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Giada Marroncini*, Serena Martinelli, Sara Menchetti, Francesco Bombardiere, Francesco Saverio Martelli

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Hyperhomocystinemia and Disease. Is 10 μmol/L a Suitable New Threshold Limit?

Giada Marroncini ^{1,2,*}, Serena Martinelli ², Sara Menchetti ¹, Francesco Bombardiere ¹ and Francesco Saverio Martelli ¹

- ¹ Biomolecular Diagnostic Laboratories, Via N. Porpora, Florence, 50144 Italy
- ² Department of Clinical and Experimental Medicine, University of Florence, 50139 Florence, Italy
- * Correspondence: giada.marroncini@bdmail.it

Abstract: Hyperhomocysteinemia (HHcy) is a medical condition characterized by an abnormally high level of homocysteine (Hcy) in the blood. Homocysteine is a toxic sulfur-containing amino acid that is produced during the metabolism of methionine. Under normal circumstances, Hcy is recycled back to methionine via the remethylation pathway, through the action of various enzymes and vitamins, particularly folic acid (vitamin B9) and B12 used when intracellular methionine levels are low, thus restoring the necessary levels to correctly maintain active protein synthesis. A second pathway, used in cases of intracellular methionine excess, (the trans-sulfuration pathway) is the one that recicles Hcy into cysteine (a precursor of glutathione), first passing through cystathionine (via the enzyme cystathionine beta-synthase), a reaction that requires vitamin B6 in its active form. HHcy has been identified as a risk factor for a variety of disorders, including cardiovascular diseases, multiple sclerosis, diabetes, Alzheimer's and Parkinson's diseases, osteoporosis and cancer. However, it remains unclear whether the slightly elevated concentration of Hcy (Hcy 7-10 µmol/L) is a causative factor or simply a marker of these pathologies. In human plasma, the concentration of Hcy ([Hcy]) is classified in mild (15 to 30 μmol/L), moderate (30 to 100 μmol/L), and severe (greater than 100 μmol/L). Interestingly, many laboratories continue to consider $25 \,\mu$ mol/L as normal. This review seeks to examine the controversial literature regarding the normal range of HHcy and emphasizes that even a [Hcy] level of 10 µmol/L may contribute to the development of several diseases, aiming to discuss whether it would be appropriate to lower the threshold of HHcy normal values.

Keywords: Hyperhomocysteinemia; reference range; cardiovascular disease; neurological disease; osteoporosis and cancer

1. Introduction

Epigenetics is the study of molecular modifications to DNA and chromatin that are heritable through cell division (mitosis) and potentially reversible, occurring without altering the underlying DNA sequence. Rather than altering the genetic code itself, these modifications dictate the timing, extent, and manner in which specific genes are expressed, functioning like a conductor that coordinates the harmony of gene expression. DNA methylation is the most extensively studied epigenetic mechanism. It plays a role in a variety of diseases, including birth defects [1] and cancers [2,3], and can be influenced by environmental factors [4], behavior, and nutrition [5]. This epigenetic modification typically occurs at cytosine residues adjacent to guanine residues (5'-CG-3'), referred to as CpG sites and plays a pivotal role in in both physiological and pathological conditions. Methionine (meth) is an essential amino-acid, consumed from the diet which can be regenerated from homocysteine (Hcy) via meth-synthase (MTR) in a reaction that requires vitamin B12 (cobalamin) as a cofactor. Disruption of MTR affects numerous methylation-dependent reactions, including epigenetic modifications, and also impacts the intracellular folate pathway (Figure 1)

Figure 1. Schematic diagram of Hcy physiological production through the meth and folate cycle. (A) Dietary meth is converted to Hcy through SAM and SAH and then back to meth via the remethylation pathway. Half of Hcy goes to the transsulfuration pathway, where it is converted to cysteine with the

help of CBS. Then cysteine is further converted to GSH; dietary folic acid (vitamin B9) enters the folate cycle after its conversion first to dihydrofolate (DHF) and then to THF. MTHFR is a key enzyme that converts 5, 10-methylene-THF to 5-methyl-THF.

One-carbon metabolism (OCM) is the metabolic process by which S-adenosylmeth (SAM), the ultimate biochemical methyl donor for processes such as DNA methylation, is generated. [6] During OCM, meth is metabolized to SAM by the methadenosyltransferase (MAT) family of enzymes (I, II, and III), supplying all of the methyl groups required for DNA methylation and protein post-translational modifications. In this regard, vitamin B6, folate (B9), and B12 are essential co-factors involved in many steps of the OCM metabolism and in the degradation of Hcy either by remethylation or by transsulfuration [7,8]. Genetic factors, nutritional vitamins deficiencies, age and life style factors may contribute to an abnormal increase of Hcy leading to hyperhomocysteinemia (HHcy).

In such cases, the toxic effects of Hcy are at least partially due to oxidative damage to proteins and DNA [9], so an efficient detoxification of Hcy is essential for maintaining genomic stability and cellular viability.

Thus, HHcy has been indicated as a risk factor for a variety of disorders, such as cardiovascular diseases [10–12] multiple sclerosis [11], diabetes [12], Altzheimer's disease [13] osteoporosis and osteoporotic fractures [14] and cancer [15–17]. To date, it is unclear whether the slightly elevated concentration of Hcy is the causative agent or merely a marker for the pathology (Figure 1). However, Hcy is now recognized as a risk factor for a wide range of diseases and conditions, affecting individuals from conception to death. In human plasma, Hcy concentration ([Hcy]) is typically below 12–15 μ M and the cysteine concentration level is 240–360 μ M [18]. Yet, despite this, the widely acknowledged reference classification of HHcy is mild (15 to 30 μ mol/L), moderate (30 to 100 μ mol/L), and severe (greater than 100 μ mol/L) [19]. Interestingly, numerous laboratories still persist to maintain 25 μ mol/L as normal.

This review aims to discuss the controversial literature about HHcy normality range and highlight that already 10 μ mol/L of [Hcy] may prompt to the development of several diseases.

2. The OCM Pathways

As already mentioned, Hcy is produced from meth through a process called demethylation, which involves the formation of two important intermediates: S-adenosylmeth(SAM) and S-adenosylhomocysteine (SAH). Under normal circumstances, Hcy is eliminated through three main pathways: (1) methylation to meth, which can take place through the action of MTR; (2) through an irreversible transsulfuration pathway, in which Hcy is transformed into cystathionine and eventually into cysteine; (3) remethylated using glycine betaine (N,N,N-trimethylglycine, TMG) to meth in the liver and kidneys via the enzyme betaine-homocysteine methyltransferase (BHMT), a Zn2+dependent thiolmethyltransferase and export to the plasma. Notably, BHMT makes up to 1.5% of all the soluble protein of the liver, and recent evidence suggests that it may have a greater influence on Meth and Hcy homeostasis than MTR [20].

In the remethylation cycle, the metabolism of homocysteine is closely linked to the folate cycle, during which circulating folate is converted into tetrahydrofolate (THF). Upon undergoing methylation, THF is transformed into 5,10-methylene THF, which is subsequently reduced to 5-methyl THF by the enzyme methylene tetrahydrofolate reductase (MTHFR). This cycle is referred to as the remethylation cycle because it involves the conversion of Hcy back to meth by incorporating a methyl group from 5-methyl THF derived from the folate cycle [21]. Thus, both folate and vitamin B12 are essential for the Hcy remethylation.

Therefore, when meth is sufficiently available, Hcy combines with serine and is then broken down into α -ketobutyrate and cysteine. Cysteine serves as a precursor for glutathione (GSH) and a deficiency in the transsulfuration pathway decreased GSH synthesis, the body's primary antioxidant [8]. Besides its antioxidant role, GSH also has anti-inflammatory properties, as it reduces the production of interleukins and the expression of TNF-alpha and iNOS synthase. However,

insufficiency in the transsulfuration pathway also leads to HHcy. This condition diminishes the activity of cellular glutathione peroxidase (GPx1), an intracellular antioxidant enzyme that converts hydrogen peroxide to water, favoring GSSH over GSH. The resulting imbalance in the GSH/GSSH ratio contributes to various cardiovascular and neurodegenerative disorders. In both cases, N-Acetyl-Cysteine (NAC) supplementation provides the cysteine needed for GSH synthesis, reduces HHcy, enhances GPx1 activity, and further alleviates oxidative stress [22]. The transsulfuration pathway, that mainly involves the cystathionine β -synthase (CBS), a key enzyme which converts Hcy to cystathionine, represents the metabolic link between antioxidant and methylation metabolism.

The primary cause of HHcy is genetic defects in the enzymes responsible for Hcy metabolism, a topic of significant scientific interest [23]. Specifically, polymorphisms in key enzymes involved in Hcy metabolism, such as MTHFR (the most common is the C677T polymorphism) [24], CBS (the most important polymorphism is T833C)[25], MTR [26] and methsynthase reductase (MTRR), have been identified as important subjects of study [27].

2.1. Homocysteine and Cardiovascular Disease

HHcy is emerging as a prevalent and strong risk factor for atherosclerotic vascular disease in the coronary, cerebral, and peripheral vessels, and for arterial and venous thromboembolism [28]. Indeed, persistent high levels of Hcy contribute to the development of atherosclerotic plaques and increase the risk of atherothrombotic events by inducing endothelial dysfunction and exacerbating inflammation, as well as creating a thrombophilic profile. Due to these effects, both the World Health Organization (WHO) and health ministries have acknowledged HHcys as a significant risk factor for cardiovascular disease (CVD), alongside traditional risk factors [29]. It is now well established the mechanisms by which Hcy may contributes to the development of CVD, such as its adverse effects on the vascular endothelium and smooth muscle cells, which lead to alterations in subclinical arterial structure and function. Therefore, HHcy is considered an independent risk factor for atherosclerosis [30,31] and several studies clearly define a strong correlation between elevated Hcy and myocardial infarction, stroke and increased cardiovascular mortality [32]. Hey affects blood vessels by regulating the contractility of vascular smooth muscle cells and the permeability of endothelial cells. The major mechanism involved is the inhibition of endothelial nitric oxide synthase (eNOS), which is responsible for producing nitric oxide (NO) [33,34]. Under normal conditions, a system of antioxidant mechanisms regulates the production of reactive oxygen species (ROS). However, during adverse and chronic conditions, an imbalance in this system disrupts the generation of both NO and ROS, resulting in endothelial dysfunction [35]. Oxidative stress, thiolactone formation and protein homocysteinylation are directly related to endothelial toxicity [36,37]. In vitro studies have proposed two main mechanisms by which Hcy contributes to the accumulation of reactive oxygen species decreasing the activity of GPx1 and by inhibiting dimethylarginine dimethylaminohydrolase (DDAH), an enzyme involved in the metabolism of asymmetric dimethylarginine (ADMA), which is an endogenous inhibitor of eNOS [33,37]. Additionally, HHcyinduced oxidative stress is known to activate matrix metalloproteinases (MMPs), which disrupt extracellular matrix (ECM) metabolism and increase collagen deposition, leading to vascular fibrosis [38]. Therefore, the proinflammatory effect of HHcy is linked to ROS generation and involves the activation of nuclear transcription factor κB (NF-κB), which regulates mainly the genes responsible for the expression of intercellular adhesion molecule-1 (ICAM-1), monocyte chemoattractant protein-1 (MCP-1), vascular adhesion molecule-1 (VCAM-1) and E-selectin leading to the progression of atherosclerosis [39-41]. The role of Hcy in the activation of factor V and tissue factor (TF) which propagate coagulation and the parallel inhibition of antithrombin III, have also been well established [42,43].

Several studies in adults indicated that the risk of coronary artery disease is directly linked to [Hcy], with significant risk observed between 10 and 15 μ mol/L. Furthermore, for every 5 μ mol/L increase in [Hcy], the risk increases by nearly 20% [44,45]. In the Third National Health and Nutrition Examination Survey (NHANES) conducted from 1988 to 1994, researchers found that serum [Hcy] were independently associated with blood pressure. Specifically, a 5 μ mol/L increase in Hcy was

associated with an increase in diastolic blood pressure of 0.5 and 0.7 mm Hg and an increase in systolic blood pressure of 0.7 and 1.2 mm Hg in men and women, respectively. These findings were based on a sample size of 5.978 participants [46]. Current guidelines do not recognize Hcy as a CVD risk stratification tool even if, prospective studies, showed that rising Hcy levels predict adverse CV events better than the Framingham Risk Score, suggesting it could be considered a "novel" CVD risk marker [47]. Carnagarin and colleagues demonstrated that in hypertensive patients with Hcy above $10 \ \mu \text{mol/L}$, ACE inhibitors may be less effective in reducing blood pressure and preventing vascular damage [48] (Table 1).

2.2. Homocysteine and Neurodegenerative Diseases

2.2.1. Multiple Sclerosis (MS)

The manipulation of methylation processes has been shown to affect the vulnerability of neurons to degeneration and apoptosis in experimental models of neurodegenerative disorders as Alzheimer's disease (AD) [49], Parkinson's disease (PD) [50] and Huntington's disease (HD) [51]. Specifically, changes in the DNA methylation patterns, which is important regulators of gene expression, have been implicated in the pathogenesis of neurodegenerative diseases [52]. Modulating these epigenetic modifications can influence the susceptibility of neurons to the detrimental effects that lead to neurodegeneration and cell death. Indeed, SAM is also involved in the methylation of proteins, phospholipids and neurotransmitters [53] suggesting a role for this enzyme in signal transduction processes in the nervous system. The accumulation of Hcy can directly damage and kill neurons and some in vitro studies showed that the toxicity is caused by the activation of N-methyl-D-aspartate (NMDA) receptor [54] or apoptosis triggered by DNA damage [49] which typically involved PARP and p53 activity.

Multiple sclerosis (MS) is a chronic inflammatory demyelinating disease of the central nervous system (CNS) that results in neurological disability. The etiology of MS is thought to be multifactorial, with genetic and environmental factors interacting to contribute to the autoimmune inflammatory process [55]. Numerous studies have examined the potential roles of Hcy, vitamin B12, and folate as contributors to the neurodegenerative process [55].

Indeed, HHcy may be a risk factor for neurological decline in MS and may have a neurodegenerative effect [56]. Several studies have reported a significant increase in plasma [Hcy] in MS patients compared to healthy controls. However, the precise mechanism by which elevated Hcy contributes to the pathogenesis of MS is not yet fully understood [57]. However, the key condition in the development of MS is the dysregulation of the blood-brain barrier (BBB) and the increased migration of leukocytes across the BBB [58]. In particular, SOD1G93A transgenic mouse model of amyotrophic lateral sclerosis (ALS) demonstrated that the level of 5-5-methyltetrahydrofolate (MTHF) significantly decreased in the plasma, spinal cord and cortex at the early stages of presymptomatic ALS and that the level of Hcy were markedly elevated even after the motor symptoms appeared [59].

In humans the levels of Hcy found in the cerebrospinal fluid (CSF) and brain tissue are reported to range from 0.5 to 10 μ m [60] and can be taken up rapidly by neurons via a specific membrane transporter (L-homocysteine sulphinate (L-HCSA) and L-homocysteate (L-HCA)) [61]. It has been demonstrated that Hcy levels increased up to 20–30 μ m in amyloid precursor protein (APP) mutant transgenic mice maintained on the methyl donor-deficient while folic acid supplementation seemed to reduce risk of sporadic forms of AD and also might suppress the neurodegenerative process [62]. Interestingly, even after adjusting for vitamin B6, vitamin B12, and folate, patients with MS had a statistically significant higher plasma [Hcy] (4.5 μ mol/l, 6.2 vs 2.7 μ mol/l) compared to healthy patients, indicating that the elevated levels might be due to increased production rather than decreased removal [63](Table 1).

2.2.2. Alzheimer's Disease (AD)

Alzheimer's diseases is a progressive and neurodegenerative disorder with typical symptoms of progressive memory loss and cognitive dysfunction [64] closely associated with a variety of risk factors such as ageing, ApoE4 genotype, hyperglycemia and HHcy [65]. A much more consistent number of studies indicated that hHcy is an important and independent risk factor and biomarker independent of B vitamins for AD [66,67]. Recently, a meta-analysis studies (28.257 partecipants) found that every 5 µmol/L increase in blood Hcy is linearly associated with a 15% increase in relative risk of Alzheimer-type dementia [68]. It has been reported that in patients with AD, Hcy levels do not increase over time following the onset of the disease; however, these levels correlate with the severity of disease [49]. Additionally, it is suggested that dementia may develop prior to HHcy in the later stages [69].

In vitro studies conducted on cultured neurons, Hcy activated tau phosphokinases (glycogen synthase kinase 3 and cyclin-dependent kinase 5), and inhibited protein phosphatase 2A (a main tau phosphatase), leading to increases in phosphorylated tubulin associated unit (TAU) levels, as well as rises of tau aggregates and truncated TAU species [70]. Accordingly, in vivo studies demonstrated that high dose of Hcy (16.4 mmol/l vs 8 mmol/l in control group) induced by diet in a widely-used mice contain three mutations (3XTg mice) associated with familial AD, were able to promoted TAU phosphorylation at T231/S235 sites.

Some cohort studies have suggested that higher amount of Hcy, meth, and SAM may accelerate cognitive decline in patients with mild cognitive impairment (MCI) or AD, and that vitamin B12 deficiency may worsen the clinical outcome [71]. An interesting case series demonstrated that decreased levels of vitamin B12 and folate and serum [Hcy] from 22.5 to 8 µmol/L directly correlates with atrophic changes in the cerebral cortex and AD progression [72] (Table 1).

Moreover, the normalization of hHcy has been shown to enhance cognitive function and reduce brain amyloidosis in a transgenic mouse model of AD [73].

2.3. Homocysteine and Diabetes Mellitus (DM)

HHcy has been implicated in the pathogenesis of DM, owing to its promotion of oxidative stress, β-cell dysfunction, and insulin resistance [74,75]. Recent in vivo and in vitro studies indicate that OCM nutrients play a crucial role in supporting energy and glucose metabolism through various mechanisms. Indeed, low levels of folate, choline, or vitamin B12 have been shown to induce HHcy, which is linked to the development of DM [76,77]. As mentioned above, HHcy elevates ROS and Creactive protein (CRP) levels, promoting oxidative stress and systemic inflammation. The activation of stress-sensitive signaling pathways can ultimately results in pancreatic β -cells dysfunction [78], glucose intolerance, and insulin resistance [79]. Changes in homocysteine metabolic enzymes have been observed in different diabetic conditions [80,81]. In rats on a high-fat sucrose diet, there was a positive correlation between plasma insulin levels and both hcy and MTHFR activity, while an inverse correlation was found with cystathionine-β-synthase (CβS) activity [82,83]. Indeed, some in vitro studies showed that folic acid supplementation during the rat juvenile-pubertal period increased methylation in the promoter regions of insulin receptor, peroxisome proliferator-activated receptor- α (PPAR- α) and glucocorticoid receptor genes involved in metabolic homeostasis [84]. Analogously, mice that were deprived of folate in utero exhibited hypomethylation at the differentially methylated region (DMR) 1 of the imprinted locus insulin-like growth factor 2 (Igf2), as well as at Slc389a4CGl1, in blood, liver, and kidney tissues [85].

HHcy is directly linked to a greater risk of developing type 2 diabetes mellitus (T2DM) [77]. Thus, a cross sectional study performed on a cohort of 75 patients with T2DM and 54 healthy control subjects clearly demonstrated that plasma Hcy was significantly higher in patients with T2DM compared with controls (Hcy 12.0 ± 0.7 vs 8.7 ± 0.3 µmol/l) [86] and that insulin resistance and renal function are independent determinants of Hcy levels in DM patients. Recent evidences showed that in subjects > 65 years with DN (n=1,845) and in a non-DM group (n=28,720) concentration of Hcy > 12 µmol/L was a good indicator to predict impaired kidney function in DN patients and that [Hcy] in DN was inversely proportional to the estimated glomerular filtration rate (eGFR) [87]. Therefore, some clinical trials demonstrated that elevated Hcy levels (>50 mol/L) and/or fenofibrate

therapy, one of the best options to treat the atherogenic lipid triad (i.e., high triglycerides, low HDL, elevated small-size LDL) of T2DM, may interfere with other atheroprotective functions of HDL particles such as anti-oxidative and anti-inflammatory actions. In particular, markedly HHcy impaired ability to facilitate cholesterol efflux from macrophages-foam cell independently on apoA-I levels [88]. Moreover, high plasma [Hcy] has been associated with increased risk for coronary heart disease (CHD) events in DM individuals. Notably, in T2DM patients, plasma Hcy of 15 μ mol/L or more at baseline had a higher risk for CHD death than those with plasma Hcy levels less than 15 μ mol/L (26.1% and 13.5%, respectively; P = 0.005) [89] (Table 1).

2.4. Homocysteine and Osteoporosis

Osteoporosis is a major health problem characterized by low bone mineral density (BMD), deterioration of bone microarchitecture, and increased risk of fracture [90]. Osteoporotic fractures (OF) are associated with increased morbidity and mortality and substantial economic costs [91–93]. Recent investigations have revealed a link between elevated plasma Hcy levels and both lower BMD and a higher risk of bone fractures [94–96]. HHcy is now recognized as an independent risk factor for osteoporosis and fractures, particularly in the elderly [97].

Two independent prospective studies examined the relationship between Hcy levels and fracture incidence in three groups of men and women aged 55 years and older. The observed association between Hcy levels and fracture risk was primarily related to BMD, as well as dietary factors such as calorie, protein, calcium, and vitamin intake [14]. This study involved 2,406 individuals and found a significant association between circulating Hcy levels and the risk of OF (RR = 1.4 per 1 SD increase in natural log-transformed Hcy level; 95% CI, 1.2-1.6) [14]. Furthermore, elevated Hcy levels (>20 µmol/L for men and >18 µmol/L for women) were associated with a substantial increase in fracture risk (4.1-fold for men and 1.9-fold for women) [98] (Table 1).

Several mechanisms have been proposed to explain the connection between HHcy and the development of osteoporosis: (i) decreased bone formation due to apoptosis in human bone marrow stromal cells [99], (ii) stimulation of osteoclastogenesis through increasing intracellular radical oxygen species (ROS) generation and activation of matrix metalloproteinases (MMPs) that can then degrade extracellular bone matrix [100], (iii) reduced bone blood flow leading to changes in bone biomechanical properties [101], (iiii) altered gene expression with reduced methylation capacity, (iiiii) and reduced bone strength through interference with collagen cross-links [95].

Hey was shown to induce apoptosis in primary human bone marrow stromal cells and the HS-5 cell line, and this apoptotic effect was caspase-dependent. Furthermore, Hey induced increase in the release of cytochrome c into the cytosol and activated caspase-9 and caspase-3, but not caspase-8, indicating that the induction of apoptosis occurred through the mitochondrial pathway [99].

HHcy significantly reduced bone blood flow, as demonstrated in studies where Hcy-treated rats exhibited a lower tibial blood flow index compared to controls [102], and this reduction in blood flow was associated with increased oxidative stress and MMPs activity, leading to extracellular matrix degradation [100]. HHcy is also linked to altered gene expression through reduced DNA methylation capacity, impacting various cellular processes mainly by increasing in the expression of 5-lipoxygenase (5LO) and subsequently inducing hypomethylation of its promoter, which is associated with elevated S-adenosylhomocysteine (SAH) levels and reduced DNA methyltransferase activity [103]. Chronic HHcy exposure can also lead to the demethylation of the human telomerase reverse transcriptase (hTERT) promoter, resulting in decreased telomerase activity and accelerated senescence in endothelial cells. This process is mediated by the repression of key transcription factors and reduced DNA methyltransferase expression [104,105]. While the predominant view highlights Hcy's detrimental effects on gene expression via hypomethylation, some studies suggest potential compensatory mechanisms, such as dietary interventions that may restore methylation capacity and mitigate these effects [106].

Blouin and colleagues investigated the relationship between plasma Hcy levels and bone material properties, specifically focusing on collagen cross-link ratios in patients who had sustained femoral neck fractures [95]. In trabecular areas undergoing bone formation, the collagen cross-link

ratio was significantly higher in the high Hcy group $(7.6 \pm 0.2; N=10)$ compared to the low Hcy group $(6.4 \pm 0.2; N=9)$. Conversely, in areas undergoing resorption, there was no significant difference in the collagen cross-link ratio between the two groups $(12.5 \pm 0.5 \text{ for high Hcy and } 11.4 \pm 0.4 \text{ for low Hcy})$. A strong correlation was found between blood Hcy levels and collagen cross-link ratio at forming trabecular surfaces (Spearman's r = 0.83, p < 0.0001), but not at resorbing surfaces (Spearman's r = 0.45, p > 0.05). The study concluded that while there is a significant correlation between plasma Hcy levels and collagen cross-link ratios in bone-forming areas, this relationship is independent of bone mineral content and distribution patterns [95] .

Although the relationship between Hcy levels and osteoporosis is increasingly recognized, future research should focus on larger cohorts and explore potential therapeutic interventions, such as probiotics, to mitigate the detrimental effects of Hcy on bone health. While current findings suggest a significant association between Hcy and osteoporosis, the lack of consensus on causality calls for further studies to clarify these relationships and explore potential treatment avenues.

2.5. Homocysteine and Cancer

Cancer cells exhibit a strong reliance on the meth cycle, leading to the production of significant amounts of Hcy. Elevated intracellular Hcy levels can be released into the bloodstream, prompting numerous studies to establish a link between high Hcy levels and cancer [16]. Numerous laboratories classified blood Hcy levels exceed 15 µmol/L as HHcy which is associated with oxidative stress, endoplasmic reticulum (ER) stress, apoptosis, protein oxidation, inflammation, and impaired angiogenesis [107,108]. Moreover, several studies have reported inconsistent associations between polymorphisms in genes related to Hcy metabolism and cancer [15,109,110], but together with dietary meth, folate, vitamin B12, B6, and alcohol consumption, genetic polymorphisms maybe responsible of tumors genesis. These polymorphisms are often linked to HHcy and different cancer types. The most common mutations in MTHFR 677C->T transition at codon 222 and 1298A->C transversion at codon 429 have been associated with cervical [111], colorectal [112], endometrial [113], and esophageal cancer [114]. Analogously, MTRR gene A66G Ile22Met is found to be associated with colorectal cancer (CRC) [115] and leukemia [15]. Likewise, the allele of MTR2756G showed a positive association with CRC and the association with MTHFR1298AA further increases the risk of developing this cancer type [116].

Therefore, strong evidences supported an association between plasma Hcy levels and progression of rectal cancer from normal, to rectal adenoma, to rectal cancer. Therefore, homocysteine (Hcy) could potentially serve as a therapeutic tool to assist clinicians in the staging of rectal cancer. Infect, plasma [Hcy] directly increased from normal control subjects to patients with low-risk and high-risk adenomas, and to patients with Stage I–IV rectal cancer [117]. Similarly, from a recent metanalysis was found that every 5μ mol/L increase in Hcy was associated with a 7% higher risk of digestive cancer occurrence and the authors concluded that [Hcy] may be a potential biomarker in this type of cancer [118] (Table 1).

Increasing evidences suggest that variations in the use of dietary nutrients significantly influence tumor metabolism, growth, and treatment results [119–121]. One-carbon metabolism, which plays a critical role in redox and nucleotide metabolism, is a key target for standard cancer treatments like 5-fluorouracil and radiation therapy [122,123]. Particularly, meth is one of the nutrients that cancer cells require to maintain cell proliferation, survival, and metastasis [124]. Cells employ various pathways to convert folic acid and 5-methyl THF into THF, which can then accept a one-carbon unit for subsequent biosynthetic reactions. Folic acid is converted to THF by the enzyme dihydrofolate reductase (DHFR), a known target of the antifolate chemotherapy drug methotrexate. On the other hand, the conversion of 5-methyl THF to THF is linked to the synthesis of meth. This process involves the transfer of a methyl group to Hcy, facilitated by the vitamin B12-dependent enzyme methsynthase (MTR). MTR is the sole enzyme recognized for using 5-methyl THF as a substrate to regenerate the THF structure. Consequently, a deficiency in cobalamin leads to a loss of MTR activity, resulting in an accumulation of intracellular 5-methyl THF, a phenomenon clinically referred to as the "methyl-folate trap". This irreversible accumulation of folates as 5-methyl THF limits the

availability of THF in cells for new one-carbon unit incorporation. Several lines of evidence suggest that MTR's role in the folate cycle may be critical for tumor growth [125].

A study revealed that cancer cells exhibit elevated levels of the meth transporter SLC43A2, facilitating their increased uptake of meth, which in turn promotes cancer progression [126]. Growing evidence suggest that dietary interventions can effectively be used as therapeutic tools for treating cancer. In this regard, lowering the intake of meth or cystine has been shown to safeguard non-tumor cells and enhance the immune surveillance system in the fight against cancer [127]. Indeed, in an in vivo study, dietary meth restriction (MR) has been shown to increase [128] and enhance metabolic health [129]. Notably, certain cancer cell lines are dependent on meth for growth [130] and limiting or reducing meth intake in the diet may exhibit anti-cancer properties in mouse models of colon carcinogenesis [131].

Table 1. Range values of Hcy in diseases. The table summarizes the range of Hcy values associated with each disease covered in the review.

Disease	Serum [Hcy]	REF
	every 5 μmol/L increase in [Hcy], the risk of CD	[44,45]
	increases by nearly 20%	
Cardiovascular disease		
	<10 µmol/L (coronary artery disease)	[44,45]
	>10 μmol/L (ACE inhibitors failure)	[48]
	every 5 µmol/L increase in blood homocysteine is	
	linearly associated with a 15% increase in relative	
	risk of AD	
Neurodegenerative		[60]
diseases	4.5 - $6.2 \ \mu mol/l$ MS patients vs 2.7 in healthy	[68]
(MS and AD)	patients	[(2]
		[63]
	From 8 to 22 µmol/L directly correlates with	[70]
	atrophic changes in the cerebral cortex and AD	[72]
	progression	
Endocrine disease:	< 15 µmol/ (CHD death in DM patients)	
(DM)		[89]
	DM patients 12.0 ± 0.7 vs healthy patients 8.7 ± 0.3	
	μmol/l	[86]
	Hcy > 12 μmol/L was a good indicator to predict	[87]
	impaired kidney function in DM patients	
Osteoporosis	>20 μmol/L(men) and >18 μmol/L(women)	[98]
	increase fracture risk	[, 0]
	every 5µmol/L increase in homocysteine was	
Cancers	associated with a 7% higher risk of digestive	[118]
Cancers	associated with a 7 /0 Higher fisk of digestive	11101

3. Discussion

The range of diseases associated with HHcy is fairly well delineated. Indeed, the relationship between Hcy and CV, neurological, endocrine and oncological diseases revolve around its biochemical pathways. As mentioned above, Hcy is a toxic sulfur-containing amino acid that results from meth metabolism, and its levels are influenced by various factors, including genetic, nutritional, and lifestyle elements. HHcy has been listed by the World Health Organization (WHO) as a

cardiovascular, cerebrovascular and peripheral vascular risk factor [132], but its correlation with dysmetabolic disease and cancer is likewise disputed. In particular, there is a lot of confusion about the definition of optimal reference ranges. Actually, regarding reference ranges, it is recommended that each laboratory centres conducting the analysis develop its own reference intervals for [Hcy]. This task is complicated by numerous variables that influence circulating concentrations, including age, gender, ethnicity, pregnancy, menopause, smoking status, nutritional status, renal function, medication use, and the fortification of food with folic acid. For adults, the reference range for Hcy is generally between 6 and 10 µmol/L; however, in countries where food fortification with folic acid is mandated, this range is typically 10-30% lower. In 2003, a consensus document defined HHcy based on plasma concentrations, categorizing it as moderate (Hcy levels of 15-30 µmol/L), intermediate (30-100 μmol/L), and severe (greater than 100 μmol/L). So, Hcy is defined as normal when <15 μmol/L [133]. Nowadays, many laboratories continue to utilize these reference ranges, despite the fact that they are outdated and do not align with contemporary scientific literature. As discussed in this review, in CVD and AD patients the normality threshold is much lower. Indeed, many studies indicated that the Hcy < 10 µmol/L is recommended and that a significant increase of CVD and neurological disorders is observed risk observed precisely between 10 and 15 µmol/L [10,30,31,48,134]. The scientific community concurs that even a 5 µmol/L rise in Hcy levels serves as a predictor for various diseases, including cardiovascular, endocrine, neurodegenerative and oncological conditions. Indeed, a dose-response meta-analysis conducted in 2017 demonstrated a linear relationship between Hcy levels and the risk of all-cause mortality, indicating that for each 5 µmol/L increase in Hcy, there is a corresponding 33.6% rise in the risk of all-cause mortality [8,44,68,118].

4. Methods

For this narrative review we searched English-language publications in PubMed published mostly in the last three years. Our main search terms, their associations and their acronyms were: "homocysteine, cardiovascular disease, diabetes mellitus, cancer, osteoporosis, neurodegenerative diseases". In this narrative review we considered the role of Hcy in the above-mentioned diseases focused on the plasma [Hcy]. English language articles related to the topic were considered if they discussed one of the issues of interest and were peer reviewed.

5. Conclusion

The Hcy metabolism is a complex network of pathways, which includes several actors and regulators. In this intricate scenario, a critical role is played by HHcy, which can affect the onset and progression of several pathologic conditions such as cardiovascular diseases, endocrine pathologies, neurodegenerative pathologies, osteoporosis and cancer. Since there is a lot of confusion about the definition of optimal Hcy reference ranges, this review seeks to emphasize the importance of a revised formulation of the reference ranges for Hcy assessment. It aims to incorporate recent literature findings, as well as the genetic alteration leading to HHcy by obstructing the disposal or regeneration pathways of meth. We believe that revised criteria should allow people to be included in risk groups for developing diseases and be more frequently and carefully monitored to avoid diseases onset.

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References

- 1. Pujadas, E.; Feinberg, A.P. Regulated noise in the epigenetic landscape of development and disease. Cell 2012, 148, 1123-1131, doi:10.1016/j.cell.2012.02.045.
- Feinberg, A.P.; Tycko, B. The history of cancer epigenetics. Nat Rev Cancer 2004, 4, 143-153, doi:10.1038/nrc1279.

- 4. Etchegaray, J.P.; Mostoslavsky, R. Interplay between Metabolism and Epigenetics: A Nuclear Adaptation to Environmental Changes. Mol Cell 2016, 62, 695-711, doi:10.1016/j.molcel.2016.05.029.
- Zhai, J.; Kongsberg, W.H.; Pan, Y.; Hao, C.; Wang, X.; Sun, J. Caloric restriction induced epigenetic effects on aging. Front Cell Dev Biol 2022, 10, 1079920, doi:10.3389/fcell.2022.1079920.
- Anderson, O.S.; Sant, K.E.; Dolinoy, D.C. Nutrition and epigenetics: an interplay of dietary methyl donors, one-carbon metabolism and DNA methylation. J Nutr Biochem 2012, 23, 853-859, doi:10.1016/j.jnutbio.2012.03.003.
- 7. Herrmann, W.; Herrmann, M.; Obeid, R. Hyperhomocysteinaemia: a critical review of old and new aspects. Curr Drug Metab 2007, 8, 17-31, doi:10.2174/138920007779315008.
- 8. Fan, R.; Zhang, A.; Zhong, F. Association between Homocysteine Levels and All-cause Mortality: A Dose-Response Meta-Analysis of Prospective Studies. Sci Rep 2017, 7, 4769, doi:10.1038/s41598-017-05205-3.
- Sibrian-Vazquez, M.; Escobedo, J.O.; Lim, S.; Samoei, G.K.; Strongin, R.M. Homocystamides promote freeradical and oxidative damage to proteins. Proc Natl Acad Sci U S A 2010, 107, 551-554, doi:10.1073/pnas.0909737107.
- 10. Guieu, R.; Ruf, J.; Mottola, G. Hyperhomocysteinemia and cardiovascular diseases. Ann Biol Clin (Paris) 2022, 80, 7-14, doi:10.1684/abc.2021.1694.
- 11. Pan, L.; Yin, Y.; Chen, J.; Ma, Z.; Chen, Y.; Deng, X.; Zhang, H.T.; Leng, H.; Wu, K. Homocysteine, vitamin B12, and folate levels in patients with multiple sclerosis in Chinese population: A case-control study and meta-analysis. Mult Scler Relat Disord 2019, 36, 101395, doi:10.1016/j.msard.2019.101395.
- Suliman, M.E.; Stenvinkel, P.; Bárány, P.; Heimbürger, O.; Anderstam, B.; Lindholm, B. Hyperhomocysteinemia and its relationship to cardiovascular disease in ESRD: influence of hypoalbuminemia, malnutrition, inflammation, and diabetes mellitus. Am J Kidney Dis 2003, 41, S89-95, doi:10.1053/ajkd.2003.50093.
- 13. Hassin-Baer, S.; Cohen, O.; Vakil, E.; Sela, B.A.; Nitsan, Z.; Schwartz, R.; Chapman, J.; Tanne, D. Plasma homocysteine levels and Parkinson disease: disease progression, carotid intima-media thickness and neuropsychiatric complications. Clin Neuropharmacol 2006, 29, 305-311, doi:10.1097/01.WNF.0000236763.16032.60.
- van Meurs, J.B.; Dhonukshe-Rutten, R.A.; Pluijm, S.M.; van der Klift, M.; de Jonge, R.; Lindemans, J.; de Groot, L.C.; Hofman, A.; Witteman, J.C.; van Leeuwen, J.P.; et al. Homocysteine levels and the risk of osteoporotic fracture. N Engl J Med 2004, 350, 2033-2041, doi:10.1056/NEJMoa032546.
- Skibola, C.F.; Smith, M.T.; Kane, E.; Roman, E.; Rollinson, S.; Cartwright, R.A.; Morgan, G. Polymorphisms in the methylenetetrahydrofolate reductase gene are associated with susceptibility to acute leukemia in adults. Proc Natl Acad Sci U S A 1999, 96, 12810-12815, doi:10.1073/pnas.96.22.12810.
- Hellmich, M.R.; Szabo, C. Hydrogen Sulfide and Cancer. Handb Exp Pharmacol 2015, 230, 233-241, doi:10.1007/978-3-319-18144-8_12.
- 17. Newman, A.C.; Maddocks, O.D.K. One-carbon metabolism in cancer. Br J Cancer 2017, 116, 1499-1504, doi:10.1038/bjc.2017.118.
- 18. Rehman, T.; Shabbir, M.A.; Inam-Ur-Raheem, M.; Manzoor, M.F.; Ahmad, N.; Liu, Z.W.; Ahmad, M.H.; Siddeeg, A.; Abid, M.; Aadil, R.M. Cysteine and homocysteine as biomarker of various diseases. Food Sci Nutr 2020, 8, 4696-4707, doi:10.1002/fsn3.1818.
- 19. Kang, S.S.; Wong, P.W.; Malinow, M.R. Hyperhomocyst(e)inemia as a risk factor for occlusive vascular disease. Annu Rev Nutr 1992, 12, 279-298, doi:10.1146/annurev.nu.12.070192.001431.
- 20. Pellanda, H. Betaine homocysteine methyltransferase (BHMT)-dependent remethylation pathway in human healthy and tumoral liver. Clin Chem Lab Med 2013, 51, 617-621, doi:10.1515/cclm-2012-0689.
- 21. Blom, H.J.; Shaw, G.M.; den Heijer, M.; Finnell, R.H. Neural tube defects and folate: case far from closed. Nat Rev Neurosci 2006, 7, 724-731, doi:10.1038/nrn1986.
- Aydın, A.F.; Kondakçı, G.; Hatipoğlu, S.; Doğru-Abbasoğlu, S.; Uysal, M. N-Acetylcysteine supplementation decreased brain lipid and protein oxidations produced by experimental homocysteine thiolactone exposure: Relevance to neurodegeneration. Pathophysiology 2018, 25, 125-129, doi:10.1016/j.pathophys.2018.02.004.
- 23. Horigan, G.; McNulty, H.; Ward, M.; Strain, J.J.; Purvis, J.; Scott, J.M. Riboflavin lowers blood pressure in cardiovascular disease patients homozygous for the 677C-->T polymorphism in MTHFR. J Hypertens 2010, 28, 478-486, doi:10.1097/HJH.0b013e328334c126.
- 24. McNulty, H.; Dowey, I.R.; Strain, J.J.; Dunne, A.; Ward, M.; Molloy, A.M.; McAnena, L.B.; Hughes, J.P.; Hannon-Fletcher, M.; Scott, J.M. Riboflavin lowers homocysteine in individuals homozygous for the MTHFR 677C->T polymorphism. Circulation 2006, 113, 74-80, doi:10.1161/CIRCULATIONAHA.105.580332.
- 25. Ding, R.; Lin, S.; Chen, D. The association of cystathionine β synthase (CBS) T833C polymorphism and the risk of stroke: a meta-analysis. J Neurol Sci 2012, 312, 26-30, doi:10.1016/j.jns.2011.08.029.

- 27. Ambrosino, P.; Lupoli, R.; Di Minno, A.; Nardo, A.; Marrone, E.; Lupoli, V.; Scaravilli, A.; Mitidieri, E.; Tufano, A.; Di Minno, M.N. Cyclic supplementation of 5-MTHF is effective for the correction of hyperhomocysteinemia. Nutr Res 2015, 35, 489-495, doi:10.1016/j.nutres.2015.02.006.
- 28. van Guldener, C.; Nanayakkara, P.W.; Stehouwer, C.D. Homocysteine and blood pressure. Curr Hypertens Rep 2003, 5, 26-31, doi:10.1007/s11906-003-0007-z.
- 29. 1991 guidelines for the prevention of hypertension and associated cardiovascular disease. Joint World Health Organization/International Society of Hypertension Meeting. J Hypertens 1992, 10, 97-99, doi:10.1097/00004872-199201000-00016.
- Catena, C.; Colussi, G.; Nait, F.; Capobianco, F.; Sechi, L.A. Elevated Homocysteine Levels Are Associated With the Metabolic Syndrome and Cardiovascular Events in Hypertensive Patients. Am J Hypertens 2015, 28, 943-950, doi:10.1093/ajh/hpu248.
- Ganguly, P.; Alam, S.F. Role of homocysteine in the development of cardiovascular disease. Nutr J 2015, 14, 6, doi:10.1186/1475-2891-14-6.
- 32. Nygård, O.; Nordrehaug, J.E.; Refsum, H.; Ueland, P.M.; Farstad, M.; Vollset, S.E. Plasma homocysteine levels and mortality in patients with coronary artery disease. N Engl J Med 1997, 337, 230-236, doi:10.1056/NEJM199707243370403.
- 33. Stühlinger, M.C.; Tsao, P.S.; Her, J.H.; Kimoto, M.; Balint, R.F.; Cooke, J.P. Homocysteine impairs the nitric oxide synthase pathway: role of asymmetric dimethylarginine. Circulation 2001, 104, 2569-2575, doi:10.1161/hc4601.098514.
- 34. Pushpakumar, S.; Kundu, S.; Sen, U. Endothelial dysfunction: the link between homocysteine and hydrogen sulfide. Curr Med Chem 2014, 21, 3662-3672, doi:10.2174/0929867321666140706142335.
- 35. Boushey, C.J.; Beresford, S.A.; Omenn, G.S.; Motulsky, A.G. A quantitative assessment of plasma homocysteine as a risk factor for vascular disease. Probable benefits of increasing folic acid intakes. JAMA 1995, 274, 1049-1057, doi:10.1001/jama.1995.03530130055028.
- Jakubowski, H.; Zhang, L.; Bardeguez, A.; Aviv, A. Homocysteine thiolactone and protein homocysteinylation in human endothelial cells: implications for atherosclerosis. Circ Res 2000, 87, 45-51, doi:10.1161/01.res.87.1.45.
- 37. Handy, D.E.; Zhang, Y.; Loscalzo, J. Homocysteine down-regulates cellular glutathione peroxidase (GPx1) by decreasing translation. J Biol Chem 2005, 280, 15518-15525, doi:10.1074/jbc.M501452200.
- 38. Sen, U.; Pushpakumar, S.B.; Amin, M.A.; Tyagi, S.C. Homocysteine in renovascular complications: hydrogen sulfide is a modulator and plausible anaerobic ATP generator. Nitric Oxide 2014, 41, 27-37, doi:10.1016/j.niox.2014.06.006.
- 39. Papatheodorou, L.; Weiss, N. Vascular oxidant stress and inflammation in hyperhomocysteinemia. Antioxid Redox Signal 2007, 9, 1941-1958, doi:10.1089/ars.2007.1750.
- Postea, O.; Krotz, F.; Henger, A.; Keller, C.; Weiss, N. Stereospecific and redox-sensitive increase in monocyte adhesion to endothelial cells by homocysteine. Arterioscler Thromb Vasc Biol 2006, 26, 508-513, doi:10.1161/01.ATV.0000201039.21705.dc.
- 41. Robinson, J.M.; Ohira, T.; Badwey, J.A. Regulation of the NADPH-oxidase complex of phagocytic leukocytes. Recent insights from structural biology, molecular genetics, and microscopy. Histochem Cell Biol 2004, 122, 293-304, doi:10.1007/s00418-004-0672-2.
- 42. Gerdes, V.E.; Hovinga, H.A.; ten Cate, H.; Macgillavry, M.R.; Leijte, A.; Reitsma, P.H.; Brandjes, D.P.; Büller, H.R.; Group, A.V.M. Homocysteine and markers of coagulation and endothelial cell activation. J Thromb Haemost 2004, 2, 445-451, doi:10.1111/j.1538-7836.2004.00674.x.
- 43. Palareti, G.; Coccheri, S. Lowered antithrombin III activity and other clotting changes in homocystinuria: effects of a pyridoxine-folate regimen. Haemostasis 1989, 19 Suppl 1, 24-28, doi:10.1159/000216092.
- 44. Jacobsen, D.W. Homocysteine and vitamins in cardiovascular disease. Clin Chem 1998, 44, 1833-1843.
- Humphrey, L.L.; Fu, R.; Rogers, K.; Freeman, M.; Helfand, M. Homocysteine level and coronary heart disease incidence: a systematic review and meta-analysis. Mayo Clin Proc 2008, 83, 1203-1212, doi:10.4065/83.11.1203.
- 46. Lim, U.; Cassano, P.A. Homocysteine and blood pressure in the Third National Health and Nutrition Examination Survey, 1988-1994. Am J Epidemiol 2002, 156, 1105-1113, doi:10.1093/aje/kwf157.
- 47. Veeranna, V.; Zalawadiya, S.K.; Niraj, A.; Pradhan, J.; Ference, B.; Burack, R.C.; Jacob, S.; Afonso, L. Homocysteine and reclassification of cardiovascular disease risk. J Am Coll Cardiol 2011, 58, 1025-1033, doi:10.1016/j.jacc.2011.05.028.
- 48. Carnagarin, R.; Nolde, J.M.; Ward, N.C.; Lugo-Gavidia, L.M.; Chan, J.; Robinson, S.; Jose, A.; Joyson, A.; Azzam, O.; Galindo Kiuchi, M.; et al. Homocysteine predicts vascular target organ damage in hypertension

- and may serve as guidance for first-line antihypertensive therapy. J Clin Hypertens (Greenwich) 2021, 23, 1380-1389, doi:10.1111/jch.14265.
- Kruman, I.I.; Culmsee, C.; Chan, S.L.; Kruman, Y.; Guo, Z.; Penix, L.; Mattson, M.P. Homocysteine elicits a DNA damage response in neurons that promotes apoptosis and hypersensitivity to excitotoxicity. J Neurosci 2000, 20, 6920-6926, doi:10.1523/JNEUROSCI.20-18-06920.2000.
- Duan, W.; Ladenheim, B.; Cutler, R.G.; Kruman, I.I.; Cadet, J.L.; Mattson, M.P. Dietary folate deficiency and elevated homocysteine levels endanger dopaminergic neurons in models of Parkinson's disease. J Neurochem 2002, 80, 101-110, doi:10.1046/j.0022-3042.2001.00676.x.
- 51. Mattson, M.P. Accomplices to neuronal death. Nature 2002, 415, 377-379, doi:10.1038/415377a.
- 52. Berson, A.; Nativio, R.; Berger, S.L.; Bonini, N.M. Epigenetic Regulation in Neurodegenerative Diseases. Trends Neurosci 2018, 41, 587-598, doi:10.1016/j.tins.2018.05.005.
- 53. Cantoni, G.L.; Mudd, S.H.; Andreoli, V. Affective disorders and S-adenosylmethionine: a new hypothesis. Trends Neurosci 1989, 12, 319-324, doi:10.1016/0166-2236(89)90038-6.
- Lipton, S.A.; Kim, W.K.; Choi, Y.B.; Kumar, S.; D'Emilia, D.M.; Rayudu, P.V.; Arnelle, D.R.; Stamler, J.S. Neurotoxicity associated with dual actions of homocysteine at the N-methyl-D-aspartate receptor. Proc Natl Acad Sci U S A 1997, 94, 5923-5928, doi:10.1073/pnas.94.11.5923.
- 55. Karussis, D. The diagnosis of multiple sclerosis and the various related demyelinating syndromes: a critical review. J Autoimmun 2014, 48-49, 134-142, doi:10.1016/j.jaut.2014.01.022.
- 56. Diaz-Arrastia, R. Homocysteine and neurologic disease. Arch Neurol 2000, 57, 1422-1427, doi:10.1001/archneur.57.10.1422.
- 57. Lyros, E.; Bakogiannis, C.; Liu, Y.; Fassbender, K. Molecular links between endothelial dysfunction and neurodegeneration in Alzheimer's disease. Curr Alzheimer Res 2014, 11, 18-26, doi:10.2174/1567205010666131119235254.
- 58. Ortiz, G.G.; Pacheco-Moisés, F.P.; Macías-Islas, M.; Flores-Alvarado, L.J.; Mireles-Ramírez, M.A.; González-Renovato, E.D.; Hernández-Navarro, V.E.; Sánchez-López, A.L.; Alatorre-Jiménez, M.A. Role of the bloodbrain barrier in multiple sclerosis. Arch Med Res 2014, 45, 687-697, doi:10.1016/j.arcmed.2014.11.013.
- 59. Zhang, X.; Chen, S.; Li, L.; Wang, Q.; Le, W. Decreased level of 5-methyltetrahydrofolate: a potential biomarker for pre-symptomatic amyotrophic lateral sclerosis. J Neurol Sci 2010, 293, 102-105, doi:10.1016/j.jns.2010.02.024.
- 60. Welch, G.N.; Upchurch, G.; Loscalzo, J. Hyperhomocyst(e)inemia and atherothrombosis. Ann N Y Acad Sci 1997, 811, 48-58; discussion 58-49, doi:10.1111/j.1749-6632.1997.tb51988.x.
- 61. Grieve, A.; Butcher, S.P.; Griffiths, R. Synaptosomal plasma membrane transport of excitatory sulphur amino acid transmitter candidates: kinetic characterisation and analysis of carrier specificity. J Neurosci Res 1992, 32, 60-68, doi:10.1002/jnr.490320108.
- 62. Kruman, I.I.; Kumaravel, T.S.; Lohani, A.; Pedersen, W.A.; Cutler, R.G.; Kruman, Y.; Haughey, N.; Lee, J.; Evans, M.; Mattson, M.P. Folic acid deficiency and homocysteine impair DNA repair in hippocampal neurons and sensitize them to amyloid toxicity in experimental models of Alzheimer's disease. J Neurosci 2002, 22, 1752-1762, doi:10.1523/JNEUROSCI.22-05-01752.2002.
- 63. Brosnan, J.T.; Jacobs, R.L.; Stead, L.M.; Brosnan, M.E. Methylation demand: a key determinant of homocysteine metabolism. Acta Biochim Pol 2004, 51, 405-413.
- 64. Ying, H.; Jianping, C.; Jianqing, Y.; Shanquan, Z. Cognitive variations among vascular dementia subtypes caused by small-, large-, or mixed-vessel disease. Arch Med Sci 2016, 12, 747-753, doi:10.5114/aoms.2016.60962.
- Leboeuf, R. Homocysteine and Alzheimer's disease. J Am Diet Assoc 2003, 103, 304-307, doi:10.1053/jada.2003.50083.
- Cascalheira, J.F.; João, S.S.; Pinhanços, S.S.; Castro, R.; Palmeira, M.; Almeida, S.; Faria, M.C.; Domingues, F.C. Serum homocysteine: interplay with other circulating and genetic factors in association to Alzheimer's type dementia. Clin Biochem 2009, 42, 783-790, doi:10.1016/j.clinbiochem.2009.02.006.
- 67. Hooshmand, B.; Solomon, A.; Kåreholt, I.; Leiviskä, J.; Rusanen, M.; Ahtiluoto, S.; Winblad, B.; Laatikainen, T.; Soininen, H.; Kivipelto, M. Homocysteine and holotranscobalamin and the risk of Alzheimer disease: a longitudinal study. Neurology 2010, 75, 1408-1414, doi:10.1212/WNL.0b013e3181f88162.
- Zhou, F.; Chen, S. Hyperhomocysteinemia and risk of incident cognitive outcomes: An updated doseresponse meta-analysis of prospective cohort studies. Ageing Res Rev 2019, 51, 55-66, doi:10.1016/j.arr.2019.02.006.
- 69. Nilsson, K.; Gustafson, L.; Hultberg, B. Plasma homocysteine concentration relates to the severity but not to the duration of Alzheimer's disease. Int J Geriatr Psychiatry 2004, 19, 666-672, doi:10.1002/gps.1140.
- 70. Li, J.G.; Chu, J.; Barrero, C.; Merali, S.; Praticò, D. Homocysteine exacerbates β-amyloid pathology, tau pathology, and cognitive deficit in a mouse model of Alzheimer disease with plaques and tangles. Ann Neurol 2014, 75, 851-863, doi:10.1002/ana.24145.
- 71. Zhao, Y.; Dong, X.; Chen, B.; Zhang, Y.; Meng, S.; Guo, F.; Guo, X.; Zhu, J.; Wang, H.; Cui, H.; et al. Blood levels of circulating methionine components in Alzheimer's disease and mild cognitive impairment: A

- Sah, R.P.; Vidya, C.S.; Pereira, P.; Jayaram, S.; Yadav, A.K.; Sujatha, P. Elevated Homocysteine Level and Brain Atrophy Changes as Markers to Screen the Alzheimer Disease: Case Series. Ann Geriatr Med Res 2024, 28, 116-120, doi:10.4235/agmr.23.0135.
- Zhuo, J.M.; Praticò, D. Normalization of hyperhomocysteinemia improves cognitive deficits and ameliorates brain amyloidosis of a transgenic mouse model of Alzheimer's disease. FASEB J 2010, 24, 3895-3902, doi:10.1096/fi.10-161828.
- 74. Zhu, J.; Wu, Y.; Tang, Q.; Leng, Y.; Cai, W. The effects of choline on hepatic lipid metabolism, mitochondrial function and antioxidative status in human hepatic C3A cells exposed to excessive energy substrates. Nutrients 2014, 6, 2552-2571, doi:10.3390/nu6072552.
- 75. Zhu, J.; Saikia, G.; Zhang, X.; Shen, X.; Kahe, K. One-Carbon Metabolism Nutrients, Genetic Variation, and Diabetes Mellitus. Diabetes Metab J 2024, 48, 170-183, doi:10.4093/dmj.2023.0272.
- Savage, D.G.; Lindenbaum, J.; Stabler, S.P.; Allen, R.H. Sensitivity of serum methylmalonic acid and total homocysteine determinations for diagnosing cobalamin and folate deficiencies. Am J Med 1994, 96, 239-246, doi:10.1016/0002-9343(94)90149-x.
- 77. Huang, T.; Ren, J.; Huang, J.; Li, D. Association of homocysteine with type 2 diabetes: a meta-analysis implementing Mendelian randomization approach. BMC Genomics 2013, 14, 867, doi:10.1186/1471-2164-14-867.
- Al-Maskari, M.Y.; Waly, M.I.; Ali, A.; Al-Shuaibi, Y.S.; Ouhtit, A. Folate and vitamin B12 deficiency and hyperhomocysteinemia promote oxidative stress in adult type 2 diabetes. Nutrition 2012, 28, e23-26, doi:10.1016/j.nut.2012.01.005.
- 79. Cheng, C.K.; Wang, C.; Shang, W.; Lau, C.W.; Luo, J.Y.; Wang, L.; Huang, Y. A high methionine and low folate diet alters glucose homeostasis and gut microbiome. Biochem Biophys Rep 2021, 25, 100921, doi:10.1016/j.bbrep.2021.100921.
- 80. Tessari, P.; Coracina, A.; Kiwanuka, E.; Vedovato, M.; Vettore, M.; Valerio, A.; Zaramella, M.; Garibotto, G. Effects of insulin on methionine and homocysteine kinetics in type 2 diabetes with nephropathy. Diabetes 2005, 54, 2968-2976, doi:10.2337/diabetes.54.10.2968.
- 81. Hultberg, B.; Agardh, E.; Andersson, A.; Brattström, L.; Isaksson, A.; Israelsson, B.; Agardh, C.D. Increased levels of plasma homocysteine are associated with nephropathy, but not severe retinopathy in type 1 diabetes mellitus. Scand J Clin Lab Invest 1991, 51, 277-282, doi:10.3109/00365519109091615.
- 82. Fonseca, V.; Dicker-Brown, A.; Ranganathan, S.; Song, W.; Barnard, R.J.; Fink, L.; Kern, P.A. Effects of a high-fat-sucrose diet on enzymes in homocysteine metabolism in the rat. Metabolism 2000, 49, 736-741, doi:10.1053/meta.2000.6256.
- 83. Kaimala, S.; Ansari, S.A.; Emerald, B.S. DNA methylation in the pathogenesis of type 2 diabetes. Vitam Horm 2023, 122, 147-169, doi:10.1016/bs.vh.2022.11.002.
- 84. Burdge, G.C.; Lillycrop, K.A.; Phillips, E.S.; Slater-Jefferies, J.L.; Jackson, A.A.; Hanson, M.A. Folic acid supplementation during the juvenile-pubertal period in rats modifies the phenotype and epigenotype induced by prenatal nutrition. J Nutr 2009, 139, 1054-1060, doi:10.3945/jn.109.104653.
- 85. McKay, J.A.; Xie, L.; Harris, S.; Wong, Y.K.; Ford, D.; Mathers, J.C. Blood as a surrogate marker for tissue-specific DNA methylation and changes due to folate depletion in post-partum female mice. Mol Nutr Food Res 2011, 55, 1026-1035, doi:10.1002/mnfr.201100008.
- 86. Emoto, M.; Kanda, H.; Shoji, T.; Kawagishi, T.; Komatsu, M.; Mori, K.; Tahara, H.; Ishimura, E.; Inaba, M.; Okuno, Y.; et al. Impact of insulin resistance and nephropathy on homocysteine in type 2 diabetes. Diabetes Care 2001, 24, 533-538, doi:10.2337/diacare.24.3.533.
- 87. Zheng, X.; Liu, Q.; Liu, Z. Serum homocysteine concentration as a marker for advanced diabetic nephropathy in a cohort of elderly patients. BMC Endocr Disord 2023, 23, 114, doi:10.1186/s12902-023-01342-1
- 88. Holven, K.B.; Aukrust, P.; Retterstøl, K.; Otterdal, K.; Bjerkeli, V.; Ose, L.; Nenseter, M.S.; Halvorsen, B. The antiatherogenic function of HDL is impaired in hyperhomocysteinemic subjects. J Nutr 2008, 138, 2070-2075, doi:10.3945/jn.108.090704.
- 89. Soinio, M.; Marniemi, J.; Laakso, M.; Lehto, S.; Rönnemaa, T. Elevated plasma homocysteine level is an independent predictor of coronary heart disease events in patients with type 2 diabetes mellitus. Ann Intern Med 2004, 140, 94-100, doi:10.7326/0003-4819-140-2-200401200-00009.
- 90. Panush, R.S.; Reynolds, R.C.; Benson, J.A.; Lacombe, M.A. Clinical medicine: Perspectives for the future. The American Journal of Medicine 1993, 95, 1-12, doi:https://doi.org/10.1016/0002-9343(93)90225-E.
- 91. Melton, L.J., III. Adverse Outcomes of Osteoporotic Fractures in the General Population*. Journal of Bone and Mineral Research 2003, 18, 1139-1141, doi:10.1359/jbmr.2003.18.6.1139.
- 92. Nuti, R.; Brandi, M.L.; Isaia, G.; Tarantino, U.; Silvestri, S.; Adami, S. New perspectives on the definition and the management of severe osteoporosis: the patient with two or more fragility fractures. J Endocrinol Invest 2009, 32, 783-788, doi:10.1007/bf03346537.

- Enneman, A.W.; van der Velde, N.; de Jonge, R.; Heil, S.G.; Stolk, L.; Hofman, A.; Rivadeneira, F.; Zillikens, M.C.; Uitterlinden, A.G.; van Meurs, J.B.J. The association between plasma homocysteine levels, methylation capacity and incident osteoporotic fractures. Bone 2012, 50, 1401-1405, doi:https://doi.org/10.1016/j.bone.2012.03.013.
- 95. Blouin, S.; Thaler, H.W.; Korninger, C.; Schmid, R.; Hofstaetter, J.G.; Zoehrer, R.; Phipps, R.; Klaushofer, K.; Roschger, P.; Paschalis, E.P. Bone matrix quality and plasma homocysteine levels. Bone 2009, 44, 959-964, doi:https://doi.org/10.1016/j.bone.2008.12.023.
- 96. van Meurs Joyce, B.J.; Dhonukshe-Rutten Rosalie, A.M.; Pluijm Saskia, M.F.; van der Klift, M.; de Jonge, R.; Lindemans, J.; de Groot Lisette, C.P.G.M.; Hofman, A.; Witteman Jacqueline, C.M.; van Leeuwen Johannes, P.T.M.; et al. Homocysteine Levels and the Risk of Osteoporotic Fracture. New England Journal of Medicine 350, 2033-2041, doi:10.1056/NEJMoa032546.
- 97. Gjesdal, C.G.; Vollset, S.E.; Ueland, P.M.; Refsum, H.; Drevon, C.A.; Gjessing, H.K.; Tell, G.S. Plasma total homocysteine level and bone mineral density: the Hordaland Homocysteine Study. Arch Intern Med 2006, 166, 88-94, doi:10.1001/archinte.166.1.88.
- 98. McLean, R.R.; Jacques, P.F.; Selhub, J.; Tucker, K.L.; Samelson, E.J.; Broe, K.E.; Hannan, M.T.; Cupples, L.A.; Kiel, D.P. Homocysteine as a predictive factor for hip fracture in older persons. N Engl J Med 2004, 350, 2042-2049, doi:10.1056/NEJMoa032739.
- 99. Kim, D.J.; Koh, J.M.; Lee, O.; Kim, N.J.; Lee, Y.S.; Kim, Y.S.; Park, J.Y.; Lee, K.U.; Kim, G.S. Homocysteine enhances apoptosis in human bone marrow stromal cells. Bone 2006, 39, 582-590, doi:10.1016/j.bone.2006.03.004.
- 100. Vacek, T.P.; Kalani, A.; Voor, M.J.; Tyagi, S.C.; Tyagi, N. The role of homocysteine in bone remodeling. Clin Chem Lab Med 2013, 51, 579-590, doi:10.1515/cclm-2012-0605.
- 101. Tyagi, N.; Kandel, M.; Munjal, C.; Qipshidze, N.; Vacek, J.C.; Pushpakumar, S.B.; Metreveli, N.; Tyagi, S.C. Homocysteine mediated decrease in bone blood flow and remodeling: role of folic acid. J Orthop Res 2011, 29, 1511-1516, doi:10.1002/jor.21415.
- 102. Behera, J.; Bala, J.; Nuru, M.; Tyagi, S.C.; Tyagi, N. Homocysteine as a Pathological Biomarker for Bone Disease. J Cell Physiol 2017, 232, 2704-2709, doi:10.1002/jcp.25693.
- 103. Rabaneda, L.G.; Geribaldi-Doldán, N.; Murillo-Carretero, M.; Carrasco, M.; Martínez-Salas, J.M.; Verástegui, C.; Castro, C. Altered regulation of the Spry2/Dyrk1A/PP2A triad by homocysteine impairs neural progenitor cell proliferation. Biochim Biophys Acta 2016, 1863, 3015-3026, doi:10.1016/j.bbamcr.2016.09.018.
- 104. Gou, Y.; Ye, Q.; Liang, X.; Zhang, Q.; Luo, S.; Liu, H.; Wang, X.; Sai, N.; Zhang, X. Homocysteine restrains hippocampal neurogenesis in focal ischemic rat brain by inhibiting DNA methylation. Neurochem Int 2021, 147, 105065, doi:10.1016/j.neuint.2021.105065.
- 105. Zhang, D.; Sun, X.; Liu, J.; Xie, X.; Cui, W.; Zhu, Y. Homocysteine accelerates senescence of endothelial cells via DNA hypomethylation of human telomerase reverse transcriptase. Arterioscler Thromb Vasc Biol 2015, 35, 71-78, doi:10.1161/atvbaha.114.303899.
- 106. Allison, J.; Kaliszewska, A.; Uceda, S.; Reiriz, M.; Arias, N. Targeting DNA Methylation in the Adult Brain through Diet. Nutrients 2021, 13, doi:10.3390/nu13113979.
- 107. Majumder, A.; Singh, M.; George, A.K.; Tyagi, S.C. Restoration of skeletal muscle homeostasis by hydrogen sulfide during hyperhomocysteinemia-mediated oxidative/ER stress condition. Can J Physiol Pharmacol 2019, 97, 441-456, doi:10.1139/cjpp-2018-0501.
- 108. Hasan, T.; Arora, R.; Bansal, A.K.; Bhattacharya, R.; Sharma, G.S.; Singh, L.R. Disturbed homocysteine metabolism is associated with cancer. Exp Mol Med 2019, 51, 1-13, doi:10.1038/s12276-019-0216-4.
- 109. Liu, P.; Zhang, M.; Xie, X.; Jin, J.; Holman, C.D. Polymorphisms of 5,10-methylenetetrahydrofolate reductase and thymidylate synthase, dietary folate intake, and the risk of leukemia in adults. Tumour Biol 2016, 37, 3265-3275, doi:10.1007/s13277-015-4168-6.
- 110. Li, X.L.; Xu, J.H. MTHFR polymorphism and the risk of prostate cancer: a meta-analysis of case-control studies. Prostate Cancer Prostatic Dis 2012, 15, 244-249, doi:10.1038/pcan.2012.5.
- 111. Wu, C.Y.; Yang, M.; Lin, M.; Li, L.P.; Wen, X.Z. MTHFR C677T polymorphism was an ethnicity-dependent risk factor for cervical cancer development: evidence based on a meta-analysis. Arch Gynecol Obstet 2013, 288, 595-605, doi:10.1007/s00404-013-2721-3.
- 112. Ma, J.; Stampfer, M.J.; Giovannucci, E.; Artigas, C.; Hunter, D.J.; Fuchs, C.; Willett, W.C.; Selhub, J.; Hennekens, C.H.; Rozen, R. Methylenetetrahydrofolate reductase polymorphism, dietary interactions, and risk of colorectal cancer. Cancer Res 1997, 57, 1098-1102.
- 113. Esteller, M.; Garcia, A.; Martinez-Palones, J.M.; Xercavins, J.; Reventos, J. Germ line polymorphisms in cytochrome-P450 1A1 (C4887 CYP1A1) and methylenetetrahydrofolate reductase (MTHFR) genes and endometrial cancer susceptibility. Carcinogenesis 1997, 18, 2307-2311, doi:10.1093/carcin/18.12.2307.

- 115. Matsuo, K.; Hamajima, N.; Hirai, T.; Kato, T.; Inoue, M.; Takezaki, T.; Tajima, K. Methionine Synthase Reductase Gene A66G Polymorphism is Associated with Risk of Colorectal Cancer. Asian Pac J Cancer Prev 2002, 3, 353-359.
- Chen, K.; Song, L.; Jin, M.J.; Fan, C.H.; Jiang, Q.T.; Yu, W.P. [Association between genetic polymorphisms in folate metabolic enzyme genes and colorectal cancer: a nested case-control study]. Zhonghua Zhong Liu Za Zhi 2006, 28, 429-432.
- 117. Liu, Z.; Cui, C.; Wang, X.; Fernandez-Escobar, A.; Wu, Q.; Xu, K.; Mao, J.; Jin, M.; Wang, K. Plasma Levels of Homocysteine and the Occurrence and Progression of Rectal Cancer. Med Sci Monit 2018, 24, 1776-1783, doi:10.12659/MSM.909217.
- 118. Xu, J.; Zhao, X.; Sun, S.; Ni, P.; Li, C.; Ren, A.; Wang, W.; Zhu, L. Homocysteine and Digestive Tract Cancer Risk: A Dose-Response Meta-Analysis. J Oncol 2018, 2018, 3720684, doi:10.1155/2018/3720684.
- 119. Maddocks, O.D.; Berkers, C.R.; Mason, S.M.; Zheng, L.; Blyth, K.; Gottlieb, E.; Vousden, K.H. Serine starvation induces stress and p53-dependent metabolic remodelling in cancer cells. Nature 2013, 493, 542-546, doi:10.1038/nature11743.
- 120. Kanarek, N.; Keys, H.R.; Cantor, J.R.; Lewis, C.A.; Chan, S.H.; Kunchok, T.; Abu-Remaileh, M.; Freinkman, E.; Schweitzer, L.D.; Sabatini, D.M. Author Correction: Histidine catabolism is a major determinant of methotrexate sensitivity. Nature 2022, 602, E17-E18, doi:10.1038/s41586-021-03487-2.
- 121. Knott, S.R.V.; Wagenblast, E.; Khan, S.; Kim, S.Y.; Soto, M.; Wagner, M.; Turgeon, M.O.; Fish, L.; Erard, N.; Gable, A.L.; et al. Asparagine bioavailability governs metastasis in a model of breast cancer. Nature 2018, 554, 378-381, doi:10.1038/nature25465.
- 122. Ser, Z.; Gao, X.; Johnson, C.; Mehrmohamadi, M.; Liu, X.; Li, S.; Locasale, J.W. Targeting One Carbon Metabolism with an Antimetabolite Disrupts Pyrimidine Homeostasis and Induces Nucleotide Overflow. Cell Rep 2016, 15, 2367-2376, doi:10.1016/j.celrep.2016.05.035.
- 123. Locasale, J.W. Serine, glycine and one-carbon units: cancer metabolism in full circle. Nat Rev Cancer 2013, 13, 572-583, doi:10.1038/nrc3557.
- 124. Cellarier, E.; Durando, X.; Vasson, M.P.; Farges, M.C.; Demiden, A.; Maurizis, J.C.; Madelmont, J.C.; Chollet, P. Methionine dependency and cancer treatment. Cancer Treat Rev 2003, 29, 489-499, doi:10.1016/s0305-7372(03)00118-x.
- 125. Altea-Manzano, P.; Cuadros, A.M.; Broadfield, L.A.; Fendt, S.M. Nutrient metabolism and cancer in the in vivo context: a metabolic game of give and take. EMBO Rep 2020, 21, e50635, doi:10.15252/embr.202050635.
- 126. Peng, H.; Yan, Y.; He, M.; Li, J.; Wang, L.; Jia, W.; Yang, L.; Jiang, J.; Chen, Y.; Li, F.; et al. SLC43A2 and NFκB signaling pathway regulate methionine/cystine restriction-induced ferroptosis in esophageal squamous cell carcinoma via a feedback loop. Cell Death Dis 2023, 14, 347, doi:10.1038/s41419-023-05860-7
- 127. Shingler, E.; Perry, R.; Mitchell, A.; England, C.; Perks, C.; Herbert, G.; Ness, A.; Atkinson, C. Dietary restriction during the treatment of cancer: results of a systematic scoping review. BMC Cancer 2019, 19, 811, doi:10.1186/s12885-019-5931-7.
- 128. Lee, B.C.; Kaya, A.; Ma, S.; Kim, G.; Gerashchenko, M.V.; Yim, S.H.; Hu, Z.; Harshman, L.G.; Gladyshev, V.N. Methionine restriction extends lifespan of Drosophila melanogaster under conditions of low aminoacid status. Nat Commun 2014, 5, 3592, doi:10.1038/ncomms4592.
- 129. Malloy, V.L.; Perrone, C.E.; Mattocks, D.A.; Ables, G.P.; Caliendo, N.S.; Orentreich, D.S.; Orentreich, N. Methionine restriction prevents the progression of hepatic steatosis in leptin-deficient obese mice. Metabolism 2013, 62, 1651-1661, doi:10.1016/j.metabol.2013.06.012.
- 130. Hoffman, R.M.; Erbe, R.W. High in vivo rates of methionine biosynthesis in transformed human and malignant rat cells auxotrophic for methionine. Proc Natl Acad Sci U S A 1976, 73, 1523-1527, doi:10.1073/pnas.73.5.1523.
- 131. Komninou, D.; Leutzinger, Y.; Reddy, B.S.; Richie, J.P. Methionine restriction inhibits colon carcinogenesis. Nutr Cancer 2006, 54, 202-208, doi:10.1207/s15327914nc5402_6.
- 132. Nygård, O.; Vollset, S.E.; Refsum, H.; Stensvold, I.; Tverdal, A.; Nordrehaug, J.E.; Ueland, M.; Kvåle, G. Total plasma homocysteine and cardiovascular risk profile. The Hordaland Homocysteine Study. JAMA 1995, 274, 1526-1533, doi:10.1001/jama.1995.03530190040032.
- 133. Stanger, O.; Herrmann, W.; Pietrzik, K.; Fowler, B.; Geisel, J.; Dierkes, J.; Weger, M.; e.V, D.-L.H. DACH-LIGA homocystein (german, austrian and swiss homocysteine society): consensus paper on the rational clinical use of homocysteine, folic acid and B-vitamins in cardiovascular and thrombotic diseases: guidelines and recommendations. Clin Chem Lab Med 2003, 41, 1392-1403, doi:10.1515/CCLM.2003.214.
- 134. Liang, Z.; Li, K.; Chen, H.; Jia, J.; Li, J.; Huo, Y.; Fan, F.; Zhang, Y. The Association of Plasma Homocysteine Concentrations with a 10-Year Risk of All-Cause and Cardiovascular Mortality in a Community-Based Chinese Population. Nutrients 2024, 16, doi:10.3390/nu16121945.

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