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Article

Phylogeography of Two Enigmatic Sulphur Butterflies, *Colias mongola* Alphéraky, 1897 and *Colias tamerlana* Staudinger, 1897 (Lepidoptera, Pieridae), with Relations to *Wolbachia* Infection

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Simple Summary: The genus *Colias* Fabricius, 1807 is a taxonomically challenging group of butterflies. Many *Colias* taxa display high level of intraspecific variation in wing pattern and are weakly differentiated with the respect to genitalia structure, therefore a conclusion on their status as distinct species, subspecies or local forms can be controversial. In such cases, it is crucial to conduct a comprehensive analysis based on various phylogenetic and biogeographical methods and a large-scale sampling dataset in order to resolve existing taxonomic and nomenclatural problems. In the present study, we focused on two enigmatic *Colias* taxa of unclear taxonomic status, *Colias mongola* Alphéraky, 1897 and *Colias tamerlana* Staudinger, 1897, restricted in distribution to Western Mongolia, Northwestern China and South Siberian part of Russia. Here, we conducted a DNA barcode-based analysis that revealed complicated genetic pattern with several differentiated haplotypes clustered in four distinct haplogroups. In addition, we found a strong association between a specific mitochondrial haplogroup and *Wolbachia* infection, suggesting that *Wolbachia* endosymbionts may have played an essential role in biology and diversification of the taxa in question and the genus *Colias* as a whole.

Abstract: The genus *Colias* Fabricius, 1807 includes numerous taxa and forms with uncertain status and taxonomic position. Among such taxa are *Colias mongola* Alphéraky, 1897 and *Colias tamerlana* Staudinger, 1897, interpreted in the literature either as conspecific forms, as subspecies of different but morphologically somewhat similar *Colias* species, or as distinct species-level taxa. Based on mitochondrial and nuclear DNA markers, we reconstruct a phylogeographic pattern of the taxa in question. We recover and include in our analysis DNA barcodes of the century-old type specimens, the lectotype of *C. tamerlana* deposited in the Natural History Museum (Museum für Naturkunde), Berlin, Germany (ZMHU), the paralectotype of *C.*

tamerlana and the lectotype of *C. mongola* deposited in the Zoological Institute, Russian Academy of Sciences, Saint-Petersburg, Russia (ZISP). Our analysis grouped all specimens within four (HP_I-HP-IV) deeply divergent but geographically poorly structured clades which did not support nonconspecificity of *C. mongola*–*C. tamerlana*. We also show that all studied females of the widely distributed haplogroup HP_II were infected with a single *Wolbachia* strain belonging to the supergroup B, while the males of this haplogroup, as well as all other investigated specimens, were not infected. Our data underline the relevance of large-scale sampling dataset analysis and the need of testing for *Wolbachia* infection to avoid erroneous phylogenetic reconstructions and species misidentification.

Keywords: *Colias*; DNA barcoding; Lepidoptera; molecular taxonomy; PCR screening; *Wolbachia*

1. Introduction

Colias Fabricius, 1807 (Lepidoptera, Pieridae) is one of the largest pierid genera, with approximately 90 described species [1–4]. The genus has a predominantly Holarctic distribution, with a few species occurring in the Afrotropical, Oriental and Neotropical biogeographical regions [4–7]. Although *Colias* butterflies are among the most spectacular and well-known Lepidoptera, which historically attracted much attention of researchers and collectors, their taxonomy is still poorly elucidated. *Colias* butterflies may exhibit a significant level of interspecific variation in the wing pattern. At the same time, many allopatric taxa traditionally treated as separate species may have very similar phenotypes. In addition, such taxonomically important characters as genitalia structures, commonly used for species delimitation in insects, are largely uniform in this group and do not possess reliable diagnostic features [1]. Consequently, systematics and real taxonomic status (species, subspecies or interspecific forms) of many *Colias* taxa are a matter of debate.

The issues mentioned above can be applied to *Colias tamerlana* Staudinger, 1897 (Figure 1a) and *Colias mongola* Alphéraky, 1897 (Figure 1b) distributed in Western Mongolia, Northwestern China and South Siberian part of Russia [8,9]. Contradictory taxonomic status has been suggested for this pair of taxa in literature, from synonyms or subspecies of several distinct species to separate species-level taxa (Figure 2). The former taxon was described as a separate species by Otto Staudinger in 1897 from China, Xinjiang, East Tian Shan, north of Hami, Karlik Shan Mts. ["im ostlichsten Thian-Schan-Gebirge (nordlich von Chamyl)"] [10]. The latter one was described by Alphéraky as a variety of *Colias nastes* Boisduval, 1832 [*Colias Nastes* B. var. *Mongola* Alph. var. nova] in 1897 [11]. The type locality of *C. mongola* has been the subject of considerable debates and contradictory interpreted in the taxonomic literature (Figure 3b). Korb [12] in his paper devoted to the taxonomy of *Colias cocandica* Erschoff, 1874 and allied taxa placed the type locality of *C. mongola* a few km south of Ulan-Bator (Mongolia). Grieshuber and co-authors [3] suggested that the type series of *C. mongola* originates from the vicinity of Uliastai (Mongolia), at a distance of ca. 750 km to the west from the locality indicated by Korb [12], but subsequently corrected the area to the SE Khangai Mts. [1]. Gorbunov [13] indicated the environs of the village Turan (Republic of Buryatia, Russia) as the type locality of *C. mongola*, an area located more than 500 km away from the localities suggested by Korb [12] and Greishuber [1,3]. Finally, Weiss [14] erroneously mentioned the South Altai (Kurray, Chuya, Kobdo) as the type locality of *C. mongola*.

The nomenclatural priority of the name *C. tamerlana* over *C. mongola* [13] and taxonomic status of these taxa are also disputable [2,12–20]. Staudinger treated *C. tamerlana* and *C. mongola* as synonyms [15]. Greishuber & Lamas [2] pointed out weakly expressed morphological differences between the two taxa and speculated that molecular analysis could provide evidences for synonymizing *C. tamerlana* and *C. mongola*. Talbot [16] in his "*Lepidopterorum Catalogus*" placed *C. tamerlana* as synonym of *C. cocandica maja* Grum-Grshimailo, 1891 without providing further explanations. Korb [12], based on examinations of genitalia, concluded that *mongola* and *tamerlana* represent distinct species. Tshikolovets and co-authors [17] regarded *C. mongola* as a subspecies of *C. cocandica*. These two taxa are thought to be biogeographically isolated (the eastern limit of *C. cocandica* distribution range lies in SE Kazakhstan (South Dzungaria) and NW China (Borohoro Mts.)) [3], thus

the conclusion of their conspecificity is rather doubtful. It should be noted that butterflies from Lake Khovsgol area (Mongolia), currently considered as easternmost known population of *C. tamerlana/mongola*, were originally referred to much westerly distributed *C. cocandica* and were described as a subspecies of the latter, *C. cocandica sidonia* Weiss, 1968 [14]. Some authors regarded this taxon as subspecies of *Colias nastes* Boisduval, 1832, without considering the relationships between *C. mongola* and *C. tamerlana* and their taxonomic status [18–21], while mentioning that *C. mongola* was formerly treated as separate species.

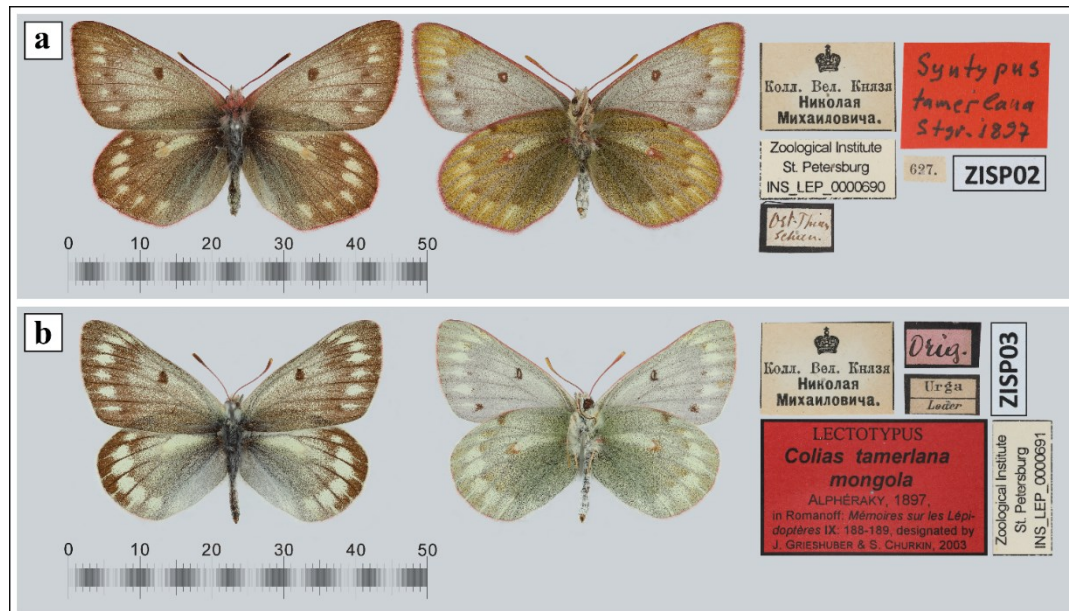


Figure 1. Type specimens of the *Colias* species deposited in the Zoological Institute, Russian Academy of Sciences, Saint-Petersburg, Russia (ZISP), barcoded in the present study: (a) paralectotype (female) of *C. tamerlana* Staudinger, 1897; (b) lectotype (male) of *C. mongola* Alphéraky, 1897.

To some extent, this assumption was based on the discovering of a large series of specimens, later described as *Colias nastes jacutica* Kurentsov 1970, which phenotypically somewhat resemble *sidonia*, the taxon that was provisionally considered a valid subspecies of *C. tamerlana/mongola*. In the original description [21], *Colias nastes jacutica* is interpreted as an intermediate link between *nastes* and *mongola* (sensu lato), yet this assumption requires further confirmation (in particular, by applying molecular methods), given that *sidonia* and *nastes* are separated by more than 2500 km without any record for the species in the gap. The second subspecies of *C. tamerlana/mongola*, *C. mongola ukokana* Korb & Yakovlev, 2000, was described from the Ukok Plateau (Republic of Altai, Russia)[22], but currently *ukokana* is considered a synonym of the nominotypical *mongola* [1–3].

The correct delineation and identification of species is not always possible based on characters of external morphology alone, especially in such taxonomically challenging groups as *Colias* [23–26]. Over the last decades, the rapid development and implementation of new molecular techniques and approaches validated mitochondrial (e.g. DNA barcodes) and nuclear DNA markers as a useful and efficient tool for species identification, detecting and analyzing cryptic diversity, revealing interspecific and deep-level relationships and phylogenetic structure in taxonomically challenging groups of insects, solving long-standing taxonomic problems [23,27–39]. Despite broad and constantly growing usage of DNA-based techniques in butterfly taxonomy and molecular systematics, surprisingly few studies focused so far on the genus *Colias* [6, 42–46]. Moreover, all but a few research aimed at inferring *Colias* phylogeny or relating to molecular aspects of the genus were based on a limited set of taxa and/or poor specimen sampling. Some recent studies and our unpublished data suggested that morphologically well-differentiated *Colias* species may have nearly identical or even shared *COI* (cytochrome c oxidase subunit 1 gene) haplotypes. For instance, D’Ercole and co-authors [47] showed that all 22 North American *Colias* species shared at least one of

the revealed barcode sequences with another species. Conversely, two or more deeply diverged mitochondrial lineages (e.g. DNA barcodes) may be found within the same species [42]. In such cases, imbalanced taxa sampling, and/or testing only a small number of specimens per species could obscure the true phylogenetic relationships within the genus, and lead to species misidentification and incorrect taxonomical conclusions. To overcome this problem, all *Colias* taxa should be analysed in detail to ascertain their actual taxonomic status and avoid erroneous interpretations.

Here, we analysed three nuclear genes, *Ca-ATPase* (sarco/endoplasmic reticulum calcium ATPase), *H3* (Histone h3) and *CAD* (Carbamoyl-Phosphate Synthetase 2, Aspartate Transcarbamylase and Dihydroorotase), as well as the barcoding region of the mitochondrial *COI* gene for a large set of *C. mongola* and *C. tamerlana* samples, including recovered for the first time DNA barcodes of the century-old type specimens. We also conducted PCR screening for three *Wolbachia* genes of the collected *C. mongola* and *C. tamerlana* butterflies in order to reveal patterns of *Wolbachia* infection.

Basing on a large-scale sampling dataset, our study aimed to (i) reveal the phylogeographic structure of *C. mongola* and *C. tamerlana*, (ii) clarify their taxonomic status, contradictory interpreted in the literature, (iii) shed light on the role of *Wolbachia* in evolutionary history and observed biogeographic patterns of the taxa in question.

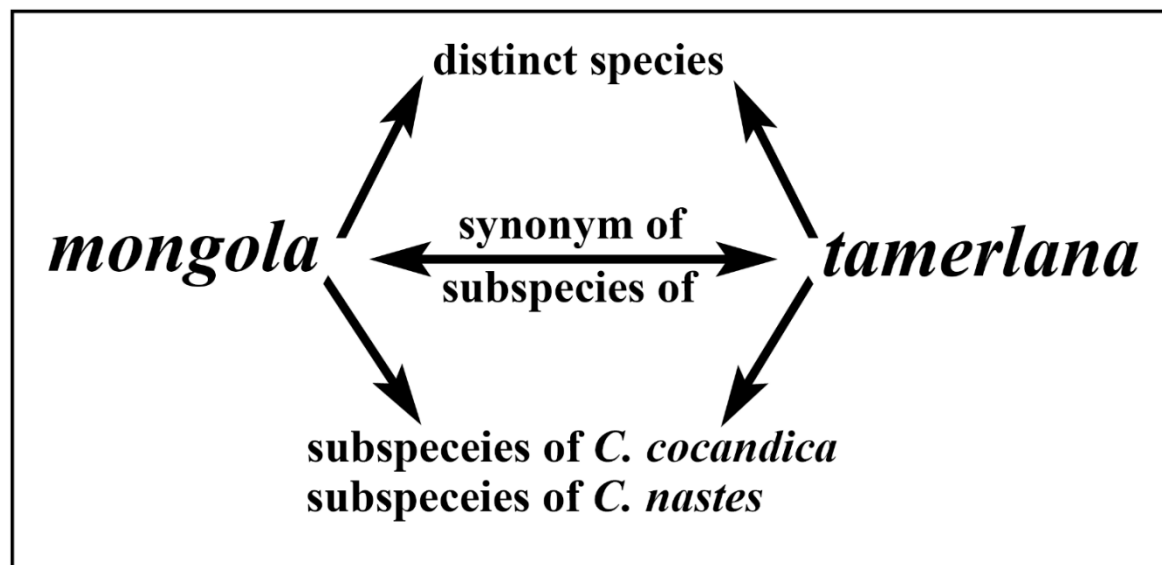


Figure 2. Relationships of *C. mongola*, *C. tamerlana* and the related taxa suggested by different authors (see text for explanation).

2. Materials and Methods

2.1. Taxon Sampling

Ninety-one specimens of *C. mongola* and *C. tamerlana* were collected in various localities of Western Mongolia, Northwestern China, and South Siberian part of Russia (Figure 3a), covering the known distribution ranges of these taxa. All sampling sites were partitioned into eighteen populations (Figure 3b). Samples either were preserved in 96% alcohol or were kept dry for subsequent molecular analysis. The lectotype of *C. tamerlana* deposited in the Natural History Museum (Museum für Naturkunde), Berlin, Germany (ZMHU), the paralectotype of *C. tamerlana* and the lectotype of *C. mongola* deposited in the Zoological Institute, Russian Academy of Sciences, Saint-Petersburg, Russia (ZISP), four specimens of *C. tamerlana sidonia* from the type locality deposited in the Zoological Museum of the Moscow State University, Moscow, Russia (ZMMU), and three samples mined from BOLD database (<http://www.boldsystems.org>, accessed on 03 September 2022) were included in the sampling dataset. Thus, the final dataset included 101 specimens. The list of the

specimens used for the molecular analysis with identification codes and collection data is given in Table 1.

Table 1. List of the studied specimens. LT – lectotype; PT – paratype; PLT – paralectotype; * – DNA barcodes, mined from BOLD; ** – numbers in brackets correspond to sampling localities, shown in Figure 3b.

Taxon	Sample ID	Sex	COI haplotype	Wolbachia			Year	Locality
				16S	wsp	ftsZ		
<i>tamerlana</i>	Akr01	♂	hp3a	-	-	-	2016	(01)** China, Xinjiang
<i>tamerlana</i>	Akr02	♂	hp3a	-	-	-	2016	(01) China, Xinjiang
<i>tamerlana</i>	Akr03	♀	hp3a	-	-	-	2016	(01) China, Xinjiang
<i>tamerlana</i>	Akr04	♀	hp3a	-	-	-	2016	(01) China, Xinjiang
<i>tamerlana</i>	CL162	♂	hp3a	-	-	-	2016	(01) China, Xinjiang
<i>tamerlana</i> LT	21128B07	♂	hp1b	n/a	n/a	n/a	1896	(01) China, Xinjiang
<i>tamerlana</i> PLT	20022A02	♀	hp1b	n/a	n/a	n/a	1896	(01) China, Xinjiang
<i>mongola</i>	Mnt03	♂	hp1b	-	-	-	2018	(02) Mongolia, Hovd
<i>mongola</i>	Mnt04	♀	hp3b	-	-	-	2018	(02) Mongolia, Hovd
<i>mongola</i>	CL34m	♂	hp1c	-	-	-	2015	(02) Mongolia, Hovd
<i>mongola</i>	Mnt54	♀	hp1b	-	-	-	2003	(17) Mongolia, Hovd
<i>mongola</i>	Mnt55	♀	hp2	+	+	+	2003	(17) Mongolia, Hovd
<i>mongola</i>	Mnt56	♀	hp1b	-	-	-	2003	(17) Mongolia, Hovd
<i>mongola</i>	Mnt57	♀	hp1b	-	-	-	2003	(17) Mongolia, Hovd
<i>mongola</i>	Mnt58	♀	hp2	+	+	+	2003	(17) Mongolia, Hovd
<i>mongola</i>	Mnt59	♀	hp3a	-	-	-	2003	(17) Mongolia, Hovd
<i>mongola</i>	Mnt60	♀	hp2	+	+	+	2003	(17) Mongolia, Hovd
<i>mongola</i>	Mnt61	♂	hp1b	-	-	-	2003	(17) Mongolia, Hovd
<i>mongola</i>	Mnt62	♂	hp2	+	+	+	2003	(17) Mongolia, Hovd
<i>mongola</i>	Mnt63	♂	hp1b	-	-	-	2003	(17) Mongolia, Hovd
<i>mongola</i>	Mnt01	♀	hp2	+	+	+	2013	(03) Mongolia, Gobi-Altai
<i>mongola</i>	Mnt02	♀	hp2	+	+	+	2013	(03) Mongolia, Gobi-Altai
<i>mongola</i>	CL41m	♀	hp2	+	+	+	2017	(03) Mongolia, Gobi-Altai
<i>mongola</i>	Nsk16	♂	hp2	-	-	-	2005	(04) Mongolia, Bayan-Ulegej
<i>mongola</i>	CL39m	♀	hp2	+	+	+	2017	(05) Mongolia, Bayan-Ulegej
<i>mongola</i>	CL40m	♀	hp3a	-	-	-	2017	(05) Mongolia, Bayan-Ulegej
<i>mongola</i>	CL43m	♀	hp2	+	+	+	2017	(05) Mongolia, Bayan-Ulegej
<i>mongola</i>	Mnt24z	♀	hp3a	-	-	-	2017	(05) Mongolia, Bayan-Ulegej
<i>mongola</i>	Mnt25z	♀	hp4a	-	-	-	2017	(05) Mongolia, Bayan-Ulegej
<i>mongola</i>	Mnt53	♀	hp2	+	+	+	2004	(16) Mongolia, Zavkhan
<i>mongola ukokana</i>	Mnt18	♀	hp2	+	+	+	2022	(06) Russia, Republic of Altai
<i>mongola ukokana</i>	Mnt19	♀	hp3c	-	-	-	2022	(06) Russia, Republic of Altai
<i>mongola ukokana</i>	Mnt20	♂	hp4a	-	-	-	2022	(06) Russia, Republic of Altai

<i>mongola ukokana</i>	Mnt21	♂	hp4a	-	-	-	2022	(06) Russia, Republic of Altai
<i>mongola ukokana</i>	Mnt22	♂	hp4a	-	-	-	2022	(06) Russia, Republic of Altai
<i>mongola ukokana</i>	Mnt23	♂	hp2	-	-	-	2022	(06) Russia, Republic of Altai
<i>mongola ukokana</i>	Nsk014	♂	hp3a	n/a	n/a	n/a	1997	(07) Russia, Republic of Altai
<i>mongola ukokana</i>	Mnt34z	♂	hp3a	n/a	n/a	n/a	1997	(07) Russia, Republic of Altai
<i>mongola ukokana</i> PT	Nsk015	♂	hp4c	n/a	n/a	n/a	1995	(08) Russia, Republic of Altai
<i>mongola ukokana</i>	Mnt24	♂	hp3a	-	-	-	2001	(09) Russia, Republic of Altai
<i>mongola ukokana</i>	Mnt25	♀	hp1a	-	-	-	2001	(09) Russia, Republic of Altai
<i>mongola ukokana</i>	Mnt26	♂	hp4a	-	-	-	2001	(09) Russia, Republic of Altai
<i>mongola ukokana</i>	Mnt27	♀	hp3a	-	-	-	2001	(09) Russia, Republic of Altai
<i>mongola ukokana</i>	Mnt28	♀	hp4a	-	-	-	2001	(09) Russia, Republic of Altai
<i>mongola ukokana</i>	Mnt29	♂	hp4a	-	-	-	2001	(09) Russia, Republic of Altai
<i>mongola ukokana</i>	Mnt30	♂	hp3a	-	-	-	2001	(09) Russia, Republic of Altai
<i>mongola ukokana</i>	Mnt31	♂	hp3a	-	-	-	2001	(09) Russia, Republic of Altai
<i>mongola ukokana</i>	Mnt32	♂	hp4a	-	-	-	2001	(09) Russia, Republic of Altai
<i>mongola ukokana</i>	Mnt33	♂	hp3a	-	-	-	2001	(09) Russia, Republic of Altai
<i>mongola ukokana</i>	Mnt34	♂	hp3a	-	-	-	2001	(09) Russia, Republic of Altai
<i>mongola ukokana</i>	Mnt35	♂	hp4a	-	-	-	2001	(09) Russia, Republic of Altai
<i>mongola ukokana</i>	Mnt36	♂	hp3a	-	-	-	2001	(09) Russia, Republic of Altai
<i>mongola ukokana</i>	Mnt37	♂	hp3a	-	-	-	2001	(09) Russia, Republic of Altai
<i>mongola ukokana</i>	Mnt38	♂	hp3a	-	-	-	2001	(09) Russia, Republic of Altai
<i>mongola ukokana</i>	Mnt39	♂	hp3a	-	-	-	2001	(09) Russia, Republic of Altai
<i>mongola ukokana</i>	Mnt44	♂	hp4a	-	-	-	2001	(09) Russia, Republic of Altai
<i>mongola ukokana</i>	Mnt45	♂	hp4a	-	-	-	2001	(09) Russia, Republic of Altai
<i>mongola ukokana</i>	Mnt46	♂	hp3a	-	-	-	2001	(09) Russia, Republic of Altai

<i>mongola ukokana</i>	Mnt47	♂	hp3d	-	-	-	2001	(09) Russia, Republic of Altai
<i>mongola ukokana</i>	Mnt48	♂	hp3a	-	-	-	2001	(09) Russia, Republic of Altai
<i>mongola ukokana</i>	Mnt49	♂	hp3a	-	-	-	2001	(09) Russia, Republic of Altai
<i>mongola ukokana</i>	Mnt50	♂	hp3d	-	-	-	2001	(09) Russia, Republic of Altai
<i>mongola ukokana</i>	Mnt51	♂	hp3a	-	-	-	2001	(09) Russia, Republic of Altai
<i>mongola ukokana</i>	Ku01	♀	hp3e	-	-	-	2022	(09) Russia, Republic of Altai
<i>mongola ukokana</i>	Ku02	♀	hp2	+	+	+	2022	(09) Russia, Republic of Altai
<i>mongola ukokana</i>	Ku03	♂	hp4a	-	-	-	2022	(09) Russia, Republic of Altai
<i>mongola ukokana</i>	Ku04	♂	hp4a	-	-	-	2022	(09) Russia, Republic of Altai
<i>mongola ukokana</i>	Mnt28z	♂	hp3a	-	-	-	2022	(09) Russia, Republic of Altai
<i>mongola ukokana</i>	Mnt29z	♀	hp3e	-	-	-	2022	(09) Russia, Republic of Altai
<i>mongola ukokana</i>	Mnt30z	♂	hp3d	-	-	-	2022	(09) Russia, Republic of Altai
<i>mongola ukokana</i>	LOWA141-06*	♂	hp3a	n/a	n/a	n/a	1999	(09) Russia, Republic of Altai
<i>mongola ukokana</i>	LOWA818-06*	♂	hp3a	n/a	n/a	n/a	1999	(09) Russia, Republic of Altai
<i>mongola ukokana</i>	LOWA142-06*	♂	hp4b	n/a	n/a	n/a	1999	(09) Russia, Republic of Altai
<i>mongola ukokana</i>	CL42m	♀	hp1a	-	-	-	2017	(09) Russia, Republic of Altai
<i>mongola ukokana</i>	Mnt17	♀	hp1a	-	-	-	2022	(10) Russia, Republic of Altai
<i>mongola ukokana</i>	Mnt40	♀	hp2	+	+	+	2003	(10) Russia, Republic of Altai
<i>mongola ukokana</i>	Mnt41	♂	hp3a	-	-	-	2003	(10) Russia, Republic of Altai
<i>mongola ukokana</i>	Mnt42	♂	hp3a	-	-	-	2003	(10) Russia, Republic of Altai
<i>mongola ukokana</i>	Mnt43	♂	hp3a	-	-	-	2003	(10) Russia, Republic of Altai
<i>mongola ukokana</i>	Mnt09	♂	hp3a	-	-	-	2002	(11) Russia, Republic of Altai
<i>mongola ukokana</i>	Mnt10	♂	hp3a	-	-	-	2002	(11) Russia, Republic of Altai
<i>mongola ukokana</i>	Mnt08	♂	hp3a	-	-	-	2003	(12) Russia, Republic of Altai
<i>mongola ukokana</i>	Mnt12	♂	hp3a	-	-	-	2003	(12) Russia, Republic of Altai

<i>mongola</i>	Mnt14	♀	hp2	+	+	+	2010	(13) Russia, Republic of Tyva
<i>mongola</i>	Mnt15	♀	hp3a	-	-	-	2010	(13) Russia, Republic of Tyva
<i>mongola</i>	Mnt07	♂	hp2	-	-	-	2002	(14) Russia, Republic of Tyva
<i>mongola</i>	CL35m	♀	hp1b	-	-	-	2002	(14) Russia, Republic of Tyva
<i>mongola</i>	CL37m	♂	hp1b	-	-	-	2002	(14) Russia, Republic of Tyva
<i>mongola</i>	CL38m	♀	hp1a	-	-	-	2002	(14) Russia, Republic of Tyva
<i>mongola</i>	CL54m	♂	hp1b	-	-	-	2002	(14) Russia, Republic of Tyva
<i>mongola</i>	CL55m	♂	hp4a	-	-	-	2002	(14) Russia, Republic of Tyva
<i>mongola</i>	CL56m	♂	hp1b	-	-	-	2002	(14) Russia, Republic of Tyva
<i>mongola</i>	CL57m	♀	hp1b	-	-	-	2002	(14) Russia, Republic of Tyva
<i>mongola</i>	Mnt26z	♂	hp1a	-	-	-	2002	(14) Russia, Republic of Tyva
<i>mongola</i>	Mnt05	♀	hp1b	-	-	-	2001	(15) Russia, Republic of Tyva
<i>mongola</i>	Mnt06	♀	hp1b	-	-	-	2001	(15) Russia, Republic of Tyva
<i>mongola</i> LT	20022A03	♂	hp1b	n/a	n/a	n/a	?	(??) see text for explanation
<i>sidonia</i>	S1-22064B02	♀	hp1b	n/a	n/a	n/a	1967	(18) Mongolia, Khovsgol
<i>sidonia</i>	S2-22064B03	♀	hp1b	n/a	n/a	n/a	1966	(18) Mongolia, Khovsgol
<i>sidonia</i>	S3-22064B04	♂	hp1b	n/a	n/a	n/a	1967	(18) Mongolia, Khovsgol
<i>sidonia</i>	S4-22064B05	♂	hp1b	n/a	n/a	n/a	1967	(18) Mongolia, Khovsgol

2.2. DNA Extraction

One leg from each specimen was taken for DNA extraction. For the samples more than 10 years old, total genomic DNA was extracted using QIAamp DNA Investigator Kit (Qiagen, Venlo, The Netherlands) following the manufacturer's protocol. For the specimens up to 10 years old, DNA extraction was performed using the CTAB-based method [48] with some modifications [39,49].

2.3. Molecular Markers, PCR Amplification, and Sequencing

One mitochondrial (*COI*) and three nuclear (*Ca-ATPase*, *H3*, and *CAD*) genes were used as molecular markers. A 658 bp fragment of the *COI* gene (mitochondrial DNA barcode) was amplified using LCO1490/HCO2198 [50] and LepF/LepR primer pairs [51]. In case standard lepidopteran barcode primers failed to yield a sufficient product, we amplified full-length barcode fragment using the primer pair combinations LepF/MH-MR1 + MH-MF1/LepR and LCO1490/MH-MR1 + MH-MF1/HCO2198 [52]. Primers CAD743nF/CADmidR, CADmidF/CAD1028R [53], H3aF/H3aR [54], and Ca-ATPase_F/Ca-ATPase_R [55] were used for nucDNA amplification and resulted in 847 bp fragment of the *CAD*, 328 bp fragment of the *H3*, and 445 bp fragment of the *Ca-ATPase* genes, respectively.

The PCR amplifications were performed in a 15 µL reaction volume per sample. Each reaction contained 1 µL template DNA (ca. 10–50 ng genomic DNA), 0.9 µL of both forward and reverse

primers diluted to a standard concentration of 10 μ M, 3 μ L of 5 \times ScreenMix (Evrogen, Moscow, Russia), and 9.2 μ L of ddH₂O. The temperature profile for *COI*, *CAD*, and *Ca-ATPase* genes was as follows: initial denaturation at 95 $^{\circ}$ C for 5 min, followed by 35 cycles of denaturation at 94 $^{\circ}$ C for 30 s, annealing at 50 $^{\circ}$ C (*COI*)/55 $^{\circ}$ C (*CAD*, *Ca-ATPase*)/60 $^{\circ}$ C (*H3*) for 30 s, and extension at 72 $^{\circ}$ C for 1 min 30 s, with a final extension at 72 $^{\circ}$ C for 10 min. The purified PCR products were subjected to the further sequencing. Sequencing of the double-stranded product was carried out at the Research Resource Center for Molecular and Cell Technologies (St. Petersburg State University, St. Petersburg, Russia) using ABI 3500xL analyzer (Applied Biosystems, Waltham, MA, USA).

2.4. Processing and Sequencing of Old Type Specimens

To obtain *COI* barcodes of the century-old type specimens we followed the protocol described in detail by Li et al. [56]. In brief, a single leg was used for DNA extraction. The DNA isolation protocol was non-destructive: the entire leg was soaked in a DNA extraction solution overnight and was preserved after the extraction. Genomic libraries were constructed from total DNA and sequenced for 150 bp from both ends on Illumina HiSeq x10 (Illumina, San Diego, CA, USA) in a pool with others. The sequences were demultiplexed to assign to each specimen by the index and indices removed. The barcodes were assembled using a reference sequence as a bait. All sequence reads matching the reference were mapped to it using DIAMOND [57]. The coverage of the barcode sequence is typically high (100 to 1000-fold or more), and overlapping reads result in an unambiguous sequence. Further details of experimental and computational protocols can be found in [56].

2.5. Detection of *Wolbachia* Endosymbionts

Colias specimens were screened for the presence of *Wolbachia* infection by amplifying three *Wolbachia* genes, 16S ribosomal RNA (16S), *Wolbachia* surface protein (wsp) and Filamentation temperature-sensitive protein Z (ftsZ). We used *Wolbachia*-specific primer pairs, W-Specf/W-Specr [58], wsp81F/wsp691R [59] and ftsZ-F/ftsZ-R [60], amplifying ~ 396 bp fragment of the 16S RNA gene, ~ 549 bp fragment of the wsp gene and ~ 510 bp fragment of the ftsZ gene (actual length of PCR fragments may vary, depending on the individual *Wolbachia* strain), respectively. The PCR amplifications were performed in a 15 μ L reaction volume. Each reaction contained 1 μ L template DNA (ca. 10–50 ng genomic DNA), 0.8 μ L of both forward and reverse primers diluted to a standard concentration of 10 μ M, 3 μ L of 5 \times ScreenMix (Evrogen, Moscow, Russia) and 9.4 μ L of ddH₂O. The temperature profile for 16S, wsp, and ftsZ genes was as follows: initial denaturation at 95 $^{\circ}$ C for 5 min, followed by 40 cycles of 30 s denaturation at 95 $^{\circ}$ C, 1 min annealing at 50 $^{\circ}$ C and extension at 72 $^{\circ}$ C for 45 s, with a final extension at 72 $^{\circ}$ C for 5 min. Each PCR reaction contained two negative (PCR mix with ddH₂O instead of DNA sample) and one positive (genomic DNA of a *Wolbachia*-infected *Colias* specimen, previously successfully amplified for 16S, wsp, and ftsZ genes) controls. PCR amplification was conducted twice for each specimen in order to avoid the technical errors. To ascertain the presence/absence of *Wolbachia*, each PCR product was checked on 1% standard agarose gel. Our personal unpublished data suggests that the standard screening procedure for *Wolbachia* allows detecting infection in host specimens up to 30–35 years old, but its sufficiency largely depends on the quality of the genomic DNA and storage conditions of the specimens. Thus, to avoid false-negative results, we excluded from the analysis samples collected more than twenty years ago (before the 2000s). Specimens positive for three *Wolbachia* genes were sequenced at the Research Resource Center for Molecular and Cell Technologies (St. Petersburg State University, St. Petersburg, Russia) using ABI3500xL Genetic Analyzer (Applied Biosystems, Waltham, MA, USA).

2.6. Molecular Data Analysis and Phylogenetic Reconstructions

All sequences were checked for errors, edited and aligned using Geneious v.8.1.6 [61] and BioEdit v.7.0.3 [62] software. Primer sequences were cropped. The final *COI* dataset (alignment length 658 bp) included 97 sequences of *C. mongola*/*C. tamerlana*. Sequences of pierids *Leptidea juvernica* (Linnaeus, 1758) and *Colias croceus* (Geoffroy, 1785) (GenBank accession numbers MT210323 and

OR178497, respectively) obtained previously were included as an outgroup to root the phylogram. *COI* sequences were collapsed to unique haplotypes using online tool FaBox v.1.61 (<https://birc.au.dk/~palle/php/fabox/>, accessed on 05 November 2022) [63]. Concatenated nuclear data set (alignment length 1620 bp) included sequences of 17 *C. mongola*/*C. tamerlana* specimens representing all revealed *COI* haplotypes. Phylogenetic reconstructions placed *C. croceus* within basal lineages of the genus *Colias*, thus it was selected as the outgroup to root the nuclear phylogram [42,43]. Nucleotide substitution models for each dataset were estimated based on the Bayesian information criterion (BIC) using jModelTest v.2.1.10 [64]. The best-fitting models were as follows: JC for *COI* data set, HKY+I for concatenated *Ca-ATPase*+*H3*+*CAD* nuclear genes fragments. Bayesian analyses were performed using the MrBayes v.3.2.7a software [65]. Parameters were estimated using two independent runs of 10 million generations each with four simultaneous chains (one cold and three heated). The sampling of trees and parameters was set to every 1000 generations. The first 10% of trees were discarded as burn-in prior to computing a consensus phylogeny and posterior probabilities. TRACER v.1.6 was used for checking stationarity and convergence of Bayesian analyses between runs [66]. The consensus of the obtained trees were visualized using FigTree v.1.4.4 (<http://tree.bio.ed.ac.uk/software/figtree/>, accessed on 05 November 2022). For the analysis of the phylogeographical structure of *C. mongola* and *C. tamerlana*, a median-joining haplotype network [67] was built using popART v.1.7 software [68]. Genetic distances among *COI* barcodes were calculated using MEGA v.7.0.14 [69]. The number of polymorphic and parsimony informative sites, the number of haplotypes, haplotype (*h*) and nucleotide (π) diversities were calculated in DnaSP v.6.12.03 [70]. DnaSP was further used to infer the demographic history of *C. mongola* and *C. tamerlana* with Tajima's D [71], Fu and Li's D [72], Fu and Li's F [72], and Fu's Fs [73] statistical tests for neutrality.

2.6.1. Molecular Characterization of *Wolbachia*

The BLAST algorithm implemented in NCBI (<https://blast.ncbi.nlm.nih.gov>, accessed on 20 July 2023) was used to search for sequence similarities in GenBank database with known DNA (BLASTN) sequences. We mined 43 *16S*, 74 *wsp*, and 71 *ftsZ* sequences with the highest percentage identity match, which were included in the alignment datasets. In the final alignments, identical *Wolbachia* sequences of the same host species were limited to one record. To estimate relationships among *Wolbachia* alleles, phylogenetic analyses were conducted for each gene independently using the Bayesian inference (BI) approach, applying GTR+G (for *16S* gene fragment) and GTR + I + G (for *wsp* and *ftsZ* genes fragments) substitution models, as suggested by jModelTest v.2.1.7 [64]. All other parameters of the Bayesian analyses were the same as for *COI* alignment.

2.7. Data Availability

All sequences obtained for *COI*, *CAD*, *Ca-ATPase* and *H3* *Colias* genes, and for *16S*, *wsp* and *ftsZ* *Wolbachia* genes were deposited to GenBank under accession numbers OP946559–OP946652, OR178498–OR178501, OR178497 (*COI*), OQ192178–OQ192194 (*CAD*), OQ1921161–OQ192177 (*Ca-ATPase*), OQ192144–OQ192151 (*H3*), OQ155222–OQ155235 (*16S*), OQ192116–OQ192129 (*wsp*) and OQ192130–OQ192143 (*ftsZ*) and listed in the electronic supplementary materials (Table S1). Voucher specimens were deposited in the Department of Karyosystematics of the Zoological Institute of the Russian Academy of Sciences and private collections of A. Kir'yanov, A. Krupitsky, A. Kurmaev, S. Churkin (Moscow, Russia), B. Khramov (Saint-Petersburg, Russia) and R. Yakovlev (Barnaul, Russia).

3. Results

3.1. Phylogeographic Structure of *Colias mongola* and *Colias tamerlana*

Haplotype analysis of a dataset of 101 *C. mongola* and *C. tamerlana* specimens revealed 12 *COI* haplotypes clustering in four distinct haplogroups (HP_I – HP_IV) (Figure 4a). Each haplogroup consists of one main haplotype and one to four satellites, differing from the main haplotypes in one nucleotide substitution. The only one exception is haplogroup HP_II, which consists of a single haplotype hp2. Low rate of genetic diversity ranging from 0% to 0.07%±0.05% detected within each

haplogroup. On the contrary, sequence divergence between haplogroups is relatively high ($1.23\% \pm 0.41\%$ – $2.56\% \pm 0.62\%$), with a maximum p -distance of $2.89\% \pm 0.62\%$ between the most divergent haplotypes (Table 2).

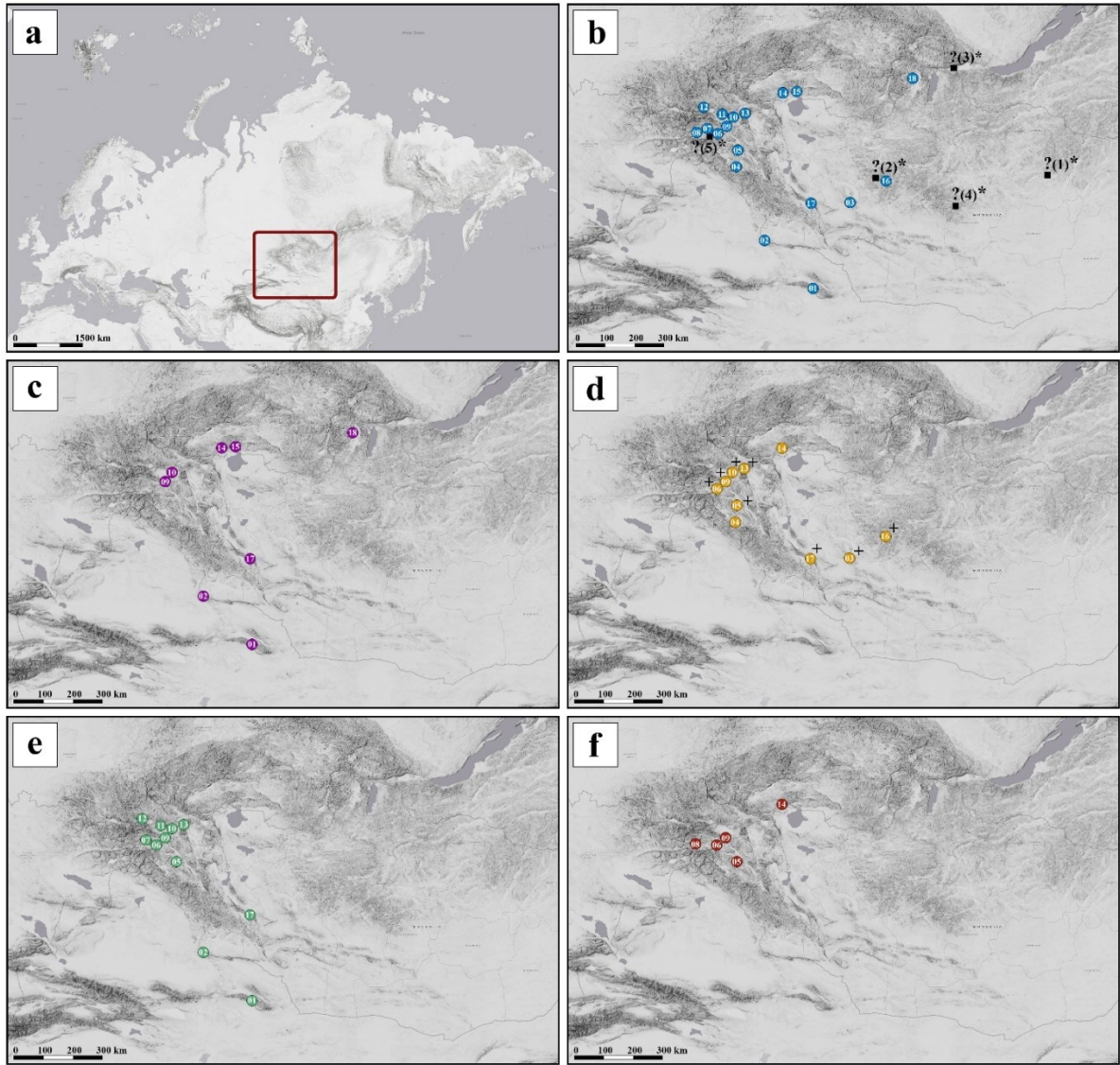


Figure 3. Maps showing location of the study area (red rectangle) (a), sampling localities of analysed specimens of *C. mongola*/*C. tamerlana* (b), and geographical distribution of revealed *COI* haplogroups of *C. mongola*/*C. tamerlana* (c-f). Question marks indicate suggested type localities of *C. mongola* (see text for explanation). *Wolbachia* infected populations indicated by "+".

In general, haplotype distribution demonstrates no clear geographical structure: haplotypes of all four haplogroups are commonly occurred in sympatry; at the same time, they can be found in the geographically remote populations (Figure 3c-f). Twenty-six specimens clustered in three haplotypes of the haplogroup HP_I which was shared by 8 populations. Seventeen specimens constituted a single haplotype of the haplogroup HP_II which occurred in 10 sampling sites. Forty-two specimens were grouped into five haplotypes of the haplogroup HP_III and sixteen specimens constituted three haplotypes of the haplogroup HP_IV, which were found in 11 and 4 sampling sites, respectively. Four haplotypes, namely hp1b, hp2, hp3a and hp4a, were the most commonly observed haplotypes out of all screened individuals. On the contrary, haplotypes hp1c, hp3b, hp3c, hp4b, hp4c consisted of only one specimen each were found in single sampling sites. Detailed data on the distribution of the haplotypes among specimens and localities is given in the Table 3.

Table 2. Summary of haplotype diversity of *C. mongola/C. tamerlana*. The number of individuals sequenced (*N*), the number of haplogroups (*HG*) and haplotypes (*H*) revealed, the number of polymorphic sites (*S*), nucleotide (π) and haplotype (*h*) diversities are given. Tajima’s *D*, Fu and Li’s *D*, Fu and Li’s *F*, Fu’s *F_s*, max. p-distance, within and between group divergence values with standard deviation (in brackets) are shown.

<i>N</i>	<i>HG</i>	<i>H</i>	<i>S</i>	π	<i>h</i>	Tajima’s <i>D</i>	Fu and Li’s <i>D</i>	Fu and Li’s <i>F</i>	Fu’s <i>F_s</i>	max. p-distance	within group divergence	between group divergence
101	4	12	28	0.0138656	0.7874	2.01246	1.21273	1.82127	7.422	2.89%	HP_I 0.04% (± 0.02%)	HP_I / HP_II 2.56% (± 0.59%)
						0.1 > p > 0.05	p > 0.1	p < 0.05	p = 0.001	(± 0.62%)	HP_II 0.00% (± 0.00%)	HP_I / HP_III 2.40% (± 0.57%)
											HP_III 0.07% (± 0.05%)	HP_I / HP_IV 2.56% (± 0.58%)
											HP_I 0.04% (± 0.03%)	HP_II / HP_III 1.39% (± 0.42%)
												HP_II / HP_IV 1.23% (± 0.41%)
												HP_III / HP_IV 1.41% (± 0.43%)

Table 3. Haplogroup and haplotype composition (the number of individuals constituted a haplogroup/haplotype (*n*), and the number of sampling sites where haplogroup/haplotype was found (*Sn*) are given) and its geographical distribution (the number of individuals (*n*), the number of haplogroups (*HGn*), the number of haplotypes (*Hn*) found in each locality and haplotype constitution are shown). * - sampling sites correspond to Figure 3b.

Haplogroup	<i>n</i>	<i>Sn</i>	COI haplotype	<i>n</i>	<i>Sn</i>	Sampling site*	<i>n</i>	<i>HGn</i>	<i>Hn</i>	COI haplotype (<i>n</i>)
HP_I	26	9	hp1a	5	4	01	7	2	2	hp1b (2), hp3a (5)
			hp1b	20	6	02	3	2	3	hp1a (1), hp1c (1), hp3b (1)
			hp1c	1	1	03	3	1	1	hp2 (3)
HP_II	17	10	hp2	17	10	04	1	1	1	hp2 (1)
			hp3a	35	9	05	5	3	3	hp2 (2), hp3a (2), hp4a (1)
			hp3b	1	1	06	6	3	3	hp2 (2), hp3c (1), hp4a (3)
HP_III	42	11	hp3c	1	1	07	2	1	1	hp3a (2)
			hp3d	3	1	08	1	1	1	hp4c (1)
			hp3e	2	1	09	35	4	7	hp1a (2), hp2 (1), hp3a (17), hp3d (3), hp3e (2), hp4a (9), hp4b (1)
HP_IV	16	4	hp4a	14	4	10	5	3	3	hp1a (1), hp2 (1), hp3a (3)
			hp4b	1	1	11	2	1	1	hp3a (2)
			hp4c	1	1	12	2	1	1	hp3a (2)
						13	2	2	2	hp2 (1), hp3a (1)
						14	9	3	4	hp1a (2), hp1b (5), hp2 (1), hp4a (1)
						15	2	1	1	hp1b (2)
						16	1	1	1	hp2 (1)
						17	10	3	3	hp1b (5), hp2 (4), hp3a (1)
						18	4	1	1	hp1b (4)

3.2. Phylogenetic Analyses of Mitochondrial and Nuclear Markers

The Bayesian phylogenetic tree for *C. mongola/C. tamerlana* COI haplotypes displayed four strongly supported (PP=1) lineages (Figure 4b) corresponding to four haplogroups revealed by haplotype network analysis. Within the *C. mongola/C. tamerlana* clade, a basal position is occupied by haplotype hp2, which is common and widespread across the geographical area studied. Further splitting recovers the clade containing specimens of HP_IV haplogroup. This clade in turn appeared as sister to the remaining two clusters, consisted of haplotypes hp3a-hp3e and hp1a-hp1c, respectively.

Combined analysis of three nuclear markers (*Ca-ATPase*, *H3*, and *CAD*) resulted in an unresolved tree for *C. mongola/C. tamerlana* that failed to recover clades revealed by BI analysis of COI barcodes (Figure 4c). In general, nuclear markers demonstrate very shallow divergence forming on

the phylogenetic reconstructions two unsupported clades (PP=0.58; PP=0.61), each consisted of specimens bearing *COI* haplotypes of different haplogroups. Both intra-individual heterogeneities and single nucleotide substitutions were found in sequenced fragments of *Ca-ATPase*, *CAD* and *H3* nuclear genes among the 17 analysed specimens of *C. mongola/C. tamerlana*. However, polymorphic sites were distributed inhomogeneously across the three genes fragments. The 847 bp fragment of the *CAD* was most variable with 23 heterogeneities/substitutions found, whereas 445 bp fragment of the *Ca-ATPase* gene and 328 bp fragment of the *H3* gene were conserved, having four and one polymorphic sites, respectively. The detailed information on the nucleotide variability of the studied nuclear genes fragments is given in the electronic supplementary (Table S2).

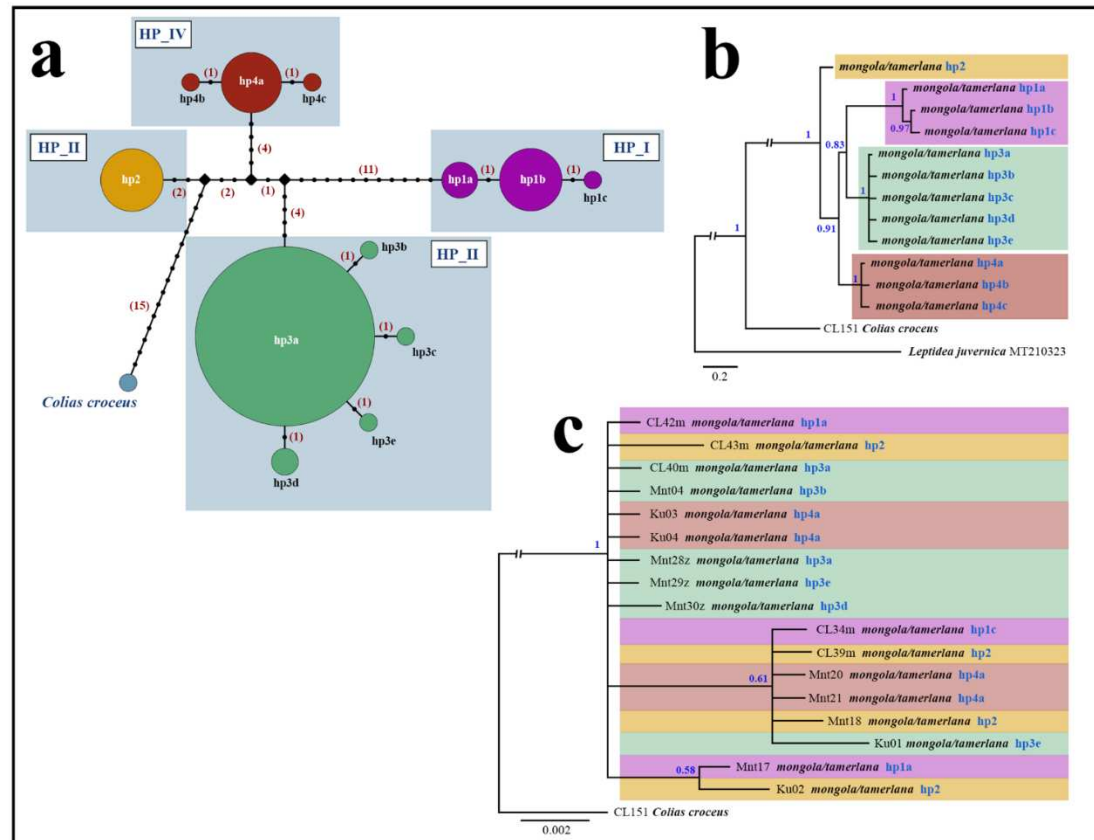


Figure 4. Phylogenetic patterns of *C. mongola/C. tamerlana* based on the analysis of mitochondrial (*COI*) and nuclear (*Ca-ATPase*, *H3*, and *CAD*) markers. (a) Median-joining haplotype network illustrating relationships of the revealed *COI* haplotypes; mutations are shown as 1-step edge; number of mutations separating haplotypes is given in parentheses. (b-c) The Bayesian consensus trees of *C. mongola/C. tamerlana* inferred from *COI* sequences (b) and concatenated alignment of three nuclear markers (c); numbers at nodes indicate Bayesian posterior probabilities.

3.3. *Wolbachia* Analysis

A total of 88 specimens of *C. mongola/C. tamerlana*, representing all four *COI* haplogroups recovered, were screened for the presence of *Wolbachia* infection. Screening for three *Wolbachia* genes (*16S*, *wsp*, and *ftsZ*) did not reveal any cases of dissimilar results (i.e., when specimen was positive for one *Wolbachia* gene, but negative for another gene/genes). In total, 14 specimens out of 88 tested were scored positive for *Wolbachia* infection (prevalence: 16%). Our analysis revealed strong sex-dependent congruence between a specific *COI* haplogroup and *Wolbachia* infection: all 14 infected specimens were females of *COI* haplogroup HP_II comprising a single haplotype hp2. Three analysed males of this haplogroup, as well as all other investigated specimens, were not infected (Figure 5). Our survey did not recover specific geographical pattern of *Wolbachia* incidence: infected specimens were randomly found in the geographically remote populations (Figure 3d).

3.3.1. Wolbachia Allele Identified in *C. mongola*/*C. tamerlana*

Wolbachia 16S, *wsp*, and *ftsZ* genes were sequenced for all infected specimens (14 females) of *C. mongola*/*C. tamerlana*. These specimens were infected by a single allele belonged to the supergroup B (grouping according to [74], which was designated as *wTam* (allele name follow the widely accepted abbreviation style [75,76]).

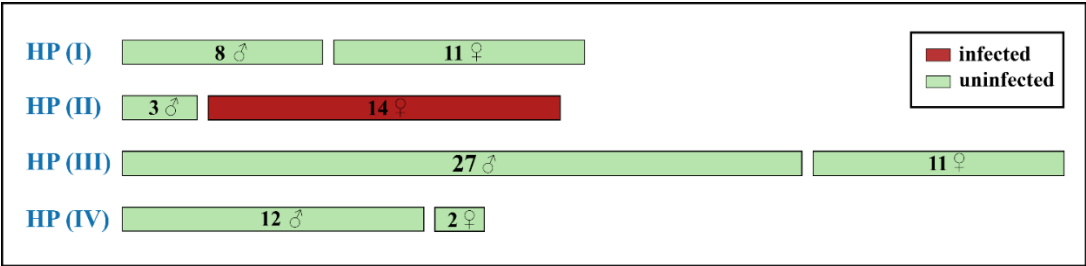


Figure 5. Infection rates of *Wolbachia* in 88 specimens of *C. mongola*/*C. tamerlana* analysed in the present study. Values and symbols in each bar indicate the number and sex of individuals bearing *COI* haplotypes of certain haplogroup. Green and red bars indicate uninfected and infected specimens, respectively.

Comparison with *Wolbachia* alleles found in other *Colias* taxa (personal unpublished data) revealed that *wTam* allele isolated from *C. mongola*/*C. tamerlana* was most similar to one of the alleles found in *C. palaeno* (Linnaeus, 1761) having with the latter identical 16S and *ftsZ* *Wolbachia* sequences and differing in one nucleotide substitution in the *wsp* gene fragment. Infected *C. mongola*/*C. tamerlana* specimens share 16S *Wolbachia* sequence with several insect taxa, belonging to orders Lepidoptera (Pieridae, Nymphalidae, Tortricidae), Orthoptera (Acrididae), Hemiptera (Issidae, Lygaeidae) and *wsp* sequence with *Phengaris nausithous* (Bergsträsser, 1779) (Lepidoptera, Lycaenidae). The closely related *Wolbachia* alleles were found in Hemiptera families Aleyrodidae, Triozidae, Delphacidae, Aphididae, Liviidae for the gene 16S; in insect orders Lepidoptera, Orthoptera, Hemiptera, Diptera, Hymenoptera and spider mites (Trombidiformes) for the gene *wsp*; in Lepidoptera, Orthoptera, Hemiptera, Diptera, Hymenoptera, Coleoptera and Trombidiformes for the gene *ftsZ* (<https://blast.ncbi.nlm.nih.gov>, accessed on 20 July 2023).

3.3.2. Phylogenetic Inferences

Bayesian analysis of the *Wolbachia* 16S gene fragment based on 43 most similar sequences with known host species mined from GenBank recovered two clades with high (clade I, BS=0.96) and low (clade II, BS=0.78) support (Figure 6). All *Wolbachia* 16S sequences isolated from infected *C. mongola*/*C. tamerlana* specimens grouped within clade II together with *Wolbachia* alleles found in three insect orders: Lepidoptera, Hemiptera and Orthoptera. The clade I included *Wolbachia* alleles found in various Lepidoptera families, Diptera, Coleoptera, Hemiptera and mites (Parasitiformes, Trombidiformes). Maximum p-distances within clade I and clade II were as high as 1.5% and 0.8%, respectively; maximum p-distances between two clades – 1.8%.

Bayesian analysis of the *Wolbachia* *wsp* gene fragment based on 74 most similar sequences with known host species mined from GenBank revealed two clades: well-supported clade I (BS=1) and weakly supported clade II (BS=0.69). The latter, in turn, subdivided into several unsupported lineages (Figure 7). Obtained in the present study *Wolbachia* *wsp* sequences from *C. mongola*/*C. tamerlana* clustered within the subclade A of the *wsp* clade II, which unites *Wolbachia* *wsp* alleles isolated from members of insect orders Lepidoptera, Orthoptera, Hemiptera, Diptera, Hymenoptera and spider mites (Trombidiformes). Maximum p-distances within *wsp* clade I and *wsp* clade II were as high as 1.8% and 5.7%, respectively; between two clades – 7.1%; within subclade A – 3%.

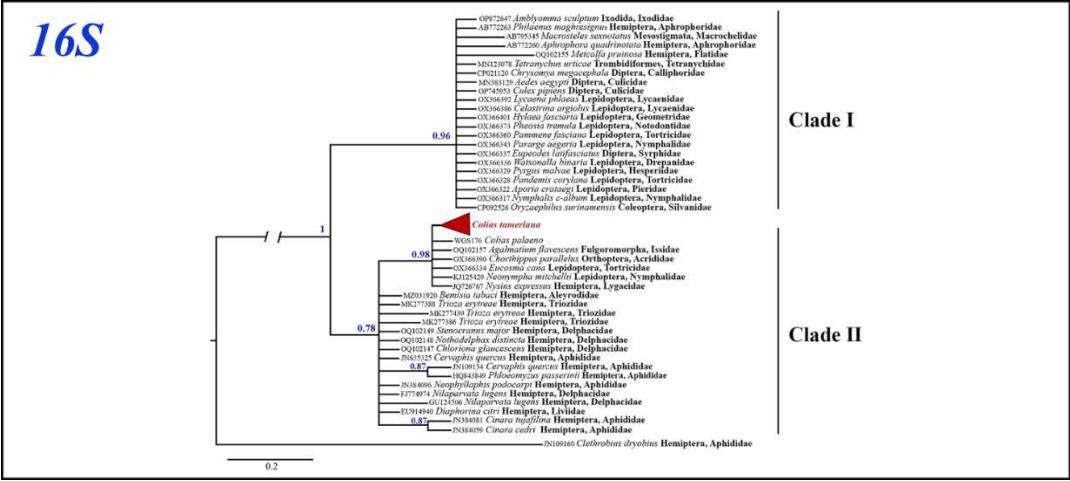


Figure 6. The Bayesian tree of 16S gene fragment inferred from 57 *Wolbachia* samples. Numbers at nodes indicate Bayesian posterior probability.

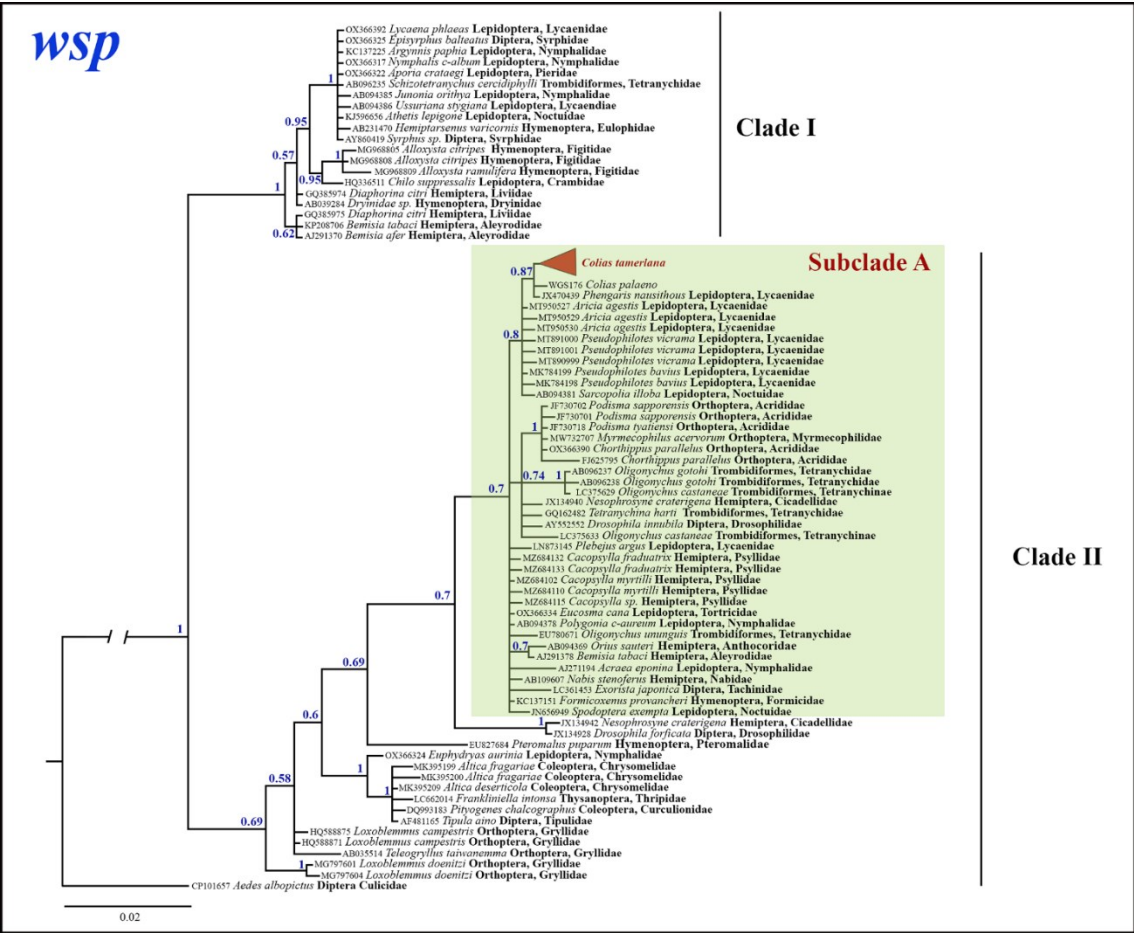


Figure 7. The Bayesian tree of *wsp* gene fragment inferred from 88 *Wolbachia* samples. Numbers at nodes indicate Bayesian posterior probability.

For the phylogenetic analysis of the *Wolbachia ftsZ* gene fragment, we used additional 71 most similar sequences with known host species obtained from GenBank. Bayesian analysis revealed several highly supported clades (Bayesian posterior probability >0.95), however deeper nodes mostly remained poorly resolved (Figure 8). *Wolbachia* samples isolated from *C. tamerlana* specimens grouped within well-supported clade (BS=0.98) together with members of insect orders Lepidoptera,

Orthoptera, Hemiptera, Diptera, Hymenoptera, Coleoptera and Trombidiformes mites. Maximum p-distances within this clade were as high as 1.77%; maximum p-distances among all analysed *ftsZ* samples reached 2.37%.

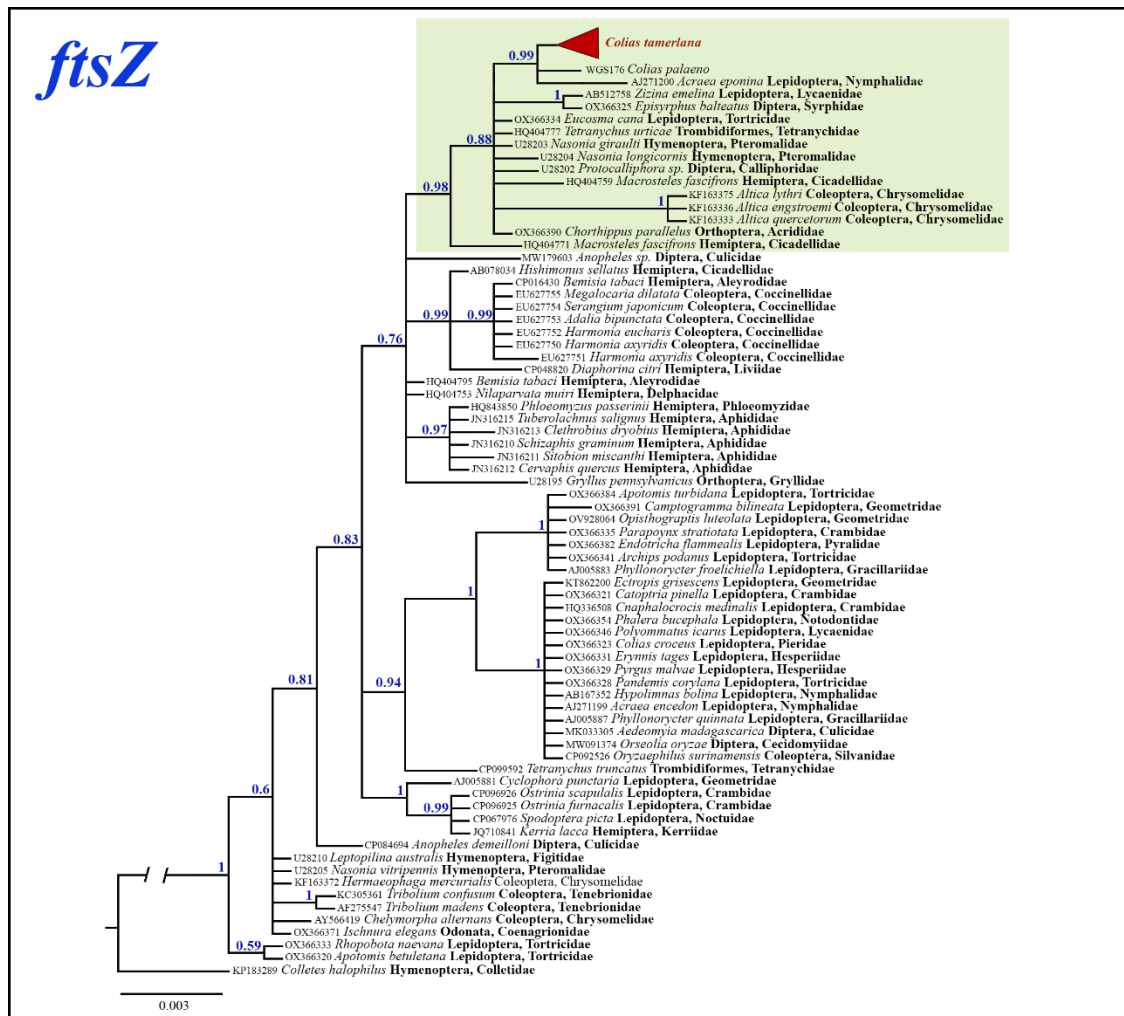


Figure 8. The Bayesian tree of *ftsZ* gene fragment inferred from 85 *Wolbachia* samples. Numbers at nodes indicate Bayesian posterior probability.

4. Discussion

4.1. Molecular Analysis and Taxonomy of *C. mongola*/*C. tamerlana*

Mitochondrial DNA is widely used in phylogenetic reconstructions, taxonomic studies and species identification and delimitation due to distinct advantages over other molecular markers: the mitochondrial genome is maternally inherited and generally does not undergo recombination; mtDNA usually evolves faster compared to nuclear DNA, potentially providing better resolution to recover low-level taxonomic relationships; mtDNA presented in cells in a high copy number, thereby allowing better and ease amplification of targeted fragment. However, numerous studies underline limited application of DNA barcoding in the context of incomplete lineage sorting, mitochondrial introgression and infection by endosymbionts, such as *Wolbachia* [77–83]. Our analysis of the mitochondrial barcodes alone revealed four differentiated clades within *C. mongola*/*C. tamerlana* with genetic distances reaching 2.89%. Such values are comparable to “standard” species-level *COI* divergence empirically estimated for Lepidoptera [84–87] and suggest relatively old separation of the recovered *COI* lineages. In fact, if we relied solely on *COI* barcodes, the possible conclusion would have been that observed lineages might represent unique cryptic species, especially taking into account comparatively young radiation of the genus *Colias* [43,44]. However, this conclusion was not

supported by subsequent analyses of the nuclear genes and phenotypic traits. Recovered mtDNA phylogenetic structure was not corroborated by nuclear data: the specimens bearing different *COI* haplotypes were randomly distributed across the nucDNA phylogenetic tree. The nuclear sequences showed no signs of significant divergence between the clades, delimited by mitochondrial barcodes, forming a single, nearly unstructured entity. Furthermore, butterflies collected from the remote geographical localities show no traces of morphological differentiation despite its phenotypic variability. The only exception is *C. tamerlana sidonia*, the geographically isolated taxon from Lake Khovsgol area (Mongolia), which differs from typical *C. mongola/C. tamerlana* in larger size, less developed dark suffusion on the fore- and hindwings and large submarginal light spots. Such prominent morphological differences even force some authors to consider *sidonia* as a subspecies of much easterly distributed taxon, *C. nastes*. However, our data unequivocally show that *sidonia* undoubtedly belongs to *C. mongola/C. tamerlana* complex sharing common and geographically widespread *COI* haplotype hb1 with the latter. It should also be noted that in the case of cryptic species we should expect a geographic isolation (in case of allopatry), or niche separation (in case of sympatry) of revealed mtDNA clusters, which we consider as putative cryptic species [88–90]. Certainly, this is not the case for *C. mongola/C. tamerlana*, where butterflies of different *COI* haplogroups were found flying together syntopically (in the same habitat) and synchronously (at the same time), in other words in complete sympatry without any niche separation.

Taxonomic status of *C. mongola* and *C. tamerlana* and relationships of this pair of taxa and allied species are also a subject of longstanding debates. Analysis of the century-old type specimens, namely the lectotype of *C. tamerlana*, the paralectotype of *C. tamerlana* and the lectotype of *C. mongola*, along with *C. tamerlana sidonia* specimens from the type locality, presumably belonging to the type series, allowed us to shed light on this very controversial issue of the *Colias* taxonomy. The type locality of *C. mongola* has been contradictory interpreted in the taxonomic literature and cannot be clearly ascertained. Thus, the limited number of specimens originated from the type series stored in the museum collections are the only reliable source of molecular data. Here, using NGS approach, we recover and analyse DNA barcodes of these old museum specimens. We confirm the conspecificity of *C. tamerlana* and *C. mongola* and show that the type specimens of these taxa share the same *COI* haplotype hb1, common and widely spread over a large geographical area from Northwestern China through South Siberian part of Russia to Western Mongolia.

4.2. *Wolbachia* Infection in *C. mongola/C. tamerlana*

Recent investigations have suggested that *Wolbachia* infection is common and widespread in Lepidoptera [91–93]; however, large-scale and comprehensive studies devoted to the incidence, patterns of *Wolbachia* infection and its impact on phylogenetic inferences of host species are still scarce [94–103]. To date, *Wolbachia* infection have been reported only for a few *Colias* species, namely *C. palaeno*, *C. hyale* (Linnaeus, 1758), *C. erate poliographus* Motschulsky, 1861 and *C. croceus* [46,93,104,105]. Here, we designated a new *Wolbachia* allele *wTam*, which had not been previously recorded in *Colias*. Surprisingly, none of the available *Wolbachia* *wsp* and *ftsZ* STs (sequence types) found in *Colias* and deposited in public databases GenBank (<https://blast.ncbi.nlm.nih.gov>, accessed on 20 July 2023) and PubMLST-*Wolbachia* (<https://pubmlst.org/organisms/wolbachia-spp>, accessed on 25 July 2023) fall within the sequences with the highest percentage identity much to *wTam*. However, our personal unpublished data suggests that alleles similar to *wTam* can be found in other *Colias* taxa.

The *Wolbachia* allele found in *C. mongola/C. tamerlana* is shared among different insect species, families and even orders. *Wolbachia* infection is mainly vertically transmitted to the progeny via maternal cytoplasm. Accordingly, one should expect the phylogeny of *Wolbachia* to be consistent with the phylogeny of their hosts. Notwithstanding, the horizontal transfer of *Wolbachia* between insect hosts have been suggested for many insect taxa [106–111], however the mechanisms of this phenomenon yet remain to be characterized. It has been suggested that *Wolbachia* can shift between distantly related hosts through host-parasitoid interactions [112–115], shared host plants [116–120], hybridisation events [98,121–124] and predator-prey associations [125].

Wolbachia 16S sequence obtained in our study is shared by two Lepidoptera species, nymphalid *Neonympha mitchellii* French, 1889 distributed in the eastern USA and tortricid moth *Eucosma cana* (Haworth, 1811), the meadow grasshopper *Chorthippus parallelus* (Zetterstedt, 1821) (Orthoptera, Acrididae), the false chinch bug *Nysius expressus* Distant, 1883 (Hemiptera, Lygaeidae) and the planthopper *Agalmatium flavescens* (Olivier, 1791) (Hemiptera, Issidae). *Wolbachia* *wsp* sequence is shared by lycaenid butterfly *Phengaris nausithous* (Lepidoptera, Lycaenidae). Our data evidenced that the same *Wolbachia* strain may occur in very distant, not closely related taxa. Thus, we confirm previous studies suggesting that horizontal transmissions events are quite common in nature [126,127]. Interestingly, the very similar *wsp* and *ftsZ* STs to *C. mongola*/*C. tamerlana* carry spider mites (Acari, Trombidiformes). Mites are known as common parasites for Lepidoptera and other insects and arthropods and have been suggested as potential vectors for *Wolbachia* transmission [128,129]. Occurrence of closely related *Wolbachia* strains in butterflies and Trombidiformes indicate that *Wolbachia* host switches in Lepidoptera might be caused by mites.

Screening for *Wolbachia* revealed infection in 14 specimens (out of 88 tested), suggesting relatively low infection rate in *C. mongola*/*C. tamerlana* (prevalence: 16%). One of the most intriguing results obtained in the present study is that we found a full congruence between sex, specific mitochondrial lineage and *Wolbachia* infection: all 14 infected specimens were females of *COI* haplogroup HP_II comprising a single haplotype hp2. To our knowledge, such sex-biased, selective and total infection of certain mitochondrial lineage have never been observed in Lepidoptera. Moreover, our personal unpublished data suggest that such patterns of selective infection can be found in other *Colias* taxa, presumably being a general characteristic of the genus. It also should be noted that surprisingly low number of publications devoted to analysis of *Wolbachia* infection in Lepidoptera consider *Wolbachia* prevalence in males and females independently [130,131], while sex-dependent impact of *Wolbachia* on its hosts is highly expected.

Interestingly, somewhat similar pattern has been observed in another pierid genus *Eurema* (Lepidoptera, Pieridae) belonging to the same subfamily Coliadinae, where sex-biased female lineages of *Wolbachia* were discovered in two Japanese species [132–134]. The *Wolbachia* allele *wFem* was found at low frequencies only in the *Eurema* females and has not been observed in the males. A causative role of *wFem* allele in feminization, a well-known manipulation effect deployed by *Wolbachia* [135], has been proposed. It has been shown that antibiotic treatment of infected larvae leads to occurrence of intersex individuals, while treatment of adult females results in all-male progeny. We cannot exclude that sex-biased *Wolbachia* infection observed in *C. mongola*/*C. tamerlana* is also a consequence of feminization, a phenomenon that has been rarely encountered within Lepidoptera [136]. However, further comprehensive studies based on the lab experiments and analysis of additional material are needed to confirm such assumptions.

5. Conclusions

Our study is the first large-scale investigation aimed at a detailed analysis of phylogeographical structure, geographical distribution and taxonomy of two enigmatic *Colias* taxa with the controversial taxonomic status, *C. mongola* and *C. tamerlana*. Our analysis cluster DNA barcodes obtained in the present study in four distinct haplogroups, however no association between nuclear genes and mitochondrial clusters, as well as between the distribution of mitochondrial haplotypes and geography have been revealed. Using NGS approach, we recover and analyse DNA barcodes of century-old *C. mongola* and *C. tamerlana* type specimens. We show that the type specimens of these taxa share the same *COI* haplotype. These results confirm the conspecificity of *C. tamerlana* and *C. mongola*, solving a longstanding question about their taxonomic status, also demonstrating that application of modern techniques is of a great importance in cases when the type locality is unknown or cannot be clearly ascertained, and old material originated from the type series is the only reliable source of molecular data. We analyse the presence and prevalence of *Wolbachia* in *C. mongola*/*C. tamerlana* and found strong correlation between sex, specific mitochondrial lineage and *Wolbachia* infection. Phylogenetic analysis placed *Wolbachia* strain of *C. mongola*/*C. tamerlana* together with members of different insect families and even orders, indicating multiple events of host shifts, thus

being consistent with the former studies, evidencing that horizontal transmission is a common mechanism of *Wolbachia* expansion. We conclude that occurrence of deep intraspecific divergences of DNA barcodes do not necessarily is a consequence of cryptic speciation but instead can be a result of *Wolbachia* infection and some other, most likely environmental factors.

6. Taxonomic conclusion

The nomenclature of *C. tamerlana* and *C. mongola* is a subject of longstanding debates [1,3,13]. Here we follow the logic of Greishuber [1], who pointed out that under the International Code of Zoological Nomenclature (Article 8.1.2.) a work is regarded as having been published only when it becomes available. The earliest confirmed date for Romanoff's book is 18.12.1897, when the copy of 9th volume have been received by the Harvard University library [1]. Consequently, the name *tamerlana* has priority over the *mongola*. Taking into account the molecular data obtained in the present study we confirm conspecificity of the taxa in question and consider *Colias nastes mongola* Alphéraky, 1897 a subjective junior synonym of *Colias tamerlana* Staudinger, 1897, syn. nov. We did not find any molecular evidences supporting subspecies status of *Colias mongola ukokana* Korb & Yakovlev, 2000 and *Colias cocandica sidonia* Weiss, 1968, despite pronounced differences in the external morphological characters of the latter. These populations do not show any signs of divergence and should be considered as the nominotypical *Colias tamerlana*.

Thus, we suggest following rearrangements here:

Colias nastes mongola Alphéraky, 1897 = *Colias tamerlana* Staudinger, 1897, syn. nov.

Colias mongola ukokana Korb & Yakovlev, 2000 = *Colias tamerlana* Staudinger, 1897, syn. nov.

Colias cocandica sidonia Weiss, 1968 = *Colias tamerlana* Staudinger, 1897, syn. nov.

Supplementary Materials: The following supporting information can be downloaded at the website of this paper posted on Preprints.org., Table S1: List of studied materials with GenBank accession numbers for sequence records. Table S2: A detailed summary of the nucleotide variability of the studied *Ca-ATPase*, *CAD*, and *H3* nuclear genes fragments among the sequenced samples of *C.mongola/C. tamerlana*.

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