Dispersive X-ray Spectroscopy) and EELS (Electron Energy Loss Spectroscopy) detectors, coupled to the scanning transmission electron microscopy (STEM) system. For this analysis, samples ZB1 and ZB4 were dispersed in isopropyl alcohol and each solution was placed on a copper (Cu) grid with carbon film. The determination of the zeta potential was performed for the ZB4 sample, by means of the aqueous dispersion of 0.001 g of the powder in 10 mL of ultrapure water (pH 7), using the Zetasizer Nano ZS90 equipment (Malvern Instruments®). The suspension was sonicated for 2 minutes before the measurements, ensuring the adequate dispersion of the particles.

2.5. Antimicrobial Activity Assay

The microorganisms were cultured in Petri dishes with BHI (Brain Heart Infusion) agar and incubated at 37 °C \pm 1 °C for 24 hours. The evaluation of the inhibition zones was performed in triplicate, with the measurement of the diameters in millimeters using a millimeter ruler, followed by the calculation of the arithmetic mean and standard deviation of the values obtained. ZB1 and ZB4 samples, in the amount of 10 mg, were dispersed in DMSO solution (10%, pH 7), and 50 μ L of each solution was applied to 6 mm diameter wells in plates seeded with pathogenic bacteria and fungi. For the control test, only DMSO was used. The microbial suspension was standardized by the McFarland scale (1.5 x 108 CFU/mL for bacteria and 6 x 108 CFU/mL for fungi). The tests followed the disk diffusion method adapted from Clinical Laboratory Standard Institute (CLSI) [28]. For this test, the strains tested were the bacteria *Corynebacterium diphtheriae* (ATCC 36053), *Staphylococcus aureus* (ATCC 29213) and *Klebsiella pneumoniae* (ATCC 700603), as well as the fungi *Candida albicans* (ATCC 14053) and *Cryptococcus neoformans* (clinical isolate).

In addition, the antimicrobial activity of the nanoparticles was evaluated by determining the Minimum Inhibitory Concentration (MIC), using an adaptation of the broth microdilution method performed in 96-well plates, according to the CLSI guidelines [29]. All materials used in the microbiological assay, such as tips, plates and tubes, were previously autoclaved, and a 2000 μ g/mL solution of the nanocomposites (ZB1 and ZB4) was prepared with 50% DMSO. In each well, 100 μ L of BHI broth and 100 μ L of the nanocomposite solution were added, followed by serial microdilutions. After that, 10 μ L of bacterial or fungal inoculum (10 5 CFU/mL) were added to the respective wells. As a positive control, only the culture medium and the microbial inoculum were used. The negative control consisted of the culture medium containing the microbial inoculum together with the drugs chloramphenicol (antibiotic) or amphotericin B (antifungal). Furthermore, a sterility control was performed to ensure the absence of contamination in the samples containing ZB1 and ZB4. All assays were performed in triplicate and incubated at 37°C ± 1°C for 24 hours.

2.6. Toxicity Analysis

The toxicity test was performed with the aim of analyzing the possible toxic effects of zinc bismuthate nanocomposite samples on *Tenebrio molitor* larvae. The Petri dishes were identified with the sample data, with one dish for each sample and a control group using PBS (Phosphate Buffered Saline).

The larvae were carefully selected, separated and subjected to an aseptic process, which consisted of rapid immersion in 70% ethanol for 30 seconds, followed by rinsing with sterile saline solution (PBS) and drying under aseptic conditions on sterile paper. After the procedure, the larvae were kept in sterile containers until the time of the experiments, ensuring that they were of similar size, weight and larval stage. Then, 10 larvae were placed on each plate and chilled on ice to induce a state of hibernation, which facilitated their handling. Using 1 mL disposable syringes and needles, $10 \, \mu$ L of the samples were injected into the third ring of each larva, forming groups of 10 larvae on each Petri dish. Subsequently, the plates with the larvae were kept at room temperature and monitored daily for a period of 10 days. The data obtained were organized and analyzed using the Origin software.

2.7. Statistical Analysis

The results are expressed as mean \pm standard deviation and were analyzed using GraphPad Prism 5.0. Statistical evaluation was performed using analysis of variance (ANOVA). The data obtained from the toxicity assay using the synthetic compounds were first subjected to the Kolmogorov-Smirnov test. The Kruskal-Wallis test was used to compare the treatments with the positive control. The survival curve was plotted based on the Kaplan-Meier analysis and the results were analyzed by the log-rank test. In all tests, a significance level of 95% was considered a significant difference (p < 0.05).

3. Results and Discussion

3.1. X-Ray Diffraction Analysis of Zinc Bismuthate-Based Nanocomposites

Figure 1 shows the diffractograms of samples ZB1 and ZB4, synthesized by hydrothermal method. For sample ZB1, the diffraction patterns indicated the predominant formation of the cubic crystal structure of zinc bismuthate (Zn0.667Bi25.333O40) with intense peaks at 2θ = 27.6°, 32.8° and 52.2° corresponding to the planes (310), (321) and (035), according to the ICDD PDF reference standard no. 01-070-1700. Similarly, sample ZB4 exhibited diffraction peaks that were also indexed according to the same pattern, corroborating the presence of the same cubic crystalline phase characteristic of zinc bismuthate. However, secondary peaks were also detected at 2θ = 31.8°, 34.5° and 36.3°, corresponding to the (100), (002) and (101) planes, characteristic of the hexagonal phase of wurtzite-type zinc oxide (ZnO), according to the ICDD PDF standard nº 36-1451 [30]. These results suggest the presence of a secondary ZnO phase in the ZB4 sample, indicating the formation of the Zn0.667Bi25.333O40/ZnO nanocomposites, which can significantly impact the structural and functional properties of the material.

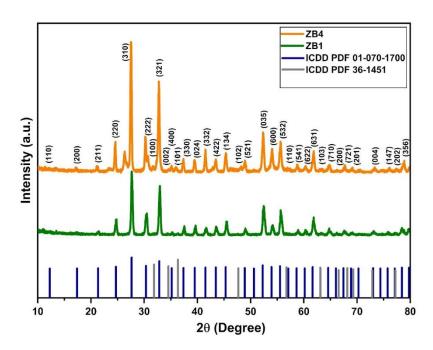


Figure 1. Diffractograms of ZB1 and ZB4 samples obtained by hydrothermal synthesis.

3.2. Vibrational Analysis by Raman Spectroscopy: Structural Confirmation and Effects Induced by ZnO e^{3+}

Figure 2 illustrates the Raman spectra of samples ZB1 and ZB4, allowing the investigation of the vibrational characteristics of the nanocomposites. The results of the factor group analysis performed for both structures, based on factor group, are presented in Table S1 (see Supplementary Material, Table S1), which shows the contribution of each type of atom to the total vibrational degree of

freedom of each crystallographic structure. The reducible symmetry equations for both phases are presented below: i) $Zn_{0.667}Bi_{25.333}O_{40}$ (cubic structure, Fd-3m), Γ optical = A_{1g} + E_{g} + $3T_{2g}$ (Raman) + $4T_{1u}$ (IR) + T_{1u} + $2A_{2u}$ + $2E_{u}$ + $2T_{2u}$ (inactive); ii) ZnO (wurtzite-like hexagonal structure, P6₃mc), Γ optical = A_{1} + $2B_{1}$ + E_{1} + $2E_{2}$, where modes A_{1} and E_{1} are active in both Raman and infrared (IR), E_{2} is exclusively Raman active and B_{1} is inactive under ideal symmetry [31]. In Figure 2, the peaks located at 90 cm⁻¹ and 128 cm⁻¹ were related to the E_{g} and T_{2g} modes, characteristic of the cubic zinc bismuthate matrix, reflecting coordinated displacements of the metal atoms in Zn–Bi–O [32,33]. The presence of ZnO with wurtzite structure was confirmed by the peaks at 203 cm⁻¹ and 326 cm⁻¹, associated with the $E_{2(low)}$ and $2E_{2}$ modes, respectively, attributed to the vibrations of the Zn–O subarray [34]. The mode at 276 cm⁻¹ was attributed to the stretching of the Bi–O bond, while the modes at 458 cm⁻¹ and 535 cm⁻¹ corresponded, respectively, to the degenerate triple bending T_{2g} due to the symmetric stretching (A_{1g}) of Bi–O and O–O [35–37]. The lower intensity peaks observed at 622 cm⁻¹, 829 cm⁻¹ and 930 cm⁻¹ indicated the presence of structural defects associated with oxygen vacancies and harmonic vibrational modes [38].

These oxygen vacancies may contribute to the non-stoichiometry of the compounds and justify the formation of ZnO. The presence of ZnO in the zinc bismuthate matrix suggests induction of structural micro deformations, evidenced by the broadening and shifting of the Raman peaks, especially in the T_{2g} and A_{1g} modes [39] and calculated by the Williamson Hall method based on the XRD data of ZB1 and ZB4, with average microstrain densities of 2.204 and 1.68 μ .nm⁻², respectively (see Supplementary Material, Table S2) [40]. These more expressive microstrain density data in sample ZB1, which decrease with increasing hydrothermal treatment temperature, were attributed to the partial replacement of Bi³⁺ by Zn²⁺, resulting in local stresses and symmetry breaking in the crystal lattice and allowing the spectroscopic activation of modes that would supposedly be inactive [41].

Such structural effects, especially local micro deformations and degeneracy of vibrational modes, are phenomena discussed in the literature on doped oxides that can favor the generation of oxidative radicals (ROS) [42]. Although no ZnO phase was detected in ZB1 sample by X-ray diffraction (XRD), possibly due to its concentration being below the detection limit of the technique, its presence was confirmed by Raman spectroscopy, which showed greater sensitivity in identifying this minority phase.

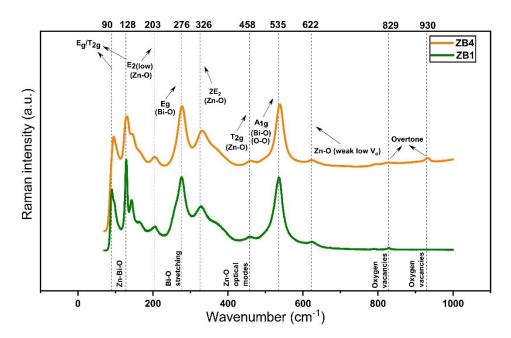


Figure 2. Raman spectra of ZB1 and ZB4.

3.3. Morphological Analysis Versus Structural Defects of Nanocomposites

Spectroscopic and microscopic analyses of samples ZB1 and ZB4 revealed the presence of oxygen vacancies (Vo), among other structural defects, which play an important role in the physicochemical and functional properties of the nanocomposites ($Zn_{0.667}Bi_{25.333}O_{40}/ZnO$), as will be detailed below.

To complement these observations, high-resolution transmission electron microscopy (HR-TEM) analysis was conducted to more accurately evaluate the morphology and crystal structure of the nanoparticles. HR-TEM images and selected electron diffraction (SAED) patterns allowed the investigation of the crystal organization of the samples, while the mean particle size distribution and elemental mapping provided additional information on the homogeneity and composition of the nanocomposites, as illustrated in Figures 3 and 4.

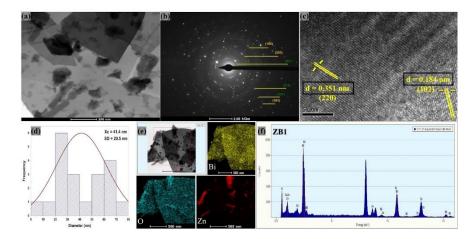


Figure 3. TEM image of ZB1 (a). SAED pattern of ZB1 (b). HR-TEM images of ZB1 (c). Average particle size distribution of ZB1 (d). Collected area of ZB1 and elemental mapping (e). EDX spectrum of ZB1 (f).

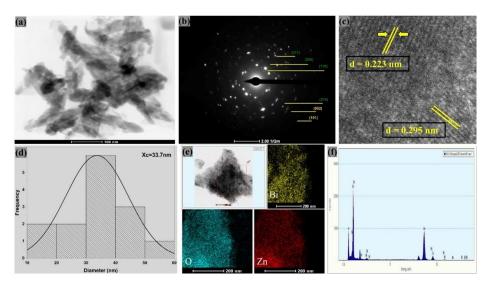


Figure 4. TEM image of ZB4 (a). SAED pattern of ZB4 (b). HR-TEM images of ZB4 (c). Average particle size distribution of ZB4 (d). Collected area of ZB4 and elemental mapping (e). EDX spectrum of ZB4 (f).

The TEM micrographs of both materials, as shown in Figures 3 (a) and 4 (a), confirm the presence of agglomerated nanoparticles, mixed morphology and lamellar structures, which are consistent with materials containing ZnO and zinc bismuthate, as they can present anisotropic growth [43]. Smaller particles have a larger surface area due to their high surface-to-volume ratio, resulting in a greater number of exposed atoms and, consequently, more dangling bonds. As a result, the surface atoms of these particles tend to interact with each other, promoting the formation of agglomerations [44].

Figure 3 (a) shows the presence of nearly spherical nanoparticles, associated with ZnO, and elongated or lamellar structures, possibly attributed to Zn_{0.667}Bi_{25,333}O₄₀, in sample ZB1. As for ZB4, as illustrated in Figure 4 (a), lamellar structures are also observed. These structures have promising potential for application as antimicrobial materials, since they can favor the interaction with microorganisms. Furthermore, the presence of these distinct forms suggests that nanocomposites may offer different mechanisms of interaction with bacteria and fungi, increasing their antimicrobial efficacy [14,17]. It is known that the synthesis process significantly influences the size and morphology of nanomaterials. Therefore, specific synthesis conditions, such as greater control of temperature, pressure and reaction time, can interfere with morphological changes [45].

The crystalline nature of the Zn_{0.667}Bi_{25.333}O₄₀/ZnO samples is confirmed by HR-TEM images. In Figure 3 (c), interplanar spacings of 0.184 nm and 0.351 nm are observed for ZB1, attributed to the (102) planes of the hexagonal structure of wurtzite ZnO and (220) of the cubic phase of Zn_{0.667}Bi_{25.333}O₄₀, respectively. Figure 4 (c) reveals spacings of 0.295 nm and 0.223 nm for ZB4, corresponding to the (100) planes of wurtzite ZnO and (024) of the cubic phase of Zn_{0.667}Bi_{25.333}O₄₀. These results indicate a strong interaction between the materials in the produced nanocomposites. In the case of ZB4, the data suggest a layered growth, similar to lamellae, associated with the (310) crystal plane, as evidenced by the X-ray diffraction (XRD) technique.

The electron diffraction patterns obtained by SAED demonstrate the crystalline structure of the ZB1 and ZB4 samples, as illustrated in Figures 3 (b) and 4 (b), revealing continuous diffraction rings and bright spots. The continuous rings are a clear sign of high crystallinity. This pronounced crystallinity, together with the polycrystalline characteristic of the material, indicates an organized atomic structure [45]. Figures 3 (d) and 4 (d) also show the average particle size (analysis by ImageJ software) being 41.4 nm for ZB1 and 33.7 nm for ZB4. These results confirm that both materials are formed by nanoparticles. This observation is consistent with the calculated average crystallite sizes, being 21.58 nm for sample ZB1 and 25.93 nm for ZB4 (see Supplementary Material, Table S2). Furthermore, only Zn, Bi and O peaks were observed in the elemental mapping analysis (EDX), thus, no impurities were detected, as shown in Figures 3 (f) and 4 (f).

Based on zeta potential analysis, the solution containing Zn_{0.667}Bi_{25.333}O₄₀/ZnO (ZB4) nanoparticles was stable, since the zeta potential value was between -30 mV and 30 mV. According to the Malvern Instruments manual [46], zeta potential values between +30 mV and -30 mV facilitate electrostatic interactions between particles and microorganisms. This range of zeta potential values leads to the formation of stable suspensions, due to the intense repulsive forces between particles in a liquid medium, which considerably reduces particle agglomeration [47].

3.3.1. Spectroscopic Evidence of Oxygen Vacancies

The Raman spectra of samples ZB1 and ZB4 (section 3.2) showed additional bands at 622 cm⁻¹, 829 cm⁻¹ and 930 cm⁻¹, attributed to vibrations associated with structural defects, including oxygen vacancies (V_o). In this context, the presence of oxygen vacancies can activate additional vibrational modes in the Raman spectrum, which under ideal crystal symmetry conditions would be inactive [48]. In support of this evidence, high-resolution transmission electron microscopy (HR-TEM) images (Figures 3C and 4C) showed regions with imperfect stacking of crystal planes, indicating the presence of structural flaws and grain boundaries. These features are consistent with the existence of oxygen vacancies, which can cause local micro deformations in the crystal structure. Furthermore, partial replacement of Bi³⁺ ions by Zn²⁺ (section 3.2) may introduce lattice strains, promoting the formation of oxygen vacancies to compensate for the charge difference [49,50].

3.3.2. Functional Implications: Potential for Generation of Reactive Oxygen Species (ROS)

The presence of oxygen vacancies (V_0) can significantly influence the electronic and optical properties of nanocomposites. In particular, V_0 can act as regions capable of capturing and retaining electrons or holes, directly interfering with the mobility of charge carriers and the functional performance of the material, affecting the electrical conductivity, optical properties, photocatalytic

and antimicrobial response of the materials. Studies also report that ZnO nanostructures containing V_{\circ} can generate reactive oxygen species (ROS) on their surface even in the absence of light [51]. Furthermore, local distortions associated with V_{\circ} can impact structural stability and interaction with adsorbed molecules, important aspects for applications in pharmaceutical devices and sensors [52,53].

The surface charge of metal oxide particles and, consequently, their stability are influenced by pH and the point of zero charge (PZC), and changes in these characteristics have a direct impact on the biological activity of the particles. PZC is the pH at which the surface charge and zeta potential are equal to zero [54]. For sample ZB4, PZC was determined at pH 7.6, indicating that its surface is electrically neutral at this value. At pHs below 7.6, the sample surface showed a positive charge, while at higher values, it showed a negative charge.

3.4. Evaluation of the Disk Diffusion Test

According to the methodology used, the presence or absence of inhibition zones against the studied pathogens is shown in Table 1.

Table 1. Inhibition zones formed in disk diffusion test (in mm). The symbol (-) indicates absence of a visible inhibition zone.

Microorganisms	ZB1	ZB4	
C. diphtheriae	10.7 ± 1.1	11.3 ± 1.1	
S. aureus	10.7 ± 1.2	11.3 ± 1.2	
C. neoformans	10.7 ± 1.1	11.7 ± 2.9	
K. pneumoniae	-	-	
C. albicans	-	-	

The analyzed samples were active against the microorganisms *Corynebacterium diphtheriae* (CD), *Staphylococcus aureus* (SA) and *Cryptococcus neoformans* (CN). Among them, ZB4 was the most promising with halos \geq 11.3 mm. It is possible to verify that sample ZB4 caused an inhibition halo of 11.7 mm for *CN*.

Determining the mechanism responsible for antimicrobial activity is challenging, as it involves several interconnected factors, such as particle size, surface charge, interaction with the bacterial cell wall and the production of reactive oxygen species (ROS) generated by metal ions [55,56].

The results show that the bactericidal activity of zinc bismuthate nanocomposites, represented by samples ZB1 and ZB4 against Gram-positive *C. diphtheriae* (CD) and *S. aureus* (SA), is strongly associated with the generation of reactive oxygen species (ROS) such as OH•, H₂O₂, O₂•- e O₂, even in the absence of light. This ROS generation by the nanocomposites can be attributed to the previously discussed structural and surface defects, representing a key mechanism for the inhibition of bacterial activity. Cell membrane damage is commonly reported as a result of an increase in ROS generation [57–59].

The ZB1 and ZB4 strains also demonstrated antifungal activity against *C. neoformans* (CN). One possible explanation for this efficacy is the effect generated by ROS that damage the cellular structures of the fungus, inhibit the biological functions of macromolecules and block DNA replication. In general, these species are effective in disrupting the functioning of efflux pumps, which reduces the ability of microorganisms to eliminate toxins and waste products from the cell [47,60].

The presence of lamellar structures, more evident in ZB4, contributes to greater adhesion to the cell surfaces of microorganisms, increasing the chance of direct interaction with the cell wall and favoring the penetration of generated ROS, which is in line with the superior performance observed in tests with *C. neoformans* [61].

Bacterial cells have a negative charge due to the abundant number of carboxyl groups that dissociate, resulting in a negatively charged cell surface. This is also observed in the fungus *C. neoformans*, which has a negatively charged polysaccharide capsule. In contrast, the ZB4 nanoparticles

have a positive charge band, as mentioned above, with a PZC of 7.6. Therefore, at pH 7, the positive charge of the sample may have caused a strong electrostatic interaction with the bacteria *C. diphtheriae* and *S. aureus*, and with the fungus *C. neoformans*, resulting in damage to the cell membrane and preventing microbial growth [62–65].

ZB1 and ZB4 did not demonstrate inhibition of the Gram-negative bacterium *K. pneumoniae* (KP), as shown in Table 1. This may be attributed to the fact that unlike Gram-positive bacteria, the cell wall structure of Gram-negative bacteria is formed by lipids, proteins and lipopolysaccharides, which offers effective protection against biocides, thus hindering the action of ROS from the nanocomposites ZB1 and ZB4. Furthermore, when compared to the other Gram-positive pathogen *S. aureus* (SA), KP has a greater negative charge, which is due to the difference in the polarity of the cell membranes between these bacteria. Thus, this would allow a more intense penetration of negatively charged free radicals, such as peroxide ions, which cause damage and cell death in *SA* [66–68].

It was also possible to observe the absence of inhibition zone for ZB1 and ZB4 against the fungus *Candida albicans* (CA). This can be attributed to the intrinsic resistance of this microorganism, since there may be changes in its morphology that consequently disable the fungal lysis process. Furthermore, the fungus Candida can demonstrate resistance mediated by efflux pumps, a process that leads to a decrease in the concentration of the antifungal agent within the fungal cell [69,70].

Furthermore, as observed in Table 1, sample ZB4 demonstrated greater efficiency in both bactericidal and fungicidal inhibition, presenting slightly superior results. This greater efficiency may be related to its average particle size of 33.7 nm. Although both samples (ZB1 and ZB4) are nanometric and have proven effective in the tests, the smaller particle size of ZB4 may have favored greater interaction with microorganisms, intensifying its antimicrobial activity.

3.5. Determination of Minimum Inhibitory Concentration (MIC) and Minimum Fungicidal Concentration (MFC)

To investigate the antimicrobial capacity of the particles, tests were also performed to determine the Minimum Inhibitory Concentration (MIC) and Minimum Fungicidal Concentration (MFC) for the microorganisms that presented the greatest inhibition in the disk diffusion test. Table 2 shows the results of the tests performed with the bacterium *Corynebacterium diphtheriae* (Gram-positive) and the fungus *Cryptococcus neoformans*.

Table 2. Minimum Inhibitory Concentration and Minimum Fungicidal Concentration (in μ g/mL) of the synthesized samples, Chloramphenicol and Amphotericin B against the chosen pathogenic microorganisms. The symbol (-) means not tested.

Microbial Strains	ZB1	ZB4	Chloramphenicol	Amphotericin B
C. diphtheriae (ATCC 36053)	1000	500	1000	-
C. neoformans (clinical isolate)	1000	500	-	1000

The ZB1 particles showed antimicrobial activity only at a concentration of up to $1000 \mu g/mL$ against the bacterium *C. diphtheriae* (*CD*) and the fungus *C. neoformans* (*CN*). These two microorganisms also showed sensitivity to ZB4, but at a concentration of up to $500 \mu g/mL$.

Figure 5 illustrates how these results were obtained through broth microdilution tests using resazurin. This reagent undergoes a redox reaction when exposed to substances produced by the cellular respiration of microorganisms and is transformed into resorufin. As a results, its color changes from blue to pink, indicating microbial growth [71].

The smaller average particle size of ZB4 (33.7 nm) compared to ZB1 (41.4 nm) results in higher specific surface area, enhancing contact with microorganisms and, consequently, favoring antimicrobial action, as evidenced by the higher inhibition halos and lower MIC observed [72].

It is worth mentioning that both synthesized samples (ZB1 and ZB4) presented sterility, with a bluish coloration, as can be observed in the sterility test performed in columns 9 to 11 of Figure 5. This attests that these samples remained free of contamination throughout the entire procedure.

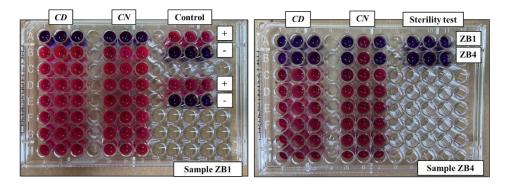


Figure 5. Broth microdilution test results of the synthesized samples against the selected pathogenic microorganisms, including control and sterility tests. The symbols (+) and (-) means positive and negative controls, respectively.

These findings align well with previous literature. For instance, Geoffrion et al. [57] highlighted the antibacterial effects of bismuth oxide nanoparticles, attributing their efficacy to the generation of reactive oxygen species (ROS). These ROS oxidize polyunsaturated fatty acids in bacterial membranes, promoting lipid peroxidation that leads to significant damage to bacterial cells and membranes [73]. ROS-mediated oxidative stress is a key mechanism behind the antimicrobial properties of bismuth oxide, and it supports the activity observed in the ZB1 and ZB4 samples.

Campos et al. [74] also demonstrated antimicrobial potential of bismuth oxide particles against common bacterial pathogens *S. aureus* and *E. coli*, reinforcing the significance of bismuth compounds in overcoming microbial resistance. Bismuth compounds are known to exhibit both antibacterial and antifungal properties, sometimes exceeding the efficacy of traditional antibiotics [10,75]. Despite this potential, their antifungal activity has not been extensively explored [75]. In comparison, studies on zinc oxide nanoparticles, such as that of Yuan et al. [76] and Alshahrani et al. [77], confirm antimicrobial (*Corynebacterium diphtheriae*) and antifungal effects (*Cryptococcus neoformans*), respectively. The promising results obtained with the zinc bismuthate particles emphasize their potential as effective antimicrobial and antifungal agents with broad-spectrum applications.

3.6. Evaluation of Toxicity Test in Alternative Model

The test and control analyses were performed using an alternative model to evaluate the toxicity of the samples on the larvae of *Tenebrio molitor* insect. These analyses produced a survival curve over a period of 10 days, as illustrated in Figure 6. The samples ZB1 and ZB4 presented a survival rate of 70% and 60%, respectively. When a Log-Rank test was performed comparing the treated groups to the control, a p-value of 0.156 was obtained, indicating that there was no statistically significant difference in toxicity between the samples and the control.

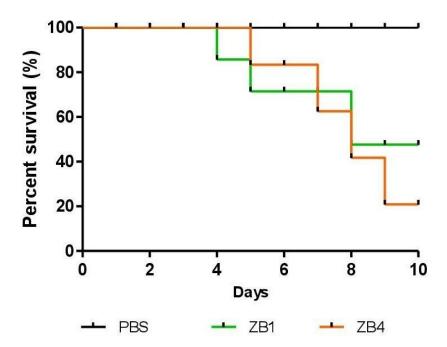


Figure 6. Survival curve of *Tenebrio molitor* larvae after a period of 10 days exposed to the samples.

In the study of Rankic et al. [78], ZnO microparticles resulted in a mortality rate of approximately 33% in Tenebrio molitor larvae. Under certain circumstances, this larvicidal effect can be triggered by inorganic particles, even when they are not formulated for pesticide purposes, as in the case with metals or metal oxides in their pure form. Furthermore, it is widely recognized that the biocompatibility of many materials, especially those at the nano or micro scale, can be limited, requiring rigorous safety assessments for biomedical applications [78,79].

The toxicity of ZnO nanoparticles was investigated by Kamaraj et al. [80]. In this study, 100% larval toxicity was observed in *Aedes aegypti* and *Anopheles stephensi*. This indicates that ZnO nanoparticles can be used in vector control and prevention of several human diseases.

Given the observed results, a possible explanation for the sublethal effects of sample ZB1 is that the particles may have caused damage to epidermal and dermal cells, leading to signs of dehydration and darkening in some larvae [78]. However, these effects did not translate into high mortality, as evidenced by the 70% survival rate. Furthermore, to the best of our knowledge, there are still no reports in the scientific literature on toxicity testing specifically involving $Z_{0.667}Bi_{25.333}O_{40}/Z_{10}O_{10}$ nanoparticles. Therefore, this work would be the first with this approach.

4. Conclusions

The synthesis of zinc bismuthate nanocomposites by hydrothermal route was shown to be feasible both in a synthesis reactor with improved temperature and pressure control and in a conventional reactor with resistive heating. In both systems, it was possible to obtain the nanocomposites under mild conditions, such as 180 °C for 1 hour and 180 °C for 12 hours, without the need for an additional calcination step. The ZB1 and ZB4 samples showed inhibitory action against *Corynebacterium diphtheriae*, *Staphylococcus aureus* and *Cryptococcus neoformans*, with emphasis on the latter, the fungus, presenting halos of 10.7 mm and 11.7 mm, respectively. No antimicrobial activity was observed against *Candida albicans* and *Klebsiella pneumoniae*. The ZB1 particles exhibited antimicrobial efficacy against *C. diphtheriae* and *C. neoformans* only at the concentration of 1000 μg/mL. These two microorganisms were also sensitive to ZB4, limited to a concentration of 500 μg/mL. Furthermore, toxicity analysis performed with *Tenebrio molitor* larvae revealed that samples ZB1 and ZB4 presented survival rates of 70% and 60%, respectively, after 10 days of exposure. Despite these differences, the Log-Rank test indicated no statistical significance in relation to the control (PBS), with

p = 0.156. These results suggest that ZB4 presents a moderate toxicity profile, making it a promising material for biomedical applications due to its effective antimicrobial activity and relative safety.

Collectively, these findings underscore the potential of ZB4 as a promising antimicrobial agent with a moderate toxicity profile, suitable for biomedical applications. The selective antimicrobial performance and favorable biocompatibility suggest that these nanocomposites could serve as effective alternatives against antibiotic-resistant pathogens, particularly *C. neoformans* and *C. diphtheriae*. Further investigations are necessary to elucidate the underlying mechanisms of antimicrobial action and to fine-tune synthesis parameters, aiming to expand the antimicrobial spectrum and foster future applications in biomedical and environmental fields.

Supplementary Materials: The following supporting information can be downloaded at the website of this paper posted on Preprints.org. Table S1: Vibrational modes identified in the Raman spectra of Zn_{0.667}Bi_{25.333}O₄₀ and ZnO nanocomposites. Table S2: Structural and microstrain parameters of ZB1 and ZB4 nanocomposites.

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Abbreviations

Abbreviations used in the manuscript:

AMR Antimicrobial Resistance
CD Corynebacterium diphtheriae
CN Cryptococcus neoformans
DMSO Dimethyl sulfoxide

MFC Minimum Fungicide Concentration
MIC Minimum Inhibitory Concentration

NPs Nanoparticles

PBS Phosphate Buffered Saline ROS Reactive Oxygen Species SA Staphylococcus aureus

ZB1 Zn_{0.667}Bi_{25,333}O₄₀/ZnO nanocomposite obtained by conventional hydrothermal synthesis

ZB4 Zn_{0,667}Bi_{25,333}O₄₀/ZnO nanocomposite obtained by Monowave 50 reactor

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