

**Article** 

Not peer-reviewed version

# The HSP70 Modulators in the Correction of Cognitive, Mnemonic, and Behavioral Disorders after Prenatal Hypoxia

<u>Olena Gennadiivna Aliyeva</u>, <u>Igor F. Belenichev</u>\*, Ivan Bilai, Iryna Duiun, Lyudmyla Makyeyeva, <u>Valentyn Oksenych</u>\*, <u>Oleksandr Kamyshnyi</u>

Posted Date: 14 March 2025

doi: 10.20944/preprints202503.1094.v1

Keywords: prenatal hypoxia; HSP70 modulators; open field test; cognitive disorders; memory disorders; behavioral disorders



Preprints.org is a free multidisciplinary platform providing preprint service that is dedicated to making early versions of research outputs permanently available and citable. Preprints posted at Preprints.org appear in Web of Science, Crossref, Google Scholar, Scilit, Europe PMC.

Copyright: This open access article is published under a Creative Commons CC BY 4.0 license, which permit the free download, distribution, and reuse, provided that the author and preprint are cited in any reuse.

Disclaimer/Publisher's Note: The statements, opinions, and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions, or products referred to in the content.

Article

## The HSP70 Modulators in the Correction of Cognitive, Mnemonic, and Behavioral Disorders after Prenatal Hypoxia

Olena Aliyeva <sup>1</sup>, Igor F. Belenichev <sup>2\*</sup>, Ivan Bilai <sup>3</sup>, Iryna Duiun <sup>3</sup>, Lyudmyla Makyeyeva <sup>1</sup>, Valentyn Oksenych <sup>5,\*</sup> and Oleksandr Kamyshnyi <sup>6</sup>

- Department of Histology, Cytology and Embryology State Medical and Pharmaceutical University, 69035 Zaporizhzhia, Ukraine
- <sup>2</sup> Department of Pharmacology and Medical Formulation with Course of Normal Physiology, Zaporizhzhia State Medical and Pharmaceutical University, 69035 Zaporizhzhia, Ukraine
- Department of Clinical Pharmacy, Pharmacotherapy, Pharmacognosy and Pharmaceutical Chemistry, Zaporizhzhia State Medical and Pharmaceutical University, 69035 Zaporizhzhia, Ukraine
- <sup>4</sup> Broegelmann Research Laboratory, Department of Clinical Science, University of Bergen, 5020 Bergen, Norway
- <sup>5</sup> Faculty of Medicine, University of Bergen, 5020 Bergen, Norway
- Department of Microbiology, Virology, and Immunology, I. Horbachevsky Ternopil National Medical University, 46001 Ternopil, Ukraine
- \* Correspondence: i.belenichev1914@gmail.com (I.F.B.) and valentyn.oksenych@uib.no (V.O.)

Abstract: Prenatal hypoxia (PH) is a major cause of nervous system disorders in early childhood, leading to reduced learning and memory capabilities, increased anxiety, and a higher risk of developing mental and neurodegenerative diseases later in life. Compensatory-adaptive mechanisms of the mother-placenta-fetus system, which enhance the fetus's CNS resilience, are known, including the activation of endogenous neuroprotection in response to hypoxic brain injury through pharmacological modulation of HSP70. To evaluate the effect of HSP70 modulators—Cerebrocurin, Angiolin, Tamoxifen, Glutaredoxin, Thiotriazoline, HSF-1 (heat shock factor 1 protein), as well as Mildronate and Mexidol—on motor, exploratory, psycho-emotional activity, learning, and memory of offspring after PH. Experimental PH was induced by daily intraperitoneal injections of sodium nitrite solution to pregnant female rats from the 16th to the 21st day of pregnancy at a dose of 50 mg/kg. Newborns received intraperitoneal injections of Angiolin (50 mg/kg), Thiotriazoline (50 mg/kg), Mexidol (100 mg/kg), Cerebrocurin (150 μL/kg), L-arginine (200 mg/kg), Glutaredoxin (200 μg/kg), HSF-1 (50 mg/kg), and Mildronate (50 mg/kg) for 30 days. At 1 month, rats were tested in the open field test, and at 2 months, they were trained and tested for working and spatial memory in the radial maze. Modeling PH led to persistent impairments in exploratory activity, psycho-emotional behavior, and a decrease in cognitive-mnestic functions of the CNS. It was found that Angiolin and Cerebrocurin had the most pronounced effects on the indicators of exploratory-activity and psychoemotional status in 1-month-old animals after PH. They also exhibited the most significant cognitiveenhancing and memory-supporting effects during training and evaluation of skill retention in the maze in 2-month-old offspring after PH. The obtained results provide experimental justification for further research into Angiolin and Cerebrocurin as promising neuroprotective agents for use after PH. Obtained results will help to expand our understanding of the effect of beta-blockers of various generations used to treat cardiovascular diseases on energy metabolism, and are also an experimental justification for the practical choice of these drugs in the complex therapy of CHF.

**Keywords:** prenatal hypoxia; HSP<sub>70</sub> modulators; open field test; cognitive disorders; memory disorders; behavioral disorders

#### 1. Introduction

The course of intrauterine development and the neonatal period largely determine the future health status and quality of life of an individual. Prenatal hypoxia (PH) is the main cause of prenatal cerebral pathology and neonatal mortality. [1-3]. In the structure of neonatal mortality, hypoxia ranks second after prematurity. Approximately 12-23% of infant mortality worldwide (0.7-1.2 million annually) is registered as a result of prenatal hypoxia, with 0.5 million children becoming disabled [4,5]. Doctors have specific medications for treating the consequences of hypoxic-ischemic damage to the central nervous system. However, the high mortality and disability rates in children indicate that these pharmacological agents are not fully effective. Therefore, a priority task in modern pharmacology is the development of new approaches for the pharmacological correction of neurological disorders resulting from PH [6-8]. In most cases, the cause of the fetus's hypoxic condition is placental insufficiency, maternal diseases, and infectious-inflammatory processes [9]. The leading mechanisms of disorders resulting from prenatal hypoxia are excitotoxicity, oxidative stress, and inflammation [10,11]. Glutamate excitotoxicity leads to nitrosative stress [12,13]. Toxic byproducts of nitric oxide metabolism, formed in cells with the involvement of nNOS and iNOS, along with reactive oxygen species (ROS), bind to important macromolecules, damaging cells and activating signaling molecules of apoptosis [14,15]. Such disturbances resulting from intrauterine hypoxia lead to neurological disorders and impairments in the nervous system in early childhood, reduced learning and memory abilities, anxiety, and an increased risk of developing mental and neurodegenerative diseases in later stages of life [16-18]. Neurons have the ability to adapt to changes caused by chemical and physical factors acting on them. The adaptation of neurons to hypoxia at the cellular and subcellular levels is regulated by the specific transcription factor HIF-1 [19-21]. The ultimate result of HIF-1 activation is an increase in oxygen delivery to the cells [22,23]. One of the primary genomic responses of cells to hypoxia and energy stress is the induction of heat shock protein (HSP), whose effect on stabilizing HIF-1 ensures the activation of processes such as proliferation, apoptosis, and angiogenesis under ischemic conditions [24,25]. Various studies, including ours, have demonstrated the compensatory-adaptive mechanisms of the mother-placenta-fetus system that enhance the fetal central nervous system's resilience. Among these mechanisms is the activation of endogenous neuroprotection in response to hypoxic brain injury, with HSP70 playing an active role. Additionally, pharmacological modulation of HSP70 is shown to be a promising direction for neuroprotection following prenatal hypoxia. [26,27]. We have obtained convincing data demonstrating the effectiveness of HSP70 modulators - Cerebrocurin, Angiolin, Tamoxifen, Glutaredoxin, Thiotriazoline, HSF-1 (heat shock factor 1 protein), as well as Mildronate and Mexidol in experimental neuroprotection following prenatal hypoxia. The use of these agents led to a reduction in mortality during the early postnatal period, a decrease in markers of oxidative and nitrative stress, and an increase in the expression and concentration of HSP70 and HIF-1 in the cytosolic and mitochondrial fractions of the brain in animals after PH [28-30]. There is evidence of the protective effect of HSP70 on higher CNS functions under various adverse influences on the brain [31,32]. All of this has served as the experimental basis for further preclinical studies of HSP70 modulators in prenatal hypoxia, specifically to investigate their impact on cognitive, memory, and behavioral disorders in offspring.

Objective of the study: to assess the impact of HSP<sub>70</sub> modulators – Cerebrocurin, Angiolin, Tamoxifen, Glutaredoxin, Thiotriazoline, HSF-1 (heat shock factor 1 protein), as well as Mildronate and Mexidol – on motor activity, exploratory behavior, psycho-emotional activity, learning, and memory performance in offspring following PH.

### 2. Materials and Methods

The study was conducted in accordance with Directive 2010/63/EU of the European Parliament and Council of September 22, 2010, on the protection of animals used for scientific purposes, as well as the national "General Ethical Principles of Animal Experimentation" (Ukraine, 2001) and the guidelines outlined in the "Basic Methods for Studying the Toxicity of Potential Pharmacological Drugs" (HEC of Ukraine, Kyiv, 2000). The experiment was approved by the Bioethics Commission of Zaporizhzhia State Medical University (protocol No. 32 dated November 16, 2021).

The experimental study was carried out on 50 female and 10 male white rats weighing 220-240 g and aged 4.5 months, obtained from the vivarium of the State Enterprise "Institute of Pharmacology and Toxicology of the National Academy of Medical Sciences of Ukraine." A model of chronic hypoxia induced by sodium nitrite during pregnancy was used for the experiment, which was reproduced by daily intraperitoneal administration of sodium nitrite solution to pregnant female rats from days 16 to 21 of pregnancy at a dose of 50 mg/kg [29]. In the control group, pregnant females were administered physiological saline in the same regimen.

The offspring were divided into groups as follows:

- 1. Group 1: Healthy animals from females with a physiologically normal weight, administered physiological saline intraperitoneally at a dose of  $5 \mu l/g$ ;
- 2. Group 2: Control group after prenatal hypoxia (PH), administered physiological saline intraperitoneally at a dose of  $5 \mu l/g$ ;
- 3. Group 3: After PH, administered Angioline intraperitoneally at a dose of 50 mg/kg;
- Group 4: After PH, administered Piracetam intraperitoneally at a dose of 500 mg/kg;
- 5. Group 5: After PH, administered Tamoxifen intranasally at a dose of 0.1 mg/kg;
- 6. Group 6: After PH, administered Thiotriazoline intraperitoneally at a dose of 50 mg/kg;
- 7. Group 7: After PH, administered Mexidol (nicomex) intraperitoneally at a dose of 100 mg/kg;
- Group 8: After PH, administered Cerebrocurin intraperitoneally at a dose of 150 μl/kg;
- 9. Group 9: After PH, administered L-arginine intraperitoneally at a dose of 200 mg/kg;
- 10. Group 10: After PH, administered Glutarodoxin intraperitoneally at a dose of 200 µg/kg;
- 11. Group 11: After PH, administered HSF-1 intraperitoneally at a dose of 50 mg/kg;
- 12. Group 12: After PH, administered Mildronate intraperitoneally at a dose of 50 mg/kg

The following pharmaceutical preparations and pharmacological agents were used in the study:

- 1. Thiotriazoline (morpholinium 3-methyl-1,2,4-triazolyl-5-thioacetic acid) (2.5% solution for injections, Arterium Corporation, Ukraine);
- 2. Tamoxifen (tablets, Finland, on the basis of which an extemporaneously prepared intranasal gel (1 mg/1 ml) was made at the Department of Pharmaceutical Technology, ZSMU);
- 3. Angioline ([(S)-2,6-diaminohexanoic acid 3-methyl-1,2,4-triazolyl-5-thioacetate]) (substance from NPO "Pharmatron," Ukraine);
  - 4. Glutaredoxin (Sigma-Aldrich, USA);
- 5. Cerebrocurin (injection solution, NPO "NIR," Ukraine). The preparation contains neuropeptides, S-100 proteins, rylin, nerve growth factor (NGF) (no less than 2 mg/ml), and amino acids;
  - 6. L-Arginine (42% solution for injections in vials, Tivortin, Yuriy-Pharm, Ukraine);
- 7. Mexidol (2-ethyl-6-methyl-3-hydroxy-pyridyl succinate, injection solution, 50 mg/ml, NPK Pharmasoft, Ellara LLC, Russia);
- 8. Mildronate (2-(2-carboxyethyl)-1,1,1-trimethylhydrazinium) (10% solution for injections in ampoules, Grindex, Latvia), 100 mg/kg, i/p;
  - 9. Piracetam (200 mg/ml, JSC "Pharmak," Ukraine);
  - 10. HSF-1 (Sigma-Aldrich, USA).

Behavioral and exploratory activity of the animals was assessed one month after birth, immediately after the experimental therapy course, while the evaluation of cognitive and mnemonic functions of the central nervous system was conducted at 2 months of age (one month after the experimental therapy course).

Assessment of motor and exploratory activity was conducted using the "Open Field" method with an arena of our own design measuring 80x80x35 cm [33]. The animal was placed in the center of one of the sides with its nose facing the wall, after which it was allowed to move freely within the arena for 8 minutes. We assessed the total distance traveled (cm), overall motor activity (cm²/s), activity structure (high activity, low activity, inactivity, %), number of freezing episodes, entries into the center, distance traveled along the wall (cm) and in the central area of the arena (cm, %), vertical exploratory activity (number of rears on the hind legs at the wall and in the center), number of short and long grooming events, and the number of defecation and urination acts.

#### 2.3. Assessment of Reference and Working Memory

The rats were subjected to food deprivation. Food was available daily for 1 hour. The animals were reduced to 85% of their original body weight by restricting their food intake while allowing free access to water.

Memory was assessed using a radial maze (LE760, AgnTho's, Sweden) [34]. The eight-arm radial maze consists of an octagonal platform (side length 22 cm), with eight numbered radial arms (1 to 8) extending from the platform. Each arm is 70 cm long and 10 cm wide, with a food cup at the end (diameter 2 cm, depth 1.5 cm). Each arm can be closed independently using a guillotine mechanism. The entire setup was positioned 70 cm above the floor. We used the methodology as described previously [34]. The experiment was conducted in complete silence.

Starting on the first day, the animals were placed on the central platform of the maze, with 4 closed arms and 4 open arms, each containing 200 mg of food pellets in the food cups. The combination of open and closed arms was unique and constant for each animal. Over the next 10 days, the animals were trained to locate the food using external visual cues. The training session lasted for 10 minutes or until the animal found all four food sources. The experiment was repeated twice daily with each animal. After the experiment, the animal was given its daily food ration.

On the 10th day, the animal was placed in the radial maze with all eight arms open, with food placed in four arms according to the usual pattern for that animal. We assessed reference memory (the long-term knowledge of the maze structure and food locations that the animal developed during the training process) and the number of reference memory errors (the first visit to a previously closed arm where the animal had never found food). We also assessed working memory (the short-term representation of the food locations in the specific trial) and the number of working memory errors (revisiting an arm where the animal had previously found or not found food). Additionally, we evaluated the total distance traveled and overall motor activity.

#### 2.4. Data Collection and Processing

The research was conducted at the Department of Experimental Pathophysiology and Functional Morphology of the Educational Medical Laboratory Center of Zaporizhzhia State Medical University. The experiments were performed in a well-lit room in complete silence. External and internal visual, olfactory, and auditory stimuli were excluded during the experiments. Animal behavior was assessed by a technician who was unaware of the animal's group assignment. Image capture and recording were performed using a color video camera SSC-DC378P (Sony, Japan). Video file analysis was conducted using Smartv 3.0 software (Harvard Apparatus, USA). Statistical processing of the results was performed using Microsoft Excel 2016 with the AtteStat 12 statistical analysis package. The Kruskal-Wallis test with Dunn's post hoc correction was used to assess the significance of differences between experimental groups. Differences were considered statistically significant at p < 0.05.

#### 3. Results

When assessing the specific indicators of the "open field" method, it was found that modeling intrauterine hypoxia negatively affected the behavioral characteristics of the animals. Intrauterine

hypoxia led to a 1.33-fold decrease (p<0.05) in the overall activity of 1-month-old animals compared to 1-month-old animals born after physiological pregnancy. Additionally, hypoxia resulted in a twofold reduction (p<0.05) in the distance traveled, alongside a 2.7-fold increase (p<0.05) in inactivity and a 1.85-fold increase (p<0.05) in immobility compared to the intact group (healthy 1-month-old rats).

Furthermore, in animals that experienced intrauterine hypoxia, there was a 1.86-fold increase (p<0.05) in the number of freezing behaviors, a 2.2-fold increase (p<0.05) in the time spent in the illuminated center, and a 1.46-fold increase (p<0.05) in the distance traveled in the darkened area (near the wall). These findings indicate reduced motor activity in the post-hypoxia animals and the development of anxiety, fear, and excitability.

Modeling intrauterine hypoxia led to a twofold increase (p<0.05) in wall-posting in 1-month-old animals compared to healthy rats of the same age. Additionally, in the control group animals, there was a fourfold decrease (p<0.05) in the number of short grooming acts, accompanied by a threefold reduction (p<0.05) in defecation acts. This also points to increased anxiety, excitability, irritability, and a reduced sense of comfort in the animals.

In the control group, a 4.14-fold decrease (p<0.05) in high activity was observed, suggesting high emotionality and excitability of the animals. The reduction in high activity may also be interpreted as a decreased ability to engage in exploratory and searching behavior, as the rat becomes inactive, fearful of entering the illuminated space, and spends more time adapting to the new environment.

When assessing the specific indicators of learning and memory reproduction in the radial arm maze in 2-month-old rats after intrauterine hypoxia, it was found that 10 days after the learning process, animals that had experienced hypoxia exhibited persistent cognitive and mnemonic impairments compared to 2-month-old animals born after physiological pregnancy. During the reproduction of the learning results in the 2-month-old rats after hypoxia, it was established that 10 days after the last training session, there was a 3.2-fold increase in the number of working memory errors and a twofold increase in the number of reference memory errors. These findings indicate a disruption of higher CNS functions after intrauterine hypoxia.

**Table 1.** The effect of the studied pharmaceutical agents on the exploratory activity of 1-month-old rats in the open field following prenatal hypoxia.

Experimental group	Overall activity, cm <sup>2</sup> /s	Time spent entering the center, sec.	High activity, %	Inactivity, %	Freezing, count	Free distance, cm
Intact	$24380.9 \pm$	$12.9\pm2.4$	$17.8 \pm 1.44$	$30.4 \pm 6.7$	$282\pm32$	$60.2 \pm 4.3$
	1242.4					
Control PH	$18167.4 \pm$	$28.3\pm3.2^{\mathrm{1}}$	$4.3\pm0.57^1$	$82.3\pm6.3^1$	$561\pm22^{1}$	$29.7\pm2.1^{1}$
	$1043.5^{1}$					
PH+	45762.2 ±	$15.4 \pm 1.5^{1,2}$	$18.2 \pm 1.55^2$	$52.2 \pm 4.2^{1,2}$	$307 \pm 17^{1,2}$	$78.3 \pm 3.5$ <sup>1,2</sup>
Cerebrocurin	$2286.5^{1,2}$					
PH + Angiolin	39863.2 ±	$12.2 \pm 1.7^{1,2}$	$14.7 \pm 1.00^{1,2}$	$37.3 \pm 3.4^{2}$	$272 \pm 12^{1,2}$	$74.7 \pm 1.7^{1,2}$
	$1022.5^{1,2}$					
PH+	27952.1 ±	$21.7 \pm 1.2^{1}$	$8.7 \pm 0.67^{12}$	$54.3 \pm 3.3^{1,2}$	$412 \pm 32^{1,2}$	$50.2 \pm 2.6^{1,2}$
Piracetam	$1103.0^{1,2}$					
PH + L-	18722.2 ±	27.7 ± 4.1 <sup>1</sup>	$4.7 \pm 0.72^{1}$	$87.3 \pm 8.2^{1}$	$532 \pm 34^{1}$	$32.4 \pm 3.2^{1}$
arginine	$978.2^{1}$					
PH+	17764.4 ±	$31.7 \pm 3.5^{1}$	$3.8 \pm 0.25^{1}$	$121.3 \pm 15.7^{1,2}$	$475 \pm 35^{1,2}$	$22.4 \pm 2.5^{1}$
Tamoxifen	$1103.3^{1}$					

PH+	$29221.2 \pm$	$23.5 \pm 2.7^{1}$	$5.8 \pm 0.35^{1}$	$78.3 \pm 4.7^{1}$	$549 \pm 34^{1}$	$53.4 \pm 4.2^{1,2}$
Glutaredoxin	$992.2^{1,2}$					
PH+	$21822.2 \pm$	$21.4 \pm 2.4^{1}$	$7.8 \pm 0.45^{1}$	$73.4 \pm 5.3^{1}$	$501 \pm 27^{1}$	$46.7 \pm 2.3^{1,2}$
Thiotriazoline	$1121.1^{1,2}$					
PH + Mexidol	19612.2 ±	$27.3 \pm 2.5^{1}$	$4.3 \pm 0.57^{1}$	$87.8 \pm 7.2^{1}$	447 ± 22 <sup>1,2</sup>	$44.2 \pm 2.7^{1,2}$
	$1231.2^{1}$					
DII . LICE1	30573.4 ±	$18.0 \pm 1.4^{1,2}$	$8.5 \pm 0.74^{1,2}$	$77.3 \pm 6.2^{1}$	$423 \pm 26^{1,2}$	$62.2 \pm 2.2^{1,2}$
PH + HSF1	911.2 <sup>1,2</sup>					
PH+	24876.2 ±	$26.5 \pm 2.2^{1}$	$4.8 \pm 0.65^{1}$	$67.3 \pm 5.3^{1}$	$511 \pm 67^{1}$	$65.1 \pm 1.4^{1,2}$
Mildronate	$914.2^{2}$					

<sup>1</sup> - changes are significant in relation to animals of the intact group (p<0.05); 2 - changes are significant in relation to the animals of control group

**Table 2.** The effect of the studied pharmaceutical agents on the exploratory activity of 1-month-old rats in the open field following prenatal hypoxia (cont.).

Experimental group	Distance along the wall, cm	Standing next to the wall, count	Short grooming, count	Defecation, count	Immobility, count
Intact	$4012.4 \pm 277.5$	4 ± 1	4 ± 1	3	$282 \pm 25$
Control PH	$5857.2 \pm 205.2^{1}$	8 ± 1	$1 \pm 1^{1}$	11	$523 \pm 17^{1}$
PH + Cerebrocurin	4521.2 ± 182.2 <sup>1,2</sup>	4 ± 1 <sup>2</sup>	4 ± 1 <sup>1,2</sup>	42	211 ± 12 <sup>2</sup>
PH + Angiolin	$4211.3 \pm 234.2^{1,2}$	$4 \pm 1^2$	$4\pm1^{1,2}$	$4^2$	$280 \pm 25^{2}$
PH + Piracetam	5922.3 ± 177.3 <sup>1</sup>	$5 \pm 1^2$	21	2	$242 \pm 18^2$
PH + L-arginine	5768.5 ± 187.31	8 ± 21	11	$1^{1}$	$515 \pm 22^{1}$
PH + Tamoxifen	6045,1 ± 312,71	61,2	11	$1^{1}$	577 ± 231
PH + Glutaredoxin	5743.5 ± 197.4 <sup>1</sup>	81	11	11	473 ± 21 <sup>1</sup>
PH + Thiotriazoline	$5634.3 \pm 223.8^{1}$	61,2	$1^1$	$1^1$	$435 \pm 21^{1}$
PH + Mexidol	5433.2 ± 211.31	$6^{1,2}$	$3^{1}$	$1^1$	$507 \pm 32^{1}$
PH + HSF1	5245.3 ± 231.3 <sup>1</sup>	61,2	31	11	$312 \pm 22^2$
PH + Mildronate	5723.5 ± 311.6 <sup>1</sup>	81	11	11	251 ± 12 <sup>2</sup>

<sup>1</sup> - changes are significant in relation to animals of the intact group (p<0.05); 2 - changes are significant in relation to the animals of control group

**Table 3.** The effect of the studied pharmaceutical agents on the exploratory activity of 1-month-old rats in the open field following prenatal hypoxia (cont.).

Experimental group	Number of reference memory errors	Number of working memory errors
Intact	2	5
Control PH	$4 \pm 1^1$	$16 \pm 1^{1}$
PH + Cerebrocurin	$2^2$	71,2
PH + Angiolin	1 <sup>1,2</sup>	$7 \pm 1^{1,2}$
PH + Piracetam	31	141,2

PH + L-arginine	5 <sup>1</sup>	16 ± 21
PH + Tamoxifen	$4^1$	$14 \pm 1^{1}$
PH + Glutaredoxin	$4^{1}$	121,2
PH + Thiotriazoline	31	121,2
PH + Mexidol	31	151
PH + HSF1	31	101,2
PH + Mildronate	41	16 <sup>1</sup>

1 - changes are significant in relation to animals of the intact group (p<0.05); 2 - changes are significant in relation to the animals of control group

The introduction of HSP<sub>70</sub> modulators, antioxidants, nootropics, and metabolotropic therapy agents immediately after birth to experimental animals following PH had varying effects on the main indicators of higher CNS functions, with some showing no improvement in behavioral reactions, emotional status, or cognitive-mnestic functions. Analyzing the results of the experimental studies, it was found that Angiolin and Cerebrocurin had the most pronounced effect on the indicators of exploratory-activity and psycho-emotional status in 1-month-old animals after PH, as observed in the "open field" test. Other drugs and pharmacological agents, such as Piracetam, Mildronate, Glutoredoxin, Thiotriazoline, and HSF-1, had a moderate effect on specific indicators of exploratory activity and psycho-emotional status in 1-month-old animals after PH. The drugs Mexidol, Tamoxifen, and L-Arginine showed no effect on the studied indicators in the "open field" test.

Cumulative administration of Cerebrocurin immediately after discontinuation of the treatment led to a 2.5-fold increase (p<0.05) in overall motor activity of animals compared to the control group (untreated animals) and a 1.87-fold increase (p<0.05) compared to the group of healthy animals (born after physiological pregnancy). In the experimental animals treated with Cerebrocurin, the free locomotion distance increased 2.6 times (p<0.05) compared to the control group and 1.3 times (p<0.05) compared to the healthy animals group, while high activity was increased 4.2 times (p<0.05) compared to the control group. This indicated a high level of motor and exploratory activity in animals after PH who were treated with Cerebrocurin. These animals were active, mobile, and intensively explored new spaces.

Cerebrocurin also positively affected the psycho-emotional status of 1-month-old animals after PH. The animals actively entered the illuminated arena, did not linger in the dark and semi-dark areas, actively groomed, and displayed reduced anxiety and aggression. In this experimental group, the number of freezing behaviors decreased 1.8 times (p<0.05), inactivity decreased 1.57 times (p<0.05), the time spent in the illuminated center of the arena decreased 1.86 times (p<0.05), the distance traveled in the darkened area decreased 1.3 times (p<0.05), and the total immobility score decreased 2.47 times (p<0.05) compared to the control group. The introduction of Cerebrocurin to animals after PH resulted in the complete restoration of psycho-emotional behavior indicators (such as grooming, defecation acts, and wall-posting behavior) to the level of the intact group (healthy animals).

1-month-old animals after PH treated with Angiolin immediately after birth also showed no difference in exploratory activity compared to healthy animals of the same age. These animals were active, with overall motor activity increasing 2.2 times (p<0.05) compared to the control group and 1.63 times (p<0.05) compared to the healthy animals group. Experimental animals treated with Angiolin traveled 2.5 times more (p<0.05) free distance compared to the control group and 1.24 times (p<0.05) compared to healthy animals, and high activity increased 3.4 times (p<0.05) compared to the control group.

Angiolin, like Cerebrocurin, positively influenced the psycho-emotional behavior of 1-monthold animals after PH. Not only did it reduce aggression and anxiety, but it also decreased the fear of entering a new illuminated space and increased comfort and empathy. In the group of animals treated with Angiolin, freezing behaviors decreased 2 times (p<0.05), inactivity decreased 2.2 times (p<0.05),

the time spent in the illuminated center decreased 2.3 times (p<0.05), the distance traveled in the darkened area decreased 1.4 times (p<0.05), and overall immobility decreased 1.86 times (p<0.05) compared to the control group. The course administration of Angiolin to animals after PH proved to be as successful as Cerebrocurin, leading to the restoration of psycho-emotional behavior indicators (grooming, defecation acts, and wall-posting behavior) to the intact level (healthy animals).

Pharmacological agents such as Piracetam, Mildronate, Glutoredoxin, Thiotriazoline, and HSF-1 significantly increased overall activity, free locomotion distance, and decreased immobility in animals after PH (Piracetam, Mildronate, Glutoredoxin, and HSF-1, p<0.05). This indicates the restoration of motor activity in experimental animals when exploring new environments. However, these pharmacological agents had no significant effect on psycho-emotional behavior (anxiety, aggression, fear, discomfort) or the exploratory component of behavior.

It is also worth noting that intranasal administration of Tamoxifen influenced fear and anxiety by reducing the number of freezing behaviors (p<0.05) and wall-posting (p<0.05).

The evaluation of the effects of the studied pharmacological agents on the cognitive-mnestic functions of the CNS in 2-month-old animals after PH revealed the following. The most pronounced mnemonic and nootropic effects were observed with Angiolin and Cerebrocurin. Thirty days after the completion of the course administration of these agents, the number of reference memory errors in animals that underwent PH decreased by 4 times (p<0.05) and 2 times (p<0.05), respectively. Also, after the course administration of Angiolin and Cerebrocurin, the number of working memory errors decreased by 2.28 times (p<0.05). Other experimental participants—Mexidol, Mildronate, Thiotriazoline, Glutoredoxin, Tamoxifen, Piracetam, L-Arginine, and HSF-1—did not significantly affect the number of reference memory errors made by experimental animals in the radial arm maze after training. However, Piracetam, Glutoredoxin, Thiotriazoline, and HSF-1 significantly reduced the number of working memory errors in the radial arm maze after training in animals that underwent PH.

Thus, it was established that modeling PH leads to persistent impairments in exploratory activity, psycho-emotional behavior, and cognitive-mnestic functions of the CNS in animals. HSP70 modulators, especially Angiolin and Cerebrocurin, had a positive effect on overall motor activity, exploratory behavior, reducing anxiety, irritability, aggression, and fear, and contributed to the improvement of learning and retention of acquired skills.

#### 4.Discussion

The data we obtained align with the concept of CNS impairments resulting from prenatal hypoxia (PH). PH leads to an increased risk of developing psychomotor and cognitive-mnestic disorders, which are often not compensated for throughout the subsequent life [35]. A number of researchers have noted that animals following prenatal hypoxia (PH) often exhibit motor and learning impairments, hyperactivity syndrome, as well as reduced memory, socialization, and attention [36]. One of the main causes of cognitive dysfunctions after PH is considered to be disturbances in energy metabolism, including the lack of energy in the synapse to maintain glutamatergic transmission, neurotransmitter reuptake, inhibition of synaptogenesis, increased production of ROS through neurochemical and energetic reactions, as well as the initiation of mechanisms of circulatory ischemia (hypoperfusion, changes in brain perfusion, intracranial pressure) [37-40]. PH leads to impairments in various types of memory (short-term memory, longterm memory, working memory, and reference memory) in the offspring of rats. This is associated with structural changes observed in the hippocampus, due to the activation of apoptosis and neuronal cell death [41-43]. PH leads to the development of persistent cognitive deficits, as well as psycho-emotional disturbances such as sluggishness, fear, anxiety, disorientation, aggression, and irritability. Several studies have shown that PH results in behavior similar to anxiety and depression, reduced social interaction, impaired sexual behavior in males, and a decrease in the acquisition of both passive and active avoidance responses [38,44]. Thus, in 1-month-old rats after PH, basal synaptic transmission in the CA3-CA1 synapses was significantly impaired, and long-term synaptic

potentiation in the hippocampus was reduced by twofold. Additionally, there was a decrease in the affinity of the GluN2B receptor, accompanied by of a significant deficit in learning and memory [45]. It has been established that PH leads to a significant increase in the production of cytotoxic products of nitrosative stress due to the elevated expression of iNOS and nNOS, in connection with deprivation of thiol-disulfide system intermediates in the CA1 hippocampus of 1- and 2-month-old rats that underwent intrauterine hypoxia [14,46].

It was also found in this age group of experimental animals that PH leads to a decrease in the expression of mRNA HIF- $1\alpha$ , mRNA HSP70, mRNA c-fos, as well as the concentration of HSP70 and HIF- $1\alpha$  in the brain and plasma, and increases offspring mortality. We established a direct correlation between the levels of HSP70 and the survival of the offspring [28,47]. HSP70 is important for memory formation as it facilitates the folding and transport of synaptic proteins, modulates signaling cascades associated with synaptic activation, and is involved in the mechanisms of neurotransmitter release [48]. C-fos plays a role in the development of cognitive and mnemonic dysfunctions of the CNS after TBI. c-fos is actively involved in the mechanisms of endogenous neuroprotection, angiogenesis, memory consolidation, expression of heat shock protein genes (HSP70 and HSP72), as well as hypoxia-inducible factor (HIF-1). TBI alters the expression of HIF- $1\alpha$  in the brains of embryonic rats depending on the stage of embryonic development, followed by a sustained increase and activation of the HPA axis, causing hypoxic disease and behavioral changes [44, 49-51]. It is known that inhibition of c-fos mRNA translation in brain structures impairs short-term memory in various learning models across different animal species [52-54] (Figure 1).

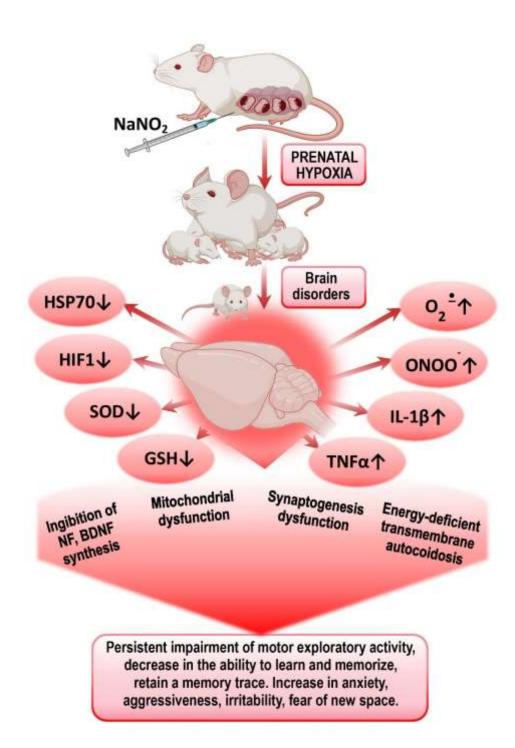
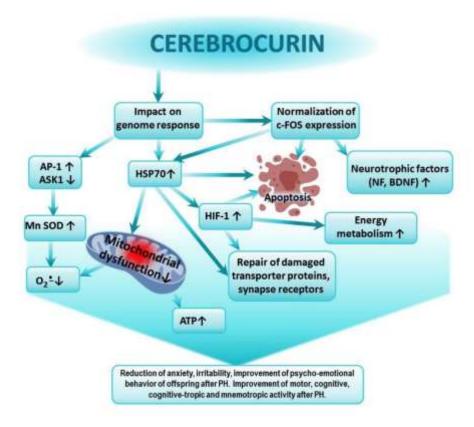


Figure 1. Mechanisms of formation of psychomotor and cognitive-mental disorders in offspring after PH.

Presumably, one of the leading mechanisms of impaired CNS cognitive-mental functions in offspring after PH is disturbances in energy metabolism, ATP deficiency leading to transmitter autocoidosis, reduced energy supply for synaptic transmission and formation of secondary mitochondrial dysfunction. The next step is increased production of ROS and NO by energetic and neurochemical reactions and triggering mechanisms of circulatory ischemia. Mitochondrial dysfunction and nitrosative and oxidative stresses initiate trigger mechanisms of neuroinflammation and neuronal apoptosis, leading to impaired mechanisms of short-term and long-term memory. Excess of ROS on the background of deprivation of antioxidant system and factors of endogenous neuroprotection (HSP70 and HIF-1) leads to desensitization of receptors (GABA, serotonin,

glutamine), suppression of transporter proteins expression and increases psycho-emotional disorders - lethargy, fear, anxiety, disorientation, aggressiveness, irritability.

In previous studies, we showed that the greatest impact on the expression of HSP70 and HIF-1 $\alpha$ in the brains of animals after TBI was exerted by Cerebrocurin and Angiolin. Agents such as HSK-1, Thiotriazoline, Glutaredoxin, and Tamoxifen moderately affected HSP70 levels, while L-arginine, Mexidol, and Piracetam showed minimal or no effect [55]. Thus, there is a certain correlation between the effect on the levels of HSP<sub>70</sub> and HIF-1 $\alpha$  and the cognitive-enhancing action of the drugs. The properties of Cerebrocurin identified by us are consistent with the findings of other researchers, who demonstrated the high antioxidant activity of the drug, aimed at increasing the expression of Mn-SOD, as well as its neuroprotective and anti-ischemic activity, realized through direct mitoprotection, increased synthesis of HSP and HIF proteins, and their mediated activation of compensatory energy shunts under acute cerebral ischemia conditions [47]. Cerebrocurin contains peptides that carry the program for analyzing the condition and construction of the CNS. Therefore, the final effect of Cerebrocurin varies due to its qualitatively distinct mechanism of action compared to other neuroprotectors. The protective effects of Cerebrocurin on brain tissue include its optimizing action on brain energy metabolism and calcium homeostasis, stimulation of intracellular synthesis of protective proteins, growth factors for neurons, and neurotrophic factors, as well as the slowing down of glutamate-calcium cascade processes and oxidative protein modification. Cerebrocurin may reduce disturbances in higher CNS functions due to alcohol intoxication, hypoxia, and cerebral circulation disorders. This action of Cerebrocurin is linked to its optimizing effect on the expression of HSP<sub>70</sub> and c-fos. The reduction in anxiety and aggressiveness in the offspring treated with Cerebrocurin is likely associated with its ability to influence the mechanisms of neurotransmitter reuptake and reduce desensitization of corresponding receptors [56,57] (Figure 2).



**Figure 2.** Possible mechanism of the positive effect of Cerebrocurin on cognitive-tropic functions of the CNS, research, motor and emotional activity of offspring after PH.

Cerebrocurin contains peptides that carry a program to analyze the state and structure of the CNS. Cerebrocurin through its effect on the expression of HSP 70 triggers regulatory mechanisms aimed at increasing the expression of antioxidant enzymes, expression of HIF proteins, activation of compensatory mitochondrial-cytosolic shunts of energy production and calcium homeostasis, stimulation of intracellular synthesis of protective proteins, neuronal growth factors and neurotrophic factor, increase in the affinity of BDNF binding to its receptors and normalization of c-fos expression. Cerebrocurin exhibits a significant antiapoptotic effect. Cerebrocurin positively affects the mechanisms of transmitter reuptake through the expression of transporter proteins.

Significant improvements in exploratory, motor, cognitive-enhancing, and mnemonic activity in the offspring after TBI, as well as the restoration of their psycho-emotional behavior following the administration of Angiolin, are ensured by the following mechanisms. First, through its antioxidant mechanism (scavenging of ROS and NO), Angiolin exerts a positive effect on the expression of genes encoding the synthesis of superoxide dismutase, glutathione peroxidase, and glutathione reductase. It regulates ROS/GSH-dependent mechanisms of HSP70 expression and, through this, promotes the prolongation of HIF-1 lifespan. Angiolin also regulates the transport of synaptic proteins, modulates signaling cascades related to the reverberation of excitation in neuronal circuits, and reduces oxidative modification of protein molecules, including receptors and ion channels [48,58]. Secondly, Angiolin, under conditions of brain ischemia, reduces the number of damaged mitochondria in the cortex and CA1 hippocampus. It is capable of activating compensatory cytosolic-mitochondrial energy shunts and increasing ATP production, which is necessary to sustain memory protein synthesis and synaptic function [58]. Many authors confirm the importance of improving brain energy metabolism in the fight against dementia [59,60]. It is also known that mitochondrial dysfunction is a hallmark of many diseases that impair brain function and affect cognitive abilities [57], while the improvement of cognitive and mnemonic functions is associated with mitoprotection [61]. All of this allows us to evaluate the cognitive-enhancing action of Angiolin through the prism of its mitoprotection. Thirdly, Angiolin, due to the L-lysine residue in its composition, increases the affinity of GABA receptors and reduces excitotoxicity [62,63]. The increase in GABA receptor affinity under the action of Angiolin may explain its anxiolytic effect (reduction of anxiety, fear, and aggressiveness) (Figure 3).

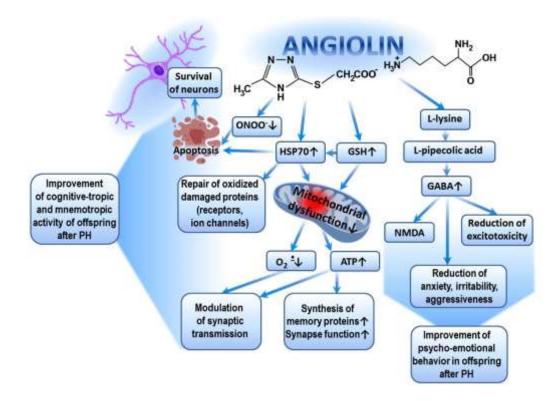


Figure 3. Possible mechanism of cognitive-tropic, mnemotropic and anti-anxiety effect of Angiolin after PH.

The pharmacological properties of Angiolin are determined by the conversion of L-lysine into pipecolic acid, which increases the affinity of the GABA-benzodiazepine receptor complex and thus reduces manifestations of glutamate excitotoxicity. Angiolin preserves the ultrastructural and functional activity of mitochondria, increases ATP production, and activates compensatory energy shunts. Angiolin activates the glutathione link of the thiol-disulfide system and thereby regulates the ROS/GSH-dependent mechanisms of HSP70 expression, which contributes to the increase in HIF-1 lifespan, regulates the transport of synaptic proteins, modulates signaling cascades associated with the reverberation of excitation in neuronal chains, reduces oxidative modification of protein molecules of receptors and ion channels. Angiolin exhibits NO scavenger properties, inhibits nitrosative stress reactions, has anti-apoptotic properties.

Activation of HSP $_{70}$  transcription in all studied organisms primarily occurs through the activation of members of the HSF family [64]. The results we obtained regarding the positive influence of HSF1 on the activity of offspring after TBI, particularly in novel space exploration and learning, are consistent with other studies. It is known that HSF1 is involved in memory formation processes. Activation of HSF1 leads to improved cognitive abilities, while its deficiency, along with the deficiency of HSP $_{70}$ , is associated with neurodegeneration [65,66]. In addition to regulating the HSP system, HSF-1 is involved in the expression of many genes and regulates processes such as metabolism, RNA splicing, apoptosis, ubiquitination, protein degradation, and intracellular signaling [67-69]. HSF-1 leads to the inhibition of TNF $\alpha$ -induced apoptosis, including in neurons, thereby preserving higher CNS functions [70,71]. In this study, Glutaredoxin demonstrated a positive effect on the motor and cognitive activity of offspring after TBI and positively influenced learning in the maze. We explain this effect from the perspective of its impact on GSH-dependent pathways of endogenous neuroprotection and the increased expression of mRNA for HSP $_{70}$ , HIF-1 $\alpha$ , and HIF-3 $\alpha$  under conditions of acute experimental cerebral ischemia, as well as the enhancement of glutathione

synthesis. It is known that the role of the glutathione component of the thiol-disulfide system in the brain is crucial for maintaining the cognitive functions of the CNS [72].

Thiotriazoline in this study demonstrated satisfactory activity in restoring motor and exploratory activity in a novel environment in the offspring after prenatal glucocorticoid exposure, as well as a positive effect on their learning ability and skill acquisition. Such effects of Thiotriazoline are not new and have been described in various studies. The ability of Thiotriazoline to modulate the antioxidant, nootropic, mnemonic, and antiamnesic activities of Piracetam has also been demonstrated, which served as the basis for the development of the drug Tiocetam (Thiotriazoline + Piracetam) [73,74]. The mnemonic and cognitive-enhancing effects of Thiotriazoline can be explained in terms of its antioxidant mechanism. Due to the specific structure of its molecule, Thiotriazoline acts as a scavenger of NO and ROS, regulating the expression of NF-kB, which may provide a significant antioxidant effect as well as regulation of the expression of protective proteins. The mechanism by which Thiotriazoline affects the expression of HSP70 has been well described in several studies and is associated with SH/SS regulation of transcription factor activity. Through its positive influence on mitochondria, Thiotriazoline improves energy metabolism, enhances performance, motor activity, and synaptic transmission of excitation [24,47,57,75]. The observed effect of Thiotriazoline on cognitive, cognitive-enhancing, and mnemonic activity in the offspring after prenatal glucocorticoid exposure is attributed to its properties as a scavenger of ROS and a regulator of their concentration. The physiological concentration of ROS, particularly superoxide, modulates synaptic plasticity at the cellular level and plays a crucial role in the formation and maintenance of memory [76]. A significant increase in ROS levels leads to disruption of neural networks essential for memory function [77,78,79,80,81].

In our study, the use of Tamoxifen as a modulator of HSP $_{70}$  to improve behavioral, exploratory, and cognitive activity in the offspring after prenatal glucocorticoid exposure did not meet expectations. A positive effect of Tamoxifen was observed in the reduction of anxiety in the 1-month-old offspring. It appears that the influence on HSP $_{70}$ -dependent mechanisms of memory formation and preservation by Tamoxifen was insufficient. The anxiolytic effect of Tamoxifen is likely related to its modulatory action on central estrogen receptors. Activation of ER $\beta$  may contribute to the generation of anxiolytic effects [82]. The anxiolytic effect of Tamoxifen in our study can also be explained by the activation of the GABAergic system through modulation of ER $\beta$  [83]. It is also possible that the enhancement of Tamoxifen's anxiolytic effect is linked to its ability to influence glutathione levels [84,85].

#### 5. Conclusions

- 1. Modeling PH leads to persistent impairments in exploratory-activity, psycho-emotional behavior of animals, as well as a decrease in their cognitive-mnestic functions of the CNS.
- 2. Administration of HSP<sub>70</sub> modulators, antioxidants, nootropics, and metabolotropic therapy agents immediately after birth to experimental animals following PH had varying effects on the main indicators of higher CNS functions, either failing to improve behavioral responses, emotional status, or cognitive-mnestic functions, or showing effects in a specific direction and with varying intensity.
- 3. Analyzing the results of experimental studies, we found that Angiolin and Cerebrocurin had the most pronounced effect on the indicators of exploratory activity in 1-month-old animals after PH, as assessed in the "open field" test. Angiolin, like Cerebrocurin, positively influenced the psychoemotional behavior of 1-month-old animals after PH, with a reduction in aggressiveness and anxiety, as well as a decrease in the fear of a new illuminated space.
- 4. Angiolin and Cerebrocurin demonstrated the most pronounced cognitive-enhancing and memory-supporting effects during training and the assessment of skill retention in a maze in 2-month-old offspring after PH.
- 5. The results obtained provide experimental justification for the clinical use of Cerebrocurin and, especially, Angiolin for the correction of anxious behavior, excitability, and cognitive-mnestic dysfunction in offspring after PH.

**Author Contributions:** Conceptualization, O.A.; I.F.B., methodology, O.A. and O.K.; validation, I.B. and O.A., I.D., O.K., L.M.; formal analysis, I.B., I.D.; investigation, O.A., I.B., I.D. and O.K.; resources, I.B., O.A.; data curation, L.M., V.O.; writing—original draft preparation, I.F.B.; writing—review and editing, I.F.B., L.M., V.O., O.K.; visualization, I.D., L.M., V.O.; supervision, I.F.B., O.A.; project administration, I.B., V.O., O.K., All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

**Institutional Review Board Statement:** The animal study protocol was approved by the Ethics Committee of Zaporizhzhia State Medical University (protocol No. 2 of 18 May 2023).

Conflicts of Interest: The authors declare no conflicts of interest.

#### References

- 1. Davis, E.P.; Narayan, A.J. Pregnancy as a period of risk, adaptation, and resilience for mothers and infants. *Dev Psychopathol.* **2020**, *32*(5), 1625-1639. doi: 10.1017/S0954579420001121.
- 2. Martín-Rodríguez, A.; Bustamante-Sánchez, Á.; Martínez-Guardado, I.; Navarro-Jiménez, E.; Plata-SanJuan, E.; Tornero-Aguilera, J.F.; Clemente-Suárez, V.J. Infancy Dietary Patterns, Development, and Health: An Extensive Narrative Review. *Children (Basel)*. **2022**, *9*(7), 1072. doi: 10.3390/children9071072.
- 3. Batey, N.; Henry, C.; Garg, S.; et al. The newborn delivery room of tomorrow: emerging and future technologies. *Pediatr Res.* **2024**, *96*, 586–594. doi: 10.1038/s41390-022-01988-y.
- 4. Dev, A.; Casseus, M.; Baptiste, W.J.; et al. Neonatal mortality in a public referral hospital in southern Haiti: a retrospective cohort study. *BMC Pediatr.* **2022**, 22, 81. doi: 10.1186/s12887-022-03141-4.
- 5. Gessesse, A.D.; Belete, M.B.; Tadesse, F. Time, cause of early neonatal death, and its predictors among neonates admitted to neonatal intensive care units at Bahir Dar City public hospitals, northwest Ethiopia: a prospective follow-up study. *Front Pediatr.* **2024**, *12*, 1335858. doi: 10.3389/fped.2024.1335858.
- 6. Ranjan, A.K.; Gulati, A. Advances in Therapies to Treat Neonatal Hypoxic-Ischemic Encephalopathy. *J Clin Med.* **2023**, 12(20), 6653. doi: 10.3390/jcm12206653.
- 7. Keam, S.J. Sovateltide: First Approval. Drugs. 2023, 83(13), 1239-1244. doi: 10.1007/s40265-023-01922-4.
- 8. Reyes-Corral, M.; Sola-Idígora, N.; de la Puerta, R.; Montaner, J.; Ybot-González, P. Nutraceuticals in the Prevention of Neonatal Hypoxia–Ischemia: A Comprehensive Review of their Neuroprotective Properties, Mechanisms of Action and Future Directions. *Int. J. Mol. Sci.* **2021**, 22, 2524. doi: 10.3390/ijms22052524.
- 9. Wardinger, J.E.; Ambati, S. Placental Insufficiency. *Treasure Island (FL): StatPearls Publishing*, **2025.** https://www.ncbi.nlm.nih.gov/books/NBK563171.
- 10. Allen, K.A.; Brandon, D.H. Hypoxic Ischemic Encephalopathy: Pathophysiology and Experimental Treatments. Newborn Infant *Nurs Rev.* **2011**, *11*(3), 125-133. doi: 10.1053/j.nainr.2011.07.004.
- 11. Nicosia, N.; Giovenzana, M.; Misztak, P.; Mingardi, J.; Musazzi, L. Glutamate-Mediated Excitotoxicity in the Pathogenesis and Treatment of Neurodevelopmental and Adult Mental Disorders. *Int. J. Mol. Sci.* **2024**, 25, 6521. doi: 10.3390/ijms25126521.
- 12. Piña-Crespo, J.C.; Sanz-Blasco, S.; Lipton, S.A. Concept of Excitotoxicity via Glutamate Receptors. In: Kostrzewa, R. (eds) Handbook of Neurotoxicity. *Springer, New York, NY*. **2014**. https://doi.org/10.1007/978-1-4614-5836-4 125.
- 13. Bertozzi, G.; Ferrara, M.; Calvano, M.; Pascale, N.; Di Fazio, A. Oxidative/Nitrosative Stress and Brain Involvement in Sepsis: A Relationship Supported by Immunohistochemistry. *Medicina*. **2024**, *60*, 1949. https://doi.org/10.3390/medicina60121949.
- 14. Martini, S.; Austin, T.; Aceti, A.; Faldella, G.; Corvaglia, L. Free radicals and neonatal encephalopathy: mechanisms of injury, biomarkers, and antioxidant treatment perspectives. *Pediatr Res.* **2020**, *87*(5), 823-833. doi: 10.1038/s41390-019-0639-6.
- 15. Belenichev, I.; Popazova, O.; Bukhtiyarova, N.; Savchenko, D.; Oksenych, V.; Kamyshnyi, O. Modulating Nitric Oxide: Implications for Cytotoxicity and Cytoprotection. Antioxidants (Basel). 2024, 13(5), 504. doi: 10.3390/antiox13050504.
- 16. Neto, A.; Fernandes, A.; Barateiro, A. The complex relationship between obesity and neurodegenerative diseases: an updated review. *Front. Cell. Neurosci.* **2023**, *17*, 1294420. doi: 10.3389/fncel.2023.1294420.
- 17. Guo, M.; Ji, X.; Liu, J. Hypoxia and Alpha-Synuclein: Inextricable Link Underlying the Pathologic Progression of Parkinson's Disease. *Front. Aging Neurosci.* **2022**, *14*, 919343. doi: 10.3389/fnagi.2022.919343.
- 18. Orzeł, A.; Unrug-Bielawska, K.; Filipecka-Tyczka, D.; Berbeka, K.; Zeber-Lubecka, N.; Zielińska, M.; Kajdy, A. Molecular Pathways of Altered Brain Development in Fetuses Exposed to Hypoxia. *Int. J. Mol. Sci.* **2023**, 24, 10401. https://doi.org/10.3390/ijms241210401.
- 19. Ludwig, P.E.; Reddy, V.; Varacallo, M.A. Neuroanatomy, Neurons. *Treasure Island (FL): StatPearls Publishing*, **2025.** https://www.ncbi.nlm.nih.gov/books/NBK441977.
- 20. Koop, L.K.; Tadi, P. Neuroanatomy, Sensory Nerves. *Treasure Island (FL): StatPearls Publishing*, **2025.** https://www.ncbi.nlm.nih.gov/books/NBK539846.

- 21. Yfantis, A.; Mylonis, I.; Chachami, G.; Nikolaidis, M.; Amoutzias, G.D.; Paraskeva, E.; Simos, G. Transcriptional Response to Hypoxia: The Role of HIF-1-Associated Co-Regulators. *Cells.* **2023**, *12*(*5*), 798. doi: 10.3390/cells12050798.
- Ziello, J.E.; Jovin, I.S.; Huang, Y. Hypoxia-Inducible Factor (HIF)-1 regulatory pathway and its potential for therapeutic intervention in malignancy and ischemia. Yale J Biol Med. 2007, 80(2), 51-60.
- 23. Aliyeva, O.G.; Belenichev, I.F.; Popazova, O.O.; Zidrashko, G.A. Pharmacological Modulation of Hif-1α in the Cerebral Cortex of Rats after Chronic Prenatal Hypoxia. *Biol. Life Sci. Forum.* **2022**, *19*, 9. doi: 10.3390/IECBS2022-12947.
- 24. Belenichev, I.F.; Kolesnik, Y.M.; Pavlov, S.V.; et al. Disturbance of HSP70 chaperone activity is a possible mechanism of mitochondrial dysfunction. *Neurochem. J.* **2011**, *5*, 251–256. doi: 10.1134/S1819712411040040.
- 25. Belenichev, I.F.; Aliyeva, O.G.; Popazova, O.O.; Bukhtiyarova, N.V. Involvement of heat shock proteins HSP70 in the mechanisms of endogenous neuroprotection: the prospect of using HSP70 modulators. *Front. Cell. Neurosci.* **2023**, *17*, 1131683. doi: 10.3389/fncel.2023.1131683.
- Aliyeva, O.; Belenichev, I.; Popazova, O. Modulation of Hsp70 in the Pharmacological Correction of Nervous System Disorders after Prenatal Hypoxia. Med. Sci. Forum. 2023, 21, 39. doi: 10.3390/ECB2023-14091.
- 27. Kim, J.Y.; Barua, S.; Huang, M.Y.; Park, J.; Yenari, M.A.; Lee, J.E. Heat Shock Protein 70 (HSP70) Induction: Chaperonotherapy for Neuroprotection after Brain Injury. *Cells.* **2020**, *9*, 2020. doi: 10.3390/cells9092020.
- 28. Belenichev, I.; Aliyeva, O.; Burlaka, B.; Burlaka, K.; Kuchkovskyi, O.; Savchenko, D.; Oksenych, V.; Kamyshnyi, O. Development and Optimization of Nasal Composition of a Neuroprotective Agent for Use in Neonatology after Prenatal Hypoxia. *Pharmaceuticals* (*Basel*). **2024**, *17*(8), 990. doi: 10.3390/ph17080990.
- 29. Belenichev, I.; Aliyeva, O.; Bukhtiyarova, N.; Semenov, D.; Voloshchuk, S. Comparative Assessment of the Effectiveness of HSP70/HIF-1α System Modulators after Prenatal Hypoxia. *Biomed. Pharmacol. J.* **2024**, *17*(1), 223-233. doi: 10.13005/bpj/2850.
- 30. Belenichev, I.F.; Aliyeva, O.G.; Gunina, L.M.; Bukhtiyarova, N.V. Evaluation of the efficiency of the neuroprotective drugs after prenatal hypoxia. *Fiziologichnyi Zhurnal*. **2023**, 69(1), 43-53. https://doi.org/10.15407/fz69.01.043.
- 31. Beretta, G.; Shala, A.L. Impact of Heat Shock Proteins in Neurodegeneration: Possible Therapeutical Targets. *Ann. Neurosci.* **2022**, *29*(1), 71-82. doi: 10.1177/09727531211070528.
- 32. Kang, Z.; Lin, Y.; Su, C.; Li, S.; Xie, W.; Wu, X. Hsp70 ameliorates sleep deprivation-induced anxiety-like behavior and cognitive impairment in mice. *Brain Res. Bull.* **2023**, 204, 110791. doi: 10.1016/j.brainresbull.2023.110791.
- 33. Varga, D.; Herédi, J.; Kánvási, Z.; Ruszka, M.; Kis, Z.; Ono, E.; Iwamori, N.; Iwamori, T.; Takakuwa, H.; Vécsei, L.; Toldi, J.; Gellért, L. Systemic L-Kynurenine sulfate administration disrupts object recognition memory, alters open field behavior and decreases c-Fos immunopositivity in C57Bl/6 mice. *Front. Behav. Neurosci.* 2015, 9, 157–157. doi: 10.3389/fnbeh.2015.00157.
- 34. Crusio, W.E.; Schwegler, H. Learning spatial orientation tasks in the radial-maze and structural variation in the hippocampus in inbred mice. *Behav. Brain Funct.* **2005**, *1*, 3. doi: 10.1186/1744-9081-1-3.
- 35. Nadel, L.; Hardt, O. Update on Memory Systems and Processes. *Neuropsychopharmacol.* **2011**, *36*, 251–273. doi: 10.1038/npp.2010.169.
- 36. Hajipour, S.; Khombi Shooshtari, M.; Farbood, Y.; Ali Mard, S.; Sarkaki, A.; Moradi Chameh, H.; Sistani Karampour, N.; Ghafouri, S. Fingolimod Administration Following Hypoxia-Induced Neonatal Seizure Can Restore Impaired Long-term Potentiation and Memory Performance in Adult Rats. *Neuroscience*. 2023, 519, 107–119. doi: 10.1016/j.neuroscience.2023.03.023.
- 37. Piešová, M.; Mach, M. Impact of perinatal hypoxia on the developing brain. *Physiol. Res.* **2020**, *69*(2), 199–213. doi: 10.33549/physiolres.934198.
- 38. Sab, I.M.; Ferraz, M.M.; Amaral, T.A.; Resende, A.C.; Ferraz, M.R.; Matsuura, C.; Brunini, T.M.; Mendes-Ribeiro, A.C. Prenatal hypoxia, habituation memory and oxidative stress. Pharmacol. *Biochem. Behav.* **2013**, 107, 24–28. doi: 10.1016/j.pbb.2013.04.004.
- 39. Robinson, S.; Petelenz, K.; Li, Q.; Cohen, M.L.; Dechant, A.; Tabrizi, N.; Bucek, M.; Lust, D.; Miller, R.H. Developmental changes induced by graded prenatal systemic hypoxic-ischemic insults in rats. *Neurobiol. Dis.* **2005**, *18*(3), 568–581. doi: 10.1016/j.nbd.2004.10.024.
- 40. Silvestro, S.; Calcaterra, V.; Pelizzo, G.; Bramanti, P.; Mazzon, E. Prenatal Hypoxia and Placental Oxidative Stress: Insights from Animal Models to Clinical Evidences. *Antioxidants (Basel)*. **2020**, *9*(*5*), 414. doi: 10.3390/antiox9050414.
- 41. Amakhin, D.V.; Soboleva, E.B.; Postnikova, T.Y.; Tumanova, N.L.; Dubrovskaya, N.M.; Kalinina, D.S.; Vasilev, D.S.; Zaitsev, A.V. Maternal Hypoxia Increases the Excitability of Neurons in the Entorhinal Cortex and Dorsal Hippocampus of Rat Offspring. Front. *Neurosci.* 2022, *16*, 867120. doi: 10.3389/fnins.2022.867120.
- 42. Cunha-Rodrigues, M.C.; Balduci, C.T.D.N.; Tenório, F.; Barradas, P.C. GABA function may be related to the impairment of learning and memory caused by systemic prenatal hypoxia-ischemia. *Neurobiol. Learn. Mem.* **2018**, 149, 20–27. doi: 10.1016/j.nlm.2018.01.004.

- 43. Wang, B.; Zeng, H.; Liu, J.; Sun, M. Effects of Prenatal Hypoxia on Nervous System Development and Related Diseases. *Front. Neurosci.* **2021**, *15*, 755554. doi: 10.3389/fnins.2021.755554.
- 44. Dubrovskaya, N.M.; Zhuravin, I.A. Ontogenetic characteristics of behavior in rats subjected to hypoxia on day 14 or day 18 of embryogenesis. *Neurosci. Behav. Physiol.* **2010**, 40(2), 231–238. doi: 10.1007/s11055-009-9235-2.
- 45. Nalivaeva, N.N.; Turner, A.J.; Zhuravin, I.A. Role of Prenatal Hypoxia in Brain Development, Cognitive Functions, and Neurodegeneration. *Front. Neurosci.* **2018**, *12*, 825. doi: 10.3389/fnins.2018.00825.
- 46. Zhuravin, I.A.; Dubrovskaya, N.M.; Vasilev, D.S.; Postnikova, T.Y.; Zaitsev, A.V. Prenatal hypoxia produces memory deficits associated with impairment of long-term synaptic plasticity in young rats. *Neurobiol. Learn. Mem.* **2019**, *164*, 107066. doi: 10.1016/j.nlm.2019.107066.
- 47. Belenichev, I.F.; Aliyeva, O.G.; Kamyshnyi, A.M. Long-term results of pharmacological correction of iNOS, eNOS, nNOS mRNA expression disorders in rat hippocampus after chronic prenatal hypoxia. Biol. Markers Fundam. *Clin. Med.* **2019**, *3*(2), 6–15. doi: 10.29256/v.03.02.2019.escbm02.
- 48. Belenichev, I.F.; Cherniy, V.I.; Nagornaya, E.A.; Bukhtiyarova, N.V.; Kucherenko, V.I. Neuroprotection and neuroplasticity; *Logos: Kiev*, **2015**, 510 p.
- 49. Zatsepina, O.G.; Evgen'ev, M.B.; Garbuz, D.G. Role of a Heat Shock Transcription Factor and the Major Heat Shock Protein Hsp70 in Memory Formation and Neuroprotection. *Cells* **2021**, *10*(7), 1638. doi: 10.3390/cells10071638.
- 50. Schlegel, C.; Liu, K.; Spring, B.; et al. Decreased expression of hypoxia-inducible factor  $1\alpha$  (HIF- $1\alpha$ ) in cord blood monocytes under anoxia. *Pediatr. Res.* **2023**, *93*, 870–877. doi: 10.1038/s41390-022-02193-7.
- 51. Shi, Y.J.; Ma, Z.Q.; Tang, J.W.; Zhao, Y.; Wang, X.; Liu, Q.; Wang, P.P.; John, C.; Chen, X.Q.; Du, J.Z. The integration of multiple signaling pathways provides for bidirectional control of CRHR1 gene transcription in rat pituitary cell during hypoxia. *Mol. Cell. Endocrinol.* **2017**, 454, 12–22. doi: 10.1016/j.mce.2017.05.031.
- 52. Wang, Q.; Zhang, L.; Zhu, C.; Lu, K.; Wu, J.; Liang, X.-F. Fosab, but not fosaa, plays an important role in learning and memory in fish—insights from zebrafish gene knockout study. Front. *Cell Dev. Biol.* **2025**, *13*, 1503066. doi: 10.3389/fcell.2025.1503066.
- 53. Lara Aparicio, S.Y.; Laureani Fierro, Á.d.J.; Aranda Abreu, G.E.; Toledo Cárdenas, R.; García Hernández, L.I.; Coria Ávila, G.A.; Rojas Durán, F.; Aguilar, M.E.H. Manzo Denes, J.; Chi-Castañeda, L.D.; et al. Current Opinion on the Use of c-Fos in Neuroscience. *NeuroSci* 2022, 3, 687–702. doi: 10.3390/neurosci3040050.
- 54. Katche, C.; Bekinschtein, P.; Slipczuk, L.; Goldin, A.; Izquierdo, I.A.; Cammarota, M.; Medina, J.H. Delayed wave of c-Fos expression in the dorsal hippocampus involved specifically in persistence of long-term memory storage. *Proc. Natl. Acad. Sci. U.S.A.* **2010**, *107*(1), 349–354. doi: 10.1073/pnas.0912931107.
- 55. Belenichev, I.F.; Aliyeva, E.G.; Kamyshny, O.M.; et al. Pharmacological Modulation of Endogenous Neuroprotection after Experimental Prenatal Hypoxia. *Neurochem. J.* **2022**, *16*, 68–75. doi: 10.1134/S1819712422010044.
- 56. Belenichev, I.F.; Sokolik, E.P.; Bukhtiyarova, N.V.; Levich, S.V. Pharmacological Modulation of Heat Shock Protein 70 (HSP70)—Dependent Mechanisms of Endogenous Neuroprotection in Conditions of Prenatal Chronic Alcoholism by Cerebrocurin and Tiocetam. Klinik Psikofarmakoloji Bülteni-Bull. *Clin. Psychopharmacol.* **2016**, *26*, 103–108.
- 57. Belenichev, I.; Popazova, O.; Bukhtiyarova, N.; Ryzhenko, V.; Pavlov, S.; Suprun, E.; Oksenych, V.; Kamyshnyi, O. Targeting Mitochondrial Dysfunction in Cerebral Ischemia: Advances in Pharmacological Interventions. *Antioxidants (Basel)* **2025**, *14*(1), 108. doi: 10.3390/antiox14010108.
- 58. Belenichev, I.F.; Mazur, I.A.; Kucherenko, L.I.; et al. The Molecular and Ultrastructural Aspects of the Formation of Mitochondrial Dysfunction in the Modeling of Chronic Cerebral Ischemia: The Mitoprotective Effects of Angiolin. *Neurochem. J.* **2016**, *10*, 131–136. doi: 10.1134/S1819712416010025.
- 59. Liu, Q.; Guo, J.; Hu, L.; Veronese, N.; Smith, L.; Yang, L.; Cao, C. Association between Intake of Energy and Macronutrients and Memory Impairment Severity in US Older Adults, National Health and Nutrition Examination Survey 2011–2014. *Nutrients* **2020**, *12*(11), 3559. doi: 10.3390/nu12113559.
- 60. Duarte, J.M.N. Loss of Brain Energy Metabolism Control as a Driver for Memory Impairment upon Insulin Resistance. *Biochem. Soc. Trans.* **2023**, *51*(1), 287–301. doi: 10.1042/BST20220789.
- 61. Sharma, C.; Kim, S.; Nam, Y.; Jung, U.J.; Kim, S.R. Mitochondrial Dysfunction as a Driver of Cognitive Impairment in Alzheimer's Disease. *Int. J. Mol. Sci.* **2021**, 22(9), 4850. doi: 10.3390/ijms22094850.
- 62. Chang, Y.F.; Gao, X.M. L-lysine is a Barbiturate-like Anticonvulsant and Modulator of the Benzodiazepine Receptor. *Neurochem. Res.* **1995**, *20*(*8*), *93*1–*93*7. doi: 10.1007/BF00970739.
- 63. Severyanova, L.A.; Lazarenko, V.A.; Plotnikov, D.V.; Dolgintsev, M.E.; Kriukov, A.A. L-Lysine as the Molecule Influencing Selective Brain Activity in Pain-Induced Behavior of Rats. *Int. J. Mol. Sci.* **2019**, 20, 1899. doi: 10.3390/ijms20081899.
- 64. Morimoto, R.I. Regulation of the Heat Shock Transcriptional Response: Cross Talk between a Family of Heat Shock Factors, Molecular Chaperones, and Negative Regulators. Genes Dev. 1998, 12(24), 3788–3796. doi: 10.1101/gad.12.24.3788.

- 65. Jiang, Y.Q.; Wang, X.L.; Cao, X.H.; Ye, Z.Y.; Li, L.; Cai, W.Q. Increased Heat Shock Transcription Factor 1 in the Cerebellum Reverses the Deficiency of Purkinje Cells in Alzheimer's Disease. *Brain Res.* **2013**, *1519*, 105–111. doi: 10.1016/j.brainres.2013.04.059.
- 66. Kondo, N.; Katsuno, M.; Adachi, H.; Minamiyama, M.; Doi, H.; Matsumoto, S.; Miyazaki, Y.; Iida, M.; Tohnai, G.; Nakatsuji, H.; Ishigaki, S.; Fujioka, Y.; Watanabe, H.; Tanaka, F.; Nakai, A.; Sobue, G. Heat Shock Factor-1 Influences Pathological Lesion Distribution of Polyglutamine-Induced Neurodegeneration. *Nat. Commun.* **2013**, *4*, 1405. doi: 10.1038/ncomms2417.
- 67. Kus-Liśkiewicz, M.; Polańska, J.; Korfanty, J.; Olbryt, M.; Vydra, N.; Toma, A.; Widłak, W. Impact of Heat Shock Transcription Factor 1 on Global Gene Expression Profiles in Cells Which Induce Either Cytoprotective or Pro-apoptotic Response Following Hyperthermia. *BMC Genomics* **2013**, *14*, 456. doi: 10.1186/1471-2164-14-456.
- 68. Erinjeri, A.P.; Wang, X.; Williams, R.; et al. HSF-1 Promotes Longevity through Ubiquilin-1-Dependent Mitochondrial Network Remodeling. *Nat. Commun.* **2024**, *15*, 9797. doi: 10.1038/s41467-024-54136-x.
- 69. Prince, T.L.; Lang, B.J.; Guerrero-Gimenez, M.E.; Fernandez-Muñoz, J.M.; Ackerman, A.; Calderwood, S.K. HSF1: Primary Factor in Molecular Chaperone Expression and a Major Contributor to Cancer Morbidity. *Cells* **2020**, *9*, 1046. doi: 10.3390/cells9041046.
- 70. Wu, L.; Hu, C.; Huang, M.; Jiang, M.; Lu, L.; Tang, J. Heat Shock Transcription Factor 1 Attenuates TNFα-Induced Cardiomyocyte Death through Suppression of NFκB Pathway. *Gene* **2013**, *527*(*1*), 89–94. doi: 10.1016/j.gene.2013.05.024.
- 71. Verma, P.; Pfister, J.A.; Mallick, S.; D'Mello, S.R. HSF1 Protects Neurons through a Novel Trimerizationand HSP-Independent Mechanism. *J. Neurosci.* **2014**, *34*(5), 1599–1612. doi: 10.1523/JNEUROSCI.3039-13.2014.
- 72. Zhang, Y.; Zhang, C.; Dai, Q.; Ma, R. Continuous Theta Burst Stimulation Inhibits Oxidative Stress-Induced Inflammation and Autophagy in Hippocampal Neurons by Activating Glutathione Synthesis Pathway, Improving Cognitive Impairment in Sleep-Deprived Mice. *Neuromolecular Med.* **2024**, *26*(1), 40. doi: 10.1007/s12017-024-08807-z.
- 73. Mazur, I.; Belenichev, I.; Kucherenko, L.; Siusiuka, V.; Reznichenko, N.; Kyryliuk, A.; Ryzhenko, V.; Shevchenko, A.; Khromylova, O. Thiotriazoline and its today. Monograph. *Dnipro, Jurfond,* **2025,** 384 p. ISBN 978-966-934-648-3.
- 74. Shevchenko, A.O.; Belenichev, I.F.; Kyryliuk, A.D. Premature Childbirth: Fundamental and Clinical Aspects. Monograph. *LAP LAMBERT Academic Publishing*, **2022**, 308 p. ISBN 978-620-5-52010-9.
- 75. Belenichev, I.F.; Vizir, V.A.; Mamchur, V.Y.; Kuriata, O.V. Place of Tiotriazoline in the Gallery of Modern Metabolitotropic *Medicines*. *Zaporozhye Medical Journal [Internet]*, **2019**. https://doi.org/10.14739/2310-1210.2019.1.155856.
- 76. Massaad, C.A.; Klann, E. Reactive Oxygen Species in the Regulation of Synaptic Plasticity and Memory. *Antioxid. Redox Signal.* **2011**, *14*(10), 2013–2054. doi: 10.1089/ars.2010.3208.
- 77. Popazova, O.; Belenichev, I.; Bukhtiyarova, N.; Ryzhenko, V.; Oksenych, V.; Kamyshnyi, A. Cardioprotective Activity of Pharmacological Agents Affecting NO Production and Bioavailability in the Early Postnatal Period after Intrauterine Hypoxia in Rats. Biomedicines 2023, 11, 2854. https://doi.org/10.3390/biomedicines11102854.
- 78. Popazova, O.; Belenichev, I.; Yadlovskyi, O.; Oksenych, V.; Kamyshnyi, A. Altered Blood Molecular Markers of Cardiovascular Function in Rats after Intrauterine Hypoxia and Drug Therapy. Curr. Issues Mol. Biol. 2023, 45, 8704–8715. https://doi.org/10.3390/cimb45110547.
- 79. Belenichev, I.; Ryzhenko, V.; Popazova, O.; Bukhtiyarova, N.; Gorchakova, N.; Oksenych, V.; et al. Optimization of the Search for Neuroprotectors among Bioflavonoids. Pharmaceuticals 2024, 17, 877. https://doi.org/10.3390/ph17070877.
- 80. Belenichev, I.; Bukhtiyarova, N.; Ryzhenko, V.; Makyeyeva, L.; Morozova, O.; Oksenych, V.; et al. Methodological Approaches to Experimental Evaluation of Neuroprotective Action of Potential Drugs. Int. J. Mol. Sci. 2024, 25, 10475. https://doi.org/10.3390/ijms251910475.
- 81. Belenichev, I.; Popazova, O.; Yadlovskyi, O.; Bukhtiyarova, N.; Ryzhenko, V.; Pavlov, S.; et al. Possibility of Using NO Modulators for Pharmacocorrection of Endothelial Dysfunction After Prenatal Hypoxia. Pharmaceuticals 2025, 18, 106. https://doi.org/10.3390/ph18010106.
- 82. Borrow, A.P.; Handa, R.J. Estrogen Receptors Modulation of Anxiety-Like Behavior. *Vitam. Horm.* **2017**, 103, 27–52. doi: 10.1016/bs.vh.2016.08.004.
- 83. Tongta, S.; Daendee, S.; Kalandakanond-Thongsong, S. Effects of Estrogen Receptor β or G Protein-Coupled Receptor 30 Activation on Anxiety-Like Behaviors in Relation to GABAergic Transmission in Stress-Ovariectomized Rats. *Neurosci. Lett.* **2022**, 789, 136885. doi: 10.1016/j.neulet.2022.136885.
- 84. Belenichev, I.F.; Odnokoz, O.V.; Pavlov, S.V.; et al. The Neuroprotective Activity of Tamoxifen and Tibolone during Glutathione Depletion in Vitro. *Neurochem. J.* **2012**, *6*, 202–212. doi: 10.1134/S181971241203004X.

85. Poladian, N.; Navasardyan, I.; Narinyan, W.; Orujyan, D.; Venketaraman, V. Potential Role of Glutathione Antioxidant Pathways in the Pathophysiology and Adjunct Treatment of Psychiatric Disorders. *Clin. Pract.* **2023**, *13*(4), 768–779. doi: 10.3390/clinpract13040070.

**Disclaimer/Publisher's Note:** The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.