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Recent Advances on SARS-CoV-2 Mpro Inhibitors: From Nirmatrelvir to Future Perspectives

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Abstract: The Main Protease (M^{pro}) plays a pivotal role in the replication of severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) and is considered a highly conserved viral target. Disruption of the catalytic activity of M^{pro} produces a detrimental effect on the course of the infection, making this target one of the most attractive for the treatment of COVID-19. The current success of the SARS-CoV-2 M^{pro} inhibitor Nirmatrelvir, the first oral drug for the treatment of severe forms of COVID-19, has further focused the attention of researchers on this important viral target, making the search for new M^{pro} inhibitors a thriving and exciting field for the development of antiviral drugs active against SARS-CoV-2 and related coronaviruses.

Keywords: COVID-19; SARS-CoV-2 Mpro; protease inhibitors; coronavirus; peptidomimetics; Paxlovid; Nirmatrelvir

1. Introduction

Since the outbreak of the COVID-19 pandemic, a considerable part of the pharmaceutical research has directed its efforts towards the discovery of new and effective antiviral drugs with the ultimate goal of preventing (or at least being prepared for) further dramatic health emergencies [1].

The following review includes the most promising scientific discoveries since 2021 concerning the synthesis and biological evaluation of SARS-CoV-2 M^{pro} inhibitors and aims to update the state-of-the-art about COVID-19 treatments with a future perspective to other related coronaviral diseases. A detailed discussion of the structure and role of SARS-CoV-2 M^{pro} during viral replication of SARS-CoV-2 has already been addressed in our previous review [2]. Then, adequate attention will be paid to the newly discovered molecules, starting from **Paxlovid®**, the first oral antiviral drug developed by Pfizer for the treatment of COVID-19 which consists of a combination of **Nirmatrelvir** (SARS-CoV-2 M^{pro} inhibitor) and **Ritonavir** (HIV-1 protease inhibitor) [3].

2. SARS-CoV-2 Mpro as a Drug Target: Structural Features and Proteolytic Mechanism

SARS-CoV-2 M^{pro} came out as a topical antiviral drug target due to several unique characteristics. In general, the combined action of SARS-CoV-2 M^{pro} and SARS-CoV-2 PL^{pro} constitutes a key aspect for the virus replication. The inhibition of their activity leads to the inability of the virus to replicate and infect other cells or subsequest hosts [4]. Indeed, M^{pro} and PL^{pro} were considered plausible drug targets since the evolving of the SARS and MERS pandemics due to their active role in post-translational processing of viral polyproteins [5]. By comparing the primary sequences of M^{pro} and PL^{pro} of SARS-CoV, MERS-CoV and SARS-CoV-2, the amino acid sequences were observed to be highly conserved among these three viruses [4]. This means that, despite the high mutation rate of coronaviruses, M^{pro} and PL^{pro} tend to be conserved as mutations of these

proteases could be fatal to the virus [6]. Therefore, the risk of developing drug-resistance is reduced for M^{pro} and PL^{pro} inihibitors [7]. PL^{pro} is endowed with a high degree of structural similarity with various human cysteine proteases. As a consequence, the development of PL^{pro} inhibitors may lead to cross-reactivity and consequent drawbacks [2]. M^{pro} instead has no homologous human proteases [8]. Thus, its proteolytic activity could be specifically inhibited with no significant impact on host cells [9]. In this perspective, M^{pro} has been considered a more attractive target as compared to PL^{pro} for anti-SARS-CoV-2 drug development. Nowadays, more than 400 crystal structures of SARS-CoV-2 M^{pro} appear in the Protein Data Bank (PDB) database [10]. The native structure of SARS-CoV-2 M^{pro} is shown in Figure 1A (PDB: 7ALH) [11]. The active form of M^{pro} is given by 2 identical monomers (A and B in Figure 1B) of approximately 34.21 kDa, oriented with a right angle to each other to create the active homodimer [10]. Monomers are formed by three domains: domains I and II are six-stranded antiparallel β-barrels which host the active site, whereas domain III contains an antiparallel globular cluster formed by five α-helices that promote the dimerization process [12]. The crystal structure of SARS-CoV-2 M^{pro} monomer is shown in Figure 1B (PDB: 1P9S) [13].

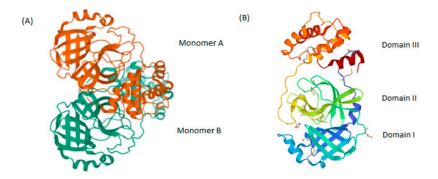


Figure 1. (A) Crystal structure of SARS-CoV-2 M^{pro} (PDB: 7ALH) enlightening the two monomers (monomer A is depicted in orange, monomer B in cyan); (B) Crystal structure of SARS-CoV-2 M^{pro} monomer (PDB: 1P9S) enlightening the three domains (domain I is depicted in blue, domain II in green, domain III in orange).

The active site of SARS-CoV-2 M^{pro} is located at the interface of domain I and II of each monomer (Figure 2). However, in the dimeric form, only one catalytic site is active [10]. This active site can be divided into six sub-pockets, named S1′, S1, S2, S3, S4, and S5 to which correspond six substrate-bindings. S1, S2 and S4 pockets are well-built in the protein cavity, while on the top of the protein surface are found the S1′, S3 and S5 subsites, which show an undefined shape [14]. SARS-CoV-2 M^{pro} is a cysteine protease that performs a hydrolytic cleveage mechanism due to a catalytic dyad formed by Cys145 and His41 residues. Normally, serine- or cysteine-proteases exploit a standard catalytic tryad, formed by Cys/Ser, His and Asp; in this case, the absence of the third element is counterbalanced by the presence of a water molecule H-bonded to His41 [2]. Another water molecule stabilizes the oxoanionic hole, establishing H-bonds with Phe140, His163, and Glu166. M^{pro} is responsible for the cleveage of 11 sites of pro-protein pp1ab, occurring when a Gln residue is found in P1 position [15]. In particular, the enzyme can recognize the sequence of Leu-Gln↓Ser-Ala-Gly, where ↓ indicates the amide bond position cleaved by M^{pro} [14]. This amino acid sequence is not recognized by any human proteases, reason why it is possible to design selctive M^{pro} inhibitors. [2].

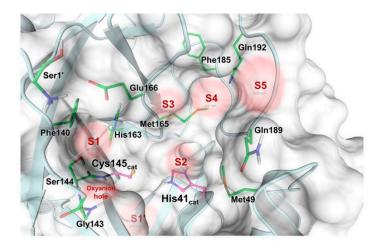


Figure 2. Active site of the SARS-CoV-2 M^{pro} with position of the catalytic dyad, oxyanion hole and subsites S1'-S5.

3. SARS-CoV-2 Mpro Inhibitors: An Overview

Having ascertained the pivotal role of M^{pro} during the replicative cycle of coronaviruses and its relevance as a therapeutic target, the efforts in the development of selective inhibitors have been enormous [16]. Firstly, inhibitors of the active site (competitive inhibitors) were found more successful in comparison to the small group of allosteric ones (non-competitive inhibitors). Indeed, many attempts have been made to develop competitive inhibitors which may interact with the residues present in the active site in a covalent or non-covalent fashion [17,18]. Beyond the reproposition of drugs (which was reasonably the most immediate countermeausure to stop the COVID-19 pandemic), the development of new SARS-CoV-2 Mpro competitive inhibitors has been mainly focused on peptidomimetics as compared to non-peptidyl compounds [2]. Peptidomimetics have the capacity of mimicking natural peptidic substrates and displaying at the same time superior pharmacokinetics characteristics [19]. During the first SARS outbreak, the peptidomimetic approach was widely used to develop new SARS-CoV Mpro inhibitors. Thus, the same approach has been proposed to counteract the new COVID-19 pandemic [20]. Despite the high similarity between the two M^{pros}, the accessibility of the active site can be quite different. Therefore, also the development of non-peptidyl small molecules turned out to be a viable strategy to develop novel SARS-CoV-2 Mpro inhibitors [21]. In general, both peptidomimetics and non-peptidyl compounds can act as covalent inhibitors due to the presence of an (C-terminal) electrophilic warhead able to bind, temporarily or permanently, the thiol group of the catalytic Cys145, leading to the inactivation of the enzyme.

On the basis of the (C-terminal) electrophilic warhead, it is possible to classify the SARS-CoV-2 M^{pro} inhibitors in different groups:

- Nitriles
- Aldehydes
- Ketones
- α-Haloacetamides
- α-Ketoamides
- Michael acceptors
- Esters and thioesters
- Selenium/sulfur-based derivatives

Besides competitive inhibitors, research also provides allosteric or non-competitive inhibitors of SARS-CoV-2 M^{pro}. In fact, six allosteric sites have been found on SARS-CoV-2 M^{pro}, which functions can be reassumed in assisting the catalytic activity through substrate processing and dimerization phenomena [22]. The latter are fundamental for the activity of the enzyme; thus, allosteric inhibitors that target sites involved in the dimerization process seem to be very interesting for the inactivation of SARS-CoV-2 M^{pro}. Three of these sites are located at the dimerization site (allosteric site 1), while

the other three sites are located between domains II and III (allosteric site 2) to confer conformational freedom (Figure 3) [22].

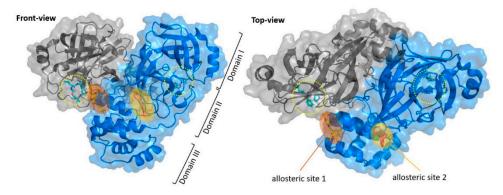


Figure 3. Schematic representation of the allosteric sites of M^{pro}. The allosteric sites are located near the dimerization site (allosteric site 1) or between domains II and III (allosteric site 2). The active site in each monomer is indicated with a yellow circle.

A complete list of the inhibitors discussed in this Review is reported in Table S1.

4. Nitriles and Nirmatrelvir

The nitrile warhead has a moderate electrophilic character. The electron-poor carbon of the nitrile group is able to undergo nucleophilic attack from the thiol group of the SARS-CoV-2 Mpro Cys145 residue to afford a reversible covalent thioimidate adduct [16]. In 2013, Chuck et al. demonstrated that nitrile-based peptidomimetics have broad-spectrum inhibition towards human coronaviruses Mpro [23]. This function has been taken into consideration by Pfizer's scientists to develop the first oral drug for the treatment of COVID-19) [24]. This drug contains a SARS-CoV-2 M^{pro} bearing a nitrile warhead in P1' called **Nirmatrelvir** (compound 1 in Figure 4). Beyond the P1' warhead, Nirmatrelvir embodies a P1 γ-lactam moiety that resembles the Gln residue of the native substrate and acts as a recognition motif for the S1 pocket. The introduction of the dimethylcyclopropyl proline moiety at the P2 position was made to achieve a deep fit within the S2 subsite. The N-terminal recognition unit is composed of a trifluoroacetamide group which provides interaction with the S3/S4 subpocket and gives the compound better pharmacokinetic properties. The discovery and development of Nirmatrelvir started from two hit compounds: PF-00835231 (2 in Figure 4), an α -hydroxy ketone-based peptidomimetic, and the corresponding phosphate prodrug PF-07304814 (3 in Figure 4), developed by Pfizer Inc. against SARS-CoV during the 2002 SARS outbreak [24]. Both compounds were selected as SARS-CoV-2 Mpro inhibitors for in vitro and in vivo biological evaluation, showing promising results as intravenously administered drug candidates. In particular, **PF-00835231** showed high potency ($K_i = 4 \text{ nM}$) and good antiviral activity (EC₅₀ = 231 nM) [25].

Figure 4. Chemical structure and biological activity of Pfizer inhibitors Nirmatrelvir, 2 and 3.

4

Owen et al. designed and synthesized a series of PF-00835231 analogues in order to improve its pharmacokinetic properties and obtain compounds suitable for oral administration [25]. The rationale of the design was to decrease the number of H-bond donors (HBD) without affecting the drug-target interaction. The first modification involved the α -hydroxy ketone warhead, replaced with two new electrophilic moieties, namely a benzothiazole-7-yl ketone and a nitrile group. The nitrile derivate (4 in Figure 5) showed higher oral absorption in rat models but lower enzymatic inhibitory and cell-based antiviral activity than 2 (K_i = 27.7 nM; EC₅₀ = 1.4 μM) [25]. Another HBD that was not critical for the drug-target interaction was the leucine residue at P2, replaced with a cyclically modified proline residue (6,6-dimethyl-3-azabicyclo[3.1.0]hexane) able to fit inside the S2 pocket. This modification, combined with the benzothiazole-7-yl ketone warhead in P1' position, led to compound 5 (Figure 5). In this case, the cell permeability increased, while enzimatic inhibitory and cell-based antiviral activity decreased as compared to compound 4 (K_i = 230 nM; EC₅₀ = 5.6 μM) probably due to the loss of a key H-bond with Gln189 in the S2 subsite. To regain potency, further investigations led to the replacement of the indole ring in P3 to reach a better fit with the S3 subsite. Two branched acyclic moieties were proposed: a methanesulfonamide and a trifluoroacetamide functionality (compounds 6 and 7, respectively; Figure 5). Compound 6 and 7 showed comparable K_i values (12.1 nM vs 7.93 nM) but different antiviral activity (6: EC₅₀ = 909 nM; 7: EC₅₀ = 85.3 nM). Moreover, 7 showed much higher oral absorption than 6 in rat and monkey in vivo models. The combination of the nitrile warhead in P1' with the trifluoroacetamide moiety in P3 eventually led to the identification of **PF-07321332**, *i.e.* **Nirmatrelvir** (1 in Figure 4).

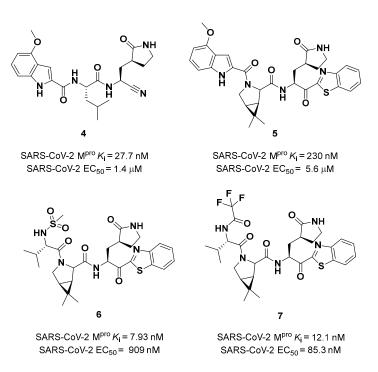


Figure 5. Chemical structure and biological activity of compounds derived from 2.

The co-crystal structure of **Nirmatrelvir** with SARS-CoV-2 M^{pro} clarified its mechanism of action. The nitrile warhead in P1' forms a reversible covalent bond with Cys145 at S1', with the formation of the thioimidate adduct that establishes H-bond with the Gly143 residue. The γ -lactam mojety interacts with His163 and Glu166 in the S1 pocket, while the aliphatic dimethyl-bicycloproline fits inside the S2 pocket via the formation of several van der Waals interactions with the surrounding apolar residues. Moreover, the carbonyl oxygen of the trifluoromethyl acetamide acts as an H-bond acceptor toward the Gln189 residue, while the trifluoromethyl group establishes interactions with the S4 pocket (Figure 6) [16].

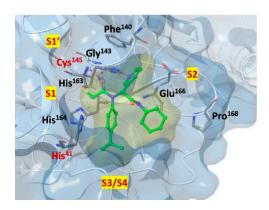


Figure 6. Crystal structure of the complex of SARS-CoV-2 Mpro/Nirmatrelvir (PDB: 7MLF).

The nitrile moiety was selected over the benzothiazole-7-yl ketone warhead due to several features: i) superior water solubility allowing for high concentration solutions of a drug candidate in pre-clinical trials; ii) reduced tendency of epimerization of the near P1 stereocenter; iii) easier synthesis for a scale up process [24]. **Nirmatrelvir** showed superior enzimatic inhibitory and antiviral activity ($K_i = 3.11$ nM; $EC_{50} = 74.5$ nM) as compared to the parent compounds (Figure 4). The introduction of a structure with less HBDs and a trifluoroacetamide moiety assured good gut permeability, oral bioavailability and proper drug clearance in $in\ vivo$ rat models. $In\ vivo$ evaluations in monkey models showed a decrease in oral bioavailability and gut permeability linked to the first pass metabolism [25]. To decrease the first pass metabolism, **Nirmatrelvir** is co-administered with the anti-HIV-1 drug **Ritonavir**, which is also an inhibitor of CYP450. Remarkable results in preclinical and clinical studies led this new combination to be authorized by the Food and Drug Administration (FDA) [26], European Medicine Agency (EMA) [27] and UK Medicines and Healthcare products Regulatory Agency (MHRA) as the only orally administered drug to treat COVID-19 [28] under the commercial name of **Paxlovid**®.

The success of the nitrile warhead on **Nirmatrelvir** lead Pfizer scientists to investigate other nitrile-based compounds as inhibitors of SARS-CoV-2 M^{pro}, resumed in a new patent published on 16th December 2021, identifying compound **8** (Figure 7) as a new SARS-CoV-2 M^{pro} inhibitor (K_i = 4 nM; EC₅₀ = 19 nM) [29]. Bai *et al.* explored other nitrile analogs, investigating the 4-methoxyindole mojety at P3 [30]. Among all products, compound **9** (Figure 7), having a 6-chloro-4-methyloxyindole function at P3, showed the best results in terms of *in vitro* enzymatic inhibitory and antiviral activity (IC₅₀ = 9 nM; EC₅₀ = 2.2 μ M) [22].

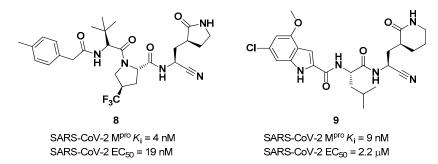


Figure 7. Chemical structure and biological activity of compounds 8 and 9.

Starting from **Nirmatrelvir**, Zhu *et al.* developed a series of novel SARS-CoV-2 M^{pro} inhibitors by optimizing the P2 and P4 positions of the peptide framework [31]. Within this series, compounds **10** and **11** (Figure 8) exhibited optimal inhibitory activity toward SARS-CoV-2 M^{pro} with IC₅₀ values of 18 nM and 22 nM, respectively. Compound **10** derived from an optimization study of the P2 site in which the dimethyl-bicycloproline of **Nirmatrelvir** was replaced with a cyclopentenyl proline. Compound **11** derived from an optimization attempt of the P4 site with the aim of improving the metabolic stability. In particular, the trifluoromethyl group of **Nirmatrelvir** was replaced with a

trifluoromethanesulfonyl group. Compounds 10 and 11 exhibited higher antiviral activity than Nirmatrelvir against SARS-CoV-2 infected VeroE6 cells, with EC50 values of 313 nM and 170 nM, respectively. They also exhibited better metabolic stability than Nirmatrelvir and similar PK properties [31].

Figure 8. Chemical structure of compounds **10** and **11** developed by Zhu *et al*. The part marked with dotted circle highlights the modifications with respect to **Nirmatrelvir**.

Inspired by the structures of hepatitis C virus NS3/4A serine protease inhibitors **Narlaprevir** and **Boceprevir**, and SARS-CoV M^{pro} inhibitors such as **GC-376** (Figure 9), Kneller *et al.* developed three SARS-CoV-2 M^{pro} inhibitors bearing a nitrile warhead [32]. Compound **12** and **13** (the two most active derivatives), called **BBH-2** and **NBH-2**, respectively (Figure 9), were designed by replacing the *C*-terminal di-ketoamide group of **Boceprevir** and **Narlaprevir**, respectively, with a nitrile warhead, whereas the hydrophobic P1 groups present in the serine protease inhibitors were replaced in both derivatives with a γ -lactam ring as per **GC-376**.

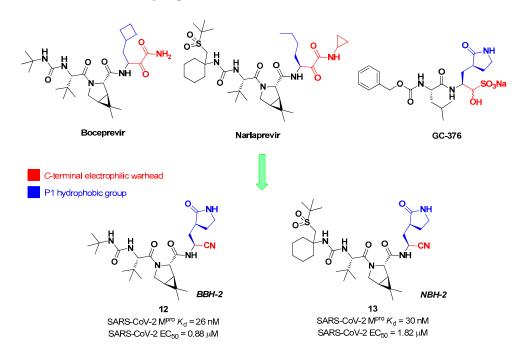


Figure 9. Chemical structure and biological activity of **Boceprevir**, **Narlaprevir**, **GC-376**, **12** and **13**. The colors highlight the chemical features the authors work on for the design of new compounds.

These compounds have been shown to have a high binding affinity toward M^{pro} , comparable to that of **Nirmatrelvir** (K_d values of 26 nM and 30 nM for **12** and **13**, respectively, and 7 nM for **Nirmatrelvir**). Furthermore, the antiviral activity of these inhibitors were measured in SARS-CoV-2

Mpro infected VeroE6 TMPRSS cells in presence of a P-glycoprotein inhibitor (CP-100346), exhibiting EC50 values of 0.88 and 1.82 μ M for **12** and **13**, respectively [32]. No cytotoxicity was detected at 10 μ M. The X-ray crystal structures of SARS-CoV-2 Mpro in complex with **12** and **13** showed that these two inhibitors bind in the same way the active site, forming a covalent bond between Cys145 and the nitrile warhead, and the newly formed thioimidate group is involved in an H-bond with the Cys145 backbone. The γ -lactam ring at P1 forms H-bonds with His163 and Glu166 main chains and an H-bond with His163 side chain. The other interactions are hydrophobic or involve H-bond with Glu166 [32].

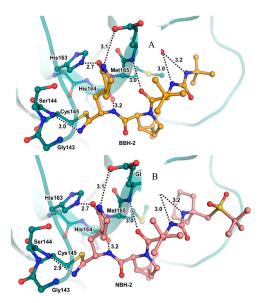


Figure 10. X-ray crystal structures of SARS-CoV-2 M^{pro} in complex with **12** (A)(PDB: 7TEH) and **13** (B)(PDB:7TFR).

Brewitz *et al.* developed different **Nirmatrelvir** derivatives by accomplishing the isoelectronic replacement nitrile/alkyne group [33]. The most promising compounds of this series turned out to be the alkyne derivate **14** and the CF₃-capped alkyne **15** (Figure 11). They displayed noteworthy SARS-CoV-2 M^{pro} inhibitory and antiviral activity, high selectivity (no activity against SARS-CoV-2 PL^{pro}) and low cytotoxicity profile. MS analyses and crystallographic studies demonstrated that, unlike nitrile derivates which inhibit M^{pro} in a reversable manner, these alkyne derivatives inhibit M^{pro} by apparent irreversible covalent mechanism forming an internal vinyl thioether adduct with the Cys145 residue [33].

Figure 11. Chemical structure and biological activity of compounds **14** and **15**. The part marked with dotted circle highlights the isoelectronic replacement of the nitrile group.

5. Aldehydes

The aldehyde functional group is considered the most widely used electrophilic warhead among covalent SARS-CoV-2 M^{pro} inhibitors, due to the susceptibility of the electron poor carbonyl carbon

to undergo nucleophilic addition by the thiol group of the Cys145 residue, with the formation of a reversible hemi-thioacetal adduct [2]. The latter highly resembles the tetrahedral intermediate of the hydrolisis of the endogenous substrate, ensuring a longer residence time and an enhanced I-E complex stability due to several H-bonds between the adduct and residues of the S1′ pocket. Furthermore, the hemithioacetal adduct is able to stabilize the ligand-target interaction acting as a H-bond donor toward the Cys145 backbone. This critical H-bond is normally provided in the natural substrate by a carbonyl oxygen of the amide group next to a Gln residue [34]. Thus, this critical interaction can be provided only by carbonyl- and ketoamide-based inhibitors, and this could explain their higher efficacy in comparison to other electrophilic warheads. The first peptidomimetics bearing an aldehyde warhead targeting SARS-CoV-2 M^{pro} were compounds **16** and **17** (Figure 16), designed and synthesized by Dai *et al.* [35]. The work was mainly devoted to explore the enzyme S2 site. Compound **16** has a cyclohexyl ring at P2, while in compound **17** we find a 3-fluorophenyl ring. Besides the aldehyde warhead at P1′, other common features are the 2-indole moiety at P3 and the (S)-γ-lactam group at P1. Also compound **18** (Figure 12) was synthetized later on by Dai *et al.*, and it turned out to be a broad-spectrum M^{pro} inhibitor of enterovirus and SARS-CoV-2 [36].

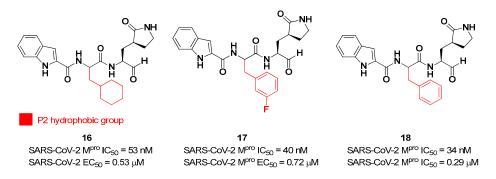


Figure 12. Chemical structure and biological activity of the first aldehyde derivatives (**16-18**) as inhibitors of SARS-CoV-2 M^{pro} developed by Dai *et al*.

The prementioned aldehyde derivatives exhibited a high enzymatic inhibitory and antiviral activity (16: IC₅₀ = 53 nM and EC₅₀ = 0.53 μ M; 17: IC₅₀ = 40 nM and EC₅₀ = 0.72 μ M; 18: IC₅₀ = 34 nM and EC₅₀ = 0.29 μ M) [36]. From the X-ray analysis of the structures of SARS-CoV-2 M^{pro} in complex with compounds 17 and 18, it was possible to observe the main interactions involved in the mechanism of action: a C-S covalent bond was detected between the carbonyl carbon of the warhead and the thiol group of the Cys145 of S1' pocket; this adduct is stabilized by an additional H-bond between the thiohemiacetal hydroxyl group and the Cys145 backbone. The (S)- γ - lactam moiety at P2 is deeply inserted into the S2 subsite forming several H-bonds with key residues of this pocket, while the indole group at P3 interacts via a H-bond with the Glu166 residue located in the surface of S3/S4 sites. The binding modes of compounds 16 and 17 are quite similar, except for the interaction with S3, involving two different P3 moieties. In compound 17, the 3-fluorophenyl group undergoes a downward rotation in the S2 pocket, unlike the cyclohexyl substituent in 16, due to an additional H-bond involving the fluorine atom and the Gln189 redisue and several hydrophobic interactions of the aromatic ring with the surrounding residues. Compound 16 was chosen as a potential drug candidate due to its better pharmacokinetic properties [35].

Other examples of aldehyde-based SARS-CoV-2 M^{pro} inhibitors are **GC373** (*i.e.* compound **19** in Figure 13A) and its bisulfite prodrug **GC376** (*i.e.* compound **20** in Figure 13A) [37]. These compounds were initially used to treat feline infectious peritonitis, caused by feline coronaviruses FCoV, and then they were repurposed as SARS-CoV-2 M^{pro} inhibitors. In both compounds there is a benzyl moiety at P3, a leucine residue at P2 and a (*S*)- γ -lactam group at P1. **19** and **20** inhibited the activity of SARS-CoV-2 M^{pro} with an IC50 value of 0.40 and 0.19 μ M, respectively. The X-ray crystal structure of SARS-CoV-2 M^{pro} in complex with both compounds confirmed the ability of the prodrug **20** to provide the aldehyde function since they showed an identical binding mode: the aldehyde warhead reacted covalently with Cys145 thiol group to give a hemithioacetal adduct which is stabilized by several H-

bonds inside the oxyanionic hole with the residues of Gly143, Ser144 and Cys145 (Figure 13B) [37]. The P1 γ - lactam moiety forms a H-bond with His163 and Glu166 and interacts with the main chain of Phe140, while the P2 leucine is inserted into the P2 pocket establishing hydrophobic interactions with Met149, His41 and Met49. The benzyl group at P3 interacts with the S3/S4 superficial sites by means of hydrophobic interactions. Both compounds exhibited high *in vitro* antiviral activity observed in VeroE6 cell infected with SARS-CoV-2 (19: EC50 = 1.5 μ M; 20: EC50 = 0.9 μ M) and low cell toxicity (CC50 > 100 μ M for both compounds). Nevertheless, the use of the bisulfite prodrug 20 has been preferred due to better outcomes in terms of bioavailability.

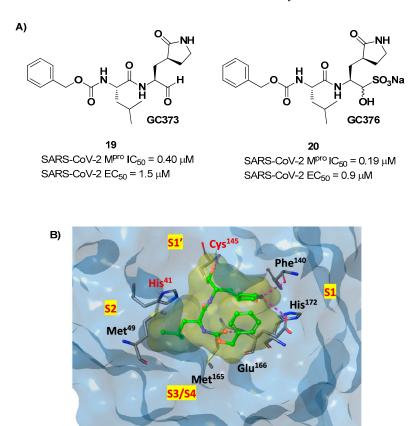


Figure 13. A) Chemical structure and biological activity of compounds **19** and **20**; B) X-ray structure of SARS-CoV-2 M^{pro} in complex with compound **19** (PDB: 6WTJ).

Structural modifications of **20** were conducted in order to improve the *in vitro* antiviral activity against SARS-CoV-2 [37]. Specifically, its P2 position was redesigned by substituting the leucine side chain with a constrained cyclopropylmethyl ring, while its benzyl group at P3 was decorated with a *meta*-F substituent (*i.e.* compound **21** in Figure 14A) or elongated with a supplementary methylene group that connects a 3-chlorophenyl ring to the carbamate linkage (*i.e.* compound **22** in Figure 14A) [38]. Both compounds displayed improved *in vitro* inhibitory activity against M^{pro} as well as antiviral activity in comparison to the lead (**21**: $IC_{50} = 0.07$ and $EC_{50} = 0.57$ μ M; **22**: $IC_{50} = 0.08$ μ M and $EC_{50} = 0.7$ μ M)[38]. The X-ray structure of the complex between M^{pro} and **21** indicated that the P2 cyclopropyl fragment is able to fit more deeply within the S2 subsite, while the substitution at P3 with a halogenated phenyl ring allowed the inhibitor to fit deeper the S3/S4 pocket instead of remaining on the surface of the protease [38].

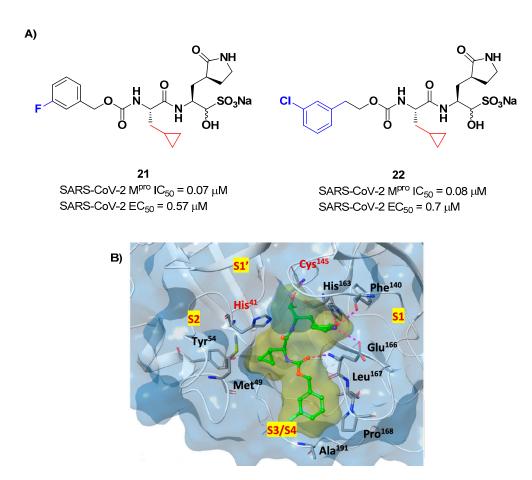


Figure 14. A) Chemical structure and biological activity of compounds **21** and **22**. Those parts marked with colors indicate modifications with respect the lead structure **20**; B) X-ray structure of SARS-CoV-2 M^{pro} in complex with compound **21** (PDB: 7LCO).

Other analogs of **20** with improved *in vitro* enzymatic inhibitory and antiviral activity are represented by compound **23** and **24** (Figure 15) [39,40]. In compound **23**, also called **UAWJ247**, structural variation in comparison to **20** involved the P2 position, where a phenyl ring replaced the isobutyl moiety. This compound exhibited an enzymatic inhibitory activity comparable to **20** (IC₅₀ = 45 nM) and a moderate antiviral activity. Compound **24**, also called **NK01-63** or **Coronastat**, contains a 3-trifluoromethylbenzyl group as a replacement of the benzyl group at P3 present in **20**. Compounds **24** showed a potent inhibitory activity against SARS-CoV-2 M^{pro} (IC₅₀ = 16 nM) and an excellent antiviral activity (EC₅₀ = 6 nM in Huh7^{ACE2} infected cell with SARS-CoV-2). The improvement in potency and antiviral activity of **24** can be explained by the presence of two additional H-bonds provided by the trifluoromethyl substituent at P3 with the Asn142 residue found in S3/S4 pocket, as observed in the X-ray structure [39,40].

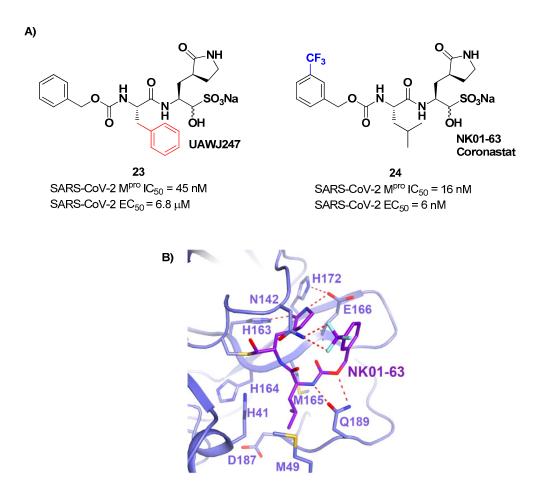


Figure 15. A) Chemical structure and biological activity of compounds **23** and **24**. Those parts marked with colors indicate modifications with respect the lead structure **20**; B) X-ray structure of SARS-CoV-2 M^{pro} in complex with compound **24** (PDB: 7TIZ).

Another interesting approach was undertaken to enhance the interaction inside the S2 pockect. It involved the introduction of a bicyclic proline derivative, a motif present in the structure of two potent inhibitors of HCV protease, *i.e.* **Boceprevir** (Figure 9) and **Telaprevir** (Figure 16). In particular, the proline residue was fused with three- (**Boceprevir**) or five-membered (**Telaprevir**) carbocyclic rings to form constrained bicyclic synthons [41]. The most interesting compounds are depicted in Figure 16. The aldehyde derivative **25** (also **MI-09** in Figure 16) contains the same P2 fragment extrapolated from **Boceprevir** and connected to a *para-*OMe benzyl carbamate functional group, while the aldehyde derivatives **26** and **27** (also **MI-23** and **MI-30**, respectively, in Figure 16) are endowed with the P2 fragment of **Telaprevir** and variously halogenated benzyl carbamate moieties at P3 [41].

Figure 16. Chemical structure of Telaprevir and chemical structure and biological activity of compounds **25-27**.

25-27 showed inhibitory activity against SARS-CoV-2 M^{pro} in the nanomolar range. The X-ray analysis of the complex **26**/M^{pro} showed that the bicyclic proline is in its *trans-exo* conformation, providing a deep fit within the S2 pocket (Figure 17) [41].

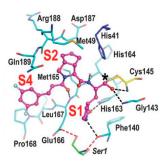


Figure 17. X-ray structure of SARS-CoV-2 Mpro in complex with compound 26 (PDB: 7D3I).

Another example of **GC376** derivative emerged from a study based on activity-guide optimization [42]. Compound **28** (Figure 18) is a novel dipeptidyl structure, bearing a constrained bicyclic ring in P3 position. This compound exhibited an inhibitory activity against SARS-CoV-2 $M^{\rm pro}$ in the micromolar range (IC50 = 0.18 μM) and antiviral activity in the sub-micromolar range (EC50 = 0.035 μM ; VeroE6 infected cells).

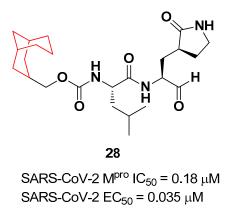


Figure 18. Chemical structure and biological activity of compound 28.

In parallel, Xia *et al.* designed and synthetized compound **29** and **30** (Figure 19), called **UAWJ9-36-1** and **UAWJ9-36-3**, respectively, by retaining the P2 bicyclic proline synthons found in **Telaprevir** and **Boceprevir** [43]. **29** was designed as a hybrid form of **GC376** and **Telaprevir**, whereas **30** was designed as a hybrid form of **GC376** and **Boceprevir**.

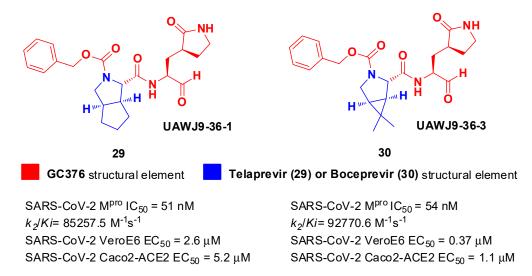


Figure 19. Chemical structure and biological activity of compounds 29 and 30.

Compounds **29** and **30** underwent enzymatic assays against all seven human coronavirus M^{pros}, providing a similar inhibition profile against SARS-CoV-2 M^{pro} (and overall for all M^{pros}) in comparison with **GC-376** (**29**: IC₅₀ = 51 nM with a k_2/K_i = 85257.5 M⁻¹s⁻¹; **30**: IC₅₀ = 54 nM with a k_2/K_i = 92770.6 M⁻¹s⁻¹). The antiviral activity was assessed against two different cell lines (VeroE6 and Caco2-ACE2) infected with SARS-CoV-2, hCoV-OC43, hCoV-229E, and hCoV-NL63. **30** showed an improved antiviral activity against SARS-CoV-2 as compared to **GC-376** (EC₅₀ = 0.37 μ M on VeroE6 and EC₅₀ = 1.1 μ M on Caco2-ACE2). The X-ray structures of SARS-CoV-2 M^{pro} in complex with both compounds confirmed the capacity of the bicyclic prolines to fit within the S2 pocket (Figure 20) [43].

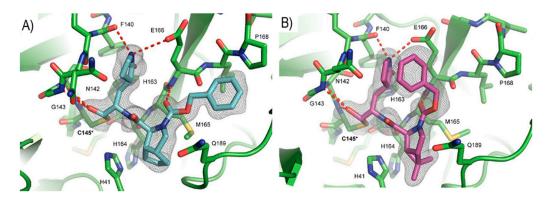


Figure 20. A) X-ray structure of SARS-CoV-2M^{pro} in complex with compound **29** (PDB: 7LIH); B) X-ray structure of SARS-CoV-2M^{pro} in complex with compound **30** (PDB: 7LIY).

Other **Boceprevir** analogs with aldehyde warhead at P1' came from the work of Alugubelli *et al.* [44]. Among the 19 synthesized compounds, the most promising derivatives were compound **31**, **32** and **33** (Figure 21), called **MPI43**, **MPI44** and **MPI46**, respectively, displaying high potency *in vitro* and *in cellulo* assays with IC50 values in the range 45-120 nM (*in vitro*) and EC50 values in the range 0.14-0.31 µM (*in cellulo*) [44]. These compounds were also evaluated for their antiviral efficacy against VeroE6 cells infected with USA-WA1/2020, Beta and Delta strains of SARS-CoV-2. Interestingly, these three compounds bear a P4 N-terminal carbamate moiety, which seems to be critical for high cellular and antiviral potency and low cytotoxicity [44].

Figure 21. Chemical structure and biological activity of compounds 31-33.

Another aldehyde-based compound was found by Günther *et al.* [45], using a large-scale X-ray crystallography to screen a library of more than 5,000 compounds among approved drugs or drugs in clinical trials. The selected compounds were then tested for their antiviral activity against SARS-CoV-2 in VeroE6 cells. Among them, **Calpeptin** (34 in Figure 22A) showed the highest antiviral activity, with an EC50 value of 72 nM and known enzymatic inhibitory activity (IC50 = 10.7 μ M) [45,46]. This compound contains an aldehyde warhead at P1', a *n*-Leu residue at P1, a Leu residue at P2 and a Boc group at P3. The X-ray crystal structure of SARS-CoV-2 M^{pro} in complex with 34 showed the standard thiohemiacetal covalent adduct Cys145/aldehyde warhead (Figure 22B). 34 also inhibits cathepsin L, making it a good anti-COVID-19 drug candidate as a dual inhibitor [45].

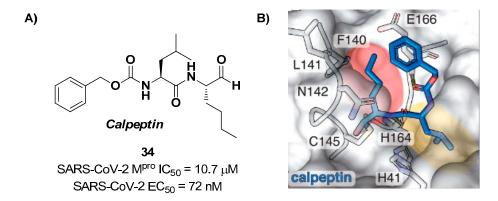


Figure 22. A) Chemical structure and biological activity of compound **34**; B) X-ray structure of SARS-CoV-2 M^{pro} in complex with **34** (PDB: 7AKU).

The aldehyde-based inhibitors discussed so far are dipeptidyl derivatives. However, there are some examples of notable tripeptidyl derivatives (Figure 23). Compound **35** and **36**, called **MPI3** and **MPI8**, respectively, are the most active tripeptides designed and synthetized by Yang *et al.* [47]. Both compounds have the aldehyde warhead at P1' and a (S)- γ -lactam group at P1. **35** is endowed with L-Leu at P2 and a Cbz-protected L-Val at P3, while in **36** we have a cyclohexyl ring at P2 and a Cbz-protected *t*-Bu Thr at P3. In the *in vitro* enzymatic inhibitory activity assay, **35** turned out to be the most active compound displaying an IC50 value of 8.5 nM. Conversely, in the *in vitro* antiviral test **35** turned out to be less active than **36** (EC50 = 2.5 μ M in VeroE6 cells) in probably due to the presence of all natural amino acids that can be hydrolyzed by cellular proteases [47]. Such a high potency could be related to the dual inhibition of SARS-Co-2 M^{pro} (IC50 = 105 nM) and cathepsin L (IC50 = 1.2 nM). This assumption led to consideration of the tripeptidyl proteasome inhibitor **MGI-132** (compound **37** in Figure 23) [48], and **calpain/cathepsin inhibitor I** and **II** (compounds **38** and **39**, respectively, in Figure 23) as potential SARS-CoV-2 M^{pro} inhibitors [39]. *In vitro* enzymatic inhibition and cell-based

antiviral tests brought about interesting results, in particular for the **calpain inhibitor II**. This could represent an interesting starting point for the development of dual inhibitors of calpain/cathepsin and SARS-CoV-2 $M^{\rm pro}$.

SARS-CoV-2 M^{PTO} IC₅₀ = 8.5 nM SARS-CoV-2 EC₅₀ = 2.5
$$\mu$$
M SARS-CoV-2 M^{PTO} IC₅₀ = 7.5 μ M SARS-CoV-2 M^{PTO} IC₅₀ = 0.97 μ M SARS-CoV-2 EC₅₀ = 2.07 μ M SARS-CoV-2 EC₅₀ = 2.07 μ M SARS-CoV-2 EC₅₀ = 2.07 μ M

Figure 23. Chemical structure and biological activity of compounds 35-39.

Ma *et al.* developed another series of tripeptidyl derivatives maintaining the aldehyde warhead at P1' and the (S)-γ-lactam group at P1, and then varying the chemical composition at both P2 and P3 positions [49]. The most interesting compounds were **40** and **41**, also called **MPI16** and **MPI17** (Figure 24). At P3 position an *O-t*-butyl-Thr residue is present in both inhibitors, while at P2 a *t*-butyl group and a cyclopropyl ring are in **40** and **41**, respectively. These compounds showed excellent enzymatic inhibitory activity (**40**: IC₅₀ = 150 nM; **41**: IC₅₀ = 60 nM), high potency in the cell-based assays (**40**: EC₅₀ = 56 nM; **41**: EC₅₀ = 97 nM) and high antiviral activity against different variants of SARS-CoV-2, suggesting that the *O-t*-butyl-Thr residue at P3 can be a key structural element for future design. From the X-ray structure of the SARS-CoV-2 M^{pro} in complex with **40** emerged that the *O-t*-butyl-Thr resudue may furnish supplementary Van der Waals interactions with Pro168 and Glu166 [49].

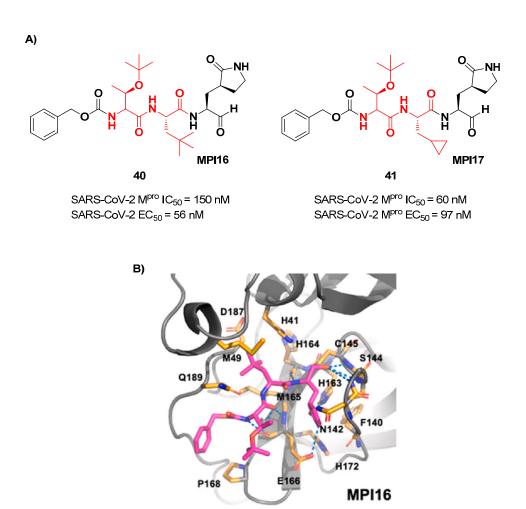


Figure 24. A) Chemical structure and biological activity of compounds **40** and **41**; B) X-ray structures of SARS-CoV M^{pro} in complex with compound **40** (PDB: 7RVQ).

6. Ketones

The ketone group is extensively expoilted as electrophilic warhead in the design of cysteine protease inhibitors including SARS-CoV-2 Mpro inhibitors. Ketones are less reactive than aldehydes; then electron withdrawing groups (especially halogens) in the α position are often needed to increase the electrophilic character of carbonyl carbon and make it more susceptible to undergo nucleophilic attack. The first example of ketone-based inhibitor was the already cited PF-00835231 (2 in Figure 4), an α-hydroxy ketone-based peptidomimetic, and its corresponding phosphate prodrug PF-07304814 (3 in Figure 4), developed by Pfizer Inc. against SARS-CoV infection during the 2002 SARS outbreak and reproposed as SARS-CoV-2 Mpro inhibitors [24]. 2 showed potent inhibitory activity toward SARS-CoV M^{pro} (*Ki* = 4 nM in SARS CoV-1 protease FRET assay) [50], as well as toward SARS-CoV-2 M^{pro} (IC₅₀ = 6.9 nM; K_i = 0.27 nM [25]). Successively, the antiviral activity of **2** was assessed on epithelial VeroE6 cells infected with SARS-CoV-2 showing promising results (EC₅₀ = 231 nM). Both 2 and 3 entered in vivo preclinical studies showing comparable enzimatic inhibitory and antiviral activity. 3 displayed superior solubility and pharmacokinetics properties as compared to 2. The X-ray structure of the covalent complex SARS-CoV-2 M^{pro}/2 evidenced the key interactions with the target: the carbonyl carbon of the α -hydroxy ketone moiety at P1' binds covalently the Cys145 thiol group to form an hemithioketal adduct that, together with the α -hydroxy group, establishes additional Hbonds with His41 and Gly143 within the S1 subpocket. The γ -lactam moiety at P1 and the Leu resdue at P2 insert into subsites S1 and S2, respectively, while the P3 indole moiety estabilishes Van der Waals interaction with residues at S3 [25].

Taking into consideration the structures of the already published SARS-CoV-2 inhibitors **PF-00835231** and **GC-376**, Bai *et al.* explored different α -acyloxymethyl ketones bearing a six-membered

lactam moiety at P1, in order to mimic the Gln residue [51]. The most promising compounds turned out to be **42**, **43** and **44** (Figure 25), which bear a 2,6-bis(trifluoro)methylbenzoate, 2,4,6-trimethylnicotinate and 4-trifluoromethyl-2,6-dimethylnicotinate fragment as C-terminal α -acyloxy moiety, respectively.

Figure 25. Chemical structure and biological activity of compounds 42, 43 and 44.

Compound **42**, which was already investigated by Kratz *et al.* as inactivator of cathepsin B [52], showed the best results in terms of enzymatic inhibitory activity against SARS-CoV-2 M^{pro} (IC₅₀ = 1.0 nM) and excellent SARS-CoV-2 replication inhibition *in vitro* (EC₅₀ = 0.16 μ M) [51]. In regard to the two nicotinic-based inhibitors, **43** exhibited a better inhibition profile (IC₅₀ = 19.0 nM; EC₅₀ = 0.30 μ M). The irreversible mechanism of action of **43** was confirmed by X-ray analysis (Figure 26): the attack of the Cys145 thiol group to the α -carbon of the ketone group and the loss of the 2,4,6-trimethylnicotinate as a leaving group. The six-membered lactam inserts into the S1 pocket establishing the same H-bonds as for the five-membered lactam of **GC-376**. Similar interactions involving other positions are in accordance with the already discussed **GC-376** [51].

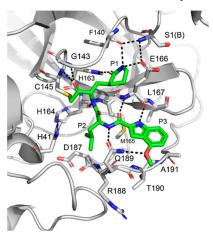


Figure 26. X-ray structure of SARS-CoV-2 M^{pro} in complex with compound 43 (PDB: 7MBI).

Other remarkable ketone-based SARS-CoV-2 M^{pro} inhibitors are represented by the **PF-00835231** benzothiazolyl analogs **5**, **6** and **7** (Figure 5), already discussed in the section of **Nirmatrelvir** [25]. Many other benzothiazolyl ketones with anti-SARS-CoV activity have been repurposed as SARS-CoV-2 M^{pro} inhibitors. One example is represented by the compound **45** (Figure 27) developed by Hattori *et al.* [53], also called **YH-53** [54]. **45** bears a P1' benzothiazolyl ketone, a P1 γ -lactame moiety, a P2 Leu residue and a P3 indole substituent. The SARS-CoV-2 M^{pro} assays reported for this compound an IC50 of 0.13 μ M and an antiviral activity of 2.6 μ M assessed in VeroE6 cells [53]. Starting from this lead compound, Higashi-Kuwata *et al.* considered the introduction of fluorine atoms to improve pharmacokinetics properties, due to the higher lipophilicity of the C-F bond compared to C-H bond [55]. Among several modifications, they explored 4-fluorinated benzothiazole ketones, obtaining compounds **46** and **47** (Figure 27), named **TKB245** and **TKB248**, respectively, as the most promising M^{pro} inhibitors. By replacing the 4-methoxy-indole ring at P3 with the trifluoroacetyl L- α -tert-butyl Gly, and the Leu residue at P2 with the 6,6-dimethyl-3-azabicyclohexane fragment, they

achieved **46** which exhibited an improved enzymatic inhibitory and antiviral activity with respect to **45** (IC₅₀ = 7 nM μ M; EC₅₀ = 0.03 μ M). To reduce the hydrolysis rate of this compound and improve its pharmacokinetic properties, the carbonyl group at P2 was replaced with a thiocarbonyl group affording compound **47**, (named **TKB248**). Pharmacokinetic studies evidenced higher T_{1/2} for **47** as compared to **46** (4.34 h vs 3.82 h), although the enzymatic inhibitory and antiviral activity were lower (IC₅₀ = 74 nM; EC₅₀ = 0.22 μ M) [55].

Figure 27. Chemical structure and biological activity of compounds 45, 46 and 47.

The *in vivo* evaluation of efficacy and pharmacokinetic parameters of both compounds were performed on human ACE2-knocked-in mice exposed to SARS-CoV-2, showing promising results. From the mass spectrometric analysis, it was found that both compounds promote dimerization of M^{pro}, which is bound to these inhibitors, preventing the entrance of the natural substrate. To elucidate the inhibition mechanism, the X-ray structures of **46** and **47** in complex with SAR-CoV-2 M^{pro} were obtained, showing that both compounds have identical binding mode with the active site [55]. As an example, Figure 28 shows the co-crystal structure of **46** with SAR-CoV-2 M^{pro}: the carbonyl carbon at P1' forms a covalent thiohemiacetal adduct with Cys145 and the thiohemiacetal-OH establish H-bonds with Gly143 and Ser144, while the 4-fluorobenzothiazole ring fills the S1' subsite with the 4-fluorine atom that points out of the subpocket. At the P1 position the γ -lactame ring establishes H-bonds with His-163, Glu166 and Phe140, while the centered amide group forms another H-bond with His164. The dimethyl-bicyclo[3.1.0]-proline moiety fits into the S2 pocket, while the trifluoromethylacetamide group at P3 forms halogen interactions with residues of the S3 pocket [55].

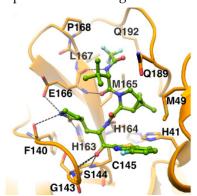


Figure 28. X-ray structure of SARS-CoV-2 M^{pro} in complex with 46 (PBD: 8DOX).

Yang *et al.* reported novel benzothiazolyl-based peptidomimetics acting as SARS-CoV-2 M^{pro} inhibitors, which were designed starting from **Nirmatrelvir** [56]. They firstly introduced the [2.2.1]azabicylic ring at P2 position and the benzothiazolyl warhead at the *C*-terminus, then they began to explore different substitutions at P3. The most promising derivative was the compound **48** (Figure 29), containing a trifluoromethanesulfonamide group at the *N*-terminus and an adamantly group at P3. **48** exhibited an interesting inhibitory activity against SARS-CoV-2 M^{pro} (IC₅₀ = 1.65 μ M), a good antiviral activity against SARS-CoV-2 infected VeroE6 cells (EC₅₀ = 0.18 μ M) and low

cytotoxicity. Most importantly, the PK properties and target selectivity of compound **48** were superior than those of other derivatives [56].

P3

A8

SARS-CoV-2
$$M^{\text{pro}}$$
 $IC_{50} = 1.65 \, \mu\text{M}$

SARS-CoV-2 $EC_{50} = 0.18 \, \mu\text{M}$

Figure 29. Chemical structure and biological activity of compound 48.

In the already mentioned work of Kneller *et al.*, beside the development of nitrile-based compounds, the authors designed a novel SARS-CoV-2 M^{pro} inhibitor bearing a benzothiazolyl ketone warhead [32]. **49**, called **BBH-1**, was obtained by replacing the P1' ketoamide group of **Boceprevir** with benzothiazolyl ketone warhead, while the hydrophobic P1 group was replaced with a γ -lactam ring found in **GC-376** (Figure 30).

Figure 30. Chemical structure and biological activity of compound 49.

The antiviral activity of this inhibitor was determined in SARS-CoV-2 M^{pro} infected VeroE6 TMPRSS cells in presence of a P-glycoprotein inhibitor, exhibiting EC₅₀ value of 1.54 μ M, while the same experiment in absence of the P-glycoprotein inhibitor displayed EC₅₀ value of 16.1 μ M. The X-ray/neutron (XN) crystal structure of SARS-CoV-2 M^{pro} in complex with **49** was obtained (Figure 31), in order to get insights on the binding mode of the inhibitor and the protonation state of **49**-M^{pro} complex. The XN structure revealed the formation of a covalent bond between the Cys145 residue and the ketone warhead with the newly formed hemithioketal group unprotonated and its negatively charged oxygen inserted into the oxyanion hole of the target. The alkoxy anion is hydrated by a water molecule and stabilized by an H-bond with the Cys145 backbone. Water molecules interact also with the aromatic rings of the benzothiazolyl moiety, which due to its bulkiness pushes the His41 residue away from its original position causing the elimination of the catalytic water molecule from the active site and deprotonation of His164. The γ -lactam of the P1 group inserts into the substrate binding subsite S1 cavity, forming an H-bond between carbonyl oxygen and His163 sidechain. The remaining amide groups and urea group of **49** are involved in several H-bonds with His164 and Glu166, while the carbonyl oxygen of the urea moiety forms an H-bond with the side chain of Gln189 [32].

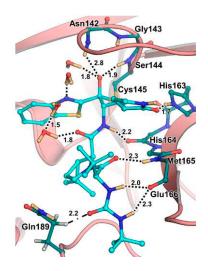


Figure 31. X-ray/neutron (XN) crystal structure of SARS-CoV-2 M^{pro} in complex with **49** (PDB: 7TD1).

Another interesting example of ketone-based SARS-CoV-2 M^{pro} inhibitors is represented by fluoromethyl ketones (FMKs). The electron-withdrawing effect exerted by the fluorine atoms in α -position of a methylketones enhances the electrophilicity of the carbonyl carbon toward nucleophilic attack of the thiol group of cysteinyl proteases [2]. Based on the number of fluorine atoms in α -position, it is possible to distinguish mono-fluoromethyl ketones (MFMKs), di-fluoromethyl ketones (DFMKs), and tri-fluoromethyl ketones (TFMKs) [57]. Zhu *et al.* reported a MFMK as SARS-CoV-2 M^{pro} inhibitor from a quantitative high throughput screening of existing compounds [58]. This compound, called **Z-FA-FMK** (50 in Figure 32A), is also an irreversible inhibitor of caspase-3. 50 showed a potent enzymatic inhibitory activity toward M^{pro} (IC50 = 11.4 μ M) and high antiviral activity (EC50 = 0.13 μ M), obtained from SARS-CoV-2 infected VeroE6 cells exploiting a CPE reduction assay [58]. Docking models of 50 with SARS-CoV-2 M^{pro} showed that the thiol group of Cys145 performs a nucleophilic attack at the α -position of the methylketone group in an Sn2-like reaction, where the fluorine atom acts as a leaving group (Figure 32B) [2]. However, MFMKs are easily catabolized forming toxic metabolites such as fluoroacetate, a feature that compromises their therapeutic utility [57].

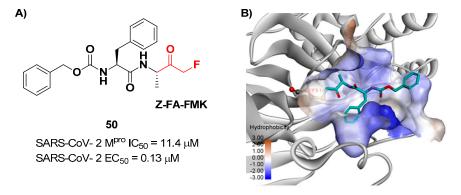


Figure 32. A) Chemical structure and biological activity of compound **50**; B) Predicted binding mode of **50** with SARS-CoV-2M^{pro}.

On the other hand, DFMKs and TFMKs seem to be more prone to the therapeutic use as they enhance the electrophilicity of the carbonyl group without manifesting metabolic issues [2]. In fact, TFMKs have been already investigated as SARS-CoV M^{pro} inhibitors, in particular with the development of compound **51**, as reported Shao *et al.* (Figure 33) [59]. Normally they act as reversible covalent inhibitors toward the target but, in their hydrated form, they can also act as transition-state competitive analogues [2]. Citarella *et al.* synthetized a new dipeptidyl DFMK as SARS-CoV-2 M^{pro}

inhibitor (*i.e.* compound **52**; Figure 33) [60], based on the corresponding TFMKs SARS-CoV M^{pro} inhibitor reported by Shao *et al.*, using a direct and chemoselective difluoromethyl unit transfer reaction [61,62]. **52** bears a Cbz-Leu-HomoPhe sequence as peptide framework linked to a *C*-terminal DFMK moiety. Compound **52** exhibited important antiviral activity toward MRC5 cell monolayer infected with hCoV-229E (EC₅₀ = 12.9 μ M). *In silico* studies showed that the two fluorine atom are able to establish halogen bond with residues of the S1' pocket [60].

Figure 33. Chemical structure and biological activity of compounds 51 and 52.

The ketone warhead was also applied in a non-peptidic compound, in particular in a bispidine-based ketone **53** (Figure 34). *In silico* studies suggested that the central carbonylic function act as an electrophilic warhead during the formation of the covalent adduct with Cys145. Enzymatic inhibitory activity of compound **53** toward SARS-CoV-2 M^{pro} showed noteworthy results (IC₅₀ = 0.75 μ M) [63].

Figure 34. Chemical structure and biological activity of compound 53.

7. α-Haloacetamides

The design of new SARS-CoV-2 M^{pro} inhibitors during these years has been also focused on α -haloacetamide-based compounds. The halogen atoms in α -position act as leaving groups during the cysteinyl nucleophilic attack on C-X, leading to an irreversible inhibition of the enzyme due to the formation of a S-C covalent bond. A first example of of α -haloacetamide SARS-CoV-2 M^{pro} inhibitor derives from the work of Stille *et al.* [64]. Starting from the non-covalent SARS-CoV-2 M^{pro} inhibitor 54 (Figure 35A), called X77, Stille and co-workers carried out a study of covalent docking by replacing the P1' imidazole ring with different electrophile warheads. From this study the most promising compound turned out to be 55 (Figure 35A), that showed an interesting enzymatic inhibitory activity against SARS-CoV-2 M^{pro} (IC50 = 0.41 μ M and K_I = 16 μ M). Structure-activity relationship (SAR) analysis of the entire panel of synthesized compounds revealed that the pyridine ring is fundamental to achieve inhibitory activity, the *tert*-butyl group is beneficial in terms of potency while the replacement of C-Cl/ C-F is detrimental [64]. The co-crystal structure of SARS-CoV-2 M^{pro}/55 is showed in Figure 35B: a covalent bond is formed between the thiol group of Cys145 and α -carbon, while the pyridyne and *tert*-butylphenyl rings fit into the S2 and S4 subpockets, respectively.

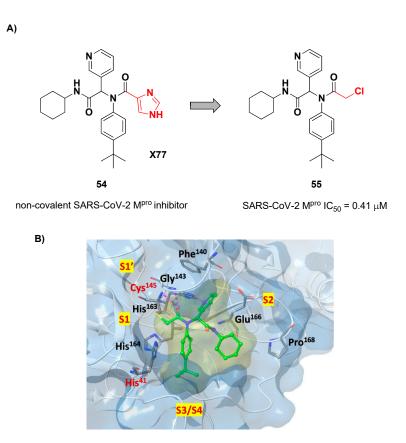


Figure 35. A) Chemical structure and biological activity of compounds **54** and **55**; B) X-ray structure of SARS-CoV-2 M^{pro} in complex with compound **55** (PDB: 7MLF).

The same approach of the previous work was used by Wang et~al. for the design of new SARS-CoV-2 α -haloacetamide inhibitors [65]. Starting from the non-covalent inhibitor **56** (Figure 36), called **23R**, they replaced its P1' furyl ring with a series of di- and tri-haloacetamides. The most promising compounds of this series turned out to be **57**, **58**, **59** and **60**, called **Jun9- 62-2R**, **Jun9-90-3R**, **Jun9-90-4R**, and **Jun9-88-6R**, respectively. Compounds **57**, **58** and **59** are endowed with an α , α -dichloroacetamide moiety as an electrophilic warhead at P1', while **60** contains an α , α , α -tribromoacetamide [65]. All compounds exhibited outstanding and selective M^{pro} inhibition (IC50 range 0.08–0.46 μ M) and good antiviral efficacy on Caco2-hACE2 cells infected with SARS-CoV-2 (EC50 range 0.58–2.07 μ M).

Figure 36. Chemical structure and biological activity of compounds 56-60.

The co-crystal structure of SARS-CoV-2 M^{pro} with compound 57 is shown in Figure 37 and confirms the ability of this electrophile warhead to establish a covalent bond with the Cys145 residue. Moreover, the R configuration of pyridine ring at P1 position seems to be fundamental for the fitting inside the S1 subsite.

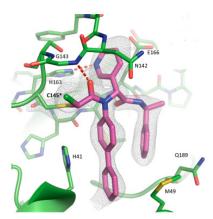


Figure 37. X-ray structure of SARS-CoV-2 M^{pro} in complex with compound 57 (PDB: 7RN1).

Based on compound **56**, a new series of α-chloro-fluoroacetamide (CFA) was proposed as covalent SARS-CoV-2 M^{pro} inhibitors by Yamane *et al.* [66]. This weak electrophilic warhead for cysteine –SH group was firstly explored in the design of tyrosine kinase inhibitors, demonstrating high selectivity with respect to other warheads. The CFA-S bond is reversable and can be hydrolyzed under neutral conditions to regenerate the cysteine residue [67]. For this reason, off-target reactions with cysteine proteases and related drawbacks are limited. The most effective compound of this series was **61** (Figure 38A), which contains a pyrimidine ring at P1 and a phenyl-pentafluorosulfanyl group at P2 [66]. Furthermore, it contains two stereocenters, thus four stereisomers. From the biological evaluation emerged the only the isomer with (R,R)-configuration exhibits outstanding and selective enzymatic inhibitory activity (IC50 = 56 nM). Docking simulations were performed for all stereoisomers of **61** (Figure 38B): the most stable pose of (R,R)-**61** suggests that the fluorine atom of

the CFA group is able to establish a H-bond with the backbone -NH of the Gly143 residue in the oxyanion hole at S1', inducing the activation of the inhibitor towards the Cys145 catalytic residue [66].

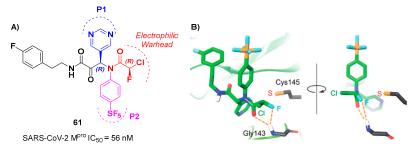


Figure 38. A) Chemical structure and biological activity of compound **61**; B) Docking of SARS-CoV-2 M^{pro} in complex with compound **61**.

Other α -haloacetamide-based SARS-CoV-2 M^{pro} inhibitors were discovered by Xiong *et al.* [68]. They first performed a virtual screening of a set of commercial non-peptidic compounds from the ChemDiv database, followed by covalent docking. From this study, 8 compounds were selected for the enzymatic inhibitory activity tests toward SARS-CoV-2 M^{pro}. Among them, 3 compounds showed remarkable activity with the piperazin-2-one-based compound **62** (Figure 39), called **Y020-9948**, as the most promising derivative (IC50 = 8.5 μ M). The discovery of this new non-peptidyl structure active toward M^{pro} led Xiong and co-workers to resolve the X-ray crystal structure of the complex SARS-CoV-2 M^{pro}/**62**, in order to understand the binding mode and in view of future developments of more potent analogs. From the X-ray analysis it is possible to observe the formation of a covalent bond between the Cys145 –SH group and the methylene group of the electrophilic warhead. Furthermore, the carbonyl carbon of the warhead establishes a H-bond with the Gly143 residue, while the *meta*-chlorophenyl moiety interacts by π - π stacking with His41 and a halogen-bond with the sulfur atom of the Met165 residue [68].

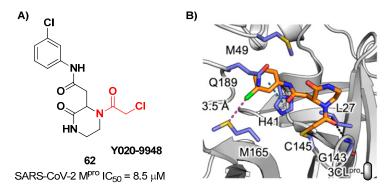


Figure 39. A) Chemical structure and biological activity of compound **62**; B) X-ray structure of SARS-CoV-2 M^{pro} in complex with compound **62** (PBD: 7VVT).

The non-covalent inhibitor **63** (Figure 40), called **MCULE-5948770040**, was discovered *via* high-throughput virtual screening by Clyde *et al.* [69]. It showed moderate inhibitory activity against M^{pro} (IC₅₀ = 4.2 μ M). From the X-ray structure of the complex SARS-CoV-2 M^{pro} /63 it can be observed that the P1-uracil and P2-dichlorobenzene moieties are inserted into the S1 and S2 subsites, respectively, linked through the piperazine ring which lays above the Cys145 catalytic residue, while the S3/S4 subsites remain unoccupied [69]. Starting from this piperazine-based hit compound, Gao *et al.* designed a panel of parent compounds from which , **64** (Figure 40), called **GC-14**, emerged as the most promising derivative [70]. It differs from the reference compound by the presence of a *N*-(thiophen-2-ylmethyl)acetamide group hanging on the piperazine core and a 3-pyridin ring in place of the pyrimidine-2,4(1*H*,3*H*)-dione nucleus at the *C*-terminus. **64** acted as a potent non-covalent inhibitor of SARS-CoV-2 M^{pro} , and also showed potent *in vitro* antiviral activity against SARS-CoV-2

(IC₅₀ = 0.4 μM and EC₅₀ = 1.1 μM) [69]. On the basis of these encouraging results, Gao *et al.* decided to introduce different electrophilic warheads on the piperazine ring of **64**, in order to develop covalent inhibitors [71]. 30 novel compounds came out from this work where the electrophilic warheads were linked to the piperazine ring via amide or sulfonamide bonds [71]. Among them, the most promising inhibitors were the α-chloroacetamide **65** (IC₅₀ = 0.18 μM) and the α-bromoacetamide **66** (IC₅₀ = 0.31 μM; Figure 40), called **GD-9** and **GD-13**, respectively (Figure 40). *In vitro* antiviral activity of these new potent SARS-CoV-2 M^{pro} inhibitors were also evaluated on VeroE6 cells infected with SARS-CoV-2, showing quite good results for compound **65** (EC₅₀ = 2.6 μM), but selectivity profile (CC₅₀ = 12.5 μM) [71].

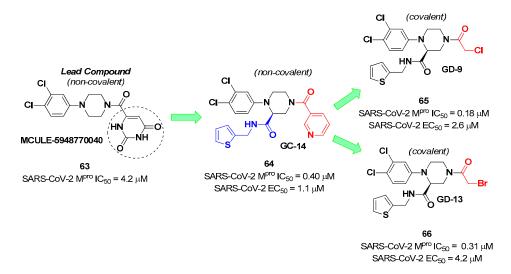


Figure 40. Chemical structure and biological activity of compounds 63-66.

The X-ray co-crystalized structure of SARS-CoV-2 M^{pro} with compound **66** confirmed the irreversible covalent binding mode which occurs between the Cys145 thiol group and the methylene carbon of the α -bromoacetamide moiety. The thiophen-2-ylmethyl substituent is surface-exposed and formed hydrophobic contacts with the Gln189 residue, the halogenated phenyl ring is inserted into the S2 pocket forming π – π stacking interactions with the His41 residue and a halogen-bond involving the *para*-chloro substituent and the Asp187 residue (Figure 41) [71].

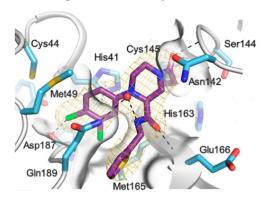


Figure 41. X-ray structure of SARS-CoV-2 Mpro in complex with compound 66 (PDB: 8B56).

Other interesting non-peptidyl SARS-CoV-2 M^{pro} inhibitors containing an α -chloroacetamide warhead are represented by the pyrazoline-based compounds **67** and **68** and the difluorinated amide **69** (Figure 42). Compound **67**, called **(R)-EN82**, was developed by Moon *et al.* and underwent activity-based protein profiling, a preliminary screening test applied on a library of 582 chloroacetamides and acrylamides [72]. From this first screening, 4 compounds emerged as promising antiviral candidates. Within this selection, **67** turned out to be the most active compound with an IC50 value of 0.53 μ M (Figure 42). Afterwards, Moon and co-workers carried out an exhaustive SAR analysis of this class of

derivatives by exploring the effect of different substituents at C-4 pyrazoline-ring. This work led to the discovery of compound **68**, called *cis*-**HW-2-010B**, which showed inhibitory activity against SARS-CoV-2 M^{pro} in the nanomolar range (IC₅₀ = 14 nM; Figure 42) [72]. In another study, El Khoury Léa *et al.* applied advanced *in silico* techniques for the design and synthesis of the cyclic α , α -difluoro-amide **69** (Figure 42), called **QUB-00006-Int-07**, acting as a covalent SARS-CoV-2M^{pro} inhibitor (IC₅₀ = 0.83 μ M) [73].

Figure 42. Chemical structure and biological activity of compounds 67-69.

8. α-Ketoamides

Another important electrophilic warhead that has been extensively explored in the design of SARS-CoV-2 M^{pro} inhibitors is the α -ketoamide moiety. The α -carbonyl group undergoes nucleophilic attack by the thiol group of the Cys145 catalytic residue, leading to the formation of a reversible covalent C-S bond. Furthermore, the α -ketoamide warhead establishes additional H-bonds with residues surrounding the active site, stabilizing the whole drug-target interaction. Examples of α ketoamide-based M^{pro} inhibitors came from the outstanding works of Zhang et al. [74,75]. As a first approach, they designed and synthetized a new series of peptidomimetic α -ketoamides acting as broad-spectrum inhibitors of enteroviruses, alpha- and beta-CoVs M^{pro} [74]. Among them, the most interesting derivative was 70 that showed noteworthy enzymatic inhibitory activity against SARS-CoV-2 M^{pro} (IC₅₀ = 0.67 μM) and antiviral activity (performed on human Calu-3 lung cells) in the micromolar range (EC₅₀ = $4-5 \mu M$). Cooper et al. demonstrated that the synthesis used to afford compound 70 led to two diastereomers differing for a single chiral center in P2 moiety (Figure 43) [76]. The diastereomeric resolution of 70, followed by the evaluation of the enzymatic inhibitory activity of the single isomers against M^{pro} led to the discovery that the diastereomer (S,S,S)-70 is more active than the diastereomer ($R_1S_1S_2$)-70 (IC50 = 0.12 μ M for ($S_1S_2S_2$)-70; IC50 > 5 μ M for ($S_1S_2S_2$)-70). The antiviral activity of (S,S,S)-70 was then assessed in different cell lines, finding that in human Calu-3 lung cells infected with SARS-CoV-2 it exhibited an EC50 value of 2.4 µM [76].

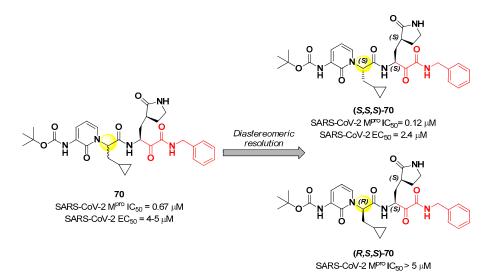


Figure 43. Structure and biological activity of 70 and resolution of the two diastereomers.

Another SARS-CoV-2 M^{pro} inhibitor bearing an α -ketoamide warhead was 71 (Figure 44A), a non-peptidic compound reported by Quan et al. [77]. This compound was obtained by multiple optimization rounds driven by in vitro and in vivo assays, involving the P1, P2, and P3 group of a bisamide bearing an α -ketoamide warhead. Afterwards, different groups linked to the α -ketoamide moiety at P1' position were explored. 71 is made up by an α -acetylketoamide at P1', a pyridyne ring at P1, a dibenzo(b,d) furan group at P2 and a 4-fluorophenyl ring at P3. During the optimization steps, the configuration of the stereocenter linking the P1–P2 units was explored, finding that the (R)epimers are always more active than the (S)-epimers. However, it was found that the (R)-epimers are prone to epimerization in vivo. In order to reduce such R/S interconversion, at the stereocenter of 71 was inserted a deuterium atom. 71 exhibited a potent inhibitory activity against SARS-CoV-2 Mpro (IC₅₀ = 8.1 nM) and outstanding oral bioavailability (92.9%, 31.9% and 85.7% in mice, rats and dogs, respectively). The in vitro antiviral activity of 71 was assessed on wild-type SARS-CoV-2 and three emerging variants: B.1.1.7 (Alpha), B.1.617.1 (Kappa) and P.3 (Theta), displaying EC50 values of 11.4 nM, 20.3 nM, 34.4 nM and 23.7 nM, respectively. Finally, the in vivo antiviral activity of the orallyadministered 71 was evaluated against Alpha and Kappa variants in infected K18-hACE2 transgenic mouse model, showing potent antiviral activity [77]. The X-ray co-crystal structure of SARS-CoV-2 M^{pro} in complex with **71** was also obtained (Figure 44B), showing a covalent bond between terminal ketone moiety and the Cys145 thiol group. The thioemiketal adduct is stabilized by two H-bonds formed by the thioemiketal-OH and the oxygen of the α -keto moiety with Cys145-amide and Gly143backbone, respectively. The pyridyne ring at P1 fits into the S1 pocket forming a H-bond with the His163 residue. The dibenzo[b,d]furan group at P2 is deeply inserted into the S2 pocket forming π - π interactions with the His41 residue. Moreover, this group is further stabilized through hydrophobic interactions with the S2 His41 and the 4-fluorophenyl ring at P3. The latter is also stabilized by hydrophobic interactions involving Met49 and Gln189 [77].

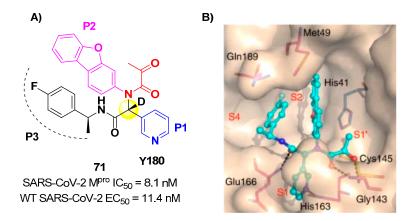


Figure 44. A) Chemical structure and biological activity of compound **71**; B) X-ray structure of SARS-CoV-2 M^{pro} in complex with **71** (PDB: 7FAZ).

An α -ketoamide-based SARS-CoV-2 M^{pro} inhibitor currently on phase III clinical trials is **72** (Figure 45A), called **RAY1216**, [78]. The design of this compound started from several SAR-optimization rounds of **Nirmatrelvir** involving the P1, P2, P3, and P4 moieties and the nitrile warhead at P1'. In particular, the α -ketoamide warhead at P1' has been functionalized with a cyclopentyl ring. Moreover, the inhibitor bears a γ -lactam moiety at P1, a cyclopentylproline at P2 (previously used in **Telaprevir**), a cyclohexylglycine at P3 and a tri-fluoroacetamide group at P4. **72** exibited a K_i = 8.4 nM in the enzymatic assay (a value comparable with **Nirmatrelvir**) [78]. The antiviral assays were performed on VeroE6 cell infected with different variants of SARS-CoV-2: the obtained EC50 were 95 nM for WT SARS-CoV-2, 130 nM for Alpha variant, 277 nM for Beta variant, 97 nM for Delta variant, 86 nM for Omicron BA.1 variant and 158 nM for Omicron BA.5 variant. The *in vivo* antiviral activity of **72** was tested at different doses in a human ACE2 transgenic mouse model, displaying similar efficacy to **Nirmatrelvir**. Pharmacokinetic properties of **72** were evaluated and compared with

Nirmatrelvir on different animal species (mice, rats and cynomolgus macaques), showing shorter elimination half-live and faster plasma clearance. The X-ray structure of **72** bound to SARS-CoV-2 M^{pro} showed the following (Figure 45B): i) in general, the structural elements in common with **Nirmatrelvir** maintain the same interactions with the target; ii) the formation of a covalent bond between the Cys145 residue and the α -ketoamide warhead is confirmed; iii) the thioemiketal-OH establish a H-bond with His41; iv) the α -ketoamide carbonyl oxygen accepts H-bonds from the oxyanion hole residues; v) the cyclopentyl moiety establishes hydrophobic interactions with Leu27; vi) P2 cyclopentylproline fits into the S2 subsite; vii) the P3 appers to have no important interactions with the target [78].

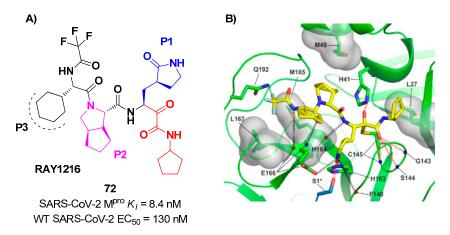


Figure 45. A) Chemical structure and biological activity of compound **72**; B) X-ray structure of SARS-CoV-2 M^{pro} with **72** (PDB: 8IGN).

A recent paper from Huang *et al.* proposed a novel α -ketoamide-based inhibitor with potent activity against SARS-CoV-2 Omicron variants [79]. They started with an *in vitro* screening of inhouse library containing more than 30,000 compounds. **73** (Figure 46A) was identified as a new hit compound, showing an IC50 value of 1.3 μ M. From the X-ray analysis of the complex SARS-CoV-2 Mpro/73 (Figure 46B) emerged that the α -ketoamide warhead binds covalently the Cys145 catalytic residue, interacting with several H-bonds with His41, His164 and residues of the oxyanion hole. The P1 benzyl and P1' thiazole moieties occupy the S1 and S1' subsites, respectively, while the P2 phenyl does not fit inside the S2 pocket, but it was observed exposed to the solvent region [53].

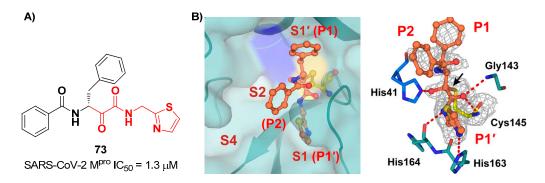


Figure 46. A) Chemical structure and biological activity of compound **73**; B) X-ray crystal structure of SARS-CoV-2 M^{pro} in complex with **73** (PDB: 8HHT).

The same research group performed a structural optimization of 73 focusing on three portions: the P1' thiazole ring, P1 and P2 phenyl rings. Each position was investigated keepin the other two fixed. The most active compound turned out to be 74 (Figure 47A), called SY110 [79]. This compound was selected based on its enzymatic inhibition activity, low cytotoxicity profile and PK parameters evaluated in Sprague-Dawley rats, displaying an IC_{50} value of 14.4 nM and higher area under the

curve and oral bioavailability as compared to other derivatives. Furthermore, **74** exhibited potent antiviral activity in plaque reduction assays against SARS-CoV-2 Omicron BA.1, its sub-lineages B.1.1.7, B.1.351 and BA.2, and against SARS-CoV-1 and MERS-CoV, demonstrating a pan-coronavirus antiviral efficacy (EC50 = 1.3, 0.38, 1.2, 2.8, 0.45 and 6.3 μ M against SARS-CoV-2 Omicron BA.1, BA.2, B.1.1.7, B.1.351, SARS-CoV-1 and MERS-CoV). *In vivo* experiments on Omicron-infected K18-hACE2 mouse models demonstrated the ability of **74** to alleviate the virus-induced pathology and, most importantly, to partially overcome **Nirmatrelvir**-resistance M^{pro} mutants in FRET assay [79]. X-ray co-crystal structure of SARS-CoV-2 M^{pro} in complex with **74** showed an atypical binding mode (Figure 47B): *i*) the α -ketoamide warhead was found covalently bound to the Cys145 catalytic residue in the (*R*)-configuration, forming several H-bonds with His41, Cys145 and Gly163; *ii*) the P1' thiazole moiety occupies the S1' subsite establishing a H-bond with His163; *iii*) the P1 benzyl moiety occupies the P1 pocket, establishing hydrophobic interactions with Thr25 and Leu27; *iv*) the P2 chiral ether is exposed to the solvent region near to a flexible loop; *v*) the P3 3,3-difluorocyclohexyl ring establishes hydrophobic interactions with the residues Met49, Met165 and Gln189 [79].

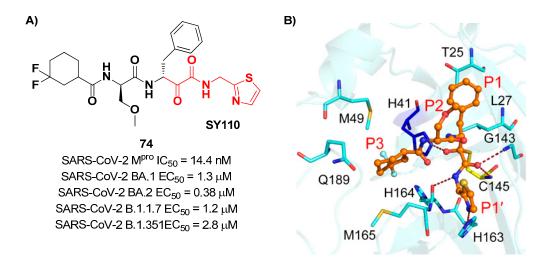


Figure 47. A) Chemical structure and biological activity of compound **74**; B) X-ray crystal structure of SARS-CoV-2 M^{pro} in complex with **74** (PDB: 8HHU).

9. Michael acceptors

Michael acceptors (MAs) such as α,β -unsaturated carbonyl, esters, vinyl, sulfonamides and nitriles have been widely used in the development of SARS-CoV-2 Mpro inhibitors. These groups exploit the electrophilic unsaturated β-position to form an irreversible covalent adduct with Cys145 through a Michael addition mechanism. The first example of MA as SARS-CoV-2 Mpro inhibitor is compound 75 (Figure 48A), called inhibitor N3, a peptidomimetic derivative previously reported as proteases inhibitor of SARS-CoV and MERS-CoV [80]. This pseudo-tetrapeptide contains a vinyl group and a benzyl-ester moiety at P1' (the MA warhead), a γ-lactam ring at P1, a Leu residue at P2, a Val residue at P3 and a N-terminal Ala residue (P4) capped with an isoxazol-3-yl group (P5). As expected, 75 showed a time-dependent enzymativ inhibition with $k_{obs} = 11,300 \,\mathrm{M}^{-1}\mathrm{s}^{-1}$, while the plaque antiviral assays on SARS-CoV-2 infected VeroE6 cells showed an EC₅₀ of 16.8 μM. The X-ray crystal structure of SARS-CoV-2 Mpro in complex with 75 (Figure 48B) confirmed the formation of a covalent bond between β-vinyl carbon and Cys145 –SH group, while the P1' benzyl group fits into the S1' subsite forming van der Waals interactions with Thr24 and Thr25. The P1 γ-lactam ring fits into the S1 subpocket and is H-bonded with His163 and Glu166. P2 Leu side chain is deeply inserted into the S2 pocket, while the P3 Val side chain is solvent-exposed. The P4 Ala side chain forms hydrophobic interactions with Met165, Leu167, Phe185 and Gln192, while the isoxazole ring at P5 establishes van der Waals interactions with Phe168. In the same study, another compound was taken into consideration for drug repurposing, i.e. CINANSERIN (76 in Figure 48A), a well-characterized hydroxytryptamine receptor antagonist discovered in 1960 (Figure 48A). **76** displayed an IC50 value of 125 μ M and an EC50 value of 20.6 μ M on infected VeroE6 cells [80]. Also in this case, the presence of an α - β unsaturated amide was supposed to be reactive toward catalytic Cys145 *via* Michael addition [22].

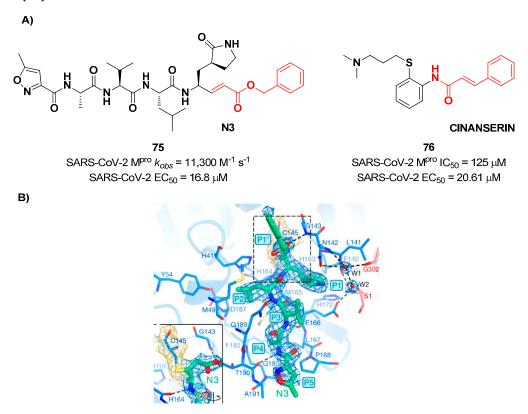


Figure 48. A) Chemical structure and biological activity of compounds **75** and **76**; B) X-ray structure of SARS-CoV-2 M^{pro} in complex with **75** (PDB: 6LU7).

Iketani *et al.* discovered compound 77 (Figure 49A), by an *in vitro* screening of existing SARS-CoV M^{pro} inhibitors [81]. It is characterized by a pseudo-tripeptide structure containing an acrylic ethyl ester moiety at P1', a γ-lactam ring at P1, a Leu residue at P2 and a *N*-Boc-*O-tert*-butyl-Thr at P3 position. 77 demonstrated to inhibit SARS-CoV-2 M^{pro} with an IC₅₀ value of 151 nM, while kinetic studies showed an inactivation rate (k_{inact}/K_i) of 4.13 × 10⁵ M⁻¹s⁻¹. Its ability to inhibit SARS-CoV-2 viral replication was tested in infected VeroE6 cells in CPE assays, showing an EC₅₀ value of 2.9 μM. Also for this compound, crystallographic studies confirmed the MA standard covalent binding mode (Figure 49B) [81].

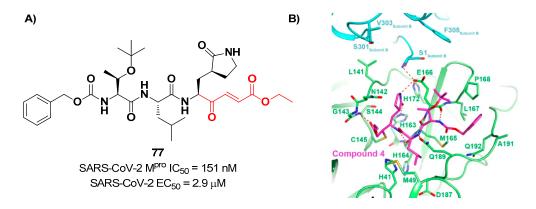


Figure 49. A) Chemical structure and biological activity of compound 77; B) X-ray structure of SARS-CoV-2 M^{pro} in complex with 77 (PDB: 7JT7).

Other examples of MAs as SARS-CoV-2 $M^{\rm pro}$ inhibitors came from a virtual screening campaign of an in-house database of ligands containing different MAs warheads, such as vinyl sulfones, vinyl amides, vinyl esters vinyl ketones, vinyl nitriles and vinyl phosphonates [82]. This strategy allowed to identify two compounds, 78 and 79 (Figure 50), containing a vinyl ketone moiety. Enzymeinhibition assays revealed that both compounds were able to modertely inhibit SARS-CoV-2 $M^{\rm pro}$, with an IC50 of 47.2 μ M and 157.5 μ M, respectively. Docking and molecular dynamics studies validated the covalent inhibition at the active site via Michael addition underlining the importance of the aliphatic residue at P2 and the aromatic ring with EWG groups in para position at P3 [82].

$$O_2$$
N

 O_2

Figure 50. Chemical structure and biological activity of compounds 78 and 79.

Lead optimization of **78** and **79** were performed by the same authors in a second work, consisting in the introduction of the γ -lactam moiety at P1 (as for most potent SARS-CoV-2 M^{pro} inhibitors) and different moieties at P2 [83]. From the enzymatic assays compounds **80**, **81** and **82** (Figure 51), called respectively **SPR38**, **SPR39** and **SPR41**, turned out to be the most promising derivativrs exhibiting activity in the sub-micromolar range ($K_i = 0.18$ –0.26 μ M). These compounds were also selected for cross-reactivity tests towards human cathepsin L and B showing activity in the micromolar range with the exception of **82**, which displayed a K_i value of 0.25 μ M against cathepsin L and was claimed as a dual inhibitor. The detected antiviral activity ranged from 1.5 to 18.5 μ M.

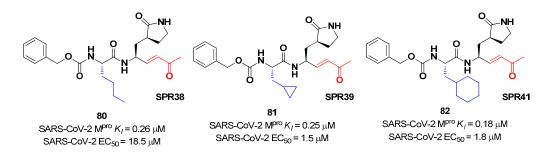


Figure 51. Chemical structure and biological activity of compounds 80-82.

Another lead optimization work (starting from a set of probes bearing an α -chloromethyl ketone warhead at P1') was performed by Mondal *et al.* and led to the identification of compounds **83** and **84** (Figure 52A) as promising MAs-based SARS-CoV-2 M^{pro} inhibitors [84]. The new MA warhead repliced the α -chloromethyl ketone moiety as the latter is associated with the already mentioned high reactivity and cytotoxicity. **83** and **84** showed inhibitory activity in micromolar range and antiviral activity in the nanomolar range (**83**: IC₅₀ = 0.9 μ M and EC₅₀ = 8.2 nM; **84**: IC₅₀ = 1.8 μ M and EC₅₀ = 14.7 nM). From a structural point of view, **84** contains a *N*-terminal Cbz group and a *para*-fluoro-D-Phe residue at P3, while **83** contains a free terminal amine and L- and D-Phe at P2 and P3, respectively. The high antiviral activity (as compared to the enzymatic inhibition activity) of these compounds may be explained by a dual-inhibition of M^{pro} and other proteases involved in SARS-CoV-2 replication cycle. Further investigation confirmed this hypothesis; indeed they inhibited cathepsin L in the nanomolar range (IC₅₀ = 60 nM for **83**, IC₅₀ = 145 nM for **84**). The good cytotoxic profile of these compounds led to *in vivo* investigations: the results showed that **83** and **84** inhibit the viral replication

in SARS-CoV-2 infected K18-hACE2 mice when administered intraperitoneally, indicating them as potential anti-SARS-CoV-2. The crystal structure of the complex $M^{pro}/83$ (Figure 52B) confirmed the formation of the Michael adduct with Cys145 and the fit of the γ -lactam ring into the S1 pocket [84].

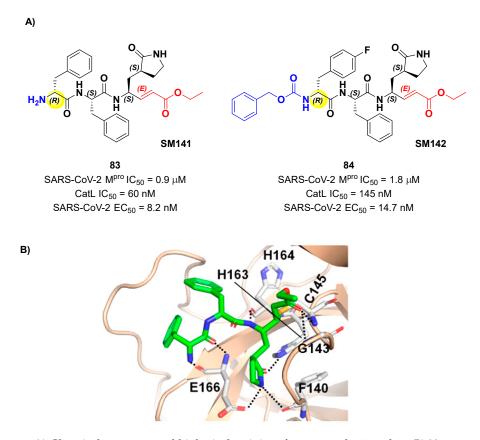


Figure 52. A) Chemical structure and biological activity of compounds **83** and **84**; B) X-ray structure of SARS-CoV-2M^{pro} in complex with **83** (PDB: 7MB0).

Citarella and co-workers explored the cinnamic ester moiety as a MA warhead for the covalent inhibition of SARS-CoV-2 M^{pro} [85]. In this work, they replaced the epoxyketone warhead in an already existing M^{pro} inhibitor, obtaining a pseudo-dipeptide with a backbone functionalized with different fragments at its *N*-terminus. From the enzymatic inhibition activity test the most promising compound turned out to be the carbamate derivate 85 (Figure 53), containing a *p*-OMe substituent on the *N*-terminal phenyl ring. The antiviral activity test on representative hCoVs demonstrated that 86 displayed EC50 values within the low micromolar range against hCoV-229E replication (α -CoV), while compounds 87 and 88 (Figure 53), both containing an indole moiety at the *N*-terminus, exhibited an interesting antiviral activity against hCoV-OC43 replication (β -CoV). Docking studies and mass experiments suggested the formation of a Michael adduct between the β position of the cinnamic ester and Cys145 [85].

Figure 53. Chemical structure and biological activity of compounds 85-88.

Stille *et al.* (already mentioned in the section of α -haloacetamides) developed a library of covalent inhibitors with different electrophile warheads, by replacing the P1′ imidazole ring in compound **54** (Figures 35A and 54A) [64]. Compounds **89** and **90**, bearing a vinyl sulphone moiety as a warhead, were selected as promising SARS-CoV-2 M^{pro} inhibitors as they showed enzymatic inhibitory activity in the sub-micromolar (IC50 = 0.42 μ M and K_i = 4.5 μ M for **89**; IC50 = 0.17 μ M and K_i = 2.3 μ M for **90**). **89** differs from **90** in having of a cyclohexyl group at the *N*-terminus in place of a 2-(*meta*-chlorophenyl)ethyl group. The binding of **89** was confirmed by X-ray crystallography (Figure 54B)[64].

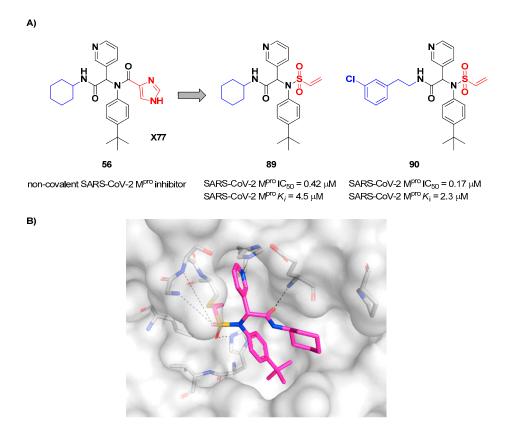


Figure 54. A) Chemical structure and biological activity of compounds **89** and **90**; B) X-ray structure of SARS-CoV-2 M^{pro} in complex with compound **89** (PDB: 7MLG).

Zaidman *et al.* designed MAs as SARS-CoV-2 inhibitors exploiting an automated pipeline, called *Covalentizer*, able to suggest new covalent inhibitors from non-covalent compounds [86]. Starting from the non-covalent SARS-CoV M^{pro} inhibitor **91** (Figure 55), called **ML-188**, they obtained a library of SARS-CoV-2 M^{pro} inhibitors by replacing its furan ring at P1' with an acrylamide warhead. The most active compound was **92** (Figure 55), which, apart from the P1' position, differs from **91** for *N*-terminus (*meta*-fluorophenetylamide moiety). This compound, obtained as a racemic mixture, exhibited M^{pro} inhibition in the micromolar range (IC₅₀ = 2.95 μ M). After chiral chromatography separation and test on single isomers, the (*S*)-enantiomer turned out to be by far the most derivative ((*S*)-**92**: IC₅₀ = 2.86 μ M; (*R*)-**92**: IC₅₀ = 86.3 μ M). The X-ray crystal structure of (*S*)-**92** in complex with SARS-CoV-2 M^{pro} confirmed the covalent binding mode at the acrylamide warhead, with the *p-tert*-butylphenyl group fitting into the S2 subsite and the fluorophenyl moiety establishing hydrophobic interactions with Met165 and Gln189 in the S4 cleft [86].

ML-188
$$SARS-CoV-2 M^{pro}$$
 inhibitor $SARS-CoV-2 M^{pro}$ iC $SO=2$ $SARS-CoV-2 M^{pro}$ iC $SO=2$ $SARS-CoV-2 M^{pro}$ iC $SO=2$ $SARS-CoV-2$ SA

Figure 55. Chemical structure and biological activity of compounds 91 and 92.

Also flavonoids were identified by *in vitro* screening as SARS-CoV-2 M^{pro} inhibitors acting as Mas. **Myrecitin** (93 in Figure 56) is one of the most important examples of M^{pro} inhibitors belonging to natural products [87]. Its pyrogallic motif *in vivo* undergoes oxidation to give an *ortho*-quinone function able to exert Michael's reactivity towards Cys145 [88]. The X-ray crystallography confirmed such hypothesis (Figure 56), highlighting the presence of a covalent bond between C6' and Cys145 – SH group. *In vitro* evaluations revealed high potency in the enzymatic assay (IC50 value 0.2– 0.6 μ M) and micromolar antiviral activity in the cell-based (EC50 = 8 μ M).

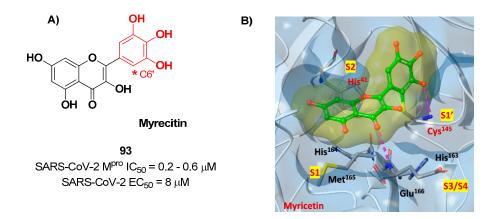


Figure 56. A) Chemical structure and biological activity of compound **93**; B) X-ray crystal structure of SARS-CoV-2 M^{pro} in complex with **93** (PDB: 7B3E).

In view of these promising results, several derivates of **Myrecitin** were synthetized (Figure 57) [88]. The introduction of a *para*-methyl group at its C7 led to compounds **94** and **95**, which showed higher potency ($IC_{50} = 0.30 \mu M$ for compound **94**; $IC_{50} = 0.26 \mu M$ for compound **95**) and good antiviral

activity (EC₅₀ = 12.6 μ M for **94**; EC₅₀ = 11.5 μ M for **95**). The addition of a phosponate group in the same position led to compound **96**, which exhibited good enzymatic inhibitory activity (IC₅₀ = 3.1 μ M) and the highest antiviral efficacy (EC₅₀ = 3.2 μ M) among these series of derivates [88].

Figure 57. Chemical structure and biological activity of compounds **94-96**.

10. Esters and Thioesters

Compounds bearing an ester moiety as electrophilic warhead are another class of M^{pro} inhbitors which has been taken into consideration during the COVID-19 outbreak. The mechanism of action of this type of inhibitors involves a nucleophilic attack by the Cys145 catalytic residue on the carbonyl carbon of the ester moiety and subsequential expulsion of the alkoxy group, resulting in an irreversible acylation of the target. Interestingly, this type of warhead was mostly applied on nonpeptidyl small molecules bearing rigid heterocycles such as pyridine and/or indole ring [22]. In fact, the first examples of ester-based SARS-CoV-2 Mpro inhibitors were indole/indoline-based chloropyridinyl compounds, previously widely reported in literature as therapeutic agents against SARS-CoV. Specifically, Ghosh et al. developed GRL-0920 and GRL-0820 (97 and 98 in Figure 58A) and both compounds exhibited good inhibitory activity against SARS-CoV-2 Mpro, displaying IC50 values of 0.25 μM and 73 nM for 97 and 98, respectively [89]. The activity on virus infected VeroE6 cells was 2.8 µM and 15 µM for 97 and 98, respectively [90]. The binding at the carbonyl group of the indole ring was demonstrated via ESI-QTOF/MS analysis and via X-ray crystal structure of SARS-CoV-2 M^{pro} in complex with 97 (Figure 58B) [89]. Another important compound is GRL-1720 (99 in Figure 58A), bearing an indoline scaffold [91]. This compound was able to irreversibly inhibit the activity of SARS-CoV-2 Mpro with IC50 value of 0.32 μ M, k_{inact} = 2.53 min⁻¹, K_i = 2.15 μ M and k_{inact} / K_i = 19,610M⁻¹s⁻¹. The antiviral assays on VeroE6 cells exposed to SARS-CoV-2 provided an EC₅₀ value of 15 µM for 99 without significant cytotoxicity [91].

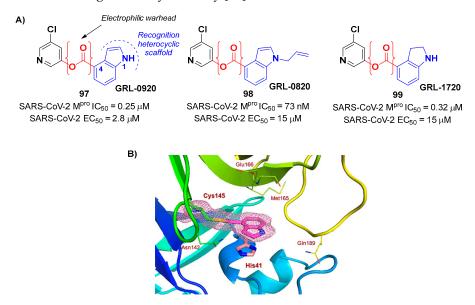


Figure 58. A) Chemical structure and biological activity of compounds **97-99**; B) X-ray crystal structure of SARS-CoV-2 M^{pro} in complex with **97** (PDB: 7RBZ).

On the basis of the structure of **97**, Breidenbach *et al.* identified other SARS-CoV-2 M^{pro} by exploring the different position of the ester moiety at the indole scaffold [92]. Compounds **100** and **101** (Figure 59), bearing the ester moiety at position 7 and 2, respectively, exhibited the highest activity (IC₅₀ = 55 nM and 34 nM, respectively).

Figure 59. Chemical structure and biological activity of compounds 100 and 101.

Interestingly, Iketani *et al.* identified another chloropyridinyl ester as potent SARS-CoV-2 M^{pro} inhibitor from a wide *in vitro* screening of already existing anti-SARS-CoV-agents targeting M^{pro} [81]. This compound, called **MAC-5576** (**102** in Figure 60A), exhibited potent inhibition activity with an IC₅₀ value of 81 nM. The X-ray crystal structure of **102** in complex with SARS-CoV-2 M^{pro} showed the covalent bond formation between Cys145 and acyl thiophene moiety (Figure 60B), although no time-dependent inhibition was observed in the enzymatic assay for this compound. The presence of the thiophene ring induces a conformational rotation of the catalytic His41 side chain, the pyridine ring rotates to arrange itself parallel to the thiophene nucleus forming π – π interactions [81].

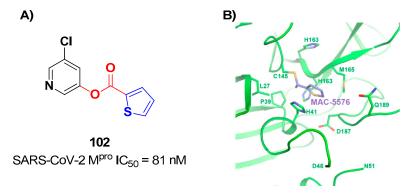


Figure 60. A) Chemical structure and biological activity of compound **102**; B) X-ray crystal structure of SARS-CoV-2 M^{pro} in complex with **102** (PDB: 7JT0).

On the indole scaffold Pillaiyar *et al.* investigated the thioester warhead [93]. First they performed a virtual screening on a library of more than 10,000 kinase inhibitors. Then, on the basis of these results, a wide panel of thioesters (*i.e.* compounds **103-109**; Figure 61A & C) were designed, synthetized and biologically tested. The top two compounds of this set were **103** (which displayed IC50 value of 11 nM, K_I value of 14 nM and k_{inact}/K_I value 58,700 M⁻¹s⁻¹) and **104** (which displayed IC50 value of 88 nM, K_I value of 33 μ M and a k_{inact}/K_I value 27,200 M⁻¹s⁻¹. All compounds showed promising results in the cell-based antiviral assay; in particular **108** that exhibited an EC50 value of 38 nM. The X-ray crystal structure of the complex SARS-CoV-2 M^{pro}/**103** showed that the Cys145 residue cleaves the inhibitor by formation of a thioester adduct, with the indole nucleus forming several hydrophobic interactions with His41, Met165, Asp187, Arg188, Gln189 and Met49 (Figure 61B) [93].

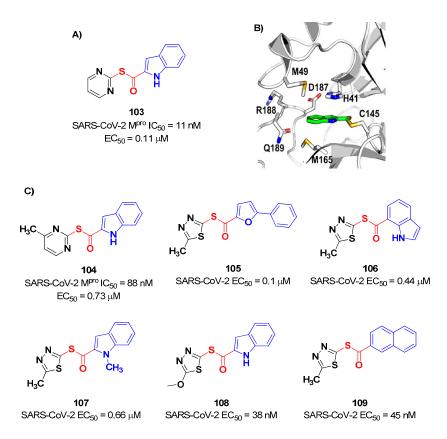


Figure 61. A & C) Chemical structure and biological activity of compounds **103-109**; B) X-ray crystal structure of SARS-CoV-2 M^{pro} in complex with **103** (PDB: 7X6K).

11. Selenium/sulfur-based derivatives

The investigation of selenium/sulfur-based derivatives as SARS-CoV-2 Mpro inhibitors emerged by the repurposing of **Ebselen** (110 in Figure 61), a non-specific binder that showed an IC50 value of 0.67 μ M in the enzymatic assay and an EC50 value of 4.67 μ M in virus infected VeroE6 cells [80,94]. Zmudzinski *et al.* screened a panel of **Ebselen** derivates with mono- or di-substitutions within the phenyl ring [95]. Four compounds (*i.e.* 111-114; Figure 62) exhibited higher activity than **Ebselen** in the enzymatic assay, highlighting the beneficial impact of such substitutions.

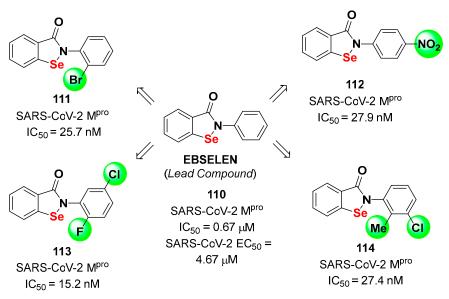


Figure 62. Chemical structure and biological activity of compounds 110-114.

Further SAR analyses on these **Elselen**-like derivatives bearing substitutions at the N-phenyl ring were carried out by Huff *et al.*(Huff et al. 2022) All compounds underwent enzymatic evaluation in a FRET-based assay and cell-based test in virus infected VeroE6 cells. Five of them (*i.e.* 115-119; Figure 63) exhibited IC50 values in the sub-micromolar range, while the most effective antiviral compound was 119 showing an EC50 of 0.8 μ M. Docking studies indicated the *meta*-substitution as the most favorable due to additional H-bond interactions with the target.

Figure 63. Chemical structure and biological activity of compounds 115-119.

Amporndanai *et al.* developed another series of **Ebselen**-like derivatives with different *N*-benzyl or *N*-aryl functionalities [94]. Among them, five derivatives (*i.e.* **120–124**; Figure 64) turned out to be more active than **Ebselen** both in the enzymatic and cell-based assay. In particular, **124** (or **MR6-31-2**) was ~3-fold more effective than **Ebselen** as SARS-CoV-2 M^{pro} inhibitor. The X-ray structure of the complex enzyme/**124** confirmed the same promiscuous mechanism of action of **Ebselen** (PDB: 7BAL) [94].

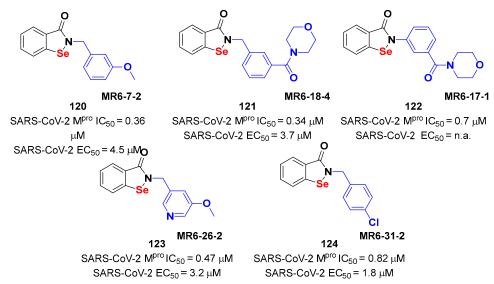


Figure 64. Chemical structure and biological activity of compounds 120-124.

Ebsulfur (*i.e.* 2-phenylbenzisothiazol-3(2*H*)-one **125**; Figure 65) is the sulfur isosteric analogue of **Ebselen**. Likewise **Ebselen**, **Ebsulfur** has been taken into consideration as a lead compound for the design of SARS-CoV-2 M^{pro} covalent inhibitors [97]. Sun *et al.* performed an *in vitro* screening of several analogous compounds, noting that derivates containing the furane substituent displayed higher inhibition against M^{pro} than **Ebselen** and **Ebsulfur**. In fact, compounds **126** and **127** (Figure 65) displayed a IC₅₀ values of 0.11 μM and 74 nM, respectively. Molecular modeling studies evidenced

that the furan ring plays a beneficial role in the binding mode with the target by forming hydrophobic interactions with Met165, Arg188, Asp187 and Met49 [97].

Figure 65. Chemical structure and biological activity of Ebsulfur, 126 and 127.

Further **Ebsulfur** analogues has been explored by Chen *et al.* providing structural key features responsible for the activity [98]. By a high-throughput screening of diverse compound libraries, they identified the hit compound **128** (IC $_{50}$ = 190 nM; Figure 66), whose optimization involved three recurring moieties, namely the benzoisothiazole core, a linker and a terminal variously substituted phenyl ring (Figure 66). The first series of compounds involved the replacement of the 3,5-dichlorophenyl ring of **128** with other substituted aromatic rings or other moieties [98]. In this first seriers, derivates **129-134** exhibited enzymatic inhibition activity higher than the hit **128** [98].

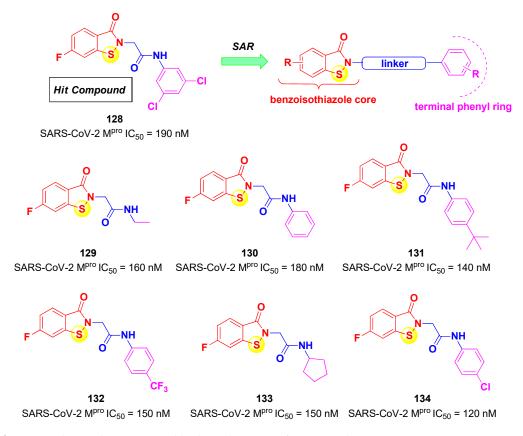


Figure 66. Chemical structure and biological activity of compounds 128-134.

The SAR analysis evidenced that the electronic effects on this ring are less important than potential establishment of π - π interaction within the target. Therefore, the simple phenyl ring was chosen later on for further optimizations. However, both modifications of the acetamide linker and benzoisothiazole core ring did not bring about more active compounds. Substitutions on the benzoisothiazole core instead led to interesting results (*i.e.* compounds **135-137**; Figure 67), in particular by switching the fluorine atom from position 6 to position 4 as for **135** (IC50 = 116 nM against SARS-CoV-2 M^{pro}) [98].

Figure 67. Chemical structure and biological activity of compounds 135-137.

12. Non-covalent inhibitors

Non-covalent inhibitors do not possess an electrophilic warhead and generally display less reactivity and improved selectivity as compared to covalent inhibitors. They establish only secondary interactions with the active site such as H-bonds, hydrophobic stackings, Van der Waals forces, electrostatic interactions and salt bridges [99]. Some examples of SARS-CoV-2 M^{pro} non-covalent inhibitors have been already introduced throughout the article as they may have acted as lead structures for the development of covalent inhibitors. These include compound **54** (**X77**), **56** (**23R**), **63** (**MCULE-5948770040**) and **64** (**GC-14**). **54** (Figure 68A), previously identified as SARS-CoV M^{pro} inhibitor [100], was investigated by Stille *et al.* in complex with SARS-CoV-2 M^{pro} *via* X-ray analysis [64]. The imidazole-amide moiety interacts via H-bonds with His41 and Gly143, while it is positioned at 3.2 Å from Cys145-SH. The pyridyne ring fits into the S2 pocket forming a H-bond with His163 side chain, while the *tert*-butylphenyl group is deeply inserted into the S4 subpocket. The carbonyl oxygen of the cyclohexyl-amide moiety accepts a H-bond from the –NH function of Glu166 (Figure 68B). Its enzymatic inhibitory activity was in the micromolar range (IC₅₀ = 4.1 µM) [64].

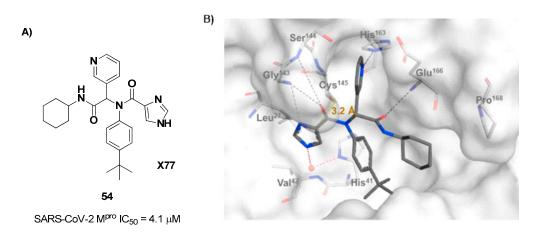


Figure 68. A) Chemical structure and biological activity of the compound **54**; B) X-Ray crystal structure of SARS-CoV-2 M^{pro} in complex with **54** (PDB: 6W63), highlighting binding site interactions.

Kitamura *et al.* developed **56** (compound **23R** in Figure 69A) which displayed an IC50 value of 0.2 μ M in the enzymatic test and an EC50 value of 1.3 μ M in the antiviral assay. The X-ray crystal structure of SARS-CoV-2 M^{pro} in complex with **56** is reported in Figure 69B. The furylamide moiety at P1' forms a bifurcated H-bond with Gly143, the biphenyl group at P2 fits into the S2 pocket forming

hydrophobic interactions, while the pyridine ring at P1 occupies the S1 pocket and establishes several hydrophobic interactions and a H-bond with the His163 residue. The amide group that links the pyridine ring to the α -methylbenzyl group establishes a H-bond with the main chain of Glu166n, while the phenyl ring of the α -methylbenzyl moiety is partially positioned in both the S2 and S4 pockets forming π - π interactions with the external phenyl of the biphenyl group [101].

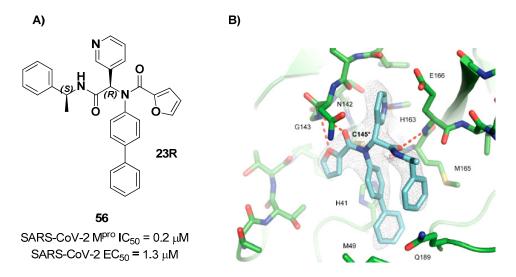


Figure 69. A) Chemical structure and biological activity of the compound **56**; B) X-ray crystal structure of SARS-CoV-2 M^{pro} in complex with **56** (PDB: 7KX5).

As already mentioned, the non-covalent inhibitor **63** (Figure 70A) was discovered *via* high-throughput virtual screening by Clyde *et al.* and it was the starting point for the design of piperazine-based covalent compounds [69]. From the X-ray structure of the complex SARS-CoV-2 M^{pro}/**63** it can be observed that the P1-uracil and P2-dichlorobenzene moieties are inserted into the S1 and S2 subsites, respectively, linked through the piperazine ring which lays above the catalytic Cys145, while the S3/S4 subsites remain unoccupied (Figure 70B) [69].

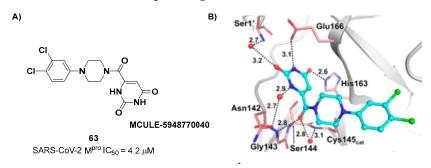


Figure 70. A) Chemical structure and biological activity of compound **63**; B) X-ray structure of SARS-CoV-2 M^{pro} in complex with **63** (PDB: 7LTJ).

Gao *et al.* performed several modifications of **63** in order to occupy each pocket of the active site of M^{pro}, and he most promising compound of this work turned out to be **64** (Figure 71A) [70]. The cocrystal structure of complex M^{pro}/**64** (Figure 71B) showed that the nicotinic group at P1 fits into the S1 subsite and establishes a H-bond with the His163 residue. The dichlorophenyl group at P2 in inserted into the S2 pocket forming π - π interactions His41 side chain and a halogen bond between the chlorine atom at C-4 and Asp187 backbone. The 2-methylthiophene group at P4 fits into the S4 pocket forming a H-bond with the Glu166 residue, while the piperazine ring shows the same binding mode as **63**. Additionally, the P4 thiophene, P2 3,4-dichlorophenyl ring and imidazole side chain of His41 are involved in a sandwich-like π - π interactions, similarly to **56** [70].

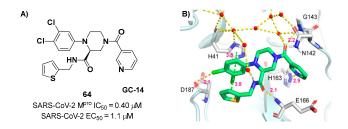


Figure 71. A) Chemical structure and biological activity of compound **64**; B) X-ray structure of SARS-CoV-2 M^{pro} in complex with **64** (PDB: 8ACL).

In early 2022, Unoh et al. discovered S-217622 (compound 140 in Figure 72A), the first oral noncovalent, nonpeptidic SARS-CoV-2 Mpro inhibitor clinical candidate [102]. This compound was obtained via screening of an in-house library subjected to a docking-based virtual screening with predefined filters (a hydrophobic moiety in S2 and two H-bond acceptors in S1) followed by biological screening. The obtained hit compound 138 (Figure 72A) was characterized by good in vitro inhibitory activity (IC₅₀ = 8.6 µM) and optimal in vitro and in vivo PK profile for oral administration. The X-ray crystal structure of SARS-CoV-2 Mpro in complex with 138 was resolved to understand important binding interactions to maintain for a further hit optimization [102]. To optimize the interaction at S1', the 4-difluoromethoxy-2-methylbenzene group was replaced with a 6-chloro-2methyl-2H-indazole scaffold, which maintained a H-bond with Thr26, achieving compound 139 (Figure 72A). This compound displayed an outstanding improvement in the inhibitory activity (IC50 = 96 nM) and a good antiviral activity against infected VeroE6 cells (EC₅₀ = 12.5 µM), maintaining the favorable PK properties as 138 [102]. Additional modifications entailed the replacement of the methyl-amide function at P1 with a 1-methyl-1H-1,2,4-triazole ring, affording the clinical candidate 140 (Figure 72A). This compound displayed excellent enzymatic inhibitory and antiviral activity (IC₅₀ = 13 nM; EC₅₀ = $0.37 \mu M$ on WT SARS-CoV-2) and optimal PK profile, especially in monkey and dog models. 140 demonstrated also to inhibit the viral replication of a series of SARS-CoV-2 variants (WT, Alpha, Beta, Gamma, Delta and Omicron with EC50 values from 0.29 to 0.5 μM) as well as different human coronaviruses (SARS-CoV, MERS-CoV, hCoV-229E and hCoV-OC43 with EC50 values from 74 nM to 5.5 μM) [102]. These promising results prompted Unoh and co-workers to test 140 in vivo in mice models infected with SARS-CoV-2 Gamma strain: 140 was administered orally immediately and 12 hours after infection, exhibiting rapid efficacy measured after 24 hours. These promising results in pre-clinical studies make the SARS-CoV-2 Mpro inhibitor 140 a potential clinical candidate for oral administration. The X-ray crystal structure of SARS-CoV-2 Mpro complexed with 140 showed that the 1-methyl-1*H*-1,2,4-triazole group occupy the S1 pocket establishing a H-bond with the His163 backbone. The 2,4,5-trifluorobenzylic group at P2 fits into the S2 pocket forming a face-to-face π - π interactions with a rotated His41 residue, while the 6-chloro-2-methyl-2H-indazole scaffold at P1' maintains the H-bond with Thr26 and establishes hydrophobic interactions with the Met49 residue (Figure 72B) [102].

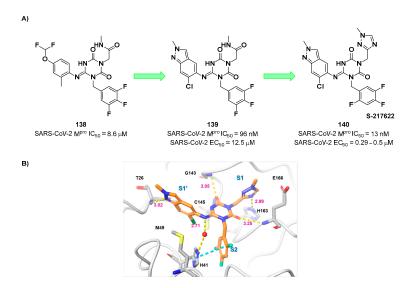


Figure 72. A) Chemical structure and biological activity of compounds **138-140**; B) X-ray crystal structure of SARS-CoV-2 M^{pro} in complex with **140** (PDB: 7VU6).

Another contribution in this field was made by Yang et al. which performed a multiple conformational-based virtual screening on a library of more than 8,000 compounds [103]. This screening was based on nine different conformations of SARS-CoV-2 Mpro substrate-binding site, and the 49 top-ranked compounds were then evaluated for their binding profiles. Six compounds were able to bind M^{pro}; therefore, they were selected for *in vitro* evaluation of their enzymatic inhibitory activity on FRET assays. All of them were able to inhibit Mpro with IC50 values ranging from 0.69 to 2.05 µM. The two most promising compounds from the enzymatic inhibitory assay, namely compound 141 (IC₅₀ = 0.73 μ M), called Z1244904919, and compound 142 (IC₅₀ = 0.69 μ M), called Z1759961356, were selected for the evaluation of their antiviral activity on SARS-CoV-2 infected VeroE6 cells in plaque reduction assays, displaying EC50 values of 5.0 and 8.5 μM, respectively (Figure 73A). The hypothetical binding modes between these compounds and SARS-CoV-2 Mpro were elucidated by MD simulation and binding free energy analysis and is depicted in Figure 73B and Figure 73C: in 141, the fluorophenol moiety fits into the S1 pocket, while the piperidine ring interacts with the residues of the S4 pocket and connect with the indoline nucleus which, in turns, establishes interactions with the residues of the S2 and S3 sites. 142 has a higher binding affinity due to suggested additional interactions with the His164 and Met165 in the S1 subsite and Asn47 in the S2 subsite [103].

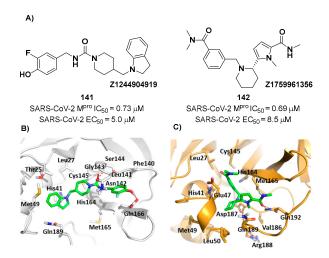


Figure 73. A) Chemical structure and biological activity of compounds **141** and **142**; B) Docking of SARS-CoV-2 M^{pro} in complex with **141**; C) Docking of SARS-CoV-2 M^{pro} in complex with **142**.

Han *et al.* developed a novel non-covalent SARS-CoV-2 M^{pro} inhibitor starting from the already known benzotriazole-based SARS-CoV inhibitor **ML-300** (**143** in Figure 74) [104]. The optimization of **143** led to compound **144**, called **CCF981** (Figure 74), the most potent compound of the work with an IC50 value of 68 nM against SARS-CoV-2 M^{pro} and EC50 values in virus infected VeroE6 cells of 0.50 and 0.56 μ M in cytopathic effect (CPE) inhibition and in a plaque reduction assay, respectively.

SARS-CoV-2 Plague reduction EC
$$_{50}$$
 = 0.56 μ M SARS-CoV-2 Plague reduction EC $_{50}$ = 0.56 μ M

Figure 74. Chemical structure and biological activity of compounds 143 and 144.

Another interesting compound was obtained by Elsegini *et al.* through a pharmacophore-based virtual screening against M^{pro}, PL^{pro} and human furin protease, an enzyme involved in the cleavage of spike protein during viral entry, with the aim to select potential dual inhibitor to maximize the antiviral efficacy [105]. After mapping more than 500,000 compounds, only 16 of them were selected for biological evaluation. Among them, compounds **145** and **146** showed inhibition activity against M^{pro} and human furin protease in the sub-micromolar range, but activity against PL^{pro} in the nanomolar range (Figure 75A). Both compounds exhibited a promising antiviral activity when tested *in vitro* against NRC-03-nhCoV, the SARS-CoV-2 strain isolated in Egypt, in VeroE6 cells, exhibiting EC₅₀ values of 0.77 and 0.11 μ M, respectively. Docking studies suggested that both compounds form H-bonds with Asn142 and hydrophobic interactions with Met49 and Ile41; compound **145** forms H-bonds with Thr24, Thr25, and Gln189, while compounds **146** forms an H-bond with Glu166 and a π - π interaction with His41 (Figure 75B and Figure 75C) [105].

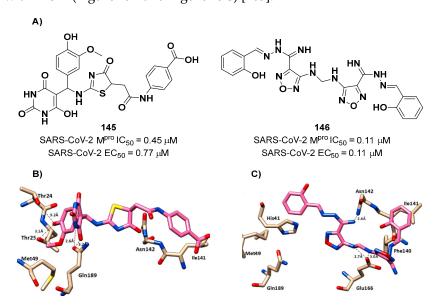


Figure 75. A) Chemical structure and biological activity of compounds **145** and **146**. B) Docking of SARS-CoV-2 M^{pro} in complex with **145**; C) Docking of SARS-CoV-2 M^{pro} in complex with **146**.

Luttens *et al.* were able to identify non-covalent M^{pro} inhibitors through a virtual screening applied on ultralarge chemical libraries [107]. This strategy brought about the selection of top-ranked compounds for enzyme inhibition assay. Three of them were able to inhibit SARS-CoV-2 M^{pro}. The hydantoin-based compounds **147** and **148** (Figure 76) were co-crystalized with SARS-CoV-2 M^{pro} and

the X-ray crystal structures (PDB: 7B2U and PDB: 7AU4) showed that the hydantoin carbonyl groups interacts with the Gly143 and Glu166 backbone via H-bonds, while the substituents on the hydantoin core fits into S2 and S1 pockets. Furthermore, compounds **147** and **148** were subjected to hit-to-lead optimization driven by guided by docking predictions and determined crystal structures [107]. This strategy was focused on the maintenance of the hydantoin core and variation of the groups at P1 and P2 positions. From this study emergerd that the isoquinoline and spyro-cyclic scaffolds were the best chemical frameworks for the P1 and P2 position, respectively. Compound **149** (Figure 76), characterized by an *ortho*-chlorophenyl ring as aromatic tail at the P2 position, displayed the best inhibitory activity against SARS-CoV-2 M^{pro} with IC50 = 77 nM and K_D = 38 nM. This compound was also subjected to CPE-based assays performed in infected Huh7 cells, showing an EC50 value of 0.11 μ M. **149** was also screened against SARS-CoV and MERS-CoV in CPE-based assays, demonstrating a broad-spectrum activity against coronaviruses (SARS-CoV-1 EC50 = 0.39 μ M in VeroE6 cells, MERS-CoV EC50 = 0.20 μ M in Huh7 cells) [107].

enzyme activity at
$$50~\mu\text{M} = 26~\%$$
 enzyme activity at $50~\mu\text{M} = 63\%$ enzyme activity at $50~\mu\text{M} = 63\%$ enzyme activity at $50~\mu\text{M} = 63\%$ sars-cov-2 EC₅₀ = 0.11 μM

Figure 76. Chemical structure and biological activity of compounds 147-149.

Another important non-covalent SARS-CoV-2 M^{pro} inhibitor was developed by Rossetti *et al.* by examination of hits derived from two *in silico* screening studies (REAL Space and ZINC compound libraries) on two different structures of M^{pro} [108]. From the top-ranked hits, a total of 486 compounds were selected on the basis of fragment screening, drug-likeness and chemical diversity. Only five of them were effective M^{pro} inhibitors. The top two are characterized by a dihydro-quinolinone core (*i.e.* **150** and **151**; Figure 77). They underwent a first round of chemical structure similarity searches that led to the discovery to three significantly more potent derivatives, *i.e.* **152**, **153** and **154** (Figure 77), called **Z228770960**, **Z393665558** and **Z225602086**, respectively, with two of them able to enhance the melting temperature of M^{pro} in a thermal-shift assay. Further optimizations were obtained after a second step of chemical structure similarity searches(*i.e.* **155-158**, called **Z222979552**, **Z228166018**, **Z222977344** and **Z222978028**, respectively; Figure 77) [108].

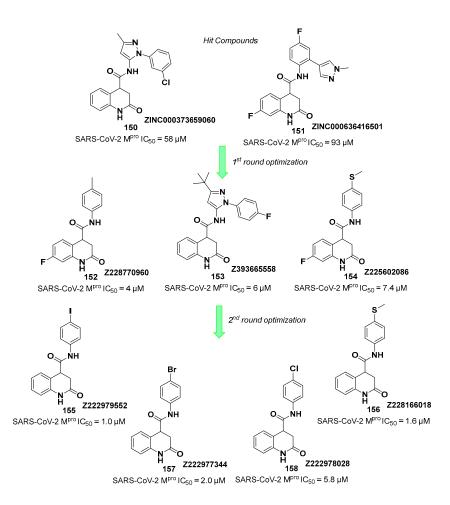


Figure 77. Chemical structure and biological activity of compounds 150-158.

The co-crystal structure of the most potent derivative **155** with SARS-CoV-2 M^{pro} confirmed the non-covalent mechanism of action (Figure 78): the dihydro-quinolinone core forms several H-bonds with Glu166, His163, His172, while the carbonyl oxygen of the amide bond accepts two H-bonds from Cys145 thiol and Glu166 backbone, respectively. The *para*-iodobenzene ring establishes π - π interactions with the His41 residue and additional hydrophobic interaction with Asn142, Met49 and Met165 [108].

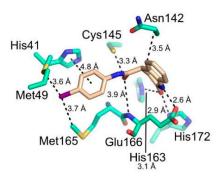


Figure 78. X-ray crystal structure of SARS-CoV-2 Mpro in complex with 155 (PDB:7P2G).

Other quinazolin-4-one-based non-covalent inhibitors of SARS-CoV-2 M^{pro} are compounds **160** and **161** (Figure 79A), proposed by Zhang *et al.* [109], which were developed starting from the lead **Baicalein** (**159** in Figure 79A) by replacing its chromen-4-one ring and varying the substitution pattern in C2 and N3 positions. The most interesting compound (**160**) contains a phenyl group both at C2 and N3 position and posses higher activity as compared to **Baicalein** [56]. Furthermore, **160** and **161** showed optimal selectivity against other human proteases and low cytotoxicity [109]. The co-

crystal structure of SARS-CoV-2 M^{pro}/**161** is reported in Figure 79B: the three hydroxy groups form a H-bond pattern with the backbone of Gly143, Ser144 and Gly145, and with the imidazole ring of the His163 residue via a water molecule, while this aromatic portion forms hydrophobic interactions with Cys145 Asn142 and His41. The carbonyl oxygen of the quinazolin-4-one core forms a H-bond with the Glu166 backbone, while the 3'-methyl-4'-fluorophenyl substituent at N3 position deeply fits into the S2 pocket. The *sec*-butyl substituent at C2 position matches a newly formed cavity called S2c pocket, a result of a ligand-induced conformation change involving the side chains of Met49 and Gln189 [109].

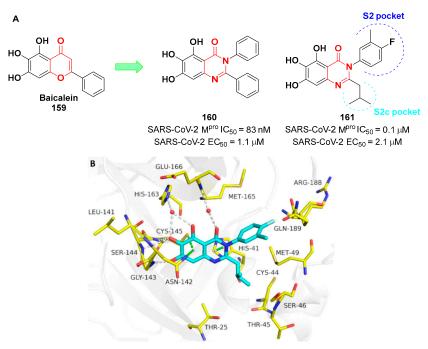


Figure 79. A) Chemical structure and biological activity of compounds **159** - **161.** B) X-ray crystal structure of **161** in complex with SARS-CoV-2 M^{pro} (PDB: 8I4S).

Using a similar rational approach, Citarella *et al.* designed a novel SARS-CoV-2 M^{pro} inhibitor characterized by a trifluoromethyl diazirine ring [110]. In the context of developing fluorinated functionalities able to target druggable enzymes [111], they replaced the epoxyketone warhead in an already existing M^{pro} inhibitor with a trifluoromethyl diazirine ring to afford compound **162**, called **MPD112** (Figure 80A). This compound showed *in vitro* inhibition activity against SARS-CoV-2 M^{pro} at low micromolar level (IC₅₀ = 4.1 µM), no cytotoxicity and selectivity toward M^{pro} against PL^{pro}. Furthermore, mass experiments with SARS-CoV-2 M^{pro} revealed no covalent interaction between target and ligand, suggesting a non-covalent mechanism of inhibition. A molecular docking approach was exploited to shed light on the binding mode of **162** within the target active site (Figure 80B): docking results showed that the trifluoromethyl diazirine ring is located in close proximity to the Cys145 thiol group forming a halogen bond with the side chain of Thr26. The benzyl carbamate moiety establishes several hydrophobic interactions with Phe140, Glu166 and a H-bond with the oxygen of the Glu166 backbone. The central phenyl group forms hydrophobic interactions with Pro168, Leu167 and Gln189 [110].

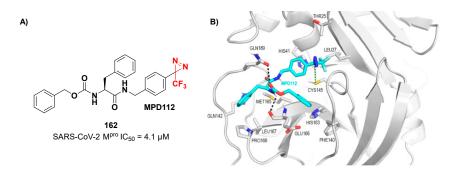


Figure 80. A) Chemical structure and biological activity of compound **159**; B) SARS-CoV-2 M^{pro}/**162** complex suggested by molecular docking.

In early 2022, Alhadrami *et al.* reported the indole alkaloid **Neoechinulin A** (**163** in Figure 81A) [112], isolated from the Red Sea fungus *Aspergillus fumigatus*, as a potential SARS-CoV-2 M^{pro} inhibitor through bio-guided screening. This compound showed a promising inhibitory activity with an IC50 value of 0.47 μ M. The mechanism of inhibition was elucidated using docking and molecular dynamics simulations: **163** is characterized by a diketopiperidine nucleus which forms four hydrogen bonds with the Leu141, Asn142 and Gly143 residues of the S1' sub-pocket and with Glu166 residue of the S2 sub-pocket. Important hydrophobic interactions were also observed with the His41 residue (Figure 81B) [112].

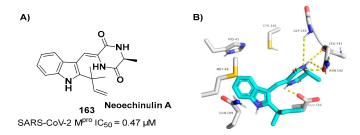


Figure 81. A) Chemical structure and biological activity of compound **163**. B) Docking of SARS-CoV-2 M^{pro} in complex with **163**.

Zhang *et al.* demonstrated that the natural product **Shikonin** (**164** in Figure 82A) is a broad spectrum anti-coronaviruses agent, with micromolar IC50 values against the M^{pro} of SARS-CoV-2, SARS-CoV, MERS-CoV, hCoV-HKU1, hCoV-NL63, and hCoV-229E [113]. The X-ray crystal structure of SARS-CoV-2 M^{pro} in complex with **Shikonin** was determined by Li *et al.* in a previous work [114]. They found out that the presence of **Shikonin** in the active site cleft of the enzyme causes a conformational change in the catalytic dyad His41-Cys45 and establishes a face-to-face π - π interaction involving the His41 residue of the target and the naphtoquinone ring of the inhibitor. One of the two –OH groups of **Shikonin** establishes H-bonds with the Cys145 –SH group and His164 backbone, while the *iso*-hexenyl group establishes H-bonds with Arg188 and Gln189 residues. Another important difference is the change in position of a flexible loop, including Cys44 to Tyr54, Asp187 to Ala191, and Leu141 to Ser144.

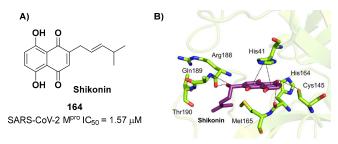


Figure 82. A) Chemical structure and biological activity of compound **164**; B) X-ray crystal structure of SARS-CoV-2 M^{pro} in complex with **164** (PDB: 7CA8).

Zhang *et al.* discovered two 9,10-dihydrophenanthrene derivatives as novel SARS-CoV-2 M^{pro} inhibitors through an *in vitro* screening of an in-house compound library based on the same scaffold [115]. The first round of optimization pointed out the importance of the *para*-bromo phenyl group at R1 and the pyridyne ring at R3 for the inhibitory activity. Then, these two units were maintained for further optimizations carried out on the pyridine ring. The aliphatic –OH group was taken into account for obtaining hydrolyzable esters. The enzymatic assay revealed that compounds **165** and **166** (Figure 83), called **C1** and **C2**, were the most promising derivatives with IC₅₀ values of 1.5 μM and 1.8 μM, respectively. Kinetic studies provided *Ki* values of 6.1 μM and 7.6 μM, for **165** and **166**, respectively, and suggested a dose-dependent mixed-inhibition mechanism of SARS-CoV-2 M^{pro}, which means that these inhibitors can bind at least two sites in the target protein, probably the substrate-binding site and the dimer interface. Indeed, molecular docking simulations confirmed this bimodal binding mechanism for **165** [115].

Figure 83. Chemical structure and biological activity of compound 165 and 166.

13. Allosteric inhibitors

Pelitinib (compound **167** in Figure 84A) is an irreversible covalent pan-inhibitor of the epidermal growth factor receptors developed as anticancer agent and repurposed as SARS-CoV-2 M^{pro} inhibitor. Surpsingly, **167** was not able to bind covalently the Cys145 –SH group. Crystallographic studies revealed instead that this compound fits into the hydrophobic pocket of the dimerization site by means of its halogenated aromatic group (Figure 84B). (Günther et al. 2021) Its 3-cianoquinoline scaffold interacts with the Ser301 residue, while the ethyl ether substituent interacts with the Tyr118 and Asn142 residues of the opposite monomer. **167** showed anti-SARS-CoV-2 activity at low micromolar level [45].

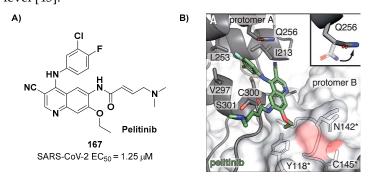


Figure 84. A) Chemical structure and biological activity of compound **167**; B) X-ray crystal structure of SARS-CoV-2 M^{pro} in complex with **167** (PDB: 7AXM).

Similarly to **164**, **Ifenoprodil** and **RS-102895** (compounds **168** and **169** in Figure 85) bind the same hydropobick pocket, showing moderate antiviral activity in VeroE6 cells (EC₅₀ = 47 μ M and CC₅₀ > 100 μ M for **168**; EC₅₀ = 19.8 μ M and CC₅₀ = 55 μ M for **169**) [45].

Figure 85. Chemical structure and biological activity of compounds 168 and 169.

In the same study, Günther and co-workers identified compound **170**, called **AT7519**, binding a different region located in a deep grove between the catalytic and dimerization domains (Figure 86A) [45]. The halogenated aromatic ring of **170** establishes Van der Walls interactions with several hydrophobic residues, while the pyrazole ring and the external piperidine ring forms important H-bonds with Gln110 and Asp153, respectively (Figure 86B). Ther H-bonding with Asp153 causes a shift of loop153-155, allowing an additional salt bridge between Asp153 and Arg298; this last residue has been found to be crucial for the dimerization process by mutational experiments, explaining the moderate antiviral activity of **170** (EC50 = 25.2 μ M, CC50 > 100 μ M) [45].

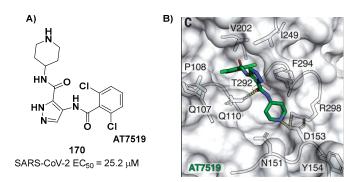


Figure 86. A) Chemical structure and biological activity of compound **170**; B) X-ray crystal structure of SARS-CoV-2 M^{pro} in complex with **170** (PDB: 7AGA).

From a FRET-based enzymatic screening of **Niclosamide** derivatives, Samrat *et al.* identified compounds **171-173** (Figure 87), called **JMX0286**, **JMX0301** and **JMX0941**, as potent inhibitor of SARS-CoV-2 M^{pro} (IC50 values 3.9–4.8 μ M) [116]. After evaluation of their antiviral activity (EC50 = 1.7–2.3 μ M) and cytotoxicity (CC50 = 10.6–53.1 μ M), kinetic experiments highlighted a non-competitive inhibition mechanism, while computational studies suggest that these compounds occupy the same allosteric site of **170** [116].

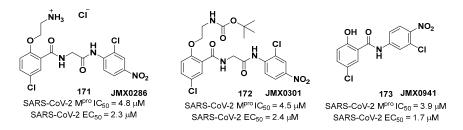


Figure 87. Chemical structure and biological activity of compounds 171-173.

Due to the similarity between the active sites of SARS-CoV-2 M^{pro}, FXa and thrombin, Chaves *et al.* tested different FXa and thrombin inhibitors against SARS-CoV-2 M^{pro} [117]. Among several anticoagulants, **Apixaban** (**174** in Figure 88) turned out to be the most potent compound, with a K_i value of 9.7 nM. From kinetic experiments, Chaves and collaborators observed a non-competitive mechanism of inhibition, due to the fact that that V_{max} decreased in presence of **174**, but the value of

the K_m was not affected. Among different hypothesis, the authors observed by molecular docking that 174 could bind the dimerization domain at allosteric site 1. The antiviral activity test performed on SARS-CoV-2 Calu-3 infected cells demonstrated that 174 showed important antiviral properties, with an EC₅₀ value of 1.8 μ M [117].

OMe
$$H_2N$$

$$N-N$$

$$O$$

$$N$$

$$Apixaban$$

$$SARS-CoV-2 M^{pro} K_i = 9.7 \text{ nM}$$

$$SARS-CoV-2 EC_{50} = 1.8 \mu\text{M}$$

Figure 88. Chemical structure and biological activity of compound 174.

14. Conclusions and future perspectives

The COVID-19 pandemic has proved to be an unexpected and unprecedented health emergency for the entire world population and has highlighted the lack of effective and available antiviral therapies to treat infected people. The SARS-CoV-2 virus has afflicted billions of humans worldwide in different aspects of life and, compared to the previous strains of the genus β-CoV MERS and SARS, it has been shown to possess significantly higher transmissibility with serious implications for the health of the most fragile patients. The lack of effective emergency treatments and the shortage of life-saving drugs prompted scientists and industrial experts to focus important efforts in the development of effective antiviral treatments. In this context, the Main Proteases emerged as the most appealing targets in the development of anticoronavirus drugs and this led to the successful marketing of Nirmatrelvir, the first oral drug to treat sever forms of COVID-19. Considering the recurrence of coronavirus infections in human history, the search for therapies directed against future pandemics should not be stopped, and this fact again places M^{pro} as the target of choice due to its low mutability among coronaviruses. Modern drug discovery aims to identify broad spectrum antiviral compounds capable of acting on a wide range of already known coronaviruses (alpha and beta genera) by acting on the preserved M^{pro}, in order to prevent future developments of pandemics. Another interesting approach that was highlighted during the discussion of the Review consists in the dual mechanism of action of some inhibitors capable of targeting both viral Mpro and host cathepsin L, a promising strategy that has already provided interesting results in the panorama of antiviral inhibitors. Taking a critical look at the future of SARS-CoV-2 Mpro inhibitors, we hope that research on this front remains a thriving field for preventing the spread of pandemic outbreaks caused by new coronaviruses in the future.

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References

 von Delft, A.; Hall, M.D.; Kwong, A.D.; Purcell, L.A.; Saikatendu, K.S.; Schmitz, U.; Tallarico, J.A.; Lee, A.A. Accelerating Antiviral Drug Discovery: Lessons from COVID-19. *Nat. Rev. Drug Discov.* 2023, 22, 585–603, doi:10.1038/s41573-023-00692-8.

- 2. Citarella, A.; Scala, A.; Piperno, A.; Micale, N. SARS-CoV-2 Mpro: A Potential Target for Peptidomimetics and Small-Molecule Inhibitors. *Biomolecules* **2021**, *11*, 607, doi:10.3390/biom11040607.
- 3. Mahase, E. Covid-19: Pfizer's Paxlovid Is 89% Effective in Patients at Risk of Serious Illness, Company Reports. *BMJ* **2021**, n2713, doi:10.1136/bmj.n2713.
- 4. Capasso, C.; Nocentini, A.; Supuran, C.T. Protease Inhibitors Targeting the Main Protease and Papain-like Protease of Coronaviruses. *Expert Opin. Ther. Pat.* **2021**, *31*, 309–324, doi:10.1080/13543776.2021.1857726.
- 5. Hilgenfeld, R. From SARS to MERS: Crystallographic Studies on Coronaviral Proteases Enable Antiviral Drug Design. *FEBS Journal* **2014**, *281*, 4085–4096, doi:10.1111/febs.12936.
- Zhang, L.; Zhao, H.; Liu, J.; He, L.; Yu, R.; Kang, C. Design of SARS-CoV-2 Mpro, PLpro Dual-Target Inhibitors Based on Deep Reinforcement Learning and Virtual Screening. *Future Med. Chem.* 2022, 14, 393–405, doi:10.4155/fmc-2021-0269.
- 7. Goyal, B.; Goyal, D. Targeting the Dimerization of the Main Protease of Coronaviruses: A Potential Broad-Spectrum Therapeutic Strategy. *ACS Comb. Sci.* **2020**, *22*, 297–305, doi:10.1021/acscombsci.0c00058.
- 8. Liu, Y.; Liang, C.; Xin, L.; Ren, X.; Tian, L.; Ju, X.; Li, H.; Wang, Y.; Zhao, Q.; Liu, H.; et al. The Development of Coronavirus 3C-Like Protease (3CLpro) Inhibitors from 2010 to 2020. Eur. J. Med. Chem. 2020, 206, 112711, doi:10.1016/j.ejmech.2020.112711.
- Jin, Z.; Du, X.; Xu, Y.; Deng, Y.; Liu, M.; Zhao, Y.; Zhang, B.; Li, X.; Zhang, L.; Peng, C.; et al. Structure of Mpro from SARS-CoV-2 and Discovery of Its Inhibitors. *Nature* 2020, 582, 289–293, doi:10.1038/s41586-020-2223-y.
- Hu, Q.; Xiong, Y.; Zhu, G.; Zhang, Y.; Zhang, Y.; Huang, P.; Ge, G. The SARS-CoV-2 Main Protease (M pro
 Structure, Function, and Emerging Therapies for COVID-19. MedComm (Beijing) 2022, 3, doi:10.1002/mco2.151.
- 11. RCSB PDB 7ALH: Crystal Structure of the Main Protease (3CLpro/Mpro) of SARS-CoV-2 at 1.65A Resolution (Spacegroup C2). Available online: https://www.rcsb.org/structure/7ALH (accessed on 28 February 2023).
- 12. Zhang, L.; Lin, D.; Sun, X.; Curth, U.; Drosten, C.; Sauerhering, L.; Becker, S.; Rox, K.; Hilgenfeld, R. Crystal Structure of SARS-CoV-2 Main Protease Provides a Basis for Design of Improved α -Ketoamide Inhibitors. *Science* (1979) **2020**, 368, 409–412, doi:10.1126/science.abb3405.
- 13. RCSB PDB 1P9S: Coronavirus Main Proteinase (3CLpro) Structure: Basis for Design of Anti-SARS Drugs Available online: https://www.rcsb.org/structure/1P9S (accessed on 28 February 2023).
- 14. Kneller, D.W.; Phillips, G.; Weiss, K.L.; Pant, S.; Zhang, Q.; O'Neill, H.M.; Coates, L.; Kovalevsky, A. Unusual Zwitterionic Catalytic Site of SARS–CoV-2 Main Protease Revealed by Neutron Crystallography. *J. Biol. Chem.* **2020**, 295, 17365–17373, doi:10.1074/jbc.AC120.016154.
- 15. Greasley, S.E.; Noell, S.; Plotnikova, O.; Ferre, R.; Liu, W.; Bolanos, B.; Fennell, K.; Nicki, J.; Craig, T.; Zhu, Y.; et al. Structural Basis for the in Vitro Efficacy of Nirmatrelvir against SARS-CoV-2 Variants. *J. Biol. Chem.* **2022**, 298, 101972, doi:10.1016/j.jbc.2022.101972.
- La Monica, G.; Bono, A.; Lauria, A.; Martorana, A. Targeting SARS-CoV-2 Main Protease for Treatment of COVID-19: Covalent Inhibitors Structure–Activity Relationship Insights and Evolution Perspectives. *J. Med. Chem.* 2022, 65, 12500–12534, doi:10.1021/acs.jmedchem.2c01005.
- 17. Rossetti, G.G.; Ossorio, M.A.; Rempel, S.; Kratzel, A.; Dionellis, V.S.; Barriot, S.; Tropia, L.; Gorgulla, C.; Arthanari, H.; Thiel, V.; et al. Non-Covalent SARS-CoV-2 Mpro Inhibitors Developed from in Silico Screen Hits. *Sci. Rep.* **2022**, *12*, 2505, doi:10.1038/s41598-022-06306-4.
- 18. Mangiavacchi, F.; Botwina, P.; Menichetti, E.; Bagnoli, L.; Rosati, O.; Marini, F.; Fonseca, S.F.; Abenante, L.; Alves, D.; Dabrowska, A.; et al. Seleno-Functionalization of Quercetin Improves the Non-Covalent Inhibition of Mpro and Its Antiviral Activity in Cells against SARS-CoV-2. *Int. J. Mol. Sci.* **2021**, *22*, 7048, doi:10.3390/ijms22137048.
- 19. Vagner, J.; Qu, H.; Hruby, V.J. Peptidomimetics, a Synthetic Tool of Drug Discovery. *Curr. Opin. Chem. Biol.* **2008**, *12*, 292–296, doi:10.1016/j.cbpa.2008.03.009.
- 20. Pillaiyar, T.; Manickam, M.; Namasivayam, V.; Hayashi, Y.; Jung, S.-H. An Overview of Severe Acute Respiratory Syndrome–Coronavirus (SARS-CoV) 3CL Protease Inhibitors: Peptidomimetics and Small Molecule Chemotherapy. *J. Med. Chem.* 2016, 59, 6595–6628, doi:10.1021/acs.jmedchem.5b01461.
- 21. Bzówka, M.; Mitusińska, K.; Raczyńska, A.; Samol, A.; Tuszyński, J.A.; Góra, A. Structural and Evolutionary Analysis Indicate That the SARS-CoV-2 Mpro Is a Challenging Target for Small-Molecule Inhibitor Design. *Int. J. Mol. Sci.* **2020**, *21*, 3099, doi:10.3390/ijms21093099.

- 22. Fischer, C.; Feys, J.R. SARS-CoV-2 Mpro Inhibitors: Achieved Diversity, Developing Resistance and Future Strategies. *Future Pharmacol.* **2023**, *3*, 80–107, doi:10.3390/futurepharmacol3010006.
- 23. Chuck, C.-P.; Chen, C.; Ke, Z.; Chi-Cheong Wan, D.; Chow, H.-F.; Wong, K.-B. Design, Synthesis and Crystallographic Analysis of Nitrile-Based Broad-Spectrum Peptidomimetic Inhibitors for Coronavirus 3C-like Proteases. *Eur. J. Med. Chem.* **2013**, *59*, 1–6, doi:10.1016/j.ejmech.2012.10.053.
- 24. Halford, B. The Path to Paxlovid. ACS Cent. Sci. 2022, 8, 405–407, doi:10.1021/acscentsci.2c00369.
- 25. Owen, D.R.; Allerton, C.M.N.; Anderson, A.S.; Aschenbrenner, L.; Avery, M.; Berritt, S.; Boras, B.; Cardin, R.D.; Carlo, A.; Coffman, K.J.; et al. An Oral SARS-CoV-2 M pro Inhibitor Clinical Candidate for the Treatment of COVID-19. *Science* (1979) **2021**, 374, 1586–1593, doi:10.1126/science.abl4784.
- 26. Palace Welcomes FDA Approval of COVID Generic Drug | Inquirer News Available online: https://newsinfo.inquirer.net/1539269/palace-welcomes-fda-ok-of-covid-generic-drug (accessed on 5 March 2023).
- 27. EMA Receives Application for Conditional Marketing Authorisation for Paxlovid (PF-07321332 and Ritonavir) for Treating Patients with COVID-19 | European Medicines Agency Available online: https://www.ema.europa.eu/en/news/ema-receives-application-conditional-marketing-authorisation-paxlovid-pf-07321332-ritonavir-treating (accessed on 5 March 2023).
- 28. Oral COVID-19 Antiviral, Paxlovid, Approved by UK Regulator GOV.UK Available online: https://www.gov.uk/government/news/oral-covid-19-antiviral-paxlovid-approved-by-uk-regulator (accessed on 5 March 2023).
- 29. WO2021250648A1 Nitrile-Containing Antiviral Compounds Google Patents Available online: https://patents.google.com/patent/WO2021250648A1/en (accessed on 5 March 2023).
- 30. Bai, B.; Arutyunova, E.; Khan, M.B.; Lu, J.; Joyce, M.A.; Saffran, H.A.; Shields, J.A.; Kandadai, A.S.; Belovodskiy, A.; Hena, M.; et al. Peptidomimetic Nitrile Warheads as SARS-CoV-2 3CL Protease Inhibitors. *RSC Med. Chem.* **2021**, *12*, 1722–1730, doi:10.1039/D1MD00247C.
- 31. Zhu, M.; Fu, T.; You, M.; Cao, J.; Yang, H.; Chen, X.; Zhang, Q.; Xu, Y.; Jiang, X.; Zhang, L.; et al. Design, Synthesis and Biological Evaluation of Covalent Peptidomimetic 3CL Protease Inhibitors Containing Nitrile Moiety. *Bioorg. Med. Chem.* **2023**, *87*, 117316, doi:10.1016/J.BMC.2023.117316.
- 32. Kneller, D.W.; Li, H.; Phillips, G.; Weiss, K.L.; Zhang, Q.; Arnould, M.A.; Jonsson, C.B.; Surendranathan, S.; Parvathareddy, J.; Blakeley, M.P.; et al. Covalent Narlaprevir- and Boceprevir-Derived Hybrid Inhibitors of SARS-CoV-2 Main Protease. *Nat. Commun.* **2022**, *13*, 2268, doi:10.1038/s41467-022-29915-z.
- 33. Brewitz, L.; Dumjahn, L.; Zhao, Y.; Owen, C.D.; Laidlaw, S.M.; Malla, T.R.; Nguyen, D.; Lukacik, P.; Salah, E.; Crawshaw, A.D.; et al. Alkyne Derivatives of SARS-CoV-2 Main Protease Inhibitors Including Nirmatrelvir Inhibit by Reacting Covalently with the Nucleophilic Cysteine. *J. Med. Chem.* **2023**, *66*, 2663–2680, doi:10.1021/acs.jmedchem.2c01627.
- 34. Yang, H.; Yang, M.; Ding, Y.; Liu, Y.; Lou, Z.; Zhou, Z.; Sun, L.; Mo, L.; Ye, S.; Pang, H.; et al. The Crystal Structures of Severe Acute Respiratory Syndrome Virus Main Protease and Its Complex with an Inhibitor. *PNAS* 2003, *100*, 13190–13195, doi:10.1073/pnas.1835675100.
- 35. Dai, W.; Zhang, B.; Jiang, X.-M.; Su, H.; Li, J.; Zhao, Y.; Xie, X.; Jin, Z.; Peng, J.; Liu, F.; et al. Structure-Based Design of Antiviral Drug Candidates Targeting the SARS-CoV-2 Main Protease. *Science* (1979) **2020**, 368, 1331–1335, doi:10.1126/science.abb4489.
- 36. Dai, W.; Jochmans, D.; Xie, H.; Yang, H.; Li, J.; Su, H.; Chang, D.; Wang, J.; Peng, J.; Zhu, L.; et al. Design, Synthesis, and Biological Evaluation of Peptidomimetic Aldehydes as Broad-Spectrum Inhibitors against Enterovirus and SARS-CoV-2. *J. Med. Chem.* 2022, 65, 2794–2808, doi:10.1021/acs.jmedchem.0c02258.
- 37. Vuong, W.; Khan, M.B.; Fischer, C.; Arutyunova, E.; Lamer, T.; Shields, J.; Saffran, H.A.; McKay, R.T.; van Belkum, M.J.; Joyce, M.A.; et al. Feline Coronavirus Drug Inhibits the Main Protease of SARS-CoV-2 and Blocks Virus Replication. *Nat. Commun.* **2020**, *11*, 4282, doi:10.1038/s41467-020-18096-2.
- 38. Vuong, W.; Fischer, C.; Khan, M.B.; van Belkum, M.J.; Lamer, T.; Willoughby, K.D.; Lu, J.; Arutyunova, E.; Joyce, M.A.; Saffran, H.A.; et al. Improved SARS-CoV-2 Mpro Inhibitors Based on Feline Antiviral Drug GC376: Structural Enhancements, Increased Solubility, and Micellar Studies. *Eur. J. Med. Chem.* **2021**, 222, 113584, doi:10.1016/j.ejmech.2021.113584.
- 39. Sacco, M.D.; Ma, C.; Lagarias, P.; Gao, A.; Townsend, J.A.; Meng, X.; Dube, P.; Zhang, X.; Hu, Y.; Kitamura, N.; et al. Structure and Inhibition of the SARS-CoV-2 Main Protease Reveal Strategy for Developing Dual Inhibitors against M pro and Cathepsin L. *Sci. Adv.* **2020**, *6*, doi:10.1126/sciadv.abe0751.

- 40. Liu, H.; Iketani, S.; Zask, A.; Khanizeman, N.; Bednarova, E.; Forouhar, F.; Fowler, B.; Hong, S.J.; Mohri, H.; Nair, M.S.; et al. Development of Optimized Drug-like Small Molecule Inhibitors of the SARS-CoV-2 3CL Protease for Treatment of COVID-19. *Nat. Commun.* **2022**, *13*, 1891, doi:10.1038/s41467-022-29413-2.
- 41. Qiao, J.; Li, Y.-S.; Zeng, R.; Liu, F.-L.; Luo, R.-H.; Huang, C.; Wang, Y.-F.; Zhang, J.; Quan, B.; Shen, C.; et al. SARS-CoV-2 M pro Inhibitors with Antiviral Activity in a Transgenic Mouse Model. *Science* (1979) **2021**, 371, 1374–1378, doi:10.1126/science.abf1611.
- 42. Dampalla, C.S.; Kim, Y.; Bickmeier, N.; Rathnayake, A.D.; Nguyen, H.N.; Zheng, J.; Kashipathy, M.M.; Baird, M.A.; Battaile, K.P.; Lovell, S.; et al. Structure-Guided Design of Conformationally Constrained Cyclohexane Inhibitors of Severe Acute Respiratory Syndrome Coronavirus-2 3CL Protease. *J. Med. Chem.* 2021, 64, 10047–10058, doi:10.1021/acs.jmedchem.1c00319.
- 43. Xia, Z.; Sacco, M.; Hu, Y.; Ma, C.; Meng, X.; Zhang, F.; Szeto, T.; Xiang, Y.; Chen, Y.; Wang, J. Rational Design of Hybrid SARS-CoV-2 Main Protease Inhibitors Guided by the Superimposed Cocrystal Structures with the Peptidomimetic Inhibitors GC-376, Telaprevir, and Boceprevir. *ACS Pharmacol. Transl. Sci.* **2021**, *4*, 1408–1421, doi:10.1021/acsptsci.1c00099.
- 44. Alugubelli, Y.R.; Geng, Z.Z.; Yang, K.S.; Shaabani, N.; Khatua, K.; Ma, X.R.; Vatansever, E.C.; Cho, C.-C.; Ma, Y.; Xiao, J.; et al. A Systematic Exploration of Boceprevir-Based Main Protease Inhibitors as SARS-CoV-2 Antivirals. *Eur. J. Med. Chem.* **2022**, *240*, 114596, doi:10.1016/j.ejmech.2022.114596.
- 45. Günther, S.; Reinke, P.Y.A.; Fernández-García, Y.; Lieske, J.; Lane, T.J.; Ginn, H.M.; Koua, F.H.M.; Ehrt, C.; Ewert, W.; Oberthuer, D.; et al. X-Ray Screening Identifies Active Site and Allosteric Inhibitors of SARS-CoV-2 Main Protease. *Science* (1979) **2021**, 372, 642–646, doi:10.1126/science.abf7945.
- 46. Hu, Y.; Ma, C.; Szeto, T.; Hurst, B.; Tarbet, B.; Wang, J. Boceprevir, Calpain Inhibitors II and XII, and GC-376 Have Broad-Spectrum Antiviral Activity against Coronaviruses. *ACS Infect. Dis.* **2021**, *7*, 586–597, doi:10.1021/acsinfecdis.0c00761.
- 47. Yang, K.S.; Ma, X.R.; Ma, Y.; Alugubelli, Y.R.; Scott, D.A.; Vatansever, E.C.; Drelich, A.K.; Sankaran, B.; Geng, Z.Z.; Blankenship, L.R.; et al. A Quick Route to Multiple Highly Potent SARS-CoV-2 Main Protease Inhibitors. *ChemMedChem* **2021**, *16*, 942–948, doi:10.1002/cmdc.202000924.
- 48. Costanzi, E.; Kuzikov, M.; Esposito, F.; Albani, S.; Demitri, N.; Giabbai, B.; Camasta, M.; Tramontano, E.; Rossetti, G.; Zaliani, A.; et al. Structural and Biochemical Analysis of the Dual Inhibition of MG-132 against SARS-CoV-2 Main Protease (Mpro/3CLpro) and Human Cathepsin-L. *Int. J. Mol. Sci.* **2021**, 22, 11779, doi:10.3390/ijms222111779.
- Ma, Y.; Yang, K.S.; Geng, Z.Z.; Alugubelli, Y.R.; Shaabani, N.; Vatansever, E.C.; Ma, X.R.; Cho, C.-C.; Khatua, K.; Xiao, J.; et al. A Multi-Pronged Evaluation of Aldehyde-Based Tripeptidyl Main Protease Inhibitors as SARS-CoV-2 Antivirals. Eur. J. Med. Chem. 2022, 240, 114570, doi:10.1016/j.ejmech.2022.114570.
- 50. Hoffman, R.L.; Kania, R.S.; Brothers, M.A.; Davies, J.F.; Ferre, R.A.; Gajiwala, K.S.; He, M.; Hogan, R.J.; Kozminski, K.; Li, L.Y.; et al. Discovery of Ketone-Based Covalent Inhibitors of Coronavirus 3CL Proteases for the Potential Therapeutic Treatment of COVID-19. *J. Med. Chem.* **2020**, *63*, 12725–12747, doi:10.1021/acs.jmedchem.0c01063.
- 51. Bai, B.; Belovodskiy, A.; Hena, M.; Kandadai, A.S.; Joyce, M.A.; Saffran, H.A.; Shields, J.A.; Khan, M.B.; Arutyunova, E.; Lu, J.; et al. Peptidomimetic α-Acyloxymethylketone Warheads with Six-Membered Lactam P1 Glutamine Mimic: SARS-CoV-2 3CL Protease Inhibition, Coronavirus Antiviral Activity, and *in Vitro* Biological Stability. *J. Med. Chem.* **2022**, *65*, 2905–2925, doi:10.1021/acs.jmedchem.1c00616.
- 52. Krantz, A.; Copp, L.J.; Coles, P.J.; Smith, R.A.; Heard, S.B. Peptidyl (Acyloxy)Methyl Ketones and the Quiescent Affinity Label Concept: The Departing Group as a Variable Structural Element in the Design of Inactivators of Cysteine Proteinases. *Biochemistry* 1991, 30, 4678–4687, doi:10.1021/bi00233a007.
- 53. Hattori, S.; Higashi-Kuwata, N.; Hayashi, H.; Allu, S.R.; Raghavaiah, J.; Bulut, H.; Das, D.; Anson, B.J.; Lendy, E.K.; Takamatsu, Y.; et al. A Small Molecule Compound with an Indole Moiety Inhibits the Main Protease of SARS-CoV-2 and Blocks Virus Replication. *Nat. Commun.* **2021**, *12*, 668, doi:10.1038/s41467-021-20900-6.
- 54. Thanigaimalai, P.; Konno, S.; Yamamoto, T.; Koiwai, Y.; Taguchi, A.; Takayama, K.; Yakushiji, F.; Akaji, K.; Chen, S.-E.; Naser-Tavakolian, A.; et al. Development of Potent Dipeptide-Type SARS-CoV 3CL Protease Inhibitors with Novel P3 Scaffolds: Design, Synthesis, Biological Evaluation, and Docking Studies. *Eur. J. Med. Chem.* 2013, *68*, 372–384, doi:10.1016/j.ejmech.2013.07.037.
- 55. Higashi-Kuwata, N.; Tsuji, K.; Hayashi, H.; Bulut, H.; Kiso, M.; Imai, M.; Ogata-Aoki, H.; Ishii, T.; Kobayakawa, T.; Nakano, K.; et al. Identification of SARS-CoV-2 Mpro Inhibitors Containing P1' 4-

- Fluorobenzothiazole Moiety Highly Active against SARS-CoV-2. *Nat. Commun.* **2023**, *14*, 1076, doi:10.1038/s41467-023-36729-0.
- 56. Yang, H.; You, M.; Shu, X.; Zhen, J.; Zhu, M.; Fu, T.; Zhang, Y.; Jiang, X.; Zhang, L.; Xu, Y.; et al. Design, Synthesis and Biological Evaluation of Peptidomimetic Benzothiazolyl Ketones as 3CLpro Inhibitors against SARS-CoV-2. *Eur. J. Med. Chem.* **2023**, 257, 115512, doi:10.1016/J.EJMECH.2023.115512.
- 57. Citarella, A.; Micale, N. Peptidyl Fluoromethyl Ketones and Their Applications in Medicinal Chemistry. *Molecules* **2020**, *25*, 4031, doi:10.3390/molecules25174031.
- 58. Zhu, W.; Xu, M.; Chen, C.Z.; Guo, H.; Shen, M.; Hu, X.; Shinn, P.; Klumpp-Thomas, C.; Michael, S.G.; Zheng, W. Identification of SARS-CoV-2 3CL Protease Inhibitors by a Quantitative High-Throughput Screening. *ACS Pharmacol. Transl. Sci.* 2020, *3*, 1008–1016, doi:10.1021/acsptsci.0c00108.
- 59. Shao, Y.M.; Yang, W. Bin; Kuo, T.H.; Tsai, K.C.; Lin, C.H.; Yang, A.S.; Liang, P.H.; Wong, C.H. Design, Synthesis, and Evaluation of Trifluoromethyl Ketones as Inhibitors of SARS-CoV 3CL Protease. *Bioorg. Med. Chem.* 2008, *16*, 4652–4660, doi:10.1016/J.BMC.2008.02.040.
- Citarella, A.; Gentile, D.; Rescifina, A.; Piperno, A.; Mognetti, B.; Gribaudo, G.; Sciortino, M.T.; Holzer, W.;
 Pace, V.; Micale, N. Pseudo-Dipeptide Bearing α,α-Difluoromethyl Ketone Moiety as Electrophilic
 Warhead with Activity against Coronaviruses. *Int. J. Mol. Sci.* 2021, 22, 1398, doi:10.3390/IJMS22031398.
- 61. Citarella, A.; Ielo, L.; Stagno, C.; Cristani, M.; Muscarà, C.; Pace, V.; Micale, N. Synthesis, Computational Investigation and Biological Evaluation of α , α -Difluoromethyl Ketones Embodying Pyrazole and Isoxazole Nuclei as COX Inhibitors. *Org. Biomol. Chem.* **2022**, *20*, 8293–8304, doi:10.1039/D2OB01382G.
- 62. Miele, M.; Citarella, A.; Micale, N.; Holzer, W.; Pace, V. Direct and Chemoselective Synthesis of Tertiary Difluoroketones via Weinreb Amide Homologation with a CHF 2 -Carbene Equivalent. *Org. Lett.* **2019**, *21*, 8261–8265, doi:10.1021/acs.orglett.9b03024.
- 63. Shcherbakov, D.; Baev, D.; Kalinin, M.; Dalinger, A.; Chirkova, V.; Belenkaya, S.; Khvostov, A.; Krut'ko, D.; Medved'ko, A.; Volosnikova, E.; et al. Design and Evaluation of Bispidine-Based SARS-CoV-2 Main Protease Inhibitors. *ACS Med. Chem. Lett.* **2022**, *13*, 140–147, doi:10.1021/acsmedchemlett.1c00299.
- 64. Stille, J.K.; Tjutrins, J.; Wang, G.; Venegas, F.A.; Hennecker, C.; Rueda, A.M.; Sharon, I.; Blaine, N.; Miron, C.E.; Pinus, S.; et al. Design, Synthesis and in Vitro Evaluation of Novel SARS-CoV-2 3CLpro Covalent Inhibitors. *Eur. J. Med. Chem.* **2022**, 229, 114046, doi:10.1016/J.EJMECH.2021.114046.
- 65. Ma, C.; Xia, Z.; Sacco, M.D.; Hu, Y.; Townsend, J.A.; Meng, X.; Choza, J.; Tan, H.; Jang, J.; Gongora, M. V.; et al. Discovery of Di- and Trihaloacetamides as Covalent SARS-CoV-2 Main Protease Inhibitors with High Target Specificity. *JACS* **2021**, *143*, 20697–20709, doi:10.1021/jacs.1c08060.
- 66. Yamane, D.; Onitsuka, S.; Re, S.; Isogai, H.; Hamada, R.; Hiramoto, T.; Kawanishi, E.; Mizuguchi, K.; Shindo, N.; Ojida, A. Selective Covalent Targeting of SARS-CoV-2 Main Protease by Enantiopure Chlorofluoroacetamide. *Chem. Sci.* **2022**, *13*, 3027–3034, doi:10.1039/D1SC06596C.
- 67. Shindo, N.; Fuchida, H.; Sato, M.; Watari, K.; Shibata, T.; Kuwata, K.; Miura, C.; Okamoto, K.; Hatsuyama, Y.; Tokunaga, K.; et al. Selective and Reversible Modification of Kinase Cysteines with Chlorofluoroacetamides. *Nat. Chem. Biol.* **2019**, *15*, 250–258, doi:10.1038/s41589-018-0204-3.
- 68. Xiong, M.; Nie, T.; Shao, Q.; Li, M.; Su, H.; Xu, Y. In Silico Screening-Based Discovery of Novel Covalent Inhibitors of the SARS-CoV-2 3CL Protease. *Eur. J. Med. Chem.* **2022**, 231, 114130, doi:10.1016/j.ejmech.2022.114130.
- 69. Clyde, A.; Galanie, S.; Kneller, D.W.; Ma, H.; Babuji, Y.; Blaiszik, B.; Brace, A.; Brettin, T.; Chard, K.; Chard, R.; et al. High-Throughput Virtual Screening and Validation of a SARS-CoV-2 Main Protease Noncovalent Inhibitor. J. Chem. Inf. Model. 2022, 62, 116–128, doi:10.1021/ACS.JCIM.1C00851/ASSET/IMAGES/LARGE/CI1C00851_0005.JPEG.
- Gao, S.; Sylvester, K.; Song, L.; Claff, T.; Jing, L.; Woodson, M.; Weiße, R.H.; Cheng, Y.; Schäkel, L.; Petry, M.; et al. Discovery and Crystallographic Studies of Trisubstituted Piperazine Derivatives as Non-Covalent SARS-CoV-2 Main Protease Inhibitors with High Target Specificity and Low Toxicity. *J. Med. Chem.* 2022, 65, 13343–13364, doi:10.1021/ACS.JMEDCHEM.2C01146/SUPPL_FILE/JM2C01146_SI_002.CSV.
- Gao, S.; Song, L.; Claff, T.; Woodson, M.; Sylvester, K.; Jing, L.; Weiße, R.H.; Cheng, Y.; Sträter, N.; Schäkel, L.; et al. Discovery and Crystallographic Studies of Nonpeptidic Piperazine Derivatives as Covalent SARS-CoV-2 Main Protease Inhibitors. *J. Med. Chem.* 2022, 65, 16902–16917, doi:10.1021/ACS.JMEDCHEM.2C01716/SUPPL_FILE/JM2C01716_SI_001.CSV.

- 72. Moon, P.; Boike, L.; Dovala, D.; Henning, N.J.; Knapp, M.; Spradlin, J.N.; Ward, C.C.; Wolleb, H.; Zammit, C.M.; Fuller, D.; et al. Discovery of Potent Pyrazoline-Based Covalent SARS-CoV-2 Main Protease Inhibitors. *bioRxiv* 2022, 2022.03.05.483025, doi:10.1101/2022.03.05.483025.
- 73. El Khoury, L.; Jing, Z.; Cuzzolin, A.; Deplano, A.; Loco, D.; Sattarov, B.; Hédin, F.; Wendeborn, S.; Ho, C.; El Ahdab, D.; et al. Computationally Driven Discovery of SARS-CoV-2 M pro Inhibitors: From Design to Experimental Validation. *Chem. Sci.* **2022**, *13*, 3674–3687, doi:10.1039/D1SC05892D.
- 74. Zhang, L.; Lin, D.; Kusov, Y.; Nian, Y.; Ma, Q.; Wang, J.; Von Brunn, A.; Leyssen, P.; Lanko, K.; Neyts, J.; et al. α-Ketoamides as Broad-Spectrum Inhibitors of Coronavirus and Enterovirus Replication: Structure-Based Design, Synthesis, and Activity Assessment. *J. Med. Chem.* **2020**, *63*, 4562–4578, doi:10.1021/ACS.JMEDCHEM.9B01828/SUPPL_FILE/JM9B01828_SI_002.CSV.
- 75. Zhang, L.; Lin, D.; Sun, X.; Curth, U.; Drosten, C.; Sauerhering, L.; Becker, S.; Rox, K.; Hilgenfeld, R. Crystal Structure of SARS-CoV-2 Main Protease Provides a Basis for Design of Improved α -Ketoamide Inhibitors. *Science* (1979) **2020**, 368, 409–412, doi:10.1126/science.abb3405.
- 76. Cooper, M.S.; Zhang, L.; Ibrahim, M.; Zhang, K.; Sun, X.; Röske, J.; Göhl, M.; Brönstrup, M.; Cowell, J.K.; Sauerhering, L.; et al. Diastereomeric Resolution Yields Highly Potent Inhibitor of SARS-CoV-2 Main Protease. *J. Med. Chem.* **2022**, 65, 13328–13342, doi:10.1021/ACS.JMEDCHEM.2C01131/SUPPL_FILE/JM2C01131_SI_001.CSV.
- 77. Quan, B.X.; Shuai, H.; Xia, A.J.; Hou, Y.; Zeng, R.; Liu, X.L.; Lin, G.F.; Qiao, J.X.; Li, W.P.; Wang, F.L.; et al. An Orally Available Mpro Inhibitor Is Effective against Wild-Type SARS-CoV-2 and Variants Including Omicron. *Nat. Microbiol.* 2022 7:5 2022, 7, 716–725, doi:10.1038/s41564-022-01119-7.
- 78. Chen, X.; Huang, X.; Ma, Q.; Kuzmič, P.; Zhou, B.; Xu, J.; Liu, B.; Jiang, H.; Zhang, W.; Yang, C.; et al. Inhibition Mechanism and Antiviral Activity of an α-Ketoamide Based SARS-CoV-2 Main Protease Inhibitor. *bioRxiv* **2023**, 2023.03.09.531862, doi:10.1101/2023.03.09.531862.
- 79. Huang, C.; Shuai, H.; Qiao, J.; Hou, Y.; Zeng, R.; Xia, A.; Xie, L.; Fang, Z.; Li, Y.; Yoon, C.; et al. A New Generation Mpro Inhibitor with Potent Activity against SARS-CoV-2 Omicron Variants. *Signal Transduct. Target. Ther.* **2023**, *8*, 1–13, doi:10.1038/s41392-023-01392-w.
- 80. Jin, Z.; Du, X.; Xu, Y.; Deng, Y.; Liu, M.; Zhao, Y.; Zhang, B.; Li, X.; Zhang, L.; Peng, C.; et al. Structure of Mpro from SARS-CoV-2 and Discovery of Its Inhibitors. *Nature* **2020**, *582*, 289–293, doi:10.1038/s41586-020-2223-v.
- 81. Iketani, S.; Forouhar, F.; Liu, H.; Hong, S.J.; Lin, F.Y.; Nair, M.S.; Zask, A.; Huang, Y.; Xing, L.; Stockwell, B.R.; et al. Lead Compounds for the Development of SARS-CoV-2 3CL Protease Inhibitors. *Nat. Commun.* **2021**, *12*, 1–7, doi:10.1038/s41467-021-22362-2.
- 82. Amendola, G.; Ettari, R.; Previti, S.; Di Chio, C.; Messere, A.; Di Maro, S.; Hammerschmidt, S.J.; Zimmer, C.; Zimmermann, R.A.; Schirmeister, T.; et al. Lead Discovery of SARS-CoV-2 Main Protease Inhibitors through Covalent Docking-Based Virtual Screening. *J. Chem. Inf. Model.* **2021**, *61*, 2062–2073, doi:10.1021/acs.jcim.1c00184.
- 83. Previti, S.; Ettari, R.; Calcaterra, E.; Di Maro, S.; Hammerschmidt, S.J.; Müller, C.; Ziebuhr, J.; Schirmeister, T.; Cosconati, S.; Zappalà, M. Structure-Based Lead Optimization of Peptide-Based Vinyl Methyl Ketones as SARS-CoV-2 Main Protease Inhibitors. *Eur. J. Med. Chem.* **2023**, 247, 115021, doi:10.1016/j.ejmech.2022.115021.
- 84. Mondal, S.; Chen, Y.; Lockbaum, G.J.; Sen, S.; Chaudhuri, S.; Reyes, A.C.; Lee, J.M.; Kaur, A.N.; Sultana, N.; Cameron, M.D.; et al. Dual Inhibitors of Main Protease (M Pro) and Cathepsin L as Potent Antivirals against SARS-CoV2. *JACS* **2022**, *144*, 21035–21045, doi:10.1021/jacs.2c04626.
- 85. Citarella, A.; Moi, D.; Pedrini, M.; Pérez-Peña, H.; Pieraccini, S.; Dimasi, A.; Stagno, C.; Micale, N.; Schirmeister, T.; Sibille, G.; et al. Synthesis of SARS-CoV-2 Mpro Inhibitors Bearing a Cinnamic Ester Warhead with in Vitro Activity against Human Coronaviruses. *Org. Biomol. Chem.* **2023**, *21*, 3811-3824, doi:10.1039/D3OB00381G.
- 86. Zaidman, D.; Gehrtz, P.; Filep, M.; Walsh, M.A.; Von Delft, F.; London Correspondence, N. An Automatic Pipeline for the Design of Irreversible Derivatives Identifies a Potent SARS-CoV-2 M pro Inhibitor. *Cell. Chem. Biol.* **2021**, *28*, 1795-1806.e5, doi:10.1016/j.chembiol.2021.05.018.
- 87. Kuzikov, M.; Costanzi, E.; Reinshagen, J.; Esposito, F.; Vangeel, L.; Wolf, M.; Ellinger, B.; Claussen, C.; Geisslinger, G.; Corona, A.; et al. Identification of Inhibitors of SARS-CoV-2 3CL-Pro Enzymatic Activity Using a Small Molecule in Vitro Repurposing Screen. *ACS Pharmacol. Transl. Sci.* 2021, 4, 1096–1110, doi:10.1021/ACSPTSCI.0C00216/ASSET/IMAGES/LARGE/PT0C00216_0007.JPEG.

- 88. Su, H.; Yao, S.; Zhao, W.; Zhang, Y.; Liu, J.; Shao, Q.; Wang, Q.; Li, M.; Xie, H.; Shang, W.; et al. Identification of Pyrogallol as a Warhead in Design of Covalent Inhibitors for the SARS-CoV-2 3CL Protease. *Nat. Commun.* **2021**, *12*, 1–12, doi:10.1038/s41467-021-23751-3.
- 89. Ghosh, A.K.; Raghavaiah, J.; Shahabi, D.; Yadav, M.; Anson, B.J.; Lendy, E.K.; Hattori, S.I.; Higashi-Kuwata, N.; Mitsuya, H.; Mesecar, A.D. Indole Chloropyridinyl Ester-Derived SARS-CoV-2 3CLpro Inhibitors: Enzyme Inhibition, Antiviral Efficacy, Structure-Activity Relationship, and X-Ray Structural Studies. *J. Med. Chem.* 2021, 64, 14702–14714, doi:10.1021/ACS.JMEDCHEM.1C01214/SUPPL_FILE/JM1C01214_SI_002.CSV.
- 90. Hattori, S.I.; Higshi-Kuwata, N.; Raghavaiah, J.; Das, D.; Bulut, H.; Davis, D.A.; Takamatsu, Y.; Matsuda, K.; Takamune, N.; Kishimoto, N.; et al. GRL-0920, an Indole Chloropyridinyl Ester, Completely Blocks SARS-CoV-2 Infection. *mBio* 2020, 11, 1–16, doi:10.1128/MBIO.01833-20/ASSET/11D12B15-E56A-4381-A5F9-AD6A79F97BE2/ASSETS/GRAPHIC/MBIO.01833-20-F0005.JPEG.
- 91. Hattori, S. ichiro; Higashi-Kuwata, N.; Hayashi, H.; Allu, S.R.; Raghavaiah, J.; Bulut, H.; Das, D.; Anson, B.J.; Lendy, E.K.; Takamatsu, Y.; et al. A Small Molecule Compound with an Indole Moiety Inhibits the Main Protease of SARS-CoV-2 and Blocks Virus Replication. *Nat. Commun.* **2021**, *12*, 1–12, doi:10.1038/s41467-021-20900-6.
- 92. Breidenbach, J.; Lemke, C.; Pillaiyar, T.; Schäkel, L.; Al Hamwi, G.; Diett, M.; Gedschold, R.; Geiger, N.; Lopez, V.; Mirza, S.; et al. Targeting the Main Protease of SARS-CoV-2: From the Establishment of High Throughput Screening to the Design of Tailored Inhibitors. *Angew. Chem. Int. Ed.* **2021**, *60*, 10423–10429, doi:10.1002/ANIE.202016961.
- 93. Pillaiyar, T.; Flury, P.; Krüger, N.; Su, H.; Schäkel, L.; Barbosa Da Silva, E.; Eppler, O.; Kronenberger, T.; Nie, T.; Luedtke, S.; et al. Small-Molecule Thioesters as SARS-CoV-2 Main Protease Inhibitors: Enzyme Inhibition, Structure–Activity Relationships, Antiviral Activity, and X-Ray Structure Determination. *J. Med. Chem.* 2022, 65, 9376–9395, doi:10.1021/acs.jmedchem.2c00636.
- 94. Amporndanai, K.; Meng, X.; Shang, W.; Jin, Z.; Rogers, M.; Zhao, Y.; Rao, Z.; Liu, Z.J.; Yang, H.; Zhang, L.; et al. Inhibition Mechanism of SARS-CoV-2 Main Protease by Ebselen and Its Derivatives. *Nat. Commun.* **2021**, *12*, 1–7, doi:10.1038/s41467-021-23313-7.
- 95. Zmudzinski, M.; Rut, W.; Olech, K.; Granda, J.; Giurg, M.; Burda-Grabowska, M.; Zhang, L.; Sun, X.; Lv, Z.; Nayak, D.; et al. Ebselen Derivatives Are Very Potent Dual Inhibitors of SARS-CoV-2 Proteases PLpro and Mpro in in Vitro Studies. *bioRxiv* **2020**, 2020.08.30.273979, doi:10.1101/2020.08.30.273979.
- 96. Huff, S.; Kummetha, I.R.; Tiwari, S.K.; Huante, M.B.; Clark, A.E.; Wang, S.; Bray, W.; Smith, D.; Carlin, A.F.; Endsley, M.; et al. Discovery and Mechanism of SARS-CoV-2 Main Protease Inhibitors. *J. Med. Chem.* 2022, 65, 2866–2879, doi:10.1021/ACS.JMEDCHEM.1C00566/SUPPL_FILE/JM1C00566_SI_001.CSV.
- 97. Sun, L.Y.; Chen, C.; Su, J.; Li, J.Q.; Jiang, Z.; Gao, H.; Chigan, J.Z.; Ding, H.H.; Zhai, L.; Yang, K.W. Ebsulfur and Ebselen as Highly Potent Scaffolds for the Development of Potential SARS-CoV-2 Antivirals. *Bioorg. Chem.* **2021**, *112*, 104889, doi:10.1016/J.BIOORG.2021.104889.
- 98. Chen, W.; Feng, B.; Han, S.; Wang, P.; Chen, W.; Zang, Y.; Li, J.; Hu, Y. Discovery of Highly Potent SARS-CoV-2 Mpro Inhibitors Based on Benzoisothiazolone Scaffold. *Bioorg. Med. Chem. Lett.* **2022**, *58*, 128526, doi:10.1016/j.bmcl.2022.128526.
- 99. Aljoundi, A.; Bjij, I.; El Rashedy, A.; Soliman, M.E.S. Covalent Versus Non-Covalent Enzyme Inhibition: Which Route Should We Take? A Justification of the Good and Bad from Molecular Modelling Perspective. *Protein Journal* **2020**, *39*, 97–105, doi:10.1007/S10930-020-09884-2/FIGURES/5.
- 100. John, S. Broad-Spectrum Non-Covalent Coronavirus Protease Inhibitors. *Angew. Chem. Int. Ed* **2017**, 797, 3168–3210.
- 101. Kitamura, N.; Sacco, M.D.; Ma, C.; Hu, Y.; Townsend, J.A.; Meng, X.; Zhang, F.; Zhang, X.; Ba, M.; Szeto, T.; et al. Expedited Approach toward the Rational Design of Noncovalent SARS-CoV-2 Main Protease Inhibitors. J. Med. Chem. 2022, 65, 2848–2865, doi:10.1021/ACS.JMEDCHEM.1C00509/SUPPL_FILE/JM1C00509_SI_002.CSV.

- 102. Unoh, Y.; Uehara, S.; Nakahara, K.; Nobori, H.; Yamatsu, Y.; Yamamoto, S.; Maruyama, Y.; Taoda, Y.; Kasamatsu, K.; Suto, T.; et al. Discovery of S-217622, a Noncovalent Oral SARS-CoV-2 3CL Protease Inhibitor Clinical Candidate for Treating COVID-19. *J. Med. Chem.* 2022, 65, 6499–6512, doi:10.1021/acs.jmedchem.2c00117.
- 103. Yang, J.; Lin, X.; Xing, N.; Zhang, Z.; Zhang, H.; Wu, H.; Xue, W. Structure-Based Discovery of Novel Nonpeptide Inhibitors Targeting SARS-CoV-2 M ^{pro}. *J. Chem. Inf. Model.* **2021**, *61*, 3917–3926, doi:10.1021/acs.jcim.1c00355.
- 104. Han, S.H.; Goins, C.M.; Arya, T.; Shin, W.J.; Maw, J.; Hooper, A.; Sonawane, D.P.; Porter, M.R.; Bannister, B.E.; Crouch, R.D.; et al. Structure-Based Optimization of ML300-Derived, Noncovalent Inhibitors Targeting the Severe Acute Respiratory Syndrome Coronavirus 3CL Protease (SARS-CoV-2 3CLpro). *J. Med. Chem.* 2022, 65, 2880–2904, doi:10.1021/ACS.JMEDCHEM.1C00598/SUPPL_FILE/JM1C00598_SI_002.CSV.
- 105. Elseginy, S.A.; Fayed, B.; Hamdy, R.; Mahrous, N.; Mostafa, A.; Almehdi, A.M.; S. M. Soliman, S. Promising Anti-SARS-CoV-2 Drugs by Effective Dual Targeting against the Viral and Host Proteases. *Bioorg. Med. Chem. Lett.* **2021**, 43, 128099, doi:10.1016/J.BMCL.2021.128099.
- 106. Zhang, C.-H.; Stone, E.A.; Deshmukh, M.; Ippolito, J.A.; Ghahremanpour, M.M.; Tirado-Rives, J.; Spasov, K.A.; Zhang, S.; Takeo, Y.; Kudalkar, S.N.; et al. Potent Noncovalent Inhibitors of the Main Protease of SARS-CoV-2 from Molecular Sculpting of the Drug Perampanel Guided by Free Energy Perturbation Calculations. ACS Cent. Sci. 2021, doi:10.1021/ACSCENTSCI.1C00039.
- 107. Luttens, A.; Gullberg, H.; Abdurakhmanov, E.; Vo, D.D.; Akaberi, D.; Talibov, V.O.; Nekhotiaeva, N.; Vangeel, L.; De Jonghe, S.; Jochmans, D.; et al. Ultralarge Virtual Screening Identifies SARS-CoV-2 Main Protease Inhibitors with Broad-Spectrum Activity against Coronaviruses. *JACS* 2022, 144, 2905–2920, doi:10.1021/JACS.1C08402/ASSET/IMAGES/LARGE/JA1C08402_0004.JPEG.
- 108. Rossetti, G.G.; Ossorio, M.A.; Rempel, S.; Kratzel, A.; Dionellis, V.S.; Barriot, S.; Tropia, L.; Gorgulla, C.; Arthanari, H.; Thiel, V.; et al. Non-Covalent SARS-CoV-2 Mpro Inhibitors Developed from in Silico Screen Hits. *Sci. Rep.* 2022 12:1 2022, 12, 1–9, doi:10.1038/s41598-022-06306-4.
- 109. Zhang, K.; Wang, T.; Li, M.; Liu, M.; Tang, H.; Wang, L.; Ye, K.; Yang, J.; Jiang, S.; Xiao, Y.; et al. Discovery of Quinazolin-4-One-Based Non-Covalent Inhibitors Targeting the Severe Acute Respiratory Syndrome Coronavirus 2 Main Protease (SARS-CoV-2 Mpro). *Eur. J. Med. Chem.* **2023**, 257, 115487, doi:10.1016/j.ejmech.2023.115487.
- 110. Citarella, A.; Moi, D.; Pedrini, M.; Pérez-Peña, H.; Pieraccini, S.; Stagno, C.; Micale, N.; Schirmeister, T.; Sibille, G.; Gribaudo, G.; et al. Discovery of a Novel Trifluoromethyl Diazirine Inhibitor of SARS-CoV-2 Mpro. *Molecules* 2023, 28, doi:10.3390/MOLECULES28020514.
- 111. Ielo, L.; Patamia, V.; Citarella, A.; Efferth, T.; Shahhamzehei, N.; Schirmeister, T.; Stagno, C.; Langer, T.; Rescifina, A.; Micale, N.; et al. Novel Class of Proteasome Inhibitors: In Silico and In Vitro Evaluation of Diverse Chloro(Trifluoromethyl)Aziridines. *Int. J. Mol. Sci.* 2022, 23, 12363, doi:10.3390/ijms232012363.
- 112. Alhadrami, H.A.; Burgio, G.; Thissera, B.; Orfali, R.; Jiffri, S.E.; Yaseen, M.; Sayed, A.M.; Rateb, M.E. Neoechinulin A as a Promising SARS-CoV-2 Mpro Inhibitor: In Vitro and In Silico Study Showing the Ability of Simulations in Discerning Active from Inactive Enzyme Inhibitors. *Mar. Drugs* **2022**, *20*, 163, doi:10.3390/MD20030163/S1.
- 113. Zhang, Y.; Gao, H.; Hu, X.; Wang, Q.; Zhong, F.; Zhou, X.; Lin, C.; Yang, Y.; Wei, J.; Du, W.; et al. Structure-Based Discovery and Structural Basis of a Novel Broad-Spectrum Natural Product against the Main Protease of Coronavirus. *J. Virol.* **2022**, *96*, 1253–1274, doi:10.1128/JVI.01253-21/SUPPL_FILE/JVI.01253-21-S0001.PDF.
- 114. Li, J.; Zhou, X.; Zhang, Y.; Zhong, F.; Lin, C.; McCormick, P.J.; Jiang, F.; Luo, J.; Zhou, H.; Wang, Q.; et al. Crystal Structure of SARS-CoV-2 Main Protease in Complex with the Natural Product Inhibitor Shikonin Illuminates a Unique Binding Mode. *Sci Bull (Beijing)* **2021**, *66*, 661–663, doi:10.1016/J.SCIB.2020.10.018.
- 115. Zhang, J.W.; Xiong, Y.; Wang, F.; Zhang, F.M.; Yang, X.; Lin, G.Q.; Tian, P.; Ge, G.; Gao, D. Discovery of 9,10-Dihydrophenanthrene Derivatives as SARS-CoV-2 3CLpro Inhibitors for Treating COVID-19. *Eur. J. Med. Chem.* **2022**, 228, 114030, doi:10.1016/J.EJMECH.2021.114030.

- 116. Samrat, S.K.; Xu, J.; Xie, X.; Gianti, E.; Chen, H.; Zou, J.; Pattis, J.G.; Elokely, K.; Lee, H.; Li, Z.; et al. Allosteric Inhibitors of the Main Protease of SARS-CoV-2. *Antiviral Res.* **2022**, 205, 105381, doi:10.1016/j.antiviral.2022.105381.
- 117. Chaves, O.A.; Sacramento, C.Q.; Fintelman-Rodrigues, N.; Temerozo, J.R.; Pereira-Dutra, F.; Mizurini, D.M.; Monteiro, R.Q.; Vazquez, L.; Bozza, P.T.; Castro-Faria-Neto, H.C.; et al. Apixaban, an Orally Available Anticoagulant, Inhibits SARS-CoV-2 Replication and Its Major Protease in a Non-Competitive Way. J. Mol. Cell. Biol. 2022, 14, doi:10.1093/jmcb/mjac039.

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