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Relevance of the Injection Model and IA Task to Study the Effects of $A\beta_{1-42}$ Oligomers on Contextual Memory and Dorsal CA1 Pyramidal Neurons

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Relevance of the Injection Model and IA Task to Study the Effects of Aβ₁₋₄₂ Oligomers on Contextual Memory and Dorsal CA1 Pyramidal Neurons

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Abstract: The transgenic models have been yielding invaluable insights into amyloid pathology as they mimic the nature of Alzheimer's disease (AD). However, there is no clear relationship between senile plaques and memory deficits. Instead, cognitive impairment and synaptic dysfunction are particularly linked to a rise in A β_{1-42} oligomer level. Thus, injection of A β_{1-42} oligomers into a specific brain region is considered as an alternative to investigate the effects of increased soluble Aβ species without any plaques owing to its controllability, credibility and validity. Despite the homogeneous cytoarchitecture, the hippocampal CA1 (Cornu ammonis 1) region displays topographical heterogeneity and is differentially affected in the early stage of AD. Moreover, specific targeting of CA1 subregion is important not to miss axis-linked implications in the behavioral consequences. Next, inhibitory avoidance (IA) task is a relevant learning paradigm to assess synaptic basis of hippocampal CA1-dependent contextual memory formation. This one-trial task is based on associating the training context with the aversive stimulus after the foot shock and triggers trainingdependent synaptic plasticity similar to in vitro HFS (High-frequency stimulation). Therefore, in this review, we will focus on why injection of $A\beta_{1-42}$ oligomers is a valid in vivo model to investigate the early stage of AD and why dorsal CA1 region is a target area to understand the adverse effects of $A\beta_{1-42}$ oligomers on contextual learning in the IA task.

Keywords: injection model; Aβ₁₋₄₂ oligomers; dorsal CA1; IA task; contextual memory

1. Introduction

Alzheimer's disease (AD) is clinically characterized by an early deficit in episodic memory, concerning spatiotemporal context [1,2], along with extracellular senile plaques and intracellular neurofibrillary tangles (NFT) [3]. The amyloid plaques harbor heterogeneous aggregates of amyloid beta (A β) peptides containing 40 (A β ₁₋₄₀) or 42 (A β ₁₋₄₂) amino acid residues. The amyloid cascade hypothesis initially suggested that A β deposition was the primary trigger for AD as it was found to precede and influence the tau pathology [4]. Later, findings revealed that cognitive impairment [5] and synaptic dysfunction [6] were correlated and coincided with a rise in the level of A β oligomers [7], even without the formation of amyloid plaques. Consequently, the earliest amyloid toxicities in AD have been modified to be associated with soluble A β species [8,9].

The generation of transgenic animals overexpressing proteins related to familial AD, mutant APP (amyloid precursor protein) and PS (presenilin) is based on the amyloid hypothesis that cognitive deficit was linked to plaque formation [10]. With an attempt to replicate the hallmarks of AD including senile plaques, these animals develop age-dependent amyloid pathology in a spatiotemporal manner which closely mimics disease progression in human [4,11]. However, both

intrinsic and extrinsic factors modulate the course and outcome of these models [12]. More importantly, the presence of amyloid plaques may not account for memory deficits. Moreover, memory impairment cannot also be directly related to a specific dysfunction of a particular brain area [11].

On the other hand, injection of $A\beta$ oligomers into a specific brain region is required as an alternative to understand the effects of increased soluble $A\beta$ species without the presence of any plaques [1]. Unlike the transgenic model, the injection model allows us to control the concentration of $A\beta$ oligomers spatiotemporally and recapitulate the cardinal features of early AD [1]. Using wild-type strain also provides an excellent in vivo model as it can circumvent the potential side effects from the gene mutations. In this context, adverse effects associated with the early $A\beta$ pathology could be identified, thereby preventing cognitive impairment and interfering with advanced neurodegenerative processes [12].

Therefore, we will review why injection of $A\beta_{1-42}$ oligomers is a valid model to investigate early stage of AD pathology and why dorsal CA1 region is a target area to understand the adverse effects of $A\beta_{1-42}$ oligomers related to contextual memory.

2. Transgenic Model vs. Injection Model

Transgenic animals, mice and rats [13,14], which express the mutant forms of APP and/or PS gene have enabled us to study AD-related pathologies. These models share a common feature of increased A β production, leading to formation and deposition of amyloid plaques [15]. By recapitulating the neuropathological features of AD [10] and exhibiting a chronic, progressive spectrum of disease [16], these models appear to phenocopy the critical aspects of human AD. Thus, they have represented a promising niche for investigating A β pathology and testing the efficacy of therapeutic interventions.

However, these models are based on familial AD-associated gene mutations which only account for < 1% of AD cases [13]. By contrast, clinical AD results from a complex interaction of multiple etiologies, rather than a single involvement of genes [17]. Next, using transgenic animals is time-consuming because it takes months or years to develop A β plaques which do not have a clear relationship with cognitive impairments. Resembling advanced AD, they are lacking early AD-associated pathological changes. The appearance of AD-related phenotypes could be confounded by several factors such as species, age, etc [12]. More importantly, they develop AD phenotypes not in chronological order and thus are not equivalent to human AD. For example, plaque deposition is preceded by cellular changes and cognitive impairments [10].

To overcome the shortcomings of the transgenic models, injection of $A\beta$ oligomers has been alternatively used. Firstly, the dose of oligomers, the route of injection and the age of animals can be adjusted to minimize the experimental contributing factors. Next, the wild-type strain can circumvent the by-stander effects of the recipient genes derived from introduction of exogenous genes' mutations [16]. Moreover, the key advantage of this model can be partly explained by the facts that any brain region can be targeted [1] and the oligomers are rapidly delivered to the area of interest [18]. Most importantly, the greatest benefit of this $A\beta$ -injected model is its credibility as pathological changes are directly triggered by the $A\beta$ oligomers.

As a rise in soluble $A\beta$ species in a certain brain area has been regarded as a starting phenomenon in AD, the oligomer injection is a valid model to investigate the early stage of AD pathology [1,15]. Injecting either natural or synthetic $A\beta_{1-42}$ oligomers can trigger synaptic dysfunction, astrogliosis and cognitive deficits [18–20]. Moreover, these features can be directly linked to specific dysfunction of a particular brain area due to amyloid deposition [18,21]. However, having considered the chronic nature of AD rather than a sudden surge of oligomers, the transgenic models are also recommended to strengthen the findings from the $A\beta$ oligomer-injected models [16].

3. Why Is Understanding Hippocampal CA1 Region Important?

A typical CA1 (Cornu ammonis 1) pyramidal neuron (CA1 PN) receives most glutamatergic inputs from the entorhinal cortex (EC). The classical trisynaptic circuitry originates from projections of the EC layer 2 stellate cells, via perforant path (PP), to dentate gyrus (DG) granule cells whose mossy fibers synapse with CA3 PNs, which in turn excite CA1 PNs through the Schaffer collateral (SC) pathway. In addition, direct glutamatergic projections from EC layer 3 PNs also reach CA1 PNs through the temporo-ammonic (TA) pathway. The trisynaptic and monosynaptic circuits convergence at the proximal and distal apical dendrites of CA1 PNs, respectively [22,23]. Finally, CA1 integrates non-spatial information from the lateral entorhinal cortex (LEC) and spatial information from the medial entorhinal cortex (MEC) and provides the primary output of the hippocampus, along with subiculum to the extrahippocampal circuits (Figure 1A) [22].

The hippocampal CA1 region shows functional, intrinsic and synaptic heterogeneities across the longitudinal (dorso-ventral), transverse (proximo-distal) and radial (deep-superficial) axes despite its homogeneous cytoarchitecture [24,25]. Firstly, dorsal CA1 is particularly engaged in the formation of spatial and contextual memories [26] whereas ventral CA1 PNs mediate social-related cues [27]. Along the transverse axis, proximal CA1 PNs exhibit specific firing towards spatial information [28] whereas activation of distal CA1 PNs corresponds to object memory encoding [29,30]. Compared to superficial PNs, deep CA1 PNs are more likely to have place fields and are highly activated during spatial processing (Figure 1B) [31]. This functional diversity reflects differential behavioral role of the hippocampal CA1 subfields in different axes.

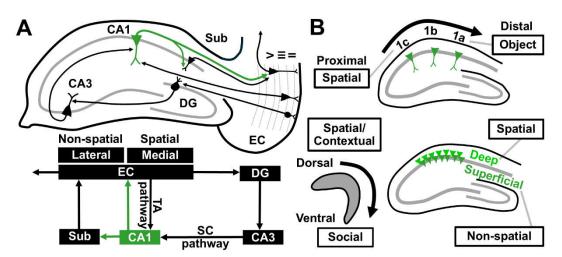


Figure 1. Schematic diagrams illustrating the hippocampal circuit and functional heterogeneity along the anatomical axes. (A) The circuit diagram of hippocampal CA1 region. The classical hippocampal circuit comprises of indirect and direct inputs from EC Layer II and III, via the SC and TA pathways, to CA1 which sends projection back to the EC Layer V. The indirect and direct pathways convergence at the proximal and distal apical dendrites of CA1 PN, respectively (upper panel). CA1 integrates non-spatial information from LEC and spatial information from MEC and provides the primary output of the hippocampus, along with subiculum (lower panel). (B) Functional heterogeneity of CA1 across the longitudinal, transverse and radial axes. The left panel illustrates that dorsal CA1 is particularly engaged in spatial/contextual memory whereas ventral CA1 mediates social-related cues. The upper right panel shows that proximal CA1 exhibits specific firing towards spatial information whereas distal CA1 PNs are more responsive to object encoding. The lower right panel represents that deep CA1 PNs are highly activated during spatial encoding whilst superficial neurons are specialized for processing of non-spatial information. CA, Cornu ammonis; DG, Dentate gyrus; EC, Entorhinal cortex; LEC, Lateral entorhinal cortex; MEC, Medial entorhinal cortex; PN, Pyramidal neuron; SC, Schaffer collateral; Sub, Subiculum; TA, Temporo-ammonic.

These in vivo functional differences can also be contributed by variations in the input-output function of CA1 PNs. Fundamentally, a neuron integrates synaptic inputs and decides whether to

fire an action potential (AP) or not. The intrinsic excitability (IE) determines its likelihood of firing in response to synaptic inputs and is therefore a crucial link between synaptic input and action potential output (Figure 2A) [32]. Though controlled by independent mechanisms, both forms of plasticity are interdependent [33]. Indeed, both are tightly coupled and bi-directionally adjusted in CA1 PNs so that neuronal activity is maintained to prevent extreme modifications [34].

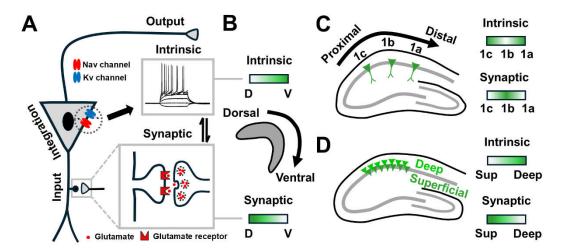


Figure 2. Schematic diagrams illustrating the input-output function and topographical heterogeneity of CA1 along its anatomical axes. (A) The input-output function of CA1 PN. A neuron receives information via synapse and integrates it at the soma. Its intrinsic properties determine the likelihood of firing AP, thus serving as a crucial link between synaptic input and AP output. Intrinsic excitability can be modulated by passive and active membrane properties, including voltage-gated ion channels. Glutamatergic transmission can be modified via either pre-synaptic glutamate release or post-synaptic glutamate receptor expression. Though controlled by independent mechanisms, both forms of plasticity are tightly coupled and bi-directionally adjusted so that neuronal activity is maintained to prevent extreme changes. (B) Intrinsic and synaptic heterogeneity of CA1 across the longitudinal axis. Ventral CA1 PNs display a weaker and less stable synaptic plasticity but are more excitable than dorsal PNs. (C) Intrinsic and synaptic heterogeneity of CA1 across the transverse axis. In the dorsal hippocampus, similar synaptic properties are found at proximal and distal CA1 PNs together with equivalent excitability whereas CA1b is distinguished by having lower intrinsic excitability with higher synaptic integrating properties. (D) Intrinsic and synaptic heterogeneity of CA1 across the radial axis. Synaptic plasticity of CA1 PNs is more robust in the superficial layer whereas deep CA1 PNs are capable of eliciting higher firing. AP, Action potential; CA, Cornu ammonis; D, Dorsal; Kv channel, Voltage-gated potassium channel; Nav, Voltage-gated sodium channel; PN, Pyramidal neuron; Sup, Superficial; V, Ventral.

Yet, how these properties can be directly translated to in vivo responses is controversial owing to the striking topographical differences [35]. For example, ventral CA1 PNs form a weaker and less stable synaptic plasticity [36,37], but are also more excitable than dorsal PNs (Figure 2B) [38]. In the dorsal hippocampus, similar synaptic and intrinsic properties are found at CA1a and CA1c PNs whereas CA1b is distinguished by having lower intrinsic excitability with higher synaptic integrating properties (Figure 2C) [35]. Next, synaptic plasticity of CA1 PNs is more robust in the superficial layer [39] whereas deep CA1 PNs elicit higher firing in the dorsal region (Figure 2D) [35]. These findings suggest that these axis-dependent differences may be related to the expression/function of ion channels as well as receptors underlying intrinsic and synaptic properties.

4. Why Is Hippocampal CA1 Relevant as a Target Area to Study AD Pathology?

The CA1 region, which integrates and processes contextual [40] and spatial information [41], is affected in in the early stage of AD. Interestingly, it shows spatiotemporal pattern of amyloid deposition along its anatomical axes [24]. For example, ventral CA1 has a higher amyloid plaque burden than its dorsal pole [42]. Across the transverse axis, amyloid deposits appear more

abundantly in the distal CA1 region than the proximal CA1 [43]. However, the impact of amyloid on the radial axis is still unknown. Taken together, CA1 PNs exhibit axis-linked differential susceptibility to AD pathology.

Selective vulnerability of CA1 PNs with non-uniform functions may underline selective loss of a particular modality of memory. In the context of functional in vivo consequences, spatial memory deficits can be related to specific dysfunction of dorsal CA1 PNs. Place cells in the dorsal CA1 became damaged along with nearby A β aggregates [2] which was accompanied by spatial memory deficits [44]. Yet, there is no clear evidence to characterize such functional impairments in ventral CA1 and across the other two axes.

Owing to the contrasting heterogeneities of CA1, specific targeting of its subfield is important not to miss axis-linked implications in the functional consequences of AD. This leads to direct injection of oligomeric A β_{1-42} into the dorsal CA1 region which shows A β_{1-42} oligomer-induced deficits in spatial working memory [1,45], in association with synaptic dysfunction [46] and A β deposition at the injected site [47]. Thus, this CA1 injection model which directly links A β oligomers with amyloid-related pathology is valid to advance our understanding of cognitive deficits in AD.

5. Effect of Injecting Aβ₁₋₄₂ Oligomers into Dorsal CA1 on Contextual Memory

Here, we will focus on contextual memory because of the rapid and stable development of associative learning and its clear relationship with dorsal CA1 [48]. The inhibitory avoidance (IA) task, a learning paradigm for contextual memory, is based on associating the training context with the aversive stimulus after the foot shock [49]. Briefly, contextual learning requires a certain period of exploration prior to shock during which spatial and non-spatial features of the environment are integrated. After receiving shock, if the animals have learnt the context-shock pairing, they will stay in the lit area which they would normally not prefer. Thus, the prolonged latency to enter the dark box indicates formation of contextual memory.

Several studies supported that encoding and retrieval of contextual memory require dorsal CA1 [26,27]. Similar to in vitro HFS (high-frequency stimulation), induction of NMDA (N-methyl-D-aspartate)-dependent LTP (long-term potentiation) [48] is followed by delivery of GluR1-containing AMPARs (α -amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid receptor) to the CA3-CA1 synapses [50], resulting in increased number of AMPARs [49] after contextual learning. Moreover, formation of contextual memory strengthens synaptic transmission at both excitatory (AMPAR) and inhibitory (GABAAR) (γ -aminobutyric acid receptor) synapses [49,51] under the cholinergic control [51]. Next, enhanced plasticity in the CA1b region suggests processing and integrating spatial and non-spatial information necessary for establishing contextual memory [52]. That is why this one-trial IA task is a relevant learning paradigm to study the adverse effects of oligomers on the dorsal CA1 region.

As CA1b displays robust processing of its mixed input [35] and assembles information necessary for IA learning [52], we aimed to elucidate the effects of $A\beta_{1-42}$ oligomers on contextual memory by injecting into this target region. Previously, we found that unilateral injection of $A\beta_{1-42}$ oligomers did not affect learning performance whereas bilateral injection shortened the latency (Figure 3) [53]. This was in accordance with our previous finding that bilateral dorsal CA1 is required for contextual learning [51]. We also assured that this contextual impairment was not influenced by motor, sensory and emotional changes. More importantly, this $A\beta_{1-42}$ oligomer-induced behavioral consequence can be directly related to dorsal CA1 dysfunction as the electrophysiological properties of CA1 PNs were adversely affected along with concomitant deposition of $A\beta_{1-42}$ oligomers in the target region [53].

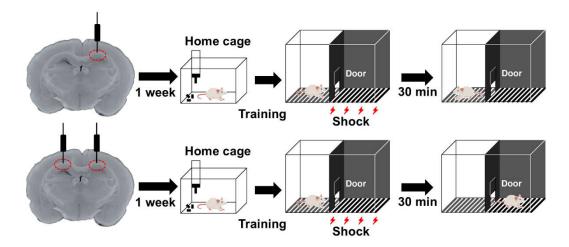


Figure 3. Effect of $A\beta_{1-42}$ oligomers on inhibitory avoidance (IA) task. On the training day, a brief electrical foot shock (1.6 mA, 2 s) was applied when the rats entered the dark side of IA box. After 30 minutes, they were placed in the light compartment and the latency to enter the dark side was measured. The prolonged latency after shock indicates formation of contextual memory. Though unilateral injection did not affect learning performance (upper panel), a shorter latency was observed after bilateral injection of $A\beta_{42-1}$ oligomers into the dorsal CA1 (lower panel).

6. Conclusions

Significant progress has been made to explore the relationship of A β oligomers with specific dysfunction of hippocampal CA1 region. Linking the diversity of CA1 PNs to their functional outcome and specifying the adverse effects of A β ₁₋₄₂ oligomers on dorsal CA1 will lead to a better understanding of episodic memory impairment in AD and will provide a niche for therapeutic interventions.

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Conflicts of Interest: The authors declare no conflicts of interest.

Abbreviations

Aβ Amyloid beta AD Alzheimer's disease

AMPAR α -amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid receptor

AP Action potential

APP Amyloid precursor protein

CA1 Cornu ammonis 1
DG Dentate gyrus
EC Entorhinal cortex

GABAAR γ-aminobutyric acid receptor
HFS High-frequency stimulation
IA task Inhibitory avoidance task
IE Intrinsic excitability

Kv channel Voltage-gated potassium channel

LEC Lateral entorhinal cortex
LTP Long-term potentiation
MEC Medial entorhinal cortex
Nav channel Voltage-gated sodium channel

NFT Neurofibrillary tangle NMDA N-methyl-D-aspartate PN Pyramidal neuron PP Perforant path

Presenilin

SC Schaffer collateral TA Temporo-ammonic

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PS

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